

CrossMark  
click for updatesCite this: *Chem. Sci.*, 2017, 8, 1134

# Reversible ratiometric detection of highly reactive hydropersulfides using a FRET-based dual emission fluorescent probe†

Ryosuke Kawagoe, Ippei Takashima, Shohei Uchinomiya and Akio Ojida\*

Hydropersulfide (R–SSH) is an important class of reactive sulfur species (RSS) involved in a variety of physiological processes in mammals. A fluorescent probe capable of real-time detection of hydropersulfide levels in living cells would be a versatile tool to elucidate its roles in cell signalling and redox homeostasis. In this paper, we report a ratiometric fluorescent probe for hydropersulfide sensing, based on a fluorescence resonance energy transfer (FRET) mechanism. This sensing mechanism involves a nucleophilic reaction of a hydropersulfide with the pyronine-unit of the probe, which modulates the intramolecular FRET efficiency to induce a dual-emission change. The reversible nature of this reaction allows us to detect increases and decreases of hydropersulfide levels in a real-time manner. The probe fluorometrically sensed highly reactive hydropersulfides, such as  $\text{H}_2\text{S}_2$  and Cys-SSH, while the fluorescence response to biologically abundant cysteine and glutathione was negligible. Taking advantage of the reversible and selective sensing properties, this probe was successfully applied to the ratiometric imaging of concentration dynamics of endogenously produced hydropersulfides in living cells.

Received 29th August 2016  
Accepted 23rd September 2016

DOI: 10.1039/c6sc03856e

[www.rsc.org/chemicalscience](http://www.rsc.org/chemicalscience)

## Introduction

Reactive sulfur species (RSS) are a family of sulfur-containing molecules endogenously produced in biological systems. Among RSS, hydropersulfides (R–SSHs) such as hydrogen persulfide ( $\text{H}_2\text{S}_2$ ) and cysteine hydropersulfide (CysSSH) are increasingly recognized as an important class of RSS that modulate a variety of physiological events in mammals. For instance, hydropersulfides are found to play a role in *S*-sulfhydration of Cys residues of proteins as signalling molecules.<sup>1,2</sup> Previous reports proposed that protein *S*-sulfhydration is mediated by the reaction of hydrogen sulfide ( $\text{H}_2\text{S}$ ) with oxidised cysteine residues such as *S*-sulfenic acid (*S*-OH) and *S*-nitrosothiol (SNO).<sup>3–7</sup> However, recent reports suggested that hydropersulfides serve as the main reactive species that directly *S*-sulfhydrate numerous proteins.<sup>8–13</sup> In particular, it has been revealed that *S*-sulfhydration regulates the functions of important classes of proteins involved in cell redox homeostasis,<sup>8–10</sup> metabolism,<sup>11</sup> and signal transduction.<sup>12</sup> Meanwhile, Akaike suggested that hydropersulfides such as *S*-sulfhydrated glutathione (GSSH) serve as potent reducing agents in redox signalling, and may provide a primary and potent antioxidant defence in cells.<sup>1</sup>

Hydropersulfides (R–SSHs) are generated by different pathways in biological systems. It has been reported that hydrogen persulfide ( $\text{H}_2\text{S}_2$ ) can be formed by the oxidation of endogenous  $\text{H}_2\text{S}$  by reactive oxygen species (ROS).<sup>14</sup> Akaike found that CysSSH is biosynthesized from cystine (CysS-SCys) by two major enzymes: cystathionine  $\beta$ -synthetase (CBS) and cystathionine  $\gamma$ -lyase (CSE).<sup>1,15</sup> He also proposed that the enzymatically generated CysSSH is converted to GSH-based hydroper-/hydropolysulfides (*e.g.*, GSSH, GSSSH, *etc.*) through persulfide interchange reactions. Meanwhile, Banerjee proposed that sulfide oxidation pathways in mitochondria are the important source of RSS, such as GSSH.<sup>16</sup> Despite the extensive study of the hydropersulfide formation pathways, regulation of their levels in cells, especially their reducing mechanism, remains largely elusive. Recent reports proposed thioredoxin (Trx) as an important enzyme that reduces CysSSH, although clear evidence for this process has not yet been provided.<sup>17</sup>

To understand the varied roles of hydropersulfides in biological systems, it is critical to develop a new analytical tool that allows us to detect the formation and consumption of these RSS species. Fluorescent probes which are available for real-time cell imaging could meet this requirement.<sup>18–23</sup> In this regard, Xian *et al.* recently reported a series of selective fluorescent probes for hydroper-/hydropolysulfides ( $\text{H}_2\text{S}_n$ ,  $n > 1$ ).<sup>18–20</sup> They ingeniously exploited the high nucleophilic activity of  $\text{H}_2\text{S}_n$  to develop reaction-based turn-on fluorescence probes, which were successfully applied to the visualization of intracellular  $\text{H}_2\text{S}_n$ . However, due to the irreversible nature of the reactions, it was intrinsically difficult to monitor reversible concentration

Graduate School of Pharmaceutical Sciences, Kyushu University, 3-1-1 Maidashi, Higashi-ku, Fukuoka, 812-8582, Japan

† Electronic supplementary information (ESI) available: Full experimental procedures, synthesis and characterization of compounds. See DOI: 10.1039/c6sc03856e

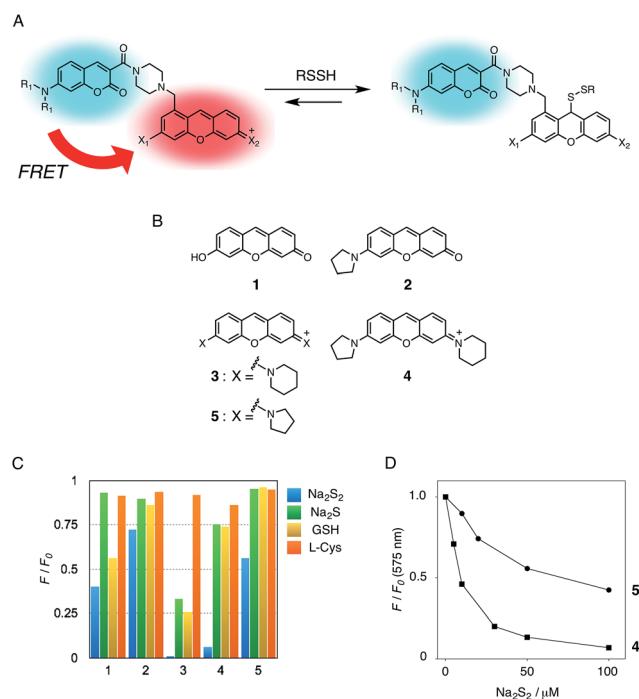


dynamics of intracellular  $H_2S_n$  using these probes. In this paper, we report the development of a ratiometric fluorescent probe for detecting hydropersulfides, based on intramolecular fluorescence resonance energy transfer (FRET) (Fig. 1A).<sup>24</sup> The sensing mechanism of this probe involves a reversible nucleophilic attack of a highly reactive hydropersulfide species on the pyronine fluorophore. This adduct formation disrupts the conjugation structure of the xantheno ring, decreasing the intramolecular FRET efficiency due to a change in the spectral overlap between the coumarin fluorescence (FRET donor) and the xantheno absorbance (FRET acceptor), which causes a clear dual-emission signal change. Taking advantage of this reversible sensing property, the probe was successfully applied to detect the concentration dynamics of hydropersulfides in living cells, demonstrating the utility of the probe as a chemical tool in RSS research.

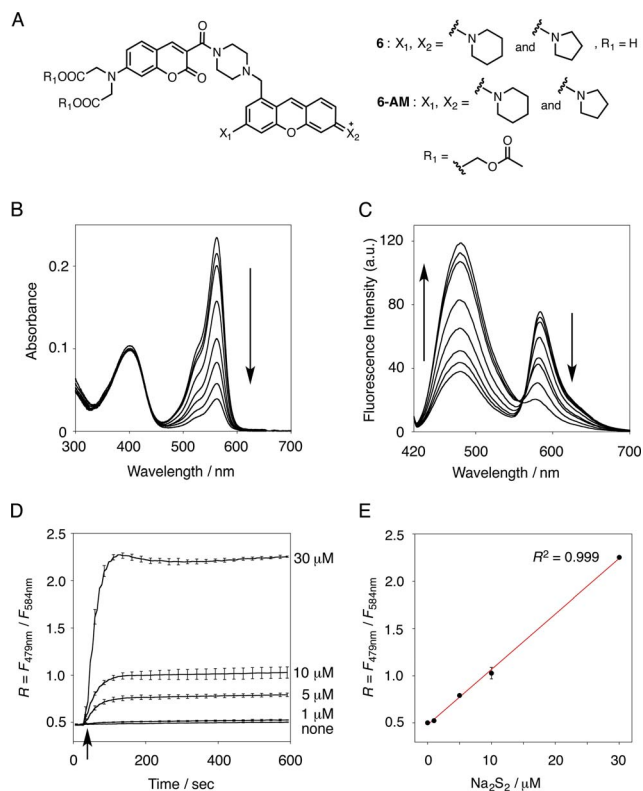
## Results and discussion

### Molecular design of the probe

In the previous study, we reported that the fluorescence of the xantheno derivative **1** significantly decreased upon addition of a large excess of glutathione (GSH,  $\sim 10$  mM) under neutral aqueous conditions.<sup>25</sup> The spectroscopic analyses revealed that



**Fig. 1** (A) Mechanism of the FRET-based ratiometric fluorescence sensing of hydropersulfide. (B) Structure of the xantheno derivatives. (C) Fluorescence quenching efficiency ( $F/F_0$ ) of **1**–**5** upon addition of  $Na_2S_2$  (50  $\mu$ M),  $Na_2S$  (50  $\mu$ M), GSH (5 mM), and L-Cys (1 mM). The data were measured 10 min after addition of the thiol compounds. Measurement conditions: [probe] = 5  $\mu$ M in 50 mM HEPES, 10 mM NaCl, 1 mM  $MgSO_4$ , pH 7.4, 25  $^\circ$ C.  $\lambda_{ex}$  = 480 nm (**1**), 500 nm (**2**), 530 nm (**3**, **4**, **5**). (D) Fluorescence titration profile of **4** (■) and **5** (●) upon addition of  $Na_2S_2$  (0–100  $\mu$ M). Measurement conditions: [probe] = 5  $\mu$ M in 50 mM HEPES, 10 mM NaCl, 1 mM  $MgSO_4$ , pH 7.4, 25  $^\circ$ C.  $\lambda_{ex}$  = 530 nm.



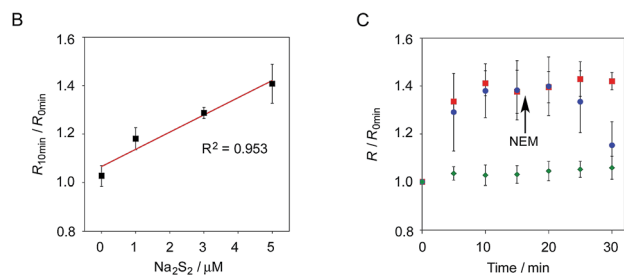
**Fig. 2** (A) Structures of probe **6** and **6-AM**. (B and C) Absorption and fluorescence spectral changes of **6** (5  $\mu$ M) upon addition of  $Na_2S_2$  (0–100  $\mu$ M). Measurement conditions: [**6**] = 5  $\mu$ M in 50 mM HEPES, 10 mM NaCl, 1 mM  $MgSO_4$ , 0.4% Tween, pH 7.4, 25  $^\circ$ C.  $\lambda_{ex}$  = 410 nm. (D) Time-dependent change of the ratio value ( $R = F_{479\text{ nm}}/F_{584\text{ nm}}$ ) of **6** (5  $\mu$ M) upon addition of  $Na_2S_2$  (0–30  $\mu$ M).  $Na_2S_2$  was added at 30 s, as indicated by the arrow. Measurement conditions: [**6**] = 5  $\mu$ M in 50 mM HEPES, 10 mM NaCl, 1 mM  $MgSO_4$ , 0.4% Tween, pH 7.4, 25  $^\circ$ C.  $\lambda_{ex}$  = 410 nm.  $n = 3$ . (E) Plot of the ratio value  $R (F_{479\text{ nm}}/F_{584\text{ nm}})$  600 s after addition of  $Na_2S_2$  (0–30  $\mu$ M).

xantheno **1**, which lacks a C9 aromatic substituent unlike fluorescein, was susceptible to nucleophilic attack by thiol species and readily converted to a non-fluorescent adduct. Since hydropersulfides (RSSHs) are more nucleophilic than stable thiols such as GSH and  $H_2S$ ,<sup>26</sup> we thought that this reaction-based fluorescence quenching could be exploited for selective detection of hydropersulfides. As an initial attempt, we synthesized a series of xantheno derivatives bearing the different substituents (Fig. 1B), and evaluated their fluorescence responses toward several biological thiol species. The results are summarized in Fig. 1C and S1.† Compound **1** showed a marked decrease in fluorescence ( $F/F_0 = 40\%$ ) upon treatment with 50  $\mu$ M sodium disulfide ( $Na_2S_2$ ), the extent of which is much larger than that induced by addition of the same concentration of  $Na_2S$  ( $F/F_0 = 93\%$ ) and a high concentration (1 mM) of cysteine ( $F/F_0 = 91\%$ ). However, **1** also responded to a biologically relevant concentration of GSH (5 mM) with a high quenching efficiency ( $F/F_0 = 56\%$ ), indicative of the low selectivity of **1** among biologically relevant thiols. The rhodol-type compound **2** exhibited a rather non-selective weak fluorescence response to the thiol species. The pyronine-type compound **3**,





Scheme 1 Synthesis of probe 6.



**Fig. 4** (A) Fluorescence images of A549 cells treated with **6-AM** (5  $\mu\text{M}$ ). (a)  $F_{430-480}$ , (b)  $F_{550-630}$ , (c) DIC, and (d) ratio image ( $R = F_{430-480}/F_{550-630}$ ). (e) and (f) Ratio image after 30 min in the presence and absence of  $\text{Na}_2\text{S}_2$  (5  $\mu\text{M}$ ), (g) and (h) ratio image after 15 min in the presence  $\text{Na}_2\text{S}_2$  (5  $\mu\text{M}$ ) and subsequent treatment with NEM (100  $\mu\text{M}$ ) for 15 min. Scale bar: 30  $\mu\text{m}$ . (B) Ratio value change of A549 cells upon the addition of various concentrations of  $\text{Na}_2\text{S}_2$  (0–5  $\mu\text{M}$ ),  $n$  = 6. (C) Time trace plot of the ratio value in A549 cells upon the treatment with  $\text{Na}_2\text{S}_2$  (5  $\mu\text{M}$ ) (red square,  $n$  = 6),  $\text{Na}_2\text{S}_2$  (5  $\mu\text{M}$ ) and the subsequent addition of NEM (100  $\mu\text{M}$ ) at 15 min (blue circle,  $n$  = 4), without  $\text{Na}_2\text{S}_2$  and NEM (green diamond,  $n$  = 6).





**Fig. 5** Ratiometric detection of endogenously produced hydropersulfide in A549 cells. (A) Production and degradation pathways of hydropersulfides in cells. (B) Ratio images of A549 cells treated with 6-AM (5  $\mu$ M) (a) before addition of cystine, (b) 30 min after addition of cystine (200  $\mu$ M), (c) 30 min after addition of cystine (200  $\mu$ M) in the presence of AOAA (1 mM). (C) Time trace plot of the ratio value ( $R = F_{430-480}/F_{550-630}$ ) change in A549 cells upon treatment with cystine (200  $\mu$ M) in the absence (red circle,  $n = 6$ ) and presence of AOAA (1 mM) (blue square,  $n = 6$ ). (D) Comparison of the ratio value  $R$  change in A549 cells upon treatment with cysteine for 30 min: (1) control (without cysteine),  $n = 6$ , (2) cystine (200  $\mu$ M),  $n = 6$ , (3) cystine (200  $\mu$ M) in the presence of AOAA (1 mM),  $n = 4$ , (4) cystine (200  $\mu$ M) in the presence of auranofin (2  $\mu$ M),  $n = 3$ . (E) Ratio images of A549 cells treated with 6-AM (5  $\mu$ M) (a) before addition of L-cysteine, (b) 30 min after addition of L-cysteine (200  $\mu$ M), (c) 30 min after addition of L-cysteine (200  $\mu$ M) in the presence of AOAA (1 mM). (F) Time trace plot of the ratio value  $R$  change in A549 cells upon treatment with L-cysteine (200  $\mu$ M) in the absence (red circle,  $n = 3$ ) and presence of AOAA (1 mM) (blue square,  $n = 3$ ). (G) Comparison of the ratio value  $R$  change in A549 cells upon treatment with L-cysteine for 30 min: (1) control (without cysteine),  $n = 6$ , (2) L-cysteine (200  $\mu$ M),  $n = 4$ , (3) L-cysteine (200  $\mu$ M) in the presence of AOAA (1 mM),  $n = 3$ , (4) L-cysteine (200  $\mu$ M) in the presence of auranofin (2  $\mu$ M),  $n = 5$ . (H) Time trace plot of the ratio value  $R$  change in A549 cells upon treatment with L-cysteine (200  $\mu$ M) (red square,  $n = 3$ ) and upon treatment with L-cysteine (200  $\mu$ M) and NaOCl (300  $\mu$ M) (blue circle,  $n = 3$ ). NaOCl was added 15 min after addition of L-cysteine. Conditions:  $\lambda_{\text{ex}} = 405$  nm,  $R = F_{430-480}/F_{550-630}$  nm. Scale bar: 30  $\mu$ m. \* $P < 0.05$ , \*\* $P < 0.01$ .

possessing two six-membered piperidine rings, is highly susceptible to thiol species with significant fluorescence quenching efficiencies ( $F/F_0 < 30\%$ ), except for L-cysteine. However, pyronines 4 and 5, which possess one and two five-membered pyrrolidine rings, respectively, showed selective fluorescence responses ( $F/F_0 = 6\%$  and  $56\%$ , respectively) toward  $\text{Na}_2\text{S}_2$  (50  $\mu\text{M}$ ). The formation of the  $\text{H}_2\text{S}_2$  adduct with 5 was confirmed by a  $^1\text{H-NMR}$  experiment (Fig. S2†). The fluorescence response of 4 and 5 toward  $\text{Na}_2\text{S}_2$  was further evaluated by titration with different concentrations of  $\text{Na}_2\text{S}_2$ . As shown in

Fig. 1D and S3,† 4 was more sensitive than 5 and showed a substantial decrease in fluorescence with a low concentration of  $\text{Na}_2\text{S}_2$  (below 10  $\mu\text{M}$ ). The varied fluorescence response of these pyronine-type probes, depending on the substituents, would be reasonably explained by the different electron donating abilities of the cyclic amines. That is, the five-membered pyrrolidine can act as a stronger electron donating substituent than the six-membered piperidine,<sup>25</sup> so that the tolerance to nucleophilic attack by  $\text{Na}_2\text{S}_2$  is in the order of  $5 > 4 > 3$ .



We selected pyronine **4** as a fluorescent subunit of the ratiometric probe for hydropersulfides, on account of its selective and sensitive detection of  $\text{Na}_2\text{S}_2$ , as shown in Fig. 1. The structure of the newly designed dual-emission probe **6** is shown in Fig. 2A. The probe possesses a coumarin as the FRET donor, which is conjugated to a pyronine unit as the FRET acceptor through a rigid linker. The two carboxylate groups are introduced into the coumarin unit in order to increase the hydrophilicity of the probe, which prevents its leakage from cells during imaging experiments. The synthesis of probe **6** is shown in Scheme 1. The radical bromination of **7** with *N*-bromosuccinimide (NBS) and the subsequent nucleophilic reaction with *N*-Boc-piperazine yielded **8**. After the deprotection, **8** was converted to bis-triflate **10**, which was sequentially reacted with pyrrolidine and piperidine to give **11** as a mixture of the substitution isomers. The keto-reduction of **11** with borane-SMe<sub>2</sub> and the subsequent oxidation using DDQ yielded the pyronine **13**. After the removal of the Boc group, **13** was subjected to a conjugation reaction with the *N*-hydroxysuccinimide ester of coumarin **14** to give **15**. Finally, the deprotection of the *tert*-butyl ester groups of **15** and the following HPLC purification provided **6** as a mixture of the isomers.

### Ratiometric fluorescence sensing of hydropersulfides

The functional analysis of probe **6** was initially conducted in a neutral aqueous solution. A solution of **6** (5  $\mu\text{M}$ ) in 50 mM HEPES buffer (pH 7.4) showed two distinct UV peaks at 400 nm and 562 nm (Fig. 2B), which correspond to the absorbance of coumarin and pyronine, respectively. Upon titration with sodium disulfide  $\text{Na}_2\text{S}_2$  (0–100  $\mu\text{M}$ ), the absorbance at 562 nm decreased gradually to ca. 25% of its original intensity, whereas the absorbance at 400 nm scarcely changed. In the fluorescence spectrum, a solution of **6** (5  $\mu\text{M}$ ) showed two distinct emissions at 479 nm and 584 nm due to the coumarin and pyronine units, respectively, when excited at 410 nm (Fig. 2C). This dual-emission spectrum changed dramatically in a see-saw manner upon addition of  $\text{Na}_2\text{S}_2$  (0–100  $\mu\text{M}$ ). The large decrease in emission at 584 nm and the concomitant increase in emission at 479 nm strongly suggest that FRET between the coumarin and xanthen units is cancelled as a result of a decrease in the spectral overlap between the coumarin emission and the pyronine absorption (Fig. S4†). The FRET efficiency of **6** was calculated to be 60% in the initial state, which decreased to 31% in the presence of 100  $\mu\text{M}$   $\text{Na}_2\text{S}_2$ . Fig. 2D shows the time-lapse detection of the fluorescence response of **6** toward  $\text{Na}_2\text{S}_2$ . The ratio value  $R$  ( $F_{479\text{ nm}}/F_{584\text{ nm}}$ ) rapidly increased upon addition of  $\text{Na}_2\text{S}_2$  (0–30  $\mu\text{M}$ ) and reached a plateau almost within 60 s. The plot of  $R$  value against the concentration of  $\text{Na}_2\text{S}_2$  (0–30  $\mu\text{M}$ ) shows a linear relationship (Fig. 2E), indicative of the highly quantitative nature of the ratiometric detection of hydropersulfides using **6**. Probe **6** was able to detect as low as 1.0  $\mu\text{M}$   $\text{Na}_2\text{S}_2$  based on the calculation of the detection limit ( $3\sigma$ ). This sensitivity is sufficiently high for detection of intracellular hydropersulfides, the concentration of which was reported to be around ten micromolar under basal conditions.<sup>1</sup>

The sensing selectivity of **6** for various biologically relevant thiol species was evaluated (Fig. 3A). In contrast to the large increase in  $R$  value induced by  $\text{Na}_2\text{S}_2$  (lane 3), **6** showed a negligible fluorescence response upon addition of the same concentration of  $\text{Na}_2\text{S}$  (lane 2). This sensing selectivity is reasonably ascribed to the lower  $\text{pK}_a$  value of  $\text{H}_2\text{S}_2$  ( $\text{pK}_a = 5.0$ ) compared to that of  $\text{H}_2\text{S}$  ( $\text{pK}_a = 6.9$ ), rendering  $\text{H}_2\text{S}_2$  highly nucleophilic as a thiolate anion ( $\text{HS}_2^-$ ) under neutral conditions (pH = 7.4) (Fig. 3B).<sup>26</sup> A moderate  $R$  value change was observed upon addition of hydropolysulfide  $\text{Na}_2\text{S}_4$  (lane 4), though this change was apparently smaller than that induced by  $\text{Na}_2\text{S}_2$  (lane 3). The weak fluorescence response of **6** for  $\text{Na}_2\text{S}_4$  might be ascribed to the rather poor nucleophilic activity of the  $\text{HS}_4^-$  anion due to its extremely low  $\text{pK}_a$  value ( $\text{pK}_a = 3.8$ ).<sup>26</sup> Probe **6** also showed a moderate increase in the  $R$  value upon treatment with CysSSH (lane 5) and GSSH (lane 6), which were generated *in situ* from Cys and GSH, respectively, by the reaction with  $\text{Na}_2\text{S}$  and NO donor (NOC7).<sup>1</sup> In a similar manner, the mixture of  $\text{Na}_2\text{S}$  (50  $\mu\text{M}$ ) and NaOCl (50  $\mu\text{M}$ ) in a basic solution (0.1 M NaOH), which generates  $\text{Na}_2\text{S}_2$  *in situ*, induced a moderate increase of  $R$  value (lane 7), while their mixture in a neutral solution (50 mM HEPES, pH 7.4), which mainly produces  $\text{Na}_2\text{S}_4$  and  $\text{Na}_2\text{S}_5$ ,<sup>14</sup> resulted in a small increase in the  $R$  value (lane 8). This difference in signal change is consistent with the results obtained by the direct titration with these thiols (lanes 3 and 4). Probe **6** showed a negligible  $R$  value change upon addition of biologically relevant concentrations of GSH (5 mM, lane 9), *L*-cysteine (1 mM, lane 10), and cystine (0.5 mM, lane 11). It is noteworthy that the sensing selectivity of **6** for  $\text{Na}_2\text{S}_2$  is apparently higher than that of **5** among these thiol species (Fig. 1C), probably due to steric hindrance and/or the electron configuration effect of the coumarin unit conjugated to the pyronine unit of **6**. Therefore, **6** was able to detect  $\text{Na}_2\text{S}_2$  with a sufficient sensitivity (detection limit = 4.4  $\mu\text{M}$ ) even in the presence of 5 mM of GSH (Fig. S5†). We also confirmed that **6** scarcely responded to various redox-relevant compounds, including ROS such as NaOCl and  $\text{H}_2\text{O}_2$ , reactive nitrogen species (NOS), ascorbic acid, and KCN (Fig. S6†).

All of these data suggest that probe **6** primarily serves as a selective fluorescent probe for hydropersulfides. It was confirmed that **6** did not show a significant ratio change over the physiological pH range (5.0 to 8.5; Fig. S7†).

The fluorescence response of **6** towards a change in the hydropersulfide level was examined. As shown in Fig. 3C, the  $R$  value of **6** (5  $\mu\text{M}$ ) increased stepwise upon the repeated addition of 30  $\mu\text{M}$   $\text{Na}_2\text{S}_2$ , and reached a plateau almost within 60 s. The subsequent addition of *N*-ethylmaleimide (NEM, 500  $\mu\text{M}$ ) to consume  $\text{Na}_2\text{S}_2$  induced a large decrease in the  $R$  value. These data clearly indicate that **6** can reversibly change its  $R$  value depending on the concentration of hydropersulfide according to the binding equilibrium shown in Fig. 1A. The reverse fluorescence response of **6** was further evaluated upon addition of other reactive species (Fig. 3D). Addition of NaOCl (100  $\mu\text{M}$ ) to the mixed solution of **6** (5  $\mu\text{M}$ ) and  $\text{Na}_2\text{S}_2$  (50  $\mu\text{M}$ ) also induced a significant decrease in the  $R$  value (lane 3), as observed with NEM (lane 4). This change is reasonably ascribed to the decrease in  $\text{Na}_2\text{S}_2$  level due to the formation of oxidized sulfurs



and hydropolysulfides ( $\text{H}_2\text{S}_n$ ,  $n > 2$ ). Conversely, a small change in the  $R$  value was induced upon addition of GSH (5 mM, lane 5) and  $\text{l-Cys}$  (1 mM, lane 6), suggesting that these biologically abundant thiols do not interfere with the hydropersulfide sensing of **6**.

### Ratio imaging of hydropersulfides in living cells

We next applied probe **6** to the ratiometric fluorescence imaging of hydropersulfides ( $\text{R-SSH}$ ) in living cells. For cell imaging, **6** was chemically modified with acetoxymethyl (AM) groups to enhance its membrane permeability.<sup>27</sup> It was expected that the AM-modified probe, **6-AM** (Fig. 2A), could be readily hydrolyzed by intracellular esterases to liberate **6**, which is unlikely to leak from cells due to its highly polar character. When A549 cells were treated with **6-AM** (5  $\mu\text{M}$ ) for 20 min, bright fluorescence of the probe was observed in the cytosolic region of the cells (Fig. 4A(a–d)). A cell viability assay revealed that **6-AM** showed low cytotoxicity to A549 cells at a concentration below 25  $\mu\text{M}$  (Fig. S8†). Addition of 5  $\mu\text{M}$   $\text{Na}_2\text{S}_2$  to the cell medium induced an obvious change in the fluorescence ratio ( $R = 430\text{--}480\text{ nm}/550\text{--}630\text{ nm}$ ) (Fig. 4A(e)) of cells, relative to that observed in the control experiment without  $\text{Na}_2\text{S}_2$  treatment (Fig. 4A(f)). Titration of the **6-AM** pre-stained cells with  $\text{Na}_2\text{S}_2$  (0–5  $\mu\text{M}$ ) showed a linear dependence between  $\text{Na}_2\text{S}_2$  concentration and  $R$  value (Fig. 4B and S9†), demonstrating the accurate hydropersulfide sensing property of **6** in living cells. The time-lapse imaging revealed that the  $R$  value increased immediately after the addition of 5  $\mu\text{M}$   $\text{Na}_2\text{S}_2$  and reached a steady state ( $R \approx 1.4$ ) after 10 min (Fig. 4C). The subsequent treatment of the cells with 100  $\mu\text{M}$   $N$ -ethylmaleimide (NEM) induced a significant decrease in the  $R$  value (Fig. 4A(g, h) and C) due to the decrease of the hydropersulfide level by the nucleophilic reaction with NEM, demonstrating the reversible sensing property of **6** in living cells.

The probe **6-AM** was further applied to the ratiometric detection of endogenous hydropersulfides produced by enzymes in living cells. It has been reported that CSE and CBS are the major enzymes responsible for generation of cysteine hydropersulfide ( $\text{CysSSH}$ ) from cystine ( $\text{CysSSCys}$ ) in A549 cells.<sup>1</sup> When A549 cells, pre-stained with **6-AM**, were treated with cystine ( $\text{CysSSCys}$ ), a gradual increase in the  $R$  value was observed (Fig. 5B and C). This change was effectively suppressed on treatment of the cells with aminooxyacetic acid (AOAA), an inhibitor of CSE and CBS (Fig. 5B and D, lane 3).<sup>28</sup> Conversely, treatment of the cells with auranofin, an inhibitor for thio-redoxin reductase (TrxR), induced a statistically significant increase in the  $R$  value (Fig. 5D, lane 4 and S10†), suggesting an increase in the intracellular hydropersulfide level as a result of the inhibition of Trx activation by TrxR (Fig. 5A). This result is consistent with the recent report that Trx catalyses reduction of  $\text{Cys-SH}$  as a major regulator of its intracellular level.<sup>17</sup> It has been reported that hydropersulfide is also produced in living cells by the oxidation of  $\text{H}_2\text{S}$ , which is generated from  $\text{l-Cys}$  by CSE and CBS.<sup>20,22</sup> To confirm this point, fluorescence imaging of A549 cells treated with  $\text{l-Cys}$  (200  $\mu\text{M}$ ) was performed using **6-AM**. As shown in Fig. 5E and F, the  $R$  value largely increased in

a time-dependent manner in the living cells. Treatment of the cells with AOAA effectively suppressed the increase in  $R$  value induced by  $\text{l-Cys}$  (Fig. 5E and G, lane 3). Unlike the case of the cysteine experiment (Fig. 5D), inhibition of TrxR by auranofin did not cause an increase in the  $R$  value (Fig. 5G, lane 4 and S11†), implying that the direct conversion of  $\text{l-Cys}$  to  $\text{CysSSH}$  is not a major pathway of the hydropersulfide formation in living cells. All of these data suggest that hydropersulfides are generated from  $\text{l-Cys}$  through the enzyme-mediated  $\text{H}_2\text{S}$  formation and subsequent oxidation in living cells. Finally, the addition of high concentration  $\text{NaOCl}$  (300  $\mu\text{M}$ ) induced a significant decrease in the  $R$  value in cells (Fig. 5H and S12†), indicating that the decrease in hydropersulfide level was a result of its oxidative degradation to oxidized sulfurs and hydrogen polysulfides ( $\text{H}_2\text{S}_n$ ,  $n > 2$ ).

## Conclusion

In conclusion, we have developed a ratiometric fluorescent probe **6** that can visualize the endogenously produced hydropersulfides in living cells. To our knowledge, probe **6** is the first example of a ratiometric fluorescent probe that can reversibly detect intracellular hydropersulfide levels. Since the research field of reactive sulfur species (RSS) is still in its infancy, probe **6** would serve as a versatile analytical tool, not only for understanding the chemical nature of hydropersulfide in biological systems, but also for elucidating the roles of hydropersulfide in cell signalling and redox homeostasis. For this purpose, further functional improvements in probe **6**, which include sensing selectivity for a single hydropolysulfide species and/or localizability to a certain cell compartment, would be desirable to realize more precise and quantitative analysis of RSS. Research along these lines is currently underway in our laboratory.

## Experimental section

### Synthesis and characterization of the compounds

The syntheses and characterization of probes **1–6** and **6-AM** are described in the ESI.†

### Fluorescence measurement

Fluorescence titration was conducted with a solution (3 mL or 0.5  $\mu\text{L}$ ) of the probe in a quartz cell. Typically, a freshly prepared aqueous stock solution of  $\text{Na}_2\text{S}_2$  was added to a solution of **6** (5  $\mu\text{M}$ ) in 50 mM HEPES, 10 mM NaCl, 1 mM  $\text{MgSO}_4$ , pH 7.4 with 0.4% Tween at 25 °C and the fluorescence emission spectra were measured after 10 min ( $\lambda_{\text{ex}} = 410\text{ nm}$ ) with a Perkin Elmer LS55 fluorescence spectrometer.

### Cell culture

A549 cells were cultured in high glucose Dulbecco's Modified Eagle's Medium (DMEM, Gibco) supplemented with 10% fetal bovine serum (FBS, Gibco) and 1% antibiotic-antimycotic solution (Gibco) at 37 °C under a humidified atmosphere of 5%  $\text{CO}_2$  in air. A subculture was performed every 3–4 days from subconfluent (<80%) cultures using a trypsin–EDTA solution.



For the fluorescence bioimaging, cells were cultured for 2 days in a 35 mm glass-bottomed dish (Iwaki Scitech).

### Fluorescence imaging of exogenous H<sub>2</sub>S<sub>2</sub> in A549 cells

Fluorescence imaging was conducted with a confocal laser scanning microscope (LSM 780, Zeiss) equipped with a 63× objective lens. The following detection channels were chosen for the ratiometric imaging; Ch1  $\lambda_{\text{ex}} = 405$  nm,  $\lambda_{\text{em}} = 430$ –480 nm, and Ch2  $\lambda_{\text{ex}} = 405$  nm,  $\lambda_{\text{em}} = 550$ –630 nm. In a glass-based dish, A549 cells in HBS buffer (107 mM NaCl, 6 mM KCl, 1.2 mM MgSO<sub>4</sub>, 2.0 mM CaCl<sub>2</sub>, 11.5 mM glucose, 20 mM HEPES, pH 7.4) were incubated with 6-AM (5  $\mu\text{M}$ ) for 20 min at 37 °C under a humidified atmosphere of 5% CO<sub>2</sub> in air. After removal of excess probe and washing with HBS buffer, the cells were treated with Na<sub>2</sub>S<sub>2</sub> (0–5  $\mu\text{M}$ , final conc.) and subjected to the fluorescence imaging. For the imaging of the endogenously produced hydropersulfides, A549 cells, pre-stained with 6-AM (5  $\mu\text{M}$ ), were treated with cystine (200  $\mu\text{M}$ ) or L-cysteine (200  $\mu\text{M}$ ). For inhibition of CSE and CBS enzymes, the cells were pre-treated with AOAA (aminooxyacetic acid, 1 mM, Sigma) in HBS buffer for 1 h before staining with 6-AM. For inhibition of TrxR, the cells were pre-treated with auranofin (2  $\mu\text{M}$ , Wako) in HBS buffer for 1 h before staining with 6-AM.

## Acknowledgements

We appreciate the technical support from the Research Support Center, Graduate School of Medical Sciences, Kyushu University. A. O. acknowledges the Naito Foundation and Toray Science Foundation for their financial support. R. K. acknowledges the Japan Society for the Promotion of Science (JSPS) Research Fellowships for Young Scientists. This work was performed under the Cooperative Research Program of “Network Joint Research Center for Materials and Devices” organized by MEXT, Japan.

## Notes and references

- 1 T. Ida, T. Sawa, H. Ihara, Y. Tsuchiya, Y. Watanabe, Y. Kumagai, M. Suematsu, H. Motohashi, S. Fujii, T. Matsunaga, M. Yamamoto, K. Ono, N. O. Devarie-Baez, M. Xian, J. M. Fukuto and T. Akaike, *Proc. Natl. Acad. Sci. U. S. A.*, 2014, **111**, 7606–7611.
- 2 B. D. Paul and S. H. Snyder, *Nat. Rev. Mol. Cell Biol.*, 2012, **13**, 499–507.
- 3 C. E. Paulsen and K. S. Carroll, *Chem. Rev.*, 2013, **113**, 4633–4679.
- 4 T. V. Mishanina, M. Libiad and R. Banerjee, *Nat. Chem. Biol.*, 2015, **11**, 457–464.
- 5 A. K. Mustafa, M. M. Gadalla, N. Sen, S. Kim, W. Mu, S. K. Gazi, R. K. Barrow, G. Yang, R. Wang and S. H. Snyder, *Sci. Signaling*, 2009, **2**, ra72.
- 6 M. R. Filipovic, J. L. Miljkovic, T. Nauser, M. Royzen, K. Klos, T. Shubina, W. H. Koppenol, S. J. Lippard and I. Ivanovic-Burmazovic, *J. Am. Chem. Soc.*, 2012, **134**, 12016–12027.
- 7 D. Zhang, I. Macinkovic, N. O. Devarie-Baez, J. Pan, C.-M. Park, K. S. Carroll, M. R. Filipovic and M. Xian, *Angew. Chem., Int. Ed.*, 2014, **53**, 575–581.
- 8 N. Sen, B. D. Paul, M. M. Gadalla, A. K. Mustafa, T. Sen, R. Xu, S. Kim and S. H. Snyder, *Mol. Cell*, 2012, **45**, 13–24.
- 9 M. Nishida, T. Sawa, N. Kitajima, K. Ono, H. Inoue, H. Ihara, H. Motohashi, M. Yamamoto, M. Suematsu, H. Kurose, A. van der Vliet, B. A. Freeman, T. Shibata, K. Uchida, Y. Kumagai and T. Akaike, *Nat. Chem. Biol.*, 2012, **8**, 714–724.
- 10 S. Koike, Y. Ogasawara, N. Shibuya, H. Kimura and K. Ishii, *FEBS Lett.*, 2013, **587**, 3548–3555.
- 11 T. V. Mishanina, P. K. Yadav, D. P. Ballou and R. Banerjee, *J. Biol. Chem.*, 2015, **290**, 25072–25080.
- 12 Y. Kimura, Y. Mikami, K. Osumi, M. Tsugane, J. Oka and H. Kimura, *FASEB J.*, 2013, **27**, 2451–2457.
- 13 R. Greiner, Z. Pálkás, K. Bäsell, D. Becher, H. Antelmann, P. Nagy and T. P. Dick, *Antioxid. Redox Signaling*, 2013, **19**, 1749–1765.
- 14 P. Nagy and C. C. Winterbourn, *Chem. Res. Toxicol.*, 2010, **23**, 1541–1543.
- 15 P. K. Yadav, M. Martinov, V. Vitvitsky, J. Seravalli, R. Wedmann, M. R. Filipovic and R. Banerjee, *J. Am. Chem. Soc.*, 2016, **138**, 289–299.
- 16 M. Libiad, P. K. Yadav, V. Vitvitsky, M. Martinov and R. Banerjee, *J. Biol. Chem.*, 2014, **289**, 30901–30910.
- 17 R. Wedmann, C. Onderka, S. Wei, I. András Szijártó, J. Miljkovic, A. Femic, M. Lange, S. Savitsky, P. Kumar, R. Torregrossa, E. G. Harrer, T. Harrer, I. Ishii, M. Gollasch, M. E. Wood, E. Galardon, M. Xian, M. Whiteman, R. Banerjee and M. Filipovic, *Chem. Sci.*, 2016, **7**, 3414–3426.
- 18 W. Chen, C. Liu, B. Peng, Y. Zhao, A. Pacheco and M. Xian, *Chem. Sci.*, 2013, **4**, 2892–2896.
- 19 C. Liu, W. Chen, W. Shi, B. Peng, Y. Zhao, M. Xian, H. Ma and M. Xian, *J. Am. Chem. Soc.*, 2014, **136**, 7257–7260.
- 20 W. Chen, E. W. Rosser, T. Matsunaga, A. Pacheco, T. Akaike and M. Xian, *Angew. Chem., Int. Ed.*, 2015, **54**, 13961–13965.
- 21 L. Zeng, S. Chen, T. Xia, W. Hu, C. Li and Z. Liu, *Anal. Chem.*, 2015, **87**, 3004–3010.
- 22 M. Gao, F. Yu, H. Chen and L. Chen, *Anal. Chem.*, 2015, **87**, 3631–3638.
- 23 X. Han, F. Yu, X. Song and L. Chen, *Chem. Sci.*, 2016, **7**, 5098–5107.
- 24 J. R. Lakowicz, *Principles of Fluorescence Spectroscopy*, Springer, New York, 3rd edn, 2006.
- 25 R. Kawagoe, I. Takashima, K. Usui, A. Kanegae, Y. Ozawa and A. Ojida, *ChemBioChem*, 2015, **16**, 1608–1615.
- 26 K. Ono, T. Akaike, T. Sawa, Y. Kumagai, D. A. Wink, D. J. Tantillo, A. J. Hobbs, P. Nagy, M. Xian, J. Lin and J. M. Fukuto, *Free Radical Biol. Med.*, 2014, **77**, 82–94.
- 27 R. Y. Tsien, *Nature*, 1981, **290**, 527–528.
- 28 A. Asimakopoulou, P. Panopoulos, C. T. Chasapis, C. Coletta, Z. Zhou, G. Cirino, A. Giannis, C. Szabo, G. A. Spyroulias and A. Papapetropoulos, *Br. J. Pharmacol.*, 2013, **169**, 922–932.

