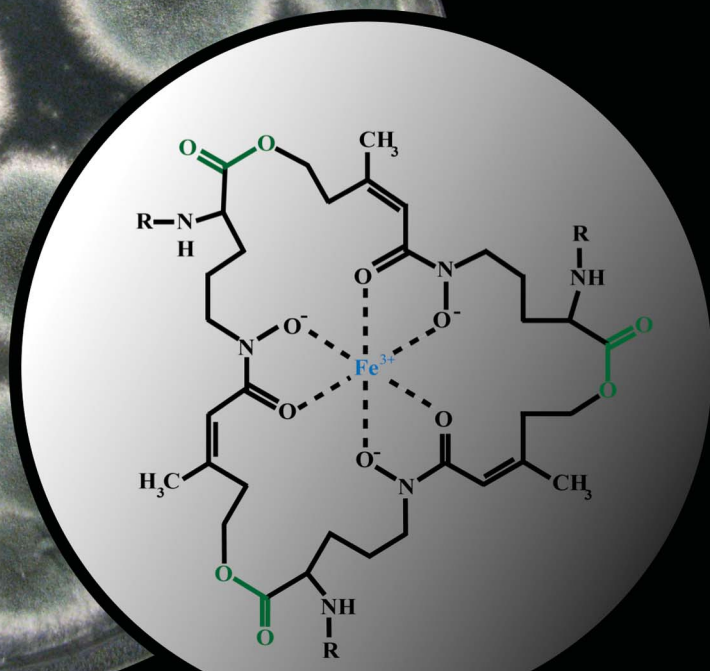


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HIGHLIGHT

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Fungal siderophore metabolism with a focus on *Aspergillus fumigatus*


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Fungal siderophore metabolism with a focus on *Aspergillus fumigatus*

Hubertus Haas*

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Siderophores are chelators synthesized by microbes to sequester iron. This article summarizes the knowledge on the fungal siderophore metabolism with a focus on *Aspergillus fumigatus*. In recent years, *A. fumigatus* became a role model for fungal biosynthesis, uptake and degradation of siderophores as well as regulation of siderophore-mediated iron handling and the elucidation of siderophore functions. Siderophore functions comprise uptake, intracellular transport and storage of iron. This proved to be crucial not only for adaptation to iron starvation conditions but also for germination, asexual and sexual propagation, antioxidative defense, mutual interaction, microbial competition as well as virulence in plant and animal hosts. Recent studies also indicate the high potential of siderophores and its biosynthetic pathway to improve diagnosis and therapy of fungal infections.

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1. Introduction

Fungi play a major role in ecology and affect human life in many ways positively and negatively: they are major players in saprobic decomposition, important symbionts of plants (mycorrhiza), pathogens of plants and animals affecting health as well as food production and preservation (decay, toxin production), serve as

food or in food production (e.g., mushrooms, alcohol, bread, mould cheese), and are important work horses in biotechnology as producers of widely used enzymes as well as primary and secondary metabolites (e.g. citric acid, statins, cyclosporine, β -lactam antibiotics). A crucial nutrient affecting all of these functions is iron and in most fungal species, iron handling involves fungal-specific ferric iron chelators, termed siderophores.

2. *Aspergillus fumigatus*

Aspergillus fumigatus is a ubiquitous saprophytic fungus typically residing in the soil or decaying organic matter. As an efficient recycler in nature, this mould possesses a versatile metabolism that meets its nutritional requirements under a wide range of environmental conditions.¹ *A. fumigatus* produces huge numbers of minute (2–3 μ m diameter) asexual spores that are efficiently released into the environment, particularly into the air. These conidia are constantly inhaled and rapidly eliminated by a functional immune system. However, immunosuppression such as in patients undergoing bone marrow transplantation, cancer chemotherapy or suffering from chronic granulomatous disease, increases the risk of invasive systemic infection, termed aspergillosis. Due to the increase in immunosuppressive regimens in medical interventions, *A. fumigatus* has become the most common air-borne fungal pathogen of humans.² The latter boosted the interest in this fungus, leading to the still ongoing detailed characterization of its physiology and metabolism, particularly of course of its virulence determinants. As a result, *A. fumigatus* is now one of the best-studied filamentous fungal species and a fungal role

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Highlight

model including a sequenced genome, which facilitates molecular analysis and allows “genome mining”.³

3. Iron homeostasis

Due to the ability to coordinate and activate oxygen and the ideal redox chemistry for electron transport, iron is an indispensable cofactor for a variety of cellular processes in all eukaryotes and most prokaryotes including respiration, amino acid metabolism, and biosynthesis of DNA and sterols. On the other hand, iron excess has the potential to catalyse the formation of cell-damaging reactive oxygen species *via* the Fenton and Haber–Weiss reactions.⁴ The iron-dependence of several enzymes involved in oxidative stress detoxification such as heme-dependent catalases and peroxidases emphasizes the complexity of cellular iron handling.

Despite its high abundance in the Earth’s crust, the bioavailability of iron is low owing to its oxidation by atmospheric oxygen into sparingly soluble ferric hydroxides. For pathogens in plant as well as animal hosts, iron is not freely available but tightly sequestered, *e.g.* in vertebrates most of the iron is associated with proteins such as transferrin, lactoferrin and ferritin and with heme that is bound by hemoglobin. The preclusion of invading microbes from iron represents a key pathway in host defence as a component of the so called “nutritional immunity”.^{5,6} Consequently, pathogenic as well as non-pathogenic organisms had to evolve sophisticated strategies to ensure iron supply, while at the same time avoiding iron toxicity. This includes diverse mechanisms for uptake and storage of iron as well as fine-tuned regulation of acquisition, storage and consumption of iron.

4. Fungal iron acquisition and storage

In fungi, four different mechanisms for acquisition of iron have been identified: (i) ferric iron uptake mediated by ferric iron-specific chelators, termed siderophores, (ii) reductive iron assimilation (RIA), (iii) low-affinity ferrous iron uptake and (iv) heme uptake and degradation. *A. fumigatus*, similar to most

fungal species, employs siderophore-mediated iron uptake, RIA and low affinity iron uptake.⁷ *A. fumigatus* appears unable to utilize heme as an iron source.⁸ Interestingly, the fungal species studied in most detail lack siderophore biosynthesis (SB) including the opportunistic pathogenic basidiomycete *Cryptococcus neoformans* and the *Saccharomycotina* including the fungal role model *Saccharomyces cerevisiae* and the opportunistic pathogenic genus *Candida* spp.⁷ Nevertheless, these siderophore non-producers are able to take up and utilize iron chelated by siderophores from other organisms, termed xenosiderophores.⁹ Interestingly, all these siderophore non-producers display predominantly yeast morphology. However, there are also yeast species that produce siderophores such as *Schizosaccharomyces pombe* or *Aureobasidium pullulans*.^{10,11} The siderophore non-producers utilize RIA, heme-bound iron and low affinity iron uptake.¹² There are also examples for species lacking RIA such as *Aspergillus nidulans* and *Coprinus* spp.^{7,13}

To overcome periods of iron-limitation as well as to avoid iron toxicity, all cells require means to store iron. In fungi, two strategies for iron storage are known: (i) vacuolar iron deposition and (ii) siderophore-mediated iron storage.^{7,12} In contrast to bacteria, plants and animals, most fungi appear to lack ferritin-mediated iron storage and detoxification. *A. fumigatus* employs both intracellular siderophores and vacuolar iron deposition for iron storage.^{14,15}

5. Fungal siderophores

The majority of fungal siderophores belong to the hydroxamate-class. Exceptions are the carboxylate-type siderophore rhizoferrin produced by various *Mucorales* and the catecholate pistillaridin produced by the marine species *Penicillium bilaii*.^{16,17} Fungal hydroxamate siderophores can be grouped into four structural families: fusarinines, coprogens, ferrichromes and rhodotorulic acid. Representatives of each family are shown in Fig. 1. The chemistry of siderophores has been comprehensively reviewed.^{18,19} Universal to all is the hydroxamate group that forms a strong iron(III)-binding bidentate. The hydroxamate group is built by acylation of the non-proteinogenic amino acid *N*⁵-hydroxy-L-ornithine, which is derived by hydroxylation of L-ornithine, with acetyl or more complex groups such as anhydromevalonyl. To increase affinity for Fe(III), most fungal siderophores include three of these moieties linked by ester or peptide bonds to form hexadentate structures. Cyclisation of the siderophore, as found in ferrichromes and some fusarinines, improves the chemical stability.

Rhodotorulic acid, produced by basidiomycetous yeasts such as *Rhodotorula* spp., is a dipeptide consisting of two *N*⁵-acetyl-*N*⁵-hydroxy-L-ornithine units linked head-to-head to form a diketopiperazine ring. Fusarinines consist of three *N*⁵-*cis*-anhydromevalonyl-*N*⁵-hydroxy-L-ornithine units linked head-to-tail by ester bonds in linear or cyclic form. Coprogens are linear di- or tri-hydroxamates composed of *trans*-fusarinine units (*i.e.* the acyl groups are *trans*-anhydromevalonyl moieties). Coprogen consist of three *N*⁵-*trans*-anhydromevalonyl-*N*⁵-hydroxy-L-ornithine units, with two units linked head-to-head to form a diketopiperazine ring (dimerum acid) and a third unit esterified to



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main research interests are pathways and regulatory circuits that are specific for fungi with a focus on siderophore and secondary metabolism.



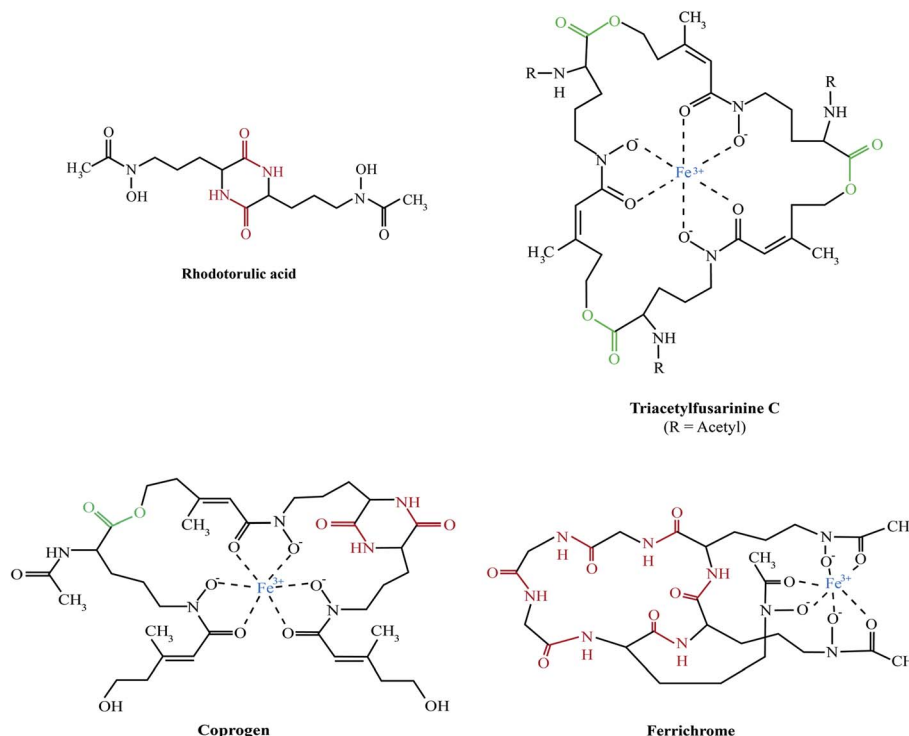


Fig. 1 Representatives of the major fungal siderophore types. With exception of rhodotorulic acid, siderophores are shown in the ferric form. Peptide bonds, ester bonds, and Fe(III) are shown in red, green and blue, respectively. In T AFC, R = acetyl, in F sC R = H.

the C-terminal OH group of dimerum acid. N^2 -Acetylation and N^2 -methylation allow for the diversity of the coprogen family. Ferrichromes are cyclic hexapeptides consisting of three N^5 -acyl- N^5 -hydroxy-L-ornithines and three additional amino acids, with two residues consisting of alanine, serine or glycine and the third residue always being a glycine.²⁰ A number of different acyl groups have been found in this family (acetyl, malonyl, *trans*- β -ethylglutaconyl, *trans*-anhydromevalonyl, and *cis*-anhydromevalonyl). Recently, a ferrichrome-type siderophore with a novel structure comprised of four glycines, a glutamine and three *N*⁵-*trans*-anhydromevalonyl- N^5 -hydroxy-L-ornithine moieties, produced by the endophyte *Epichloë festucae*, termed epichloënin A, was identified.²¹ The function of hexadecydroastechrome, a tryptophan-derived and iron-binding hydroxamate recently isolated from *A. fumigatus*, is still unknown.²²

6. The siderophore biosynthetic pathway

A. fumigatus produces four siderophores: two extracellular fusarinine-type siderophores, fusarinine C (F sC) and its derivative triacetylfulsarinine C (T AFC) for iron uptake as well as two intracellular ferrichrome-type siderophores, ferricrocin (F C) for distribution and storage of iron in hyphae and hydroxyferricrocin (H F C) for storage of iron in asexually-produced spores, termed conidia.²³ A scheme of T AFC-mediated iron uptake and F C-mediated iron storage is displayed in Fig. 2.

F sC consists of three N^5 -anhydromevalonyl- N^5 -hydroxy-L-ornithine residues cyclically linked by ester bonds. T AFC is the

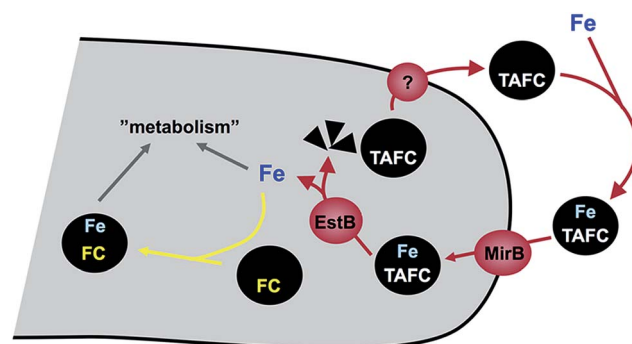


Fig. 2 Schematic view of T AFC-mediated iron acquisition and F C-mediated iron storage in *A. fumigatus*. The transporters for secretion of siderophores including T AFC (marked by a question mark) have not been identified yet.

N^2 -acetylated F sC. F C is a cyclic hexapeptide with the structure Gly-Ser-Gly-(N^5 -acetyl- N^5 -hydroxy-L-ornithine)₃ and H F C is the hydroxylated F C with a single unidentified hydroxylation site.⁷ Based on biochemical studies, the fungal siderophore biosynthetic pathway was postulated by Plattner and Diekmann.²⁴ In *A. fumigatus*, the siderophore biosynthetic pathway has been characterized at the molecular level by reverse genetics, *i.e.* generation and characterization of mutant strains with defects in putative siderophore biosynthetic genes identified by their upregulation during iron starvation.²³ The siderophore biosynthetic pathway of *A. fumigatus* is depicted in Fig. 3. The first committed step in the biosynthesis of all four siderophores is the



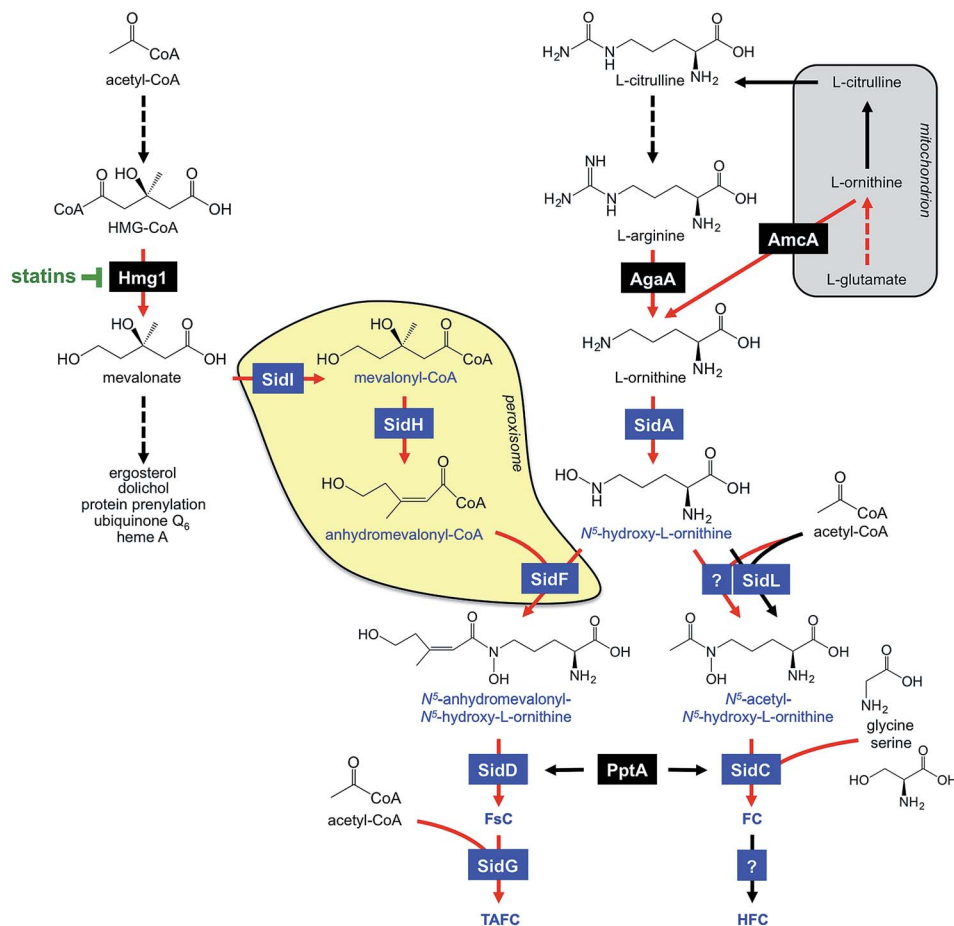


Fig. 3 *A. fumigatus* SB (in blue) and its links to the isoprenoid biosynthesis as well as ornithine/arginine metabolism. Involved enzymes are boxed and described in the text. Enzymatic steps transcriptionally upregulated during iron starvation are marked by red arrows. Broken arrows denote reactions involving more than one enzyme. Mitochondrion and peroxisome are shaded in grey and yellow, respectively.

*N*⁵-hydroxylation of L-ornithine catalysed by the L-ornithine-*N*⁵-monooxygenase SidA.⁸ SidA orthologues have been shown to be responsible for SB in various other fungal species including the ascomycetes *A. nidulans*, *Aspergillus oryzae*, *Fusarium graminearum*, *Magnaporthe grisea*, *Histoplasma capsulatum*, *A. pullulans* and the basidiomycete *Ustilago maydis*.^{11,13,25–29} These fungal ornithine-*N*⁵-monooxygenases show significant similarity at the protein level to bacterial siderophore-biosynthetic enzymes such as the ornithine-*N*⁵-monooxygenases PvdA from *Pseudomonas aeruginosa* and the lysine-*N*⁵-monooxygenases IucD from *Escherichia coli*.^{30,31} PvdA and IucD are essential for biosynthesis of pyoverdine and aerobactin, respectively. All these enzymes require oxygen as substrate and NADPH and FAD as co-factors.²⁹

Subsequently, the pathways for biosynthesis of extra- and intracellular siderophores, *i.e.* fusarinines and ferrichromes, split due to attachment of different acyl residues. For extracellular fusarinine-type siderophores, the transacylase SidF transfers anhydromevalonyl to *N*⁵-hydroxy-L-ornithine.²³ The required anhydromevalonyl-CoA moiety is derived from mevalonate by CoA-ligation and dehydration catalysed by the enzymes SidI and SidH, respectively, thereby linking isoprenoid and siderophore biosynthetic pathways.³² SidI belongs to the acyl-CoA synthetase protein family with most

members being involved in lipid metabolism. In *Neurospora crassa*, the SidI orthologue Acs5 was shown to be involved in coprogen biosynthesis.³³ SidH belongs to the enoyl-CoA hydratase protein family, again, with most members being involved in lipid metabolism.³⁴ The *N*⁵-acetylation of *N*⁵-hydroxy-L-ornithine for the intracellular ferrichrome-type siderophores requires two transacylases: the constitutively expressed enzyme SidL and an unidentified enzyme, the activity of which is upregulated by iron starvation.³⁵ Both SidF and SidL belong to the “GCN5-related *N*-acetyltransferases (GNAT)”. *A. fumigatus* possesses several GNAT proteins, but SidF and SidL appear to be the only two carrying the pfam10331/AlcB domain that is also conserved in bacterial siderophore biosynthetic enzymes such as the *N*⁵-hydroxylysine:acetyl coenzyme A *N*⁵-transacylase IucB, which is involved in biosynthesis of the hydroxamate aerobactin.³⁶ The only other functionally characterized fungal SidF orthologue is Fer5 from *U. maydis*, which catalyses *N*⁵-acylation of *N*⁵-hydroxy-L-ornithine with methylglutaconyl-CoA for biosynthesis of ferrichrome A.³⁷ Similar to fusarinine biosynthesis in *A. fumigatus*, ferrichrome A biosynthesis is also linked to isoprenoid biosynthesis as methylglutaconyl-CoA is derived from HMG-CoA *via* dehydration by the enoyl-CoA-hydratase Fer4.³⁷



In *A. fumigatus*, fusarinines and ferrichromes are then assembled by two different non-ribosomal peptide synthetases (NRPSs), an enzyme family that is best known from secondary metabolism. The NRPS SidD links three *N*⁵-*cis*-anhydromevalonyl-*N*⁵-hydroxy-L-ornithine residues by ester bonds, yielding FsC.²³ The NRPS SidC links three *N*⁵-acetyl-*N*⁵-hydroxy-L-ornithine, two L-glycine and one L-serine residue by peptide bonds, yielding the hexapeptide ferricrocin. In other fungal species, SidC homologs were confirmed to be essential for synthesis of ferricrocin or other ferrichrome-type siderophores such as ferrichrome, ferrichrome A and ferrirhodin in *S. pombe*, *U. maydis* and *Fusarium sacchari*, respectively.^{13,37–40} Consistently, phylogenetic analysis predicts monogenetic origin of ferrichrome-type NRPS.⁴¹ Similarly, SidD orthologs in other species were found to be responsible not only for synthesis of fusarinine- but also coprogen-type siderophores, e.g. *Alternaria brassicicola*, *C. heterostrophus* and *Colletotrichum gramini-cola*.^{42,43} This indicates a common evolutionary origin of fusarinine- and coprogen-synthetic NRPSs. SidE, another NRPS predicted to be involved in SB in *A. fumigatus*,⁴⁴ was recently demonstrated to be responsible for synthesis of fumarylalanine but not siderophores.⁴⁵

NRPSs, similar to polyketide synthetases and the α -amino-adipate reductase, which is involved in lysine-biosynthesis, depend on activation by the Sfp-type 4'-phosphopantetheinyl transferase, termed PptA in *A. fumigatus*.⁴⁶ Consistently, this enzyme was confirmed to be essential for SB in various fungal species, including *A. nidulans*, *Cochliobolus sativus*, *Fusarium fujikuroi* and *C. gramini-cola*.^{47–50}

T AFC is derived by triple *N*²-acetylation of FsC catalysed by SidG.²³ SidG belongs to the GNAT protein family and its activity is particularly required for biosynthesis of T AFC but not FsC, coprogens and ferrichromes. In agreement, SidG orthologs are found only in few *Aspergillus*, *Fusarium* and *Nectria* species but not in species lacking T AFC production.

HFC is derived from FC by single hydroxylation catalysed by an unidentified gene product.²³ The hydroxylation site in HFC remains to be identified. So far, HFC has been detected only in *A. fumigatus* and appears to be missing in other FC-producing fungal species such as *A. nidulans* and *N. crassa*.^{51,52}

7. Subcellular localization of SB

In agreement with the presence of putative peroxisomal targeting motifs, the three enzymes linking isoprenoide and T AFC biosynthetic pathways, SidI, SidH and SidF, were recently found to be sequestered in peroxisomes.³⁴ This study also demonstrated that artificial delocalization of one or two but not of all three enzymes into the cytosol impairs T AFC biosynthesis suggesting that not all T AFC biosynthesis intermediates can efficiently cross the peroxisomal membrane. The peroxisomal targeting motifs are conserved in fungal orthologues of SidF, SidH and SidI.³⁴ In agreement with the evolutionary conservation of the partial peroxisomal compartmentalization of fungal SB, the SidI orthologue of *N. crassa* and the SidF ortholog of *Penicillium chrysogenum*, which are both involved in coprogen synthesis, were identified to be peroxisomal matrix proteins.^{34,53}

Exceptions are the SidH and SidF orthologs Fer4 and Fer5 from *U. maydis*, which lack peroxisomal targeting motifs.³⁷

8. Siderophore uptake and degradation

Cellular uptake of siderophore-iron complexes is mediated by "siderophore-iron transporters (SITs)", which belong to a subfamily of the major facilitator protein superfamily. SITs act most likely as proton symporters energized by the plasma membrane potential.^{54,55} SIT-mediated iron uptake is universally conserved in the fungal kingdom, even in species not producing siderophores such as *S. cerevisiae*, *Candida* spp. and *C. neoformans*.^{7,55–57} Functional characterization of SITs started in *S. cerevisiae*, which possesses four SITs with different substrate specificity: Sit1p/Arn3p for ferrioxamine B (a hydroxamate-type siderophore produced by various *Streptomyces* species), Arn1p for ferrichromes, Taf1p/Arn2p for T AFC, Enb1p/Arn4p for enterobactin (a catecholate-type siderophore produced by various *Enterobacteriaceae*). In contrast, *C. albicans* contains only a single broad range substrate specific SIT, CaArn1p/CaSit1p.⁵⁸ Similar to the siderophore non-producer *S. cerevisiae*, most fungal species including siderophore producers possess multiple SIT, e.g. *A. fumigatus* and *A. nidulans* encode seven and ten putative SITs, respectively.⁷ Heterologous expression in *S. cerevisiae* indicated that the *A. nidulans* SITs MirB and MirA transport T AFC and enterobactin, respectively.^{54,55} Similarly, the *A. fumigatus* MirB orthologue was confirmed to transport T AFC and FgSit1 from *F. graminearum* was suggested to transport ferrichrome.^{59,60}

Subsequent to uptake, the ester bonds of T AFC- and FsC-iron complexes are hydrolysed in the cytosol. Two of the enzymes involved have been identified, the esterases EstB and SidJ, which belong to two different protein families.^{61,62} Interestingly, EstB is homologous to bacterial siderophore degrading enzymes such as *E. coli* Fes, which is involved in release of iron from the catecholate-type siderophore enterobactin.⁶³ EstB is specific for T AFC and SidJ is specific for FsC.^{61,62} However, both enzymes are not essential for but rather optimize siderophore hydrolysis and consequently release of iron; e.g. inactivation of EstB ($\Delta estB$ mutant) decreases the growth rate during iron starvation, decreases the transfer of iron from T AFC to the metabolism and to the intracellular siderophore FC and consequently delays iron sensing. Lack of FC, vacuolar storage, and in particular lack of both increases the cellular content of iron chelated by siderophore breakdown products,¹⁵ indicating that the transfer of iron from extracellular siderophores to the metabolism, FC or the vacuole precedes recycling of siderophore breakdown products.

9. Genomic organization of genes involved in siderophore metabolism

In *A. fumigatus*, most of the identified siderophore metabolic genes are organized in three gene clusters, i.e. they are neighbouring each other in the genome, which most likely facilitates



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co-regulation of their expression.¹⁴ Exceptions are the genes encoding SidA, SidL, PptA and five of the seven putative siderophore transporters. In this respect interesting to note, expression of the genes encoding SidL and PptA is, in contrast to the other siderophore metabolic genes, not regulated by iron availability. Moreover, PptA is not exclusively involved in SB. Siderophore metabolic genes are usually also genomically clustered in other fungal species, whereby the number of clusters and gene combinations vary significantly,^{14,34} e.g., SidA ortholog-encoding genes are clustered with siderophore NRPS in various other fungi including *U. maydis* and *N. crassa*.

10. Regulation of the siderophore system

Cellular adaptation to iron starvation requires upregulation of high-affinity iron acquisition and downregulation of both iron storage and iron consumption, while adaptation to iron excess requires the inverse response. In *A. fumigatus*, changes in iron availability cause an extensive transcriptional remodeling affecting about 13% of the protein-encoding genes.¹⁴ The two central regulators mediating iron regulation at the transcriptional level in this fungus are the transcription factors SreA and HapX.^{14,64} During iron sufficiency, the GATA-factor SreA, which binds to the consensus sequence ATCWGATAA, represses high-affinity iron uptake, including the siderophore system, to avoid iron toxicity. During iron starvation, the bZIP-transcription factor HapX represses iron-consuming pathways such as respiration, heme biosynthesis and iron-sulfur cluster-dependent processes to spare iron. Furthermore, HapX activates synthesis of the ribotoxin AspF1 and the siderophore system. HapX functions *via* physical interaction with the DNA-binding CCAAT-binding complex (CBC).⁶⁵ The CBC is a heterotrimeric DNA-binding complex, which is conserved in all eukaryotes. In *A. nidulans*, inactivation of either one of its subunits phenocopies HapX-deficiency with respect to the defects in adaptation to iron starvation.⁶⁵ However, the CBC has HapX-independent functions and is speculated to affect expression of about 30% of all genes. SreA and HapX are interconnected in a negative feedback loop: SreA represses expression of *hapX* during iron sufficiency, while HapX represses *sreA* during iron starvation. Additionally, both SreA and HapX are regulated posttranslationally with iron blocking HapX function and activating SreA function.^{65,66} Inactivation of both HapX and SreA is synthetically lethal underlining the critical role of iron homeostasis in cellular survival.^{14,64,65}

Most fungal species possess orthologs to SreA and HapX and the role of SreA in regulation of SB has been confirmed in *N. crassa*, *H. capsulatum*, *C. heterostrophus*, *S. pombe*, *A. pullulans* and *U. maydis*.^{67–72} In *S. pombe*, posttranslational iron sensing by the HapX and SreA orthologs involves glutathione and the monothiol glutaredoxin Grx4, indicating sensing of the cellular iron status *via* sensing the cellular iron-sulfur cluster availability.⁷⁰ In agreement with a similar mode of iron-sensing in *A. fumigatus*, extreme starvation for sulfur, which is essential for biosynthesis of both iron-sulfur clusters and glutathione, was

found to cause derepression of the siderophore system leading to detrimental excessive iron uptake.⁷³ Remarkably, *S. cerevisiae* lacks orthologs of SreA and HapX. It employs entirely different regulators, Aft1/2 and Cth1/2, which are conserved only in closely related *Saccharomycotina* species.¹² Nevertheless, *S. cerevisiae* Aft1/2 also senses the cellular iron status *via* iron sulfur cluster biogenesis involving glutathione and glutaredoxin.⁷⁴ These data indicate a universal conservation of the iron-sensing machinery executed by different types of transcription factors in different organisms.

Due to the central metabolic role of iron, it is not surprising that a variety of regulatory circuits affect cellular iron handling. For example, adaptation to hypoxia of the obligate aerobic *A. fumigatus* includes upregulation of glycolysis, the TCA-cycle, respiration and ergosterol biosynthesis and as a consequence increased cellular iron and heme contents.^{75,76} Hypoxic adaptation in *A. fumigatus* is coordinated by the transcription factor Srba, a member of the “sterol regulatory element binding protein (SREBP)” family, which is conserved in most eukaryotes.⁷⁶ These transcription factors are proteolytically activated by cellular sterol-depletion to maintain sterol homeostasis.⁷⁷ In *A. fumigatus*, Srba was found to be crucial not only for ergosterol-feedback regulation, resistance against triazole drugs and hypoxic growth, but also for activation of siderophore-mediated iron uptake in response to hypoxia and iron starvation.⁷⁸ The defects in hypoxic growth and azole resistance caused by Srba-deficiency were partially cured by high-iron supplementation, which underlines the involvement of Srba in iron homeostasis. Srba is transcriptionally upregulated during both hypoxia and iron starvation, most likely in response to sterol-depletion. These data implicate that *A. fumigatus* might sense iron not only *via* HapX and SreA but also *via* Srba. *S. cerevisiae* lacks an Srba ortholog and the regulators mediating hypoxic adaptation in this yeast are not conserved in most other fungal species.⁷⁶

Further regulatory proteins suggested to affect siderophore metabolism are the pH regulatory transcription factor PacC, the gluconeogenesis-activating transcription factor AcuM, the unfolded protein-responsive IreA-HacA pathway, the cross-pathway control mediating transcription factor CpcA, and the mitogen-activated protein kinase (MAPK) MpkA, which is involved in maintaining cell wall integrity, protection against ROS, and secondary metabolism.^{79–83}

11. Precursor supply for SB

During iron starvation, siderophore production reaches up to 10% of the biomass. This condition dramatically remodels the free amino acid pool of *A. fumigatus* with eight amino acids increasing and three amino acids decreasing more than 1.5-fold.⁶⁴ The 7-fold increase of the siderophore precursor L-ornithine during iron starvation compared to sufficiency indicates that the massive L-ornithine demand for SB is matched by active up-regulation of its biosynthesis. In agreement, blocking siderophore-mediated ornithine consumption by inactivation of SidA (Δ sidA mutant) increases the cellular L-ornithine pool another 2.9-fold.⁶⁴



The biosynthesis of L-ornithine in the mitochondrial matrix as well as its export *via* the transporter AmcA to the cytosol are transcriptionally upregulated under iron deprivation.⁶⁴ L-Ornithine is a precursor of L-arginine, and the cytosolic localized conversion of L-arginine to ornithine by the arginase AgaA is likewise upregulated in response to iron starvation (Fig. 3). Nevertheless, the metabolic commitment for siderophore production must be balanced against other essential cellular functions, *e.g.* L-arginine is vital for protein biosynthesis and ornithine is a precursor for polyamine biosynthesis. Recently, L-arginine was found to allosterically activate SidA enzyme activity,⁸⁴ *i.e.* is stimulated only if the cytosolic L-arginine pool is sufficient for all cellular needs. In agreement, studies with arginine-auxotrophic *A. fumigatus* mutants, which allowed manipulation of the cellular L-arginine and L-ornithine levels, revealed prioritization of these amino acids for cellular needs other than SB, *e.g.* arginine for protein biosynthesis and ornithine for polyamine biosynthesis.⁸² The rationale for this regulation might be that *A. fumigatus* possesses RIA as an alternative high-affinity iron acquisition system that is independent of L-ornithine. Furthermore, these studies indicated that SB is mainly fueled by mitochondrial rather than cytosolic ornithine production, at least with L-glutamine as the nitrogen source.

Consistent with mevalonate being a precursor for TAFC biosynthesis (Fig. 2), overexpression of the hydroxymethylglutaryl-CoA (HMG-CoA) reductase Hmg1 increases TAFC production, while inhibition of Hmg1 with lovastatin reduces TAFC biosynthesis in *A. fumigatus*.³² Mevalonate is an intermediate of the isoprenoid biosynthetic pathway with ergosterol as one important product. Ergosterol is the major target of antifungal treatment, *i.e.* amphotericin B and azoles. In contrast to siderophore production, iron starvation decreases the cellular ergosterol level due to the iron-requirement of ergosterol biosynthesis.³² Concordant with bilateral demand for mevalonate, genetic inactivation of mevalonate consumption for TAFC biosynthesis altered the sterol composition and increased lovastatin resistance during iron starvation.³² These data also revealed a novel consequence of statin treatment in fungi that produce siderophores, the synthesis of which depend on HMG-CoA, such as fusarinines and coprogens.

12. The role of siderophores in physiology and interaction with other organisms

In *A. fumigatus*, genetic elimination of extracellular siderophores (Δ sidI, Δ sidH, Δ sidF, and Δ sidD mutants) decreases growth, conidiation and oxidative stress resistance during iron limitation but not during iron sufficiency, which enables compensation by other iron acquisition systems, mainly RIA.²³ In agreement, simultaneous inactivation of RIA and SB blocks growth without high ferrous iron supplementation, which supports growth *via* low-affinity ferrous iron uptake.⁸ Eliminating intracellular siderophores (Δ sidC and Δ sidL, mutants) reduces conidiation and blocks sexual development, as shown in *A. nidulans*, *Cochliobolus heterostrophus* and *Gibberella*

zeae,^{51,85} most likely due to the crucial role of FC in intracellular iron transport from substrate-contacting hyphae into aerial hyphae.⁸⁶ The lack of HFC (Δ sidC and Δ sidL, mutants) decreases the conidial iron content by about 50%, which impairs iron-dependent enzymes such as aconitase and catalase A, and thereby decreases conidial size and conidial resistance to oxidative stress.^{23,35,86} Moreover, the lack of HFC delays germination during iron starvation.²³ Intracellular iron excess, *e.g.* due to derepression of iron uptake caused by SreA-deficiency, induces massive hyphal accumulation of ferri-FC and transcriptional induction of the transporter CccA, which mediates vacuolar iron deposition in both *A. fumigatus* and *A. nidulans*.^{14,51,87} Nevertheless, vacuolar iron deposition rather than FC-mediated iron storage appears to represent the major iron detoxification system.¹⁵ Inactivation of the entire siderophore system (Δ sidA mutant) combines the defects caused by inactivation of either extra- or intracellular SB and renders *A. fumigatus* extremely sensitive to iron starvation.^{8,23}

Both extra- and intracellular siderophores are crucial for virulence as elimination of the entire SB (Δ sidA mutant) results in absolute avirulence of *A. fumigatus* in a murine model of invasive pulmonary aspergillosis,^{8,88} while deficiency in either extracellular (Δ sidI, Δ sidH, Δ sidF or Δ sidD mutants) or intracellular siderophores (Δ sidC mutants) causes partial attenuation of virulence.^{23,32} The fact that mutants lacking extracellular siderophores are not avirulent indicates partial compensation by RIA *in vivo*. Consistently, both RIA and the siderophore system are transcriptionally upregulated during murine infection.⁸⁹ Conidial HFC appears to play a particularly crucial role during initiation of infection because restoration of the conidial HFC content by supplementation with FC during conidiation partially cures the virulence defect of Δ sidA conidia.²³ SidG-deficiency, which eliminates TAFC production with concomitant increase of F_sC production, affects neither growth nor virulence, indicating that the structural differences between these two siderophores are irrelevant in the settings analysed.²³ The fact that *A. fumigatus* siderophores are able to remove iron from host sources, such as transferrin underlines their crucial role in virulence.^{90,91}

During infection, the siderophore system is important not only for extra-, but most likely also for intracellular growth as defects in the siderophore system decrease intracellular growth and survival of *A. fumigatus* after phagocytosis by murine alveolar macrophages.⁹² Consistently, impairment of SB alters the immune response of macrophages after phagocytosis of *A. fumigatus*,⁹³ and the gene encoding SidA was among the highest upregulated genes after internalization of *A. fumigatus* by epithelial cells.⁹⁴ Moreover, the siderophore system was also found to be important for virulence of *Histoplasma capsulatum*, a dimorphic fungal pathogen replicating in the yeast form within macrophages.²⁸ The evolutionary conserved role of siderophores in virulence of *A. fumigatus* has been confirmed in various infection models apart from pulmonary murine infection, *i.e.* a murine cutaneous model, *Drosophila melanogaster* and *Galleria mellonella*.^{95–97}

SB is also a virulence determinant of a number of phytopathogenic ascomycetes with a necrotrophic life style such as *C. heterostrophus*, *Cochliobolus miyabeanus*, *F. graminearum*,



Highlight

A. brassicicola, *Alternaria alternata* and *M. grisea* on different plant hosts.^{25,42,98,99} In contrast, SB is dispensable and RIA is essential for phytopathogenicity of biotrophic *U. maydis*.^{29,100} Recent studies with the hemibiotrophic fungus *C. graminicola* shed light on the apparent discrepancy of the role of siderophores in virulence of necro- and biotrophic pathogens.^{43,101} In this maize pathogen, SB was found to be induced and important for virulence during the necrotrophic phase but to be specifically downregulated during the biotrophic phase, compensated by RIA, in order to repress the plant immune response induced by fungal siderophores.

Consistent with the crucial role of adaptation to iron starvation, including SB as virulence determinant, involved regulators such as HapX and SrbA as well as NRPS-activating Sfp-type 4'-phosphopantetheinyl transferase is essential for virulence of different siderophore producers.^{49,102}

In *Epichloë festucae*, extracellular siderophore biosynthesis is important to maintain mutualistic symbiotic grass-endophyte associations as its inactivation transformed this mutualistic fungus into an antagonistic one.¹⁰³

Apart from their roles in virulence and symbiosis, siderophores might also play a role in competitive and cooperative interactions among microorganisms because chelation of environmental iron by siderophore-types that are not recognized by competitors might be used to starve them for iron. This is counteracted by the evolution of transporters recognizing these xenosiderophores. The latter turns a growth-suppressing into a growth-promoting molecule. This scenario is supported by the fact that most siderophore-producing bacteria and fungi possess xenosiderophore-specific SITs.⁷ The central role of iron and siderophores in microbial interaction is underlined by two recent findings.

Patients with cystic fibrosis contain a mixed microbial community including the bacterium *P. aeruginosa* and *A. fumigatus*. *In vitro* mass spectrometry imaging revealed that *P. aeruginosa* stimulates production by *A. fumigatus* via secretion of 1-hydroxyphenazine TAFC.¹⁰⁴ The mechanism and rationale of this microbial interaction is currently unknown but it underlines the central role of siderophores. Along these lines, HapX-mediated iron regulation was demonstrated to be crucial for iron competition of *F. oxysporum* with siderophores of a biocontrol strain of *Pseudomonas putida*.¹⁰⁵

13. Applied aspects: potential use in treatment and diagnosis of fungal infections

Fungal infections such as aspergillosis are difficult to diagnose and treat, which is reflected by the high mortality rates that continue to be associated with this disease.¹⁰⁶ The crucial role of the siderophore system for fungal virulence and the differences to the mammalian iron acquisition mechanisms might help to improve therapy and diagnosis of fungal infections. Specifically, the fungal siderophore biosynthetic pathway represents a promising target for selective therapeutic intervention. In this respect, the structural elucidation of the *A. fumigatus* SidA

enzyme might facilitate design of selective inhibitors.¹⁰⁷ Recently, treatment of fungal keratitis in a murine model by dual topical therapy with the iron chelator deferiprone and statins, which target biosynthesis of both isoprenoids and extracellular siderophores, showed promising restriction of fungal growth.¹⁰⁸

Furthermore, SITs represent one of few protein families that are unique to the fungal kingdom and are not present in prokaryotes or other eukaryotes.¹⁰⁹ Consequently, this protein family is an attractive drug target by potentially mediating fungal-specific drug delivery by a Trojan horse approach,¹¹⁰ in which antifungal agents are covalently attached to siderophores for selective import into fungi.

Moreover SITs represent an attractive target to improve diagnosis of fungal infections.⁶⁸ Gallium chelated by TAFC or the bacterial siderophore ferrioxamine E was found to enable *in vivo* imaging of aspergillosis in animal models by Positron Emission Tomography (PET) due to selective accumulation in fungal cells.^{111–114}

The understanding of the fungal siderophore system has advanced significantly in recent years. Nevertheless, its potential application in treatment and diagnosis of fungal infections still requires deeper insights.

Moreover, the recombinant *A. fumigatus* enzymes SidI and SidH were employed for biosynthesis of anhydromevalonolacton, an intermediate in the production of bioplastic in *E. coli*, which indicates the biotechnological potential of enzymes involved in the fungal siderophore pathway.¹¹⁵

14. Abbreviations

CBC	CCAAT binding complex
CoA	Coenzyme A
FC	Ferricrocin
FsC	Fusarinine C
GNAT	GCN5-related N-acetyltransferases
HFC	Hydroxyferricrocin
RIA	Reductive iron assimilation
SB	Siderophore biosynthesis
SIT	Siderophore-iron transporter
TAFC	Triacetyl-fusarinine C

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16. References

- 1 K. J. Kwon-Chung and J. A. Sugui, *PLoS Pathog.*, 2013, **9**, e1003743.



- 2 F. Tekaia and J. P. Latge, *Curr. Opin. Microbiol.*, 2005, **8**, 385–392.
- 3 W. C. Nierman, A. Pain, M. J. Anderson, J. R. Wortman, H. S. Kim, J. Arroyo, M. Berriman, K. Abe, D. B. Archer, C. Bermejo, J. Bennett, P. Bowyer, D. Chen, M. Collins, R. Coulsen, R. Davies, P. S. Dyer, M. Farman, N. Fedorova, T. V. Feldblyum, R. Fischer, N. Fosker, A. Fraser, J. L. Garcia, M. J. Garcia, A. Goble, G. H. Goldman, K. Gomi, S. Griffith-Jones, R. Gwilliam, B. Haas, H. Haas, D. Harris, H. Horiuchi, J. Huang, S. Humphray, J. Jimenez, N. Keller, H. Khouri, K. Kitamoto, T. Kobayashi, S. Konzack, R. Kulkarni, T. Kumagai, A. Lafon, J. P. Latge, W. Li, A. Lord, C. Lu, W. H. Majoros, G. S. May, B. L. Miller, Y. Mohamoud, M. Molina, M. Monod, I. Mouyna, S. Mulligan, L. Murphy, S. O'Neil, I. Paulsen, M. A. Penalva, M. Perlea, C. Price, B. L. Pritchard, M. A. Quail, E. Rabinowitsch, N. Rawlins, M. A. Rajandream, U. Reichard, H. Renault, G. D. Robson, S. Rodriguez de Cordoba, J. M. Rodriguez-Pena, C. M. Ronning, S. Rutter, S. L. Salzberg, M. Sanchez, J. C. Sanchez-Ferrero, D. Saunders, K. Seeger, R. Squares, S. Squares, M. Takeuchi, F. Tekaia, G. Turner, C. R. Vazquez de Aldana, J. Weidman, O. White, J. Woodward, J. H. Yu, C. Fraser, J. E. Galagan, K. Asai, M. Machida, N. Hall, B. Barrell and D. W. Denning, *Nature*, 2005, **438**, 1151–1156.
- 4 B. Halliwell and J. M. Gutteridge, *Biochem. J.*, 1984, **219**, 1–14.
- 5 T. Ganz, *Curr. Opin. Immunol.*, 2009, **21**, 63–67.
- 6 E. D. Weinberg, *Biochim. Biophys. Acta*, 2009, **1790**, 600–605.
- 7 H. Haas, M. Eisendle and B. G. Turgeon, *Ann. Rev. Phytopathol.*, 2008, **46**, 149–187.
- 8 M. Schrettl, E. Bignell, C. Kragl, C. Joechl, T. Rogers, H. N. Arst, Jr, K. Haynes and H. Haas, *J. Exp. Med.*, 2004, **200**, 1213–1219.
- 9 C. C. Philpott, *Biochim. Biophys. Acta*, 2006, **1763**, 636–645.
- 10 M. Schrettl, G. Winkelmann and H. Haas, *Biomaterials*, 2004, **17**, 647–654.
- 11 Z. Chi, X. X. Wang, Z. C. Ma, M. A. Buzdar and Z. M. Chi, *Biomaterials*, 2012, **25**, 219–230.
- 12 C. D. Kaplan and J. Kaplan, *Chem. Rev.*, 2009, **109**, 4536–4552.
- 13 M. Eisendle, H. Oberegger, I. Zadra and H. Haas, *Mol. Microbiol.*, 2003, **49**, 359–375.
- 14 M. Schrettl, H. S. Kim, M. Eisendle, C. Kragl, W. C. Nierman, T. Heinekamp, E. R. Werner, I. Jacobsen, P. Illmer, H. Yi, A. A. Brakhage and H. Haas, *Mol. Microbiol.*, 2008, **70**, 27–43.
- 15 F. Gsaller, M. Eisendle, B. E. Lechner, M. Schrettl, H. Lindner, D. Muller, S. Geley and H. Haas, *Metallomics*, 2012, **4**, 1262–1270.
- 16 A. Thieken and G. Winkelmann, *FEMS Microbiol. Lett.*, 1992, **73**, 37–41.
- 17 R. J. Capon, M. Stewart, R. Ratnayake, E. Lacey and J. H. Gill, *J. Nat. Prod.*, 2007, **70**, 1746–1752.
- 18 J. C. Renshaw, G. D. Robson, A. P. J. Trinci, M. G. Wiebe, F. R. Livens, D. Collison and R. J. Taylor, *Mycol. Res.*, 2002, **106**, 1123–1142.
- 19 R. C. Hider and X. Kong, *Nat. Prod. Rep.*, 2010, **27**, 637–657.
- 20 M. B. Hossain, M. A. Jalal and D. van der Helm, *Acta Crystallogr., Sect. C: Cryst. Struct. Commun.*, 1997, **53**(Pt 6), 716–718.
- 21 A. Koulman, T. V. Lee, K. Fraser, L. Johnson, V. Arcus, J. S. Lott, S. Rasmussen and G. Lane, *Phytochemistry*, 2012, **75**, 128–139.
- 22 W. B. Yin, J. A. Baccile, J. W. Bok, Y. Chen, N. P. Keller and F. C. Schroeder, *J. Am. Chem. Soc.*, 2013, **135**, 2064–2067.
- 23 M. Schrettl, E. Bignell, C. Kragl, Y. Sabiha, O. Loss, M. Eisendle, A. Wallner, H. N. Arst, K. Haynes and H. Haas, *PLoS Pathog.*, 2007, **3**, e128.
- 24 H. J. Plattner and H. Diekmann, in *Metal ions in fungi*, ed. G. Winkelmann and D. R. Winge, Marcel Dekker, Inc., New York, N.Y., 1994, pp. 99–117.
- 25 D. Greenshields, L. Guosheng, J. Feng, G. Selvaraj and Y. Wei, *Mol. Plant Pathol.*, 2007, **8**, 411–421.
- 26 O. Yamada, S. Na Nan, T. Akao, M. Tominaga, H. Watanabe, T. Satoh, H. Enei and O. Akita, *J. Biosci. Bioeng.*, 2003, **95**, 82–88.
- 27 C. Hof, K. Eisfeld, L. Antelo, A. J. Foster and H. Anke, *Fungal Genet. Biol.*, 2009, **46**, 321–332.
- 28 L. H. Hwang, J. A. Mayfield, J. Rine and A. Sil, *PLoS Pathog.*, 2008, **4**, e1000044.
- 29 B. Mei, A. D. Budde and S. A. Leong, *Proc. Natl. Acad. Sci. U. S. A.*, 1993, **90**, 903–907.
- 30 P. Visca, A. Ciervo and N. Orsi, *J. Bacteriol.*, 1994, **176**, 1128–1140.
- 31 M. Herrero, V. de Lorenzo and J. B. Neilands, *J. Bacteriol.*, 1988, **170**, 56–64.
- 32 S. Yasmin, L. Alcazar-Fuoli, M. Grundlinger, T. Puempel, T. Cairns, M. Blatzer, J. F. Lopez, J. O. Grimalt, E. Bignell and H. Haas, *Proc. Natl. Acad. Sci. U. S. A.*, 2012, **109**, E497–E504.
- 33 C. M. Roche, H. W. Blanch, D. S. Clark and N. L. Glass, *Eukaryot. Cell*, 2013, **12**, 1244–1257.
- 34 M. Grundlinger, S. Yasmin, B. E. Lechner, S. Geley, M. Schrettl, M. Hynes and H. Haas, *Mol. Microbiol.*, 2013, **88**, 862–875.
- 35 M. Blatzer, M. Schrettl, B. Sarg, H. H. Lindner, K. Pfaller and H. Haas, *Appl. Environ. Microbiol.*, 2011, **77**, 4959–4966.
- 36 V. de Lorenzo, A. Bindereif, B. H. Paw and J. B. Neilands, *J. Bacteriol.*, 1986, **165**, 570–578.
- 37 B. Winterberg, S. Uhlmann, U. Linne, F. Lessing, M. A. Marahiel, H. Eichhorn, R. Kahmann and J. Schirawski, *Mol. Microbiol.*, 2010, **75**, 1260–1271.
- 38 W. M. Yuan, G. D. Gentil, A. D. Budde and S. A. Leong, *J. Bacteriol.*, 2001, **183**, 4040–4051.
- 39 T. Schwecke, K. Gottling, P. Durek, I. Duenas, N. F. Kaufer, S. Zock-Emmenthal, E. Staub, T. Neuhof, R. Diekmann and H. von Dohren, *ChemBioChem*, 2006, **7**, 612–622.
- 40 A. Munawar, J. W. Marshall, R. J. Cox, A. M. Bailey and C. M. Lazarus, *ChemBioChem*, 2013, **14**, 388–394.



- 41 K. E. Bushley, D. R. Ripoll and B. G. Turgeon, *BMC Evol. Biol.*, 2008, **8**, 328.
- 42 S. Oide, W. Moeder, H. Haas, S. Krasnoff, D. Gibson, K. Yoshioka and B. G. Turgeon, *Plant Cell*, 2006, **18**, 2836–2853.
- 43 E. Albarouki, L. Schafferer, F. Ye, N. von Wiren, H. Haas and H. B. Deising, *Mol. Microbiol.*, 2014, **92**, 338–355.
- 44 K. Reiber, E. P. Reeves, C. M. Neville, R. Winkler, P. Gebhardt, K. Kavanagh and S. Doyle, *FEMS Microbiol. Lett.*, 2005, **248**, 83–91.
- 45 W. Steinchen, G. Lackner, S. Yasmin, M. Schrettl, H. M. Dahse, H. Haas and D. Hoffmeister, *Appl. Environ. Microbiol.*, 2013, **79**, 6670–6676.
- 46 G. Allen, M. Bromley, S. J. Kaye, D. Keszenman-Pereyra, T. D. Zucchi, J. Price, M. Birch, J. D. Oliver and G. Turner, *Fungal Genet. Biol.*, 2011, **48**, 456–464.
- 47 H. Oberegger, M. Eisendle, M. Schrettl, S. Graessle and H. Haas, *Curr. Genet.*, 2003, **44**, 211–215.
- 48 Y. Leng and S. Zhong, *Mol. Plant Pathol.*, 2012, **13**, 375–387.
- 49 R. Horbach, A. Graf, F. Weihmann, L. Antelo, S. Mathea, J. C. Liermann, T. Opatz, E. Thines, J. Aguirre and H. B. Deising, *Plant Cell*, 2009, **21**, 3379–3396.
- 50 P. Wiemann, S. Albermann, E. M. Niehaus, L. Studt, K. W. von Barga, N. L. Brock, H. U. Humpf, J. S. Dickschat and B. Tudzynski, *PLoS One*, 2012, **7**, e37519.
- 51 M. Eisendle, M. Schrettl, C. Kragl, D. Muller, P. Illmer and H. Haas, *Eukaryot. Cell*, 2006, **5**, 1596–1603.
- 52 B. F. Matzanke, E. Bill, A. X. Trautwein and G. Winkelmann, *J. Bacteriol.*, 1987, **169**, 5873–5876.
- 53 J. A. Kiel, M. A. van den Berg, F. Fusetti, B. Poolman, R. A. Bovenberg, M. Veenhuis and I. J. van der Klei, *Funct. Integr. Genomics*, 2009, **9**, 167–184.
- 54 H. Haas, M. Schoeser, E. Lesuisse, J. F. Ernst, W. Parson, B. Abt, G. Winkelmann and H. Oberegger, *Biochem. J.*, 2003, **371**, 505–513.
- 55 C. C. Philpott and O. Protchenko, *Eukaryot. Cell*, 2008, **7**, 20–27.
- 56 W. H. Jung and J. W. Kronstad, *Cell. Microbiol.*, 2008, **10**, 277–284.
- 57 T. Nevitt and D. J. Thiele, *PLoS Pathog.*, 2011, **7**, e1001322.
- 58 O. Ardon, H. Bussey, C. Philpott, D. M. Ward, S. Davis-Kaplan, S. Verroneau, B. Jiang and J. Kaplan, *J. Biol. Chem.*, 2001, **276**, 43049–43055.
- 59 I. Raymond-Bouchard, C. S. Carroll, J. R. Nesbitt, K. A. Henry, L. J. Pinto, M. Moinzadeh, J. K. Scott and M. M. Moore, *Eukaryot. Cell*, 2012, **11**, 1333–1344.
- 60 Y. S. Park, T. H. Kim, H. I. Chang, H. C. Sung and C. W. Yun, *Biochem. Biophys. Res. Commun.*, 2006, **345**, 1634–1642.
- 61 C. Kragl, M. Schrettl, B. Abt, B. Sarg, H. H. Lindner and H. Haas, *Eukaryot. Cell*, 2007, **6**, 1278–1285.
- 62 M. Grundlinger, F. Gsaller, M. Schrettl, H. Lindner and H. Haas, *Appl. Environ. Microbiol.*, 2013, **79**, 7534–7536.
- 63 T. J. Brickman and M. A. McIntosh, *J. Biol. Chem.*, 1992, **267**, 12350–12355.
- 64 M. Schrettl, N. Beckmann, J. Varga, T. Heinekamp, I. D. Jacobsen, C. Jochl, T. A. Moussa, S. Wang, F. Gsaller, M. Blatzer, E. R. Werner, W. C. Niermann, A. A. Brakhage and H. Haas, *PLoS Pathog.*, 2010, **6**, e1001124.
- 65 P. Hortschansky, M. Eisendle, Q. Al-Abdallah, A. D. Schmidt, S. Bergmann, M. Thon, O. Kniemeyer, B. Abt, B. Seeber, E. R. Werner, M. Kato, A. A. Brakhage and H. Haas, *EMBO J.*, 2007, **26**, 3157–3168.
- 66 H. Haas, I. Zadra, G. Stoffler and K. Angermayr, *J. Biol. Chem.*, 1999, **274**, 4613–4619.
- 67 L. W. Zhou, H. Haas and G. A. Marzluf, *Mol. Gen. Genet.*, 1998, **259**, 532–540.
- 68 L. H. Hwang, E. Seth, S. A. Gilmore and A. Sil, *Eukaryot. Cell*, 2012, **11**, 16–25.
- 69 N. Zhang, N. A. MohdZainudin, K. Scher, B. J. Condon, B. A. Horwitz and B. G. Turgeon, *Mol. Plant-Microbe Interact.*, 2013, **26**, 1473–1485.
- 70 S. Labbe, B. Pelletier and A. Mercier, *Biometals*, 2007, **20**, 523–537.
- 71 Z. Chi, X. X. Wang, Q. Geng and Z. M. Chi, *Biometals*, 2013, **26**, 955–967.
- 72 C. Voisard, J. Wang, J. L. McEvoy, P. Xu and S. A. Leong, *Mol. Cell. Biol.*, 1993, **13**, 7091–7100.
- 73 J. Amich, L. Schafferer, H. Haas and S. Krappmann, *PLoS Pathog.*, 2013, **9**, e1003573.
- 74 H. Li and C. E. Outten, *Biochemistry*, 2012, **51**, 4377–4389.
- 75 M. Vodisch, K. Scherlach, R. Winkler, C. Hertweck, H. P. Braun, M. Roth, H. Haas, E. R. Werner, A. A. Brakhage and O. Kniemeyer, *J. Proteome Res.*, 2011, **10**, 2508–2524.
- 76 S. D. Willger, S. Puttikamonkul, K. H. Kim, J. B. Burritt, N. Grahl, L. J. Metzler, R. Barbuch, M. Bard, C. B. Lawrence and R. A. Cramer, Jr, *PLoS Pathog.*, 2008, **4**, e1000200.
- 77 C. M. Bien and P. J. Espenshade, *Eukaryot. Cell*, 2010, **9**, 352–359.
- 78 M. Blatzer, B. M. Barker, S. D. Willger, N. Beckmann, S. J. Blosser, E. J. Cornish, A. Mazurie, N. Grahl, H. Haas and R. A. Cramer, *PLoS Genet.*, 2011, **7**, e1002374.
- 79 M. Eisendle, H. Oberegger, R. Buttinger, P. Illmer and H. Haas, *Eukaryot. Cell*, 2004, **3**, 561–563.
- 80 H. Liu, F. N. Gravelat, L. Y. Chiang, D. Chen, G. Vanier, D. E. Ejzykiewicz, A. S. Ibrahim, W. C. Nierman, D. C. Sheppard and S. G. Filler, *Mol. Microbiol.*, 2010, **78**, 1038–1054.
- 81 X. Feng, K. Krishnan, D. L. Richie, V. Aimanianda, L. Hartl, N. Grahl, M. V. Powers-Fletcher, M. Zhang, K. K. Fuller, W. C. Nierman, L. J. Lu, J. P. Latge, L. Woollett, S. L. Newman, R. A. Cramer, Jr, J. C. Rhodes and D. S. Askew, *PLoS Pathog.*, 2011, **7**, e1002330.
- 82 N. Beckmann, L. Schafferer, M. Schrettl, U. Binder, H. Talasz, H. Lindner and H. Haas, *PLoS One*, 2013, **8**, e67426.
- 83 R. Jain, V. Valiante, N. Remme, T. Docimo, T. Heinekamp, C. Hertweck, J. Gershenzon, H. Haas and A. A. Brakhage, *Mol. Microbiol.*, 2011, **82**, 39–53.
- 84 R. E. Frederick, J. A. Mayfield and J. L. DuBois, *J. Am. Chem. Soc.*, 2011, **133**, 12338–12341.



- 85 S. Oide, S. B. Krasnoff, D. M. Gibson and B. G. Turgeon, *Eukaryot. Cell*, 2007, **6**, 1339–1353.
- 86 A. Wallner, M. Blatzer, M. Schrettl, B. Sarg, H. Lindner and H. Haas, *Appl. Environ. Microbiol.*, 2009, **75**, 4194–4196.
- 87 H. Oberegger, M. Schoeser, I. Zadra, B. Abt and H. Haas, *Mol. Microbiol.*, 2001, **41**, 1077–1089.
- 88 A. H. Hissen, A. N. Wan, M. L. Warwas, L. J. Pinto and M. M. Moore, *Infect. Immun.*, 2005, **73**, 5493–5503.
- 89 A. McDonagh, N. D. Fedorova, J. Crabtree, Y. Yu, S. Kim, D. Chen, O. Loss, T. Cairns, G. Goldman, D. Armstrong-James, K. Haynes, H. Haas, M. Schrettl, G. May, W. C. Nierman and E. Bignell, *PLoS Pathog.*, 2008, **4**, e1000154.
- 90 A. H. Hissen, J. M. Chow, L. J. Pinto and M. M. Moore, *Infect. Immun.*, 2004, **72**, 1402–1408.
- 91 A. H. Hissen and M. M. Moore, *J. Biol. Inorg. Chem.*, 2005, **10**, 211–220.
- 92 M. Schrettl, O. Ibrahim-Granet, S. Droin, M. Huerre, J. P. Latge and H. Haas, *Microbes Infect.*, 2010, **12**, 1035–1041.
- 93 M. Seifert, M. Nairz, A. Schroll, M. Schrettl, H. Haas and G. Weiss, *Immunobiology*, 2008, **213**, 767–778.
- 94 J. L. Oosthuizen, P. Gomez, J. Ruan, T. L. Hackett, M. M. Moore, D. A. Knight and S. J. Tebbutt, *PLoS One*, 2011, **6**, e20527.
- 95 R. Ben-Ami, R. E. Lewis, K. Leventakos, J. P. Latge and D. P. Kontoyiannis, *Antimicrob. Agents Chemother.*, 2010, **54**, 1848–1854.
- 96 G. Chamilos, E. M. Bignell, M. Schrettl, R. E. Lewis, K. Leventakos, G. S. May, H. Haas and D. P. Kontoyiannis, *Med. Mycol.*, 2010, **48**, 506–510.
- 97 J. L. Slater, L. Gregson, D. W. Denning and P. A. Warn, *Med. Mycol.*, 2011, **49 Suppl. 1**, S107–113.
- 98 L. H. Chen, C. H. Lin and K. R. Chung, *Mol. Plant Pathol.*, 2013, **14**, 497–505.
- 99 C. Hof, K. Eisfeld, K. Welzel, L. Antelo, A. J. Foster and H. Anke, *Mol. Plant Pathol.*, 2007, **8**, 163–172.
- 100 H. Eichhorn, F. Lessing, B. Winterberg, J. Schirawski, J. Kamper, P. Muller and R. Kahmann, *Plant Cell*, 2006, **18**, 3332–3345.
- 101 E. Albarouki and H. B. Deising, *Mol. Plant Microbe In.*, 2013, **26**, 695–708.
- 102 S. A. Leong and G. Winkelmann, *Met. Ions Biol. Syst.*, 1998, **35**, 147–186.
- 103 L. J. Johnson, A. Koulman, M. Christensen, G. A. Lane, K. Fraser, N. Forester, R. D. Johnson, G. T. Bryan and S. Rasmussen, *PLoS Pathog.*, 2013, **9**, e1003332.
- 104 W. J. Moree, V. V. Phelan, C. H. Wu, N. Bandeira, D. S. Cornett, B. M. Duggan and P. C. Dorrestein, *Proc. Natl. Acad. Sci. U. S. A.*, 2012, **109**, 13811–13816.
- 105 M. S. Lopez-Berges, J. Capilla, D. Turra, L. Schafferer, S. Matthijs, C. Jochl, P. Cornelis, J. Guarro, H. Haas and A. Di Pietro, *Plant Cell*, 2012, **24**, 3805–3822.
- 106 W. J. Steinbach, *PLoS Pathog.*, 2013, **9**, e1003642.
- 107 S. Franceschini, M. Fedkenheuer, N. J. Vogelaar, H. H. Robinson, P. Sobrado and A. Mattevi, *Biochemistry*, 2012, **51**, 7043–7045.
- 108 S. M. Leal, Jr, S. Roy, C. Varechon, S. Carrion, H. Clark, M. S. Lopez-Berges, A. Di Pietro, M. Schrettl, N. Beckmann, B. Redl, H. Haas and E. Pearlman, *PLoS Pathog.*, 2013, **9**, e1003436.
- 109 T. Hsiang and D. L. Baillie, *J. Mol. Evol.*, 2005, **60**, 475–483.
- 110 M. J. Miller, H. Zhu, Y. Xu, C. Wu, A. J. Walz, A. Vergne, J. M. Roosenberg, G. Moraski, A. A. Minnick, J. McKee-Dolence, J. Hu, K. Fennell, E. Kurt Dolence, L. Dong, S. Franzblau, F. Malouin and U. Mollmann, *Biometals*, 2009, **22**, 61–75.
- 111 M. Petrik, G. M. Franssen, H. Haas, P. Laverman, C. Hortnagl, M. Schrettl, A. Helbok, C. Lass-Flörl and C. Decristoforo, *Eur. J. Nucl. Med. Mol. Imaging*, 2012, **39**, 1175–1183.
- 112 M. Petrik, H. Haas, G. Dobrozemsky, C. Lass-Flörl, A. Helbok, M. Blatzer, H. Dietrich and C. Decristoforo, *J. Nucl. Med.*, 2010, **51**, 639–645.
- 113 M. Petrik, H. Haas, P. Laverman, M. Schrettl, G. M. Franssen, M. Blatzer and C. Decristoforo, *Mol. Imaging Biol.*, 2014, **16**, 102–108.
- 114 M. Petrik, H. Haas, M. Schrettl, A. Helbok, M. Blatzer and C. Decristoforo, *Nucl. Med. Biol.*, 2012, **39**, 361–369.
- 115 M. Xiong, D. K. Schneiderman, F. S. Bates, M. A. Hillmyer and K. Zhang, *Proc. Natl. Acad. Sci. U. S. A.*, 2014, **111**, 8357–8362.

