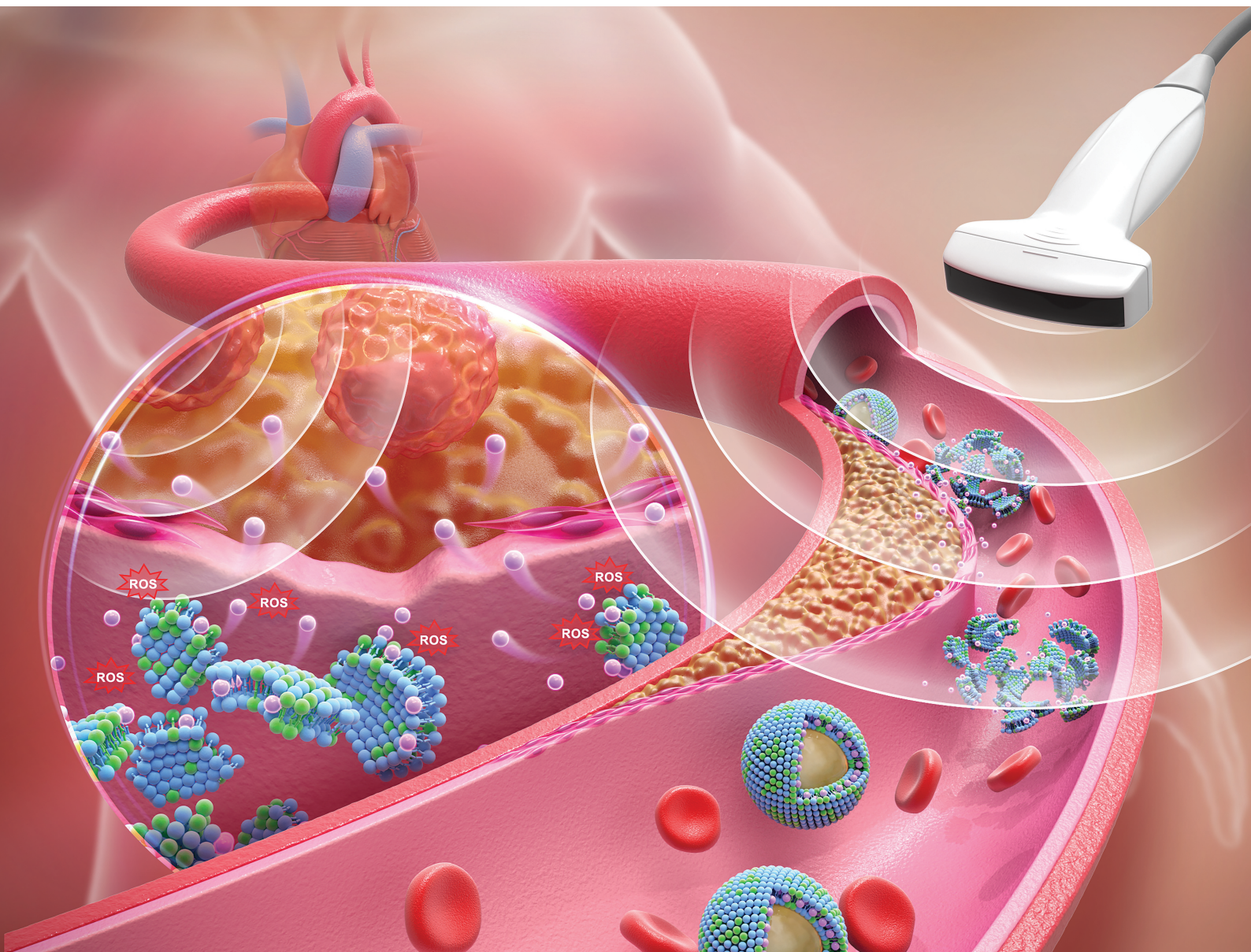


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A study of saponin-encapsulated ultrasound microbubbles $Rb_3NPs@MBs$ for atherosclerosis targeted treatment†

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Atherosclerosis remains a leading disease posing significant threats to human health and life. Oxidative stress plays a critical role in the initiation of early atherosclerosis. Ginsenoside Rb_3 has been shown to exert potential therapeutic effects against atherosclerosis due to its antioxidant properties. However, its clinical utility remains constrained to the nanometer scale, offering insufficient targeting capability for atherosclerosis treatment. To address this limitation, we designed a novel Rb_3 -loaded microbubble system $Rb_3NPs@MBs$. This microbubble system effectively encapsulates Rb_3 nanoparticles and, via ultrasound-targeted microbubble destruction (UTMD), facilitates their targeted accumulation in the aortic arch of atherosclerotic mice. Subsequently, $Rb_3NPs@MBs$ reduce oxidative stress, attenuate endothelial cell apoptosis and foam cell formation, and ultimately diminish plaque development at the lesion site. This strategy holds promise as a therapeutic approach for atherosclerosis. These findings suggest that $Rb_3NPs@MBs$ represent a promising therapeutic strategy for atherosclerosis.

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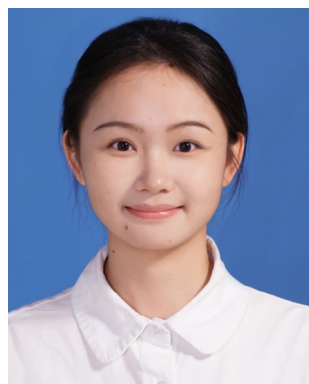
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1. Introduction

Cardiovascular disease (CVD) remains the leading cause of mortality among non-communicable diseases globally, exhibiting a high incidence rate.¹ Atherosclerosis (AS) is characterized by the formation of fibrofatty lesions within the arterial walls, leading to cardiovascular and cerebrovascular conditions such



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as coronary heart disease and stroke. AS serves as the pathological foundation for the onset and progression of CVD.²

AS is a complex, multicellular, and staged disease that typically begins with apoptosis and dysfunction of endothelial cells.³ When vascular endothelial cells are subjected to abnormal shear stress within blood vessels, the intracellular response to this disturbance results in the upregulation and increased activity of vascular cell adhesion molecule-1 (VCAM-1), as well as increased permeability to lipoproteins, particularly to low intensity lipoprotein (LDL) in the circulation.^{4,5} LDL traverses the endothelial barrier and undergoes oxidation to form oxidized LDL (ox-LDL) within the intimal layer of blood vessels, where it accumulates.⁶ Concurrently, monocytes are recruited from the bloodstream into the intimal region, where they differentiate into macrophages.^{7,8} These macrophages take up ox-LDL and transform into foam cells, which accumulate and ultimately form plaques.⁹ In summary, AS is a fibrofatty lesion in the arterial wall mediated by oxidative stress and inflammation.^{10,11} At present, various symptomatic treatments are available for different clinical phenotypes of AS, including antithrombotic drugs, antilipemic agents, and anti-myocardial ischemia medications. Additionally, targeted therapies such as PCSK9 inhibitors and inflammatory factor monoclonal antibodies like Canakinumab have progressed to clinical trial stages.^{12–16}

Ginseng, a traditional medicine in ancient China, is widely used for vascular protection in traditional medicine practices.^{17–19} Saponin Rb₃, an extract derived from ginseng, has been investigated for its potential in treating AS due to its potent antioxidant effects.²⁰ Rb₃ can regulate energy metabolism and apoptosis in cardiomyocytes by activating the peroxisome proliferator-activated receptor alpha (PPAR α) pathway, thereby enhancing cardiovascular function.^{21,22} However, existing studies are limited to the incorporation of Rb₃ into nano-systems and the efficiency of targeted delivery is often suboptimal, particularly in regions with severe vascular plaques or specific localized areas.^{23,24} In order to solve the problem of

drug targeting, this study employed ultrasound-targeted microbubble destruction (UTMD) technology.^{25–27} Typically, microbubbles—vacuole-like structures encapsulating a gas core—are utilized as ultrasound contrast agents.²⁸ By modifying the components of microbubbles, such as replacing the core gas with therapeutic gases like oxygen or carbon monoxide,^{28,29} or by incorporating therapeutic agents (e.g., genes, proteins, and nanomedicines),^{30,31} therapeutic purposes can be achieved. UTMD facilitates the targeted explosion of microbubbles at specific sites under ultrasound guidance.³² The resultant physical effects include the opening of tight junctions between cells, increased tissue permeability, enhanced cellular uptake of drugs, and augmented drug delivery efficiency.^{33–40} As a non-invasive treatment modality,⁴¹ UTMD has been validated for applications such as opening the blood–brain barrier,⁴² enhancing the enhanced permeability and retention (EPR) effect in tumour therapy,^{43,44} and treating AS.^{45–48}

Therefore, in this study, a saponin-encapsulated microbubble system Rb₃NPs@MBs was constructed and combined with UTMD, so that the drug could be enriched in the targeted region of AS mice under ultrasound guidance. Using this drug delivery system, we verified the feasibility of Rb₃NPs@MBs for reducing aortic plaques in AS mice and explored the related mechanism of Rb₃NPs@MBs for the treatment of AS.

2. Materials and methods

2.1. Materials

1,2-Distearoyl-*sn*-glycero-3-phosphoethanolamine-*N*-[amino (polyethylene glycol)-2000] (DSPE-PEG2000) and 1,2-distearoyl-*sn*-glycero-3-phosphocholine (DSPC) were purchased from Avanti (816-94-4 & 474922-77-5, USA). Saponin Rb₃ was obtained from Chengdu Pufei (JOT-10260, China). Lipopolysaccharides (LPS) were obtained from Solarbio (L8880, China). Dulbecco's minimum essential medium (DMEM) and fetal bovine serum were purchased from Gibco (USA). Endothelial cell medium (ECM) was obtained from ScienCell (1001, USA). The mouse macrophage cell line (RAW264.7) was obtained from the Laboratory Center of Shenzhen University; the human umbilical vein endothelial cell line (HUVEC) was obtained from BeNa Culture Collection (BNCC378266, China).

2.2. Preparation and characterization of Rb₃NPs@MBs

DSPC and DSPE-PEG2000 were dissolved in chloroform at 7.90 mg ml⁻¹ and 29.92 mg ml⁻¹ respectively. Rb₃ was dissolved in a solvent mixture of chloroform and methanol (1 : 1, v/v) at a concentration of 2 mg ml⁻¹. To synthesize Rb₃NPs@MBs, 272 μ l of DSPC, 30 μ l of DSPE-PEG2000, and 500 μ l of Rb₃ were combined in a round-bottom flask and subjected to rotary evaporation in a fume hood for 40 min to form a lipid film. The resultant lipid film was hydrated with 1 ml of phosphate-buffered saline (PBS) and ultrasonicated in a water bath maintained at 65 °C for 5–6 min. The hydrodynamic diameter of Rb₃NPs@Lipos was measured using dynamic light



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scattering (DLS) with a Zetasizer Nano (Malvern, England), and the structural morphology was characterized by transmission electron microscopy (TEM).

Subsequently, the $\text{Rb}_3\text{NPs@Lipos}$ solution was transferred to a vial connected for gas exchange *via* syringes through rubber caps to produce $\text{Rb}_3\text{NPs@MBs}$. Initially, the gas within the vial was evacuated by vacuuming for 5 min, followed by the introduction of perfluoropropane (C_3F_8) for 1 min to facilitate gas mixing. To ensure complete gas exchange, this procedure was repeated 3–5 times. Subsequently, additional C_3F_8 was added, and the mixture was agitated using a shaker for 45 seconds to form $\text{Rb}_3\text{NPs@MBs}$. The size, particle size distribution, and concentration of $\text{Rb}_3\text{NPs@MBs}$ were quantified using a particle analyzer (AccuSizer 780AD, USA) prior to structural examination under an optical microscope.

Conventional microbubbles (MBs) were prepared similarly to $\text{Rb}_3\text{NPs@MBs}$, excluding the addition of Rb_3 . After establishing a standard curve for Rb_3 *via* ultraviolet (UV) spectrophotometry, uncoated and coated Rb_3 were separated by centrifuging the $\text{Rb}_3\text{NPs@MBs}$ solution (5000 rpm for 10 min) in an ultrafiltration tube. The encapsulation efficiency and drug loading of $\text{Rb}_3\text{NPs@MBs}$ were subsequently determined using UV spectrophotometry.

To assess blood compatibility, the hemolysis rate (HR) of $\text{Rb}_3\text{NPs@MBs}$ after contact with blood was calculated. After the whole blood of fresh heparin sodium anticoagulated C57BL/6J mice was centrifuged and washed, the red blood cells were resuspended in a 2% concentration red blood cell suspension in PBS. According to the concentration gradient (10, 50, 100, and 1000 $\mu\text{g ml}^{-1}$), $\text{Rb}_3\text{NPs@MBs}$ and the red blood cell suspension were gently mixed (volume ratio 1:1), and each concentration was divided into two groups: the UTMD group (given ultrasound operation) and the free group (without ultrasound operation), and incubated for 1 h at 37 °C. NS and the red blood cell suspension (volume ratio 1:1) were used as negative controls, and ddH_2O and the red blood cell suspension (volume ratio 1:1) were used as positive controls. The ultrasonic conditions were as follows: output frequency: 1.0 MHz; duty cycle: 20%; and pulse frequency: 100 Hz. The output sound intensity was 100 mW cm^{-2} . The time was 1 min. At the end of the incubation, the mixture was centrifuged at 3000 rpm for 10 min and the supernatant was removed. The supernatant was transferred to a 96-well plate, and the absorbance value at the specific wavelength of 541 nm of hemoglobin was measured using a microplate reader. Each well was set up with three repeat wells. The HR was calculated using the following formula:

$$\text{HR}(\%) = \frac{\text{OD}_{\text{test}} - \text{OD}_{\text{negative control}}}{\text{OD}_{\text{positive control}} - \text{OD}_{\text{negative control}}} \times 100\%.$$

2.3. Evaluation of the therapeutic effect *in vitro*

RAW264.7 cells were cultured in DMEM with 10% FBS and 1% penicillin/streptomycin (1% penicillin and 1% streptomycin). HUVEC cells were cultured in ECM with 10% FBS, 1% ECGs, and 1% penicillin/streptomycin (1% penicillin and 1% strepto-

mycin). Cells were maintained in an incubator at 37 °C under a humidified atmosphere and 5% CO_2 .

2.3.1. *In vitro* biocompatibility assay of $\text{Rb}_3\text{NPs@MBs}$. To evaluate cell viability, RAW264.7 cells and HUVEC cells were seeded in 96-well plates at a density of 2×10^4 cells per well. The cells were treated with varying concentrations of $\text{Rb}_3\text{NPs@MBs}$ (0, 1, 5, 10, 25, 50, and 100 $\mu\text{g ml}^{-1}$) in combination with UTMD. Ultrasound was induced using a LIPUSTIM® device (SX Ultrasonic, China) with the following parameters: frequency of 1.0 MHz, duty cycle of 50%, pulse rate of 1000 Hz, intensity of 500 mW cm^{-2} , and duration of 3 min. Subsequently, the cells were incubated for 24 h. Cell viability across different concentration groups was then assessed using the CCK-8 assay kit (Biosharp, China).

2.3.2. Selection of *in vitro* $\text{Rb}_3\text{NPs@MBs}$ therapeutic concentrations. To determine the optimal concentration of $\text{Rb}_3\text{NPs@MBs}$ to be used in *in vitro* experiments, confocal microscopy was utilized to evaluate the effect of varying drug concentrations on the scavenging of LPS-induced reactive oxygen species (ROS). After 24 h of incubation, the original medium was aspirated, and the cells were treated with different concentrations of $\text{Rb}_3\text{NPs@MBs}$ (0, 5, 10, 25, and 50 $\mu\text{g ml}^{-1}$) in combination with UTMD using the following settings: frequency of 1.0 MHz, duty cycle of 20%, pulse rate of 100 Hz, intensity of 100 mW cm^{-2} , and duration of 1 min. Subsequently, the cells were co-cultured with 1 $\mu\text{g ml}^{-1}$ LPS for 6 hours. After treatment, the cells were collected and incubated at 37 °C for 30 min using 10 $\mu\text{mol L}^{-1}$ of ROS probes (Invitrogen, USA). Then, the cells were stained with DAPI (Beyotime, China) for observation under a confocal microscope (LSM719, Carl Zeiss Jena, Germany).

2.3.3. Assessment of ultrasound enhanced cellular uptake. To explore whether UTMD promoted the uptake of $\text{Rb}_3\text{NPs@MBs}$ by RAW264.7 cells and HUVEC cells, Nile red (NR) fluorescent dye was used to replace Rb_3NPs to prepare fluorescent microbubbles NRNPs@MBs . Flow cytometry was used to quantitatively measure the uptake of fluorescence by cells at different time points.

The NRNPs concentration, NRNPs@MBs preparation, and subsequent characterization of the NRNPs@MBs were the same as that in section 2.2. RAW264.7 cells and HUVEC cells in good growth condition were plated in 6-well plates, and 2 ml of $1 \times 10^5 \text{ ml}^{-1}$ cell suspension was added to each well and cultured for 24 h. The cells were divided into the control group, NRNPs group (10 $\mu\text{g ml}^{-1}$), and $\text{NRNPs@MBs} + \text{UTMD}$ (10 $\mu\text{g ml}^{-1}$) group, and the UTMD parameters were the same as those in section 2.3.2. After treatment, the cells were collected at different time points (30 min, 1 h, 3 h, 6 h, 12 h, and 24 h), washed with PBS three times, and centrifuged at 1000 rpm for 5 min each time. After the last wash, the cells were resuspended in 500 μl PBS for flow cytometry, and the fluorescence channel was the PE channel.

2.3.4. Examination of DiI-ox-LDL uptake of RAW264.7 cells. RAW264.7 cells were seeded in a confocal dish at a density of 1×10^5 cells per well. After 24 h of culture, the original medium was removed. The cells were divided into five



2.4.2 Validation of aortic arch enrichment and 24 h metabolism of Rb₃NPs@MBs + UTMD. To validate that the combined application of Rb₃NPs@MBs and UTMD facilitates targeted disruption and drug enrichment within the target area, *in vivo* small animal imaging was conducted using an IVIS Lumina LT system (PerkinElmer). Since Rb₃NPs@MBs are inherently non-fluorescent, Cy7.5 maleimide (MedChemExpress, USA) was employed as a fluorescent substitute for Rb₃ to fabricate Cy7.5@MBs. The experiments were divided into three groups ($n = 3$): control group (normal saline), Cy7.5 group (Cy7.5-NS), and Cy7.5@MBs + UTMD group (Cy7.5@MBs and UTMD). The drug dosage and UTMD procedures for all experimental groups were consistent with those described in section 2.4.1.

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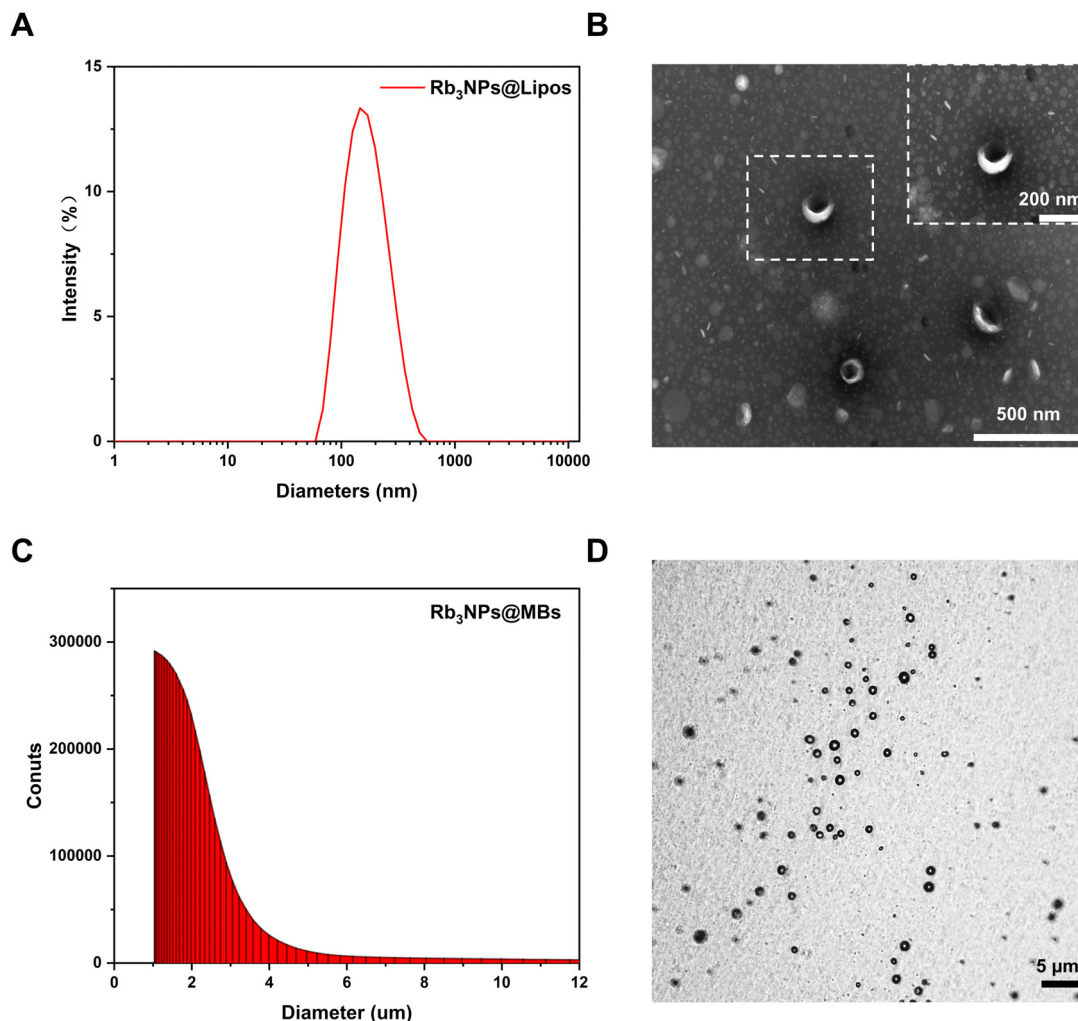


Fig. 1 The characterization of Rb₃NPs@Lipos and Rb₃NPs@MBs. (A) The particle size of Rb₃NPs@Lipos was measured by DLS. (B) The morphological structure of Rb₃NPs@Lipos was observed by TEM. (C) The concentration and size distribution of Rb₃NPs@MBs were measured by PSS. (D) The structures of Rb₃NPs@MBs were observed light microscope.

in fluorescence intensity between the NRNPs group and the NRNPs@MBs + UTMD group at 24 h.

In summary, UTMD promoted the uptake of fluorescence, and it can also be assumed that when NR was replaced by Rb₃NPs, UTMD would also promote the uptake of the drug.

3.3 The therapeutic effect of Rb₃NPs@MBs *in vitro*

In order to elucidate the therapeutic effects of the prepared formulation on early AS events, RAW264.7 cells and HUVECs were employed *in vitro*. Additionally, a microbubble-only (MBs) group was included to confirm that any therapeutic benefits were attributable to Rb₃NPs@MBs rather than the mechanical effects associated with microbubble destruction.

First, a CCK-8 assay was performed to assess the impact of UTMD combined with Rb₃NPs@MBs on cell viability. As a widely used assay reflecting proliferative capacity, the CCK-8 results revealed that the combination of UTMD and Rb₃NPs@MBs at concentrations ranging from 1 to 100 μg ml⁻¹

did not significantly affect RAW264.7 cells and HUVEC viability (Fig. 3A).

Subsequently, confocal microscopy was utilized to examine the intracellular ROS-scavenging capacity of Rb₃NPs@MBs at various concentrations in order to determine the optimal concentration for *in vitro* experiments. Here, LPS was used to induce an inflammatory response, reflecting its well-established role as a cost-effective, highly reproducible stimulus for creating *in vitro* inflammatory disease models, including those relevant to AS.⁵¹ Confocal images showed that when the treatment of Rb₃NPs@MBs combined with UTMD was absent (0 μg ml⁻¹, only LPS treatment), green fluorescence indicative of ROS was most pronounced. Among the 5–50 μg ml⁻¹ Rb₃NPs@MBs treatment groups, the 10 and 25 μg ml⁻¹ conditions demonstrated the most substantial reduction in fluorescence (Fig. 3B), with subsequent ImageJ-based quantitative analysis corroborating that 10 and 25 μg ml⁻¹ yielded optimal ROS-scavenging activity (Fig. 3C). Based on the principle of



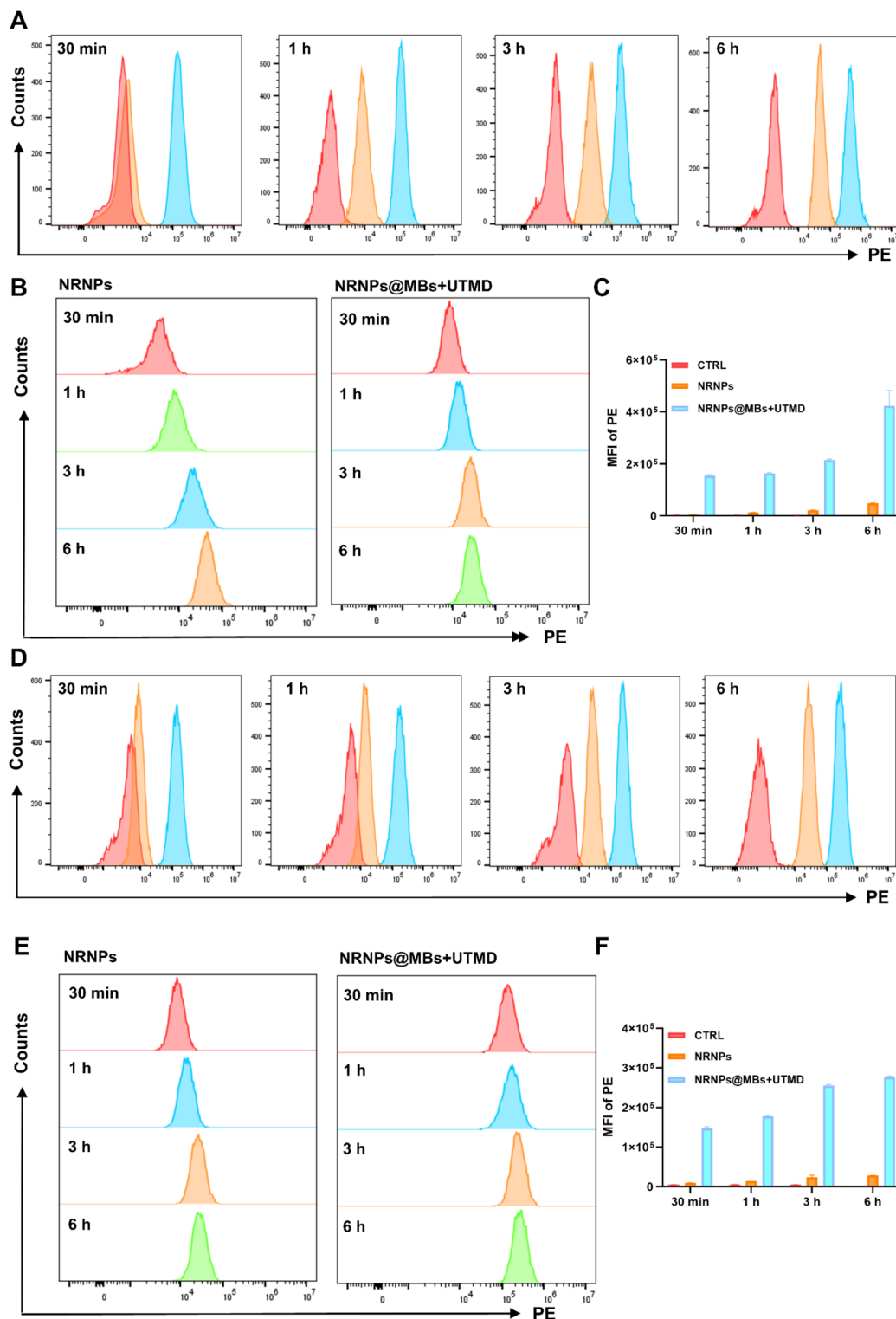


Fig. 2 Cellular uptake at different time points was determined by flow cytometry. (A) Fluorescence flow diagram of RAW264.7 cells at 30 min, 1 h, 3 h, and 6 h, and the group color corresponds to the legend color in (C). (B) Comparison of fluorescence flow images of RAW264.7 cells with the NRNPs group and NRNPs@MBs + UTMD group at different time points. (C) Quantitative analysis of flow fluorescence intensity in (A) ($n = 3$). (D) Fluorescence flow diagram of HUVEC cells at 30 min, 1 h, 3 h, and 6 h, and the grouping colors correspond to the legend colors in (F). (E) Comparison of fluorescence flow images of HUVEC cells at different time points between the NRNPs group and the NRNPs@MBs + UTMD group. (F) Quantitative analysis of flow fluorescence intensity in (D) ($n = 3$).



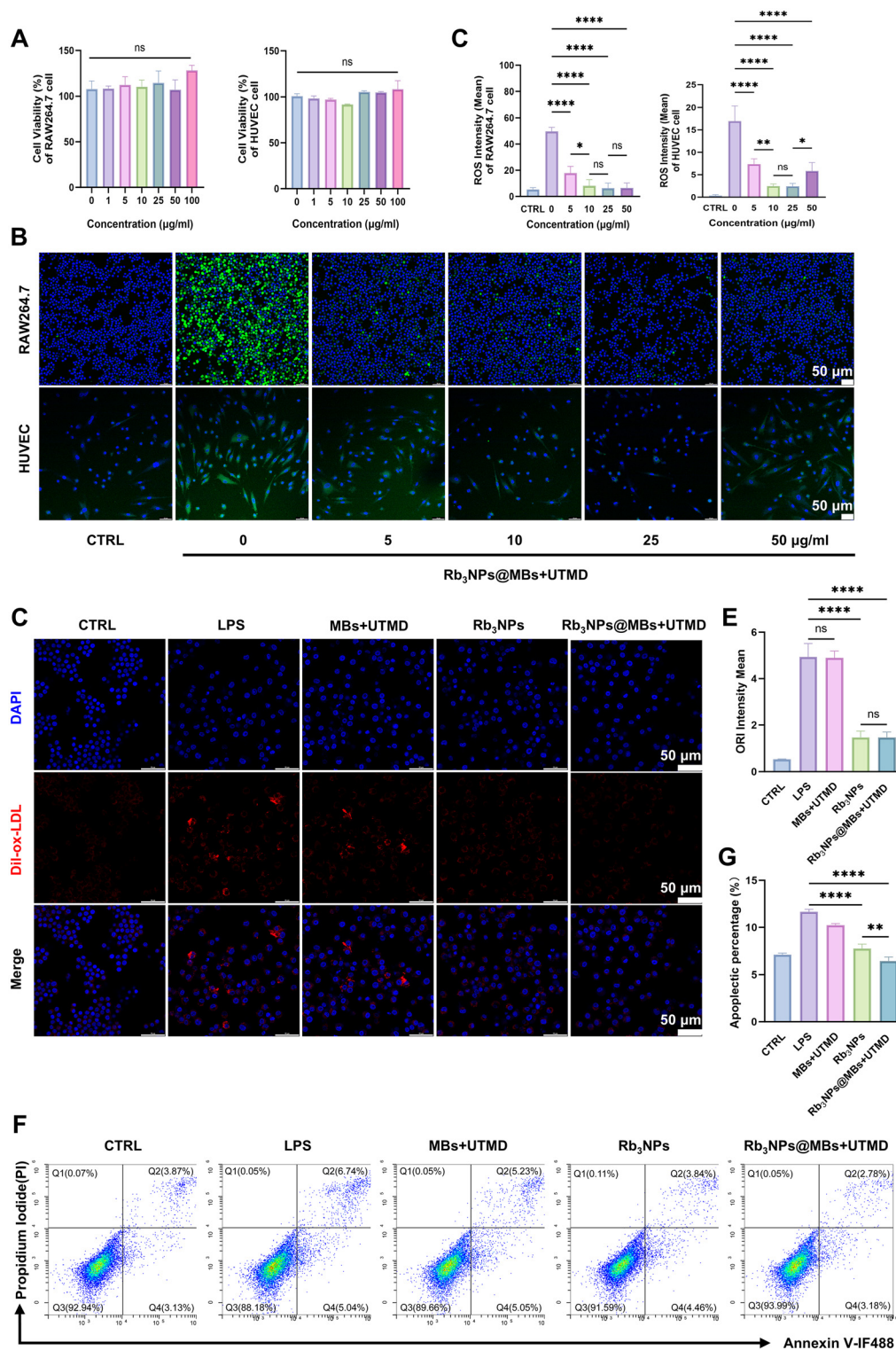
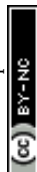


Fig. 3 The therapeutic effect of Rb₃NPs@MBs *in vitro*. (A) The effect on cell viabilities were evaluated by the CCK-8 assay ($n = 3$). The concentration of Rb₃NPs@MBs from 1 to 100 $\mu\text{g ml}^{-1}$. (B) Confocal microscopy was used to observe intracellular ROS fluorescence intensity. Green fluorescence represents ROS and blue fluorescence represents cell nuclei. Scale bar = 50 μm . (C) Quantification of ROS fluorescence intensity in (B) ($n = 3$). (D) The uptake of Dil-ox-LDL by RAW264.7 cells was observed by confocal microscopy. Red fluorescence represents Dil-ox-LDL, and blue fluorescence represents the cell nucleus. Scale bar = 50 μm . (E) Quantitative analysis of Dil-ox-LDL fluorescence intensity in (D) ($n = 3$). (F) The apoptosis of HUVEC cells was analyzed by flow cytometry. (G) Statistical analysis of the proportion of apoptosis in (F) ($n = 3$).



administering the minimum effective dose, $10\ \mu\text{g ml}^{-1}$ was chosen for all subsequent *in vitro* experiments.

To investigate whether $\text{Rb}_3\text{NPs@MBs}$ could diminish ox-LDL uptake by RAW264.7 cells and thereby attenuate foam cell formation, cells were incubated with Dil-ox-LDL, an oxidized low-density lipoprotein labeled red fluorescence.⁵² Confocal microscopy and quantitative fluorescence analysis revealed that MBs combined with UTMD did not significantly reduce Dil-ox-LDL uptake compared to the LPS-only group. In contrast, both the Rb_3NPs and the $\text{Rb}_3\text{NPs@MBs} + \text{UTMD}$ treatments exhibited a marked decrease in red fluorescence (Fig. 3D and E), indicating a significant inhibition of ox-LDL uptake and foam cell formation. Notably, no statistically significant difference was observed between the Rb_3NPs and $\text{Rb}_3\text{NPs@MBs} + \text{UTMD}$ groups. Then, the effect of $\text{Rb}_3\text{NPs@MBs}$ on HUVEC apoptosis was examined. Flow cytometry demonstrated that both Rb_3NPs and $\text{Rb}_3\text{NPs@MBs} + \text{UTMD}$ treatments reduced LPS-induced HUVEC apoptosis, with a more pronounced anti-apoptotic effect observed in the $\text{Rb}_3\text{NPs@MBs} + \text{UTMD}$ group (Fig. 3F and G). These findings suggest that the combination of $\text{Rb}_3\text{NPs@MBs}$ and UTMD effectively mitigates two hallmarks of early AS events—foam cell formation and endothelial cell apoptosis.

Moreover, the therapeutic benefit in HUVECs was superior when Rb_3NPs were delivered *via* MBs combined with UTMD relative to Rb_3NPs alone.

3.4 $\text{Rb}_3\text{NPs@MBs}$ reduce oxidative stress in cells

Numerous studies have demonstrated that oxidative stress plays a pivotal role in the pathogenesis of AS. As an inflammatory disease, the entire progression of AS is often accompanied by an imbalance between intracellular oxidative and antioxidant mechanisms. To determine whether $\text{Rb}_3\text{NPs@MBs}$ can modulate key cellular events through the reduction of oxidative stress, we performed a series of experiments to assess the oxidative stress levels in HUVECs and RAW264.7 cells.

First, in HUVECs, we assessed mitochondrial membrane potential ($\Delta\Psi_m$) using JC-1 staining. Mitochondria constitute the primary center of cellular energy and metabolism, with the mitochondrial respiratory chain serving as a principal source of reactive oxygen species (ROS).⁵³ The extent of mitochondrial mass is closely linked to ROS levels, and the decline in $\Delta\Psi_m$ reflects mitochondrial impairment. Under conditions of high mitochondrial membrane potential, the JC-1 dye accumulates within the mitochondrial matrix, forming aggregates that exhibit red fluorescence. Conversely, when the membrane potential is low, JC-1 remains in its monomeric form and produces green fluorescence. In this study, we observed that the green fluorescence intensity in the LPS group was comparable to that of the CCCP (positive control) and MBs + UTMD groups, whereas the $\text{Rb}_3\text{NPs@MBs} + \text{UTMD}$ groups displayed markedly weaker green fluorescence (Fig. 4A). These findings indicate that $\text{Rb}_3\text{NPs@MBs} + \text{UTMD}$ effectively mitigated the LPS-induced reduction in mitochondrial membrane potential in HUVECs.

In RAW264.7 cells, intracellular ROS levels were quantified by flow cytometry. As shown in Fig. 4B and C, the MBs + UTMD treatment alone did not attenuate LPS-induced ROS generation, while both Rb_3NPs and $\text{Rb}_3\text{NPs@MBs} + \text{UTMD}$ substantially reduced ROS levels. This result is consistent with our previous observations. Notably, no significant difference was detected between the Rb_3NPs and $\text{Rb}_3\text{NPs@MBs} + \text{UTMD}$ groups in terms of ROS suppression.

Finally, we measured superoxide dismutase (SOD) and glutathione (GSH) levels in HUVECs and RAW264.7 cells using commercial assay kits. SOD is a major antioxidant enzyme responsible for catalyzing the dismutation of superoxide anion radicals (O_2^-) into oxygen and hydrogen peroxide (H_2O_2), while GSH is a key antioxidant that directly scavenges free radicals and participates in multiple antioxidant reactions. Intracellular antioxidant capacity can therefore be evaluated by measuring both SOD and GSH. In HUVECs, the $\text{Rb}_3\text{NPs@MBs} + \text{UTMD}$ group exhibited significantly higher SOD and GSH levels compared with the LPS group, and this effect exceeded that observed in the Rb_3NPs group. Similarly, in RAW264.7 cells, $\text{Rb}_3\text{NPs@MBs} + \text{UTMD}$ markedly enhanced SOD and GSH content relative to the LPS group, although no statistically significant difference was found between the Rb_3NPs and $\text{Rb}_3\text{NPs@MBs} + \text{UTMD}$ groups (Fig. 4D).

Collectively, these data suggest that $\text{Rb}_3\text{NPs@MBs}$ effectively mitigate oxidative stress in both RAW264.7 and HUVEC cells. However, whether the combined delivery of $\text{Rb}_3\text{NPs@MBs}$ and UTMD confers additional therapeutic benefits in RAW264.7 cells requires further investigation.

3.5 Biodistribution of microbubbles combined with UTMD *in vivo*

The effects of 2 weeks $\text{Rb}_3\text{NPs@MBs} + \text{UTMD}$ on liver and kidney biochemical indicators (ALT, AST, BUN, and CRE), blood cell count (white blood cell count, neutrophil count, lymphocyte count, and monocyte count) and important organs (heart, liver, spleen, lungs, and kidneys) of mice were not different from those of normal mice. We then set out to explore whether UTMD similarly promotes drug absorption *in vivo* (Fig. S7†). Ultrasound is a commonly used imaging modality, particularly valuable for vascular imaging. It enables the visualization of multidimensional parameters, such as hemorheology, vascular plaque formation, and vascular stenosis.⁵⁴ Among various ultrasound-based techniques, UTMD has emerged as a promising drug delivery strategy, owing to its safety, noninvasive nature, ease of operation, and high targeting efficiency.

To evaluate the *in vivo* targeting efficiency of UTMD and the distribution and metabolism of microbubbles, we substituted Rb_3NPs with Cy7.5 (a fluorescent probe) and prepared Cy7.5@MBs following the same protocol and concentration used for $\text{Rb}_3\text{NPs@MBs}$. The UTMD-induced disruption process of Cy7.5@MBs is depicted in Scheme 1, while the operational details are demonstrated in Video 1 of the ESI.† Briefly, after locating the aortic arch *via* ultrasound, Cy7.5@MBs were administered through the tail vein of AS mice. When the microbubbles had adequately filled the vessels and enhanced the ultrasound



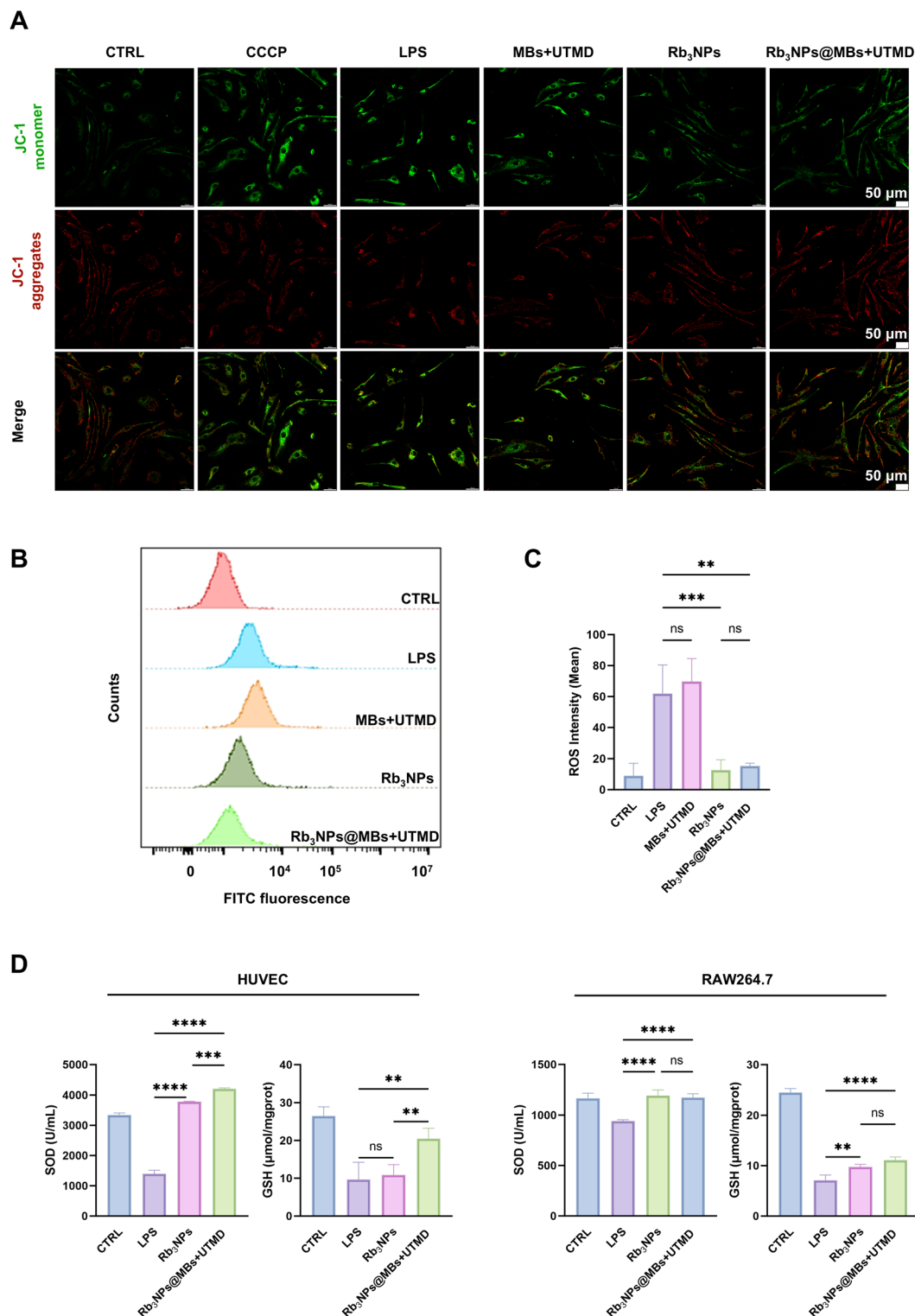
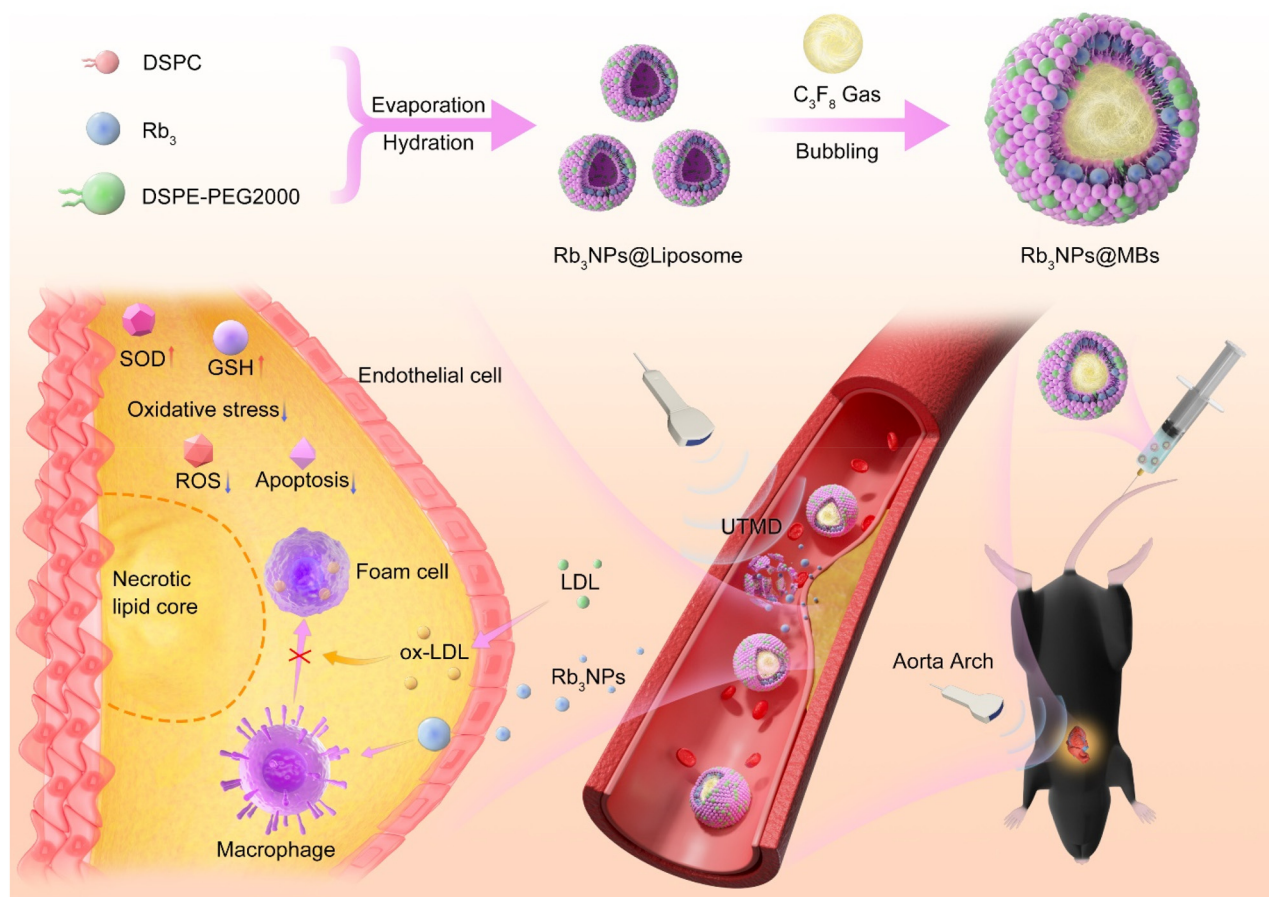


Fig. 4 Rb₃NPs@MBs reduce oxidative stress in cells. (A) JC-1 staining of HUVEC cells. The CCCP group is the positive control group. Green fluorescence represents the JC-1 monomer and red fluorescence represents JC-1 aggregates ($n = 3$). Scale bar = 50 μm . (B) ROS flow analysis in RAW264.7 cells. (C) Quantitative analysis of the data in B ($n = 3$). (D) SOD and GSH oxidation factors in HUVEC cells and RAW264.7 cells ($n = 3$).

contrast, UTMD was applied at the aortic arch. One hour later, *in vivo* fluorescence imaging was performed ($n = 3$), and after 24 hours of circulation, the major organs were collected for

ex vivo imaging ($n = 3$). As shown in Fig. 4A, no fluorescence was detected in the aortic arch for either the control or the Cy7.5 groups, whereas strong fluorescence signals were observed





Scheme 1 The scheme of $Rb_3NPs@MBs$ combined with UTMD for the targeted treatment of atherosclerosis. The $Rb_3NPs@MBs$ burst in the targeted aortic arch under the guidance of ultrasound, and the energy generated by the explosion promotes drug penetration into the plaque, reduces the degree of oxidative stress in the plaque, decreases ROS, and upregulates the oxidation factors SOD and GSH, reduces the apoptosis of endothelial cells and the formation of foam cells, and ultimately reduces the plaque area and increases the stability of the plaque.

in the aortic arch of the $Cy7.5@MBs$ + UTMD group. This conclusion was also verified by fluorescent section staining of the aortic arch (Fig. S8†). After 24 hours of circulation, fluorescence imaging of the major organs revealed that, in the $Cy7.5$ group, the liver exhibited the highest fluorescence intensity, whereas the $Cy7.5@MBs$ + UTMD group showed minimal fluorescence in both the liver and the lungs (Fig. 4B).

Moreover, the overall fluorescence intensity in these organs was substantially lower than that in the $Cy7.5$ group. These findings suggest reduced hepatic retention of $Cy7.5$ in the $Cy7.5@MBs$ + UTMD group, while residual fluorescence in the lungs confirms that microbubbles with small diameters can be metabolized *via* pulmonary clearance. Notably, the fluorescence indicated that UTMD successfully concentrated $Cy7.5$ at the ultrasound-targeted site and demonstrated the feasibility of this targeted delivery approach to overcome blood circulation.

3.6 $Rb_3NPs@MBs$ in combination with UTMD targeted reduction of arterial plaques in AS mice

Apolipoprotein E knockout ($ApoE^{-/-}$) mice, also referred to as APOE-deficient mice, are widely recognized as a classical

model for investigating AS.^{55,56} Accordingly, five- to six-week-old $ApoE^{-/-}$ mice were maintained on a high-fat diet for 12 weeks to establish an AS model. In this study, simvastatin (SV) was selected as the positive control drug, given that statins remain the first-line therapy for AS. Additionally, to confirm that any therapeutic benefit did not result solely from the mechanical effects of MBs under ultrasound disruption, an MB-only group was included. Thus, C57BL/6 mice were assigned to the control group, whereas AS mice were allocated to the following groups, each containing eight animals: model, MBs + UTMD, SV, Rb_3NPs , and $Rb_3NPs@MBs$ + UTMD. After 12 weeks of high-fat feeding, treatments were initiated, and following two weeks of intervention, the mice were euthanized for subsequent analyses (Fig. 6A).

3.6.1 Oil red O staining of aortas. Oil red O staining was performed on the entire aorta. Under the same dose and treatment frequency, the $Rb_3NPs@MBs$ + UTMD group exhibited the smallest plaque area in the whole aorta, followed by the Rb_3NPs group. By contrast, the plaque areas in the MBs + UTMD group and the SV group did not significantly differ from the model group (Fig. 6B and D). Further analysis of the



aortic arch, where UTMD was targeted, showed no difference in plaque area between the Rb_3NPs group and the model group, whereas the $\text{Rb}_3\text{NPs@MBs}$ + UTMD group displayed a marked reduction in plaque area relative to both the model and Rb_3NPs groups (Fig. 6C and D). Hence, oil red O analysis for both the aorta and the aortic arch demonstrated that $\text{Rb}_3\text{NPs@MBs}$ + UTMD specifically reduced the plaque area in the targeted region and exhibited stronger anti-atherosclerotic efficacy compared with Rb_3NPs alone, with a targeting efficiency of 31.9%.

3.6.2 Evaluation of serum inflammatory factors, oxidative stress, and lipids. AS is a systemic inflammatory disease, and quantifying inflammatory factors helps determine disease severity. Enzyme-linked immunosorbent assay (ELISA) results showed that IL-6 levels were significantly lower in the $\text{Rb}_3\text{NPs@MBs}$ + UTMD group than in the model group, with a more pronounced decrease than in the Rb_3NPs group. Concurrently, TGF- β levels in the $\text{Rb}_3\text{NPs@MBs}$ + UTMD group were significantly higher than those in the model group (Fig. 6E). Taken together, these findings reinforce the anti-inflammatory effects of $\text{Rb}_3\text{NPs@MBs}$ + UTMD on AS.

SOD and GSH are key intracellular antioxidants, whereas malondialdehyde (MDA), a lipid peroxidation byproduct, is an indicator of oxidative stress. Biochemical assays demonstrated that SOD and GSH concentrations in the $\text{Rb}_3\text{NPs@MBs}$ + UTMD group were significantly elevated compared with the model group, whereas MDA levels were markedly reduced, notably surpassing the reduction observed in the Rb_3NPs group (Fig. 6F). These results indicate that $\text{Rb}_3\text{NPs@MBs}$ + UTMD ameliorates oxidative stress by increasing levels of the antioxidant factors SOD and GSH while decreasing MDA.

Next, to examine lipid profiles—a key reflection of abnormal lipid metabolism in AS—serum levels of triglycerides (TG), cholesterol (CHO), LDL, and HDL were measured. Levels of these lipids were significantly reduced in the $\text{Rb}_3\text{NPs@MBs}$ + UTMD group compared with the model group (Fig. 6G), confirming that $\text{Rb}_3\text{NPs@MBs}$ + UTMD can effectively modulate dyslipidaemia in AS.

3.6.3 Pathological staining of aortic plaques. Further pathological staining of the aortic plaques was performed to elucidate the direct effects of the treatment on plaque characteristics. VCAM-1, an endothelial adhesion molecule induced upon endothelial cell activation, is a pivotal factor in AS pathogenesis and a prime target for anti-AS therapies. Immunohistochemical staining showed that the VCAM-1-positive area was minimized in the $\text{Rb}_3\text{NPs@MBs}$ + UTMD group (Fig. 7A), indicating significantly downregulated VCAM-1 expression. Oil red O staining of the aortic valve plaques revealed that the $\text{Rb}_3\text{NPs@MBs}$ + UTMD group exhibited the smallest positively stained area (Fig. 7B), suggesting a notable reduction in plaque size following treatment. Masson staining demonstrated a higher proportion of blue-stained collagen in the plaques of the $\text{Rb}_3\text{NPs@MBs}$ + UTMD group, indicative of enhanced plaque stability. HE staining likewise confirmed the reduced aortic root plaque area under $\text{Rb}_3\text{NPs@MBs}$ + UTMD treatment.

Collectively, these data demonstrate that $\text{Rb}_3\text{NPs@MBs}$ + UTMD successfully released the drug at the aortic arch targeted by ultrasound, thereby reducing local plaque burden, downregulating VCAM-1 expression, mitigating inflammation, and lowering both lipid and oxidative stress levels in treated mice.

4. Discussion and conclusions

In this study, we developed a novel ultrasound-visualized drug delivery system, termed $\text{Rb}_3\text{NPs@MBs}$, using saponin Rb_3 . Both *in vitro* and *in vivo* experiments demonstrated that the combination of $\text{Rb}_3\text{NPs@MBs}$ with UTMD effectively alleviated arterial plaques in AS.

Firstly, we successfully prepared microbubbles with a diameter of about 1.70 μm , and the $\text{Rb}_3\text{NPs@MBs}$ were uniformly dispersed with good properties. The results of cellular uptake indicate that the mechanism of UTMD promoting cellular uptake of drugs is feasible (Fig. 2). Then at the cellular level, a CCK-8 assay confirmed that $\text{Rb}_3\text{NPs@MBs}$ in conjunction with UTMD (1–100 $\mu\text{g ml}^{-1}$) did not compromise cell viability in both HUVECs and RAW264.7 cells. Following this screening, 10 $\mu\text{g ml}^{-1}$ was chosen as the optimal *in vitro* concentration based on confocal microscopy analyses (Fig. 3A–C). Subsequent experiments indicated that $\text{Rb}_3\text{NPs@MBs}$ reduced Dil-ox-LDL uptake in RAW264.7 cells, thereby inhibiting foam cell formation (Fig. 3D and E). In addition, flow cytometry revealed that $\text{Rb}_3\text{NPs@MBs}$ suppressed HUVEC apoptosis (Fig. 3F and G). To investigate whether these beneficial effects were mediated through modulation of oxidative stress, we evaluated the mitochondrial membrane potential (*via* JC-1 staining) in HUVECs, measured ROS levels in RAW264.7 cells, and quantified key antioxidant factors (SOD and GSH) (Fig. 4). The results showed that $\text{Rb}_3\text{NPs@MBs}$ preserved mitochondrial membrane potential in HUVECs, mitigated mitochondrial damage and ROS generation, and reduced ROS levels in RAW264.7 cells. Moreover, $\text{Rb}_3\text{NPs@MBs}$ therapy elevated both SOD and GSH in endothelial cells and macrophages. Thus, $\text{Rb}_3\text{NPs@MBs}$ inhibited early pathological events in AS, primarily by decreasing intracellular oxidative stress.

Because the aortic arch is predisposed to plaque formation (due to its angulation, increased branching, and abnormal shear forces),⁵⁷ we selected it as the primary target for UTMD-mediated drug delivery *in vivo*. *In vivo* imaging of small animals confirmed that UTMD overcame blood flow constraints, facilitated traversal of the endothelial barrier, and effectively accumulated Cy7.5 in the targeted aortic arch (Fig. 5A). ApoE^{−/−} mice were treated for two weeks. Notably, the MBs + UTMD and SV groups did not show significant differences in biochemical indices or plaque burden relative to untreated AS controls (model group). Although Rb_3NPs alone reduced total plaque area in the aorta, its effect on the aortic arch, where the plaque was most advanced, remained limited. In contrast, $\text{Rb}_3\text{NPs@MBs}$ + UTMD not only effectively reduced the overall plaque burden in the aorta but also signifi-



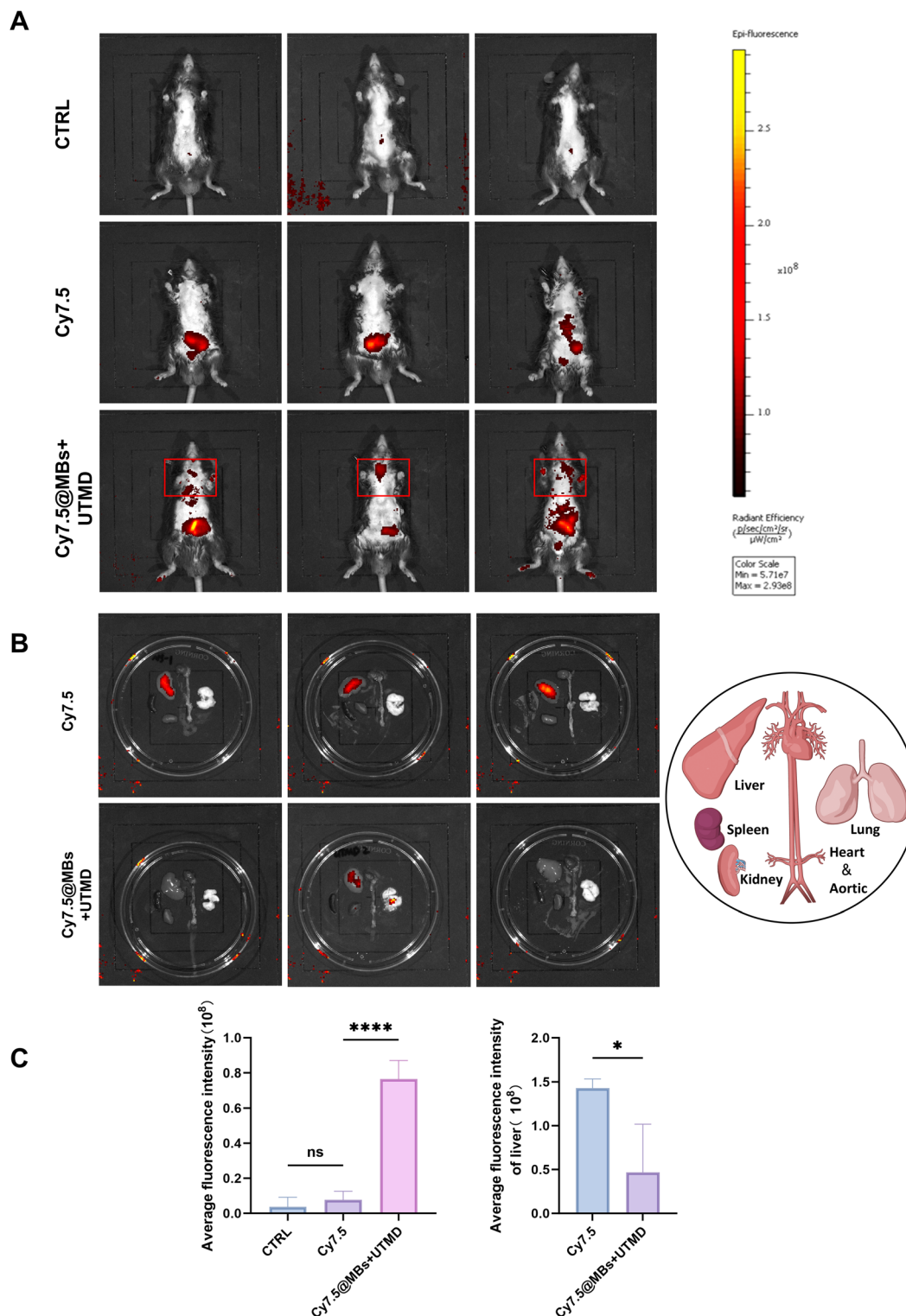


Fig. 5 Biodistribution of microbubbles combined with UTMD *in vivo* ($n = 3$). (A) *In vivo* fluorescence imaging of mice after UTMD (1 h). Color scale: min = 5.71×10^7 , max = 2.93×10^8 . (B) Fluorescence imaging of the mouse liver, spleen, lungs, kidneys, heart, and aorta after 24 h. Color scale: min = 1.12×10^8 , max = 1.68×10^8 . (C) Fluorescence quantitative analysis of (A) and (B) ($n = 3$).

cantly diminished plaques in the targeted aortic arch (Fig. 6B–D). The combination therapy simultaneously lowered inflammatory markers, oxidative stress, and lipid levels in AS mice

(Fig. 6E–G). Pathological assessments of aortic valve plaques demonstrated that $\text{Rb}_3\text{NPs@MBs} + \text{UTMD}$ reduced VCAM-1 expression, decreased lipid accumulation, increased plaque



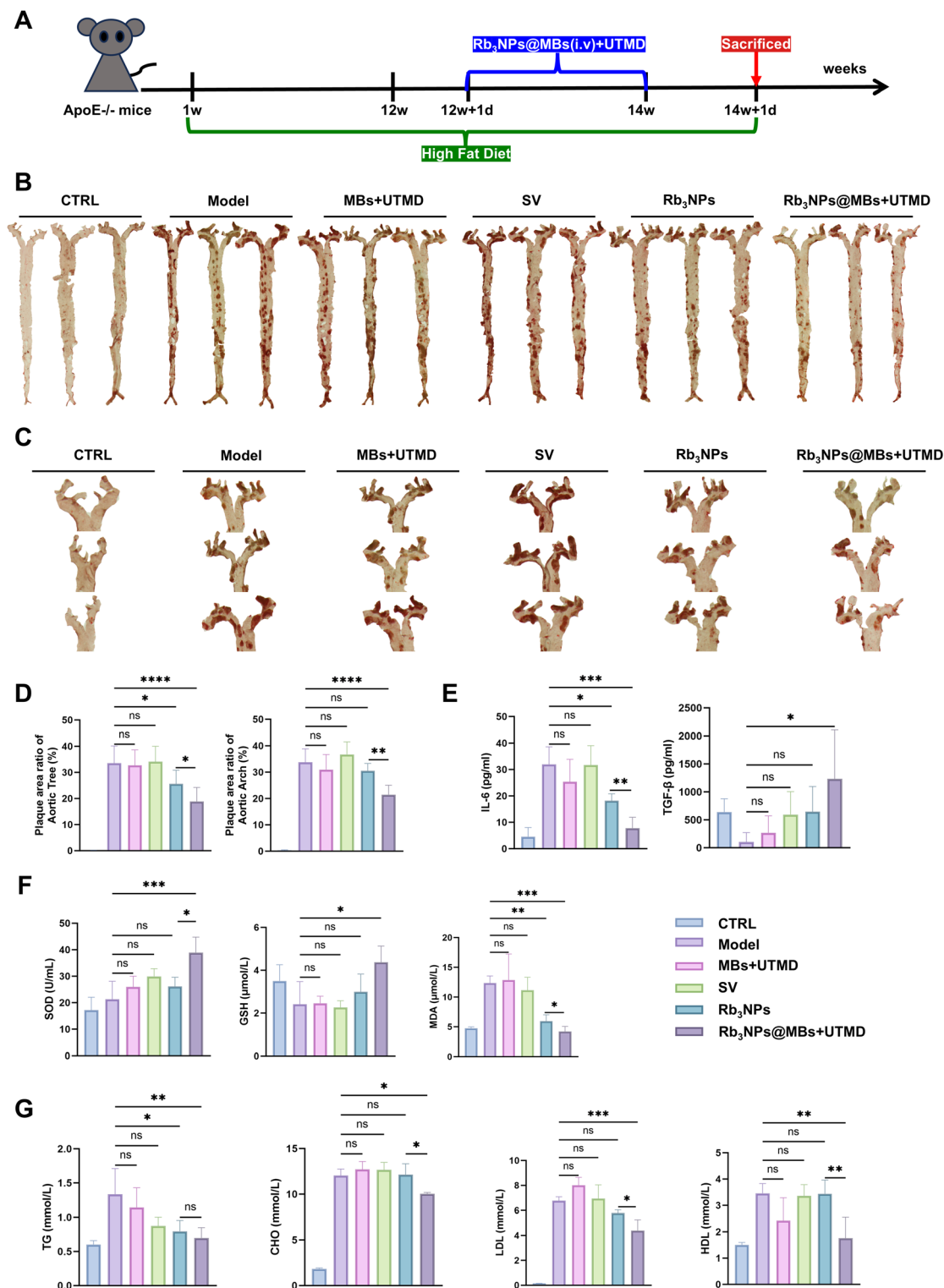
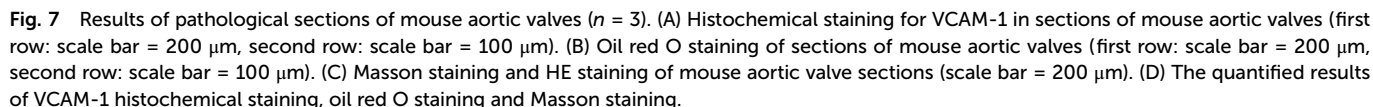


Fig. 6 Rb₃NPs@MBs in combination with UTMD targeted reduction of arterial plaque in AS mice. (A) Schematic diagram of the treatment cycle of mice ($n = 8$). (B) Aortic tree oil red O staining ($n = 8$). (C) Oil red O staining of aortic arch in mice. (D) Quantitative analysis of (B) & (C). (E) ELISA results of IL-6 and TGF- β in mouse serum ($n = 5$). (F) The biochemical detection results of oxidative stress factors SOD, GSH, and MDA in mouse plasma ($n = 5$). (G) Results of biochemical measurements of lipid levels of TG, CHO, LDL, and HDL in mouse serum ($n = 5$).

collagen content, and diminished plaque area, thereby corroborating its enhanced therapeutic efficacy (Fig. 7). Together, these results suggest that Rb₃NPs@MBs with UTMD can effec-

tively deliver the drug to the targeted aortic arch, reduce plaque formation, and improve pathological indices relevant to AS.





therapeutic effect is not obvious mainly because the active uptake of Rb₃NPs by cells also increases with time, accumulating to a level comparable to that of the Rb₃NPs@MBs + UTMD group at 24 h. Therefore, the final intracellular drug concentration was not significantly different between the two groups.

This phenomenon may be related to the high affinity of macrophages for fat-soluble drugs, and further research is needed in the future. However, *in vitro* models cannot fully replicate the complexity of *in vivo* pathophysiology, and our *in vivo* results confirmed that Rb₃NPs@MBs combined with UTMD outperformed Rb₃NPs alone.

In conclusion, we successfully constructed a targeted drug delivery system—Rb₃NPs@MBs with UTMD—that demonstrated feasibility, efficacy, and safety in both *in vitro* and *in vivo* models. These findings highlight its potential for clinical application as a promising therapeutic strategy for AS. Ongoing research will aim to further optimize this system and evaluate its effectiveness in diverse AS models and clinical settings.

Author contributions

Manuscript drafting, data curation, investigation, and visualization: Chunting Zhong, Xiaoting Yang, Jianhua Bai, Yiran Ji, Xiao Tan, Xiaoyu Chen and Lijun Xing; methodology, validation, and project administration: Bingxuan Xu and Dianhuan Tan; *in vitro* experiments: Chunting Zhong and Jianhua Bai; *in vivo* imaging experiments: Chunting Zhong; *in vivo* experiments: Chunting Zhong, Xiaoting Yang, Jianhua Bai, Yiran Ji, Xiao Tan and Xiaoyu Chen; study design and conceptualization: Tingting Zheng; supplementary experimental data: Chunting Zhong, Jianhua Bai and Xiaoting Yang; resources and funding acquisition: Yun Chen and Tingting Zheng; supervision and manuscript editing: Tingting Zheng. All authors have approved the manuscript and agreed to its submission to *Biomaterials Science*.

Data availability

The data supporting this article have been included as part of the ESI.†

Conflicts of interest

There are no conflicts to declare.

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References

- 1 V. Junaid, A. M. K. Minhas, M. Inam, C. Hinkamp, K. M. Talha, C. Meloche, S. Sheikh, A. Khoja, C. Krittanawong, E. M. Vaughan, D. K. Kalra, L. Slipczuk and S. S. Virani, *Curr. Atheroscler. Rep.*, 2024, **27**, 14.
- 2 P. Libby, J. E. Buring, L. Badimon, G. K. Hansson, J. Deanfield, M. S. Bittencourt, L. Tokgözoğlu and E. F. Lewis, *Nat. Rev. Dis. Primers*, 2019, **5**, 56.
- 3 L.-L. Bu, H.-H. Yuan, L.-L. Xie, M.-H. Guo, D.-F. Liao and X.-L. Zheng, *Int. J. Mol. Sci.*, 2023, **24**, 15160.
- 4 S. Jebari-Benslaiman, U. Galicia-García, A. Larrea-Sebal, J. R. Olaetxea, I. Alloza, K. Vandenbroeck, A. Benito-Vicente and C. Martín, *Int. J. Mol. Sci.*, 2022, **23**, 3346.
- 5 E. Niki, *Free Radicals Biol. Med.*, 2018, **120**, 425–440.
- 6 S. Zhong, L. Li, X. Shen, Q. Li, W. Xu, X. Wang, Y. Tao and H. Yin, *Free Radicals Biol. Med.*, 2019, **144**, 266–278.
- 7 Y. Li, M. Zhou, H. Li, C. Dai, L. Yin, C. Liu, Y. Li, E. Zhang, X. Dong, H. Ji and Q. Hu, *Eur. Heart J.*, 2024, **45**, 268–283.
- 8 L. Zhu, Y. Zhong, M. Yan, S. Ni, X. Zhao, S. Wu, G. Wang, K. Zhang, Q. Chi, X. Qin, C. Li, X. Huang and W. Wu, *ACS Appl. Mater. Interfaces*, 2024, **16**, 32027–32044.
- 9 P. Libby, *Nature*, 2021, **592**, 524–533.
- 10 P. Marchio, S. Guerra-Ojeda, J. M. Vila, M. Aldasoro, V. M. Victor and M. D. Mauricio, *Oxid. Med. Cell. Longevity*, 2019, **2019**, 1–32.
- 11 H. Ait-Oufella and P. Libby, *Arterioscler., Thromb., Vasc. Biol.*, 2024, **44**, 1899–1905.
- 12 X. Zhang, F. Centurion, A. Misra, S. Patel and Z. Gu, *Adv. Drug Delivery Rev.*, 2023, **194**, 114709.
- 13 L. H. Opie, *Trends Cardiovasc. Med.*, 2015, **25**, 216–225.
- 14 C. P. Cannon, *Eur. Heart J.*, 2019, **40**, 3526–3528.
- 15 A. Ajoolabady, D. Pratico, M. Mazidi, I. G. Davies, G. Y. H. Lip, N. Seidah, P. Libby, G. Kroemer and J. Ren, *Metabolism*, 2025, **163**, 156064.
- 16 P. M. Ridker, B. M. Everett, T. Thuren, J. G. MacFadyen, W. H. Chang, C. Ballantyne, F. Fonseca, J. Nicolau, W. Koenig, S. D. Anker, J. J. P. Kastelein, J. H. Cornel, P. Pais, D. Pella, J. Genest, R. Cifkova, A. Lorenzatti, T. Forster, Z. Kobalava, L. Vida-Simiti, M. Flather, H. Shimokawa, H. Ogawa, M. Dellborg, P. R. F. Rossi, R. P. T. Troquay, P. Libby and R. J. Glynn, *N. Engl. J. Med.*, 2017, **377**, 1119–1131.
- 17 M. A. Potenza, M. Montagnani, L. Santacroce, I. A. Charitos and L. Bottalico, *J. Ginseng Res.*, 2023, **47**, 359–365.
- 18 W. Fan, Y. Huang, H. Zheng, S. Li, Z. Li, L. Yuan, X. Cheng, C. He and J. Sun, *Biomed. Pharmacother.*, 2020, **132**, 110915.
- 19 J. Chen, X. Wei, Q. Zhang, Y. Wu, G. Xia, H. Xia, L. Wang, H. Shang and S. Lin, *Acta Pharm. Sin. B*, 2023, **13**, 1919–1955.
- 20 W. Jin, C. Li, S. Yang, S. Song, W. Hou, Y. Song and Q. Du, *Front. Pharmacol.*, 2023, **14**, 1166898.
- 21 S.-J. Oh, Y. Oh, I. W. Ryu, K. Kim and C.-J. Lim, *Biosci., Biotechnol., Biochem.*, 2016, **80**, 95–103.



- 22 X. Chen, Q. Wang, M. Shao, L. Ma, D. Guo, Y. Wu, P. Gao, X. Wang, W. Li, C. Li and Y. Wang, *Biomed. Pharmacother.*, 2019, **120**, 109487.
- 23 T. Dai, W. He, C. Yao, X. Ma, W. Ren, Y. Mai and A. Wu, *Biomater. Sci.*, 2020, **8**, 3784–3799.
- 24 Y. Zhang, H. Ji, O. Qiao, Z. Li, L. Pecoraro, X. Zhang, X. Han, W. Wang, X. Zhang, S. Man, J. Wang, X. Li, C. Liu, L. Huang and W. Gao, *Biomed. Pharmacother.*, 2021, **139**, 111630.
- 25 S. Lu, P. Zhao, Y. Deng and Y. Liu, *Pharmaceutics*, 2022, **14**, 480.
- 26 W. He, X. Xing, X. Wang, D. Wu, W. Wu, J. Guo and S. Mitragotri, *Adv. Funct. Mater.*, 2019, **11**, 1855–1863.
- 27 S. Huo, P. Zhao, Z. Shi, M. Zou, X. Yang, E. Warszawik, M. Loznik, R. Göstl and A. Herrmann, *Nat. Chem.*, 2021, **13**, 131–139.
- 28 Q. Deng, J. Mi, J. Dong, Y. Chen, L. Chen, J. He and J. Zhou, *ACS Nano*, 2022, **17**, 263–274.
- 29 W. Guo, S. Huang, J. An, J. Zhang, F. Dong, J. Dang and J. Zhang, *ACS Appl. Mater. Interfaces*, 2022, **14**, 50664–50676.
- 30 X. Wang, F. Li, J. Zhang, L. Guo, M. Shang, X. Sun, S. Xiao, D. Shi, D. Meng, Y. Zhao, C. Jiang and J. Li, *J. Controlled Release*, 2024, **367**, 45–60.
- 31 H. Yang, Y. Sun, J. Wei, L. Xu, Y. Tang, L. Yang, X. Zhang and Y. Lu, *Biomed. Pharmacother.*, 2019, **118**, 109161.
- 32 A. Bouakaz and J. Escoffre, *Adv. Drug Delivery Rev.*, 2024, **206**, 115199.
- 33 W. Wei, Y. Wang, Z. Wang and X. Duan, *TrAC, Trends Anal. Chem.*, 2023, **160**, 116958.
- 34 Y. Yang, Q. Li, X. Guo, J. Tu and D. Zhang, *Ultrason. Sonochem.*, 2020, **67**, 105096.
- 35 S. Keller, M. Bruce and M. A. Averkiou, *Ultrasound Med. Biol.*, 2019, **45**, 833–845.
- 36 S. M. Chowdhury, L. Abou-Elkacem, T. Lee, J. Dahl and A. M. Lutz, *J. Controlled Release*, 2020, **326**, 75–90.
- 37 I. Lentacker, I. De Cock, R. Deckers, S. C. De Smedt and C. T. W. Moonen, *Adv. Drug Delivery Rev.*, 2014, **72**, 49–64.
- 38 I. De Cock, E. Zagato, K. Braeckmans, Y. Luan, N. de Jong, S. C. De Smedt and I. Lentacker, *J. Controlled Release*, 2015, **197**, 20–28.
- 39 B. van Elburg, J. Deprez, M. van den Broek, S. C. De Smedt, M. Versluis, G. Lajoinie, I. Lentacker and T. Segers, *J. Controlled Release*, 2023, **363**, 747–755.
- 40 K.-H. Song, A. C. Fan, J. T. Bransky, T. Trudeau, A. Gutierrez-Hartmann, M. L. Calvisi and M. A. Borden, *Theranostics*, 2015, **5**, 1419–1427.
- 41 M. A. O'Reilly, *Science*, 2024, **385**, eadp7206.
- 42 Z. Zhang, B. Xu, T. Lv, Y. Shi, M. Wang, D. Hu, A. Hu, P. Li, S. Lin, S. Zhang, R. Yao, L. Luo, L. Wang, Y. Zhang, Y. Han, H. Hu, X. Shuai, J. Shi, Y. Chen and T. Zheng, *Adv. Ther.*, 2023, **6**, 2300056.
- 43 L. Duan, L. Yang, J. Jin, F. Yang, D. Liu, K. Hu, Q. Wang, Y. Yue and N. Gu, *Theranostics*, 2020, **10**, 462–483.
- 44 S. Liu, Y. Zhang, Y. Liu, W. Wang, S. Gao, W. Yuan, Z. Sun, L. Liu and C. Wang, *Br. J. Cancer*, 2022, **128**, 715–725.
- 45 X. Li, S. Guo, T. Xu, X. He, Y. Sun, X. Chen, S. Cao, X. Si, W. Liao, Y. Liao, Y. Han and J. Bin, *Theranostics*, 2020, **10**, 2522–2537.
- 46 B. G. Brown and X.-Q. Zhao, *J. Am. Coll. Cardiol.*, 2007, **49**, 933–938.
- 47 H. Yuan, H. Hu, J. Sun, M. Shi, H. Yu, C. Li, Y. U. Sun, Z. Yang and R. M. Hoffman, *In Vivo*, 2018, **32**, 1025–1032.
- 48 F. Liu, Y. Mao, J. Yan, Y. Sun, Z. Xie, F. Li, F. Yan, H. Zhang and P. Zhang, *Research*, 2022, **2022**, 9830627.
- 49 J. Pazzi and A. B. Subramaniam, *J. Colloid Interface Sci.*, 2024, **661**, 1033–1045.
- 50 L. Fournier, T. de La Taille and C. Chauvierre, *Biomaterials*, 2023, **294**, 122025.
- 51 Y. Wen, Y. Ji, S. Zhang, B. Xu, S. Sun, Y. Chen, X. Shuai and T. Zheng, *Aging*, 2024, **16**, 10784–10798.
- 52 Z. He, W. Chen, K. Hu, Y. Luo, W. Zeng, X. He, T. Li, J. Ouyang, Y. Li, L. Xie, Y. Zhang, Q. Xu, S. Yang, M. Guo, W. Zou, Y. Li, L. Huang, L. Chen, X. Zhang, Q. Saiding, R. Wang, M.-R. Zhang, N. Kong, T. Xie, X. Song and W. Tao, *Nat. Nanotechnol.*, 2024, **19**, 1386–1398.
- 53 Z.-C. Wang, K.-M. Niu, Y.-J. Wu, K.-R. Du, L.-W. Qi, Y.-B. Zhou and H.-J. Sun, *Cell Death Dis.*, 2022, **13**, 824.
- 54 R. Cannella, G. Pilato, M. Mazzola and T. V. Bartolotta, *Radiol. Med.*, 2023, **128**, 1023–1034.
- 55 C. Lane-Donovan, W. M. Wong, M. S. Durakoglugil, C. R. Wasser, S. Jiang, X. Xian and J. Herz, *J. Neurosci.*, 2016, **36**, 10141–10150.
- 56 E. Asimakidou, E. N. Saipuljumri, C. H. Lo and J. Zeng, *Neural Regener. Res.*, 2024, **20**, 1069–1076.
- 57 P. Libby and G. K. Hansson, *Circ. Res.*, 2015, **116**, 307–311.

