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Chloride anion transporters inhibit growth of methicillin-resistant *Staphylococcus aureus* (MRSA) *in vitro*†

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A series of aminopyrrolic receptors were tested as anion transporters using POPC liposome model membranes. Many were found to be effective Cl⁻ transporters and to inhibit clinical strains of *Staphylococcus aureus* growth *in vitro*. The best transporters proved effective against the methicillin-resistant *Staphylococcus aureus* (MRSA) strains, Mu50 and HP1173. Tris-thiourea tren-based chloride transporters were also shown to inhibit the growth of *S. aureus in vitro*.

There is a tremendous interest in novel antibiotics that can combat highly resistant bacterial strains such as MRSA.¹ Recent studies of compounds that can mediate the transport of anions demonstrated significant biological activity. Several have shown antiproliferative activity in cancer cell lines.^{2,3} For instance, the strapped calixpyrrole (**1**) displays modest NaCl transport in liposome models and is able to inhibit growth of cancer cell lines *in vitro*.⁴ This effect was ascribed to a combination of receptor mediated chloride transport and sodium transport involving endogenous ion channels. This and other recent developments in the anion transport field^{5,6} have led us to consider that other biologically active species may be mediating their effect in whole or in part by an ability to transport chloride anions⁷ into cells. §¶

As a first test of this hypothesis, two known antibiotic agents, thiocarlidate and trichlorocarbanalidate were tested for their ability to transport chloride anions through a standard 1-palmitoyl-2-oleoyl-*sn*-glycero-3-phosphocholine (POPC) liposomal model membrane. They were found to be modest NaCl (or HCl||) co-transporters and



Fig. 1 Chloride efflux caused by the addition of the antibiotics thiocarlidate or trichlorocarbanalidate (0.675 mol%), with and without NaNO₃ added to the exterior of POPC vesicles loaded with 0.5 M NaCl, 0.1 M Na₂SO₄, and 20 mM phosphate buffer, pH = 7.2. 100% chloride efflux was determined by lysing the liposomes with Triton X.

effective Cl⁻/NO₃⁻ antiporters (Fig. 1). This finding, as well as the ability of several cation ionophores to act as antibiotics,⁸ has led us to consider that anion transporters might show antibiotic activity.



With this view in mind, we selected a family of aminopyrrolic compounds, namely **2–15** (Scheme 1), and investigated their chloride anion transport properties. The antibiotic activity of these compounds, as well as the known Cl⁻/HCO₃⁻ anion antiport agents **16–18** (Scheme 2), were also tested. The present study

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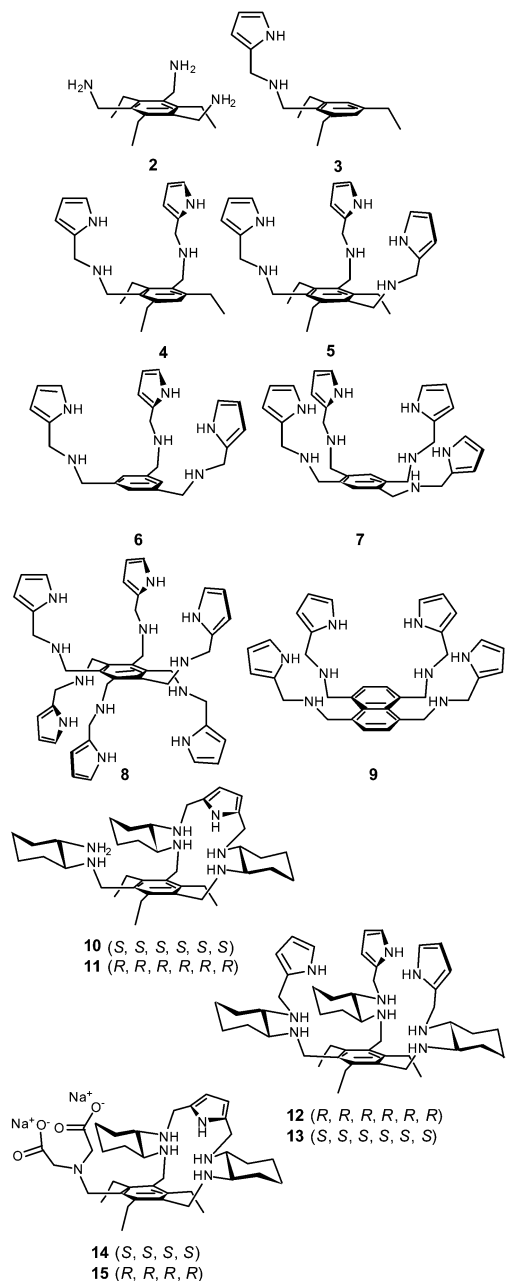
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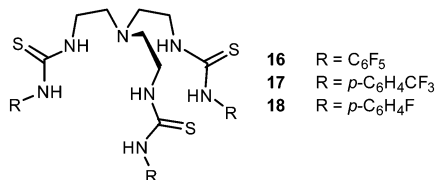
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Scheme 1 Structures of compounds 2–15.



Scheme 2 Structures of compounds 16–18.

was prompted by previous work, showing that several aminopyrrolic compounds were biologically active vs. pathogenic yeasts,¹⁰ exhibited antiviral activity,¹¹ induced apoptosis in HeLa and PLC/PRF/5 cells,¹² and were found to recognize the chloride anion well

in polar organic solvents.^{13,14} Here, we report the finding that for the aminopyrrolic compounds 2–15 chloride anion transport in liposomal model correlates well with *in vitro* antibiotic activity against the *S. aureus* strains UAMS1, HP1173, and Mu50.

Compounds 2–15 were examined for their ability to transport chloride anions across a phospholipid bilayer. In a typical experiment, 160 nm POPC vesicles that contained NaCl (500 mM), Na₂SO₄ (100 mM), and HEPES buffer (5 mM) were suspended in a solution of Na₂SO₄ (100 mM) and HEPES buffer (5 mM). The compound subject to study (0.675 mol%, compared to POPC concentration) was added to the solution and the rate of chloride efflux from the vesicles was monitored using a Cl⁻ ion selective electrode (ISE). After 9 minutes, the liposomes were lysed with polyethylene glycol *tert*-octylphenyl ether (Triton X-114), and a final reading from the ISE corresponding to 100% chloride release was taken. High levels of transport activity were seen in descending order for compounds 12, 13, 10, 8, and 11 (Fig. 2). In these studies the solutions external to the liposomes contained only SO₄²⁻ and HEPES. These are hydrophilic species¹⁵ that are typically not transported through POPC membranes.¹⁶ Therefore, we infer that under these experimental conditions, anion antiport should not contribute to the efflux of chloride from the liposome. Studies involving compound 5 were also carried out when the external sulphate anion was replaced by either NO₃⁻ or HCO₃⁻ (as the sodium salts). The nitrate and bicarbonate anions are considerably more lipophilic than the sulphate anion and can facilitate the transport of chloride through the liposomal membrane by promoting Cl⁻/NO₃⁻ or Cl⁻/HCO₃⁻ antiport processes. In these experiments a higher concentration of NaCl (1 M) inside the liposome was used to give a larger response with the ISE. The rate of chloride efflux is slightly raised on the addition of the more lipophilic anions nitrate and bicarbonate to the external solution (Fig. 3). Nevertheless, this difference only accounts for a small percentage of the total efflux of chloride from the liposomes. We thus conclude that anion antiport only plays a minor role in chloride efflux and that the receptors of the present study are functioning primarily as cation–anion cotransporters. ||



Fig. 2 Chloride efflux was initiated by the addition of 2–15 (0.675 mol%) to a solution of POPC vesicles (1 mM POPC, internal solution: 0.5 M NaCl, 0.1 M Na₂SO₄, 5 mM HEPES, pH = 7.2, external solution: 0.1 M Na₂SO₄, 5 mM HEPES pH = 7.2).





Fig. 3 Chloride efflux promoted by the addition of **5** (0.675 mol%) and either NaNO_3 or NaHCO_3 to the liposome solution (1 mM POPC, internal solution: 1 M NaCl, 0.1 M Na_2SO_4 , 5 mM HEPES, pH = 7.2, external solution: 0.1 M Na_2SO_4 , 5 mM HEPES pH = 7.2).



Fig. 4 A 96 well plate bacterial assay of compounds **18** (top row) and **17** (middle row) against *S. aureus* grown in 99 μL brain–heart infusion media and DMSO (bottom row) after 16 hours of bacterial growth. Each successive well (from left to right) represents a two-fold dilution of compound. Clear wells indicate inhibition of bacterial growth, while cloudy wells signify unhindered bacterial growth.

Studies of the antibiotic activity of compounds **2–18** were carried out on 96 well plates using the broth dilution method (*cf.* Fig. 4). Antibiotic activity was tested against both Gram positive (*S. aureus*) and Gram negative (*Pseudomonas aeruginosa*) bacteria. The bacterial laden plates were incubated for 16 hours at 37 °C. The plates were then examined visually. The wells that remained clear were deemed to reflect bacterial growth inhibition. Wells that were cloudy were considered to contain active colonies of bacteria. The MIC was taken as the concentration present in the last well that remained clear as judged in this manner. Experiments were repeated 3 times and gave concordant results within the range of a single serial dilution (Tables S1–S5, ESI†).

Good activity against *S. aureus* was seen for many compounds. However, except for compounds **5** and **8**, no activity was seen against Gram negative bacteria.** The most active aminopyrrole compounds proved to be in descending order **5**, **8**, **10**, **11**, **12**, **13**. The lowest activity was found for **2**, **14**, and **15** (Table 1). A plot of MIC vs. chloride anion transport revealed a general correlation with bacterial inhibition in *S. aureus* (Fig. 5). This trend was found to hold best for the compounds with intermediate activity.

While the ion transport rates correlate with the antibiotic activity of the compounds, the deviations of the most active compounds may reflect the fact that ion transport ability is only part of the story underlying the observed antibiotic activity. In particular, the best chloride anion transporters, **12** and **13** have significantly higher transport rates than compounds **10** and **11**, but only modestly better antibacterial activity. Previous studies of these compounds revealed that compounds **12** and **13** can bind to mannosides on the cell glycolyx,¹⁰ and that this may underlie their observed *in vitro* anticancer activity, as opposed to anion transport *per se*. Such findings provide support for the notion that the antibiotic activity of **12** and **13** may be due to the compounds acting by multiple mechanisms. Additionally, compounds **5** and **8** were the only compounds to be active against Gram negative bacteria. This leads to the conclusion that, at least in the case of compounds **5**, **8**, **12**, and **13**, ion transport is not the only factor responsible for antibiotic activity.

Nevertheless, we believe that an ability to promote the through-membrane transport of the chloride anion may be a useful harbinger of antibiotic activity. The fact that chloride anion exchange, rather than cation–anion cotransport, was seen for the known antibiotics, thiocarlide and trichlorocarbanalide (*cf.* Fig. 1), leads us to suggest that chloride anion transport can inhibit bacterial growth *in vitro*. Further evidence for this suggestion came from the finding that the synthetic thioureas **16–18** proved quite active *in vitro* (MIC = 1.23, 0.93, and 1.78 $\mu\text{g mL}^{-1}$ for the Mu50 *S. aureus*, respectively). These species are known to act primarily as anion exchangers (*e.g.*, Cl^- for HCO_3^- and Cl^- for SO_4^{2-}), rather than as ion pair cotransporters,^{2,16} although this class of transporter has been shown to be capable of dissipating pH gradients across lipid bilayers.^{2,17}

A notable feature of the present compounds is that they are active against HP1173 and Mu50 methicillin resistant strains of *S. aureus*, as well as the non-methicillin resistant UAMS1 strain. This leads us to suggest that receptors that can act as highly effective through-membrane anion carriers (both exchangers and ion pair cotransporters) may have a role to play as rationally designed antibiotic agents. Nevertheless, it is important to appreciate that the correlation between chloride anion transport and observed antibiotic activity found for **2–15** in the case of *S. aureus* is not perfect. While a general trend holds, deviations are seen in the case of the most active compounds. This may indicate that there are other mechanisms aside from ion

Table 1 MIC values for compounds **2–15** against the Mu50 strain of *S. aureus*. Average of ≥ 3 independent studies

Compound	2	3	4	5	6	7	8	9	10	11	12	13	14	15
MIC ($\mu\text{g mL}^{-1}$)	> 100	50	66.67	6.25	25	50	4.69	25	12.5	12.5	4.69	3.13	> 100	> 100





Fig. 5 Plot of chloride efflux promoted by 2–13 in liposomal transport models as detailed in Fig. 2 against their effectiveness in reducing the growth of the Mu50 (resistant) strain of *S. aureus*.

transport that are responsible for the antibacterial activity in *S. aureus* or that there are differences between the properties of the model membranes used in this study and the bacterial membrane.¹⁸ Further study of these and other anion transporters are on-going in an effort to elucidate more fully the underlying mechanisms of action and to develop more active receptor-based antibiotic agents.

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Notes and references

§ Simple cation transporters often function as antibiotics.⁸

¶ During the course of this work, Schmitzer and coworkers reported that certain imidazolium-based anion transporters likewise displayed strong antibiotic effects.⁹

|| The counter cation accompanying chloride anion egress was not monitored directly. The observation of a cation dependence on the rate when the inner solution was changed from NaCl to KCl so as to provide a less hydrated cation¹⁴ is consistent with a Na⁺/Cl⁻ cotransport mechanism (see Supporting Information). However, the presence of readily

protonated amino functionality in receptors 2–15 makes it difficult to discount an alternative mechanism involving H⁺/Cl⁻ cotransport. Through-membrane proton transport is expected to be most important at early times before a proton gradient is built up across the liposomal membrane. An effort to distinguish between limiting NaCl vs. HCl cotransport mechanisms has not been made in the case of thiocarlide and trichlorocarbanalide.

** Although further study is needed, it is likely that artificial receptors such as those of the present study will prove more effective as both ion carriers and antibiotics in the case of Gram positive bacteria than Gram negative bacteria since Gram positive bacteria have a single cellular membrane, whereas Gram negative bacteria have 2 cell membranes.

- M. E. Stryjewski and G. R. Corey, *Clin. Infect. Dis.*, 2014, **58**, S10–S19.
- N. Busschaert, M. Wenzel, M. E. Light, P. Iglesias-Hernández, R. Pérez-Tomás and P. A. Gale, *J. Am. Chem. Soc.*, 2011, **133**, 14136–14148.
- (a) S. J. Moore, M. Wenzel, M. E. Light, R. Morley, S. J. Bradberry, P. Gómez-Iglesias, V. Soto-Cerrato, R. Pérez-Tomás and P. A. Gale, *Chem. Sci.*, 2012, **3**, 2501–2509; (b) S. J. Moore, C. J. E. Haynes, J. González, J. L. Sutton, S. J. Brooks, M. E. Light, J. Herniman, G. J. Langley, V. S. Cerrato, R. Pérez-Tomás, I. Marques, P. J. Costa, V. Félix and P. A. Gale, *Chem. Sci.*, 2013, **4**, 103–117.
- S.-K. Ko, S. K. Kim, A. Share, V. M. Lynch, J. Park, W. Namkung, W. V. Rossom, N. Busschaert, P. A. Gale, J. L. Sessler and I. Shin, *Nat. Chem.*, 2014, **6**, 885–892.
- N. Busschaert and P. A. Gale, *Angew. Chem., Int. Ed.*, 2013, **52**, 1374–1382.
- A. V. Jentzsch, A. Henning, J. Mareda and S. Matile, *Acc. Chem. Res.*, 2013, **46**, 2791–2800.
- (a) H. Li, H. Valkenier, L. W. Judd, P. R. Brotherhood, S. Hussain, J. A. Cooper, O. Jurček, H. A. Sparkes, D. N. Sheppard and A. P. Davis, *Nat. Chem.*, 2014, **8**, 24–32; (b) J. T. Davis, O. Okunola and R. Quesada, *Chem. Soc. Rev.*, 2010, **39**, 3843–3862; (c) G. W. Gokel and S. Negin, *Acc. Chem. Res.*, 2013, **46**, 2824–2833.
- D. A. Kevin II, D. A. F. Meujo and M. T. Hamann, *Expert Opin. Drug Discovery*, 2009, **4**, 109–146.
- C. R. Elie, G. David and A. R. Schmitzer, *J. Med. Chem.*, 2015, **58**, 2358–2366.
- C. Nativi, O. Francesconi, G. Gabrielli, I. De Simone, B. Turchetti, T. Mello, L. D. C. Mannelli, C. Ghelardini, P. Buzzini and S. Roelens, *Chem. – Eur. J.*, 2012, **18**, 5064–5072.
- O. Francesconi, C. Nativi, G. Gabrielli, I. De Simone, S. Noppen, J. Balzarini, S. Liekens and S. Roelens, *Chem. – Eur. J.*, 2015, **21**, 10089–10093.
- S.-H. Park, Y. P. Choi, J. Park, A. Share, O. Francesconi, C. Nativi, W. Namkung, J. L. Sessler, S. Roelens and I. Shin, *Chem. Sci.*, 2015, **6**, 7284–7292.
- S. Roelens, A. Vacca and C. Venturi, *Chem. – Eur. J.*, 2009, **15**, 2635–2644.
- S. Roelens, A. Vacca, O. Francesconi and C. Venturi, *Chem. – Eur. J.*, 2009, **15**, 8296–8302.
- Y. Marcus, *J. Chem. Soc., Faraday Trans.*, 1991, **87**, 2995–2999.
- N. Busschaert, L. E. Karagiannidis, M. Wenzel, C. J. E. Haynes, N. J. Wells, P. G. Young, D. Makuc, J. Plavec, K. A. Jolliffe and P. A. Gale, *Chem. Sci.*, 2014, **5**, 1118–1127.
- X. Wu, L. W. Judd, E. N. W. Howe, A. M. Withecombe, V. Soto-Cerrato, H. Li, N. Busschaert, H. Valkenier, R. Pérez-Tomás, D. N. Sheppard, Y. Jiang, A. P. Davis and P. A. Gale, *Chem*, 2016, DOI: 10.1016/j.chempr.2016.04.002.
- M. J. Spooner and P. A. Gale, *Chem. Commun.*, 2015, **51**, 4883–4886.

