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# Continuous flow-ultrasonic synergy in click reactions for the synthesis of novel 1,2,3-triazolyl appended 4,5-unsaturated L-ascorbic acid derivatives†

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A combination of flow chemistry and batch-based synthetic procedures has been successfully applied to the assembly of novel 4,5-unsaturated L-ascorbic acid series **6a–6n** with diverse C-6-substituted 1,2,3-triazole moiety. We report herein the first Cu(I)-catalyzed 1,3-dipolar cycloaddition of azido functionalized L-ascorbic acid derivative and selected alkynes to provide target 1,2,3-triazolyl appended 4,5-didehydro-5,6-dideoxy-L-ascorbic acid library **6a–6n** under both micro-flow and batch conditions. Implementation of ultrasound with flow chemistry accelerated hour-scale reaction conditions in batch to the minute range in micro-flow device and considerably improved the yields for the flow syntheses of **6a–6n**. Moreover, the synergistic use of microreactor technology and ultrasonic irradiation highlights the sustainable eco-friendly aspect of utilized method.

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## Introduction

L-Ascorbic acid (AA) or vitamin C is known as a ubiquitous carbohydrate of vital importance in the living beings. Since the discovery of vitamin C, the number of its known biological functions is continually expanding.<sup>1</sup> Thus, it plays an important role in a number of cellular processes, including collagen synthesis, cellular oxidation, and as a cofactor in several important hydroxylation reactions.<sup>2</sup> AA is one of the most important biomolecules, which protects cellular components against oxidative damage caused by free radicals and oxidants that are involved in the development of a multitude of chronic diseases. Furthermore, some L-ascorbic acid derivatives showed to be effective antitumoral agents,<sup>3</sup> immunostimulants<sup>4</sup> and protecting agents against peroxidation of lipids of the biomembranes.<sup>5</sup> In previous study we demonstrated that some pyrimidine and purine derivatives of 4,5-didehydro-5,6-dideoxy- and 6-deoxy-L-ascorbic acid exhibited selective and efficient cytostatic activities and displayed antiviral activities against varicella zoster virus (TK<sup>+</sup>VZV and TK<sup>−</sup>VZV) and cytomegalovirus (CMV).<sup>6</sup> Structurally related  $\gamma$ -(triazolyl ethylidene)

butenolides exhibited most potent activity against breast cancer cells and preliminary due to reactive oxygen species (ROS) generation, subsequent activation of p38, leading to apoptosis and inhibition of cancer cells.<sup>7</sup> Furthermore, several preclinical and clinical trials on vitamin C have also supported its promising efficacy against a number of cancer types and low toxicity of high-dose vitamin C, that has been demonstrated in anti-cancer therapy.<sup>8</sup>

On the other hand, 1,2,3-triazoles have been the nuclei of choice in recent years because of their excellent pharmacokinetic characteristics, favourable safety profile, latent ability for the formation of hydrogen bonds, moderate dipole character, rigidity and stability under *in vivo* conditions.<sup>9</sup> Therefore, triazole chemistry has recently received significant impulse from the pioneering work of Sharpless and Meldal on Cu(I)-catalyzed alkyne-azide cycloaddition (CuAAC).<sup>10</sup> The CuAAC reaction has become the cream of the crop for click chemistry and has been broadly used in many areas of modern chemistry, such as polymer and materials sciences,<sup>11</sup> supramolecular chemistry,<sup>12</sup> bioconjugation,<sup>13</sup> and combinatorial chemistry.<sup>14</sup> This reaction is also significant from the viewpoint of drug discovery,<sup>15</sup> because 1,2,3-triazoles possess a wide range of biological properties, such as anticancer,<sup>16</sup> antiviral,<sup>17</sup> antibacterial<sup>18</sup> and antifungal activities.<sup>19</sup> Therefore, the 1,2,3-triazole moiety is extensively used in medicinal chemistry as a pharmacophore to modify known bioactive molecules and to potentiate their biological activities.<sup>9a,20</sup> A growing effort has been devoted to the employment of click chemistry for the synthesis of functionally diverse

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† Electronic supplementary information (ESI) available: <sup>1</sup>H and <sup>13</sup>C NMR spectra of **6a–6n**, and 2D NMR spectra of **6a** and **6n**, HPLC chromatograms for micro-flow reactions of **6a–6n**. See DOI: 10.1039/c6ra25244c

carbohydrate derivatives as bioactive compounds and fluorescent glycoprobes.<sup>21</sup>

Although click reactions are traditionally conducted in batch process, the progress toward increased sustainability that requires novel approaches with reduced environmental impact opened up in recent years continuous-flow as a novel alternative to conventional batch-based synthesis.<sup>22</sup> Flow chemistry technique performs a chemical reaction in a continuously flowing stream in a network of interconnecting tubes. The reagents coming from different tubes are mixed together in the reactor to start the chemical reaction. Due to its many advantages over batch reaction processes, flow chemistry has become an emerging field in organic synthesis, and has recently been employed to enhance the selectivity, safety, and efficiency of chemical syntheses, including the rapid synthesis of natural products and pharmaceuticals.<sup>23</sup> Microreactor technology has the potential to revolutionize the pharmaceutical industry because the number of potential drug candidates that can be prepared and screened can be considerably increased, hence the likelihood of developing new drugs is considerably enhanced.<sup>24</sup> Besides being more economical and environmentally friendly, microreactor technology (MRT) is even more advantageous than the macroscopic batch reactor method, mainly due to the high surface/volume ratio, reduced utilization of reagents, and also better control over mass as well as heat transfer.<sup>25</sup> Many previously challenging or hazardous reactions can now be safely and efficiently conducted in flow.<sup>26</sup> In addition, flow chemical synthesis can be performed under non-classical reaction conditions, such as applying ultrasound and microwave irradiation, to significantly improve efficiency of reactions.<sup>27</sup> It was demonstrated that the use of ultrasound, an alternative green source of energy, to promote chemical reactions allowed a decrease in the reaction time and side reactions. On the other hand, with a well-defined inner micro-structure, a microreactor provides an ideal environment to investigate and control the acoustic cavitation process, which is the main mechanism causing sonochemical effects.<sup>28</sup> Although the advantages of the continuous flow reactor technology for the synthesis of 1,4-disubstituted 1,2,3-triazoles over their batch reaction counterparts have been demonstrated recently<sup>29</sup> further improvement in this direction is still mandatory. Despite the increasingly importance of flow chemistry in discovery of biologically active molecules, to our knowledge no studies are hitherto performed on the use of continuous flow to provide L-ascorbic acid and its derivatives, as molecules with unique biological properties. Realizing the benefits of flow chemistry and in continuation of our study towards the synthesis of 1,2,3-triazole-containing heterocycle pharmacophores,<sup>15b,30</sup> our aim was to improve the practical applicability of Cu(I)-catalyzed azide-alkyne 1,3-dipolar cycloaddition (CuAAC) to provide diverse C-6-substituted 1,2,3-triazolyl 4,5-unsaturated L-ascorbic acid derivatives **6a–6n** using innovative enabling ultrasound (US) and continuous flow hybrid system (Fig. 1).

Herein we report classical multistep synthesis to afford azido functionalized L-ascorbic acid for clicking with terminal alkynes

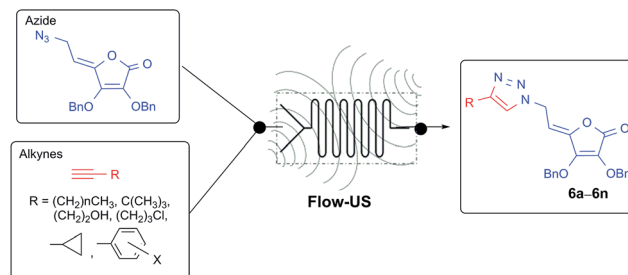


Fig. 1 Synthesis of C-6-substituted 1,2,3-triazolyl 4,5-unsaturated L-ascorbic acid derivatives **6a–6n** utilizing ultrasonic (US)-assisted continuous flow in click reactions.

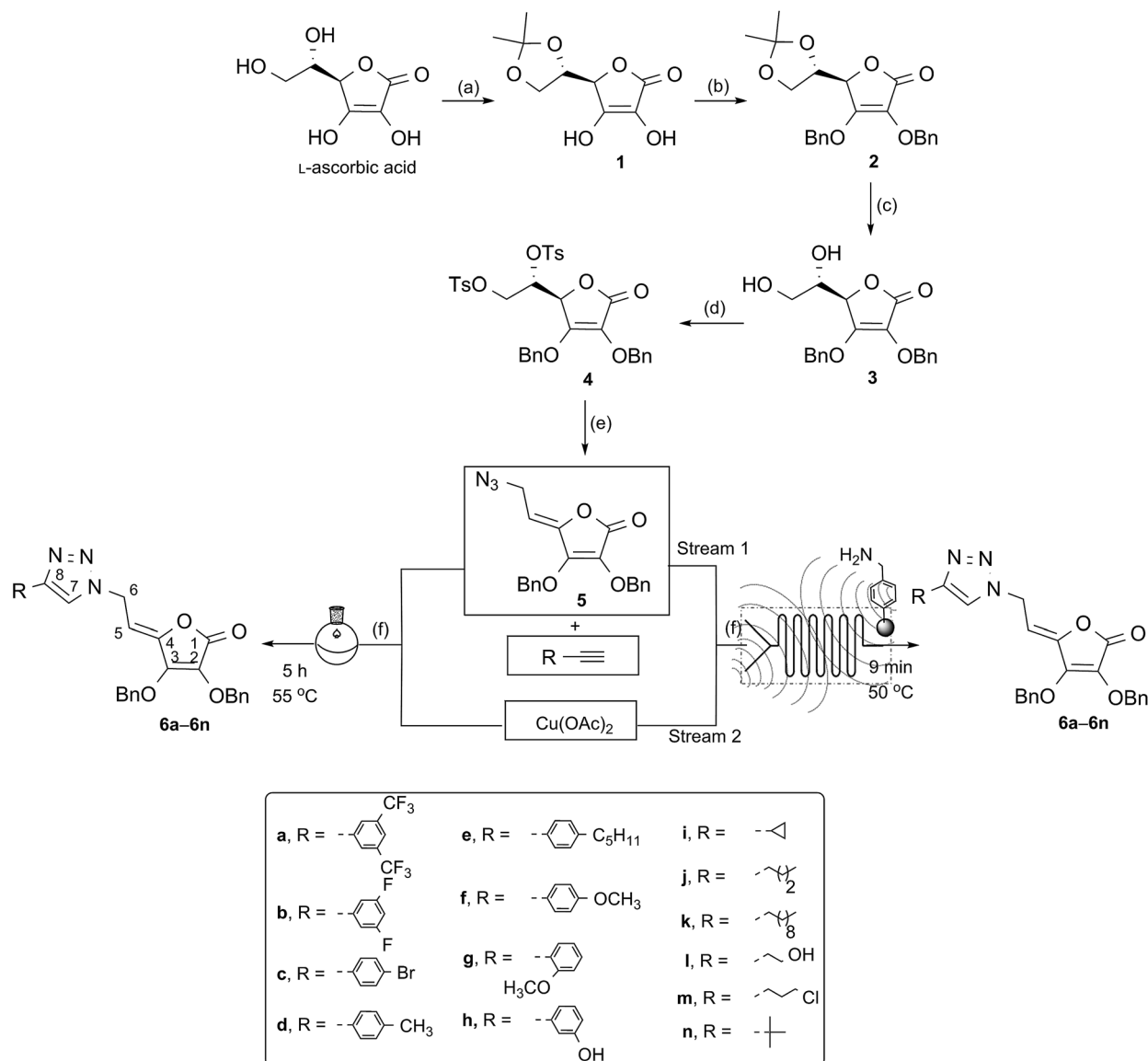
and investigate both continuous flow-ultrasound and batch approach for CuAAC reaction.

## Results and discussion

### Synthesis of the 1,2,3-triazolyl appended 4,5-unsaturated L-ascorbic acid library under batch conditions

In order to synthesize C-6-substituted 1,2,3-triazolyl 4,5-dihydro-5,6-dideoxy-L-ascorbic acid derivatives, synthetic transformations of L-ascorbic acid that included selective protection and deprotection of 2,3- and 5,6-dihydroxyl groups were performed as described in the literature.<sup>31</sup> Thus, 5,6,*O*-isopropylidene-L-ascorbic acid (**1**) was synthesized *via* reaction of L-ascorbic acid with acetyl chloride, while the remaining C-2 and C-3 hydroxyl groups of the lactone ring were protected by reaction of compound **1** with benzyl chloride in the presence of potassium carbonate to afford compound **2**. To apply molecular hybridization by coupling of L-ascorbic acid derivative with 1,2,3-triazole ring, we used the strategy that involved the functionalization of L-ascorbic acid with an azide substituent at the C-6 (Scheme 1). Therefore, isopropylidene protection in **2** was removed in acidic condition to give compound **3** with free C-5 and C-6 hydroxyl functionalities, which were subsequently converted to tosylate, as a good leaving group, using *p*-toluenesulfonyl chloride (*p*-TsCl) in pyridine. The 5,6,*O*-ditosyl derivative of L-ascorbic acid (**4**) was used as the parent compound for the preparation of the C-6 azido derivative of 4,5-unsaturated L-ascorbic acid **5**. Compound **5** was successfully synthesized from compound (**4**) through elimination to an exocyclic allylic tosylate which through a subsequent S<sub>N</sub>2 reaction with sodium azide (NaN<sub>3</sub>) directly afforded azide **5** in a moderate yield of 51%. 1,2,3-Triazolyl appended 4,5-unsaturated L-ascorbic acid derivatives **6a–6n** with various substituents at C-8 of the triazole ring were obtained through the copper mediated 1,3-dipolar cycloaddition of C-6 azido L-ascorbic acid derivative **5** and terminal alkynes (Scheme 1). In order to assess the influence of structural modifications at C-8 of the triazole on activity, selected alkynes **a–n** were used in click reaction with the azide **5**. The synthesis of **6a–6n** was performed using Cu(OAc)<sub>2</sub>, which enabled the reduction of Cu(II) to Cu(I) ions through reaction with methanol.<sup>32</sup> (*Z*)-Configuration around the C-4=C-5 double bond was verified by the ROESY spectra which displayed the cross peak between the *O*-methylene at C-3 and methine C-5 protons. In the hypothetical (*E*)-configuration, where the spatial





**Scheme 1** Synthesis of novel 1,4-disubstituted-1,2,3-triazole L-ascorbic acid derivatives by conventional batch and flow-ultrasound conditions. Reagents and conditions: (a) acetyl chloride, acetone; (b) benzyl chloride,  $K_2CO_3$ ; (c) 50% acetic acid,  $CH_3OH$ ; (d) *p*-TsCl,  $CH_2Cl_2$ , pyridine, 0 °C; (e)  $NaN_3$ , acetonitrile, 80 °C; (f)  $CH_3OH$ .

distance between C-3 and C-5 protons is too large, mentioned interactions would not be expected. These findings are in agreement with the determined (*Z*)-configuration of closely related pyrimidine and purine derivatives of L-ascorbic acids.<sup>33</sup>

### Synthesis of the 1,2,3-triazolyl appended 4,5-unsaturated L-ascorbic acid library under continuous flow process

In order to investigate the advantages of flow chemistry over batch methods, the Cu(I)-catalyzed click reactions of azido functionalized L-ascorbic acid derivative 5 and alkynes were performed in a microreactor system. Two enabling technologies, micro-flow chemistry and ultrasound irradiation (US) have been combined and compared to batch mode synthesis. To combine ultrasound irradiation and microfluidic system, we applied a simple and inexpensive technique, which was at the

same time safe and efficient, by directly immersing the micro-reactor in the ultrasonic bath.

For the optimization of reaction parameters, click chemistry of azido derivative of L-ascorbic acid 5 and 3,5-di(trifluoromethyl)phenylacetylene and 3-butyne-1-ol were chosen as test reactions to obtain 1,4-disubstituted 1,2,3-triazoles 6a and 6l, respectively, with the highest yield (Scheme 1 and Table 1, entries 1–4). The highest conversion of azide 5 to 1,2,3-triazole products was observed under ultrasound-assisted micro-flow using the 0.1 M solution of the starting azide and alkyne, flow rate of  $0.5 \mu L \min^{-1}$  and a slightly elevated temperature of 50 °C (Table 1, entry 3). The residence time for the compounds prepared under these reaction conditions amounted 9 min. Higher reaction temperatures typically resulted in the formation of unidentified side-products.



**Table 1** The effect of the flow rate and ultrasound irradiation on the reactions between azido derivative of L-ascorbic acid **5** and corresponding alkynes in a flow microreactor<sup>a</sup>

| Entry | Conditions                      | Yield <sup>b</sup> [%] |                 |
|-------|---------------------------------|------------------------|-----------------|
|       |                                 | Compd <b>6a</b>        | Compd <b>6l</b> |
| 1     | 1 $\mu\text{L min}^{-1}$        | 35                     | 51              |
| 2     | 0.5 $\mu\text{L min}^{-1}$      | 50                     | 72              |
| 3     | 0.5 $\mu\text{L min}^{-1}$ , US | 99                     | 83              |
| 4     | 0.2 $\mu\text{L min}^{-1}$      | Clogging               | Clogging        |

<sup>a</sup> 0.1 M solution of substrate, 1 equiv. alkyne and temperature of 50 °C.

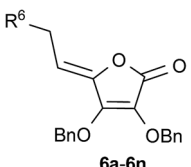
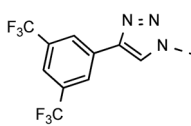
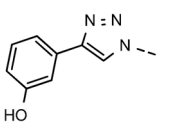
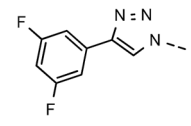
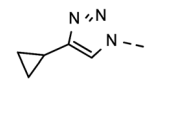
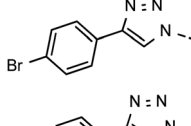
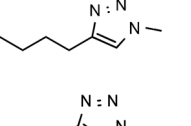
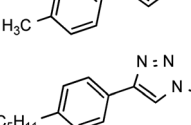
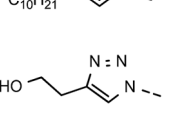
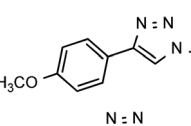
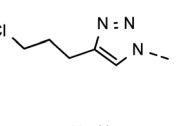
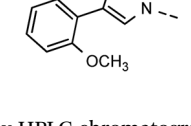
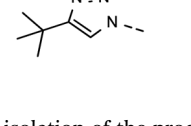
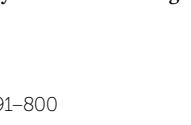
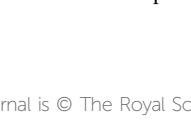
<sup>b</sup> Yields were determined by HPLC chromatography.

Higher concentration of the starting compounds and lower flow rate (0.2  $\mu\text{L min}^{-1}$ ) led to the precipitation of the triazole product in the flow microreactor. However, when we attempted

to decrease the residence time by increasing the flow rate to 1  $\mu\text{L min}^{-1}$  the yields of **6a** and **6l** decreased.

As we can observe from Table 2, the yields of both flow-ultrasound and batch-based reactions were dependent on the C-8 substituents at 1,2,3-triazole scaffold. Interestingly, the effects of C-8 substituents in continuous micro-flow and batch syntheses were similar. Notably, all tested reactions using combination of micro-flow and ultrasound method were more successful than those under classical synthetic conditions. Among the aromatic alkynes, type of the substituents and their position on the phenyl ring were nicely tolerated in click reactions. In general, electron-donating groups gave target 1,2,3-triazolyl appended L-ascorbic acid derivatives (**6e–6h**) under micro-flow in excellent yields. Only exception was the click reaction with alkyne containing *p*-methylphenyl moiety, that afforded 4-tolyl-1,2,3-triazolyl appended L-ascorbic acid (**6d**) using both micro-flow and batch conditions in the lowest yields

**Table 2** A comparison of continuous micro-flow and batch approaches for the synthesis of 1,2,3-triazolyl appended 4,5-unsaturated L-ascorbic acid library **6a–6n**

|  |           |                                                                                     |                    |                   |       |           |                                                                                       |                    |                   |
|-----------------------------------------------------------------------------------|-----------|-------------------------------------------------------------------------------------|--------------------|-------------------|-------|-----------|---------------------------------------------------------------------------------------|--------------------|-------------------|
| Entry                                                                             | Compd     | R <sup>6</sup>                                                                      | Yield [%]          |                   | Entry | Compd     | R <sup>6</sup>                                                                        | Yield [%]          |                   |
|                                                                                   |           |                                                                                     | Batch <sup>a</sup> | Flow <sup>b</sup> |       |           |                                                                                       | Batch <sup>a</sup> | Flow <sup>b</sup> |
| 1                                                                                 | <b>6a</b> |  | 81                 | 99                | 8     | <b>6h</b> |  | 79                 | 99                |
| 2                                                                                 | <b>6b</b> |  | 62                 | 89                | 9     | <b>6i</b> |  | 70                 | 76                |
| 3                                                                                 | <b>6c</b> |  | 49                 | 63                | 10    | <b>6j</b> |  | 64                 | 74                |
| 4                                                                                 | <b>6d</b> |  | 20                 | 45                | 11    | <b>6k</b> |  | 83                 | 98                |
| 5                                                                                 | <b>6e</b> |  | 46                 | 88                | 12    | <b>6l</b> |  | 61                 | 83                |
| 6                                                                                 | <b>6f</b> |  | 49                 | 98                | 13    | <b>6m</b> |  | 36                 | 86                |
| 7                                                                                 | <b>6g</b> |  | 82                 | 94                | 14    | <b>6n</b> |  | 26                 | 62                |

<sup>a</sup> Yield was determined by HPLC chromatograms of the reaction mixture. <sup>b</sup> Yields were determined by isolation of the product.



of 45% and 20%, respectively. While the position of the electron-donating substituents did not affect the CuAAC reaction in flow method, *o*- and *m*-substituted electron-donating group significantly improved the reactions (**6g** and **6h**) in comparison to *p*-substituted phenyl (**6e** and **6f**) using classical synthetic approach. From the halogen and trifluoromethyl, as electron-withdrawing substituents on the phenyl ring in selected alkynes, we can note that *p*-bromo substituent caused the moderate yield in the synthesis of the desired 4-(*p*-bromophenyl)-1,2,3-triazole appended L-ascorbic acid (**6c**). In contrast to that, 1,3-dipolar reaction with dipolarophile containing stronger electron-withdrawing fluorine and trifluoromethyl substituents at 3 and 5 position of the phenyl moiety proceeded under micro-flow with excellent yields (**6a** and **6b**). From the aliphatic alkynes, we found that unbranched alkynes afforded 1,2,3-triazolyl appended L-ascorbic acid (**6j–6m**) with side chains at C-8 of triazole in higher yields than branched alkyne (**6n**). Thus, long-chain dodecyne was successfully applied, as a dipolarophile in CuAAC reaction, affording the 4-decyl-1,2,3-triazole appended L-ascorbic acid (**6k**) in the highest yields of 98% and 83% under micro-flow and batch conditions, respectively. Overall, it was demonstrated that the ultrasonic-assisted flow method applied in click reactions to provide target 1,2,3-triazolyl appended 4,5-unsaturated L-ascorbic acid derivatives gave improved yields and considerably shortened the reaction time of click reactions. Thereby, the applied methodology demonstrated to be a more efficient and sustainable approach than the batch-based synthesis.

## Conclusions

We have performed the multistep synthesis of the 6-azido derivative of L-ascorbic acid **5** as a precursor for Cu(I)-catalyzed 1,3-dipolar cycloaddition (CuAAC) with alkynes. Click reactions of the azide **5** and selected terminal alkynes were subsequently carried out in the presence of Cu(OAc)<sub>2</sub>, that was employed as a catalyst to generate the required Cu(I)-ions *in situ*, to provide the novel C-6-substituted 1,2,3-triazolyl 4,5-didehydro-5,6-dideoxy-L-ascorbic acid series **6a–6n**. (*Z*)-Configuration across the C-4=C-5 double bond in **6a–6n** was deduced from connectivities in ROESY spectra. To evaluate the potential benefits of continuous micro-flow on CuAAC reactions, the batch conditions of click chemistry were successfully translated into a flow regime. Moreover, for the environmentally benign synthesis of **6a–6n** two enabling technologies, ultrasound irradiation and flow microreactors, have been combined and compared to batch mode synthesis. The CuAAC reactions leading to triazole–L-ascorbic acid hybrids **6a–6n** were greatly facilitated through the beneficial features of continuous micro-flow and ultrasound. Namely, reduced reaction time from hour-scale reaction to minute range and significantly improved yields in flow click reactions demonstrated the excellent synthetic capabilities of the applied method. Finally, we may conclude that translating batch click chemistry to ultrasound-assisted micro-flow processes in the present study highlights the successful advancement of click chemistry to generate new hybrids with L-ascorbic acid and 1,2,3-triazole units. Taking into

account the potential synergy of biological activity of 1,2,3-triazole and L-ascorbic acid, we anticipate that L-ascorbic acid and 1,2,3-triazole pharmacophores if linked together would generate novel molecular entities which are likely to exhibit promising biological properties. Therefore, further antitumoral evaluations of described compounds are envisaged and will be reported on in due course.

## Experimental part

### General

Melting points (uncorrected) were determined with a Kofler micro hot-stage (Reichert, Wien, Austria). Precoated Merck (Darmstadt, Germany) silica gel 60F-254 plates were used for thin layer chromatography and the spots were detected under UV light (254 nm). Column chromatography was performed using Fluka (Buchs, Switzerland) silica gel (0.063–0.2 mm); glass columns were slurry-packed under gravity. Compounds **6a–6n** were additionally filtered through a short column of Merck (Darmstadt, Germany) aluminium oxide (Al<sub>2</sub>O<sub>3</sub>, 0.063–0.2 mm) in order to scavenge the copper residues from the click reactions.

All continuous flow reaction were performed using tubular glass microreactor (length: width: depth = 330 mm: 250 μm: 50 μm with the internal volume of 6 μL). Two syringe pumps (PHD 4400 Syringe Pump Series, Harvard Apparatus, USA) equipped with high pressure stainless steel syringes (8 cm<sup>3</sup>, Harvard Apparatus, USA) were used to feed substrates to the inlet streams of microreactor. Total flow velocity ( $\Phi = 0.2\text{--}2\text{ }\mu\text{L min}^{-1}$ ) was altered and the influence on the reaction yield was monitored. When the reaction was performed in a microreactor in combination with ultrasound, microreactor was completely emerged in to an ultrasonic bath (Bandelin Sonorex, Germany) under the continuous frequency of 35 kHz and temperature of 50 °C. The samples were analyzed by high performance liquid chromatography on a Prominence HPLC system (Shimadzu, Germany) equipped with a LC-20AT pump, CBM-20A system controller, DGU-20A5 degasser, SPD-M20A photodiode array detector using an Agilent Eclipse XDB-C18 reverse-phase analytical column (4.6 × 150 mm). UV absorption was monitored at 254 nm wavelength. Water (A: 99.9% H<sub>2</sub>O, 0.1% formic acid) and acetonitrile (B: 475 mL acetonitrile, 50 mL H<sub>2</sub>O, 0.5 mL formic acid) were used for elution at a flow rate of 1 mL min<sup>−1</sup>. For the analysis of the reaction mixtures **6a–6n**, a gradient method starting with 40% B within 20 min, increasing to 60% B within next 25 min and then to 90% B within 25 min (total run time 70 min) and a gradient method starting with 40% B within 28 min, increasing to 60% B within next 32 min and then to 90% B within 5 min (total run time 65 min) were used. Elemental analyses were performed in the Central Analytic Service, Ruđer Bošković Institute, Zagreb. All elemental compositions were within the 0.4% of the calculated values.

<sup>1</sup>H and <sup>13</sup>C NMR spectra were acquired on a Bruker 300 and 600 MHz NMR spectrometer (Bruker Biospin, Rheinstetten, Germany). All data were recorded in DMSO-*d*<sub>6</sub> at 298 K. Chemical shifts were referenced to the residual solvent signal of DMSO at  $\delta$  2.50 ppm for <sup>1</sup>H and  $\delta$  39.50 ppm for <sup>13</sup>C. Individual



resonances were assigned on the basis of their chemical shifts, signal intensities, multiplicity of resonances and H–H coupling constants.

### Synthetic procedures

5,6-*O*-Isopropylidene-L-ascorbic acid (**1**), 2,3-*O*,*O*-dibenzyl-5,6-*O*-isopropylidene-L-ascorbic acid (**2**), 2,3-*O*,*O*-dibenzyl-L-ascorbic acid (**3**), 2,3-*O*,*O*-dibenzyl-4,5-didehydro-5,6-*O*-ditosyl-L-ascorbic acid (**4**) were synthesized in accordance with procedures given in the literature.<sup>29</sup>

**Preparation of (Z)-C-6-azido-2,3-*O*,*O*-dibenzyl-4,5-didehydro-5,6-dideoxy-L-ascorbic acid (**5**).**<sup>7</sup> Ditosylated L-ascorbic acid derivative **4** (2.03 g, 3.056 mmol) was refluxed in acetonitrile (55 mL) together with NaN<sub>3</sub> (1.19 g, 18.34 mmol) at 80 °C for 4 h. The reaction mixture was then allowed to stir overnight at room temperature. After purification with column chromatography on silica gel (CH<sub>2</sub>Cl<sub>2</sub>) compound **5** was isolated as yellow oil (569 mg, 51%).

<sup>1</sup>H NMR (300 MHz, DMSO) δ 7.65–7.04 (m, 10H, OCH<sub>2</sub> Ph), 5.53 (t, *J* = 7.7 Hz, 1H, H-5), 5.34 (s, 2H, OCH<sub>2</sub>Ph), 5.15 (s, 2H, OCH<sub>2</sub>Ph), 4.09 (d, *J* = 7.7 Hz, 2H, H-6) ppm. <sup>13</sup>C NMR (151 MHz, DMSO) δ 163.6 (C-1), 147.9 (C-3), 143.6 (C-4), 135.7 (Ph-q), 135.5 (Ph-q), 128.9–127.9 (OCH<sub>2</sub> Ph), 123.3 (C-2), 102.2 (C-5), 74.0 (OCH<sub>2</sub>Ph), 73.0 (OCH<sub>2</sub>Ph), 44.5 (C-6) ppm.

### General procedure for the preparation of 1,2,3-triazole-L-ascorbic acid derivatives (**6a**–**6n**) under batch conditions

The C-6 azido L-ascorbic acid derivative **5** (200–350 mg) and Cu(OAc)<sub>2</sub> (0.05 equiv.) were dissolved in CH<sub>3</sub>OH. The corresponding alkyne (1.2 equiv.) was then added and the reaction mixture was heated for 5 h at 55 °C. The reaction mixture was stirred overnight at room temperature and the solvent was evaporated *in vacuo*. The product was purified with column chromatography on silica gel (CH<sub>2</sub>Cl<sub>2</sub> : CH<sub>3</sub>OH = 200 : 1–40 : 1) and Al<sub>2</sub>O<sub>3</sub>. The oily product was triturated with *n*-hexane in order to obtain compounds **6a**–**6i**.

**Preparation of (Z)-2,3-*O*,*O*-dibenzyl-4,5-didehydro-5,6-dideoxy-6-[4-(3,5-di(trifluoromethyl)phenyl)-1,2,3-triazole-1-yl]-L-ascorbic acid (**6a**).** Compound **6a** was synthesized according to the general procedure using compound **5** (300 mg, 0.826 mmol), Cu(OAc)<sub>2</sub> (7.50 mg, 0.041 mmol) and 3,5-di(trifluoromethyl)phenylacetylene (0.18 mL, 0.991 mmol) in CH<sub>3</sub>OH (16.5 mL). Compound **6a** (401 mg, 81%, mp 89–90 °C) was isolated as white powder.

<sup>1</sup>H-NMR (300 MHz, DMSO) δ 9.00 (s, 1H, H-7), 8.51 (s, 2H, Ph-2', Ph-6'), 8.07 (s, 1H, Ph-4'), 7.44–7.32 (m, 10H, OCH<sub>2</sub> Ph), 5.72 (t, *J* = 7.7 Hz, 1H, H-5), 5.34 (2H, s, OCH<sub>2</sub>Ph), 5.34 (d, *J* = 7.6 Hz, 2H, H-6), 5.18 (s, 2H, OCH<sub>2</sub>Ph) ppm. <sup>13</sup>C NMR (151 MHz, DMSO) δ 163.5 (C-1), 147.9 (C-3), 143.9 (C-8), 143.7 (C-4), 135.6 (Ph-q), 135.3 (Ph-q), 133.2 (Ph-1'), 131.3; 131.1; 130.9; 130.7 (q, *J*<sub>CF</sub> = 33.1 Hz, Ph-3', Ph-5'), 128.8–127.9 (OCH<sub>2</sub> Ph), 125.3; 125.3 (d, *J*<sub>CF</sub> = 3.6 Hz, Ph-2', Ph-6'), 125.9; 124.1; 122.3; 120.5 (q, *J*<sub>CF</sub> = 272.7 Hz, CF), 123.5 (C-2), 123.4 (C-7), 121.2–121.0 (m, Ph-4'), 101.4 (C-5), 74.0 (OCH<sub>2</sub>Ph), 73.1 (OCH<sub>2</sub>Ph), 44.5 (C-6) ppm. Anal. calcd for C<sub>30</sub>H<sub>21</sub>F<sub>6</sub>N<sub>3</sub>O<sub>4</sub>: C, 59.90; H, 3.52; N, 6.99. Found: C, 59.98; H, 3.51; N, 6.99.

**Preparation of (Z)-2,3-*O*,*O*-dibenzyl-4,5-didehydro-5,6-dideoxy-6-[4-(3,5-difluorophenyl)-1,2,3-triazole-1-yl]-L-ascorbic acid (**6b**).** Compound **6b** was synthesized according to the general procedure using compound **5** (300 mg, 0.826 mmol), Cu(OAc)<sub>2</sub> (7.50 mg, 0.041 mmol) and 3,5-difluorophenylacetylene (0.12 mL, 0.991 mmol) in CH<sub>3</sub>OH (16.5 mL). Compound **6b** (257 mg, 62%, mp 74–76 °C) was isolated as white powder. <sup>1</sup>H-NMR (300 MHz, DMSO) δ 8.75 (s, 1H, H-7), 7.58–7.18 (m, 13H, OCH<sub>2</sub> Ph, Ph-2', Ph-4', Ph-6'), 5.69 (t, *J* = 7.2 Hz, 1H, H-5), 5.34 (s, 2H, OCH<sub>2</sub>Ph), 5.29 (d, *J* = 7.8 Hz, 2H, H-6), 5.18 (s, 2H, OCH<sub>2</sub>Ph) ppm. <sup>13</sup>C-NMR (75 MHz, DMSO) δ 163.5 (C-1), 165.0; 164.6; 161.3; 161.1 (dd, *J*<sub>CF</sub> = 246.3, 13.6 Hz, Ph-3', Ph-5'), 147.9 (C-3), 144.6 (C-8), 143.6 (C-4), 135.7 (Ph-q), 135.3 (Ph-q), 134.3; 134.1; 133.9 (t, *J*<sub>CF</sub> = 11.0 Hz, Ph-1'), 128.8–127.9 (OCH<sub>2</sub> Ph), 123.4 (C-2), 123.0 (C-7), 108.3; 108.1; 108.0; 107.9 (dd, *J*<sub>CF</sub> = 17.9, 8.6 Hz, Ph-2', Ph-6'), 103.5; 103.1; 102.8 (t, *J*<sub>CF</sub> = 26.0 Hz, Ph-4'), 102.8 (C-5), 74.0 (OCH<sub>2</sub>Ph), 73.1 (OCH<sub>2</sub>Ph), 44.4 (C-6) ppm. Anal. calcd for C<sub>28</sub>H<sub>21</sub>F<sub>2</sub>N<sub>3</sub>O<sub>4</sub>: C, 67.06; H, 4.22; N, 8.38. Found: C, 67.12; H, 4.21; N, 8.40.

**Preparation of (Z)-2,3-*O*,*O*-dibenzyl-6-[4-(4-bromophenyl)-1,2,3-triazole-1-yl]-4,5-didehydro-5,6-dideoxy-L-ascorbic acid (**6c**).** Compound **6c** was synthesized according to the general procedure using compound **5** (300 mg, 0.826 mmol), Cu(OAc)<sub>2</sub> (7.50 mg, 0.041 mmol) and 4-bromophenylacetylene (179 mg, 0.991 mmol) in CH<sub>3</sub>OH (16.5 mL). Compound **6c** (220 mg, 49%, mp = 126–131 °C) was isolated as white powder. <sup>1</sup>H NMR (300 MHz, DMSO) δ 8.66 (s, 1H, H-7), 7.80 (d, *J* = 8.5 Hz, 2H, Ph-2', Ph-6'), 7.64 (d, *J* = 8.6 Hz, 2H, Ph-3', Ph-5'), 7.51–7.27 (m, 10H, OCH<sub>2</sub> Ph), 5.69 (t, *J* = 7.5 Hz, 1H, H-5), 5.33 (s, 2H, OCH<sub>2</sub>Ph), 5.27 (d, *J* = 7.5 Hz, 2H, H-6), 5.17 (s, 2H, OCH<sub>2</sub>Ph) ppm. <sup>13</sup>C NMR (151 MHz, DMSO) δ 163.5 (C-1), 147.9 (C-3), 145.4 (C-8), 143.4 (C-4), 135.7 (Ph-q), 135.3 (Ph-q), 131.8 (Ph), 129.9 (Ph-1'), 128.8–127.1 (OCH<sub>2</sub> Ph, Ph'), 123.4 (C-2), 121.9 (C-7), 120.8 (Ph-4'), 101.7 (C-5), 74.0 (OCH<sub>2</sub>Ph), 73.0 (OCH<sub>2</sub>Ph), 44.3 (C-6) ppm. Anal. calcd for C<sub>28</sub>H<sub>22</sub>BrN<sub>3</sub>O<sub>4</sub>: C, 61.77; H, 4.07; N, 7.72. Found: C, 61.83; H, 4.08; N, 7.74.

**Preparation of (Z)-2,3-*O*,*O*-dibenzyl-4,5-didehydro-5,6-dideoxy-6-(4-tolyl-1,2,3-triazole-1-yl)-L-ascorbic acid (**6d**).**<sup>7</sup> Compound **6d** was synthesized according to the general procedure using compound **5** (345 mg, 0.950 mmol), Cu(OAc)<sub>2</sub> (8.62 mg, 0.047 mmol) and 4-tolylacetylene (0.15 mL, 1.140 mmol) in CH<sub>3</sub>OH (19 mL). Compound **6d** (91 mg, 20%, mp = 108–111 °C, in literature colorless oil) was isolated as white powder. <sup>1</sup>H NMR (300 MHz, DMSO) δ 8.54 (s, 1H, H-7), 7.72 (d, *J* = 8.1 Hz, 2H, Ph-2', Ph-6'), 7.46–7.20 (m, 12H, OCH<sub>2</sub> Ph, Ph-3', Ph-5'), 5.69 (t, *J* = 7.5 Hz, 1H, H-5), 5.33 (s, 2H, OCH<sub>2</sub>Ph), 5.26 (d, *J* = 7.5 Hz, 2H, H-6), 5.17 (s, 2H, OCH<sub>2</sub>Ph), 2.32 (s, 3H, CH<sub>3</sub>) ppm. <sup>13</sup>C NMR (151 MHz, DMSO) δ 163.6 (C-1), 148.0 (C-3), 146.6 (C-8), 143.3 (C-4), 137.2 (Ph-4'), 135.7 (Ph-q), 135.4 (Ph-q), 129.5–127.9 (OCH<sub>2</sub> Ph, Ph'), 127.9 (Ph-1'), 125.1 (Ph'), 123.4 (C-2), 121.2 (C-7), 102.0 (C-5), 74.1 (OCH<sub>2</sub>Ph), 73.1 (OCH<sub>2</sub>Ph), 44.2 (C-6), 20.8 (CH<sub>3</sub>) ppm. Anal. calcd for C<sub>29</sub>H<sub>25</sub>N<sub>3</sub>O<sub>4</sub>: C, 72.64; H, 5.25; N, 8.76. Found: C, 72.74; H, 5.26; N, 8.77.

**Preparation of (Z)-2,3-*O*,*O*-dibenzyl-4,5-didehydro-5,6-dideoxy-6-[4-(4-pentylphenyl)-1,2,3-triazole-1-yl]-L-ascorbic acid (**6e**).** Compound **6e** was synthesized according to the general procedure using compound **5** (300 mg, 0.826 mmol), Cu(OAc)<sub>2</sub>



(7.50 mg, 0.041 mmol) and 4-pentylphenylacetylene (0.20 mL, 0.991 mmol) in CH<sub>3</sub>OH (16.5 mL). Compound **6e** (202 mg, 46%, mp = 97–99 °C) was isolated as white powder. <sup>1</sup>H NMR (300 MHz, DMSO) δ 8.54 (s, 1H, H-7), 7.73 (d, *J* = 8.1 Hz, 2H, Ph-2', Ph-6'), 7.53–7.14 (m, 12H, OCH<sub>2</sub>Ph, Ph-3', Ph-5'), 5.69 (t, *J* = 7.5 Hz, 1H, H-5), 5.33 (s, 2H, OCH<sub>2</sub>Ph), 5.26 (d, *J* = 7.5 Hz, 2H, H-6), 5.17 (s, 2H, OCH<sub>2</sub>Ph), 2.58 (t, 2H, H-1'), 1.66–1.44 (m, 2H, H-2'), 1.32–1.15 (m, 4H, H-3', H-4'), 0.86 (t, *J* = 6.8 Hz, 3H, CH<sub>3</sub>) ppm. <sup>13</sup>C NMR (151 MHz, DMSO) δ 163.5 (C-1), 147.9 (C-3), 146.6 (C-8), 143.3 (C-4), 142.1 (Ph-4'), 135.7 (Ph-q), 135.3 (Ph-q), 128.8–128.1 (OCH<sub>2</sub>Ph, Ph'), 127.8 (Ph-1'), 125.1 (Ph'), 123.4 (C-2), 121.1 (C-7), 101.9 (C-5), 74.0 (OCH<sub>2</sub>Ph), 73.0 (OCH<sub>2</sub>Ph), 44.2 (C-6), 34.8 (C-1'), 30.8 (CH<sub>2</sub>), 30.4 (CH<sub>2</sub>), 21.9 (C-4'), 13.8 (CH<sub>3</sub>) ppm. Anal. calcd for C<sub>33</sub>H<sub>33</sub>N<sub>3</sub>O<sub>4</sub>: C, 74.00; H, 6.21; N, 7.84. Found: C, 74.16; H, 6.20; N, 7.83.

**Preparation of (Z)-2,3-O,O-dibenzyl-4,5-didehydro-5,6-dideoxy-6-[4-(4-methoxyphenyl)-1,2,3-triazole-1-yl]-L-ascorbic acid (6f).** Compound **6f** was synthesized according to the general procedure using compound **5** (291 mg, 0.801 mmol), Cu(OAc)<sub>2</sub> (7.27 mg, 0.048 mmol) and 4-methoxyphenylacetylene (0.13 mL, 0.961 mmol) in CH<sub>3</sub>OH (16 mL). Compound **6f** (193 mg, 49%, mp = 131–134 °C) was isolated as white powder. <sup>1</sup>H NMR (300 MHz, DMSO) δ 8.49 (s, 1H, H-7), 7.76 (d, *J* = 8.7 Hz, 2H, Ph-2', Ph-6'), 7.54–7.24 (m, 10H, OCH<sub>2</sub>Ph), 7.00 (d, *J* = 8.8 Hz, 2H, Ph-3', Ph-5'), 5.69 (t, *J* = 7.5 Hz, 1H, H-5), 5.33 (s, 1H, OCH<sub>2</sub>Ph), 5.25 (d, *J* = 7.5 Hz, 1H, H-6), 5.17 (s, 1H, OCH<sub>2</sub>Ph), 3.78 (s, 3H, OCH<sub>3</sub>) ppm. <sup>13</sup>C NMR (75 MHz, DMSO) δ 163.6 (C-1), 159.0 (Ph-4'), 148.0 (C-3), 146.5 (C-8), 143.3 (C-4), 135.7 (Ph-q), 135.4 (Ph-q), 128.8–126.5 (OCH<sub>2</sub>Ph, Ph-2', Ph-6'), 123.4 (C-2), 123.2 (Ph-1'), 120.6 (C-7), 114.3 (Ph-3', Ph-5'), 102.0 (C-5), 74.1 (OCH<sub>2</sub>Ph), 73.1 (OCH<sub>2</sub>Ph), 55.1 (OCH<sub>3</sub>), 44.2 (C-6) ppm. Anal. calcd for C<sub>29</sub>H<sub>25</sub>N<sub>3</sub>O<sub>5</sub>: C, 70.29; H, 5.09; N, 8.48. Found: C, 70.31; H, 5.08; N, 8.47.

**Preparation of (Z)-2,3-O,O-dibenzyl-4,5-didehydro-5,6-dideoxy-6-[4-(2-methoxyphenyl)-1,2,3-triazole-1-yl]-L-ascorbic acid (6g).** Compound **6g** was synthesized according to the general procedure using compound **5** (350 mg, 0.963 mmol), Cu(OAc)<sub>2</sub> (8.75 mg, 0.048 mmol) and 2-methoxyphenylacetylene (0.15 mL, 1.156 mmol) in CH<sub>3</sub>OH (19 mL). Compound **6g** (392 mg, 82%) was isolated as yellow oil. <sup>1</sup>H NMR (300 MHz, DMSO) δ 8.45 (s, 1H, H-7), 8.12 (dd, *J* = 7.7 Hz, 1.6 Hz, 1H, Ph-6'), 7.52–7.24 (m, 11H, OCH<sub>2</sub>Ph, Ph-4'), 7.13 (d, *J* = 8.8 Hz, 1H, Ph'), 7.08–7.01 (m, 1H, Ph'), 5.69 (t, *J* = 7.5 Hz, 1H), 5.33 (s, 2H, OCH<sub>2</sub>Ph), 5.25 (d, *J* = 7.5 Hz, 2H, H-6), 5.17 (s, 2H, OCH<sub>2</sub>Ph), 3.78 (s, 3H, OCH<sub>3</sub>) ppm. <sup>13</sup>C NMR (151 MHz, DMSO) δ 163.6 (C-1), 155.3 (Ph-2'), 148.0 (C-3), 143.0 (C-4), 141.9 (C-8), 135.7 (Ph-q), 135.4 (Ph-q), 128.9–126.5 (OCH<sub>2</sub>Ph, Ph'), 124.1 (C-7), 123.3 (C-2), 120.6 (Ph-5'), 119.0 (Ph-1'), 111.5 (Ph-3'), 102.4 (C-5), 74.0 (OCH<sub>2</sub>Ph), 73.0 (OCH<sub>2</sub>Ph), 55.4 (OCH<sub>3</sub>), 44.1 (C-6) ppm. Anal. calcd for C<sub>29</sub>H<sub>25</sub>N<sub>3</sub>O<sub>5</sub>: C, 70.29; H, 5.09; N, 8.48. Found: C, 70.36; H, 5.10; N, 8.50.

**Preparation of (Z)-2,3-O,O-dibenzyl-4,5-didehydro-5,6-dideoxy-6-[4-(3-hydroxyphenyl)-1,2,3-triazole-1-yl]-L-ascorbic acid (6h).** Compound **6h** was synthesized according to the general procedure using compound **5** (300 mg, 0.826 mmol), Cu(OAc)<sub>2</sub> (7.50 mg, 0.041 mmol) and 3-hydroxyphenylacetylene (0.11 mL, 0.991 mmol) in CH<sub>3</sub>OH (16.5 mL). Compound **6h** (314 mg, 79%, mp 160–161 °C) was isolated as white powder. <sup>1</sup>H-NMR (300 MHz, DMSO) δ 9.53 (s, 1H, OH), 8.6 (s, 1H, H-7), 7.54–7.11 (m, 13H,

OCH<sub>2</sub>Ph, Ph'), 6.74–6.71 (m, 1H, Ph-4'), 5.70 (t, *J* = 7.5 Hz, 1H, H-5), 5.33 (s, 2H, OCH<sub>2</sub>Ph), 5.26 (d, *J* = 7.5 Hz, 2H, H-6), 5.17 (s, 2H, OCH<sub>2</sub>Ph) ppm. <sup>13</sup>C NMR (151 MHz, DMSO) δ 163.7 (C-1), 157.8 (Ph-3'), 148.1 (C-3), 146.7 (C-8), 143.4 (C-4), 135.7 (Ph-q), 135.4 (Ph-q), 131.9 (Ph-1'), 130.0–128.0 (OCH<sub>2</sub>Ph, Ph'), 123.4 (C-2), 121.6 (C-7), 116.1 (Ph'), 115.0 (Ph'), 111.9 (Ph'), 102.1 (C-5), 74.1 (OCH<sub>2</sub>Ph), 73.1 (OCH<sub>2</sub>Ph), 44.3 (C-5) ppm. Anal. calcd for C<sub>28</sub>H<sub>23</sub>N<sub>3</sub>O<sub>5</sub>: C, 69.84; H, 4.81; N, 8.73. Found: C, 69.70; H, 4.81; N, 8.73.

**Preparation of (Z)-2,3-O,O-dibenzyl-6-[4-cyclopropyl-1,2,3-triazole-1-yl]-4,5-didehydro-5,6-dideoxy-L-ascorbic acid (6i).** Compound **6i** was synthesized according to the general procedure using compound **5** (330 mg, 0.908 mmol), Cu(OAc)<sub>2</sub> (8.25 mg, 0.045 mmol) and cyclopropylacetylene (0.1 mL, 1.090 mmol) in CH<sub>3</sub>OH (18 mL). Compound **6i** (273 mg, 70%, mp = 65–67 °C, in literature colorless oil) was isolated as white powder. <sup>1</sup>H-NMR (300 MHz, DMSO) δ 7.85 (s, 1H, H-7), 7.44–7.30 (m, 10H, OCH<sub>2</sub>Ph), 5.60 (t, *J* = 7.5 Hz, 1H, H-5), 5.32 (s, 2H, OCH<sub>2</sub>Ph), 5.14 (s, 2H, OCH<sub>2</sub>Ph), 5.12 (s, 2H, H-6), 1.96–1.87 (m, 1H, H-1'), 0.91–0.84 (m, 2H, CH<sub>2</sub>), 0.71–0.66 (m, 2H, CH<sub>2</sub>) ppm. <sup>13</sup>C-NMR (151 MHz, DMSO) δ 163.5 (C-1), 149.2 (C-8), 147.9 (C-3), 143.1 (C-4), 135.7 (Ph-q), 135.3 (Ph-q), 128.8–127.9 (OCH<sub>2</sub>Ph), 123.3 (C-2), 120.8 (C-7), 102.2 (C-5), 74.0 (OCH<sub>2</sub>Ph), 73.0 (OCH<sub>2</sub>Ph), 43.9 (C-6), 7.5 (C-2', C-3'), 6.5 (C-1') ppm. Anal. calcd for C<sub>25</sub>H<sub>23</sub>N<sub>3</sub>O<sub>4</sub>: C, 69.92; H, 5.40; N, 9.78. Found: C, 70.04; H, 5.41; N, 9.77.

**Preparation of (Z)-2,3-O,O-dibenzyl-6-(4-butyl-1,2,3-triazole-1-yl)-4,5-didehydro-5,6-dideoxy-L-ascorbic acid (6j).** Compound **6j** was synthesized according to the general procedure using compound **5** (300 mg, 0.826 mmol), Cu(OAc)<sub>2</sub> (7.50 mg, 0.041 mmol) and 1-hexyne (0.12 mL, 0.991 mmol) in CH<sub>3</sub>OH (16.5 mL). Compound **6j** (236 mg, 64%, mp = 64–67 °C, in literature colorless oil) was isolated as white powder. <sup>1</sup>H NMR (300 MHz, DMSO) δ 7.87 (s, 1H, H-7), 7.63–7.14 (m, 10H, OCH<sub>2</sub>Ph), 5.61 (t, *J* = 7.5 Hz, 1H, H-5), 5.32 (s, 2H, OCH<sub>2</sub>Ph), 5.22–5.03 (m, 4H, OCH<sub>2</sub>Ph, H-6), 2.59 (t, *J* = 7.5 Hz, 2H, H-1'), 1.63–1.50 (m, 2H, H-2'), 1.41–1.14 (m, 2H, H-3'), 0.88 (t, *J* = 7.3 Hz, 3H, CH<sub>3</sub>) ppm. <sup>13</sup>C NMR (75 MHz, DMSO) δ 163.6 (C-1), 148.0 (C-3), 147.1 (C-8), 143.0 (C-4), 135.7 (Ph-q), 135.4 (Ph-q), 128.8–127.9 (OCH<sub>2</sub>Ph), 123.3 (C-2), 121.9 (C-7), 102.3 (C-5), 74.0 (OCH<sub>2</sub>Ph), 73.0 (OCH<sub>2</sub>Ph), 43.9 (C-6), 31.0 (C-1'), 24.6 (CH<sub>2</sub>), 21.6, (CH<sub>2</sub>), 13.7 (CH<sub>3</sub>) ppm. Anal. calcd for C<sub>26</sub>H<sub>27</sub>N<sub>3</sub>O<sub>4</sub>: C, 70.09; H, 6.11; N, 9.43. Found: C, 69.99; H, 6.08; N, 9.40.

**Preparation of (Z)-2,3-O,O-dibenzyl-6-[4-decyl-1,2,3-triazole-1-yl]-4,5-didehydro-5,6-dideoxy-L-ascorbic acid (6k).** Compound **6k** was synthesized according to the general procedure using compound **5** (351 mg, 0.966 mmol), Cu(OAc)<sub>2</sub> (8.77 mg, 0.048 mmol) and 1-dodecyne (0.25 mL, 1.159 mmol) in CH<sub>3</sub>OH (19 mL). Compound **6k** (425 mg, 83%, mp 88–70 °C) was isolated as white powder.

<sup>1</sup>H-NMR (300 MHz, DMSO) δ 7.87 (s, 1H, H-7), 7.44–7.31 (m, 10H, OCH<sub>2</sub>Ph), 5.61 (t, *J* = 7.2 Hz, 1H, H-5), 5.32 (s, 2H, OCH<sub>2</sub>Ph), 5.19–5.15 (m, 4H, OCH<sub>2</sub>Ph, H-6), 2.57 (t, *J* = 7.5 Hz, 2H, H-1'), 1.60–1.51 (m, 2H, H-2'), 1.23 (s, 14H, CH<sub>2</sub> × 7), 0.84 (t, *J* = 6.9 Hz, 3H, CH<sub>3</sub>) ppm. <sup>13</sup>C-NMR (151 MHz, DMSO) δ 163.5 (C-1), 147.9 (C-3), 147.2 (C-8), 143.0 (C-3), 135.7 (Ph-q), 135.4 (Ph-q), 128.8–127.9 (OCH<sub>2</sub>Ph), 123.3 (C-2), 121.8 (C-7), 102.2 (C-



5), 74.0 (OCH<sub>2</sub>Ph), 73.0 (OCH<sub>2</sub>Ph), 43.8 (C-6), 31.2 (C-1'), 28.9 (CH<sub>2</sub>), 28.9 (CH<sub>2</sub>), 28.9 (CH<sub>2</sub>), 28.7 (CH<sub>2</sub>), 28.6 (CH<sub>2</sub>), 28.5 (CH<sub>2</sub>), 24.9 (CH<sub>2</sub>), 22.1 (CH<sub>2</sub>), 13.9 (CH<sub>3</sub>) ppm. Anal. calcd for C<sub>32</sub>H<sub>39</sub>N<sub>3</sub>O<sub>4</sub>: C, 72.56; H, 7.42; N, 7.93. Found: C, 72.69; H, 7.43; N, 7.95.

**Preparation of (Z)-2,3-O,O-dibenzyl-4,5-didehydro-5,6-dideoxy-6-[4-(2-hydroxyethyl)-1,2,3-triazole-1-yl]-L-ascorbic acid (6l).** Compound **6l** was synthesized according to the general procedure using compound **5** (300 mg, 0.826 mmol), Cu(OAc)<sub>2</sub> (7.50 mg, 0.041 mmol) and 3-butyne-1-ol (0.08 mL, 0.991 mmol) in CH<sub>3</sub>OH (16.5 mL). Compound **6l** (218 mg, 61%, mp = 59–60 °C) was isolated as white powder. <sup>1</sup>H NMR (300 MHz, DMSO) δ 7.89 (s, 1H, H-7), 7.56–7.17 (m, 10H, OCH<sub>2</sub>Ph), 5.61 (t, *J* = 7.5 Hz, 1H, H-5), 5.32 (s, 2H, OCH<sub>2</sub>Ph), 5.24–5.09 (m, 4H, OCH<sub>2</sub>Ph, H-6), 4.67 (t, *J* = 5.0 Hz, 1H, OH), 3.61 (dd, *J* = 11.6, 6.7 Hz, 2H, H-2'), 2.75 (t, *J* = 6.9 Hz, 1H, H-1') ppm. <sup>13</sup>C NMR (151 MHz, DMSO) δ 163.5 (C-1), 147.9 (C-3), 144.7 (C-8), 143.1 (C-4), 135.7 (Ph-q), 135.4 (Ph-q), 128.8–127.9 (OCH<sub>2</sub>Ph), 123.3 (C-2), 122.5 (C-7), 102.2 (C-5), 74.0 (OCH<sub>2</sub>Ph), 73.0 (OCH<sub>2</sub>Ph), 60.3 (C-2'), 43.8 (C-6), 29.1 (C-1') ppm. Anal. calcd for C<sub>24</sub>H<sub>23</sub>N<sub>3</sub>O<sub>5</sub>: C, 66.50; H, 5.35; N, 9.69. Found: C, 66.74; H, 5.36; N, 9.69.

**Preparation of (Z)-2,3-O,O-dibenzyl-6-(4-chloropropyl-1,2,3-triazole-1-yl)-4,5-didehydro-5,6-dideoxy-L-ascorbic acid (6m).** Compound **6m** was synthesized according to the general procedure using compound **5** (347 mg, 0.955 mmol), Cu(OAc)<sub>2</sub> (8.67 mg, 0.048 mmol) and 5-chloro-1-pentyne (0.12 mL, 1.146 mmol) in CH<sub>3</sub>OH (19 mL). Compound **6m** (158 mg, 36%) was isolated as white powder.

<sup>1</sup>H NMR (300 MHz, DMSO) δ 7.94 (s, 1H, H-7), 7.54–7.17 (m, 10H, OCH<sub>2</sub>Ph), 5.62 (t, *J* = 7.5 Hz, 1H, H-5), 5.32 (s, 2H, OCH<sub>2</sub>Ph), 5.25–5.06 (m, 4H, OCH<sub>2</sub>Ph, H-6), 3.67 (t, *J* = 6.5 Hz, 2H, H-3'), 2.74 (t, *J* = 7.4 Hz, 2H, H-1'), 2.14–1.88 (m, 2H, H-2') ppm. <sup>13</sup>C NMR (151 MHz, DMSO) δ 163.5 (C-1), 147.9 (C-3), 145.7 (C-8), 143.1 (C-3), 135.7 (Ph-q), 135.3 (Ph-q), 128.8–127.9 (OCH<sub>2</sub>Ph), 123.3 (C-2), 122.2 (C-7), 102.1 (C-5), 74.0 (OCH<sub>2</sub>Ph), 73.0 (OCH<sub>2</sub>Ph), 44.6 (C-3'), 43.9 (C-6), 31.7 (C-1'), 22.2 (C-2') ppm. Anal. calcd for C<sub>25</sub>H<sub>24</sub>ClN<sub>3</sub>O<sub>4</sub>: C, 64.44; H, 5.19; N, 9.02. Found: C, 64.58; H, 5.20; N, 9.04.

**Preparation of (Z)-2,3-O,O-dibenzyl-6-(4-tert-butyl-1,2,3-triazole-1-yl)-4,5-didehydro-5,6-dideoxy-L-ascorbic acid (6n).** Compound **6n** was synthesized according to the general procedure using compound **5** (300 mg, 0.826 mmol), Cu(OAc)<sub>2</sub> (7.50 mg, 0.041 mmol) and 3,3-dimethyl-1-butyne (0.12 mL, 0.991 mmol) in CH<sub>3</sub>OH (16.5 mL). Compound **6n** (96 mg, 26%) was isolated as a transparent oil. <sup>1</sup>H NMR (300 MHz, DMSO) δ 7.89 (s, 1H, H-7), 7.61–7.18 (m, 10H, OCH<sub>2</sub>Ph), 5.62 (t, *J* = 7.5 Hz, 1H, H-5), 5.33 (s, 2H, OCH<sub>2</sub>Ph), 5.23–5.09 (m, 4H, OCH<sub>2</sub>Ph, H-6), 1.25 (s, 9H, 3 × CH<sub>3</sub>) ppm. <sup>13</sup>C NMR (151 MHz, DMSO) δ 163.5 (C-1), 156.4 (C-8), 147.9 (C-3), 143.0 (C-4), 135.7 (Ph-q), 135.3 (Ph-q), 128.8–127.9 (OCH<sub>2</sub>Ph), 123.3 (C-2), 119.9 (C-7), 102.3 (C-5), 74.0 (OCH<sub>2</sub>Ph), 73.0 (OCH<sub>2</sub>Ph), 43.9 (C-6), 30.5 (C-1'), 30.2 (CH<sub>3</sub>) ppm. Anal. calcd for C<sub>26</sub>H<sub>27</sub>N<sub>3</sub>O<sub>4</sub>: C, 70.09; H, 6.11; N, 9.43. Found: C, 69.97; H, 6.11; N, 9.44.

## General flow procedure for the synthesis of 1,2,3-triazolyl L-ascorbic acid derivatives 6a–6n in a microreactor

Solution 1: C-6 azido L-ascorbic acid derivative (**5**) (0.1 M) and terminal alkyne (1 equiv.) were diluted in CH<sub>3</sub>OH.

Solution 2: Cu(OAc)<sub>2</sub> was diluted in CH<sub>3</sub>OH (0.005 M) and homogenized by sonication.

Solution 1 and 2 were loaded into an injection loop and injected into the micro-flow reactor set at 50 °C at a rate of 0.5 μL min<sup>−1</sup> (9 min residence time). The microreactor was positioned into an ultrasound bath. Outflows from the microreactors were collected in vials placed on ice and filled with QuadraPure BZA copper-scavenger resin (polymer supported benzylamine) to stop the reaction. Reaction mixture was filtered *via* cotton. Prior to the HPLC analysis the sample (10 μL) was diluted in H<sub>2</sub>O (300 μL) and acetonitrile (400 μL). Conversion of compound **5** in flow reactions was determined using a prepared calibration curve of compound **5**.

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