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Benchmark of a functional-group database for distributed polarizability and dipole moment in biomolecules†

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The extraction of functional-group properties in condensed phases is very useful for predicting material behaviors, including those of biomaterials. For this reason, computational approaches based on partitioning schemes have been developed aiming at rapidly and accurately estimating properties from chemically meaningful building blocks. A comprehensive database of group polarizabilities and dipole moments is useful not only to predict the optical properties of biomacromolecules but also to improve molecular force fields focused on simulating biochemical processes. In this work we benchmark a database of distributed polarizabilities and dipole moments for functional groups extracted from a series of polypeptides. This allows reconstruction of a variety of relevant chemical environments. The accuracy of our database was tested to predict the electro-optical properties of larger peptides and also simpler amino acids for which density functional theory calculations at the M06-HF/aug-cc-pVDZ level of theory was chosen as the reference. This approach is reasonably accurate for the diagonal components of the polarizability tensor, with errors not larger than 15–20%. The anisotropy of the polarizability is predicted with smaller efficacy though. Solvent effects were included explicitly by surrounding the database entries by a box of water molecules whose distribution was optimized using the CHARMM force field.

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1 Introduction

Biomaterials are either artificial substances incorporated in living systems, or natural bioproducts with broader applications in materials science and nanotechnology.^{1,2} From an optical point of view, biomaterials are evaluated based on refractive indices and transparency, whereas mechanical, chemical and biological properties may strongly depend on the specific application. Despite their satisfactory performance, current biomaterials are mainly developed based on a trial-and-error optimization, rather than being properly engineered.³ For optimizing the materials efficiency, it is vital to develop accurate methodologies aiming at rapid screening of the best candidates for a given functionality.⁴ The long-term goal of our work is to develop algorithms for optimizing the optical efficiency of functional biomaterials, based on their breakdown into smaller

building blocks which could be amino acids, peptides, lipids, nucleic acids, *etc.* This implies additive schemes for the rapid prediction of bulk properties.

A variety of schemes has been proposed to calculate the dipole moments and distributed functional-group polarizabilities.^{5–12} Bader chose the topological partitioning of the charge density based on Quantum Theory of Atoms In Molecules (QTAIM).¹³ Keith generalized the Bader approach within the “atomic response theory”,¹⁴ which was later implemented in the *PolaBer* program.¹⁵ One of the advantages is the removal of origin-dependent terms that are particularly troublesome for the transferability of the functional groups. Indeed, QTAIM offers advantages because it leads to an exact partitioning of the charge density in real space. This contrasts with schemes such as the one proposed by Stone in the framework of the distributed multipole analysis¹⁶ and with the method proposed by Angyan, in which atomic polarizabilities are obtained by differentiating the energy, rather than dipole moments.^{17–20}

The modelling of biomolecules through charge-density based parameters has become an important research field. Databases of functional groups have been created aiming at having a more realistic reconstruction of the electro-optical and electrostatic properties of molecules.^{21,22} The hypothesis of transferability in biomolecules is strongly supported.²³ More recently, generalized

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databases which include polarizability tensors of various relevant functional groups have been proposed.²⁴ In that work, the polarizability of a transferable functional group is reconstructed from a set of previously selected molecular entries rotated to a common coordinate system and then clustered appropriately using a genetic algorithm. A combination of the building blocks thus allows prediction of the electro-optical properties with high accuracy. Methods based on artificial intelligence can also be used to precisely estimate atomic polarizabilities from a very large molecular training set.²⁵

We note that the inclusion of polarizabilities as parameters to describe part of the molecular energy in classical force-field simulations could significantly improve the accuracy, in particular if one is interested in taking into account effects due to long-range interactions such as intermolecular contacts. Traditionally, additive force fields have been developed including point atomic charges to compute the electrostatic contribution of intermolecular interactions, thus neglecting details of the charge density distribution. This results in an incomplete inclusion of polarization effects due to the vicinity. New developments in molecular mechanics have started to correct for such perturbations by considering distributed polarizabilities within a set of parameters.²⁶ Therefore, an accurate database of polarizability tensors would also be desirable from this perspective.

While building blocks can be extracted from very accurate gas-phase calculations, their properties are only representative of the actual biomaterial when the chemical environment is taken into account. For biomolecules, this necessarily implies careful consideration of intermolecular interactions. Because these contacts are typically weaker than covalent bonds, semi-empirical approaches based on classical electrostatics have long been used to estimate the opto-electronic properties of the biomaterial from gas-phase calculations.^{27–31} In such methods, the electric field experienced by a particular molecule in the condensed phase has additive contributions from an externally applied field and from the field produced by all vicinal dipoles. Most of these implementations reduce entire molecules to point dipoles, thus do not explicitly treat functional groups. One of these implementations is known as the dipole interaction model (DIM), and uses a dipole field tensor whose components depend on each interacting pair.³² We have built a modified version of DIM which employs a distributed functional-group algorithm.^{33,34} Here, we propose a systematic building-block database for estimating electro-optical properties of biomolecules, particularly proteins, in both gas and condensed phase. Given the importance of aqueous medium in biochemistry, the database entries are suitable for biomolecules solvated with water. We used the database to estimate the properties of a series of amino acids and polypeptides, and benchmarked the results against quantum-mechanical calculations.

2 Computational methods

We have used a protein of the melanoma-antigen gene family, MAGE-1,³⁵ as a reference to create entries for our database.

A fragment consisting of nine amino acid fragments, namely H-Glu-Ala-Asp-Pro-Thr-Gly-His-Ser-Tyr-OH, was selected. Then, the residues were randomly combined to create seven peptides, each one consisting of five amino acid residues: H-Asp-Ala-Glu-Gly-Ser-OH, H-Thr-Gly-Pro-Tyr-Ser-OH, H-His-Ala-Glu-Pro-Tyr-OH, H-Thr-Asp-His-Pro-Ala-OH, H-Tyr-Ser-Glu-Asp-His-OH, H-Ala-Thr-Ser-Gly-His-OH and H-Glu-Pro-Asp-Tyr-Thr-OH. Zwitterions were always chosen. The molecular geometries for these peptides were optimized in the gas phase using the CHARMM additive force field.³⁶ Building blocks were extracted after calculating the dipole moment and polarizability for each atom. Database entries were generated by averaging those quantities for each functional group.

To assess the quality of the database, a testing set containing twenty four different molecules was chosen. It includes eight amino acids, namely Ala, Gly, Glu, Asp, His, Ser, Tyr, and Thr, and some of their corresponding di- and tri-peptides. Geometries were again optimized in the gas phase and zwitterionic form, using the CHARMM force field.

2.1 Quantum-mechanical calculations

Although the gold standard for wave-function calculations in small organic molecules is coupled-cluster, inclusion of an electronic-correlation level higher than CCSD typically improves the polarizabilities only slightly.³⁷ Hybrid and meta-hybrid DFT functionals proved to be very efficient at reproducing CCSD polarizabilities³⁸ for organic molecules, including amino acids and peptides.^{31,33} In particular, functionals of the M06 class³⁹ give very consistent quantities when compared to the CCSD results. Thus, in this work, M06-HF is used.

We have also benchmarked some correlation-consistent basis functions for small organic molecules, ranging from cc-pVDZ to cc-pVQZ and include many of their augmented versions.³³ The combination of basis-set polarization and diffusion was shown to be extremely important for obtaining good-quality polarizabilities. The inclusion of diffuse functions is usually more important than the valence splitting. For this reason, aug-cc-pVDZ returns similar values to cc-pVQZ, a much larger basis, and to aug-cc-pVQZ, the largest basis tested. Inclusion of more functions, as in the case of d-aug-cc-pVDZ, did not show any significant improvement. Quantum calculations in this work were performed at the aug-cc-pVDZ level of theory. Gaussian 16⁴⁰ was used for DFT calculations. AIMAll⁴¹ software was employed to partition the charge density according to QTAIM.

2.2 Dipole-moment and polarizability calculations

PolaBer¹⁵ was used to compute origin-independent dipole moments and distributed atomic polarizabilities. The dipole moment of an atomic basin Ω is the sum of a polarization component $\mu_p(\Omega)$ and a charge-translation contribution $\mu_c(\Omega)$:

$$\mu(\Omega) = \mu_p(\Omega) + \mu_c(\Omega) = - \int_{\Omega} [r - R_{\Omega}] \rho(r) dr + [R_{\Omega} - R_0] q(\Omega) \quad (1)$$

$q(\Omega)$ is the atomic-basin charge, R_{Ω} is the vector position of Ω , and R_0 is the origin of an arbitrary coordinate system.



The charge-translation term can be rewritten as an origin-independent contribution that allows for the transferability of properties from one molecular system to another:¹⁴

$$\mu_c(\Omega) = \sum_A q(\Omega|A)[R_{BCP} - R_\Omega] \quad (2)$$

where $q(\Omega|A)$ is the charge induced on Ω due to its bonding to A , and R_{BCP} is the position vector of the bond critical point which links nuclei Ω and A , measured with respect to the arbitrary origin R_0 . The summation runs over all A basins bonded to Ω .

Calculations were performed using a static field with a magnitude of 0.001 a.u. applied along each one of the positive and negative Cartesian directions. The polarizability tensor components of a given basin are calculated by numerically differentiating the dipole moment with respect to the applied field. This procedure is exact provided that the perturbation is small enough to guarantee a linear response. The $\alpha_{ij}(\Omega)$ component of the tensor is given by:

$$\alpha_{ij}(\Omega) = \lim_{F_j^{ext} \rightarrow 0} \frac{\mu_i^j(\Omega) - \mu_i^0(\Omega)}{F_j^{ext}} \quad (3)$$

in which $\mu_i^j(\Omega)$ is the dipole component in the i direction calculated with the uniform field F^{ext} applied along the j direction.

Atomic polarizability tensors can be visualised in the same space as the molecule assuming $1 \text{ \AA}^3 \equiv 1 \text{ \AA}$. A scaling factor, usually of 0.2 \AA^{-2} , may be necessary to reduce the size of ellipsoids for visualization purposes. The isotropic polarizability is computed as the arithmetical average of the main diagonal components and the anisotropy of the tensor is estimated as:

$$\Delta\alpha = \left\{ \frac{1}{2} [3\text{Tr}(\alpha^2) - (\text{Tr}\alpha)^2] \right\}^{1/2} \quad (4)$$

2.3 Database

After calculation and functional-group extraction, dipole-moment vectors and polarizability tensors are oriented according to a specific framework defined by the geometry of each functional group. Because they are orientation dependent, the first step in order to generate an exportable building block is to rotate equivalent groups to a common framework. This can be achieved by employing a rotation matrix obtained from diagonalization of the charge tensor \mathbf{Q} , analogous to the inertia tensor where masses are replaced by atomic numbers Z :²⁴

$$\mathbf{Q} = \begin{bmatrix} \sum_i Z_i(y_i^2 + z_i^2) & -\sum_i Z_i y_i x_i & -\sum_i Z_i z_i x_i \\ -\sum_i Z_i y_i x_i & \sum_i Z_i(x_i^2 + z_i^2) & -\sum_i Z_i z_i y_i \\ -\sum_i Z_i z_i x_i & -\sum_i Z_i z_i y_i & \sum_i Z_i(x_i^2 + y_i^2) \end{bmatrix} \quad (5)$$

x_i , y_i , z_i are the atomic coordinates of a given building block referring to an arbitrary coordinate system where the origin is the center of charges of the molecule from which it came from.



Fig. 1 Twenty five transferable building blocks extracted from the prototypical peptides and their correspondent dummy atoms (marked with yellow crosses).

The charge-density distribution of a building block, hence its associated dipole moment and polarizability, depends on the charge distribution of the surroundings. For example, the properties of a $-\text{CH}_2-$ group extracted from a highly symmetrical molecule such as methane would be different from those extracted from a less symmetrical molecule such as glycine. In the former, the charge density of the group possesses the same symmetry as the idealized $-\text{CH}_2-$ group which is C_{2v} . In the latter, although the idealized functional group should still have C_{2v} symmetry, the presence of different atoms directly attached to it (C and N atom) lowers its effective symmetry to C_s . For a C_{2v} charge density distribution, the polarizability tensor must have all off-diagonal terms equal to zero but for the C_s symmetry, the values for off-diagonal tensor components can be either positive or negative. In order to ensure consistent off-diagonal components after group rotation, we have explicitly included neighboring atoms to compute the transferable functional group. This implies that our $-\text{CH}_2-$ building block is actually a $R_1-\text{CH}_2-R_2$ group, where R_1 and R_2 are dummies (Fig. 1).

2.4 Classical molecular dynamics

Fifty eight water molecules were added surrounding a central glycine molecule, using the CHARMM-GUI⁴² feature. The geometry of the aggregate was optimized using the Adopted Basis Newton Raphson Method, implemented on the CHARMM³⁶ additive force field, free version 46b1. Further equilibration was performed and 100 000 steps were computed using the NVT ensemble, with an interval of 1 fs. A Nose-Hoover thermostat^{43,44} was used for temperature control, at 303.15 K. Afterwards, molecular dynamics were performed using the NPT ensemble. Berendsen's⁴⁵ thermostat was used to control the temperature and pressure at 303.15 K



and 1 atm. 200 000 molecular frames were calculated with a 2 fs interval, giving an overall time of 400 ps. The geometries were extracted every 20 ps and further used for estimation of group polarizabilities with the database approach.

3 Results and discussion

The importance of taking into account both inter- and intramolecular chemical environments of a building block when estimating the opto-electronic properties of molecular biomaterials from transferable functional groups was discussed in our previous works.^{33,34} Covalent bonds are certainly the most relevant interactions to be considered in case highly accurate polarizability tensors are desired. Intermolecular contacts, on the other hand, such as hydrogen bonds or other non-covalent contacts, play a smaller role and therefore can be efficiently taken into account by means of semi-empirical approaches such as a DIM. When transferable building blocks were extracted from simple amino acids or amino acid residues, we needed to properly correct for intramolecular environment effects. In this work we propose an alternative that is to extract building blocks from larger systems, namely, polypeptides containing five amino acid residues. In this case, the most relevant intramolecular polarization effects are already properly considered at the electronic-structure calculation stage without the need for further corrections. As we show in the next paragraphs, the functional groups are capable of reconstructing dipole moments and polarizabilities of other biomolecules with acceptable quality. As for the effect of intermolecular interactions, our database is expanded to explicitly consider water molecules solvating the functional groups.

Fig. 2 shows how the Cartesian components of dipole moments for thirty two molecules reconstructed using our database compare with the quantum-mechanical results. Deviations from the quantum reference are more pronounced in the cases where no peptide bonds are present, as for the amino acids. The correlation coefficient (R^2) is just 0.87. Because the dipole moments directly depend on charges of the atomic basins, they are very sensitive to the chemical environment, either intra- or intermolecular. The proximity between the carboxylate and the ammonium groups in the zwitterionic forms of the amino acids induces some charge-density redistribution when compared to the same terminal groups in larger peptides, where they are more distant apart. Upon adding one or two peptide bonds, thus generating di- and tripeptides, the accuracy of the database reconstruction improves, with correlation coefficients of $R^2 = 0.96$ and $R^2 = 0.92$, respectively. This is justified by the fact that the terminal charges are more distant to each other, as occurs for the pentapeptides used to calculate the transferable building blocks. Just as a consistency check, we have verified how the database reconstruction replicates the dipole moments of these polypeptides as well. The correlation coefficient is of course almost perfect, $R^2 = 0.99$. Since the database entries were generated by averaging data, each functional group possesses a certain level of molecular flexibility that depends on the similarity of the chemical environments



Fig. 2 Cartesian components of dipole moments of amino acids (A), dipeptides (B), tripeptides (C) and polypeptides (D), reconstructed using the database benchmarked against quantum-mechanical calculations at the M06-HF/aug-cc-pVDZ level of theory. The chart D refers to the peptides used to create entries for the database. All values are in atomic units.



Table 1 Building blocks dipole moments and polarizabilities used as entries for the database (atomic units). The values represent the mean quantity of a given building block, with their respective standard deviation (SD) after the rotation procedure. SD was calculated using the following expression:

$$\left(\frac{\sum (x_n - \langle x \rangle)^2}{n-1} \right)^{\frac{1}{2}}$$

The functional groups terminology follows Fig. 1; CTER and NTER refer to the terminal groups of the peptide chain

	μ_x	μ_y	μ_z	α_{xx}	α_{yy}	α_{zz}	α_{xy}	α_{xz}	α_{yz}
CH-ASP	-1.5655	0.1040	-0.0917	10.620	5.995	7.889	1.615	1.345	1.479
SD	0.2944	0.2009	0.1943	1.5984	0.723	0.556	0.675	0.691	0.560
CHCH ₃ -ALA	-0.2090	-1.4430	0.1223	25.238	22.684	18.067	-0.339	1.531	0.811
SD	0.1998	0.2016	0.0436	1.3220	1.153	0.993	1.715	0.547	0.728
CH ₂ -ASP	-0.2088	0.0164	-0.2216	12.198	9.135	11.303	-0.276	0.445	1.446
SD	0.2584	0.0260	0.1996	1.0385	0.420	0.606	0.700	0.516	0.713
CO-ACID	-1.2924	0.9607	-0.0470	14.503	16.399	8.159	-4.667	-0.702	0.479
SD	0.4615	0.3995	0.1601	2.6708	1.356	0.385	0.923	1.552	1.314
COO ⁻ -CTER	-0.3515	2.5330	0.1233	27.600	29.122	16.258	-2.463	0.223	0.233
SD	0.5143	0.4160	0.2218	2.1888	2.686	1.589	2.029	2.627	1.114
CH-GLU	-1.5930	0.2158	0.1305	10.323	6.928	9.562	0.783	0.746	1.803
SD	0.3344	0.1405	0.1565	1.5714	1.083	0.951	1.314	0.669	0.693
CH ₂ CH ₂ -GLU	0.0421	0.2309	0.1488	27.357	23.330	18.707	-0.483	0.164	3.406
SD	0.4350	0.2592	0.1294	1.4756	1.143	0.599	1.104	1.247	1.152
CH ₂ -GLY	-1.8027	0.1847	-0.0246	12.766	9.243	8.219	1.312	0.720	0.393
SD	0.1652	0.0759	0.0396	1.5685	1.170	0.728	1.342	0.579	0.615
CONH	-0.1341	3.7732	-0.0563	30.210	34.645	12.955	-9.197	0.859	0.442
SD	0.4193	0.5946	0.1156	2.1619	2.626	1.156	2.962	2.177	1.988
CH-HSD	-1.4580	0.2301	0.1644	10.228	6.897	9.030	0.923	0.301	1.877
SD	0.3587	0.0959	0.1612	1.6542	0.773	0.612	1.001	0.842	0.259
CH ₂ -HSD	0.0369	-0.0406	0.0420	12.121	8.658	13.974	0.104	0.815	-0.420
SD	0.2009	0.0244	0.1004	0.4177	0.605	1.165	0.843	0.449	0.751
IMI	-1.2614	1.1101	-0.0789	48.111	58.992	31.601	-0.254	2.149	-0.319
SD	0.2862	0.5789	0.2798	6.1780	1.119	1.245	2.549	3.349	1.519
NH ₃ ⁺ -NTER	1.3408	0.2149	-0.0512	13.455	8.353	8.412	2.494	0.598	0.132
SD	0.2141	0.1208	0.2050	0.8095	0.530	0.864	0.335	0.482	0.356
OH-ACID	-0.4716	0.8199	0.0162	11.910	6.428	6.472	1.172	-0.146	-0.110
SD	0.1968	0.1430	0.0709	1.5458	0.356	1.444	0.823	0.670	0.717
OH-PHE	-0.8353	0.4923	0.0347	16.206	7.757	5.301	-0.220	-0.264	-0.434
SD	0.2606	0.2314	0.0501	1.4456	2.393	0.605	1.420	0.471	0.744
OH-SER	-0.8759	0.5959	-0.0045	13.490	8.420	6.000	0.401	-0.070	-0.183
SD	0.3459	0.2484	0.0492	1.0083	0.947	0.619	1.743	0.894	1.209
OH-THR	-0.4889	0.8809	-0.0242	12.488	8.162	5.411	0.690	0.111	0.019
SD	0.1739	0.2544	0.0325	0.3543	1.719	0.676	1.147	0.675	0.767
PHE	-0.8746	0.0284	-0.3525	82.087	67.753	39.262	1.487	5.254	0.906
SD	0.5222	0.0903	0.2786	7.0060	1.932	3.821	2.518	3.156	1.627
PRO-ring	1.0803	-4.8003	1.1733	81.985	70.247	49.983	-7.335	-2.320	-5.746
SD	0.5198	0.4051	0.3166	4.0049	3.267	2.515	4.250	2.373	1.666
CH-SER	-1.0798	-0.0184	-0.1136	9.496	6.870	8.256	1.603	1.217	1.060
SD	0.2107	0.1878	0.1927	0.6564	0.313	0.527	0.670	0.546	0.711
CH ₂ -SER	-0.2809	0.0089	0.0546	12.324	9.314	7.800	-0.253	-0.604	0.492
SD	0.1998	0.0487	0.1454	0.6112	0.331	0.353	0.355	0.253	0.721
CH-THR	-1.2957	0.2127	0.1282	8.716	7.077	7.794	0.673	0.618	1.258
SD	0.2277	0.1119	0.1374	0.6678	0.491	0.307	1.042	0.318	0.686
CH ₃ -THR	0.0817	0.1038	0.4302	26.699	19.869	20.134	0.530	0.024	-0.365
SD	0.1841	0.2011	0.2361	3.0589	0.782	1.801	1.086	0.844	1.510
CH-TYR	-1.4295	0.0024	-0.1510	10.949	7.953	9.053	2.089	1.324	2.357
SD	0.2783	0.1611	0.1704	1.4963	1.027	0.886	0.611	1.383	1.073
CH ₂ -TYR	-0.2982	-0.0052	-0.1210	13.720	8.411	15.348	-0.521	1.923	0.807
SD	0.1636	0.0831	0.1877	0.8230	0.706	0.897	0.606	1.608	0.269

among the set of prototypical molecules used in the definition of the building block, as shown in Table 1. Molecular polarizabilities estimated with the database are very accurate, as shown in Fig. 3. Each component of the diagonalized tensor is accurately reproduced as well (numerical data can be found in the ESI†).

Fig. 4 compares the magnitude and the direction of the molecular dipole moment vectors obtained using the database with those calculated from quantum mechanics. Our database is efficient to estimate not only the magnitude of the dipoles, with a mean percentual deviation of 14%, but also their directions, with

an average difference of around 6°. Fig. 5 stresses the quality of the building blocks to reproduce the overall isotropic polarizability α_{iso} and its anisotropy $\Delta\alpha$. The latter quantity is, as one would expect, more variable, emphasizing the importance of intramolecular environment effects on determining the shape and orientation of the polarizability ellipsoids.

3.1 Electrostatic potential maps

Aiming to test the efficiency of our database to estimate other properties in biomolecules, we have used the transferable





Fig. 3 Diagonalized polarizability tensor components for amino acids (A), dipeptides (B), tripeptides (C) and polypeptides (D), obtained using the database benchmarked against quantum-mechanical calculations at the M06-HF/aug-cc-pVDZ level of theory. The chart D at the bottom refers to the peptides used to create entries for the database. All values are in atomic units.

functional groups to reconstruct the dipolar electric potential for the MAGE-1 fragment (Fig. 6), which is a good approximation to the total electric potential.⁴⁶ The dipolar potential of an atomic group defining a basin Ω at a position \mathbf{r} was therefore calculated from the ground-state dipole moment of the functional group:

$$V(\mathbf{r}; \Omega) = \frac{\boldsymbol{\mu}(\Omega) \cdot \mathbf{r}}{|\mathbf{r}|^3} \quad (6)$$

The sum over all functional groups provides the molecular electrostatic potential map. The set of distributed atomic dipoles could in principle be used to provide accurate electrostatic mapping. However, replacing them with larger building blocks and locating their dipole moments at each center of charge gives a very good overview of the charge distribution in the polypeptides. Electrostatic potential maps are relevant benchmarks because they are fundamental to understanding and interpreting the sources of electric properties and reactivity.

Electrostatic potential maps estimated using the dipole moments coming from the building-block database do not superimpose perfectly with the ones calculated using an atomic distribution of dipoles. Nevertheless, our database provides a satisfactory panorama, being particularly useful when a quick assessment is desired. The difference map shown in Fig. 6 indicates that the largest deviations occur in regions associated to the peptide bonds and the carboxyl terminal group. It does not necessarily imply that the dipole moments of such groups are ill defined. The inaccuracy may be a deficiency of the dipolar simplification itself, which reduces entire functional groups to point dipoles. The map constructed using the database presents a largely negative region around the $-\text{COO}^-$ group that is concentrated between the oxygen atoms. This almost coincides with the direction of the dipole moment vector for the group. An alternative to improve the description of the electrostatic potentials could be to split the group dipole vector into bond contributions.

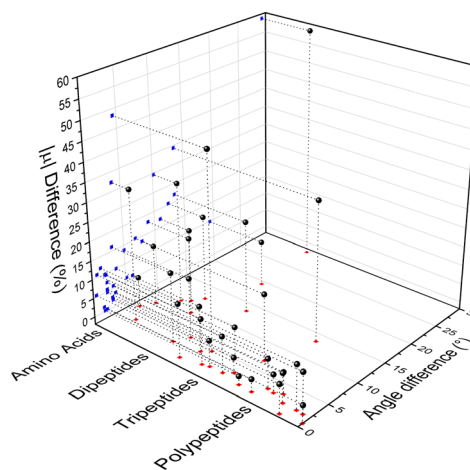


Fig. 4 Dipole moment deviation (percent), and angle differences (degrees) between quantum quantities and those obtained using the database.





Fig. 5 Comparison between quantum and database properties for the module of dipole moments (left), the polarizabilities isotropy, (middle) and anisotropy (right). All values are in atomic units.



Fig. 6 Electrostatic potential map in atomic units of the MAGE-1 fragment plotted over a 0.01 a.u. isodensity surface calculated from eqn (6) using a functional group (database) and an atomic distribution approach.

3.2 Condensed-phase properties

Among several approaches to compute the influence of the intermolecular chemical environment in the electro-optical properties, calculations under periodic boundary conditions are certainly considered the most powerful ones. However, besides being restricted to crystalline systems, the amount of electronic correlation one can introduce is still quite limited. As an alternative, finite molecular aggregates can be used to estimate the properties of condensed phases, either crystalline or non-crystalline. In some cases, this can be done by means of an electrostatic dipole interaction model (DIM),⁴⁷ in others, by quantum-mechanically calculating the charge density of the entire cluster.⁴⁸

Consideration of a condensed medium has a profound impact on the properties derived from the charge density distribution. The presence of strong hydrogen bonds may induce severe changes in both the orientation and size of polarizability tensors and dipole moments.^{33,48} Here, we explicitly considered interactions between the peptides used to build the database and a solvent, represented by water molecules.

The semi-empirical DIM method is very fast, yet capable of returning accurate results compared to quantum-mechanical calculations. The idea is to correct gas-phase properties for effects of the medium a posteriori. It is based on electrostatic

interactions between pairs of dipole moments that simulate the effective electric field felt by a molecule, an atom or a building block. In our previous work, we demonstrated that replacing atoms or functional groups, instead of entire molecules, by point dipoles is more accurate.³³ According to our implementation, the i component of the local (applied plus induced) field on the basin Ω due to all neighboring basins Λ can be written as:

$$F_i^\Omega = F_i^{\text{ext}} - \sum_{\Lambda \neq \Omega} \sum_j T_{ij}^{\Omega\Lambda} \mu_j^\Lambda \quad (7)$$

where $T_{ij}^{\Omega\Lambda}$ is the ij component of the dipole field tensor between the atoms or functional groups Ω and Λ , defined as:

$$T_{ij}^{\Omega\Lambda} = \frac{-3}{r_{\Omega\Lambda}^5} \begin{pmatrix} x_{\Omega\Lambda}^2 - \frac{r_{\Omega\Lambda}^2}{3} & x_{\Omega\Lambda}y_{\Omega\Lambda} & x_{\Omega\Lambda}z_{\Omega\Lambda} \\ x_{\Omega\Lambda}y_{\Omega\Lambda} & y_{\Omega\Lambda}^2 - \frac{r_{\Omega\Lambda}^2}{3} & y_{\Omega\Lambda}z_{\Omega\Lambda} \\ x_{\Omega\Lambda}z_{\Omega\Lambda} & y_{\Omega\Lambda}z_{\Omega\Lambda} & z_{\Omega\Lambda}^2 - \frac{r_{\Omega\Lambda}^2}{3} \end{pmatrix} \quad (8)$$

where $x_{\Omega\Lambda} = (x_\Omega - x_\Lambda)$ is the difference in the Cartesian x coordinate between the basins Ω and Λ , and $r_{\Omega\Lambda}$ is the corresponding interatomic distance. Functional-group positions are again taken at the center of charge of the group. From the total electric field and the unperturbed polarizability tensors, dipole moments can be recalculated:

$$\mu_j^{F_i}(\Omega) = \mu_j^0 + \alpha_{ij}(\Omega) \left(F_i^{\text{ext}} - \sum_{\Lambda \neq \Omega} \sum_j T_{ij}^{\Omega\Lambda} \mu_j^\Lambda \right) \quad (9)$$

Polarizabilities are obtained by finite differentiation. Cycles are very efficient on reaching the convergence criterion of 10^{-4} a.u. for the magnitude of the dipole moment.

We used glycine as a model. To ensure that the intermolecular-interaction sphere around a given central molecule is complete and that all relevant interactions are included, we explicitly added fifty eight water molecules around a glycine. This aggregate was subjected to geometry optimization using the CHARMM force field. A representation of the aggregate together with polarizability ellipsoids is given in Fig. 7. To complement our analysis, we also performed a quantum-mechanical calculation using an implicit





Fig. 7 Glycine molecule surrounded by water. Ellipsoids compare functional-group polarizabilities in the gas-phase (green/light blue) and in the condensed phase after using the DIM (purple/pink).

polarizable continuum medium (PCM). The results can be found in Table 2.

It is known that including an isotropic polarization due to the environment by means of implicit solvents may provide unreliable properties, in particular for those that depend most on the anisotropic polarization of the charge density.^{49,50} Nevertheless, both DIM and PCM showed an increment in the dipole moment with respect to the glycine molecule in isolation, about 10–30% when focusing on individual vector components. The magnitude of the dipole changes from 4.96 a.u. in the isolated situation to 6.5 a.u. when using DIM, and to 6.0 a.u. when employing PCM. The former method gives a dipole moment which deviates 0.9° from the vector direction obtained in the gas phase, whereas the latter provides a smaller deviation of 0.3°. This small reorientation is expected from PCM because it lacks directional intermolecular interactions.

Simulating the chemical environment of a molecule increases the electric field felt by it, which increases the charge-density polarization. However, it is not uncommon to observe a decrement in the molecular polarizability when moving from gas to



Fig. 8 Glycine molecular polarizabilities and dipole moments, evaluated after molecular dynamics simulation, with an overall time of 400 ps. The dashed lines represent the average values of a given quantity.

condensed phases. This is justified by a volume restriction imposed on the central molecule when it is explicitly surrounded by solvent molecules.³¹ Since PCM does not include any kind of volume restriction, we observed an increment of about 20–25% in each component of the diagonalized polarizability tensor whereas, for DIM, the components changed only slightly with respect to glycine in the gas phase. For the sake of clarity and to make sure that the obtained values of polarizabilities are representative for possible conformers of glycine, we performed additional molecular dynamic simulations to consider more than one geometry (for Cartesian coordinates and molecular geometries, the reader is referred to the ESI† pdb file). Once those geometries were obtained, we calculated molecular dipole moments and polarizabilities and compared them to other models used in this work. Graphical representation of the time evolution of dipole moments and polarizabilities is given in Fig. 8. To evaluate how reasonable the database-derived electric properties are, we used polarizabilities estimated *via* the Clausius–Mossotti equation as benchmark values, which shows the relationship between the experimentally derived dielectric constant and atomic polarizabilities. One must be however aware that the above-mentioned relationship works best for gases and is only approximately true for liquids or solids, particularly if the dielectric constant is large. In fact, to the best of our knowledge, there is no experimental polarizability data available, in particular for amino-acid systems

Table 2 Dipole moments and polarizability components for glycine calculated in isolation, using both explicit (DIM) and implicit (PCM) solvent models, and experimentally determined quantities. MD refers to the average value after molecular dynamics, and the respective standard deviation. All values are in atomic units

	μ_x	μ_y	μ_z	$ \mu $	α_{11}	α_{22}	α_{33}	α_{iso}
Isolated	2.78	−1.42	−3.85	4.96	34.5	45.8	53.9	44.7
DIM	3.55	−1.91	−5.05	6.47	35.7	44.9	53.1	44.6
PCM	3.36	−1.74	−4.70	6.02	43.3	56.7	64.1	54.7
MD				6.28 ± 0.31	34.4 ± 1.4	47.6 ± 1.4	53.1 ± 2.0	44.9 ± 0.4
EXP				6.2 ± 0.1^a				44.3 ± 0.6^b

^a 25 °C, 1 M, pH 6.7 using Kirkwoods dielectric mixture theory.⁵² ^b From molar refraction measured in aqueous solution at $\lambda = 589$ nm and 25 °C using the Clausius–Mossotti equation.⁵³



which would allow us to accurately compare the full anisotropy of the polarizability tensor. Thus our goal here is rather to make sure that the database delivers values of the same order of magnitude, rather than benchmark against exact numbers. The results are summarized in Table 2. The values referring to MD simulation agree well with both experimental values and the frozen local-minima geometry obtained by DIM, proving that for small systems such as glycine, the conformational changes are not significantly affecting the discussed electric properties.

4 Conclusions

In this work, we proposed a database to quickly and accurately estimate dipole moments and distributed polarizabilities of polypeptides. The database entries consist of twenty five unique building blocks. We discussed two applications. The first one is related to the reconstruction of electrostatic potential maps for biomolecules. The second is associated with the determination of properties for the condensed phase which includes water molecules within a region around an amino acid. A dipole interaction model was employed to take into account the polarization effects due to solvent molecules. Although we have used medium-size peptides as precursors for generating the database, it was proved to be useful to quickly obtain dipole moments and polarizabilities for smaller molecules, having a slightly different chemical nature, and larger peptides (up to nine amino acid residues). The next step of our work is to increase the variety of database entries allowing estimation of the properties of larger peptides and proteins. Certainly that will require inclusion of dynamic solvation medium, to better describe the influence of the solvent on the electric properties of the studied system. Furthermore, since our approach delivers currently only static polarizability tensors, neglecting the effects from oscillating electric fields, our intention is to eventually introduce frequency dependent polarizabilities, following for example the procedure already present in the literature.⁵¹ This approach will for sure be more beneficial for benchmarking database-derived polarizabilities against experimentally available parameters.

Conflicts of interest

There are no conflicts to declare.

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Notes and references

- 1 S. Shabahang, S. Kim and S.-H. Yun, *Adv. Funct. Mater.*, 2018, **28**, 1706635.
- 2 D. Shan, E. Gerhard, C. Zhang, J. W. Tierney, D. Xie, Z. Liu and J. Yang, *Bioact. Mater.*, 2018, **3**, 434–445.
- 3 B. D. Ratner, *J. Biomed. Mater. Res.*, 1993, **27**, 837–850.
- 4 A. D. Costache, J. Ghosh, D. D. Knight and J. Kohn, *Adv. Eng. Mater.*, 2010, **12**, B17.
- 5 J. Applequist, *Acc. Chem. Res.*, 1977, **10**, 79–85.
- 6 K. J. Miller, *J. Am. Chem. Soc.*, 1990, **112**, 8533–8542.
- 7 T. Zhou and C. E. Dykstra, *J. Phys. Chem. A*, 2000, **104**, 2204–2210.
- 8 C. S. Ewig, M. Waldman and J. R. Maple, *J. Phys. Chem. A*, 2002, **106**, 326–334.
- 9 M. in het Panhuis, R. W. Munn and P. L. A. Popelier, *J. Chem. Phys.*, 2004, **120**, 11479–11486.
- 10 D. Geldof, A. Krishtal, P. Geerlings and C. V. Alsenoy, *J. Phys. Chem. A*, 2011, **115**, 13096–13103.
- 11 N. Otero, C. V. Alsenoy, C. Pouchan and P. Karamanis, *J. Comput. Chem.*, 2015, **36**, 1831–1843.
- 12 Y. Mei, A. C. Simmonett, F. C. Pickard, R. A. DiStasio, B. R. Brooks and Y. Shao, *J. Phys. Chem. A*, 2015, **119**, 5865–5882.
- 13 K. E. Laidig and R. F. W. Bader, *J. Chem. Phys.*, 1990, **93**, 7213–7224.
- 14 T. A. Keith, in *The Quantum Theory of Atoms in Molecules*, ed. C. F. Matta and R. J. Boyd, Wiley-VCH, Weinheim, 2007, ch. 3, pp. 61–94.
- 15 A. Krawczuk, D. Perez and P. Macchi, *J. Appl. Crystallogr.*, 2014, **47**, 1452–1458.
- 16 A. J. Stone and M. Alderton, *Mol. Phys.*, 2002, **100**, 221–233.
- 17 J. G. Angyan, G. Jansen, M. Loos, C. Hättig and B. A. Heß, *Chem. Phys. Lett.*, 1994, **219**, 267–273.
- 18 J. G. Angyan, C. Chipot, F. Dehey, C. Hättig and C. Millot, *J. Comput. Chem.*, 2003, **24**, 997–1008.
- 19 G. Jansen, C. Hättig, B. A. Heß and J. G. Angyan, *Mol. Phys.*, 1996, **88**, 69–92.
- 20 C. Hättig, G. Jansen, B. A. Heß and J. G. Angyan, *Mol. Phys.*, 1997, **91**, 145–160.
- 21 P. M. Dominiak, A. Volkov, A. P. Dominiak, K. N. Jarzemska and P. Coppens, *Acta Crystallogr., Sect. D: Biol. Crystallogr.*, 2009, **65**, 485–499.
- 22 B. Dittrich, T. Koritsanszky and P. Luger, *Angew. Chem., Int. Ed.*, 2004, **43**, 2718–2721.
- 23 C. F. Matta and R. W. F. Bader, *Proteins: Struct., Funct., Genet.*, 2000, **40**, 310–329.
- 24 M. Ernst, L. H. R. Dos Santos, A. Krawczuk and P. Macchi, in *Understanding Intermolecular Interactions in the Solid State: Approaches and Techniques*, ed. D. Chopra, The Royal Society of Chemistry, London, 2019, ch. 7, pp. 211–242.
- 25 A. Kumar, P. Pandey, P. Chatterjee and A. D. MacKerell Jr., *J. Chem. Theory Comput.*, 2022, **18**, 1711–1725.
- 26 C. M. Baker, *Wiley Interdiscip. Rev.: Comput. Mol. Sci.*, 2015, **5**, 241–254.
- 27 D. A. Dunmur, *Mol. Phys.*, 1972, **23**, 109–115.
- 28 H. Reis, S. Raptis, M. G. Papadopoulos, R. H. C. Jansen, D. N. Theodorou and R. W. Munn, *Theor. Chem. Acc.*, 1998, **99**, 384–390.
- 29 M. A. Spackman, P. Munshi and D. Jayatilaka, *Chem. Phys. Lett.*, 2007, **443**, 87–91.
- 30 A. M. Mkadmh, A. Hinchliffe and F. M. Abu-Awwad, *THEOCHEM*, 2009, **901**, 9–17.



- 31 L. H. R. Dos Santos, A. Krawczuk and P. Macchi, *J. Phys. Chem. A*, 2015, **119**, 3285–3298.
- 32 M. Guillaume and B. Champagne, *Phys. Chem. Chem. Phys.*, 2005, **7**, 3284–3289.
- 33 R. F. Ligorio, A. Krawczuk and L. H. R. Santos, *J. Phys. Chem. A*, 2020, **124**, 10008–10018.
- 34 R. F. Ligorio, A. Krawczuk and L. H. R. Santos, *J. Phys. Chem. A*, 2021, **125**, 4152–4159.
- 35 J. C. Cheville and P. C. Roche, *Mod. Pathol.*, 1999, **12**, 974–978.
- 36 B. R. Brooks, C. L. B. III, A. D. Mackerell, L. Nilsson, R. J. Petrella, B. Roux, Y. Won, G. Archontis, C. Bartels, S. Boresch, A. Cafflich, L. Caves, Q. Cui, A. R. Dinner, M. Feig, S. Fischer, J. Gao, M. Hodoseck, W. Im, K. Kuczera, T. Lazaridis, J. Ma, V. Ovchinnikov, E. Paci, R. W. Pastor, C. B. Post, J. Z. Pu, M. Schaefer, B. Tidor, R. M. Venable, H. L. Woodcock, X. Wu, W. Yang, D. M. York and M. Karplus, *J. Comput. Chem.*, 2009, **30**(10), 1545–1614.
- 37 J. R. Hammond, N. Govind, K. Kowalski, J. Autschbach and S. S. Xantheas, *J. Chem. Phys.*, 2009, **131**, 214103.
- 38 D. Hait and M. Head-Gordon, *Phys. Chem. Chem. Phys.*, 2018, **20**, 19800.
- 39 Y. Zhao and D. G. Truhlar, *Theor. Chem. Acc.*, 2008, **120**, 215–241.
- 40 M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, G. A. Petersson, H. Nakatsuji, X. Li, M. Caricato, A. V. Marenich, J. Bloino, B. G. Janesko, R. Gomperts, B. Mennucci, H. P. Hratchian, J. V. Ortiz, A. F. Izmaylov, J. L. Sonnenberg, D. Williams-Young, F. Ding, F. Lipparini, F. Egidi, J. Goings, B. Peng, A. Petrone, T. Henderson, D. Ranasinghe, V. G. Zakrzewski, J. Gao, N. Rega, G. Zheng, W. Liang, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, K. Throssell, J. A. Montgomery, Jr., J. E. Peralta, F. Ogliaro, M. J. Bearpark, J. J. Heyd, E. N. Brothers, K. N. Kudin, V. N. Staroverov, T. A. Keith, R. Kobayashi, J. Normand, K. Raghavachari, A. P. Rendell, J. C. Burant, S. S. Iyengar, J. Tomasi, M. Cossi, J. M. Millam, M. Klene, C. Adamo, R. Cammi, J. W. Ochterski, R. L. Martin, K. Morokuma, O. Farkas, J. B. Foresman and D. J. Fox, *Gaussian 16, Revision C.01*, Gaussian, Inc., Wallingford CT, 2016.
- 41 T. A. Keith, *AIMAll (Version 19.10.12)*, TK Gristmill Software, Overland Park KS, USA, 2019, aim.tkgristmill.com.
- 42 S. Jo, T. Kim, V. Iyer and W. Im, *J. Comput. Chem.*, 2008, **29**, 1859–1865.
- 43 S. Nose, *Mol. Phys.*, 1986, **57**, 187–191.
- 44 W. G. Hoover, *Phys. Rev. A: At., Mol., Opt. Phys.*, 1985, **31**, 1965–1967.
- 45 H. J. C. Berendsen, J. P. M. Postma, W. F. van Gunsteren, A. DiNola and J. R. Haak, *J. Chem. Phys.*, 1984, **81**, 3684.
- 46 A. Jabluszewska, A. Krawczuk, L. H. R. Dos Santos and P. Macchi, *ChemPhysChem*, 2020, **21**, 2155–2165.
- 47 L. Silberstein, *Philos. Mag.*, 1917, **33**, 92–128.
- 48 L. H. R. Dos Santos and P. Macchi, *Crystals*, 2016, **6**, 43.
- 49 J. Zhang, H. Zhang, T. Wu, Q. Wang and D. van der Spoel, *J. Chem. Theory Comput.*, 2017, **13**, 1034–1043.
- 50 J. R. Spaeth, I. G. Kevrekidis and A. Z. Panagiotopoulos, *J. Chem. Phys.*, 2011, **134**, 164902.
- 51 T. Seidler, A. Krawczuk, B. Champagne and K. Stadnicka, *J. Phys. Chem. C*, 2016, **120**, 4481–4494.
- 52 M. W. Aaron and E. H. Grant, *Trans. Faraday Soc.*, 1963, **59**, 85–89.
- 53 T. L. McMeekin, M. L. Groves and N. J. Hipp, *Refractive indices of amino acids, proteins, and related substances*, in *Amino Acids and Serum Proteins*, ed. J. Stekol, American Chemical Society, Washington DC, 1964, pp. 54–66.

