



Cite this: *Nanoscale*, 2024, **16**, 12820

Advances in engineered nanosystems: immunomodulatory interactions for therapeutic applications

Rupam Khatua,^a Bibrita Bhar,^{†a} Souradeep Dey,^b Chitra Jaiswal,^a Victoria J^a and Biman B. Mandal^{*,a,b,c}

Advances in nanotechnology have led to significant progress in the design and fabrication of nanoparticles (NPs) with improved therapeutic properties. NPs have been explored for modulating the immune system, serving as carriers for drug delivery or vaccine adjuvants, or acting as therapeutics themselves against a wide range of deadly diseases. The combination of NPs with immune system-targeting moieties has facilitated the development of improved targeted immune therapies. Targeted delivery of therapeutic agents using NPs specifically to the disease-affected cells, distinguishing them from other host cells, offers the major advantage of concentrating the therapeutic effect and reducing systemic side effects. Furthermore, the properties of NPs, including size, shape, surface charge, and surface modifications, influence their interactions with the targeted biological components. This review aims to provide insights into these diverse emerging and innovative approaches that are being developed and utilized for modulating the immune system using NPs. We reviewed various types of NPs composed of different materials and their specific application for modulating the immune system. Furthermore, we focused on the mechanistic effects of these therapeutic NPs on primary immune components, including T cells, B cells, macrophages, dendritic cells, and complement systems. Additionally, a recent overview of clinically approved immunomodulatory nanomedicines and potential future perspectives, offering new paradigms of this field, is also highlighted.

Received 17th February 2024,

Accepted 27th May 2024

DOI: 10.1039/d4nr00680a

rsc.li/nanoscale

1. Introduction

The immune system plays a crucial role in the surveillance of our body. The system consists of specialized cells, tissues, and organs working together to encounter the threats posed by deadly diseases, including influenza, human immunodeficiency virus (HIV), SARS-CoV-2 infections, atherosclerosis, cancers, multiple sclerosis (MS), systemic lupus erythematosus (SLE), diabetes, *etc.* Conventionally, the immune system is divided into two components, referred to as (a) innate and (b) adaptive components. The innate immune system consists of phagocytes (dendritic cells (DCs) and macrophages) and granulocytes (neutrophils, eosinophils, basophils, and mast

cells), contributing to the formation of the first line of defence in the body. DCs and macrophages, as well as other host cells, including epithelial cells, fibroblasts, and endothelial cells, play primary roles in the recognition of pathogens during the innate immune response. They recognize pathogen-associated molecular patterns (PAMPs) or damage-associated molecular patterns (DAMPs) *via* pattern recognition receptors (PRRs) and get activated quickly to recruit themselves to the assault sites (infected, inflamed, and damaged tissues).¹ The classical, lectin, and alternative pathways of the complement system are the additional components of the innate immune system. The classical pathway is activated by antigen-antibody interaction; the lectin pathway *via* microbial molecules (mannose residues) by soluble mannose-binding lectins; and the alternative pathway by any recognizing surfaces.² On the other hand, the adaptive immune component governs the activities of T cells and B cells that together confer “specificity” and “memory” to the immune responses. In response to pathogenic changes in the body, the innate and adaptive immune cells collectively drive immunological responses and restore normal physiological activities. Aberrant immune implications can result in either

^aBiomaterials and Tissue Engineering Laboratory, Department of Biosciences and Bioengineering, Indian Institute of Technology Guwahati, Guwahati – 781039, Assam, India. E-mail: biman.mandal@iitg.ac.in, mandal.biman@gmail.com

^bCentre for Nanotechnology, Indian Institute of Technology Guwahati, Guwahati – 781039, Assam, India

^cJyoti and Bhupat Mehta School of Health Sciences and Technology, Indian Institute of Technology Guwahati, Guwahati – 781039, Assam, India

[†]These authors contributed equally.

immune suppression or overactivation of immune components. In many cases, the hyperactive cellular and molecular components of the immune system play pathogenic roles, thereby promoting disease progression, while individuals also suffer from immunosuppressive diseases due to the compromised activities of T cells, B cells, macrophages, DCs, and other immune components. Importantly, it has been observed that even different phenotypes of a specific immune cell are also engaged in different sets of diseases. For instance, M1 phenotype macrophages play proinflammatory roles (implicated in autoimmune disorders), while M2 phenotypes are associated with anti-inflammatory responses (implicated in immunosuppressive disorders). Thus, understanding the functionalities of the immune system and clinically harnessing the modulation of immune components are imperative in the fight against a variety of immune disorders.

Advances in science and nanotechnology now allow us to manipulate cellular and molecular immune components. Progress in biomedical science has led to the development of engineered nanostructured materials for therapeutic delivery. Due to their high surface area-to-volume ratio, nanoparticles (NPs) exert greater biological interaction with respect to their bulk counterparts.³ This enhanced biological interaction thereby augments better therapeutic efficacy by concentrating the therapeutic cargo at pathogenic sites. In combination with specific therapeutic agents, NPs can be employed in both activation as well as suppression of immune responses. To achieve immune-specific therapeutic delivery, the surface of NPs can be functionalized with antibodies, peptides, oligosaccharides, antigens, *etc.*^{4–6} The clinical and preclinical data also suggest that immunotherapeutic nanomedicines are rapidly emerging and promising novel therapeutic platforms for targeting diseases characterized by aberrant immune functions. This review discusses, in detail, the fundamental properties of various types of NPs and their interaction with physiological and immune components, as well as their contribution in targeting immune components for therapeutic applications. Furthermore, we illustrate how these therapeutic nanomedicines mechanistically act on different components of the immune system, including T cells, B cells, DCs, macrophages, and complement systems. We also provided a current update on clinically approved nanomedicines with their immunomodulatory properties. Furthermore, a comprehensive overview of the challenges in the field and future perspectives is presented, elucidating avenues for further research and development.

2. The interplay between fundamental morphological features of NPs and immune systems

The interactions between fundamental morphological features of NPs and the immune components, including size, shape, surface charge, and modifications, are crucial in determining

their responses within biological systems (Fig. 1). Understanding fundamental features of different NPs and their interactions with immune components is essential in order to rationally design nanomedicines for optimized therapeutic outcomes.

2.1. Size

Size is a key factor that governs the pharmacokinetic and pharmacodynamic behavior of therapeutic NPs. Particles smaller than 5 nm undergo either rapid renal clearance or get cleared by the extravasation process.^{7,8} Increasing the size of NPs beyond a certain limit leads to their accumulation and deposition in multiple organs and tissues. The particles greater than 200 nm in diameter activate and provoke the complement system, which can lead to rapid removal of the NPs from circulatory systems and accumulation in organs like the liver and spleen.^{9–11} Although the cellular uptake of NP depends on cell types, it has been observed that particles of 50 nm in size get internalized by the cells with higher efficiency at a greater uptake rate.¹² 100–200 nm rigid and spherical NPs exhibit prolonged circulation time, avoid hepatic uptake, and are also protected from being engulfed by the spleen cells.¹³ The NPs of 20–200 nm in size exhibit greater accumulation in tumor tissues by avoiding the reticuloendothelial system and renal filtration, which helps in achieving greater therapeutic concentration at the tumor site.^{14,15} Therefore, variations in particle size trigger immune responses differentially depending on the immune cell types. In comparison with small particles, large particles induce greater immune responses by locating themselves in immune cells and delivering therapeutic payloads.¹⁶ On the other hand, the smaller particles have also been reported to trigger potent immune responses by modulating helper T-cell subtypes compared with the larger NPs.¹⁷ Thus, multiple studies suggest that 20–200 nm is the most effective size range that can be considered in the development of nanotherapeutics.¹⁸

2.2. Shape

The variation in shape governs different cellular uptake patterns of NPs by mammalian cells and influences their systemic circulation and binding affinity behavior. Cell membranes exhibit different sets of cellular responses with particle shape alterations due to changes in membrane integrity.¹⁹ Hence, altering the shape of NPs can improve their therapeutic outcomes.²⁰ In particular, *in vitro* cellular study based on a macrophage cell line, RAW264.7, revealed that the uptake efficiency of triangular particles is greater than that of rod and star-shaped particles.²¹ The rod-shaped and spherical particles can also induce different sub-populations of helper T cells. *In vivo* results depicted that spherical particles favor the helper T cell 1 (Th-1) subtypes, while rod-shaped particles promote Th-2 cell-induced immune responses.¹⁷ The shape variations have significant effects on adjuvanticity and cytokine production. The specific adjuvant engineered on gold nanorods and nanospheres provoked the release of different levels of inflammatory cytokines. In comparison with nanospheres, the nanorods displayed a

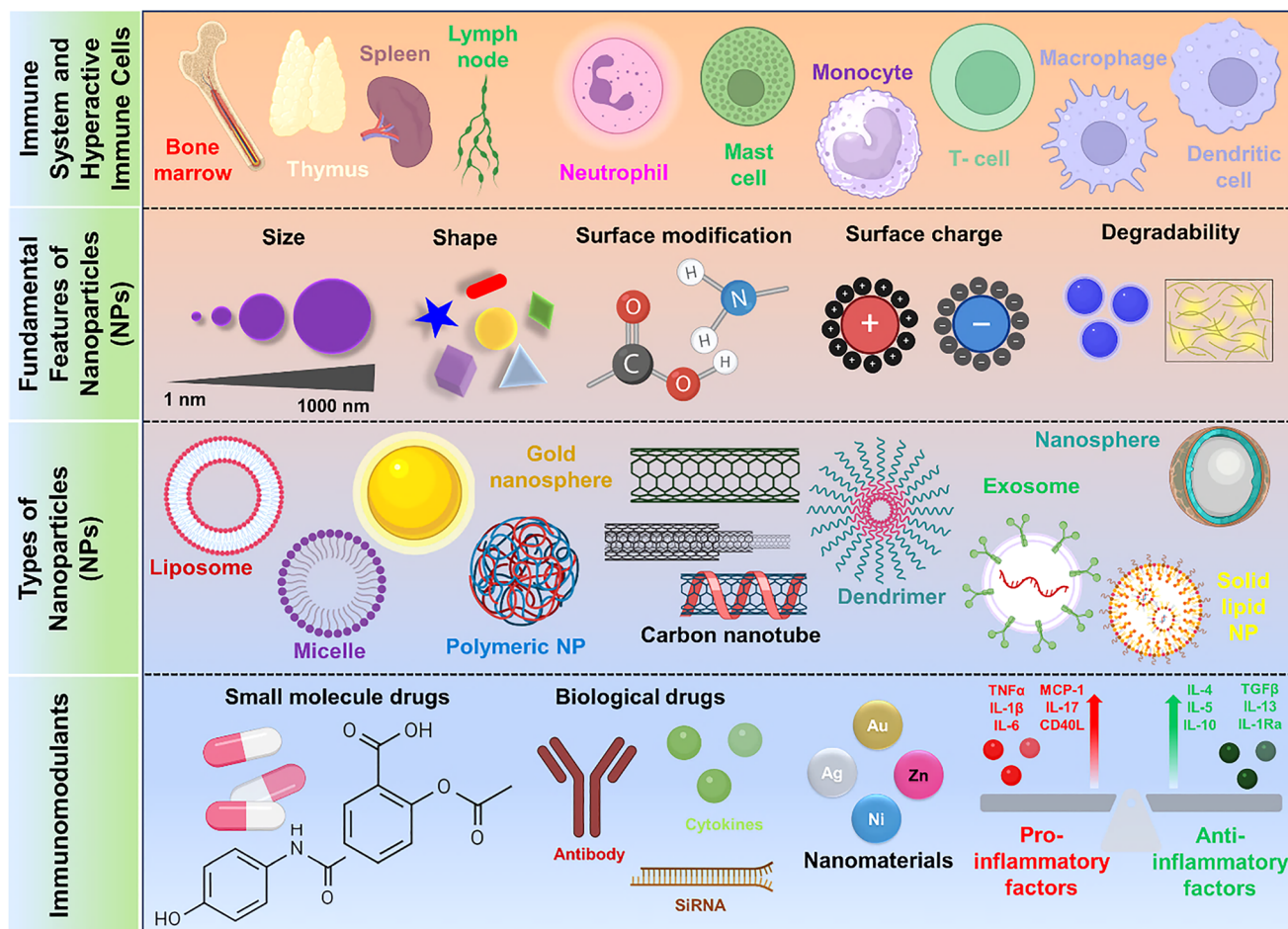


Fig. 1 Schematic illustration depicting immune components and fundamental attributes of nanosystems in modulating immune response for immunotherapy. Image created with BioRender.com.

greater ability to potentiate the adjuvanticity of chemical species with minimal inflammatory cytokine production.²² Even at the same therapeutic doses, the antibody-coated nanorod exerts multi-fold greater therapeutic efficacy against breast cancer cells than the nanospheres.²³ Contradictory to these results, a recent study also demonstrated that the mammalian cell studied with spherical gold NP exhibited better uptake efficacy in comparison with rod-shaped NPs.²⁴

2.3. Surface charge

The charge on surface of the NP is very crucial for its bioactivity, and its variation induces different sets of biological responses. NPs with a positive charge show more rapid cellular uptake than neutral and negatively charged NPs.^{25,26} Negatively charged cell membranes promote the cellular internalization of positively charged NPs.¹² However, positive charge also affects the structural integrity of the phospholipid bilayer of the plasma membrane, and greater charge density leads to disordering of the phospholipid bilayer structure.²⁷ The positive and negative charges of the NPs also reflect their mode of entry into the cells. Positively charged particles follow the macropinocytosis mode of entry, while negatively charged

particles make their entry into the cell *via* the clathrin- or caveolae-independent endocytosis pathway.²⁸ Differently charged particles with similar size and shape provoke varied immune responses. Positively charged NPs show a greater influence on the activation of antigen-presenting cells (APCs) like DCs and macrophages than uncharged particles. Thus, the loading of an anionic species like nucleic acids or other polymers can mask the extent of positive charge of the particles that slackens the undesirable immunogenic outputs.^{29,30} The positively charged gold NPs (AuNPs) stimulated the monocyte cells of immune systems at a significant level by inducing the expressions of pro-inflammatory cytokine (IL-1 β) and anti-inflammatory cytokine (TGF- β), while the negatively charged AuNPs promoted pro-inflammatory (TNF- α) expression.³¹ The deleterious immune responses that are produced by cationic charges can be advantageous in immunotherapeutic applications. Recent studies suggested that the positive charge can play beneficial roles in the development of vaccines where the cationic surface moieties can potentiate the therapeutic efficacies of adjuvants like ovalbumin (OVA) *via* complement activation, T-cell activation, enhanced antibody production, and cytokine secretion.³²

2.4. Surface modification

The surface modification of NPs is carried out to improve biocompatibility and blood circulation time, targeting the disease pathology, and achieving greater therapeutic retention in desired sites with minimal toxicity hazards.³³ In achieving specific nano-bio interaction as well as targeted delivery of therapeutics, the NP surface can be modified with a variety of ligands such as polyethylene glycol (PEG), polyethyleneimine (PEI), cell-penetrating peptides, targeting antibodies, antigens, *etc.*^{34–37} PEG is a chemically inert hydrophilic polymer widely used for surface modification, which helps in preventing direct exposure of the NP surface to the biosystems and inhibits particle aggregation, opsonization, and phagocytosis.³⁸ The phagocytosis of NPs by macrophages is considered one of the major clinical limitations that occurs in the development of therapeutics. Surface modification with antibodies has proved to be an excellent strategy to target immune components and prevent macrophage-mediated phagocytosis. Besides PEG, CD47 antibody conjugation with NPs can selectively block the signal regulatory protein- α (SIRP- α), which is found on macrophages. The CD47-SIRP- α coupling interaction and subsequent blocking of SIRP- α target protein with CD47 antibody-modified nanoplatfoms prevents macrophagic phagocytosis of the NPs.³⁷ CD11c monoclonal antibody has also been used as a surface-modifying agent to bind to DCs *via* specific intercellular adhesion molecules, such as 3-grabbing-non-integrin (SIGN) surface proteins.³⁹ Hence, CD11c antibody modification of NPs loaded with immunosuppressive agents can deliver therapeutic cargo to DCs by specifically targeting the DC-SIGN.³⁹ Tailoring the surface of NPs with antigen molecules is a key strategy in the development of nano-vaccines. OVA, the model antigen, can be attached to the NP surface, and the delivery of OVA-conjugated nano-vaccine showed antigen-induced immunological responses.⁴⁰ Furthermore, the modification of NPs with amine functional groups enables the OVA-loaded nano-vaccines to activate the complement systems at a significant level.³² Recently, cytokine modifications have been made in the surface engineering of NPs to target specific components of the immune system. Interleukin-2 receptors (IL-2R) that are expressed on the surface of T cells can be targeted to deliver therapeutics precisely and specifically. In this regard, the IL-2 cytokine can be used as a surface-modifying ligand to mediate IL-2R-dependent T-cell targeting and delivery of immunotherapeutic agents.⁴¹ Chemical species such as hyaluronic acid, folic acid, and biotins have been engineered onto the NP surface to specifically bind with the respective CD44 and biotin receptors that are overexpressed in cancer cells. Thus, modifying the particle surfaces with these ligands can selectively eliminate the cancer cells while minimally affecting the normal and healthy cell population.^{42–44} The surface modifying agents employed in the fabrication of various types of NPs for target-specific immune activities are listed in Table 1.

3. Types of NPs and their immunotherapeutic applications

NPs are broadly classified into polymeric, metallic, ceramic, carbon-based, lipid-based, and other types.⁵⁹ Another class of

NPs that has gained great attention in therapeutic application is biological NPs. The category includes viral components, exosomes, ferritin, lipoproteins, and magnetite, all of which are utilized for therapeutic delivery purposes.⁶⁰ Different types of such NPs, along with their immunotherapeutic applications, are discussed below.

3.1. Polymeric NPs

Polymeric NPs are crucial for therapeutic delivery due to their stability, biocompatibility, biodegradability, and water-soluble properties.⁶¹ As the by-products are relatively less toxic, polymers of both natural and synthetic origin are preferred for the fabrication of NPs.⁶² The polymeric NPs are mainly divided into two forms: nanocapsules and nanospheres. The nanocapsules possess a reservoir-like structure, and the nanospheres provide a solid matrix structure where the therapeutics are loaded by entrapment and adsorption mechanisms.^{63,64} These two large categories of polymeric NPs are further sub-categorized into different shapes like dendrimers, polymerosomes, and micelles.⁶¹ Dendrimers are hyperbranched synthetic polymeric 3D nanostructures bearing multiple functional groups that help in the binding of various therapeutics.^{65,66} Polymerosomes are stable polymeric amphiphilic vesicles capable of encapsulating both hydrophilic and hydrophobic therapeutic agents while maintaining their stability during the delivery process.⁶⁷ Micelles are generally spherical self-assembled NPs composed of amphiphilic block co-polymers characterized by hydrophobic core (accommodate hydrophobic drugs) and hydrophilic shell structures in an aqueous medium.⁶⁸ A variety of hydrophobic chemotherapeutic agents can be accommodated within polymeric micelles for their active and passive delivery.⁶⁹ Naturally derived polymers, such as proteins and polysaccharides, are extensively utilized in nanomedicine development. Polysaccharide-based NPs are favored for their biocompatibility and biodegradability, serving in various therapeutic roles, including cancer immunotherapy, inflammatory bowel disease (IBD) therapy, atherosclerosis treatment, vaccine delivery, macrophage polarization, and arthritis management.^{70–74} Similarly, various protein-based NPs are also used in immune modulation and therapeutic delivery in the treatment of various diseases such as acute pancreatitis, gastritis, cancer, arthritis, *etc.*^{73,75–77} Besides natural polymeric NPs, synthetic polymer-based NPs are pivotal in immunotherapy, offering promise due to their distinct characteristics like weak functional groups, chemical inertness, and tunable mechanical properties.^{78–82} The detail of polymer-based NPs are listed in following Table 2.

3.2. Metallic NPs

The metallic NPs have emerged as potential carriers of therapeutics due to their unique optical, magnetic, catalytic, and photocatalytic properties.¹⁰² The metallic NPs can be categorized into pure metallic NPs, metal oxide NPs, doped metal/metal oxide/metal NPs, metal sulfides, and metal-organic frameworks (MOFs).¹⁰³ Among pure metal NPs, gold NPs (AuNPs) with different shapes, like nanorods, nanostars, and nanoclusters, are most widely used for immunotherapeutic

Table 1 List of surface modifying agents used to fabricate NPs for target-specific immune activities

| Nanoparticle involved | Surface-modifying agent | Method of modification | Target receptor | Purpose of modification | Ref. |
|---|--|--|--|--|-----------|
| LNPs | Murine anti-DEC205 single chain antibody (scFv) | Thiol-maleimide chemical attachment | DEC205 receptors on DCs | DC-targeted siRNA delivery for immunosuppression | 45 |
| Carboxylated polystyrene NPs | PEG, CD47 antibody | EDC-NHS coupling reaction | SIRP α on macrophages | Macrophage targeting in the prevention of phagocytosis | 37 |
| Silicon NPs | Anti-CD11c/anti-DC-SIGN antibodies | Direct conjugation of periodate oxidized mAb | CD11c/DC-SIGN | DC-specific drug delivery | 39 |
| SPIONs (superparamagnetic iron oxide NPs) | IL-2 (interleukin-2) cytokine | Biotin-streptavidin non-covalent interaction | IL-2R on T-cells | T-cell targeted immunotherapy | 41 |
| Chitosan NPs | Mannose | Electrostatic interaction | Mannose receptors on immature DCs | DC-targeting in tumor immunotherapy | 46 |
| Gold NPs | Shikimoyl-ligand | Covalent attachment <i>via</i> 6-amino hexane thiol spacer | Mannose receptors on DCs | DC-targeted genetic immunization | 47 and 48 |
| Gold NPs | HIV Gag p17 and CMV pp65 peptides | EDC-NHS coupling | DC-SIGN receptors | DC-targeting in anti-HIV therapy | 49 |
| Polystyrene NPs | CD200 glycoprotein | Attachment <i>via</i> streptavidin rDNA technique | CD200R on macrophages | CD200R targeting, preventing the phagocytosis and inflammatory cytokines | 50 |
| PLGA NPs | M2pep peptide | Tannic acid-Fe ³⁺ complexation | CD206R on macrophages | M2-phenotype macrophage targeting in tumor immunotherapy | 51 |
| PLGA- <i>b</i> -PEG NPs | Herceptin® antibody/trastuzumab | NHS-PEG-alkyne linker mediated NHS-esterification | HER2 ⁺ receptors on cancer cells | Targeting the HER2 ⁺ breast cancer cells | 52 |
| Iron oxide NPs | Neu antibody/trastuzumab | Thiol-maleimide reaction | HER2 ⁺ receptors on cancer cell | HER2 ⁺ anti-cancer immunotherapy | 6 |
| PLGA NPs | Anti-CD8a F(ab') ₂ obtained from IgG antibody | Thiol-maleimide reaction | CD8a receptors on T-cells | CD8 ⁺ T-cell targeting in cancer immunotherapy | 53 |
| Lipid-dendrimer-calcium phosphate NPs | SP94 peptide | Thiol-maleimide reaction | PD1 (programmed cell death protein 1)-receptors on T-cells | Targeted delivery of drugs in hepatocellular carcinoma therapy | 54 |
| Liposome | IL-2-Fc, anti-CD137 Fab2 protein | Thiol-maleimide reaction | IL-2, CD137 receptors on CD8 ⁺ T-cells | T-cell, NK-cell activation in tumor immunotherapy | 55 |
| LNPs | Anti-human CD45RO primary antibody | Biotin-streptavidin non-covalent interaction | CD45RO on memory T-cells (Tm) cells | CD8 ⁺ Tm-cell targeting in lupus nephritis therapy | 56 |
| ZIF-8 metal-organic frameworks (MOFs) | Anti-CD16/32 antibody | Electrostatic adsorption | CD16/32 on M1-macrophages | M1-macrophage targeted drug delivery in osteoarthritis therapy | 57 |
| PLGA- <i>b</i> -PEG NPs | Anti-CD19, anti-CD220 mAbs | EDC-NHS coupling | CD19, B220 (CD45R) on B-cells | B-cell targeted drug delivery | 58 |

applications. Previous studies also demonstrated that bare metal NPs without any therapeutic agents can stimulate the immune system for the induction of effective immune responses against viral infections.¹⁰⁴ In the treatment of viral disease, the metallic NPs were observed to deliver specific nucleic acid cargos in hosts where subsequent expression of nucleic acid elicits potent anti-viral immune responses.¹⁰⁵ Cytokine-induced killer (CIK) cells are the emerging choice of therapeutic cells in the development of cell-drug therapy against cancer. The metallic NPs have shown the ability to form CIK cell-mediated cell-drug nanotherapeutics for exhibiting anti-tumor immune responses.^{106,107} Silver NP (AgNP) is a metallic category of NPs known for its pronounced antimicrobial properties, and researchers are exploiting AgNPs for immunotherapeutic purposes. *In vivo* animal studies provided several insights into the immunotherapeutic role played by

AgNPs, and polyphenol-modified AgNPs exert immune protection against the highly infectious herpes simplex virus-2.¹⁰⁸ The cell death of distant and deep-tissue tumors is quite challenging to target, and therefore researchers have successfully designed an immunogenic cell death therapy using palladium NPs for efficient deep-tissue tumor cell targeting. The Pd-based nanotherapeutic system triggers the release of “danger” signaling molecules that recruit T cells of immune systems in tumor tissues and efficiently arrest tumor growth.¹⁰⁹

3.3. Ceramic NPs

The ceramic NPs are considered promising therapeutic delivery vehicles due to their high heat resistance, mechanical integrity, stability, and chemical inertness. They are involved in the delivery of drugs, genes, proteins, peptides etc.^{110,111} Ceramic NPs are composed of oxides, carbides, carbonates,

Table 2 List of natural and synthetic polymers used for fabrication of NPs for immunotherapeutic applications

| Polymer source | Properties | Immunotherapeutic applications of fabricated NPs | Ref. |
|-----------------------------------|---|---|-----------------|
| Natural polymers | | | |
| Chitosan | U.S. FDA-approved, cationic, highly basic, polysaccharide, biocompatible. | Anti-cancer immunotherapy. | 74 and 83 |
| Hyaluronic acid | Poly-anionic, non-sulphated polysaccharide, non-toxic, biocompatible, biodegradable. | Inflammatory bowel disease (IBD) therapy, anti-inflammatory activities in atherosclerosis. | 70 and 84 |
| Gelatin | Protein obtained from collagen, biodegradable, biocompatible, non-antigenic. | Antigen delivery for immune stimulation, delivery of immunostimulant CpG oligonucleotides. | 85 and 86 |
| Silk fibroin | Protein obtained from skin cocoons or larvae, excellent biocompatibility, biodegradability, and low immunogenicity. | Immune system modulating drug delivery for cancer therapy, macrophage modulation, immunosuppressive therapeutic delivery in acute pancreatitis. | 87–89 |
| Albumin | Plasma protein, high biocompatibility, biodegradability, non-immunogenicity. | Immunomodulation in glioblastoma therapy, immune system activation and sonodynamic anti-tumor therapy, macrophage modulation in treatment of gastritis, checkpoint blockade-based metastatic pancreatic cancer immunotherapy, neutrophil-targeted drug delivery in rheumatoid arthritis (RA). | 75–77,90 and 91 |
| Cyclodextrin | Amphiphilic cyclic oligosaccharide, excellent biocompatibility. | Anti-inflammatory activity in treatment of Atherosclerosis, immune checkpoint blockade-based cancer immunotherapy. | 71 and 92 |
| Alginate | Anionic polysaccharide, good biodegradability, biocompatibility, non-toxic. | Antigen delivery for eliciting immune response against influenza, therapeutic delivery targeting macrophage polarization in treatment of RA, dendritic cell-targeted antigen delivery in cancer immunotherapy. | 4,72 and 73 |
| Synthetic polymers | | | |
| PLA-poly(lactic acid)/polylactide | Aliphatic polyester molecule, biocompatible, low toxicity, controlled hydrolytic degradation. | Macrophage cell-mediated anticancer drug delivery, hepatitis B vaccine delivery for cell-mediated immunity, improving immunogenicity of polysaccharide antigens and vaccine delivery, DC-targeted mRNA vaccine delivery. | 78 and 93–95 |
| PLGA-poly(lactide-co-glycolide) | Copolymer of PLA and PGA, FDA-approved, biocompatible, tunable mechanical property, a wide range of erosion times. | Macrophage-stimulated immune modulation and drug delivery, immune induction and therapeutic delivery. | 79 and 96–98 |
| Poly(caprolactone) | Polyester molecule, FDA-approved, biocompatible, non-toxic. | pH-responsive antigen delivery for humoral immune induction, vaccine delivery, immunity induction, immune modulation and anti-inflammatory drug delivery. | 80,99 and 100 |
| Polyanhydrides | Excellent biocompatibility, sustained drug delivery. | Protective and sustained immunisation, oral antigen delivery. | 82 and 101 |
| Polyorthoesters | Biocompatible, non-toxic, sustained release of drugs. | Immune cell-targeted antigen delivery. | 81 |

phosphates of different metals, and metalloids.¹¹⁰ Iron oxide NPs (IONPs) are one of the most popular and promising NPs that are employed in numerous immunotherapeutic applications. Starting from immunosuppressive drugs to various antigens, a wide range of therapeutics can be delivered with the help of IONPs.^{112,113} Due to their elemental compositions, the IONPs are involved in neutrophil modulation in the therapy of iron-deficiency disorders.¹¹⁴ The superior thermal conductivity of IONPs makes them a perfect candidate for photoimmunotherapy. In combination with other therapeutic agents, IONPs modulated anti-inflammatory macrophage phenotypes into tumor-suppressive proinflammatory macrophages, which facilitated immunotherapeutic restriction of tumor progression.¹¹⁵ Similarly, a variety of other ceramic NPs, such as silica NPs, hydroxyapatite NPs, titanium oxide, zinc oxide, and copper oxide NPs are also harnessed in multiple immunotherapeutic applications.

3.4. Carbon NPs

Carbon-based NPs are widely used nanostructured vehicles that have great potential for the delivery of therapeutics. The carbon family of NPs exhibits several characteristic features,

such as high mechanical integrity, excellent thermal conductivity, and great optical and magnetic properties. Therapeutic agents such as anti-cancer drugs, therapeutic peptides, genetic materials, antioxidants, protective agents, *etc.*, are either encapsulated or conjugated for their effective delivery.¹¹⁶ Among multiple subtypes, carbon nanotubes (CNTs), nanodiamonds (ND), carbon dots (CD), and graphene are the most used carbon NPs. The CNTs are sp² hybridized NPs, which are further divided into two types: single-walled CNT (SWCNT) and multi-walled CNT (MWCNT). The use of CNTs in near-infrared light-triggered conductive nanomaterial-assisted photothermal therapy (PTT) for cancer cell ablation has gained attention. Conjugating checkpoint blockers to CNTs enhances their anti-tumor immunotherapeutic efficacy, making CNT-led PTT more efficient.¹¹⁷ CNTs have also been used to deliver various immune-stimulating agents to provoke the immune components and to fight against cancer pathologies.^{118–120} Additionally, in the context of cancer vaccines, CNTs have been used to develop nanocomposite vaccines, such as by combining NY-ESO-1 antigen and CpG-ODN (TLR9 agonist) adjuvant molecule. NY-ESO-1, a cancer-testis antigen found in various cancers such as lung cancer, melanoma, and prostate

cancer, elicits effective T-cell responses due to its potential immunogenicity, making it a promising vaccine candidate.¹²¹ Early phase clinical trials have shown that immunotherapy using NY-ESO-1 could lead to the mitigation of cancers. The addition of a vaccine adjuvant molecule to NY-ESO-1 ameliorated the immunological responses against cancer. The nano-vaccine exhibited rapid internalisation by DCs, and elicited strong antitumor immunological responses by promoting humoral and cellular (CD4⁺ and CD8⁺ T-cell responses).¹²¹ In the manipulation of immune components to treat cancer, other carbon-based nanomaterials such as carbon dots and graphene are also employed to promote T-cell infiltration, modulation of DCs, and polarization of macrophages that suppress the disease pathologies.^{122–128} The immunotherapeutic applications of different types of NPs are listed in Table 3.

3.5. Lipid-based NPs (LNPs)

LNPs belong to the most important class of NPs that are receiving growing clinical approvals due to their suitable physico-chemical properties, payload flexibility, greater bioavailability, superior biocompatibility, biodegradability, and facile mode of fabrication. The LNPs can be either natural or synthetic in origin. The polar hydrophilic head and non-polar hydrophobic tails of the lipid molecules are assembled to give rise to a variety of LNPs. A wide range of lipid materials, such as triglycerides, mixtures of triglycerides, waxes, hard fats, and other lipids, are harnessed in the fabrication of LNPs. Additional components like emulsifiers and/or co-emulsifiers are also used for the fabrication of LNPs. Based on the lipid assembly features, the LNPs are further classified into liposomes, cationic LNPs, solid-lipid NPs (SLNs), nanostructured lipid carriers (NLCs), non-lamellar LNPs, ethosomes, cubosomes, *etc.* Within the aqueous interior, hydrophilic therapeutic agents can be loaded while maintaining drug stability, whereas hydrophobic drugs can be entrapped in the non-polar hydrophobic chains of the lipid components. LNPs can be functionally modified with a variety of agents, such as monoclonal antibodies, peptides, small molecule ligands, *etc.*, for the targeted delivery of therapeutic cargo.¹⁵⁹ Liposomes were developed in the earliest phases and were considered the simplest drug delivery vehicle to target and modulate the components of immune systems. For anti-cancer immunotherapy, the liposomes were loaded with various therapeutic agents that aimed at modulating the activities of macrophages, DCs, T cells, and NK cells residing in the tumor microenvironment (TME).^{160–163} In addition to anti-inflammatory drugs, therapeutic RNAs have also been delivered using liposomes for the treatment of diseases such as atherosclerosis, bacterial infections, and RA.^{164–166} Immune cell-specific small interfering RNA (siRNA) delivery and downregulation of gene expression of pathogenic stimulatory molecules are attractive ways to regulate the hyperactive immune systems. Hence, protective and targeted delivery of siRNA without further provoking the immune cells is quite challenging and demanding. In this regard, cationic LNPs were heralded as safe and potent nano-carriers of immune system-targeted therapeutic siRNA delivery

that results in a reduction of hyperactive, dysregulated immune responses.⁴⁵ Preventing the premature release of cargo siRNA in systemic circulation promoted greater accumulation of the therapeutic in the TME. Hence, a higher level of therapeutic siRNA at the TME raised the pH of the acidic TME by silencing the specific gene, which consequently decreased the number of immunosuppressive cells, promoted T-cell infiltration, and restored immune activities to retard the growth of tumor tissues.¹⁶⁷ Other types of LNPs, such as SLNs, NLCs, ethosomes, and cubosomes, are widely used as drug-delivery vehicles to modulate the immune components in several diseases. The immunotherapeutic applications of different types of LNPs are listed in Table 4.

3.6. Biologic NPs (BNPs)

The BNPs are naturally obtained NPs that are formed in various biological systems. These BNPs are composed of different organic and inorganic materials and can be intracellular as well as extracellular in origin. Among the different BNPs, exosomes, viruses and virus-like particles (VLPs), ferritin, magnetite, *etc.*, are widely used in the delivery of therapeutics (Table 5). The structural uniformity, immune system encompassing capability, and lower level of toxicity make them a suitable candidate for drug delivery applications.⁶⁰ Exosome nanovesicles are secreted by most cells (endothelial cells, adipocytes, B cells, DCs, neurons, mast cells, tumor cells, *etc.*) and are also found in several bodily fluids such as saliva, plasma, breast milk, amniotic fluid, cerebrospinal fluid, *etc.*^{179,180} Individual exosomes can elicit both positive and negative immune responses. Particularly in multiple cases of cancer immunotherapies, the exosome-based nanomedicines have greatly evolved. The chimeric antigen receptor-T cell (CAR-T) cell-derived CAR-exosomes are known to express cytotoxic signal molecules that can be useful in triggering anti-cancer immune responses. Unlike CAR-T cells used in CAR-T therapy, the CAR-exosome nanovesicle does not possess programmed cell death protein (PD1) and is relatively safe to use. Thus, the use of CAR-exosome reduces the risk of slackening of anti-tumor efficacy during recombinant programmed death ligand 1 (PD-L1) therapy of cancer.¹⁸¹ DC-targeted exosome-based vaccines represent a novel strategy in immunogenic cell death therapy. The surface-modified exosomes loaded with antigen and adjuvant exhibit promising results in DC activation and subsequent modulation of the tumor-reactive CD8⁺ T cells.¹⁸² Exosome nanovesicles of different cellular origins display bystander activities on macrophage polarization. Repolarizing the tumor-associated macrophages by macrophage-derived exosomes has become a new tool in antitumor immunotherapy. M2 macrophages assist in tumor progression by releasing anti-inflammatory cytokines and angiogenic factors. Thus, macrophage transition from the M2 to M1 phenotype is essential to release proinflammatory cytokines, thereby eliciting anti-tumor immune responses. In this regard, M1 proinflammatory macrophage-derived exosome NPs were successfully used as a therapeutic agent to repolarize the M2 macrophage into the M1 phenotype that effectively restricts

Table 3 List of different types of nanoparticles synthesized for immunotherapeutic applications

| Types of NPs | Therapeutic payload | Application | Ref. |
|---------------------------|--|---|-------------|
| Metallic NPs | | | |
| Gold nanorod (AuNR) | ssRNA | Antiviral therapy against pandemic influenza. | 104 |
| | None | Immune stimulation and inhibition of Respiratory Syncytial Virus (RSV). | 105 |
| Gold nanostar (AuNS) | Immunoadjuvant imiquimod (R837) | Immunotherapy in melanoma | 129 |
| | Cytokine-induced killer (CIK) cells | Anticancer immunotherapy | 106 |
| Gold nanocluster (AuNC) | CIK cells | Anticancer immunotherapy | 107 |
| | Antigenic peptide, cytosine-phosphate-guanine (CpG) oligodeoxynucleotides (ODNs) | Vaccine-mediated immune stimulation | 130 |
| Silver NP (AgNP) | Technetium-99 m, lutecium-177 | Cancer radio-immunotherapy | 131 |
| | None | Enhances immune protection against Herpes simplex virus-2 (HSV-2) infection. | 108 |
| Palladium NP (PdNP) | None | Immune stimulation in cancer immunotherapy. | 132 |
| | None | Immune modulation for cancer therapy | 133 |
| Ceramic NPs | Doxorubicin (DOX) | Anticancer chemoimmunotherapy | 109 |
| | Iron oxide NP (IONP) | | |
| Iron oxide NP (IONP) | Ovalbumin (OVA) | Anti-tumor vaccine immunotherapy | 112 |
| | Mycophenolic acid | Immunosuppressive drug delivery. | 113 |
| | None | Neutrophil modulation in iron deficient anaemia. | 114 |
| | Sulfasalazine | Anti-cancer immunotherapy | 115 |
| | DOX, polyinosinic: polycytidylic acid (poly (I:C)) | Anti-cancer immunotherapy | 134 |
| | Ferumoxytol | Immune activation in cancer immunotherapy | 135 |
| | Poly (I:C) | Vaccine delivery to lymph nodes. | 136 |
| Silica-based NP (SiNP) | Nucleic acid and DOX | Co-delivery of therapeutics for immune stimulation and cancer cell targeting. | 137 |
| | Peptide neoantigen, CpG oligodeoxynucleotide | Anticancer immunotherapy. | 138 |
| | anti-PD1 antibody | Anticancer immunotherapy. | 139 |
| | Gardiquimod | Anticancer photoimmunotherapy. | 140 |
| | Cyclic diguanylate monophosphate(cdGMP) | Dysregulated APCs specific drug delivery in glioblastoma immunotherapy. | 141 |
| Hydroxyapatite NP (HApNP) | Peptide antigen B ₂ T | Vaccine delivery | 142 |
| | Methylprednisolone acetate | Immunosuppressive and anti-inflammatory drug delivery in RA. | 143 |
| | Lactoferrin | Immunomodulation for the treatment of <i>Helicobacter pylori</i> infection. | 144 |
| | OVA | Immune stimulation, anti-cancer immunity | 145 |
| Titanium oxide (TiO) NP | Bacterial lipopolysaccharide | Macrophage modulation and immune stimulation | 146 |
| | Chito-oligosaccharides | Anticancer immunotherapy | 147 |
| | None | Anticancer immunotherapy | 148 |
| Zinc oxide (ZnO) NP | None | DC and helper T-cell modulation | 149 |
| | DOX | DOX-induced macrophage polarization in cancer immunotherapy | 150 |
| Copper oxide (CuO) NP | None | T-cell differentiation, immune modulation | 151 |
| | Chitosan | Macrophage activation in cancer immunotherapy | 152 |
| Carbon-based NPs | None | Immunomodulation in the treatment of inflammatory ulcerative colitis. | 153 |
| | CNTs (SWCNT and MWCNT) | | |
| CNTs (SWCNT and MWCNT) | Anti-cytotoxic T-lymphocyte-associated protein 4 | Immune stimulation in anti-tumor therapy | 117 |
| | CpG, OVA and anti-CD40 Ig (α CD40) | Adjuvant delivery for immune induction for cancer therapy | 118 |
| Nanodiamonds (NDs) | None | Immune cell recruitment for anticancer therapy | 119 |
| | CpG | Macrophage activation and immune stimulation against glioblastoma. | 120 |
| | Indolicidin | Immune activation and modulation in the treatment against antibiotic resistance. | 154 |
| | None | Immunomodulation in bone remodelling | 155 |
| Carbon dots (CDs) | None | Immune cell induction for anti-tumor immunotherapy | 156 |
| | Octadecylamine, dexamethasone | Macrophage-specific immunomodulation in the treatment of RA. | 157 and 158 |
| Graphene-nanoplatfrom | Fe ion, DOX, and Losartan | T-cell infiltration, chemoimmunotherapy against cancer | 122 |
| | None | DC-targeted danger signal-specific anti-cancer immunotherapy | 123 |
| Graphene-nanoplatfrom | None | Induction of CD8 ⁺ T cells, mature macrophages, and natural killer cells for anti-cancer immunotherapy | 124 |
| | None | Immunotherapy against melanoma | 125 |
| Graphene-nanoplatfrom | None | Anticancer immunotherapy | 126–128 |

Table 4 List of LNPs used for immunotherapy of various diseases

| Types of LNPs | Lipid composition | Therapeutic payload | Applications | Ref. |
|---------------|---|---|--|-------------|
| Liposomes | SPC, cholesterol, DSPE-PEG2000 | Honokiol and disulfiram-copper complex | Activation of macrophage, DCs, T cells, NK cells, immunogenic cell death, anti-tumor immunotherapy against glioblastoma. | 162 |
| | Phosphatidylcholine, DSPE-PEG carboxy, CSF ₁ R-inhibiting amphiphile | Anti-PDL ₁ and BLZ ₉₄₅ (CSF ₁ R-inhibitor) | Anti-cancer immunotherapy | 160 |
| | HSPC, cholesterol, DSPE-PEG2000 | Ursolic acid | Immunomodulation and anti-cancer immunotherapy | 161 |
| | Cholesterol, phosphatidylcholine, DPPE, rhodamine red-labelled DPPE | siRNA | NK cell-targeted anti-tumor therapy | 163 |
| | DSPE | Methotrexate | Anti-inflammatory drug delivery in the treatment of atherosclerosis | 164 |
| | DOTAP, DMPC, DSPE-PEG | siRNA | Macrophage modulation, immunogene therapy against staphylococcus infection | 166 |
| | (DC _{8,9} PC), DSPE-PEG2000 | Dexamethasone | Anti-inflammatory drug delivery in the treatment of RA | 165 |
| Cationic LNPs | DSPC, DLinDMA, cholesterol, DSPE-PEG-MAL | siRNA | DC-targeted siRNA delivery | 45 |
| | DOTAP, PEG5000-block-PLGA11000 | siRNA | T-cell modulation and anti-cancer immunotherapy | 167 |
| | AMPA-O16B, DOPE, DSPE-PEG2000, cholesterol | Shigella bacteria-derived effector OspF | Macrophage modulation in anti-cancer therapy | 168 |
| | Amino lipid (lipid D), cholesterol, DSPC, DMG-PEG2000 | Dengue envelope proteins (DEN-80E) | Vaccine delivery and immunostimulation against dengue | 169 |
| | DODMA, DSPC, DMG-PEG | mRNA | Immunomodulation | 170 |
| SLNs and NLCs | Stearic acid, lecithin, poloxamer-188 | Paclitaxel | Immunomodulation in melanoma | 171 |
| | Naringenin and linolenic acid | Cyclosporin | Immunosuppressive drug delivery in psoriasis | 172 |
| | Cetyl palmitate, polysorbate-60 (Tween-60), miglyol-812 | Resveratrol | DC-targeted anti-inflammatory drug delivery and immunomodulation | 173 |
| | DOTAP, Span, Tween-80, Squalene, dynasan114 | Zika virus antigen encoding viral RNA | Zika vaccine delivery | 174 |
| Ethosomes | Lecithin, cholesterol, octadecyl amine | Tyrosinase-related protein-2 (TRP-2), CpG, mRNA, siRNA delivery | DC stimulation in tumor immunotherapy | 175 and 176 |
| Cubosomes | Phytantriol, propylene glycol, Pluronic F127 | Immunostimulant polysaccharide (PS) obtained from <i>Ganoderma lucidum</i> | DC activation, T-cell modulation | 177 |
| | <i>Achyranthes bidentata</i> polysaccharide (ABS), Monooleate, Pluronic F127 | ABPs delivery | Immunomodulation | 178 |

Abbreviation: SPC, soybean phosphatidylcholine; DSPE-PEG2000, 1,2-distearoyl-*sn*-glycero-3-phosphoethanolamine-*N*-[methoxy(polyethylene glycol)-2000]; HSPC, hydrogenated soy phosphatidylcholine; DSPC, distearoylphosphatidylcholine; DPPE, 1,2-dipalmitoyl-*sn*-glycero-3-phosphoethanolamine; DMPC, dimyristoyl phosphatidylcholine; DOTAP, 1,2-dioleoyl-3-(trimethylammonium)propane; DC_{8,9}PC, 1,2-bis(10,12-tricosadiynoyl)-*sn*-glycero-3-phosphocholine; DOPE, dioleoylphosphatidylethanolamine; DLinDMA, 1,2-dilinoleoyloxy-3-dimethylaminopropane; DSPE-PEG-MAL, 1,2-distearoyl-*sn*-glycero-3-phosphoethanolamine-*N*-[maleimide(polyethylene glycol)-2000]; PEG200-DMG, polyethylene glycol-2000-dimyristoyl glycerol.

tumor progression.¹⁸³ On the other hand, it is interesting that exosomes from olfactory ensheathing cells have also been shown to suppress proinflammatory macrophages.¹⁸⁴ Thus, the specific-cell-derived exosome repolarizes the M1 macrophage into an anti-inflammatory M2-phenotype and plays excellent immunomodulatory roles in the therapy of neuroinflammation. In the prevention of autoimmune uveitis, specific circulatory exosomes were used as direct anti-inflammatory nanotherapeutics to reduce the disease-relevant inflammatory immune mediators.¹⁸⁵ Viruses and VLPs are nanostructured non-pathogenic BNPs that are resistant to temperature and pH and have emerged as highly efficient vaccine delivery tools. The non-pathogenic viruses and genetic

material-free viral capsid protein-based VLPs possess a striking ability to elicit immune responses. Particularly, these BNPs act as engineered nanovaccines in cancer immunotherapy, where they can solely or in association with therapeutic immunogens provoke anti-tumor immune responses by recruiting immune cells and elevating the level of cytokines.^{186–188} Ferritin BNPs are protein nanocages found in the biological systems of eukaryotes, bacteria, and archaea that have evolved mainly to store iron species. Recent studies claimed that ferritin BNPs could meet the need for novel antiviral vaccines. The ferritin NPs obtained from diverse viral and bacterial sources exhibited antiviral immune responses against the pandemic SARS-CoV-2 virus.¹⁸⁹ Moreover, the ferritin BNPs are also found to act as

Table 5 List of BNPs used for immunotherapeutic applications

| Types of BNPs | Source of origin | Therapeutic payload | Therapeutic application | Ref. |
|------------------|---|--------------------------------|--|------|
| Exosomes | Serum of hypopharyngeal cancer patients | | CD8 ⁺ T-cell suppression, hypopharyngeal cancer immunotherapy. | 192 |
| | Bone marrow-derived dendritic cells | Ag/anti-CD64 antibody complex | Suppression of polymorphonuclear neutrophils in the treatment of asthma. | 193 |
| | Cancer cells | Chlorin e6 | Immune stimulation in cancer immunotherapy. | 194 |
| | Dendritic cells | TGF- β 1 cytokine | Immunomodulation in the treatment of inflammatory lung diseases. | 195 |
| | Olfactory ensheathing cells | | Pro-inflammatory macrophage suppression, immunomodulation in treatment of spinal cord injury. | 184 |
| Viruses and VLPs | Alfalfa mosaic virus | | Immune stimulation <i>via</i> recruitment of immune cells and increasing cytokine level in cancer immunotherapy. | 187 |
| | Cowpea mosaic virus | | Anti-tumor immunotherapy. | 186 |
| | Hepatitis B virus | OVA-antigen, gp100-antigen | DC maturation, T-cell stimulation in anticancer immunotherapy. | 188 |
| Ferritin | SARS-CoV-2 Spike-protein(S), S1, and RBD domain | | Immunization against SARS-CoV-2 virus. | 189 |
| | Engineered human ferritin | CpG ODNs | Tumor-associated M2 macrophage-targeted therapeutic delivery for cancer immunotherapy. | 196 |
| | <i>Helicobacter pylori</i> -derived ferritin | Recombinant soluble E2 subunit | Immunization against hepatitis C virus. | 190 |
| | <i>Helicobacter pylori</i> -derived ferritin | HIV-1 envelope proteins | Immunization against HIV-1 virus. | 191 |

potential vaccine delivery agents against viral infections caused by hepatitis virus C, HIV-1, and other viruses.^{190,191}

4. Key immune components and their modulation with therapeutic nanoplatforms

As the immune system plays a complex role in the pathogenesis of several diseases like allergic rhinitis (AR), RA, cancer, multiple sclerosis, HIV, and other viral infections, significant attention has been paid to the discovery of novel immunotherapeutic nanomedicines to restore physiological normalcy. In various human disorders, T cells, B cells, DCs, macrophages, and complement components of the immune system play bystander roles where aberrant immune activities are directly associated with poor disease prognosis. Hence, various immunomodulators (activators and suppressors) can be delivered to re-establish optimal immune responses by selectively targeting the immune components using novel nanomedicines. Specifically, in the treatment of autoimmune diseases like RA, type-1 diabetes mellitus, and allergic inflammation, the auto-reactive immune cells can be suppressed, while therapeutic NPs have also been used to activate the immune cell responses during the treatment of cancer and other immunosuppressive diseases.

4.1 Nanotherapeutics for targeting T cells

T cells are one of the centrally important cells of the immune system and play critical roles in regulating normal immune responses. This dynamic effector cell coordinates with other immune components for the detection and encounter of anti-

gens and other foreign invaders. Researchers have been manipulating T cells to eradicate several immune disorders. The engineered NPs are either conjugated or encapsulated with therapeutic agents that are delivered to T cells by targeting T-cell receptors (TCRs). Modulating T cells has therapeutic consequences, leading to the remission of immune disorders.

4.1.1. Nanotherapeutics in activation and recruitment of T cells. Multiple immune disorders, ranging from viral to cancer, are characterized by diminished activity of T cells. Hence, activation of T cells is necessary to deal with suppressed or compromised immunity and is a key strategy for the restoration of effective immune activities. For instance, in cancer, T cells are greatly affected due to the presence of different inhibitory signals in the TME. The reasons behind poor T-cell activities are inhibitory receptors present on abnormal T cells, inhibitory cells in the TME, immunosuppressive mediators, *etc.* Specific activation of T cells represents a targeted and precise modality in the clinical settings of antitumor immunotherapy. Activation of T cells directly, or supply of genetically modified engineered T cells from an external source pave the way for the restoration of T-cell functionalities.^{197,198} PD1 and its predominant form, PDL1, are the immune checkpoints associated with the phenomenon of tumor-immune tolerance. The induction of immunological tolerance leads to poor infiltration of cytotoxic T cells, causing the TME to be immunosuppressive in nature, which subsequently results in the failure of immune checkpoint blockade (ICB) therapy. Blocking the PD-1/PDL1 protein with an anti-PDL1 antibody and subsequent elicitation of antigen-induced recruitment of T cells in tumor tissues has become an effective approach for fighting against cancers.¹⁹⁹ The anti-PDL1 antibody assembled NPs was developed to achieve T-cell targeted therapeutic cargo delivery. The engineered antibody

nanoplatfoms, loaded with TME-responsive peptides and antigen-generating species, abrogate immunological tolerance and accelerate greater T-cell infiltration in the TME to deplete the cancer cells.¹⁹⁹ The surface of T cells expresses the costimulatory receptor OX40 and PD-1 receptor, which can be significant targets for activation of T cells.²⁰⁰ Administration of agonistic antibodies aOX40 and aPD-1 individually or in combination showed activation of T cells but at a sub-optimal level. Hence, dual antibody (aOX40 and aPD-1) delivery is necessary to synergistically activate a large population of T cells. With the help of thiol-maleimide chemistry, conjugating the aOX40 and aPD-1 antibodies on the surface of poly(lactide-co-glycolide)-*b*-poly(ethylene glycol) (PEG-PLGA) NPs provided the formation of stable antibody co-delivery nanoplatfoms that confer greater T-cell activation, improved therapeutic efficacy, and immunological memory.²⁰⁰ Apart from OX40 receptor modulation, the stimulation of Toll-Like receptors 7/8 (TLR7/8) receptors in T cells by external agonists is a key strategy to activate the T-cell population. PD-1 positive T cells are also characterized by immunosuppressive TGF β cell signalling, and their blockade prevents the establishment of an immune-deficient cold TME. Hence, the co-administration of a TGF β inhibitor (SD-208) and a TLR7/8 receptor agonist (R848 or resiquimod) *via* antibody-modified NPs enables precise targeting and activation of T cells.⁵³ The anti-PD-1 F(ab')₂ antibody fragment-modified PEG-PLGA NPs loaded with SD-208 and R848 significantly target the PD-1-positive T cells. The T-cell-specific targeted co-delivery of immunomodulatory agents ensured a higher accumulation of therapeutic cargos that expanded the activation of CD8⁺ T cells.⁵³ The direct application of therapeutic nanoscale artificial antigen-presenting cells (naAPCs) is a new approach that has been employed in the activation of T cells. Mechanistically, to activate the T cells, aAPCs act on several cellular and molecular components, such as MHC-I/T-cell receptor stimulation, CD80/CD28 costimulatory signaling, and cytokine release. The T-cell activation potential of nano aAPCs is relatively less than that of micro aAPCs, which can be a therapeutic barrier in this strategy. Hence, antigen-based pre-activation of CD8⁺ T cells and subsequently inducing high redox potential augments the T-cell activation efficacy of nano aAPCs in a size-convertible manner. The pre-activated CD8⁺ T-cell-mediated redox potential converted the nano aAPCs into highly efficient micro aAPCs at the tumor site and significantly activated the CD8⁺ T cells²⁰¹ (Fig. 2). The poor efficacy of triggering T-cell activation in the regression of tumor growth is observed in vaccine-monotherapy treatment modalities. Thus, vaccine efficiency can be accelerated by promoting the activation of T cells by employing adjuvant nanotherapeutics. In combination with vaccine antigen, the polymeric cationic NPs act as adjuvants to increase the immunogenicity of the administered vaccine, thereby recruiting a greater population of tumor-associated antigen-specific CD8⁺ T cells.²⁰² Since specific surface antigen receptors expressed by cancer cells are recognized by the TCRs that aid in directing T cells toward specific cancer cells, the insufficient expression of cancer cell-recognizing receptors

fails to effectively eliminate the cancer cells. Hence, CAR T-cell therapy has emerged as a tremendous strategy to target cancer cells by modulating the activity of T cells. The gene encoding the chimeric antigen receptors (CARs) can be successfully transported to the T cells by NPs, where specific releases of cargo genes express CARs on T cells and enable them to reach cancer cells for effective elimination. Smith *et al.* employed biocompatible and biodegradable poly(β -amino ester) NPs in the delivery of CAR genes to T cells.²⁰³ To achieve T-cell-specific therapeutic cargo gene delivery, the NPs were surface-modified with anti-CD3eF(ab')₂ antibody fragments by means of electrostatic interaction. Furthermore, the modification with microtubule-associated sequence and nuclear localization signals conferred microtubule-mediated uptake and nuclear transport of genetic cargo to T cells, respectively.²⁰³ Similarly, in an alternative approach, NP-based bispecific T-cell engagers, or nanoBiTEs, were developed as nanotherapeutic platfoms that interplay between T cells and cancer cells for better T-cell recruitment.²⁰⁴ The liposomal NPs were decorated with anti-CD3 antibodies for T-cell-specific binding and conjugated with monoclonal antibodies (mAbs) to bind to tumor-associated antigens (TAAs). Thus, the nanoBiTEs with dual functionality served as a linker or engager to redirect the T cells toward the antigen-displaying cancer cells. Despite being great tools for targeted activation and recruitment of T cells, nanoBiTEs and CAR-T therapeutic strategies are facing several limitations. The cancer cells might express multiple TAAs, where only targeting the monotypic antigen is a major limitation associated with both CAR-T and nanoBiTEs therapies. To overcome the limitations associated with CAR-T and nanoBiTEs therapies, the researchers have developed NP-based multi-specific T-cell engagers (nanoMuTEs).²⁰⁴ First, nanoliposomes were decorated with a variety of mAbs to bind with multi-antigenic receptors present on cancer cells, and then further modification of the liposomes with mAbs (anti-CD3) governed T-cell binding. Thus, target-specific multiple antibody-conjugated nanoMuTEs not only targeted multi-antigenic cancer cells, but also inhibited the progression of antigen-less tumors. Hence, the novel nanotherapeutic system showed higher activation of CD4⁺/CD8⁺ T cells and could be a potential solution in clinical settings of cancer immunotherapy.²⁰⁴

Modulation of immune components has become an urgent survival option for an individual suffering from the COVID-19 viral infection. Activation of different T-cell subsets and eliciting strong immune responses against COVID-19 by specific antigens could be an immune-protective, life-saving strategy. Engineered mRNA-loaded LNPs were chosen as novel nanotherapeutic vaccines to provoke strong immune responses against SARS-CoV-2 infection.²⁰⁵ The safe and well-tolerated LNPs precisely delivered genetic information encoded within the mRNA cargo. The released mRNA expressed receptor-binding domains of the COVID-19 viral spike protein, which subsequently activated CD4⁺, CD8⁺, and favourable helper type-1 T-cell (Th1) subsets. Hence, the activation of CD4⁺ and CD8⁺ T cells facilitated the establishment of prolonged immunological memory, which helped in the prevention of SARS-CoV-2 viral disease.²⁰⁵

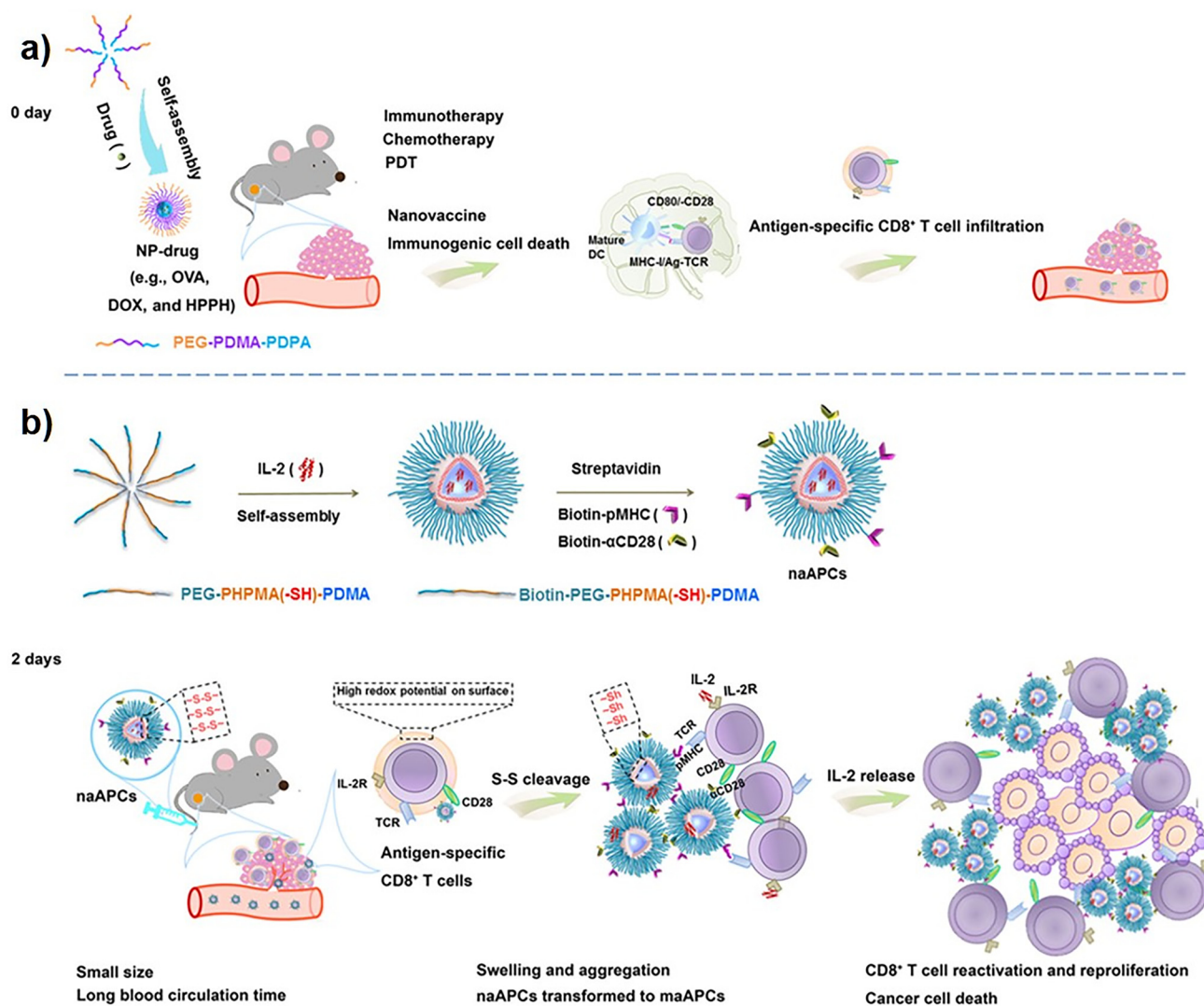


Fig. 2 Schematic illustration demonstrating nanosized artificial antigen-presenting cells (naAPCs) for immunotherapy. (a) Nanoparticles (NPs) self-assembled from copolymer PEG-PDMA-PDPA could elicit host immunity in EG7-OVA tumor-bearing mice. NP encapsulated with OVA DOX, or HPPH (NP-drug) acts as nano-vaccines. This NP-drug induces immunogenic cell death (ICD), promotes maturation of dendritic cells (DCs), antigen processing, and T-cell presentation. This ultimately leads to activation of antigen-specific CD8⁺ T cells and infiltration into tumor tissue. (b) Scheme representing formation of IL-2-loaded size-transformable naAPCs through self-assembly of copolymer biotin-PEG-PHPMA(-SH)-PDMA. The surface of naAPCs is decorated with peptide-loaded MHC (pMHC) monomer and αCD28. High redox potential on preactivated antigen-specific T-cell surface results in cleavage of disulfide bonds of naAPCs into thiols. As a consequence, conversion of naAPC from nanosize to microsize leads to the formation of an aggregate in tumor tissue due to its large size, while secreting IL-2 to enhance immune response. Reproduced with permission from ref. 201, Copyright 2020, American Association for the Advancement of Science. Abbreviations used: PEG-PDMA-PDPA polyethylene glycol-block-poly(2-dimethylaminoethyl methacrylate)-block-poly(2-diisopropylaminoethyl methacrylate).

CD8⁺ T cells are superior immune cells that ensure protective immunity against viral diseases caused by the zika virus (ZIKV), dengue virus (DENV), and others. Although various antibody-inducing vaccines are used in immunotherapeutic paradigms, their suboptimal antibody response necessitates alternative and more potent immunotherapeutic platforms. Activating the CD8⁺ T cells with nanovaccines is one of the major strategies to fight against these deadly viruses. Recent study revealed that the NP-based delivery of antigen-expressing replicon RNA induced CD8⁺ T cells significantly and prevented the fatality caused by ZIKV infection.²⁰⁶ Activation and expansion

of regulatory T cells (Tregs) have shown great clinical significance in controlling autoimmune diseases. Since autoreactive T cells play pathogenic roles with deleterious outcomes against the body's own immune system, targeting the vicinity of autoreactive T cells could be an effective approach for halting disease progression. Ligating the TCRs present on cognate T cells with an external peptide-based major histocompatibility complex (pMHC) can result in the activation and expansion of immunoregulatory Tregs *via* TCR-pMHC interaction. Considering this molecular event, the administration of pMHC-decorated NPs showed promising results in the

differentiation of disease-driving autoreactive T cells into Tregs.²⁰⁷ Other than TCRs, aryl hydrocarbon receptors (AhRs) are highly expressed by several immune cells and could be a potential target to promote a greater Treg population. This could be achieved by antigen-specific immune tolerance induction and the promotion of AhR signaling. The co-delivery of AhR agonist ligand and myelin oligodendrocyte glycoprotein (MOG)_{35–55}-derived T-cell epitope led to myelin-specific T-cell modulation and facilitated anti-inflammatory tolerogenic gene expression. However, the biodegradation and clearance of therapeutic agents limit the treatment efficacy when co-administered freely. Hence, LNPs were successfully deployed in the protective delivery of therapeutic AhR-agonist and (MOG)_{35–55} T-cell epitopes at a time when the concurrent delivery of encapsulated therapeutic cargos exhibited a greater number of Forkhead box protein P3 (Foxp3⁺) Tregs and type-1 Tregs.²⁰⁸ Thus, the nanomedicine-assisted activation and expansion of Tregs could be a life-saving approach for the treatment of autoimmune disorders.²⁰⁸ Foxp3⁺ Tregs also play pivotal roles in maintaining atheroprotection; therefore, elevating the level of Tregs could be a therapeutic option in the regimen of cardiovascular immunotherapy.²⁰⁹ The modulation of vitamin D nuclear receptor (VDR) in tolerogenic DCs is a potential therapeutic target to promote greater proliferation of Tregs. In this context, the application of synthetic anti-inflammatory drugs to modulate VDR has been explored to induce disease-preventing Tregs. However, a lower therapeutic index is a major limiting factor associated with systemic administration of these agents, which ultimately fails to recruit an efficient level of Tregs. Hence, micelle NPs have been engaged in the delivery of VDR-modulating anti-inflammatory drugs to obtain a greater therapeutic index. The micelle-assisted sustained delivery of immunomodulatory drugs maintained high levels of Foxp3⁺ Tregs in atherosclerotic lesions as well as in lymphoid organs, and enhanced cardioprotectivity.²⁰⁹

4.1.2. Nanotherapeutics in T-cell suppression. The autoimmune diseases are characterized by hyperactivation of the body's own effector T cells that leads to loss of self-tolerance, and self-organ and tissue damage. Inhibiting the autoreactive CD4⁺ and CD8⁺ T-cell responses could be a beneficial therapeutic approach for inducing immunological tolerance. Several antigen-specific immunotherapies have been followed to modulate T-cell activities, but exacerbation of already existing inflammatory events is a potential risk associated with antigen-based immunotherapy. Thus, the development of tolerogenic NP-based therapeutics with the ability to co-deliver antigen and immunosuppressive agents can dampen the risk factors associated with antigen-based immunotherapies. The application of tolerogenic NPs has shown excellent ability to inhibit activation of CD4⁺ and CD8⁺ T cells as well as suppress anti-drug antibody responses, which can prevent autoimmune diseases.²¹⁰ Leveraging the fact that immune cells possess preferential uptake of NPs, the T-cell favoured the preferential uptake of PEG-modified antioxidant hydrophilic carbon nanoclusters (PEG-HCC) and showed therapeutic efficacy in the management of autoimmune encephalomyelitis.²¹¹ The

antigen-mediated pre-stimulation of T cells showed preferential internalization of antioxidant carbon NPs, which in turn promoted ROS-species scavenging and reversibly inhibited T-cell proliferation.²¹¹ The autoimmune CD8⁺ T cells are known to have β -cell destructive properties that lead to severe pathogenicity in type-1 diabetes mellitus. Thus, the inhibition of cytotoxic CD8⁺ T cells is necessary to halt the disease progression. The cocktail of HLA-A*02:01-restricted epitopes decorated on NPs exhibited clinical induction of antigen-specific immune tolerance.²¹² The epitope peptide carrying therapeutic NPs introduced T-cell tolerance mainly by promoting the activity of Tregs like CD4⁺CD25⁺, CD4⁺Foxp3⁺, and the release of anti-inflammatory IL-10 cytokines that collectively ablate cytotoxic T-cell proliferation.²¹² Similarly, autoimmune CD4⁺ T cells are also found to be involved in the pathogenicity of type-1 diabetes and could be potentially targeted for the induction of immune tolerance. In the induction of tolerogenic responses, the hybrid insulin antigen-loaded tolerogenic NP has gained attention due to its impaired ability to inhibit T-cell activities. The tolerogenic NPs act on Tregs to foster their proliferation. As a result, in comparison with cytotoxic IFN- γ effector T cells, a greater population of immunoregulatory Foxp3⁺ Tregs was induced by tolerogenic NPs.²¹³

Primary biliary cholangitis is an autoimmune disease where CD8⁺ T cells play pathogenic roles and essentially need to be suppressed to prevent disease severity. The effective delivery and subsequent cross-presentation of peptide auto-antigens to MHC-I complexes that are expressed on cholangiocyte cells could be an effective approach for aborting the infiltration of pathogenic T cells in the liver. The ovalbumin peptide SIINFEKL antigen-modified NPs were successfully used to deliver the tolerogenic peptide antigen to liver cells that were consequently cross-presented by the MHC-I complex.²¹⁴ As a result, the antigen-peptide nano-assembly downregulated the infiltration of autoreactive T cells into the liver, thereby protecting the liver from being damaged by cytotoxic T cells²¹⁴ (Fig. 3A & B). A greater population of T cells has also been implicated in hypersensitive allergic contact dermatitis. Previous studies revealed that the topical application of negatively charged SiNPs can alleviate inflammatory allergic dermatitis by reducing the high level of CD3⁺ and CD8⁺ cytotoxic T cells. Hence, the low-dose topical application of negatively charged SiNPs acts as an immunomodulator and decreases cytotoxic T-cell infiltration and inflammatory cytokine production.²¹⁵ The helper T-17 (Th-17) cells are another subset of T cells that play pathogenic roles in the development of chronic inflammatory psoriasis. The enhanced differentiation of native CD4⁺ T cells into pro-inflammatory cytokine (IL-17) producing helper T-17 cells drives strong inflammatory responses. Particularly, intracellular ROS-species-mediated stress conditions in skin keratinocytes promote high differentiation of native CD4⁺ T cells into helper T-17. Hence, halting the differentiation and proliferation of IL-17-producing Th-17 cells from their native T cells could be an effective strategy in the management of psoriasis. Endogenous bilirubin-based NPs have shown the potential to scavenge intracellular ROS

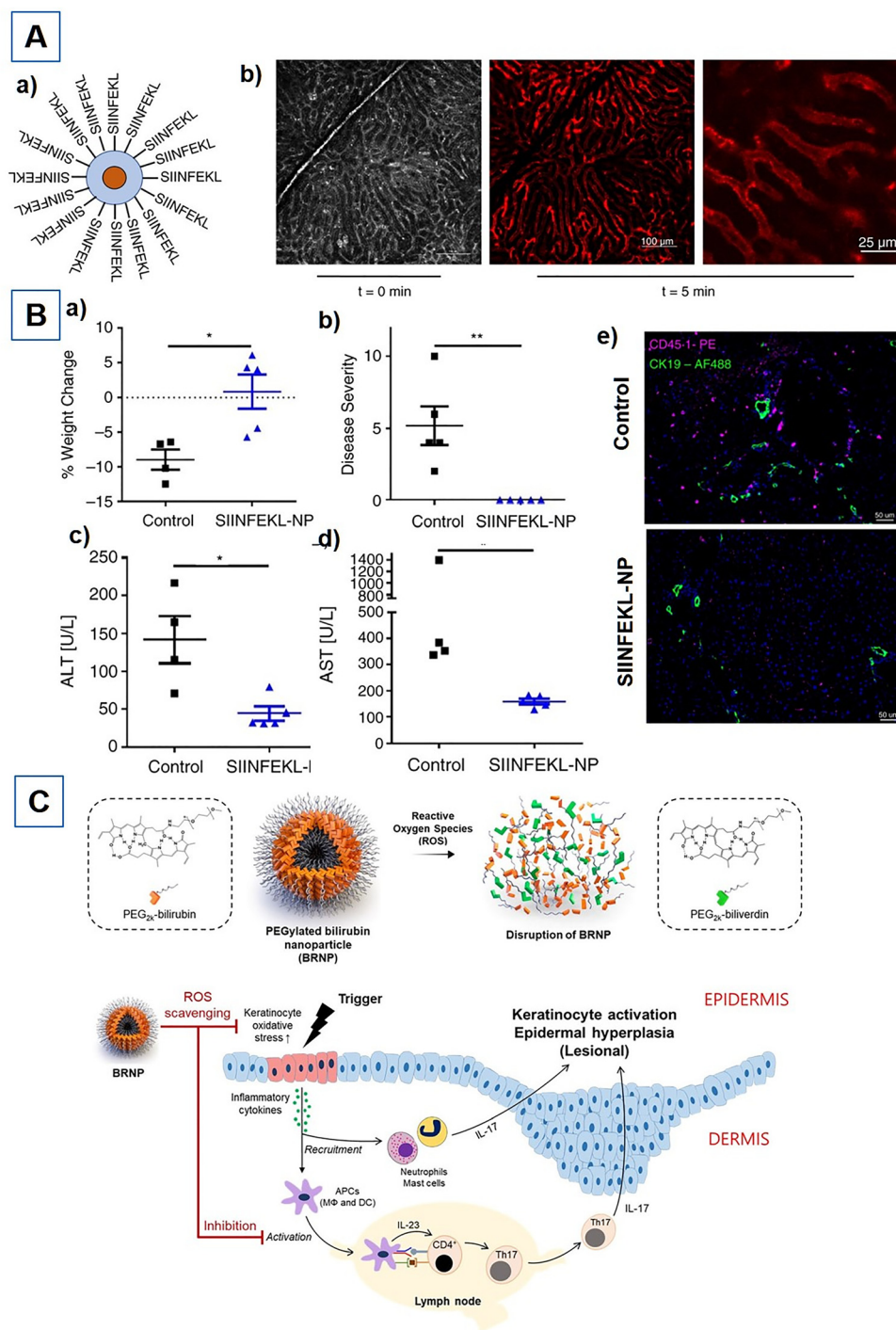


Fig. 3 (A) Pictorial representation of SIINFEKL-decorated nanoparticle (NP) and its cellular uptake: (a) Illustration of SIINFEKL peptides covalently conjugated to NP, having a monodisperse iron oxide or quantum dot core of about 7 nm diameter; (b) NP uptake in liver sinusoids (red fluorescent quantum dot core) at $t = 0$ min and 5 min post-tail vein injection. Images captured and assessed by intravital microscopy. (B) SIINFEKL peptide-loaded NP prevents CD8⁺-mediated autoimmune cholangitis in K14-OVAp mice; (a) percentage change in body weight at day 5 compared with weight at the time of OT-1 T-cell transfer; (b) Severity of autoimmune cholangitis at day 5; (c and d) Serum levels of liver enzymes: alanine aminotransferase (ALT) and aspartate aminotransferase (AST); (e) Immunofluorescence (IF) staining of CD45.1+ liver-infiltrating OT-1 cells (pink fluorescence) and CK19+ cholangiocytes (green fluorescence) shows reduced infiltration of autoreactive T cells. Reproduced with permission from ref. 214, Copyright 2021, John Wiley & Sons. (C) Schematic representation of PEGylated bilirubin nanoparticles (BRNPs) and their proposed mechanism of action in psoriasis. Oxidative stress, due to redox imbalance, leads to production of ROS and inflammatory mediators and autoantigens mediated through epidermal keratinocytes. This increases the recruitment and maturation of APCs, which further activates differentiation of naïve CD4⁺ T cells to Th1 and Th17 cells in skin lesions and lymphoid organs. Subsequent release of interferon- γ (IFN- γ) or IL-17 results in aberrant proliferation of keratinocytes. The use of BRNPs helps in mitigating the ROS production and activation of APCs through its antioxidant and anti-inflammatory action. Reproduced with permission from ref. 216, Copyright 2020, Elsevier.

species, therefore reducing intracellular stress levels. Thus, topical administration of PEG-modified bilirubin NPs (BRNPs) acted as an immunomodulator that dampened the activity of pathogenic Th-17 cells and attenuated the proinflammatory events²¹⁶ (Fig. 3C). The hyperactivity of pathogenic Th-17 cells is also directly linked with the progression of RA. Therefore, suppressing the IL-17-releasing Th-17 cells can also provide beneficial effects for the treatment of RA. The researchers claimed that targeting the signal transducer and activator of transcription 3 (STAT3) signalling pathway by delivering antioxidant CoQ10 or ubiquinone can block the inflammatory activities of Th-17 cells.²¹⁷ Hence, the immunotherapeutic hybrid NP-loaded CoQ10 delivery downregulates the IL-17 level and reduces Th-17 cell-mediated inflammation.²¹⁷ SLE is a fatal autoimmune disorder characterized by higher infiltration of pathogenic T cells, autoreactive antibody production, and loss of self-tolerance. Altered Ca²⁺ signalling has also been implicated in the pathogenesis of SLE, where elevated Ca²⁺ levels affect T-cell receptor signalling with deleterious autoimmune responses.⁵⁶ The voltage-gated potassium Kv 1.3 channels that are greatly expressed by activated effector Tm cells play a crucial role in maintaining Ca²⁺ balance *via* regulation of membrane potential. Targeted depletion of potassium Kv 1.3 channels with therapeutic NPs enables the correction of autoreactive immune responses in the treatment of SLE. Khodoun *et al.* developed a novel nanoplatfrom that disrupts Ca²⁺ signalling by selectively downregulating the potassium Kv 1.3 channels of CD8⁺ effector Tm cells. Thus, targeted depletion of potassium ion channels shows an effective reduction in Ca²⁺-mediated T-cell stimulation, CD40L, and IFN- γ levels that collectively alleviate the disease progression.⁵⁶

4.2. B-cell targeting nanotherapeutics

B cells are the antibody-producing lymphocytes that serve humoral immunity in the body. To maintain an appropriate immune balance, the activity of B cells can be harnessed in different ways. In the treatment of several immune disorders, B cells need to be activated, while in other cases, B-cell activities are essentially required to be downregulated.

4.2.1. B-cell activation. The activation of B cells and the generation of effective broadly neutralizing antibodies (bnAbs) is one of the central strategies for developing vaccines against multiple diseases. The use of NPs offers several advantages in the design and development of effective vaccines that can target the immune system and facilitate the activity of B cells. Hence, researchers are striving to develop effective nano-vaccines by loading different immunogens in NPs. Depending on the properties of loaded immunogens and nanomaterials, therapeutic nano-vaccines exhibit varied levels of B-cell stimulation to trigger antibody responses. Following the fact that disease-relevant antigen species can activate B cells, several approaches have been made for developing anti-HIV vaccines. The HIV-1 envelope trimeric protein fragments are considered as strong immunogens to induce antibody responses by activating B cells.²¹⁸ The administration of free and soluble protein antigens often fails to provoke significant B-cell acti-

vation. To overcome this limitation, Ni²⁺ ion-bound liposomal NPs surface tethered with histidine-tagged HIV-1 spike protein trimers were developed as novel nano-vaccines that strikingly stimulate B cells.²¹⁸ As a result, a strong elevation in neutralizing antibody (nAb) level was generated by the stable VLP nano-vaccines as compared with their bare protein antigen counterpart (Fig. 4). Hence, controlling the stability of trimeric antigens displayed on liposomal NPs is crucial for their immunological impacts on the host. The covalent attachment of viral immunogen spike protein onto the NP surface provides higher stability to the immunogens and, therefore, helps in obtaining the long-term clinical efficacy of the nano-vaccine. Besides antigen, the sphingomyelin constituents of the LNPs also contribute to immunogenicity and can synergize the activity of nano-vaccines to stimulate germinal center B-cell response and antibody production to encounter HIV strains.²¹⁹ Similarly, a study has been carried out with HIV envelope protein-conjugated ferritin NPs that also demonstrated the anti-retroviral efficacy of particulate nano-vaccines over the soluble free-form of antigens.²²⁰ In another study, Moyer *et al.* demonstrated direct activation of B cells and production of bnAb with the help of modified HIV envelope protein-decorated NPs.²²¹ The engineered eOD immunogen (outer domain of the HIV-1 glycoprotein-120), tethered on the surface of NPs, promoted direct internalization by B cells, thereby displaying better antigen processing, antigen presentation, and enhanced activation efficiency of B cells.²²¹

The activation of B cells and subsequent release of bnAbs could be the same method of molecular intervention for protection against deadly strains of influenza. The immunogenic properties of multiple viral protein fragments are harnessed to develop nano-vaccines with the ability to activate B cells and trigger the generation of bnAbs. Previous studies demonstrated that the extracellular domain of ion channel matrix membrane protein 2 (M2e) of influenza virus A exhibits immunogenic properties but has limited efficacy in inducing significant immunological responses. The combination of soluble adjuvant CpG oligonucleotide and M2e-peptide antigen produces a several-fold increase in immune stimulation by eliciting greater antibody responses. Thus, the AuNPs arrayed with M2e-peptide and CpG adjuvant on their surface offered superior activation of B cells, therefore producing broad-spectrum antibody responses against several strains of influenza, such as H1N1, H5N1, and H3N2.²²² Targeting the B-cell receptors (BCRs) with ligand antigens is another advanced strategy employed in the activation of B cells. Haemagglutinin (HA) antigen fragment obtained from the influenza virus acts as an immunogenic ligand that binds to sialic acid residues of BCRs. The coupling interaction between ligand-HA and sialic acid target residues of BCRs leads to significant activation of B cells.²²³ Thus, several novel approaches have been attempted for developing HA-displaying NPs that can recognize the BCRs and exhibit strong antigen-specific immune responses against influenza infection. Following this rationale, recombinant gene-expressed HA-arrayed ferritin NPs have emerged as novel nano-vaccine candidates for protection against influenza. The

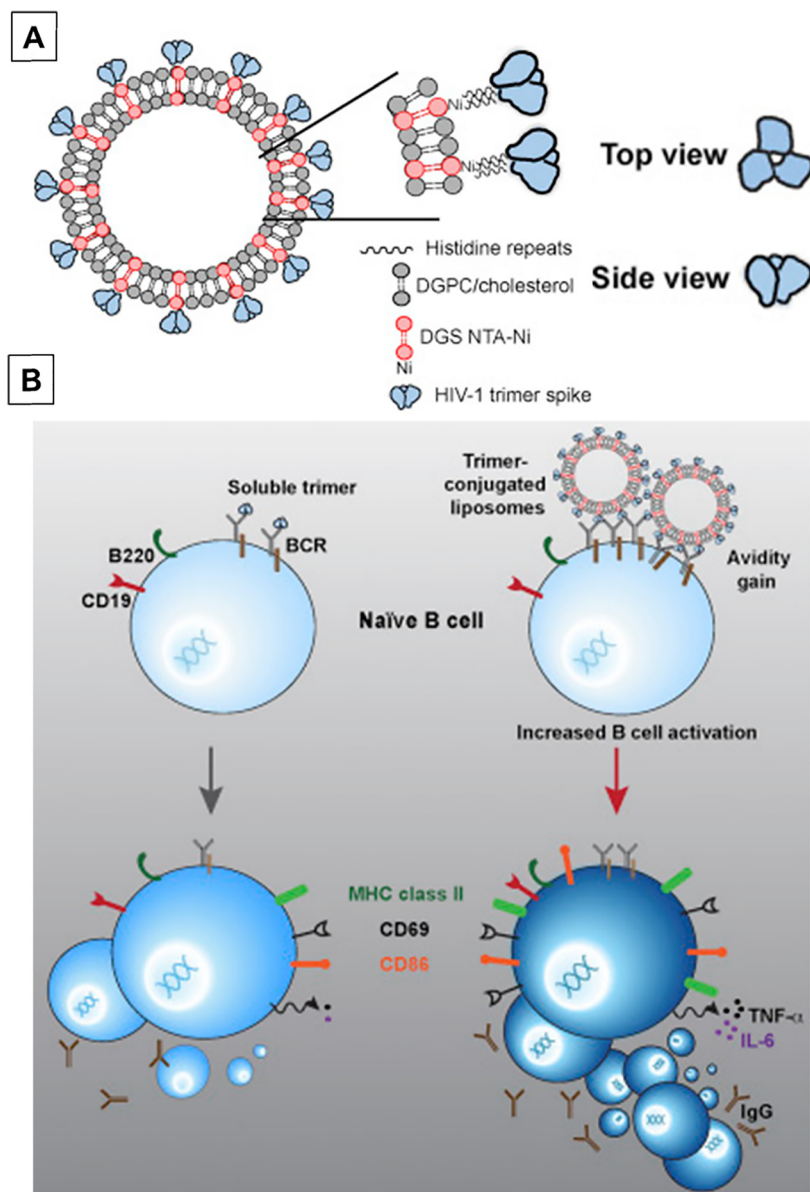


Fig. 4 (A) Pictorial representation of liposomes decorated with HIV-1 trimer spike. Magnified part illustrates binding of the 6-histidine repeats (His6 tag) present as a fusion on the C terminus of each protomer within each trimer to the Ni + 2 chelated at the hydrophilic head group of the DGS-NTA (Ni) polar lipid. (B) Schematic representation of activation of B cell using liposome decorated with HIV-1 trimer spike. Liposome conjugated with HIV-1 trimer spike leads to enhanced B-cell activation as compared with soluble factor. Reproduced with permission from ref. 218, Copyright 2016, Elsevier.

highly ordered intact HA-ferritin nano-vaccine showed strong germinal center stimulation as well as direct activation of B cells *via* receptor ligand coupling interaction.²²⁴ The effective stimulation of BCRs consequently results in a higher level of nAb production. However, instead of displaying homotypic HA antigens, the colocalization of heterotypic antigens confers superior qualitative and quantitative B-cell activation and broad antibody responses. The engineered gene construct-encoded mosaic heterotypic HA-decorated immunotherapeutic ferritin NPs showed excellent cross-reactive antibody production. Hence, the multipronged immunogenicity of mosaic NPs shows significant neutralization of the H1N1 strain of

influenza virus.²²⁵ Formation of germinal centers (GC) and activation of B cells are necessary for a robust antibody response in protective immunity against hepatitis B virus (HBV) infection. The surface protein originated in the preS1-domain of HBV and is known for its immunogenic properties that can be utilized to activate B cells in anti-HBV immunotherapy. Recent studies revealed that the concurrent delivery of antigen into DCs and lymphatic macrophages strikingly enhanced B-cell activation in coordination with follicular helper T cells.²²⁶ Hence, the ferritin NPs displaying preS1 surface functionalization targeted dual cells and elicited a robust antibody response against HBV.²²⁶

4.2.2. B-cell suppression. Malfunctioning of B cells often leads to the initiation and progression of diseases ranging from autoimmune to inflammatory disorders. To protect the internal organs and tissues of the body, autoreactive B-cell-driven activities need to be downregulated. Mantle cell lymphoma (MCL) is a form of non-Hodgkin lymphoma (NHL) characterized by pathogenic expressions of BCRs. Inhibition of BCRs is of great clinical significance for halting B-cell lymphoma progression.²²⁷ CD38 is a BCR found to be overexpressed on MCL B-cells and is correlated with the expansion of B-cell malignancy. Targeting CD38 receptors with anti-CD38 antibodies holds great promise for the remission of B-cell lymphomas. Previous clinical trials with anti-CD38 mAb have also demonstrated the therapeutic efficacy of anti-CD38 mAb, which is quite promising against multiple myeloma cells.²²⁸ Therefore, arraying the surface of NPs with anti-CD38 mAbs helps in developing therapeutic nanoplateforms that can be useful in targeting CD38-expressing B cells. Furthermore, to enhance the immunotherapeutic efficacy against MCL B-cells, the mAb-conjugated, therapeutic siRNA entrapped NPs are successfully used to downregulate tumorigenic cyclin D1 biomarkers (overexpressed in MCL).²²⁹ Targeting the CD20 receptors of NHL B-cells with clinically approved mAbs like Rituximab (anti-CD20 antibody) often shows antibody resistance. To overcome this therapeutic barrier, Li *et al.* developed novel NPs conjugated with distinct anti-CD20 mAbs that efficiently target CD20⁺ B cells.²³⁰ The NPs bearing both type I anti-CD20 mAb Rituximab and type II mAb 11B8 led to significant depletion of antibody-resistant B-cell lymphomas²³⁰ (Fig. 5A). Diffuse large B-cell lymphoma (DLBCL) is the most frequent form of NHL and exhibits greater levels of CXCR4 receptor expression, which could be a potential target for aberrant B-cell depletion. The T22-GFP-H6 protein nano-carrier is known for its selective CXCR4⁺ B-cell targeting potential. The conjugation of microtubule-targeting agents such as monomethyl auristatin E with T22-GFP-H6 forms a strong nano-assembly that selectively targets the B cells and reduces the disease progression.²³¹ The dysregulation of mitogen-activated protein kinase (MAPK) intracellular signalling cascades often leads to B-cell malignancies. CD19 is an immunoglobulin family-based transmembrane glycoprotein receptor that is strongly expressed by B cells in almost all stages of their maturation. In response to molecular signals, the CD19 receptors establish communication between lymphoid B cells and micro-environments. Selective targeting of CD-19 receptors and subsequent inhibition of MAPK signalling represents an excellent approach for suppressing leukemic B cells. The anti-CD19 antibody-modified NPs, loaded with bacterial lethal factor toxin (MAPK inhibitor), have shown the targeted delivery of cargo toxins to CD19⁺ B cells and effectively abrogate leukemic B cells²³² (Fig. 5B). By secreting autoantibodies and rheumatoid factors, the autoreactive B cells play pathogenic roles in the development of RA. Being an APC, the B cell also promotes the activation of T cells that promote inflammatory cytokine release in the joints of the body. Thus, selective targeting of autoreactive B cells is essential in the treatment of RA. The

earlier research findings suggested that the B-cell activating factor receptor (BAFF-R) plays a constructive role in regulating the growth and maturation of B cells. In targeted downregulation of BAFF-R to reduce the population of B cells, RNA interference has become a successful tool in the management of arthritis. Hence, Wu *et al.* demonstrated that BAFF-R siRNA encapsulated therapeutic NPs protectively deliver cargo into B cells. As a result, the suppression of B cells showed immunomodulation and inhibition of inflammatory symptoms in RA.²³³ Researchers also found that autoantibodies specific for citrullinated proteins (ACPA) are produced by autoreactive B cells that display pathogenic activities in the development of RA. Hence, ACPA-producing B cells could be another potential target in the prevention of RA. In this regard, therapeutic NPs having both autoreactive B-cell selectivity and B-cell targeted complement-mediated cytotoxic properties can potentially eliminate pathogenic B cells. Fibrin-derived β 60-74cit synthetic peptide modification on NPs acts as a specific ligand in recognizing the B cells, while another peptide (derived from gp120 of HIV-1) drives complement-mediated cytotoxicity to eliminate autoreactive B cells. Therefore, dual peptide-modified NPs can be specifically engaged in B-cell-targeted immunotherapies. Activating the complement system of the immune system accelerates the attenuation efficacy of B cells. In this regard, the modified effector peptide gp120 can be used to activate the complement component. Therefore, administration of NPs conjugated with multiple copies of the β 60-74cit and gp120 peptides significantly exhibits the targeted depletion of B cells *via* activation of C5b-9 membrane attack complex.²³⁴ Bruton's tyrosine kinase (BTK) is a non-receptor protein molecule that is critically required by B cells for their growth and development. In several aberrant B-cell immune diseases, the BTK plays pathogenic roles that lead to poor disease prognosis. Some BTK-inhibitor therapeutics are already clinically approved, while others are undergoing clinical trials. To facilitate the efficacy of these BTK inhibitors, researchers are developing smart and targeted nanomedicines that specifically deliver these approved drugs. Thus, dysregulated B-cell targeting and precise delivery of BTK inhibitors have become possible by employing antibody-modified NPs. A recent study demonstrated that different BCR-targeted BTK inhibitors delivered by antibody-modified NPs significantly kill dysregulated B cells. The anti-CD19-modified, drug-loaded NPs bind to CD19 receptors and then deliver the therapeutic cargo intracellularly, while anti-B220-coupled NPs recognize B220 receptors and mediate surface localization of loaded BTK-inhibitors.⁵⁸

4.3. Macrophage-targeting nanotherapeutics

Macrophages exhibit diverse phenotypic variations; generally, two common types of macrophage (M1 and M2) play significant roles during immune surveillance. M1 macrophages are known to have pro-inflammatory responses, while M2 macrophages exhibit anti-inflammatory responses. Upon stimulation, M0 macrophages (resting macrophages) give rise to M1 and M2 macrophages by a process called macrophage

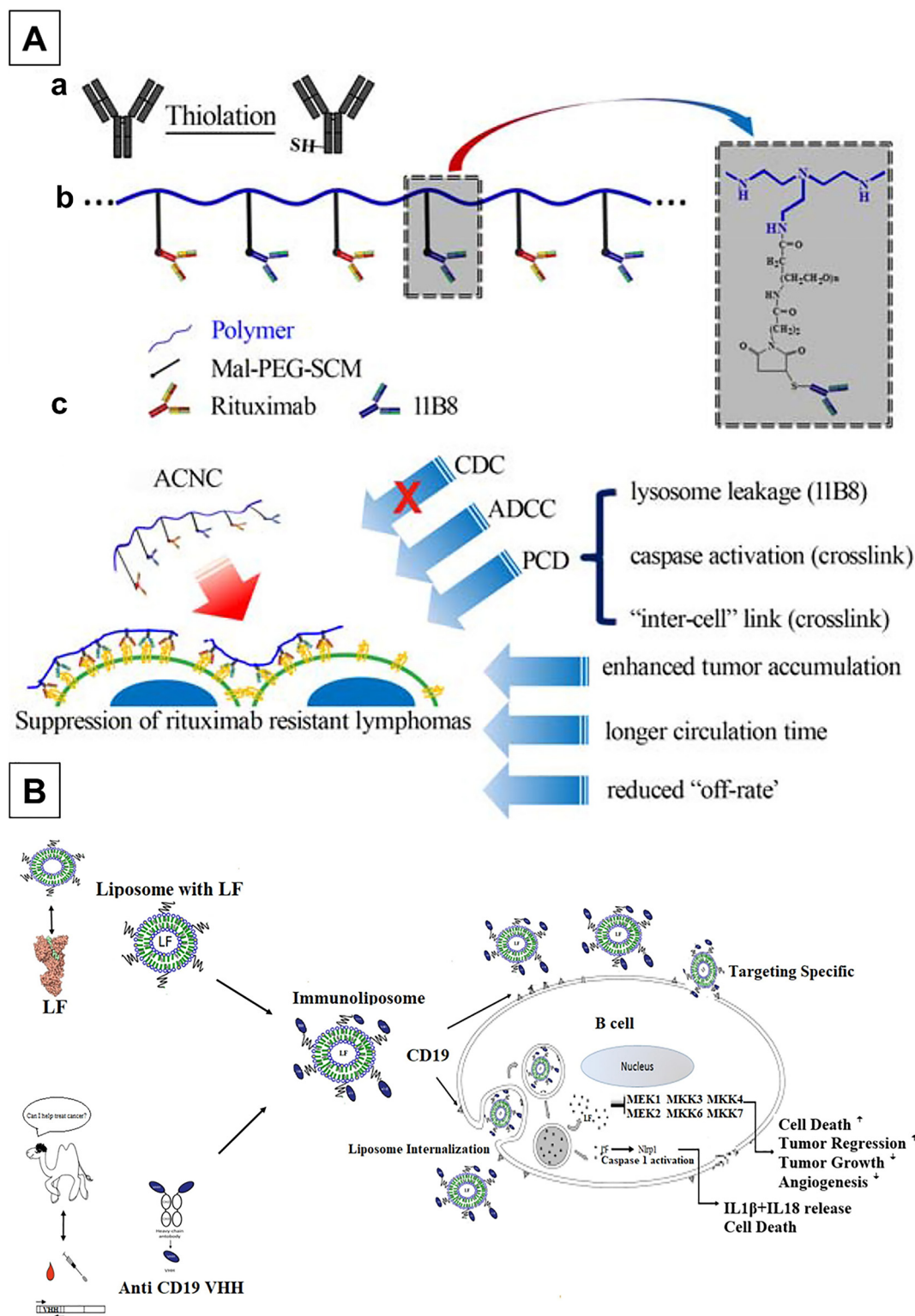


Fig. 5 (A) Fabrication and characterization of anti-CD20 nanocluster (ACNC). (a and b) Schematic depiction of nanoclusters; (c) 3 proposed major pathways through which ACNC causes B-cell depletion are complement-dependent cytotoxicity (CDC), antibody-dependent cellular cytotoxicity (ADCC) and induction of programmed cell death (PCD). These effective mechanisms cause suppression of rituximab-resistant non-Hodgkin lymphoma (NHL). Reproduced with permission from ref. 230, Copyright 2016, Elsevier. (B) Scheme representing the steps involved in the design and production of the immunoliposome. Lethal factor (LF)-loaded immunoliposome was internalized the cell by the surface receptor CD19 followed by blocked phosphorylation of MAPK pathway. Reproduced with permission from ref. 232, Copyright 2021, Elsevier.

polarization.²³⁵ Different molecular stimuli influence the specific mood of macrophage polarization, which is ultimately reflected in either pro-inflammatory or anti-inflammatory immune responses.^{236–238} Thus, targeting the event of macrophage polarization eases the advance of therapeutic intervention against several immune disorders.²³⁹

4.3.1. Macrophage activation. The tumor-associated macrophages (TAMs) are predominantly present in the TME, where they preferably express into M2-phenotype macrophages. Being anti-inflammatory in nature, the M2 macrophage forms an immunosuppressive TME and promotes the cancer pathology to progress further.²³⁸ The greater expressions of TAMs into M2-macrophages resist several anti-tumor immunotherapies, thereby necessitating novel strategies that target the TAMs and polarize them into M1 macrophages. Stimulation of immune cells with clinically approved NPs is considered safe to use in disease therapy. Ferumoxytol, an iron oxide NP formulation, has been approved by the US FDA in the therapy of iron-deficient anemia.¹³⁵ Previous studies demonstrated that ferumoxytol can modulate and activate TAMs into the pro-inflammatory tumor-suppressive M1 phenotype.¹³⁵ Particularly, the selective targeting of immunosuppressive TAMs and polarizing into M1 phenotype has always been in demand. CD206, or macrophage mannose receptor 1, is a transmembrane receptor found to be highly expressed in TAMs and could be heralded as a prime target in the specific delivery of therapeutics. Currently, targeted NPs carrying synthetic mRNA drugs have been considered as emerging classes of new therapeutics with superior ability to modulate the activities of M2-directing TAMs without any systemic toxicity concerns. Studies demonstrated that di-mannose-targeting ligand-decorated and *in vitro*-transcribed synthetic mRNA-loaded therapeutic NPs specifically target the TAMs.²⁴⁰ Hence, targeted intracellular delivery of engineered mRNA expresses interferon regulatory factor 5 and IKK- β kinase proteins that cooperatively modulate the TAMs to convert into anti-tumor M1 phenotypes.²⁴⁰ The SIRP- α receptor on macrophages interacts with the “don't eat me” signal receptor CD47 of cancer cells, thereby inhibiting macrophage-based phagocytosis. Hybrid extracellular nanovesicles (hNVs) characterized by SIRP- α receptor variants showed greater affinity for binding to CD47 receptors and blocked the immunosuppressive CD47–SIRP- α interaction. In combination with a stimulator of interferon gene (STING) agonist, the hNVs disrupted the CD47–SIRP- α interaction axis and activated M1 macrophages by modulating the TAMs, which resulted in the reduction of the immunosuppressive nature of the TME²³⁵ (Fig. 6A).

The imbalance between two phenotypic macrophages and the higher number of colon-resident M1 phenotypes drives prolonged local tissue inflammation due to its proinflammatory activities. Ulcerative colitis (UC) is an inflammatory disease where higher levels of M1-phenotypic macrophages are associated with local tissue inflammation. Although traditional therapeutics are used to treat UC, the non-specific drug delivery leads to poor therapeutic outcomes. Activation and polarization of anti-inflammatory M2 phenotypes in local

inflammatory tissues is an effective approach for treating UC. Macrophage membrane-coated biomimetic nanomedicines entrapped with synthetic drugs significantly polarized and activated the M2 phenotype from M1 macrophages.²⁴¹ β -CD NPs loaded with rosiglitazone exhibited ROS-responsive drug release in epithelial cells of the colon, where inflammation-mediated cellular stress is elevated. Moreover, the macrophage coating on NPs not only mimics to enable infiltration into the inflammatory vicinity, but also absorbs inflammatory cytokines to abolish pathogenic inflammation.²⁴¹ Activation of macrophages has also been observed in the augmentation of osteogenic differentiation. Mechanistically, stimulating the macrophages releases Oncostatin M (OSM), which acts as a potential inducer of osteoblast differentiation. In this regard, Hirata *et al.* have illustrated that activating the monocyte-derived macrophages with carbon nanohorns (CNHs) promoted OSM expression and enhanced matrix mineralization by modulating the STAT3 signalling pathway.²⁴² As a result, a greater level of alkaline phosphatase, an osteoblast differentiation marker, has been observed. Hence, CNH-based macrophage activation and release of osteoinductive factors can promote the development of potential regenerative nanomedicines.²⁴²

The regenerative or pro-healing properties of M2 phenotypes have been explored in treatment for wound healing. On exposure to external magnetic fields, the superparamagnetic iron oxide NPs (SPIONS) catalyzed M2 macrophage polarization from M1 phenotypes, which resulted in enhanced regeneration and self-healing of wounds.²⁴³ Mechanistically, the SPION-based magnetic field-driven M2-macrophage activation promoted better organization of actin filaments and enlargement in cellular topology, therefore enhancing the regenerative functionality of the wounded tissues.²⁴³ Immunosuppressed individuals are more prone to fungal infections. Hence, polarization and activation of proinflammatory M1 macrophages could be clinically beneficial for the prevention of fungal infection. Targeting the activation of TLR-4 receptors on macrophages and re-programming the M2 phenotypes into M1 phenotypes can result in significant stimulation of the immune system that provides protective immunity. Recently, Gao *et al.* developed a novel therapeutic nanopatform that targets several molecular pathways to polarize the M1 macrophage from its M2 phenotypes.²⁴⁴ Therapeutic NPs coated with mannosylated chitosan polymer specifically activated TLR-4 receptors, where TLR agonist chitosan promoted the M2 to M1 macrophage transition. The mannose modification of NPs confers mannose receptor-mediated selective uptake of imatinib-loaded nanotherapeutics by macrophages. Hence, targeted delivery of imatinib drugs blocked STAT6 signalling and reduced the immunosuppressive M2-macrophage population. Thus, the use of multifunctional NPs accelerated M1-macrophage functionalities and, therefore, stimulated the immune system to fight against fungal infection caused by *Candida albicans*.²⁴⁴ Previous studies also demonstrated that the internalization of antigen-loaded NPs can greatly activate macrophages and trigger the release of immunostimulatory cyto-

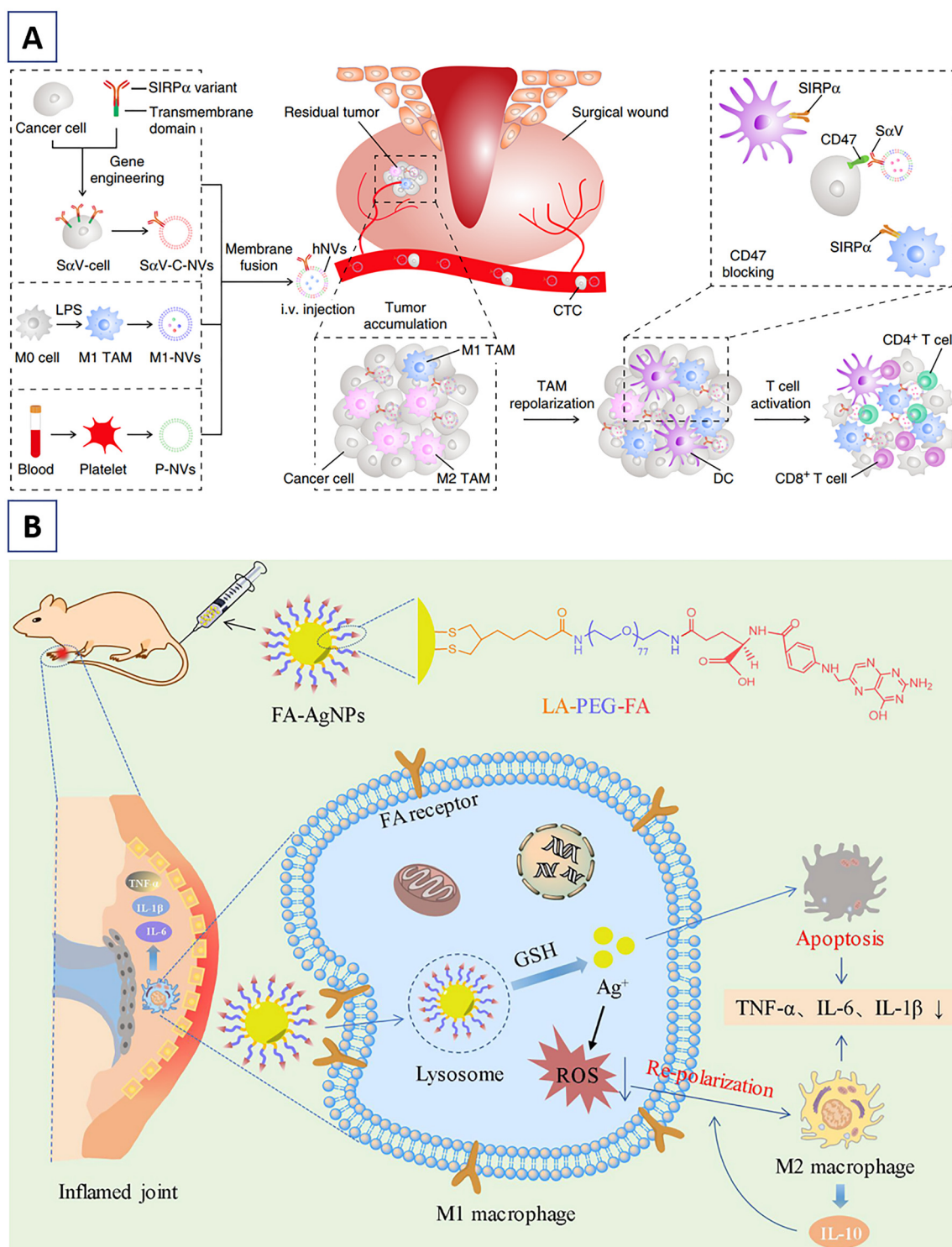


Fig. 6 (A) Schematic illustration of the hybrid cell membrane nanovesicles (hNVs) comprising engineered cancer cell-derived nanovesicles overexpressing signal regulatory protein alpha (SIRP α), platelet-derived nanovesicles and M1 macrophage-derived nanovesicles. The engineered hNVs, on interaction with circulating tumor cells, accumulate in the post-surgical tumor bed, repolarize tumor-associated macrophages towards M1 phenotype, and block the CD47-SIRP α 'don't eat me' pathway for effectively enhancing macrophage-based phagocytosis of cancer cells, as well as promote antitumor T-cell immunity. Reproduced with permission from ref. 235, Copyright 2020, Springer Nature. (B) Scheme representing the mechanism of action of folic acid-modified silver nanoparticles (FA-AgNPs) against rheumatoid arthritis. In response to the intracellular GSH, the developed FA-AgNPs release Ag⁺ to synergistically induce apoptosis of M1 macrophages and scavenge reactive oxygen species to promote M2 macrophage polarization in inflamed synovial joints. Reproduced with permission from ref. 246, Copyright 2021 Elsevier.

kines. Polymerosome NPs co-loaded with OVA antigen and MPLA adjuvant stimulated macrophage cells. The activation of immune macrophages by dual immunogens triggered the release of potent immune stimulatory IL-6 and TNF- α cytokines that helped in generating strong Ag-specific antibody responses.²⁴⁵

4.3.2. Macrophage suppression. In some treatment strategies, M1 macrophages need to be abolished to avoid pro-inflammatory immune responses, while suppressing the M2 phenotype leads to immune stimulation in the treatment of immunosuppressive conditions. Although polarizing the TAMs towards M1 macrophages with nanomedicines prevents the immune-suppressive environment found in the TME, direct inhibition of TAMs is another key strategy in re-establishing the immune balance in the TME. Displaying “eat me” signals on the surface of NPs attracts the TAMs for phagocytosis, and this event can be useful in the design of nanotherapeutics to target and deplete the pathogenic TAMs specifically.²⁴⁷ Negatively charged phospholipid phosphatidylserine (PS) naturally acts as an “eat me” signal during apoptosis, and was engineered onto the surface of dasatinib-loaded NPs to promote phagocytosis by macrophages *via* PS receptor. The internalization of PS-modified nanomedicines by TAMs resulted in the delivery of cargo dasatinib within the macrophages, leading to the depletion of TAMs.²⁴⁷ Local macrophage accumulation and proliferation are observed in the inflammatory pathogenic plaque of atherosclerosis, where mid and large-size arteries are generally affected.²⁴⁸ Additionally, ablation of the macrophages without recruiting monocytes could be an appealing strategy to prevent the prevalence of atherosclerosis. Specifically, delivering statin drugs to atherosclerotic plaque-forming macrophages can help to reduce the inflammatory activities exerted by local macrophages. The administration of biomimetic high-density lipoprotein (HDL)-based NPs loaded with HMG-Co-A reductase inhibitor simvastatin restrained the proliferation of local macrophages by halting the mevalonate metabolic pathway without attracting the blood monocytes.²⁴⁸ Moreover, in combination with oral statin delivery, the administration of simvastatin-loaded HDL NPs prolonged macrophage suppression and curtailed the disease pathology.²⁴⁸

Greater M1-macrophage infiltration and macrophage-mediated proinflammatory autoimmune responses are observed in RA.²⁴⁶ The highly reactive M1-phenotypic macrophages release several proinflammatory mediators, such as TNF- α , IL-6, and IL-1 β cytokines, that together aggravate synovial tissue inflammation. The anti-rheumatic AgNPs with folic acid conjugation selectively target M1 macrophages that are characterized by overexpression of folate receptors.²⁴⁶ The selective uptake and reduced glutathione (GSH)-responsive intracellular liberation of Ag⁺ ions led to M1-macrophage apoptosis, which consequently reduced the level of inflammatory cytokines. In addition to the M1-macrophage apoptosis, the released Ag⁺ ion also scavenged reactive oxygen species (ROS) in inflammatory tissues to augment M2-macrophage polarization and ameliorate the inflammatory responses in

RA²⁴⁶ (Fig. 6B). Other than inflammatory macrophages, bone-resorbing osteoclast cells (OCs) are found to be greatly associated with the persistent pathogenesis of RA.²⁴⁹ These OCs additionally trigger the release of proinflammatory cytokines and exacerbate synovial inflammation. Moreover, both inflammatory macrophages and OCs are known to express high levels of MMP-9 protease in an inflammatory microenvironment that facilitates the severe progression of the disease. Earlier studies have claimed that the $\alpha\beta3$ integrin receptors are highly expressed by both macrophages and OCs that play a pathogenic role in the development of RA.²⁴⁹ Thus, selective elimination of inflammatory macrophages and bone-resorptive OCs is essential to attenuate the persistent and advanced form of RA. PEG- and RGD-tripeptide-modified PLGA NPs loaded with cytotoxic agents were demonstrated to eliminate macrophages and OCs selectively by targeting integrin receptors. In the RA microenvironment, MMP-9-responsive PEG cleavage and subsequent internalization of RGD-tagged, drug-loaded PLGA NPs occurred by both macrophages and OCs due to the RGD- $\alpha\beta3$ integrin interaction. Thus, multifunctional NP-mediated dual-targeted delivery of a cytotoxic agent (celastrol) significantly ablated both inflammatory macrophages and OCs in the prevention of RA.²⁴⁹ Specific targeting of synovial M1 macrophages is believed to reduce the pathogenicity of osteoarthritis (OA), which is considered to be another globally prevalent form of arthritis. The abundance of CD16/32 receptors on M1 macrophages is a potential target for the delivery of therapeutics to suppress the synovial macrophages. Due to metabolic dysregulation, the active M1 macrophage releases inducible nitric oxide synthase (iNOS), which catalyzes the biosynthesis of gaseous nitric oxide (NO).⁵⁷ In turn, NO affects the intracellular O₂ level by interfering with mitochondrial respiratory machinery and introducing mitochondrial stress events, thereby leading to the production of hydrogen peroxide (H₂O₂) and ROS. Targeted metabolic reprogramming of M1 macrophages and suppressing their activity by inhibiting gaseous inflammatory mediators (NO, H₂O₂) ameliorated the dominance of synovial macrophages. The anti-CD16/32 antibody-modified ZIF-8 MOF-mediated delivery of therapeutics has successfully been employed in the prevention of NO and H₂O₂ release.⁵⁷ Synovial macrophages targeting ZIF-8 NPs simultaneously delivered *S*-methylisothiourea hemisulfate salt (SMT) and catalase (CAT) to reprogram the metabolic events. Intracellular pH-responsive delivery of SMT from ZIF-8 halted the generation of NO by blocking the iNOS, while CAT neutralized H₂O₂ and facilitated the production of O₂. The released O₂ also diminished the activity of the M1-polarizing inducer molecule hypoxia-inducible factor α (HIF- α) and inhibited osteoarthritic pathogenic inflammation.⁵⁷ Being a prime reservoir of viruses, macrophages are also involved in the further spread of viruses to other cellular compartments. The greater antigen presentation and prolonged viral infectivity of CD4/CD8 T cells *via* interaction between virus-reserved macrophages and T cells possess immunological implications in HIV infection. Hence, the greater reservation of viruses by macrophages in the pre-infection stage sets therapeutic barriers in

the way of antiretroviral therapy. Preventing the viral-reservoir macrophages at the pre-infection stage using nanomedicines has shown beneficial therapeutic outcomes for halting the cell-to-cell spreading of viruses. The 2nd generation carbosilane dendrimer, composed of a silica core surrounded by 16 sulfonate moieties (G2-S16), directly suppressed the viruses within macrophages and also inhibited viral spreading from the macrophage to other immune cells.²⁵⁰ Thus, the therapeutic dendrimer NPs exert the ability to inhibit infected macrophage reservoirs and could be used in antiretroviral therapy. Excessive inflammation and multiorgan failure due to proinflammatory cytokine-mediated cytokine storms can result in fatal consequences in patients suffering from the recent pandemic SARS-CoV-2 virus. Virus-infected cells surrounding macrophages release proinflammatory cytokines like IL-6, IL-1 β and recruit other immune cells in the lungs, which together lead to the initiation of cytokine storms. The hyperactive macrophage also stimulates the invaded neutrophils to secrete neutrophil extracellular traps (NETs) to exacerbate the cytokine storms with more deleterious effects. Preventing macrophages and abolishing cytokine storms is an essential strategy to alleviate the lethal inflammatory immune responses in COVID-19 infection. A recent study demonstrated that macrophage membrane-coated biomimetic NPs, loaded with antiviral drugs, have great potential to inhibit macrophages and arrest the events of cytokine storms.²⁵¹ The coating of macrophage membrane on NPs absorbed inflammatory cytokines and chemokines, and inhibited the actual macrophage population from being activated. The inhibition of macrophages downregulated the secretion of NETs from neutrophils. Moreover, the macrophage membrane-coated NPs, displaying angiotensin converting 2 (ACE-II) receptors, help in specific binding to spike proteins of SARS-CoV-2 and deliver the antiviral therapeutics to encounter the viruses.²⁵¹ Even after recovery from COVID-19 infection, individuals are always at high risk of pulmonary fibrosis (PF). The transition of monocyte macrophages into alveolar macrophages and the release of inflammatory profibrotic cytokines are known to play pathogenic roles in the progression of PF. Interrupting the profibrogenic macrophage transition and abrogating the cytokine release has been proved to be an effective approach for the alleviation of PF.²⁵² The CD206 receptor-expressed profibrogenic macrophage subpopulation targeted NP-based protective delivery of TGF β 1 siRNA and showed promising results in controlling PF. The mannose ligand-modified therapeutic albumin NPs targeted the CD206⁺ macrophage subsets and effectively delivered therapeutic cargo siRNA, which downregulated the fibrogenic TGF β 1 level to prevent lung fibrosis.²⁵²

4.5. Nanotherapeutics targeting DCs

DCs are another class of key immune cells that regulate immune functions *via* a subset of mechanisms like antigen presentation and T-cell modulation and, therefore, play a vital role in controlling immune tolerance and autoimmune responses in an organism. Aberrant DC activities are reported in many diseases, like type 1 diabetes, RA, multiple sclerosis,

etc. Furthermore, the DC also governs the acceptance and rejection of transplanted tissues in the host. Thus, tuning the activities of tolerogenic dendritic cells (toDCs) has central importance in the therapy of diseases.²⁵³

4.5.1. DC activation. Targeting the PRRs on DCs is an emerging approach to activate and mature the DCs to induce potent antitumor responses. The DCs express PRRs that recognise different PAMPs on antigens and promote the antigen presentation to trigger immune responses. TLR4 is one of the PRRs found on DC surfaces and is reported to be a prime target for stimulating the DCs. In this context, Rajput *et al.* developed antigen-loaded inulin acetate NPs to target DCs and stimulate the TLR4 signalling cascades.²⁵⁴ The developed nano-vaccine significantly modulated the activation and maturation of DCs by acting as TLR4 agonist. Moreover, encapsulation of antigen within the NPs resulted in a strong antibody response and an elevated level of immunomodulatory cytokines.²⁵⁴ The application of aggregation-induced emission (AIE) luminogen-coupled upconversion nanoparticles (AUNPs) in regulating the the ROS level in deep tissues made it feasible to restrict tumor growth.²⁵⁵ The high-intensity NIR irradiation directly kills the tumor cells by generating high levels of ROS, while the low-intensity NIR irradiation-based lower-level ROS promotes activation of DCs that induce immunogenic cell death in solid tumors. The initial high level of ROS generation by high-intensity NIR-exposure ablated the tumor cells and released TAA that eventually gets loaded onto administered UCNPs. The TAA-bound NPs were internalized by DCs, and on exposure to low-intensity NIR light the lower ROS production activated the DCs. Hence, the activation of DCs and antigen-bearing NP cross-presentation promoted CD8⁺ T cells and effectively inhibited tumor growth²⁵⁵ (Fig. 7A). Stimulating the TLR4 receptors and upregulating the NF- κ B signalling pathway in DCs promotes inflammatory cytokine production that helps in greater activation and maturation of DCs, thereby augmenting better antigen processing and inducing anti-tumor immune responses.²⁵⁶ In this context, antigen-expressing therapeutic mRNA nano-vaccines were developed to target DCs and activate them for enhanced antigen presentation as well as recruitment of T cells for immune stimulation. The cationic C1 LNP-based DC-targeted delivery of antigen-expressing mRNA exhibited strong activation of DCs *via* stimulation of TLR4 receptors and modulation of the NF- κ B signalling axis.²⁵⁶ The delivery of C1-mRNA nano-vaccines also resulted in greater expression of costimulatory CD80, CD86, and CD40 molecules on DCs that collectively elicited potent inflammatory responses toward immune-cold tumors.²⁵⁶ The cytosolic and endosomal PRRs of DCs are considered to be prime targets in the induction of innate as well as adaptive immunity. The NP-based antigen-expressing mRNA delivery to DCs and PRR-triggered induction of a strong immune response ease the way for the discovery of vaccines against infectious diseases caused by the HIV virus. Following this rationale, Coolen *et al.* developed mRNA nano-vaccines to stimulate the PRRs of DCs.⁹⁵ The PLA-NPs, decorated with cell-penetrating peptide and therapeutic mRNA, were efficiently internalized by

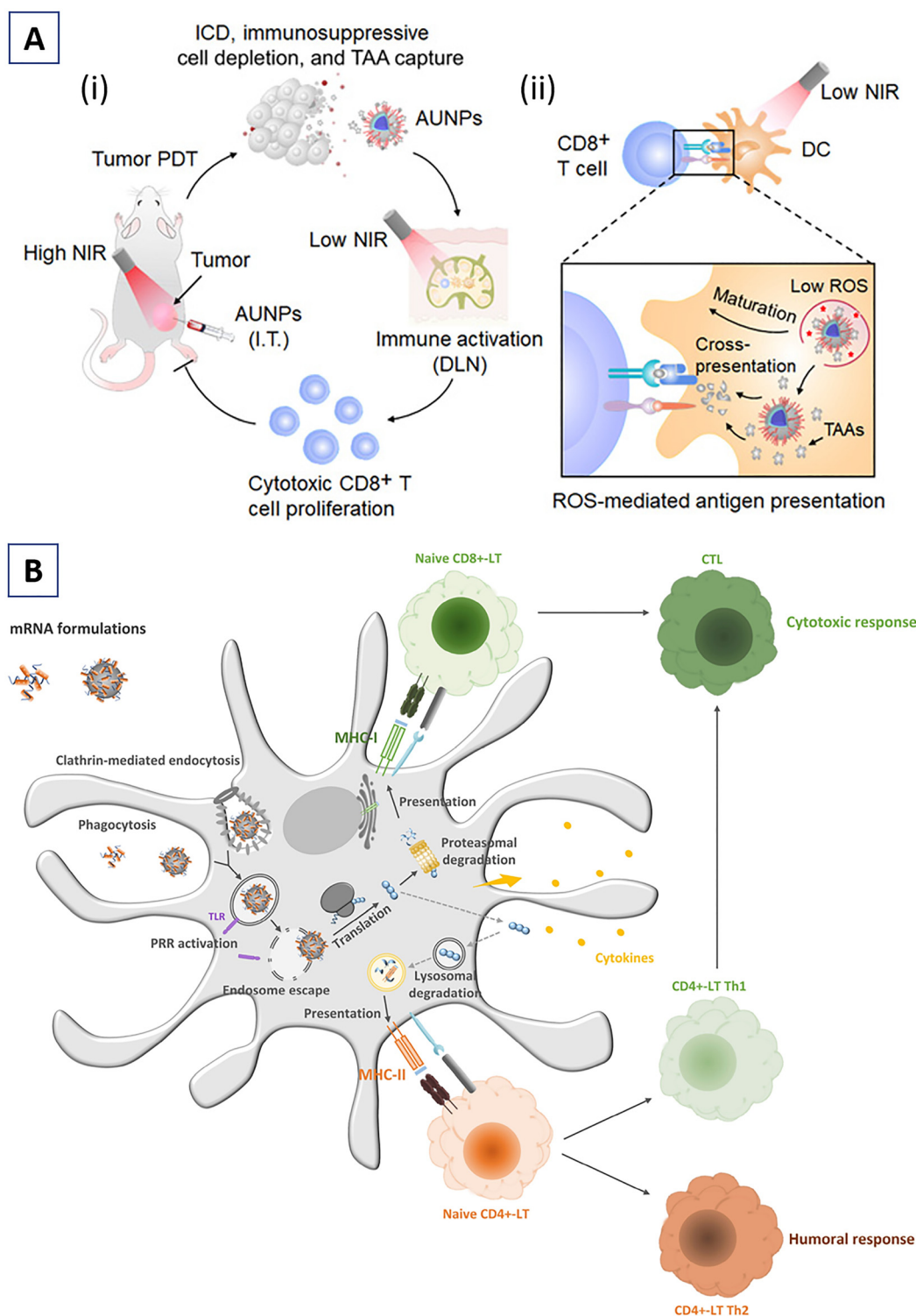


Fig. 7 (A) Schematic representation of dual-mode reactive oxygen species (ROS)-driven immunotherapy: (i) High-power near-infrared (NIR) light irradiation on the intratumorally injected aggregation-induced emission luminogen (AIE)-coupled nanoparticles (AUNPs) induces immunogenic cell death, reduces immunosuppressive cells and captures tumor-associated antigens (TAA). Furthermore, the TAA-loaded AUNPs are captured by the dendritic cells in the draining lymph node. Moreover, low-power NIR light promotes dendritic cell function through low-level production of ROS. Overall, this process induces the expansion of CD8⁺ T cells and inhibits the growth of residual tumor. (ii) Schematic illustration of the mechanism of dendritic cell activation through near-infrared light in draining lymph node. Reproduced with permission from ref. 255. Copyright 2020 American Association for the Advancement of Science. (B) Schematic illustration of the model for trafficking of LAH4-L1/mRNA polyplexes and PLA-NP/LAH4-L1/mRNA nanocomplexes into dendritic cells to induce immune response. Overall, the LAH4-L1/mRNA polyplexes and PLA-NP/LAH4-L1/mRNA nanocomplexes represent a potential platform for *ex vivo* treatment and mRNA vaccine development. Reproduced with permission from ref. 95. Copyright 2019, Elsevier. Abbreviations used: LAH4-L1: amphipathic cationic peptides and PLA-NP: poly (lactic acid) nanoparticles.

DCs through phagocytosis and clathrin-mediated endocytosis pathways, facilitating the delivery of Gag protein antigen-encoding cargo mRNA. The recognition of mRNA sequences by PRRs and simultaneous antigen expression of mRNA aided in the activation of PRRs. Specifically, the expression of Gag protein antigens significantly stimulated the endosomal TLR3 as well as DDX58 or RIG-I PRRs in DCs, thereby eliciting both innate and adaptive immune responses against retroviral infection⁹⁵ (Fig. 7B). Activation of lung-specific DCs and antigen cross-presentation are associated with stimulation of T cells that play a protective role in the prevention of bacterial and fungal infection. The intranasal administration of several vaccines is cited as having adverse immune reactions, and applications are restricted due to poor compliance.²⁵⁷ Recently, adjuvant-free pE α antigen-loaded self-assembled Q11-peptide nano-vaccine has been demonstrated to activate lung DCs with minimal inflammatory response. In particular, the intranasal delivery of nano-vaccine activated lung CD11b⁺ and CD103⁺ DCs that are essentially required for effector CD8⁺ and CD4⁺ T-cell responses, respectively.²⁵⁷ Moreover, the nanofiber E α Q11 vaccine stimulated the DCs and preferentially modulated the CD4⁺ T-cell functionality to secrete the IL-17 cytokine, which is known for offering immune protective activities in the respiratory mucosa of the lungs. Thus, intranasal delivery of nano-vaccine provided protective immunity against bacterial and fungal infections.

4.5.2. DC suppression. Autoreactive T-cell responses are upregulated due to the greater antigen cross-presentation by DCs that leads to aberrant immune disorders. Thus, suppressing DC-based antigen cross-presentation could be a potential intervention to abolish the induction of robust mixed lymphocytic reaction (MLR). Importantly, the DC-specific targeting and delivery of therapeutics to downregulate MLR is a formidable task for the scientific community. Earlier, Katakowski *et al.* demonstrated that single-chain antibody fragment-modified LNPs can particularly bind to DEC205⁺ receptors on DCs and efficiently deliver therapeutic cargos in a target-specific manner.⁴⁵ The research findings revealed that the delivery of therapeutic siRNA blunted the expression of costimulatory molecules such as CD40, CD80, and CD86 found on DCs, therefore halting the antigen cross-presentation to T cells. As a result, the DC-targeted therapeutic LNPs exerted efficacy in controlling the immunological events implicated in robust MLR.⁴⁵ The dominance of Th2 has been strongly observed in the pathogenesis of AR, where DCs play a crucial role in enhancing the differentiation of Th2 cells from naïve T lymphocytes. The costimulatory molecules CD80, CD86, and MHC II (I-A/I-E) on mature DCs participate in the differentiation of Th2 cells from the naïve T cells. Specific inhibition of DCs by downregulating the co-stimulator functionalities has shown DC resistance, which can result in the abrogation of Th2 activities. The NGR cyclic peptide ligand-modified therapeutic NPs, loaded with anti-inflammatory xanthatin molecules, exhibited suppression of CD80, CD86, and I-A/I-E costimulatory receptors.²⁵⁸ The NGR-peptide modification of NPs facilitated specific binding to CD13 receptors that are overexpressed on

DCs. Hence, DC-targeted anti-inflammatory drug delivery and subsequent resistance of DCs ameliorate the dominance of autoimmune T-cell responses and could be a beneficial modality for controlling AR. The CD40 receptors on DCs also act as a stimulatory signal in the activation of T cells and exhibit deleterious autoreactive responses during tissue or organ transplantation. Blocking the activities of DCs by inhibiting CD40 receptor signalling has great clinical significance in depleting T-cell activities toward the transplanted grafts. Recently, CRISPR-Cas9 molecular scissors were used to target and edit the specific gene that encodes CD40 receptors on DCs and reprogram the DCs by disrupting CD40 molecules.²⁵⁹ PEG-*b*-PLGA-based cationic lipid-assisted NPs were used to encapsulate and deliver the Cas9 mRNA (mCas9) and CD40 directing guide RNA (gCD40).²⁵⁹ Thus, NP-assisted delivery of the CRISPR/Cas9 tool disrupted CD40 signalling and consequently restricted T-cell-mediated transplant rejection. In multiple sclerosis, autoreactive immune responses are implicated by T-cell priming of DCs after being activated by myelin antigens. The myelin membrane of the central nervous system (CNS) gets affected by myelin-specific pathogenic Th-17 cells, where DCs play a crucial role in generation of Th-17 from CD4⁺ T cells. Recent studies revealed that ROS promotes the maturation of DCs by modulating several costimulatory receptors that ultimately provoke autoreactive CD4⁺ T-cell responses.²⁶⁰ Being antioxidant in nature, bilirubin NPs (BLNPs) showed promising therapeutic outcomes in the treatment of MS.²⁶⁰ Specifically, BLNPs abstract ROS and suppress the maturation of DCs, thereby further diminishing the conversion of CD4⁺ T cells to myelin-specific Th-17 cells. Thus, the use of therapeutic BLNPs could be a preventive strategy for blunting DC maturation and limiting the differentiation of naïve CD4⁺ T cells into autoreactive effector CD4⁺ T cells in the management of MS.

4.6. NP-assisted cytokine therapy

A subset of therapeutic cytokines ranging from interferons to interleukins is widely used in the treatment of several immune diseases. The administration of such therapeutic cytokines requires potential carriers for their protective delivery. IL-12 is one of the proinflammatory cytokines that drive antitumor immune responses against the immune-cold TME. Several pre-clinical studies have also been performed for the successful therapeutic translation of IL-12 in the treatment of cancer.²⁶¹ Researchers developed nanoplateform-based delivery systems for cytokines to reduce systemic toxicity issues, and that can be successfully employed in antitumor therapy. Mechanistically, the delivery of IL-12 molecules from engineered NPs stimulates the release of INF- γ in a downstream molecular cascade fashion, which in turn potentiates T-cell-mediated antitumor responses.²⁶² Lai *et al.* described an excellent alternative methodology to deliver the IL-12 cytokine *in vivo*.²⁶³ The IL-12-encoding specific mRNA sequence was delivered by LNPs, and subsequent expression of engineered mRNA to IL-12 stimulated CD44⁺ CD3⁺ CD4⁺ T cells, therefore augmenting the greater production of INF- γ .²⁶³ Not only did

IL-12 produce monotypic mRNA, but dual cytokine delivery approaches have also been carried out for synergistic tumor immunotherapy.²⁶⁴ Liu *et al.* have demonstrated the dual delivery of therapeutic mRNAs using LNPs. The IL-12 and IL-27-encoding mRNA-encapsulated LNPs showed striking ability against tumor growth by promoting the infiltration of CD8⁺ T cells, INF- γ , and TNF- α -secreting NK cells.²⁶⁴ Being a natural endotoxic cytokine, TNF- α is considered to be an excellent therapeutic molecule used in the remission of tumor progression. Encapsulation of TNF- α within porous NPs enables protective delivery, therefore conferring minimal acute toxicities.²⁶⁵ The release of cytokines from NPs preferably interrupted different phases of cell cycles (S/G2/M) and damaged the cancer cells directly to promote the retardation of tumor growth. Addition of cytokines to the therapeutic avenue of chemotherapy is an emerging approach for stimulating immune components that significantly limit the tumor progress. Wu *et al.* demonstrated that co-delivery of DOX and IL-2 *via* nanovesicles has a great immunomodulatory influence on the activities of DCs, T cells, and NK cells that confer anti-tumor immune responses.²⁶⁶ The anti-inflammatory and regenerative properties of IL-10 have shown promise in treating degenerative diseases. IL-10-producing exosomes effectively target injured kidney tubular epithelial cells *via* specific integrin receptors, modulating anti-inflammatory macrophage phenotypes and downregulating the mTOR pathway to maintain mitochondrial integrity. Thus, IL-10-expressing therapeutic exosomes could be a potent nanomedicine for managing acute kidney injuries (Fig. 8A).²⁶⁷ The prevalence of CNS-resident astrocytes and microglial cells has been reported to play a pathogenic role in the development of MS.²⁶⁸ The clinical application of the immunomodulatory cytokine INF- β is a promising choice of therapy. However, the low therapeutic index of the cytokine has always been a concern in the treatment of MS. Recently, González *et al.* developed INF- β loaded NPs for the intranasal delivery of cytokines to overcome the barriers of systemic administration.²⁶⁹ The efficient delivery of INF- β from the NPs alleviated MS pathogenesis by inhibiting the dominance of astrocytes and microglial cells and also depleting the activities of APCs. Furthermore, the IL-4 cytokine has been widely used in the treatment of several autoimmune diseases because of its anti-inflammatory activities. The IL-4 cytokine shows the ability to polarize the M1 macrophage into M2 phenotypes, thereby conferring a pro-regenerative immune response.²⁷⁰ However, the shorter half-life of IL-4 cytokine necessitates high-dose administration, which eventually causes toxicity issues. IL-4-conjugated AuNPs have been demonstrated to skew macrophage polarization towards the M2 phenotype effectively with minimal toxicity concerns.²⁷⁰ Hence, the NP-assisted delivery of IL-4 could be a novel therapeutic option for tissue regeneration, and is highly relevant in the management of muscle injuries.²⁷⁰ The proinflammatory activities of IL-4 and IL-13 cytokines exhibited deleterious effects in the development of asthmatic lung inflammation.²⁷¹ In particular, the IL4R α domain of IL-4 and IL-13 cytokines participated in their cognate receptor interaction during

airway inflammation. Hence, targeting and blocking the IL4R α subunit of these cytokines is a novel strategy for mitigating the symptoms of inflammatory lung diseases. Therapeutic NPs with anti-IL4R α antibodies tethered to their surface have been demonstrated to block the receptor-triggering activity of IL-4 and IL-13 cytokines, which consequently resulted in suppression of lung inflammation.²⁷¹ Recent studies identified IL-11 as one of the potent profibrogenic cytokine molecules engaged in several lung fibrotic disorders.²⁷² The binding of IL-11 to its cognate receptors stimulates the fibroblast activation and release of ECM *via* extracellular signal-regulated kinase (ERK) pathways. Greater deposition of ECM stiffens the lung tissues and drives fibrotic pathogenesis. Recently, researchers have developed siRNA-loaded therapeutic NPs for targeting the ERK pathways and downregulating IL-11 cytokine functionalities.²⁷³ Intranasal administration of siRNA-entrapped NPs diminished fibroblast activation and suppressed ECM deposition by interfering with the ERK and SMAD2 pathways. Thus, NP-mediated delivery of siRNA showed significant abrogation in lung fibrosis and improved pulmonary activities²⁷³ (Fig. 8B). The combined pathogenic effect conferred by viruses and inflammatory cytokines is known to have highly deleterious health consequences. However, targeting both viruses and inflammatory cytokines has become challenging for the scientific community. However, recently, Rao *et al.* developed an engineered novel decoy NP with dual targeting capacity.²⁷⁴ Uniquely, the decoy NPs competed with host cells for virus absorption and also captured inflammatory cytokines such as IL-6 and granulocyte macrophage-colony stimulating factor (GM-CSF). Thus, the powerful nano-decoy therapeutics show excellent ability to effectively protect individuals from viral infections caused by pseudo-viruses and SARS-CoV-2 virus.²⁷⁴

4.7. Nanotherapeutics for targeting complement systems (CS)

The CS is a part of the innate immune system, formed by the assembly of more than 30 small proteins including plasma proteins and cell surface receptors. CS plays a significant role in maintaining homeostasis by critically regulating the balance between different immune components. The specific interplay between antigen receptors, antibodies, and protein components of the CS activates the CS in a molecular cascade mechanism. The activation of the CS results in the formation of C3 convertase, which generates potential anaphylatoxins to encounter the pathogens and mediates multiple immune responses^{275,276} (Fig. 9A). In most cases, the incorrect activation of the CS results in severe immune complications. The use of nanomedicine for targeting the CS is an emerging treatment modality to restore the normalcy of the CS. NETs serve as a sanctuary for inflammatory IL-17 cytokines; they have autoimmune implications in the pathogenesis of psoriasis.²⁷⁷ The release of IL-17 from NETs occurs *via* activation of the C5b-9 complement component. Zhang *et al.* demonstrated that topical administration of human mesenchymal stem cell-derived exosome NPs showed excellent ability to inhibit C5b-9 complement activation and prevent inflammatory IL-17 cyto-

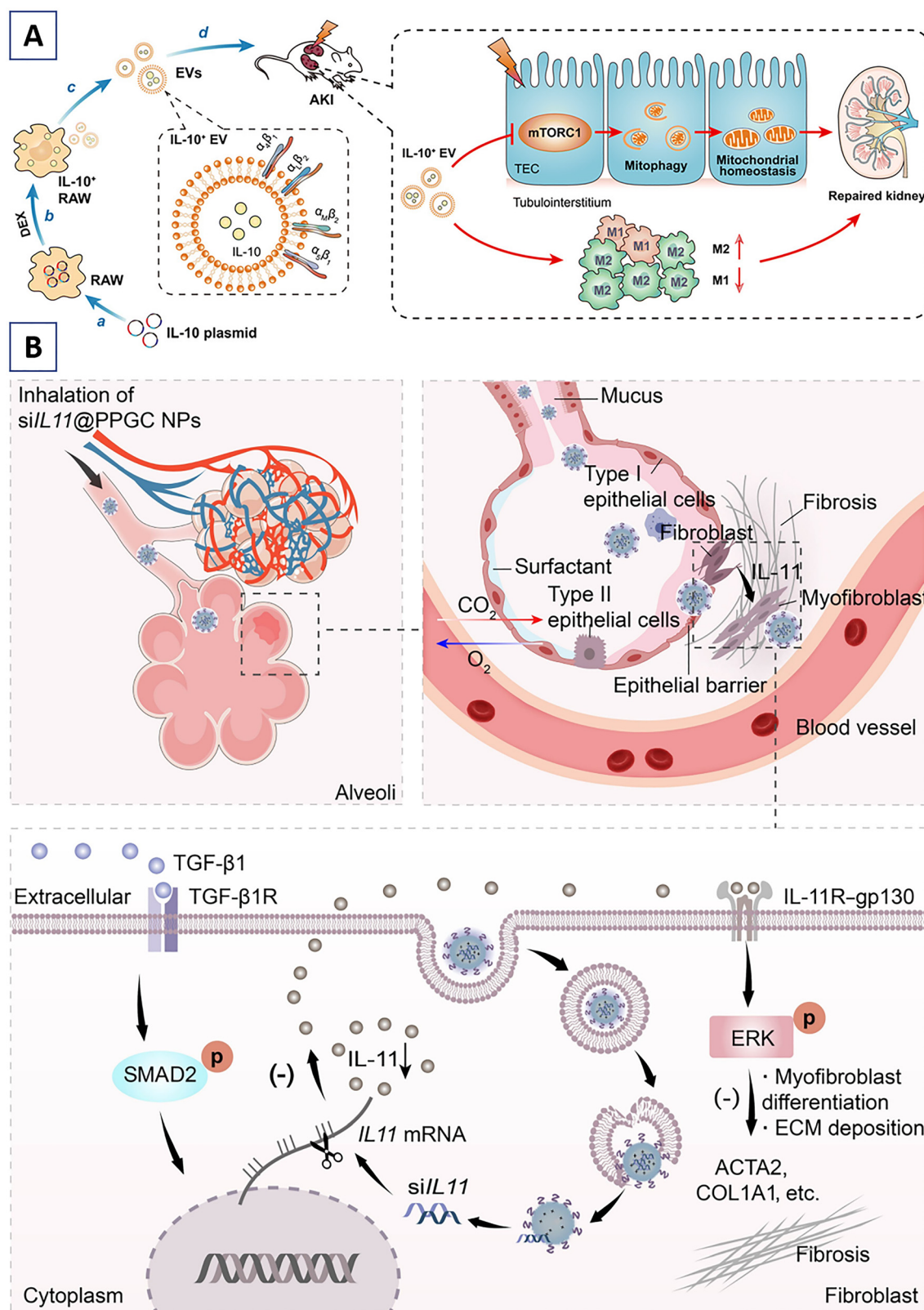


Fig. 8 (A) Schematic representation of the fabrication of interleukin-10 (IL-10)-loaded extracellular vesicles for the treatment of ischemic acute kidney injury. Reproduced with permission from ref. 267. Copyright 2020, American Association for the Advancement of Science. (B) Schematic illustration of the delivery of small interfering RNA encapsulated in self-assembled nanoparticles composed of poly(lactide-co-glycolide)-*b*-poly(ethylene glycol) (PLGA-PEG) diblock copolymer and cationic lipid-like molecules G0-C14 to mouse lung fibroblasts for the treatment of Idiopathic pulmonary fibrosis. Reproduced with permission from ref. 273. Copyright 2022, American Association for the Advancement of Science.

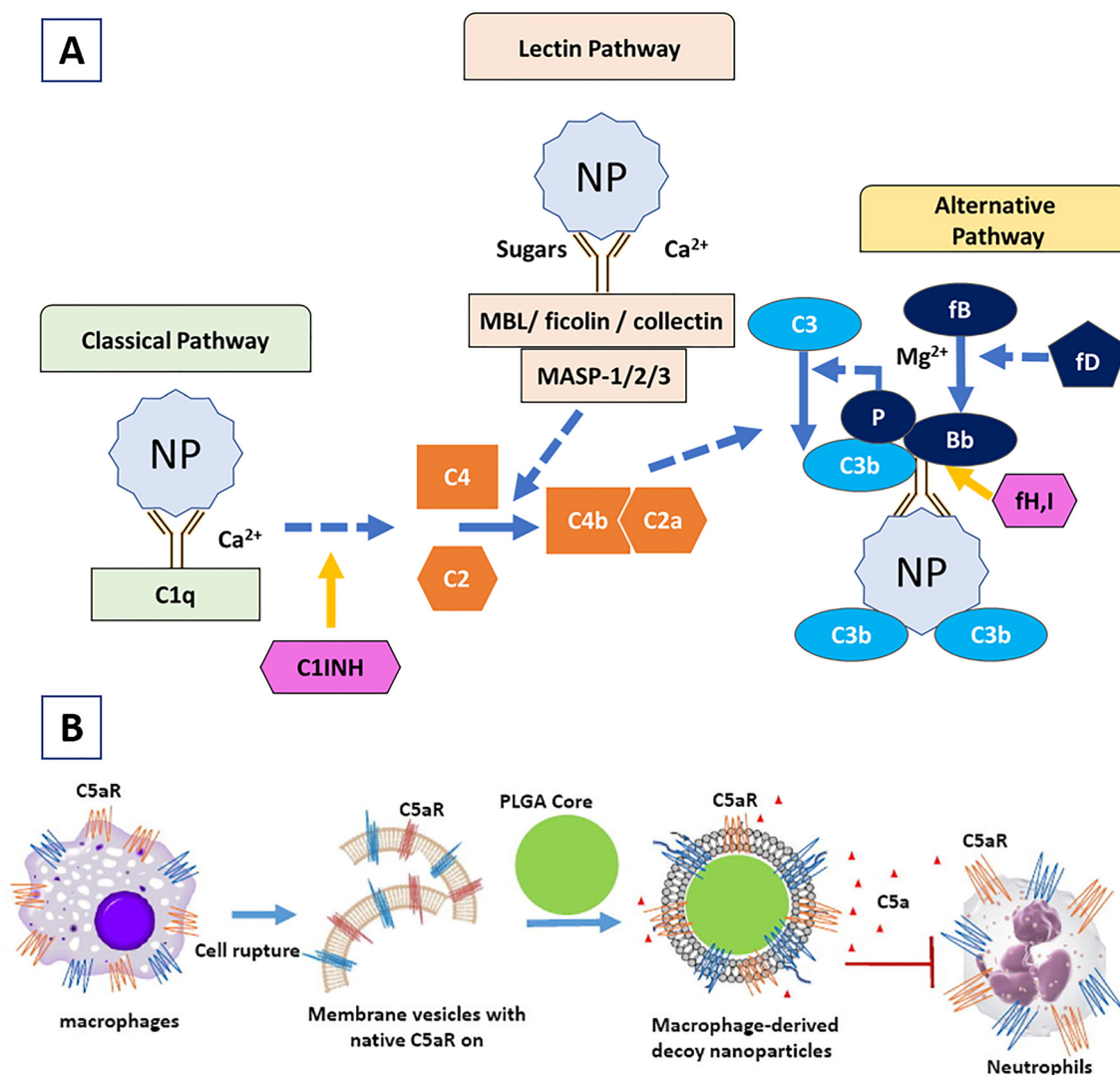


Fig. 9 (A) Schematic representation of the activation of the upstream part of the complement cascade through nanoparticle-bound immunoglobulins. The three initiating complement pathways include the classical, lectin and alternative pathway. The activation of the classical pathway occurs through C1q binding to the Fc portion of the surface-bound antibodies, the lectin pathway through the binding of the glycosylated regions of the antibodies to MBL/MASP-2 and the alternative pathway through the deposition of antibody-bound C3. The natural inhibitors of the complement cascade are shown in pink with yellow arrows. (B) Schematic illustration of the inhibition of neutrophil activation through C5aR-displaying decoy nanoparticles for improved mesenchymal stem cell-based therapies. Reproduced with permission from ref. 278. Copyright 2019, Elsevier. Abbreviations used: NP-nanoparticles; C1, C2, C3, C4-complement proteins; MBL-mannose-binding lectin; MASP-MBL-associated serine proteases; fB, fD, fI, fH: complement factors.

kinase release.²⁷⁷ In alleviation of NETosis-driven inflammatory psoriasis, the therapeutic exosome also reduced various side-effects that are observed in traditional drug use. During cell-based therapy, the administration of MSCs into the body provokes the CS to mobilize C5a, which in turn binds to the C5aR receptors present on neutrophils. Thus, escorting neutrophils to the proximity of MSCs by complement activation leads to potential damage to MSC therapeutics. Researchers found that the macrophage membrane exhibits a high expression of C5aR receptors that could be targeted in the development of nanotherapeutics by blunting the C5a-C5aR interaction and preventing the activation of neutrophils.²⁷⁸ Hence, macro-

phage membrane-coated engineered PLGA NPs were developed that effectively block the C5a-C5aR signalling, thereby preventing the C5a-mediated neutrophil targeting of therapeutic MSCs. Thus, the application of macrophage membrane-wrapped nanotherapeutics holds a promising approach for the regenerative and immunotherapeutic paradigm²⁷⁸ (Fig. 9B). The C3 component of the CS is also considered to be one of the key pathogenic mediators responsible for autoimmune myocarditis. In particular, C3-dependent liver-specific STAT3 signalling drives the pathogenesis of myocarditis and could be a potential target for therapeutic intervention. Avalle *et al.* demonstrated that liver-specific STAT3/C3-targeting siRNA NPs

Table 6 List of clinically approved/in clinical trials nanomedicines for immunotherapy

| Type of NPs | Components | Name | Manufacturer | Therapeutic payload | Mode of action and effect on immune system | Application | NCT number |
|-----------------|---|----------------------|--|-----------------------------------|--|--|-------------|
| Lipid NP | Ionizable lipid (proprietary), SM-102, cholesterol, DSPC, PEG2000 DMG | mRNA1273 | Sponsor: National Institute of Allergy and Infectious Diseases (NIAID) Collaborator: Moderna TX, Inc. | mRNA | The mRNA encoded spike (S) protein acts as an immunogen and generates antibodies, which provides immunity against the viral infection. | Covid-19 vaccine | NCT04470427 |
| Lipid NP | ALC-0315, ALC-0159, DSPC, cholesterol | BNT162b2/Tozinameran | Sponsor: BioNTech Collaborator: Pfizer | Nucleoside modified mRNA (modRNA) | The modRNA-encoded mutated full-length spike protein of SARS-CoV-2 drives immunity by serving as an immunogen. | Covid-19 vaccine | NCT04368728 |
| Liposome | Cholesterol, HSPC, PEG2000-DSPE | Doxil/Caelyx | Janssen Research & Development, LLC | Doxorubicin (DOX) | Primarily, DOX acts as a chemotherapeutic agent. In association with monoclonal antibody (Pembrolizumab), DOX can contribute to immunotherapy against sarcoma. | Anti-cancer | NCT00103506 |
| Liposome | Cholesterol, DSPC | DaunoXome | Sponsor: Nexstar Pharmaceuticals | Daunorubicin | Primarily, it acts as DNA topoisomerase-II poison. It induces cGAS-dependent innate immune response to suppress Hepatitis B virus production. | Anti-cancer | NCT00002093 |
| Protein NP | Human albumin protein | Abraxane | Sponsor: Celgene | Paclitaxel (PTX) | It inhibits dissociation of microtubules, assembles tubulin into microtubules, thus, arresting cancer cell growth. PTX also possesses tumor immunotherapy by regulating the activities of T cells, DCs, NK cells, Tregs and Macrophages. | Anti-cancer | NCT02027428 |
| Liposome | Soybean oil, egg lecithin, glycerol | Diprivan® | Sponsor: R-Pharm Collaborator: Synergy Research, Inc. | Propofol | Propofol mainly acts as GABA-receptor agonist. It also augments NK cell functions, inhibits PGE2 and COX-2 in postoperative immune protection. | Anaesthetic | NCT03669484 |
| Liposome | Cholesterol, HSPC, DSPG | AmBisome | Sponsor: Drugs for Neglected Diseases Collaborator: Shaheed Suhrawardy Medical College and Hospital International centre for Diarrhoeal Disease Research, Bangladesh Sponsor: QLT Inc. | Amphotericin B (Amp-B) | Amp-B binds to Ergosterol of the fungal cell membrane and creates pores and cells die eventually. Amp-B can modulate the immune system but also possess toxicity. | Antifungal, Antiparasitic | NCT01122771 |
| Liposome | DMPC, EPG | Visudyne | | Verteporfin | Verteporfin exerts its activity <i>via</i> light-induced PDT. Verteporfin inhibits PD-L1 and exerts antitumor efficacy. | In treatment of eye diseases caused by age-related macular degeneration (AMD) and other pathological conditions. | NCT00121407 |
| (Non-PEGylated) | DOPS, POPC | MEPACT | Sponsor: UNICANCER | Mifamurtide | Increased production of cytokines (TNF- α , IL-1, IL-6, IL-8, IL-12, and others) for cancer immunotherapy | For the treatment of osteosarcoma | NCT03643133 |

Table 6 (Contd.)

| Type of NPs | Components | Name | Manufacturer | Therapeutic payload | Mode of action and effect on immune system | Application | NCT number |
|---------------------------|------------------------------|------------------|---|---------------------|---|--|-------------|
| Liposome (Non-PEGylated) | EPC, cholesterol | Myocet | Sponsor: Sopherion Therapeutics | Doxorubicin | DNA intercalating agent | For the treatment of breast cancer | NCT00294996 |
| Hafnium oxide NP | Hafnium oxide | NBTXR3 | Sponsor: Nanobiotix | | Radiation-induced cancer cell killing and immune stimulation-based immunogenic cell death (ICD) | Radiotherapy for solid tumor | NCT04892173 |
| Fe-dextran colloid | Iron(III)-hydroxide, dextran | CosmoFer®/INFeD® | Sponsor: Pharmacosmos A/S | Fe | Overcomes iron deficiency | In the treatment of congestive heart failure | NCT00537186 |
| Fe-sucrose colloid | Iron, sucrose | Venofer | Sponsor: American Regent, Inc. | Fe | Replacement of iron | In the treatment of anemia with chronic kidney disease | NCT00236938 |
| Fe-gluconate colloid | Iron, gluconate | Ferrlecit® | Sponsor: Watson Pharmaceuticals | Fe | Replacement of iron | In the treatment of anemia with chronic kidney disease | NCT00224003 |
| Fe-carboxymaltose colloid | Iron, carboxymaltose | Ferinject® | Sponsor: Vifor Pharma Collaborator: Syneos Health ClimStar, LLC | Fe | Replacement of iron | Chronic heart failure and iron deficiency | NCT00520780 |

Abbreviation: SM-102, sphingomyelin-102; DSPC, distearoylphosphatidylcholine; PEG2000-DMG, polyethylene glycol-2000-dimethylol glycerol; HSPC, hydrogenated soy phosphatidylcholine; PEG2000-DSPE, polyethylene glycol-2000-distearoyl-*sn*-phosphoethanolamine; SM, sphingomyelin; MPEG2000-DSPE, methoxy polyethylene glycol 2000-distearoyl-*sn*-glycero-phosphoethanolamine; DSPG, distearoylphosphatidylglycerol; DMPC, dimyristoyl phosphatidylcholine; EPG, esterified propoxylated glycerol; EPC, egg phosphatidylcholine; DOPS, dioleoylphosphatidylcholine; POPC, palmitoyloleoylphosphatidylcholine.

interfered with STAT3 signalling and complement production, that provided protection against autoimmune cardiomyopathies.²⁷⁹ Autoreactive complement activities have also been reported in paroxysmal nocturnal hemoglobinuria (PNH) where the body's C5 complement proteins conduct attacks and lyse the erythrocytes.²⁸⁰ Currently, mAbs are being clinically used to counteract C5-complement protein in the management of PNH. In parallel, researchers have been trying to develop novel nano-vaccines for eliciting strong anti-C5 antibody responses. In this regard, recombinant VLPs are novel choices of nanomaterials for the design and development of complement-inhibiting nano-vaccine candidates. Zhang *et al.* have developed bacteriophage Q β plasmid-derived VLPs, surface engineered with C5 peptide epitopes to trigger anti-C5 auto-antibody responses.²⁸¹ The administration of VLP-C5 nano-vaccine efficiently produced anti-C5 autoantibodies that blocked the haemolytic C5 complement protein and could be an alternative to molecular medicines in the therapy of PNH.

Overall, several treatment strategies discussed have advanced to clinical trials, in some cases, achieved approval for immunotherapeutic applications, as listed in Table 6.

5. Challenges and future perspectives

NPs and immune cells form a unique physico-chemical environment that can be impacted by several factors of NPs, including source, variable catalysis residues, storage conditions, variability in synthesis, and endotoxin contamination.²⁸² Successful translation necessitates a meticulously streamlined synthesis protocol, standardized experimental tools and *in vitro* assays, and rigorous validation, including nanosafety assessment of endotoxins. Furthermore, NPs fabricated from specific materials, including metals, polymers, and ceramics, may show different levels of immunotoxicity that raise potential concerns over their usage. One effective strategy to mitigate material-mediated toxicity involves the utilization of inert materials that can serve as a protective coating, suppressing immune toxicity by preventing the formation of a protein corona. In addition, instead of targeting a single antigenic receptor, dual or more receptor targeting could be one of the advanced strategies for precise therapeutic intervention. NPs decorated with multiple ligands against multi-antigenic receptors can significantly reduce the associated off-target nano-immune toxicity concerns. The encapsulation of multiple therapeutic agents within a single nanocarrier and the subsequent co-delivery strategy represents a synergistic approach for achieving higher therapeutic efficacy and effectively overcoming multidrug resistance. Several practical challenges limit the clinical availability of these nanoparticle-based technologies in the market. These include scalability, reproducibility, prolonged regulatory approval processes, and patient heterogeneity.^{283,284} Several clinical data indicate that developed nanomedicines may only be effective in certain subpopulations of patients. Therefore, effective clinical implemen-

tation of these therapeutics depends on exploring strategies for patient stratification and treatment identification for trials. The selection of patient-specific immunological biomarkers can be a crucial step in advancing biomarker-guided nano-immunotherapeutic interventions. Moreover, the integration of existing knowledge of targeted regulation and stimuli-responsive release with advanced scientific theories from different fields, such as artificial intelligence, big data analysis, bioinformatics, and 3D printing, presents new opportunities for further advancement in this field. This integration holds significant potential to provide multi-immunoreceptor-targeted, micro-environment-responsive controlled delivery of drugs with minimal immunotoxicity hazards.

6. Conclusion

The increasing prevalence of life-threatening immune diseases necessitates the development of novel therapeutic platforms. The frequency and intricacy of such diseases have raised major concerns about the efficacy of traditional therapeutic regimens. In many cases, conventional therapeutic strategies have failed to deal with disease pathology for multiple reasons, such as non-specific disease targeting, low therapeutic index, rapid clearance of therapeutics, difficulty in bypassing several physiological barriers, and immune toxicities. Therapeutic NPs display better payload flexibility that helps to overcome the therapeutic limitations of traditional medications. The specific immune receptor-targeted drug-delivery strategy provides breakthroughs in target-specific modulation of pathogenic T cells, B cells, macrophages, and DCs. The combinations of engineered nanostructured vehicles and therapeutic payloads have produced several nanomedicines that can easily target these specific immune components. While some limitations exist for their clinical applications, the therapeutic efficiency of immunomodulatory NPs can be enhanced through further research and design of safer nanocarriers. The advancements in the field necessitate multidisciplinary collaboration among researchers, scientists, healthcare professionals, engineers, and regulatory experts to adapt immunomodulatory NPs to meet market demands. Such collaboration can transform health care by improving immune targeting precision and facilitating the successful clinical translation of immunomodulatory nanomedicines.

Conflicts of interest

There are no conflicts of interest to declare.

Acknowledgements

B. B. M. acknowledges the funding support from the Department of Science and Technology (DST) and the Department of Biotechnology (DBT), Government of India. R. K., B. B., S. D. and V. J. acknowledge the Ministry of

Education (MoE), Government of India, for providing fellowship. C. J. acknowledges the Council of Scientific & Industrial Research (CSIR), Government of India for fellowship.

References

- 1 K. C. Ma, E. J. Schenck, M. A. Pabon and A. M. K. Choi, *Am. J. Respir. Crit. Care Med.*, 2018, **197**, 300–309.
- 2 V. Afshar-Kharghan, *J. Clin. Invest.*, 2017, **127**, 780–789.
- 3 C. Bantz, O. Koshkina, T. Lang, H. J. Galla, C. J. Kirkpatrick, R. H. Stauber and M. Maskos, *Beilstein J. Nanotechnol.*, 2014, **5**, 1774–1786.
- 4 C. Zhang, G. Shi, J. Zhang, H. Song, J. Niu, S. Shi, P. Huang, Y. Wang, W. Wang, C. Li and D. Kong, *J. Controlled Release*, 2017, **256**, 170–181.
- 5 J. Conde, C. Bao, Y. Tan, D. Cui, E. R. Edelman, H. S. Azevedo, H. J. Byrne, N. Artzi and F. Tian, *Adv. Funct. Mater.*, 2015, **25**, 4183–4194.
- 6 P. Korangath, J. D. Barnett, A. Sharma, E. T. Henderson, J. Stewart, S. H. Yu, S. K. Kandala, C. T. Yang, J. S. Caserto, M. Hedayati, T. D. Armstrong, E. Jaffee, C. Gruettner, X. C. Zhou, W. Fu, C. Hu, S. Sukumar, B. W. Simons and R. Ivkov, *Sci. Adv.*, 2020, **6**, eaay1601.
- 7 H. S. Choi, W. Liu, P. Misra, E. Tanaka, J. P. Zimmer, B. I. Ipe, M. G. Bawendi and J. V. Frangioni, *Nat. Biotechnol.*, 2007, **25**, 1165–1170.
- 8 S. V. Vinogradov, T. K. Bronich and A. V. Kabanov, *Adv. Drug Delivery Rev.*, 2002, **54**, 135–147.
- 9 A. L. B. de Barros, A. Tsourkas, B. Saboury, V. N. Cardoso and A. Alavi, *EJNMMI Res.*, 2012, **2**, 1–15.
- 10 S. A. Kulkarni and S. S. Feng, *Pharm. Res.*, 2013, **30**, 2512–2522.
- 11 A. H. Faraji and P. Wipf, *Bioorg. Med. Chem.*, 2009, **17**, 2950–2962.
- 12 P. Foroozandeh and A. A. Aziz, *Nanoscale Res. Lett.*, 2018, **13**, 1–12.
- 13 R. A. Petros and J. M. Desimone, *Nat. Rev. Drug Discovery*, 2010, **9**, 615–627.
- 14 M. J. Ernsting, M. Murakami, A. Roy and S. D. Li, *J. Controlled Release*, 2013, **172**, 782–794.
- 15 W. Wu, L. Luo, Y. Wang, Q. Wu, H. B. Dai, J. S. Li, C. Durkan, N. Wang and G. X. Wang, *Theranostics*, 2018, **8**, 3038–3058.
- 16 S. Kang, S. Ahn, J. Lee, J. Y. Kim, M. Choi, V. Gujrati, H. Kim, J. Kim, E. C. Shin and S. Jon, *J. Controlled Release*, 2017, **256**, 56–67.
- 17 S. Kumar, A. C. Anselmo, A. Banerjee, M. Zakrewsky and S. Mitragotri, *J. Controlled Release*, 2015, **220**, 141–148.
- 18 M. F. Bachmann and G. T. Jennings, *Nat. Rev. Immunol.*, 2010, **10**, 787–796.
- 19 C. Graf, D. Nordmeyer, C. Sengstock, S. Ahlberg, J. Diendorf, J. Raabe, M. Epple, M. Köller, J. Lademann, A. Vogt, F. Rancan and E. Rühl, *Langmuir*, 2018, **34**, 1506–1519.

- 20 R. Toy, P. M. Peiris, K. B. Ghaghada and E. Karathanasis, *Nanomedicine*, 2014, **9**, 121–134.
- 21 X. Xie, J. Liao, X. Shao, Q. Li and Y. Lin, *Sci. Rep.*, 2017, **7**, 1–9.
- 22 T. Tazaki, K. Tabata, A. Ainai, Y. Ohara, S. Kobayashi, T. Ninomiya, Y. Orba, H. Mitomo, T. Nakano, H. Hasegawa, K. Ijiro, H. Sawa, T. Suzuki and K. Niikura, *RSC Adv.*, 2018, **8**, 16527–16536.
- 23 S. Barua, J. W. Yoo, P. Kolhar, A. Wakankar, Y. R. Gokarn and S. Mitragotri, *Proc. Natl. Acad. Sci. U. S. A.*, 2013, **110**, 3270–3275.
- 24 B. D. Chithrani, A. A. Ghazani and W. C. W. Chan, *Nano Lett.*, 2006, **6**, 662–668.
- 25 D. L. J. Thorek and A. Tsourkas, *Biomaterials*, 2008, **29**, 3583–3590.
- 26 I. Slowing, B. G. Trewyn and V. S. Y. Lin, *J. Am. Chem. Soc.*, 2006, **128**, 14792–14793.
- 27 S. Behzadi, V. Serpooshan, W. Tao, M. A. Hamaly, M. Y. Alkawareek, E. C. Dreaden, D. Brown, A. M. Alkilany, O. C. Farokhzad and M. Mahmoudi, *Chem. Soc. Rev.*, 2017, **46**, 4218–4244.
- 28 J. Dausend, A. Musyanovych, M. Dass, P. Walther, H. Schrezenmeier, K. Landfester and V. Mailänder, *Macromol. Biosci.*, 2008, **8**, 1135–1143.
- 29 L. Liu, F. Cao, X. Liu, H. Wang, C. Zhang, H. Sun, C. Wang, X. Leng, C. Song, D. Kong and G. Ma, *ACS Appl. Mater. Interfaces*, 2016, **8**, 11969–11979.
- 30 A. K. Dey, A. Nougarede, F. Clément, C. Fournier, E. Jouvin-Marche, M. Escudé, D. Jary, F. P. Navarro and P. N. Marche, *Front. Immunol.*, 2021, **12**, 722411.
- 31 S. Srijampa, S. Buddhisa, S. Ngerpimai, C. Leelayuwat, S. Prongvitaya, A. Chompoosor and P. Tippayawat, *Bioconjugate Chem.*, 2020, **31**, 1133–1143.
- 32 Y. Pan, Y. Qi, N. Shao, A. C. Tadde and Y. Huang, *Biomacromolecules*, 2019, **20**, 3575–3583.
- 33 J. W. Shreffler, J. E. Pullan, K. M. Dailey, S. Mallik and A. E. Brooks, *Int. J. Mol. Sci.*, 2019, **20**, 6056.
- 34 V. Patsula, D. Horák, J. Kučka, H. Macková, V. Lobaz, P. Francová, V. Herynek, T. Heizer, P. Páral and L. Šefc, *Sci. Rep.*, 2019, **9**, 1–12.
- 35 Q. Feng, Y. Liu, J. Huang, K. Chen, J. Huang and K. Xiao, *Sci. Rep.*, 2018, **8**, 1–13.
- 36 V. Juang, C.-H. Chang, C.-S. Wang, H.-E. Wang, Y.-L. Lo, V. Juang, C.-H. Chang, C.-S. Wang, Y.-L. Lo and H.-E. Wang, *Small*, 2019, **15**, 1903296.
- 37 Y. Qie, H. Yuan, C. A. Von Roemeling, Y. Chen, X. Liu, K. D. Shih, J. A. Knight, H. W. Tun, R. E. Wharen, W. Jiang and B. Y. S. Kim, *Sci. Rep.*, 2016, **6**, 1–11.
- 38 J. S. Suk, Q. Xu, N. Kim, J. Hanes and L. M. Ensign, *Adv. Drug Delivery Rev.*, 2016, **99**, 28–51.
- 39 S. O. Stead, S. Kireta, S. J. P. McInnes, F. D. Kette, K. N. Sivanathan, J. Kim, E. J. Cueto-Diaz, F. Cunin, J. O. Durand, C. J. Drogemuller, R. P. Carroll, N. H. Voelcker and P. T. Coates, *ACS Nano*, 2018, **12**, 6637–6647.
- 40 J. Xiang, L. Xu, H. Gong, W. Zhu, C. Wang, J. Xu, L. Feng, L. Cheng, R. Peng and Z. Liu, *ACS Nano*, 2015, **9**, 6401–6411.
- 41 V. S. S. A. Ayyadevara, A. Ahmadi and K. H. Roh, *Bioconjugate Chem.*, 2021, **32**, 1675–1687.
- 42 L. Jing, S. Shao, Y. Wang, Y. Yang, X. Yue and Z. Dai, *Theranostics*, 2016, **6**, 40–53.
- 43 A. Gangrade and B. B. Mandal, *ACS Biomater. Sci. Eng.*, 2019, **5**, 2365–2381.
- 44 J. Luo, X. Meng, J. Su, H. Ma, W. Wang, L. Fang, H. Zheng, Y. Qin and T. Chen, *J. Agric. Food Chem.*, 2018, **66**, 9219–9230.
- 45 J. A. Katakowski, G. Mukherjee, S. E. Wilner, K. E. Maier, M. T. Harrison, T. P. Di Lorenzo, M. Levy and D. Palliser, *Mol. Ther.*, 2016, **24**, 146–155.
- 46 G. N. Shi, C. N. Zhang, R. Xu, J. F. Niu, H. J. Song, X. Y. Zhang, W. W. Wang, Y. M. Wang, C. Li, X. Q. Wei and D. L. Kong, *Biomaterials*, 2017, **113**, 191–202.
- 47 S. K. Gulla, B. R. Rao, G. Moku, S. Jinka, N. V. Nimmu, S. Khalid, C. R. Patra and A. Chaudhuri, *Biomater. Sci.*, 2019, **7**, 773–788.
- 48 R. R. Meka, S. Mukherjee, C. R. Patra and A. Chaudhuri, *Nanoscale*, 2019, **11**, 7931–7943.
- 49 N. Climent, I. García, M. Marradi, F. Chiodo, L. Miralles, M. J. Maleno, J. M. Gatell, F. García, S. Penadés and M. Plana, *Nanomedicine*, 2018, **14**, 339–351.
- 50 J. Zhang and C. A. Peng, *Sci. Rep.*, 2020, **10**, 1–13.
- 51 L. Pang, Y. Pei, G. Uzunalli, H. Hyun, L. T. Lyle and Y. Yeo, *Pharm. Res.*, 2019, **36**, 65.
- 52 A. Badkas, E. Frank, Z. Zhou, M. Jafari, H. Chandra, V. Sriram, J. Y. Lee and J. S. Yadav, *Colloids Surf., B*, 2018, **162**, 271–278.
- 53 D. Schmid, C. G. Park, C. A. Hartl, N. Subedi, A. N. Cartwright, R. B. Puerto, Y. Zheng, J. Maiarana, G. J. Freeman, K. W. Wucherpfennig, D. J. Irvine and M. S. Goldberg, *Nat. Commun.*, 2017, **8**, 1–12.
- 54 K. W. Huang, F. F. Hsu, J. T. Qiu, G. J. Chern, Y. A. Lee, C. C. Chang, Y. T. Huang, Y. C. Sung, C. C. Chiang, R. L. Huang, C. C. Lin, T. K. Dinh, H. C. Huang, Y. C. Shih, D. Alson, C. Y. Lin, Y. C. Lin, P. C. Chang, S. Y. Lin and Y. Chen, *Sci. Adv.*, 2020, **6**, eaax5032.
- 55 Y. Zhang, N. Li, H. Suh and D. J. Irvine, *Nat. Commun.*, 2018, **9**, 1–15.
- 56 M. Khodoun, A. A. Chimote, F. Z. Ilyas, H. J. Duncan, H. Moncrieffe, K. S. Kant and L. Conforti, *Sci. Adv.*, 2021, **6**, eabd1471.
- 57 F. Zhou, J. Mei, S. Yang, X. Han, H. Li, Z. Yu, H. Qiao and T. Tang, *ACS Appl. Mater. Interfaces*, 2020, **12**, 2009–2022.
- 58 S. N. Khan, P. Han, R. Chaudhury, S. Bickerton, J. S. Lee, B. Calderon, A. Pellowe, A. Gonzalez and T. Fahmy, *Mol. Pharm.*, 2021, **18**, 850–861.
- 59 I. Khan, K. Saeed and I. Khan, *Arabian J. Chem.*, 2019, **12**, 908–931.
- 60 S. Stanley, *Curr. Opin. Biotechnol.*, 2014, **28**, 69–74.
- 61 M. J. Mitchell, M. M. Billingsley, R. M. Haley, M. E. Wechsler, N. A. Peppas and R. Langer, *Nat. Rev. Drug Discovery*, 2021, **20**, 101–124.
- 62 V. Taghipour-Sabzevar, T. Sharifi and M. M. Moghaddam, *Ther. Delivery*, 2019, **10**, 527–550.

- 63 S. Kumar, N. Dilbaghi, R. Saharan and G. Bhanjana, *BioNanoScience*, 2012, **4**, 227–250.
- 64 I. R. Khalil, A. T. H. Burns, I. Radecka, M. Kowalczyk, T. Khalaf, G. Adamus, B. Johnston and M. P. Khechara, *Int. J. Mol. Sci.*, 2017, **18**, 313.
- 65 A. P. Sherje, M. Jadhav, B. R. Dravyakar and D. Kadam, *Int. J. Pharm.*, 2018, **548**, 707–720.
- 66 C. C. Lee, J. A. MacKay, J. M. J. Fréchet and F. C. Szoka, *Nat. Biotechnol.*, 2005, **23**, 1517–1526.
- 67 T. Anajafi and S. Mallik, *Ther. Deliv.*, 2015, **6**, 521–534.
- 68 N. Majumder, N. G. Das and S. K. Das, *Ther. Deliv.*, 2020, **11**, 613–635.
- 69 A. Varela-Moreira, Y. Shi, M. H. A. M. Fens, T. Lammers, W. E. Hennink and R. M. Schiffelers, *Mater. Chem. Front.*, 2017, **1**, 1485–1501.
- 70 Y. Lee, K. Sugihara, M. G. Gilliland, S. Jon, N. Kamada and J. J. Moon, *Nat. Mater.*, 2020, **19**, 118–126.
- 71 Y. Wang, L. Li, W. Zhao, Y. Dou, H. An, H. Tao, X. Xu, Y. Jia, S. Lu, J. Zhang and H. Hu, *ACS Nano*, 2018, **12**, 8943–8960.
- 72 S. Dehghan, M. T. Kheiri, K. Abnous, M. Eskandari and M. Tafaghodi, *Microb. Pathog.*, 2018, **115**, 74–85.
- 73 S. Jain, T. H. Tran and M. Amiji, *Biomaterials*, 2015, **61**, 162–177.
- 74 H. D. Han, Y. Byeon, J. H. Jang, H. N. Jeon, G. H. Kim, M. G. Kim, C. G. Pack, T. H. Kang, I. D. Jung, Y. T. Lim, Y. J. Lee, J. W. Lee, B. C. Shin, H. J. Ahn, A. K. Sood and Y. M. Park, *Sci. Rep.*, 2016, **6**, 1–13.
- 75 Y. Wu, J. Gu, S. Zhang, Y. Gu, J. Ma, Y. Wang, L. W. Zhang and Y. Wang, *Anal. Chem.*, 2021, **93**, 6414–6420.
- 76 Q. Yu, X. Tang, W. Zhao, Y. Qiu, J. He, D. Wan, J. Li, X. Wang, X. He, Y. Liu, M. Li, Z. Zhang and Q. He, *Acta Biomater.*, 2021, **133**, 244–256.
- 77 J. Lyu, L. Wang, X. Bai, X. Du, J. Wei, J. Wang, Y. Lin, Z. Chen, Z. Liu, J. Wu and Z. Zhong, *ACS Appl. Mater. Interfaces*, 2021, **13**, 266–276.
- 78 Z. Xie, Y. Su, G. B. Kim, E. Selvi, C. Ma, V. Aragon-Sanabria, J. T. Hsieh, C. Dong and J. Yang, *Small*, 2017, **13**, 1603121.
- 79 M. Tukulula, R. Hayeshi, P. Fonteh, D. Meyer, A. Ndamase, M. T. Madziva, V. Khumalo, P. Lubuschagne, B. Naicker, H. Swai and A. Dube, *Pharm. Res.*, 2015, **32**, 2713–2726.
- 80 S. Vemireddy, P. P. Madhurantakam, M. N. Talati and H. M. S. Kumar, *ACS Appl. Bio Mater.*, 2019, **2**, 4837–4846.
- 81 Y. Xing, Z. Xu, T. Liu, L. Shi, D. Kohane and S. Guo, *Angew. Chem.*, 2020, **132**, 7302–7306.
- 82 A. Thukral, K. Ross, C. Hansen, Y. Phanse, B. Narasimhan, H. Steinberg and A. M. Talaat, *npj Vaccines*, 2020, **5**, 1–10.
- 83 F. Walter, E. Winter, S. Rahn, J. Heidland, S. Meier, A. M. Struzek, M. Lettau, L. M. Philipp, S. Beckinger, L. Otto, J. L. Möller, O. Helm, D. Wesch, R. Scherließ and S. Sebens, *PLoS One*, 2020, **15**, e0239369.
- 84 T. J. Beldman, M. L. Senders, A. Alaarg, C. Pérez-Medina, J. Tang, Y. Zhao, F. Fay, J. Deichmüller, B. Born, E. Desclos, N. N. Van Der Wel, R. A. Hoebe, F. Kohen, E. Kartvelishvily, M. Neeman, T. Reiner, C. Calcagno, Z. A. Fayad, M. P. J. De Winther, E. Lutgens, W. J. M. Mulder and E. Kluza, *ACS Nano*, 2017, **11**, 5785–5799.
- 85 S. F. Lin, P. L. Jiang, J. S. Tsai, Y. Y. Huang, S. Y. Lin, J. H. Lin and D. Z. Liu, *J. Biomed. Mater. Res., Part B*, 2019, **107**, 1228–1237.
- 86 K. Ziorek, C. Bourquin, J. Battiany, G. Winter, S. Endres, G. Hartmann and C. Coester, *Pharm. Res.*, 2008, **25**, 551–562.
- 87 C. Bourquin, *Chimia*, 2019, **73**, 69–72.
- 88 J. D. Totten, T. Wongpinyochit, J. Carrola, I. F. Duarte and F. P. Seib, *ACS Appl. Mater. Interfaces*, 2019, **11**, 14515–14525.
- 89 P. Hassanzadeh, E. Arbabi and F. Rostami, *Life Sci.*, 2021, **281**, 119772.
- 90 C. Ji, J. Si, Y. Xu, W. Zhang, Y. Yang, X. He, H. Xu, X. Mou, H. Ren and H. Guo, *Theranostics*, 2021, **11**, 8587.
- 91 J. V. Gregory, P. Kadiyala, R. Doherty, M. Cadena, S. Habel, E. Ruoslahti, P. R. Lowenstein, M. G. Castro and J. Lahann, *Nat. Commun.*, 2020, **11**, 1–15.
- 92 D. Sun, Y. Zou, L. Song, S. Han, H. Yang, D. Chu, Y. Dai, J. Ma, C. M. O'Driscoll, Z. Yu and J. Guo, *Acta Pharm. Sin. B*, 2022, **12**, 378–393.
- 93 C. Thomas, A. Rawat, L. Hope-Weeks and F. Ahsan, *Mol. Pharm.*, 2011, **8**, 405–415.
- 94 J. Meena, R. Kumar, M. Singh, A. Ahmed and A. K. Panda, *Eur. J. Pharm. Biopharm.*, 2020, **152**, 270–281.
- 95 A. L. Coolen, C. Lacroix, P. Mercier-Gouy, E. Delaune, C. Monge, J. Y. Exposito and B. Verrier, *Biomaterials*, 2019, **195**, 23–37.
- 96 B. Boltnarova, J. Kubackova, J. Skoda, A. Stefela, M. Smekalova, P. Svacinova, I. Pavkova, M. Dittrich, D. Scherman, J. Zbytovska, P. Pavek and O. Holas, *Nanomaterials*, 2021, **11**, 749.
- 97 A. Kaur, J. Rathee, R. Kanwar, D. Kaushik, D. B. Salunke and S. K. Mehta, *Colloids Surf., A*, 2022, **647**, 129084.
- 98 M. Igartua, R. M. Hernández, A. Esquisabel, A. R. Gascón, M. B. Calvo and J. L. Pedraz, *J. Controlled Release*, 1998, **56**, 63–73.
- 99 V. Bansal, M. Kumar, A. Bhardwaj, H. G. Brahmne and H. Singh, *Vaccine*, 2015, **33**, 5623–5632.
- 100 A. Ahmad, E. Fauzia, M. Kumar, R. K. Mishra, A. Kumar, M. A. Khan, S. S. Raza and R. Khan, *ACS Biomater. Sci. Eng.*, 2019, **5**, 683–695.
- 101 H. H. Salman, J. M. Irache and C. Gamazo, *Vaccine*, 2009, **27**, 4784–4790.
- 102 K. H. Huynh, X. H. Pham, J. Kim, S. H. Lee, H. Chang, W. Y. Rho and B. H. Jun, *Int. J. Mol. Sci.*, 2020, **21**, 1–29.
- 103 A. A. Yaqoob, H. Ahmad, T. Parveen, A. Ahmad, M. Oves, I. M. I. Ismail, H. A. Qari, K. Umar and M. N. M. Ibrahim, *Front. Chem.*, 2020, **8**, 528583.
- 104 K. V. Chakravarthy, A. C. Bonoiu, W. G. Davis, P. Ranjan, H. Ding, R. Hu, J. B. Bowzard, E. J. Bergey, J. M. Katz, P. R. Knight, S. Sambhara and P. N. Prasad, *Proc. Natl. Acad. Sci. U. S. A.*, 2010, **107**, 10172–10177.

- 105 S. S. Bawage, P. M. Tiwari, A. Singh, S. Dixit, S. R. Pillai, V. A. Dennis and S. R. Singh, *Nanomedicine*, 2016, **12**, 2299–2310.
- 106 S. Liang, M. Sun, Y. Lu, S. Shi, Y. Yang, Y. Lin, C. Feng, J. Liu and C. Dong, *J. Mater. Chem. B*, 2020, **8**, 8368–8382.
- 107 F. Xia, W. Hou, Y. Liu, W. Wang, Y. Han, M. Yang, X. Zhi, C. Li, D. Qi, T. Li, J. M. de la Fuente, C. Zhang, J. Song and D. Cui, *Biomaterials*, 2018, **170**, 1–11.
- 108 P. Orłowski, A. Kowalczyk, E. Tomaszewska, K. Ranaszek-Soliwoda, A. Węgrzyn, J. Grzesiak, G. Celichowski, J. Grobelny, K. Eriksson and M. Krzyzowska, *Viruses*, 2018, **10**, 524.
- 109 Y. Wen, X. Chen, X. Zhu, Y. Gong, G. Yuan, X. Qin and J. Liu, *ACS Appl. Mater. Interfaces*, 2019, **11**, 43393–43408.
- 110 S. Thomas, B. S. P. Harshita, P. Mishra and S. Talegaonkar, *Curr. Pharm. Des.*, 2015, **21**, 6165–6188.
- 111 D. Singh, P. Dubey, M. Pradhan and M. R. Singh, *Expert Opin. Drug Delivery*, 2013, **10**, 241–259.
- 112 Y. Zhao, X. Zhao, Y. Cheng, X. Guo and W. Yuan, *Mol. Pharm.*, 2018, **15**, 1791–1799.
- 113 J. Hwang, E. Lee, J. Kim, Y. Seo, K. H. Lee, J. W. Hong, A. A. Gilad, H. Park and J. Choi, *Colloids Surf., B*, 2016, **142**, 290–296.
- 114 G. Garcia, M. H. Kim, V. A. Morikis and S. I. Simon, *Front. Immunol.*, 2020, **11**, 571489.
- 115 Q. Jiang, K. Wang, X. Zhang, B. Ouyang, H. Liu, Z. Pang and W. Yang, *Small*, 2020, **16**, 2001704.
- 116 M. Mohajeri, B. Behnam and A. Sahebkar, *J. Cell Physiol.*, 2018, **234**, 298–319.
- 117 P. McKernan, N. A. Virani, G. N. F. Faria, C. G. Karch, R. P. Silvy, D. E. Resasco, L. F. Thompson and R. G. Harrison, *Nanoscale Res. Lett.*, 2021, **16**, 1–9.
- 118 H. A. F. M. Hassan, L. Smyth, J. T. W. Wang, P. M. Costa, K. Ratnasothy, S. S. Diebold, G. Lombardi and K. T. Al-Jamal, *Biomaterials*, 2016, **104**, 310–322.
- 119 M. R. M. Radzi, N. A. Johari, W. F. A. W. M. Zawawi, N. A. Zawawi, N. A. Latiff, N. A. N. N. Malek, A. A. Wahab, M. I. Salim and K. Jemon, *Biomater. Adv.*, 2022, **134**, 112586.
- 120 D. Alizadeh, E. E. White, T. C. Sanchez, S. Liu, L. Zhang, B. Badie and J. M. Berlin, *Bioconjugate Chem.*, 2018, **29**, 1659–1668.
- 121 P. C. B. D. Faria, L. I. D. Santos, J. P. Coelho, H. B. Ribeiro, M. A. Pimenta, L. O. Ladeira, D. A. Gomes, C. A. Furtado and R. T. Gazzinelli, *Nano Lett.*, 2014, **14**, 5458–5470.
- 122 L. Hou, D. Chen, R. Wang, R. Wang, H. Zhang, Z. Zhang, Z. Nie and S. Lu, *Angew. Chem.*, 2021, **133**, 6655–6666.
- 123 Q. Zhou, N. Gong, D. Zhang, J. Li, X. Han, J. Dou, J. Huang, K. Zhu, P. Liang, X. Liang and J. Yu, *ACS Nano*, 2021, **15**, 2920–2932.
- 124 D. H. Kim, J. Seo and K. Na, *Mol. Pharm.*, 2020, **17**, 2532–2545.
- 125 F. H. Horst, C. V. d. S. Rodrigues, P. H. P. R. Carvalho, A. M. Leite, R. B. Azevedo, B. A. D. Neto, J. R. Corrêa, M. P. Garcia, S. Alotaibi, M. Henini, S. B. Chaves and M. O. Rodrigues, *RSC Adv.*, 2021, **11**, 6346–6352.
- 126 X. Zhang, H. Li, C. Yi, G. Chen, Y. Li, Y. Zhou, G. Chen, Y. Li, Y. He and D. Yu, *Int. J. Nanomed.*, 2020, **15**, 9627.
- 127 K. Lategan, H. Alghadi, M. Bayati, M. F. de Cortalezzi and E. Pool, *Nanomaterials*, 2018, **8**, 125.
- 128 M. J. Feito, R. Diez-Orejas, M. Cicuéndez, L. Casarrubios, J. M. Rojo and M. T. Portolés, *Colloids Surf., B*, 2019, **176**, 96–105.
- 129 B. Zhou, J. Song, M. Wang, X. Wang, J. Wang, E. W. Howard, F. Zhou, J. Qu and W. R. Chen, *Nanoscale*, 2018, **10**, 21640–21647.
- 130 Y. Tao, Y. Zhang, E. Ju, H. Ren and J. Ren, *Nanoscale*, 2015, **7**, 12419–12426.
- 131 P. Pei, W. Shen, H. Zhou, Y. Sun, J. Zhong, T. Liu and K. Yang, *Nano Today*, 2021, **38**, 101144.
- 132 B. Chakraborty, R. Pal, M. Ali, L. M. Singh, D. S. Rahman, S. K. Ghosh and M. Sengupta, *Cell. Mol. Immunol.*, 2015, **13**, 191–205.
- 133 B. Vuković, Ž. Cvetić, K. Bendelja, R. Barbir, M. Milić, B. Dobrošević, V. Šerić and I. V. Vrček, *J. Biol. Inorg. Chem.*, 2021, **26**, 817–831.
- 134 Q. Mu, G. Lin, M. Jeon, H. Wang, F. C. Chang, R. A. Revia, J. Yu and M. Zhang, *Mater. Today*, 2021, **50**, 149–169.
- 135 S. Zanganeh, G. Hutter, R. Spitler, O. Lenkov, M. Mahmoudi, A. Shaw, J. S. Pajarinen, H. Nejadnik, S. Goodman, M. Moseley, L. M. Coussens and H. E. Daldrup-Link, *Nat. Nanotechnol.*, 2016, **11**, 986.
- 136 M. Cobaleda-Siles, M. Henriksen-Lacey, A. R. De Angulo, A. Bernecker, V. G. Vallejo, B. Szczupak, J. Llop, G. Pastor, S. Plaza-Garcia, M. Jauregui-Osoro, L. K. Meszaros and J. C. Mareque-Rivas, *Small*, 2014, **10**, 5054–5067.
- 137 R. Juneja, H. Vadarevu, J. Halman, M. Tarannum, L. Rackley, J. Dobbs, J. Marquez, M. Chandler, K. Afonin and J. L. Vivero-Escoto, *ACS Appl. Mater. Interfaces*, 2020, **12**, 38873–38886.
- 138 C. Xu, J. Nam, H. Hong, Y. Xu and J. J. Moon, *ACS Nano*, 2019, **13**, 12148–12161.
- 139 P. Zhao, L. Qiu, S. Zhou, L. Li, Z. Qian and H. Zhang, *Int. J. Nanomed.*, 2021, **16**, 2107.
- 140 A. Seth, H. G. Derami, P. Gupta, Z. Wang, P. Rathi, R. Gupta, T. Cao, J. J. Morrissey and S. Singamaneni, *ACS Appl. Mater. Interfaces*, 2020, **12**, 42499.
- 141 P. A. Bielecki, M. E. Lorkowski, W. M. Becicka, P. U. Atukorale, T. J. Moon, Y. Zhang, M. Wiese, G. Covarrubias, S. Ravichandran and E. Karathanasis, *Nanoscale Horiz.*, 2021, **6**, 156–167.
- 142 W. An, S. Defaus, D. Andreu and P. Rivera-Gil, *Front. Immunol.*, 2021, **12**, 684612.
- 143 S. Jafari, N. Maleki-Dizaji, J. Barar, M. Barzegar-Jalali, M. Rameshrad and K. Adibkia, *Eur. J. Pharm. Sci.*, 2016, **91**, 225–235.
- 144 A. Fulgione, N. Nocerino, M. Iannaccone, S. Roperto, F. Capuano, N. Roveri, M. Lelli, A. Crasto, A. Calogero, A. P. Pilloni and R. Capparelli, *PLoS One*, 2016, **11**, e0158646.

- 145 X. Wang, X. Li, A. Ito, Y. Watanabe and N. M. Tsuji, *Chem. Commun.*, 2016, **52**, 7078–7081.
- 146 Y. Hua, J. Wu, H. Wu, C. Su, X. Li, Q. Ao, Q. Zeng, X. Zhu and X. Zhang, *Acta Biomater.*, 2021, **135**, 650–662.
- 147 Y. Zhang, W. Sha, X. Zhang, M. Cheng, Q. Wu, W. Wang and Z. Yuan, *Biomater. Sci.*, 2019, **7**, 5027–5034.
- 148 F. Zhao, C. Wang, Q. Yang, S. Han, Q. Hu and Z. Fu, *Life Sci.*, 2018, **202**, 44–51.
- 149 B. C. Schanen, S. Das, C. M. Reilly, W. L. Warren, W. T. Self, S. Seal and D. R. Drake, *PLoS One*, 2013, **8**, e62816.
- 150 J. Wang, J. S. Lee, D. Kim and L. Zhu, *ACS Appl. Mater. Interfaces*, 2017, **9**, 39971–39984.
- 151 H. Moratin, P. Ickrath, A. Scherzad, T. J. Meyer, S. Naczanski, R. Hagen and S. Hackenberg, *Nanomaterials*, 2021, **11**, 629.
- 152 A. Dey, S. Manna, S. Kumar, S. Chattopadhyay, B. Saha and S. Roy, *Cytokine*, 2020, **127**, 154958.
- 153 R. K. Mishra, A. Selim, V. Gowri, A. Ahmad, A. Nadeem, N. Siddiqui, S. S. Raza, G. Jayamurugan and R. Khan, *ACS Biomater. Sci. Eng.*, 2022, **8**, 2088–2095.
- 154 A. Sur, B. Pradhan, A. Banerjee and P. Aich, *PLoS One*, 2015, **10**, e0123905.
- 155 H. Li, D. He, X. Xiao, G. Yu, G. Hu, W. Zhang, X. Wen, Y. Lin, X. Li, H. Lin, Y. Diao and Y. Tang, *ACS Appl. Mater. Interfaces*, 2021, **13**, 25290–25305.
- 156 L. P. Suarez-Kelly, A. R. Campbell, I. V. Rampersaud, A. Bumb, M. S. Wang, J. P. Butchar, S. Tridandapani, L. Yu, A. A. Rampersaud and W. E. Carson, *Nanomedicine*, 2017, **13**, 909–920.
- 157 A. E. Pentecost, C. E. Witherel, Y. Gogotsi and K. L. Spiller, *Biomater. Sci.*, 2017, **5**, 2131.
- 158 A. Pentecost, M. J. Kim, S. Jeon, Y. J. Ko, I. C. Kwon, Y. Gogotsi, K. Kim and K. L. Spiller, *Regen. Biomater.*, 2019, **6**, 163–174.
- 159 R. Tenchov, R. Bird, A. E. Curtze and Q. Zhou, *ACS Nano*, 2021, **15**, 16982–17015.
- 160 A. Ramesh, V. Malik, H. A. Ranjani, H. Smith and A. A. Kulkarni, *Drug Deliv. Transl. Res.*, 2021, **11**, 2317–2327.
- 161 N. Zhang, S. Liu, S. Shi, Y. Chen, F. Xu, X. Wei and Y. Xu, *J. Controlled Release*, 2020, **320**, 168–178.
- 162 Z. Zheng, J. Zhang, J. Jiang, Y. He, W. Zhang, X. Mo, X. Kang, Q. Xu, B. Wang and Y. Huang, *J. Immunother. Cancer.*, 2020, **8**, e000207.
- 163 G. Biber, B. Sabag, A. Raiff, A. Ben-Shmuel, A. Puthenveetil, J. I. C. Benichou, T. Jubany, M. Levy, S. Killner and M. Barda-Saad, *EMBO Mol. Med.*, 2021, **14**, e14073.
- 164 V. Di Francesco, D. Gurgone, R. Palomba, M. F. M. M. Ferreira, T. Catelani, A. Cervadoro, P. Maffia and P. Decuzzi, *ACS Appl. Mater. Interfaces*, 2020, **12**, 37943–37956.
- 165 Q. Wang, L. He, D. Fan, W. Liang and J. Fang, *J. Mater. Chem. B*, 2020, **8**, 1841–1851.
- 166 B. Kim, H. B. Pang, J. Kang, J. H. Park, E. Ruoslahti and M. J. Sailor, *Nat. Commun.*, 2018, **9**, 1–13.
- 167 Y. X. Zhang, Y. Y. Zhao, J. Shen, X. Sun, Y. Liu, H. Liu, Y. Wang and J. Wang, *Nano Lett.*, 2019, **19**, 2774–2783.
- 168 S. Yang, Q. Tang, L. Chen, J. Chang, T. Jiang, J. Zhao, M. Wang and P. R. Chen, *Angew. Chem., Int. Ed.*, 2020, **59**, 18087–18094.
- 169 G. Swaminathan, E. A. Thoryk, K. S. Cox, J. S. Smith, J. J. Wolf, M. E. Gindy, D. R. Casimiro and A. J. Bett, *Sci. Rep.*, 2016, **6**, 1–17.
- 170 S. S. Nogueira, A. Schlegel, K. Maxeiner, B. Weber, M. Barz, M. A. Schroer, C. E. Blanchet, D. I. Svergun, S. Ramishetti, D. Peer, P. Langguth, U. Sahin and H. Haas, *ACS Appl. Nano Mater.*, 2020, **3**, 10634–10645.
- 171 I. Banerjee, M. De, G. Dey, R. Bharti, S. Chattopadhyay, N. Ali, P. Chakrabarti, R. L. Reis, S. C. Kundu and M. Mandal, *Biomater. Sci.*, 2019, **7**, 1161–1178.
- 172 S. Trombino, C. Servidio, A. S. Laganà, F. Conforti, M. Marrelli and R. Cassano, *Molecules*, 2020, **25**, 3535.
- 173 J. P. Barbosa, A. R. Neves, A. M. Silva, M. A. Barbosa, M. S. Reis and S. G. Santos, *Int. J. Nanomed.*, 2016, **11**, 3501.
- 174 J. H. Erasmus, A. P. Khandhar, J. Guderian, B. Granger, J. Archer, M. Archer, E. Gage, J. Fuerte-Stone, E. Larson, S. Lin, R. Kramer, R. N. Coler, C. B. Fox, D. T. Stinchcomb, S. G. Reed and N. Van Hoeven, *Mol. Ther.*, 2018, **26**, 2507.
- 175 X. Song, Y. Jiang, W. Zhang, G. Elfawal, K. Wang, D. Jiang, H. Hong, J. Wu, C. He, X. Mo and H. Wang, *Acta Biomater.*, 2022, **140**, 247–260.
- 176 K. Wang, X. Wang, D. Jiang, Y. Pei, Z. Wang, X. Zhou, J. Wu, X. Mo and H. Wang, *Compos. B Eng.*, 2022, **233**, 109648.
- 177 Z. Liu, L. Luo, S. Zheng, Y. Niu, R. Bo, Y. Huang, J. Xing, Z. Li and D. Wang, *Int. J. Nanomed.*, 2016, **11**, 3571.
- 178 N. Ou, Y. Sun, S. Zhou, P. Gu, Z. Liu, R. Bo, Y. Hu, J. Liu and D. Wang, *Int. J. Biol. Macromol.*, 2018, **109**, 748–760.
- 179 G. Kibria, E. K. Ramos, Y. Wan, D. R. Gius and H. Liu, *Mol. Pharm.*, 2018, **15**, 3625–3633.
- 180 Y. Liang, L. Duan, J. Lu and J. Xia, *Theranostics*, 2021, **11**, 3183.
- 181 W. Fu, C. Lei, S. Liu, Y. Cui, C. Wang, K. Qian, T. Li, Y. Shen, X. Fan, F. Lin, M. Ding, M. Pan, X. Ye, Y. Yang and S. Hu, *Nat. Commun.*, 2019, **10**, 1–12.
- 182 L. Huang, Y. Rong, X. Tang, K. Yi, P. Qi, J. Hou, W. Liu, Y. He, X. Gao, C. Yuan and F. Wang, *Mol. Cancer*, 2022, **21**, 1–19.
- 183 Y. W. Choo, M. Kang, H. Y. Kim, J. Han, S. Kang, J. R. Lee, G. J. Jeong, S. P. Kwon, S. Y. Song, S. Go, M. Jung, J. Hong and B. S. Kim, *ACS Nano*, 2018, **12**, 8977–8993.
- 184 H. Fan, Z. Chen, H. Bin Tang, L. Q. Shan, Z. Y. Chen, X. H. Wang, D. G. Huang, S. C. Liu, X. Chen, H. Yang and D. Hao, *Bioeng. Transl. Med.*, 2022, **7**, e10287.
- 185 G. Jiang, J. Yun, H. J. Kaplan, Y. Zhao, D. Sun and H. Shao, *Clin. Exp. Ophthalmol.*, 2021, **49**, 1069–1077.

- 186 C. E. Boone, C. Wang, M. A. Lopez-Ramirez, V. Beiss, S. Shukla, P. L. Chariou, D. Kupor, R. Rueda, J. Wang and N. F. Steinmetz, *ACS Appl. Nano Mater.*, 2020, **3**, 8037–8051.
- 187 M. Shahgolzari, M. Pazhouhandeh, M. Milani, S. Fiering and A. Y. Khosroushahi, *Nanomedicine*, 2021, **16**, 97–107.
- 188 K. Cheng, T. Du, Y. Li, Y. Qi, H. Min, Y. Wang, Q. Zhang, C. Wang, Y. Zhou, L. Li, S. Ye, X. Zhou, S. Bi, J. Yang and L. Ren, *ACS Appl. Mater. Interfaces*, 2020, **12**, 53682–53690.
- 189 G. M. Joyce, W.-H. Chen, R. S. Sankhala, M. Rao, N. L. Michael, K. Modjarrad, M. Gordon Joyce, A. Hajduczek, P. V. Thomas, M. Choe, E. J. Martinez, W. C. Chang, C. E. Peterson, E. B. Morrison, C. Smith, R. E. Chen, A. Ahmed, L. Wiczorek, A. Anderson, J. Brett Case, Y. Li, T. Oertel, L. Rosado, A. Ganesh, C. Whalen, J. M. Carmen, L. Mendez-Rivera, C. P. Karch, N. Gohain, Z. Villar, D. McCurdy, Z. Beck, J. Kim, S. Shrivastava, O. Jobe, V. Dussupt, S. Molnar, U. Tran, C. B. Kannadka, S. Soman, C. Kuklis, M. Zemil, H. Khanh, W. Wu, M. A. Cole, D. K. Duso, L. W. Kummer, T. J. Lang, S. E. Muncil, J. R. Currier, S. J. Krebs, V. R. Polonis, S. Rajan, P. M. McTamney, M. T. Esser, W. W. Reiley, M. Rolland, N. de Val, M. S. Diamond, G. D. Gromowski, G. R. Matyas, M. Rao, N. L. Michael and K. Modjarrad, *Cell Rep.*, 2021, **37**, 110143.
- 190 Y. Yan, X. Wang, P. Lou, Z. Hu, P. Qu, D. Li, Q. Li, Y. Xu, J. Niu, Y. He, J. Zhong and Z. Huang, *J. Infect. Dis.*, 2020, **221**, 1304–1314.
- 191 A. A. Murji, J. S. Qin, T. Hermanus, L. Morris and I. S. Georgiev, *Viruses*, 2021, **13**, 1296.
- 192 Q. Gao, H. T. Liu, Y. Q. Xu, L. Zhang, Y. R. Liu, Q. Ren, J. P. Sheng and Z. X. Zhang, *Cancer Cell Int.*, 2021, **21**, 1–12.
- 193 M. Z. Zhao, Y. Li, H. Y. Han, L. H. Mo, G. Yang, Z. Q. Liu, C. Ma, P. C. Yang and S. Liu, *Mol. Immunol.*, 2021, **129**, 103–111.
- 194 Y. Jang, H. Kim, S. Yoon, H. Lee, J. Hwang, J. Jung, J. H. Chang, J. Choi and H. Kim, *J. Controlled Release*, 2021, **330**, 293–304.
- 195 M. Elashiry, R. Elsayed, M. M. Elashiry, M. H. Rashid, R. Ara, A. S. Arbab, A. R. Elawady, M. Hamrick, Y. Liu, W. Zhi, R. Lucas, J. Vazquez and C. W. Cutler, *Front. Immunol.*, 2021, **12**, 636222.
- 196 H. Shan, W. Dou, Y. Zhang and M. Qi, *Nanoscale*, 2020, **12**, 22268–22280.
- 197 A. Xia, Y. Zhang, J. Xu, T. Yin and X. J. Lu, *Front. Immunol.*, 2019, **10**, 1719.
- 198 A. K. Singh and J. P. McGuirk, *Lancet Oncol.*, 2020, **21**, e168–e178.
- 199 D. Wang, T. Wang, H. Yu, B. Feng, L. Zhou, F. Zhou, B. Hou, H. Zhang, M. Luo and Y. Li, *Sci. Immunol.*, 2019, **4**, eaau6584.
- 200 Y. Mi, C. C. Smith, F. Yang, Y. Qi, K. C. Roche, J. S. Serody, B. G. Vincent and A. Z. Wang, *Adv. Mater.*, 2018, **30**, 1706098.
- 201 W. Yang, H. Deng, S. Zhu, J. Lau, R. Tian, S. Wang, Z. Zhou, G. Yu, L. Rao, L. He, Y. Ma and X. Chen, *Sci. Adv.*, 2020, **6**, eabb2712.
- 202 R. Smith, E. I. Wafa, S. M. Geary, K. Ebeid, S. O. Alhaj-Suliman and A. K. Salem, *Sci. Adv.*, 2022, **8**, 3150.
- 203 T. T. Smith, S. B. Stephan, H. F. Moffett, L. E. McKnight, W. Ji, D. Reiman, E. Bonagofski, M. E. Wohlfahrt, S. P. S. Pillai and M. T. Stephan, *Nat. Nanotechnol.*, 2017, **12**, 813–822.
- 204 K. Alhallak, J. Sun, K. Wasden, N. Guenther, J. O'Neal, B. Muz, J. King, D. Kohlen, R. Vij, S. Achilefu, J. F. DiPersio and A. K. Azab, *Leukemia*, 2021, **35**, 2346–2357.
- 205 U. Sahin, A. Muik, E. Derhovanessian, I. Vogler, L. M. Kranz, M. Vormehr, A. Baum, K. Pascal, J. Quandt, D. Maurus, S. Brachtendorf, V. Lörks, J. Sikorski, R. Hilker, D. Becker, A. K. Eller, J. Grützner, C. Boesler, C. Rosenbaum, M. C. Kühnle, U. Luxemburger, A. Kemmer-Brück, D. Langer, M. Bexon, S. Bolte, K. Karikó, T. Palanche, B. Fischer, A. Schultz, P. Y. Shi, C. Fontes-Garfias, J. L. Perez, K. A. Swanson, J. Loschko, I. L. Scully, M. Cutler, W. Kalina, C. A. Kyratsous, D. Cooper, P. R. Dormitzer, K. U. Jansen and Ö. Türeci, *Nature*, 2020, **586**, 594–599.
- 206 A. E. Ngonu, T. Syed, A. V. Nguyen, J. A. Regla-Nava, M. Susantono, D. Spasova, A. Aguilar, M. West, J. Sparks, A. Gonzalez, E. Branche, J. L. DeHart, J. B. Vega, P. P. Karmali, P. Chivukula, K. Kamrud, P. Aliahmad, N. Wang and S. Shresta, *Sci. Adv.*, 2020, **6**, eabb2154.
- 207 S. Singha, K. Shao, Y. Yang, X. Clemente-Casares, P. Solé, A. Clemente, J. Blanco, Q. Dai, F. Song, S. W. Liu, J. Yamanouchi, C. S. Umeshappa, R. H. Nanjundappa, P. Detampel, M. Amrein, C. Fandos, R. Tanguay, S. Newbigging, P. Serra, A. Khadra, W. C. W. Chan and P. Santamaria, *Nat. Nanotechnol.*, 2017, **12**, 701–710.
- 208 J. E. Kenison, A. Jhaveri, Z. Li, N. Khadse, E. Tjon, S. Tezza, D. Nowakowska, A. Plasencia, V. P. Stanton, D. H. Sherr and F. J. Quintana, *Proc. Natl. Acad. Sci. U. S. A.*, 2020, **117**, 32017–32028.
- 209 S. Yi, N. B. Karabin, J. Zhu, S. Bobbala, H. Lyu, S. Li, Y. Liu, M. Frey, M. Vincent and E. A. Scott, *Front. bioeng. biotechnol.*, 2020, **8**, 544669.
- 210 R. A. Maldonado, R. A. LaMothe, J. D. Ferrari, A. H. Zhang, R. J. Rossi, P. N. Kolte, A. P. Griset, C. O'Neil, D. H. Altreuter, E. Browning, L. Johnston, O. C. Farokhzad, R. Langer, D. W. Scott, U. H. Von Andrian and T. K. Kishimoto, *Proc. Natl. Acad. Sci. U. S. A.*, 2015, **112**, E156–E165.
- 211 R. Huq, E. L. G. Samuel, W. K. A. Sikkema, L. G. Nilewski, T. Lee, M. R. Tanner, F. S. Khan, P. C. Porter, R. B. Tajhya, R. S. Patel, T. Inoue, R. G. Pautler, D. B. Corry, J. M. Tour and C. Beeton, *Sci. Rep.*, 2016, **6**, 1–15.
- 212 X. Xu, L. Bian, M. Shen, X. Li, J. Zhu, S. Chen, L. Xiao, Q. Zhang, H. Chen, K. Xu and T. Yang, *Diabetologia*, 2017, **60**, 2418–2431.

- 213 B. L. Jamison, T. Neef, A. Goodspeed, B. Bradley, R. L. Baker, S. D. Miller and K. Haskins, *J. Immunol.*, 2019, **203**, 48–57.
- 214 A. Carambia, C. Gottwick, D. Schwinge, S. Stein, R. Digigow, M. Şeleci, D. Mungalpara, M. Heine, F. A. Schuran, C. Corban, A. W. Lohse, C. Schramm, J. Heeren and J. Herkel, *Immunology*, 2021, **162**, 452–463.
- 215 B. C. Palmer, S. Jatana, S. J. Phelan-Dickinson and L. A. DeLouise, *Sci. Rep.*, 2019, **9**, 1–11.
- 216 H. Keum, T. W. Kim, Y. Kim, C. Seo, Y. Son, J. Kim, D. Kim, W. Jung, C. H. Whang and S. Jon, *J. Controlled Release*, 2020, **325**, 359–369.
- 217 J. Jhun, J. Moon, J. Ryu, Y. Shin, S. Lee, K. H. Cho, T. Kang, M. La Cho and S. H. Park, *PLoS One*, 2020, **15**, e0241080.
- 218 J. Ingale, A. Stano, J. Guenaga, S. K. Sharma, D. Nemazee, M. B. Zwick and R. T. Wyatt, *Cell Rep.*, 2016, **15**, 1986–1999.
- 219 T. Tokatlian, D. W. Kulp, A. A. Mutafyan, C. A. Jones, S. Menis, E. Georgeson, M. Kubitz, M. H. Zhang, M. B. Melo, M. Silva, D. S. Yun, W. R. Schief and D. J. Irvine, *Sci. Rep.*, 2018, **8**, 1–13.
- 220 K. Sliepen, G. Ozorowski, J. A. Burger, T. Van Montfort, M. Stunnenberg, C. LaBranche, D. C. Montefiori, J. P. Moore, A. B. Ward and R. W. Sanders, *Retrovirology*, 2015, **12**, 1–5.
- 221 T. J. Moyer, Y. Kato, W. Abraham, J. Y. H. Chang, D. W. Kulp, N. Watson, H. L. Turner, S. Menis, R. K. Abbott, J. N. Bhiman, M. B. Melo, H. A. Simon, S. Herrera-De la Mata, S. Liang, G. Seumoiso, Y. Agarwal, N. Li, D. R. Burton, A. B. Ward, W. R. Schief, S. Crotty and D. J. Irvine, *Nat. Med.*, 2020, **26**, 430–440.
- 222 W. Tao, B. L. Hurst, A. K. Shakya, M. J. Uddin, R. S. J. Ingrole, M. Hernandez-Sanabria, R. P. Arya, L. Bimler, S. Paust, E. B. Tarbet and H. S. Gill, *Antiviral Res.*, 2017, **141**, 62–72.
- 223 R. F. Villar, J. Patel, G. C. Weaver, M. Kanekiyo, A. K. Wheatley, H. M. Yassine, C. E. Costello, K. B. Chandler, P. M. McTamney, G. J. Nabel, A. B. McDermott, J. R. Mascola, S. A. Carr and D. Lingwood, *Sci. Rep.*, 2016, **6**, 1–11.
- 224 H. G. Kelly, H. X. Tan, J. A. Juno, R. Esterbauer, Y. Ju, W. Jiang, V. C. Wimmer, B. C. Duckworth, J. R. Groom, F. Caruso, M. Kanekiyo, S. J. Kent and A. K. Wheatley, *JCI Insight*, 2020, **5**, e136653.
- 225 M. Kanekiyo, M. G. Joyce, R. A. Gillespie, J. R. Gallagher, S. F. Andrews, H. M. Yassine, A. K. Wheatley, B. E. Fisher, D. R. Ambrozak, A. Creanga, K. Leung, E. S. Yang, S. Boyoglu-Barnum, I. S. Georgiev, Y. Tsybovsky, M. S. Prabhakaran, H. Andersen, W. P. Kong, U. Baxa, K. L. Zephir, J. E. Ledgerwood, R. A. Koup, P. D. Kwong, A. K. Harris, A. B. McDermott, J. R. Mascola and B. S. Graham, *Nat. Immunol.*, 2019, **20**, 362.
- 226 W. Wang, X. Zhou, Y. Bian, S. Wang, Q. Chai, Z. Guo, Z. Wang, P. Zhu, H. Peng, X. Yan, W. Li, Y. X. Fu and M. Zhu, *Nat. Nanotechnol.*, 2020, **15**, 406–416.
- 227 M. Fichtner, M. Dreyling, M. Binder and M. Trepel, *J. Hematol. Oncol.*, 2017, **10**, 1–12.
- 228 C. Phipps, Y. Chen, S. Gopalakrishnan and D. Tan, *Ther. Adv. Hematol.*, 2015, **6**, 120.
- 229 S. Weinstein, I. A. Toker, R. Emmanuel, S. Ramishetti, I. Hazan-Halevy, D. Rosenblum, M. Goldsmith, A. Abraham, O. Benjamini, O. Bairey, P. Raanani, A. Nagler, J. Lieberman and D. Peer, *Proc. Natl. Acad. Sci. U. S. A.*, 2016, **113**, E16–E22.
- 230 H. Li, G. Zhang, C. Jiang, F. Zhang, C. Ke, H. Zhao, Y. Sun, M. Zhao, D. Chen, X. Zhu, L. Zhang, B. Li, J. Dai and W. Li, *Oncotarget*, 2015, **6**, 24192–24204.
- 231 A. Falgàs, V. Pallarès, U. Unzueta, Y. Núñez, J. Sierra, A. Gallardo, L. Alba-Castellón, M. A. Mangués, P. Álamo, A. Villaverde, E. Vázquez, R. Mangués and I. Casanova, *Int. J. Nanomed.*, 2021, **16**, 1869–1888.
- 232 S. R. Banihashemi, F. Rahbarizadeh, A. Z. Hosseini, D. Ahmadvand and S. K. Nikkhoi, *Int. Immunopharmacol.*, 2021, **100**, 107927.
- 233 H. Wu, S. Su, Y. Wu, Y. Wu, Z. Zhang and Q. Chen, *Int. Immunopharmacol.*, 2020, **88**, 106933.
- 234 J. Pozsgay, F. Babos, K. Uray, A. Magyar, G. Gyulai, É. Kiss, G. Nagy, B. Rojkovich, F. Hudecz and G. Sármay, *Arthritis Res. Ther.*, 2016, **18**, 1–12.
- 235 L. Rao, L. Wu, Z. Liu, R. Tian, G. Yu, Z. Zhou, K. Yang, H. G. Xiong, A. Zhang and G. T. Yu, *Nat. Commun.*, 2020, **11**, 4909.
- 236 P. M. K. Tang, D. J. Nikolic-Paterson and H. Y. Lan, *Nat. Rev. Nephrol.*, 2019, **15**, 144–158.
- 237 S. Watanabe, M. Alexander, A. V. Misharin and G. R. S. Budinger, *J. Clin. Invest.*, 2019, **129**, 2619–2628.
- 238 S. C. Funes, M. Rios, J. Escobar-Vera and A. M. Kalergis, *Immunology*, 2018, **154**, 186–195.
- 239 X. Miao, X. Leng and Q. Zhang, *Int. J. Mol. Sci.*, 2017, **18**, 336.
- 240 F. Zhang, N. N. Parayath, C. I. Ene, S. B. Stephan, A. L. Koehne, M. E. Coon, E. C. Holland and M. T. Stephan, *Nat. Commun.*, 2019, **10**, 1–16.
- 241 T. Sun, C. H. T. Kwong, C. Gao, J. Wei, L. Yue, J. Zhang, R. D. Ye and R. Wang, *Theranostics*, 2020, **10**, 10106–10119.
- 242 E. Hirata, E. Miyako, N. Hanagata, N. Ushijima, N. Sakaguchi, J. Russier, M. Yudasaka, S. Iijima, A. Bianco and A. Yokoyama, *Nanoscale*, 2016, **8**, 14514–14522.
- 243 J. Wu, J. Zhu, Q. Wu, Y. An, K. Wang, T. Xuan, J. Zhang, W. Song, H. He, L. Song, J. Zheng and J. Xiao, *ACS Appl. Mater. Interfaces*, 2021, **13**, 2230–2244.
- 244 Q. Gao, J. Zhang, C. Chen, M. Chen, P. Sun, W. Du, S. Zhang, Y. Liu, R. Zhang, M. Bai, C. Fan, J. Wu, T. Men and X. Jiang, *ACS Nano*, 2020, **14**, 3980–3990.
- 245 J. W. Lim, W. Na, H. O. Kim, M. Yeom, A. Kang, G. Park, C. Park, J. Ki, S. Lee, B. Jung, H. H. Jeong, D. Park, D. Song and S. Haam, *J. Mater. Chem. B*, 2020, **8**, 5620–5626.
- 246 Y. Yang, L. Guo, Z. Wang, P. Liu, X. Liu, J. Ding and W. Zhou, *Biomaterials*, 2021, **264**, 120390.

- 247 Y. Liu, J. Wang, J. Zhang, S. Marbach, W. Xu and L. Zhu, *ACS Appl. Mater. Interfaces*, 2020, **12**, 52402–52414.
- 248 J. Tang, M. E. Lobatto, L. Hassing, S. Van Der Staay, S. M. Van Rijs, C. Calcagno, M. S. Braza, S. Baxter, F. Fay, B. L. Sanchez-Gaytan, R. Duivenvoorden, H. B. Sager, Y. M. Astudillo, W. Leong, S. Ramachandran, G. Storm, C. Pérez-Medina, T. Reiner, D. P. Cormode, G. J. Strijkers, E. S. G. Stroes, F. K. Swirski, M. Nahrendorf, E. A. Fisher, Z. A. Fayad and W. J. M. Mulder, *Sci. Adv.*, 2015, **1**, e1400223.
- 249 C. Deng, Q. Zhang, P. He, B. Zhou, K. He, X. Sun, G. Lei, T. Gong and Z. Zhang, *Nat. Commun.*, 2021, **12**, 1–15.
- 250 I. Relaño-Rodríguez, M. D. L. S. Espinar-Buitrago, V. Martín-Cañadilla, R. Gómez-Ramírez and M. Á. Muñoz-Fernández, *Int. J. Mol. Sci.*, 2021, **22**, 8366.
- 251 Q. Tan, L. He, X. Meng, W. Wang, H. Pan, W. Yin, T. Zhu, X. Huang and H. Shan, *J. Nanobiotechnology*, 2021, **19**, 173.
- 252 A. Singh, S. Chakraborty, S. W. Wong, N. A. Hefner, A. Stuart, A. S. Qadir, A. Mukhopadhyay, K. Bachmaier, J. W. Shin, J. Rehman and A. B. Malik, *Proc. Natl. Acad. Sci. U. S. A.*, 2022, **119**, e2121098119.
- 253 A. Cifuentes-Rius, A. Desai, D. Yuen, A. P. R. Johnston and N. H. Voelcker, *Nat. Nanotechnol.*, 2021, **16**, 37–46.
- 254 M. K. S. Rajput, S. S. Kesharwani, S. Kumar, P. Muley, S. Narisetty and H. Tummala, *ACS Appl. Mater. Interfaces*, 2018, **10**, 27589–27602.
- 255 D. Mao, F. Hu, Z. Yi, K. Kenry, S. Xu, S. Yan, Z. Luo, W. Wu, Z. Wang, D. Kong, X. Liu and B. Liu, *Sci. Adv.*, 2020, **6**, eabb2712.
- 256 H. Zhang, X. You, X. Wang, L. Cui, Z. Wang, F. Xu, M. Li, Z. Yang, J. Liu, P. Huang, Y. Kang, J. Wu and X. Xia, *Proc. Natl. Acad. Sci. U. S. A.*, 2021, **118**, e2005191118.
- 257 Y. Si, Q. Tian, F. Zhao, S. H. Kelly, L. S. Shores, D. F. Camacho, A. I. Sperling, M. S. Andrade, J. H. Collier and A. S. Chong, *Sci. Adv.*, 2020, **6**, 995–1002.
- 258 X. Zheng, C. Sun, R. Yu, X. Chu, J. Xu, C. Liu, M. Zhao, X. Xu, M. Xia and C. Wang, *Int. J. Pharm.*, 2020, **577**, 119034.
- 259 Y. Zhang, S. Shen, G. Zhao, C. F. Xu, H. B. Zhang, Y. L. Luo, Z. T. Cao, J. Shi, Z. Bin Zhao, Z. X. Lian and J. Wang, *Biomaterials*, 2019, **217**, 119302.
- 260 T. W. Kim, Y. Kim, W. Jung, D. E. Kim, H. Keum, Y. Son and S. Jon, *J. Controlled Release*, 2021, **331**, 74–84.
- 261 U. e. Habiba, M. Rafiq, M. B. Khawar, B. Nazir, G. Haider and N. Nazir, *Adv. Cancer Biol.: Metastasis*, 2022, **5**, 100053.
- 262 A. E. Barberio, S. G. Smith, S. Correa, C. Nguyen, B. Nhan, M. Melo, T. Tokatlian, H. Suh, D. J. Irvine and P. T. Hammond, *ACS Nano*, 2020, **14**, 11238–11253.
- 263 I. Lai, S. Swaminathan, V. Baylot, A. Mosley, R. Dhanasekaran, M. Gabay and D. W. Felsher, *J. Immunother. Cancer*, 2018, **6**, 1–11.
- 264 J. Q. Liu, C. Zhang, X. Zhang, J. Yan, C. Zeng, F. Talebian, K. Lynch, W. Zhao, X. Hou, S. Du, D. D. Kang, B. Deng, D. W. McComb, X. F. Bai and Y. Dong, *J. Controlled Release*, 2022, **345**, 306–313.
- 265 A. Kienzle, S. Kurch, J. Schlöder, C. Berges, R. Ose, J. Schupp, A. Tuettenberg, H. Weiss, J. Schultze, S. Winzen, M. Schinnerer, K. Koynov, M. Mezger, N. K. Haass, W. Tremel and H. Jonuleit, *Adv. Healthc. Mater.*, 2017, **6**, 1700012.
- 266 T. Wu, Q. Qiao, X. Qin, D. Zhang and Z. Zhang, *Nanomedicine*, 2019, **18**, 66–77.
- 267 T. T. Tang, B. Wang, M. Wu, Z. L. Li, Y. Feng, J. Y. Cao, D. Yin, H. Liu, R. N. Tang, S. D. Crowley, L. L. Lv and B. C. Liu, *Sci. Adv.*, 2020, **6**, 748–760.
- 268 C. Baecher-Allan, B. J. Kaskow and H. L. Weiner, *Neuron*, 2018, **97**, 742–768.
- 269 L. F. González, E. Acuña, G. Arellano, P. Morales, P. Sotomayor, F. Oyarzun-Ampuero and R. Naves, *J. Controlled Release*, 2021, **331**, 443–459.
- 270 T. M. Raimondo and D. J. Mooney, *Sci. Adv.*, 2021, **7**, 3693–3716.
- 271 R. Halwani, A. S. Shaik, E. Ratemi, S. Afzal, R. Kenana, S. Al-Muhsen and A. Al Faraj, *Exp. Mol. Med.*, 2016, **48**, e262.
- 272 B. Ng, J. Dong, G. D'Agostino, S. Viswanathan, A. A. Widjaja, W. W. Lim, N. S. J. Ko, J. Tan, S. P. Chothani, B. Huang, C. Xie, C. J. Pua, A. M. Chacko, N. Guimarães-Camboa, S. M. Evans, A. J. Byrne, T. M. Maher, J. Liang, D. Jiang, P. W. Noble, S. Schafer and S. A. Cook, *Sci. Transl. Med.*, 2019, **11**, 1237.
- 273 X. Bai, G. Zhao, Q. Chen, Z. Li, M. Gao, W. Ho, X. Xu and X. Q. Zhang, *Sci. Adv.*, 2022, **8**, 7162.
- 274 L. Rao, S. Xia, W. Xu, R. Tian, G. Yu, C. Gu, P. Pan, Q. F. Meng, X. Cai, D. Qu, L. Lu, Y. Xie, S. Jiang and X. Chen, *Proc. Natl. Acad. Sci. U. S. A.*, 2020, **117**, 27141–27147.
- 275 A. Zarantonello, H. Pedersen, N. S. Laursen and G. R. Andersen, *Biomolecules*, 2021, **11**, 1–24.
- 276 N. Maisha, T. Coombs and E. Lavik, *ACS Biomater. Sci. Eng.*, 2020, **6**, 4903–4915.
- 277 B. Zhang, R. C. Lai, W. K. Sim, A. B. H. Choo, E. B. Lane and S. K. Lim, *Int. J. Mol. Sci.*, 2021, **22**, 1–13.
- 278 Y. Li and F. Lin, *Acta Biomater.*, 2019, **99**, 330–338.
- 279 L. Avalle, F. Marino, A. Camporeale, C. Guglielmi, D. Viavattene, S. Bandini, L. Conti, J. Cimino, M. Forni, C. Zanini, A. Ghigo, R. L. Bogorad, F. Cavallo, P. Provero, V. Koteliansky and V. Poli, *Mol. Ther.-Methods Clin. Dev.*, 2020, **18**, 62–72.
- 280 M. A. Colden, S. Kumar, B. Munkhbileg and D. V. Babushok, *Front. Immunol.*, 2022, **12**, 830172.
- 281 L. Zhang, W. Qiu, S. Crooke, Y. Li, A. Abid, B. Xu, M. G. Finn and F. Lin, *ACS Chem. Biol.*, 2017, **12**, 539–547.
- 282 C. F. Jones and D. W. Grainger, *Adv. Drug Delivery Rev.*, 2009, **61**, 438–456.
- 283 S. Wahab, M. Ghazwani, U. Hani, A. R. Hakami, A. A. Almezizia, W. Ahmad, M. Z. Ahmad, P. Alam and S. Annadurai, *Molecules*, 2023, **28**, 1216.
- 284 J. M. Metselaar and T. Lammers, *Drug Delivery Transl. Res.*, 2020, **10**, 721–725.