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Synthetic pathways to create asymmetric center at C1 position of 1-substituted-tetrahydro- β -carbolines – a review

Md. Moaz Ahmed Asif, ^{†a} Susmita Roy Lisa ^{†b} and Nazmul Qais ^{*c}

The 2,3,4,9-tetrahydro-1*H*-pyrido[3,4-*b*]indoles or tetrahydro- β -carbolines (TH β Cs) are tricyclic compounds that are found in various natural sources that exhibit a wide range of important pharmacological activities. Chiral 1-substituted-TH β Cs, which have an asymmetric center at C1, have attained significant interest due to their possible Monoamine Oxidase (MAO) inhibitory activity, benzodiazepine receptor binding activity, and antimalarial effectiveness against chloroquine-resistant *Plasmodium falciparum*. This review highlights and summarizes various novel stereoselective approaches to introduce chirality at the C1 position of 1-substituted-TH β Cs in good yield and enantiomeric excess (ee) or diastereomeric excess (de). These methods include the Pictet–Spengler reaction, chiral auxiliary, Asymmetric Transfer Hydrogenation (ATH) with chiral catalysts, asymmetric addition reaction, and enzymatic catalysis. The syntheses of chiral TH β Cs are reviewed comprehensively, emphasizing their role in drug development from 1977 to 2024.

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1. Introduction

The tetrahydro- β -carbolines (TH β Cs) are a group of compounds found in a variety of natural and synthetic compounds containing a unique tricyclic pyrido[3,4-*b*]indole ring and renowned for their promising biological actions. Originating from tryptamine or tryptophan, these compounds are widespread in nature and have been isolated from various sources including plants, fungi, animals, and marine organisms.¹ TH β Cs exhibit a broad spectrum of pharmacological activities; including phosphodiesterase 5 (PDE5)-inhibitory,² antitumor,^{3,4} antiviral,^{5,6} and antiprotozoal⁷ especially antimalarial effects.^{8,9} Chiral 1-substituted-TH β Cs **1** (Fig. 1), having an asymmetric center present at the C1 position, are still being sought even after being discovered more than a hundred years ago.¹⁰ They are mainly MAO inhibitors or work by binding to benzodiazepine receptors.^{11,12} They have gained particular interest due to their potential antimalarial efficacy against a *Plasmodium falciparum* strain (FcB1-Colombia) that is chloroquine-resistant.¹³

Some of the specific 1-substituted-TH β Cs (Fig. 1) that have biological importance are given below:

- Justine 2 (HR22C16) induces mitotic arrest and blocking cell division in taxol-resistant cancer cells.^{4,14}
- The African rhacophorid frog *Kassina senegalensis*¹⁵ is the source of tryptargine **3a**, a highly poisonous TH β C alkaloid. It was recently discovered in a hitherto unknown ground ascidian *Eudistoma* sp.¹⁶ A very similar chemical, 6-hydroxy-tryptargine, was shown to be a strong neurotoxic in the venom of the Brazilian web spider *Parawixia bistriata*.¹⁷
- Tadalafil **4** is an orally active PDE5 inhibitor and also highly potent.^{2,18}
- Vincamine **5** aided in mild to moderate dementia patients.¹⁹
- Yohimbine **6**, an α_2 -adrenoceptor blocker that helps in erectile dysfunction.^{20,21}
- Jaddifine **7** collected from *Vinca difformis*.²²
- Ajmalicine **8** (ref. 23) and reserpine **10** (ref. 24) (Scheme 1) used as an antihypertensive.
- Neonaucleoside C **9** collected from *Neonauclea sessilifolia*.²⁵
- Fumitremorgins are found in fungi that have antiviral²⁶ and cell-cycle inhibitory activities.²⁷ They also worked as protein kinase and topoisomerase II inhibitors.²⁸

Synthetic methodologies to introduce chirality at the C1 position in TH β Cs have been extensively studied.²⁹ These methods include the Pictet–Spengler reaction,³⁰ asymmetric alkylation using N2-auxiliary as a directing group,³¹ and acid-induced epimerization in conjunction with the Pictet–Spengler reaction.³² Additionally, the Bischler–Napieralski reaction³³ and classical Noyori ATH conditions³⁴ have been highlighted as key synthetic routes to create chiral 1-substituted-TH β Cs. The C1 stereocenter in TH β Cs plays a crucial role in their

^aDepartment of Pharmacy, Faculty of Science & Engineering, University of Information Technology & Sciences, Holding 190, Road 5, Block J, Baridhara, Maddha Nayanagar, Vajara, Dhaka-1212, Bangladesh

^bDepartment of Chemistry, Syracuse University, Syracuse, NY 13244, USA

^cDepartment of Clinical Pharmacy and Pharmacology, Faculty of Pharmacy, University of Dhaka, Dhaka-1000, Bangladesh. E-mail: nqais@du.ac.bd

[†] Co-first authors.

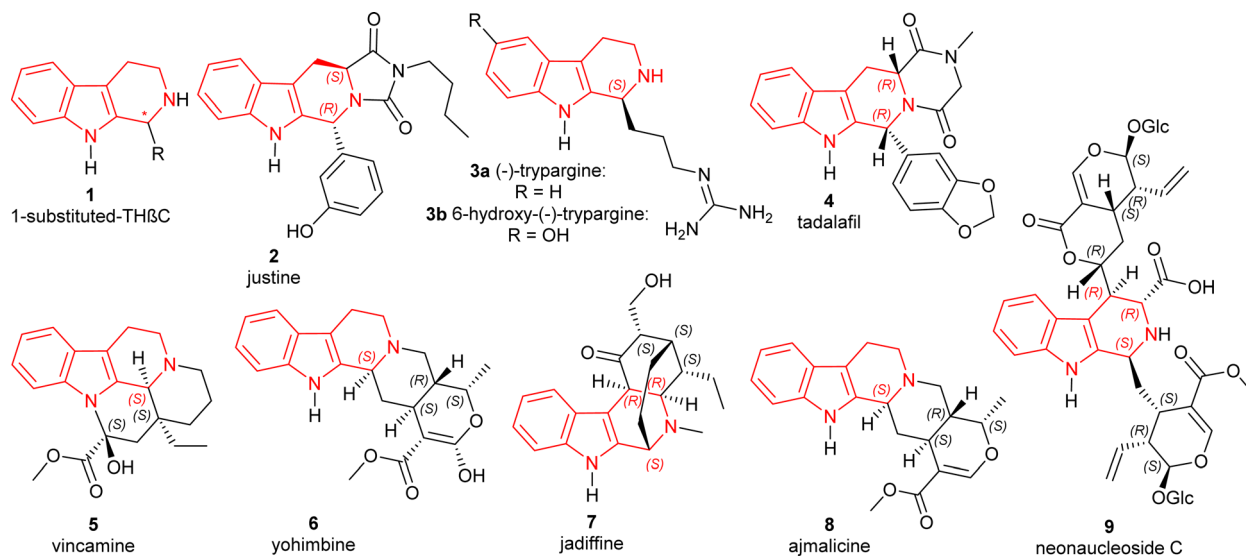



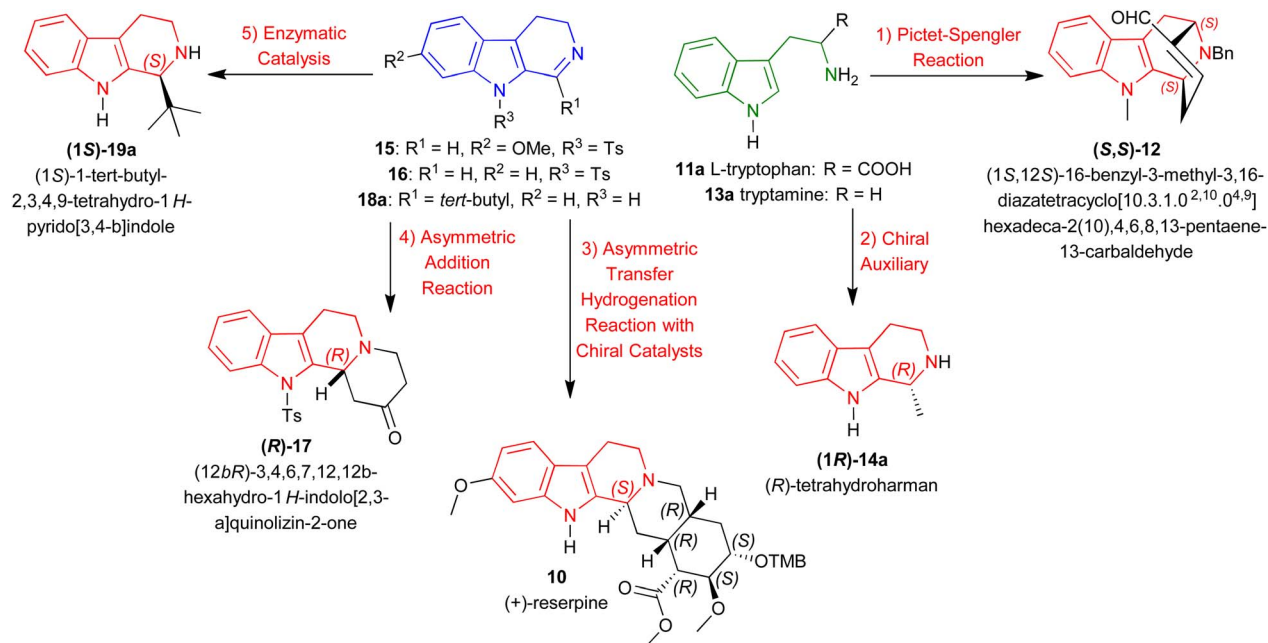
Fig. 1 Structures of important chiral 1-substituted-THβCs.

pharmacological properties, influencing their activity in various therapeutic areas. With a ubiquitous presence in both natural sources and synthetic derivatives, these compounds have significant attention in medicinal chemistry for their potential therapeutic applications. The intricate interplay of their chemical structure and biological effects underscores their pivotal role in drug discovery endeavors, accentuating the paramount importance of advancing synthetic methodologies to access these compounds efficiently.

In the last 11 years, Laine *et al.*, Maity *et al.*, Szabó *et al.*, Wang *et al.*, and Du *et al.* published reviews that emphasized on the pharmacological importance, overall synthetic methods, biological activities, and applications of THβCs,^{35–39} but our

review does not comprise any of the above-mentioned perspective wholeheartedly. This review neither talks about pharmacological importance, nor biological activities; neither gives all of the synthetic methodologies, nor the applications of THβCs also.

Instead, this review intends to offer a complete overview of the asymmetric synthesis of 1-substituted-THβCs, focusing on the synthetic methods to introduce chirality at the C1 position and their implications for drug development. Here, we discussed about five methods to create an asymmetric center at the C1 position of 1-substituted-THβCs reported from as early as 1977 to as latest as 2024. With representative examples (Scheme 1), they are:



Scheme 1 Representative examples of asymmetric methods for synthesizing chiral 1-substituted-tetrahydro-β-carbolines.



(1) *Pictet–Spengler reaction*: From L-tryptophan **11a** to synthesize (1*S*,12*S*)-16-benzyl-3-methyl-3,16-diazatetracyclo[10.3.1.0^{2,10}.0^{4,9}]hexadeca-2(10),4,6,8,13-pentaene-13-carbaldehyde (**S,S**)-**12**.⁴⁰

(2) *Chiral auxiliary*: From tryptamine **13a** to synthesize (*R*)-tetrahydroharman (**1R**)-**14a**.⁴¹

(3) *ATH with chiral catalysts*: From 7-methoxy-9-(4-methylphenyl)sulfonyl-3,4-dihydropyrido[3,4-*b*]indole (7-methoxy-9-tosyl-DHβC) **15** to synthesize (+)-reserpine **10**.⁴²

(4) *Asymmetric addition reaction*: From 9-tosyl-DHβC **16** to synthesize (12*bR*)-3,4,6,7,12,12*b*-hexahydro-1*H*-indolo[2,3-*a*]quinolizin-2-one (**R**)-**17**.⁴³

(5) *Enzymatic catalysis*: From 1-*tert*-butyl-4,9-dihydro-3*H*-pyrido[3,4-*b*]indole **18a** to synthesize (1*S*)-1-*tert*-butyl-THβC (**1S**)-**19a**.⁴⁴

2. Enantioselective synthesis of 1-substituted-tetrahydro-β-carbolines

The enantioselective synthesis of 1-substituted-THβCs **1** can be conducted by the following five methods:

Method 1. Pictet–Spengler reaction.

Method 2. Chiral auxiliary.

Method 3. Asymmetric transfer hydrogenation reaction with chiral catalysts.

Method 4. Asymmetric addition reaction.

Method 5. Enzymatic catalysis.

Method 1. Pictet–Spengler reaction

More than 113 years ago from now in 1911, Amé Pictet and Theodor Spengler devised a novel way to produce 1,2,3,4-tetrahydroisoquinoline by heating β-phenylethylamine and formaldehyde dimethylacetal in the presence of hydrochloric acid.⁴⁰ This reaction is known as the Pictet–Spengler reaction. In 1928, Tatsui used the basis of this reaction to be the first to produce 1-methyl-THβC from tryptamine and ethanal in the presence of sulphuric acid.⁴⁵

Example 1. Asymmetric formal syntheses of (–)-koumine, (–)-taberpsychine, and (–)-koumidine intermediates from L-tryptophan. Bailey and McLay asymmetrically synthesized intermediates of naturally occurring (+)-koumine,⁴⁶ (+)-taberpsychine⁴⁷ & (+)-koumidine.^{46,48}

First, L-tryptophan methyl ester **11b** was condensed with methyl 4-oxobutanoate at 0 °C with excess 2,2,2-trifluoroacetic acid (TFA) in dichloromethane (DCM) to get (1*S*,3*S*)-**20** (predominating than its (1*R*,3*S*)-diastereomer by 4:1 diastereomeric ratio or dr) by Pictet–Spengler reaction under kinetic control⁴⁹ with a total yield of 61%.

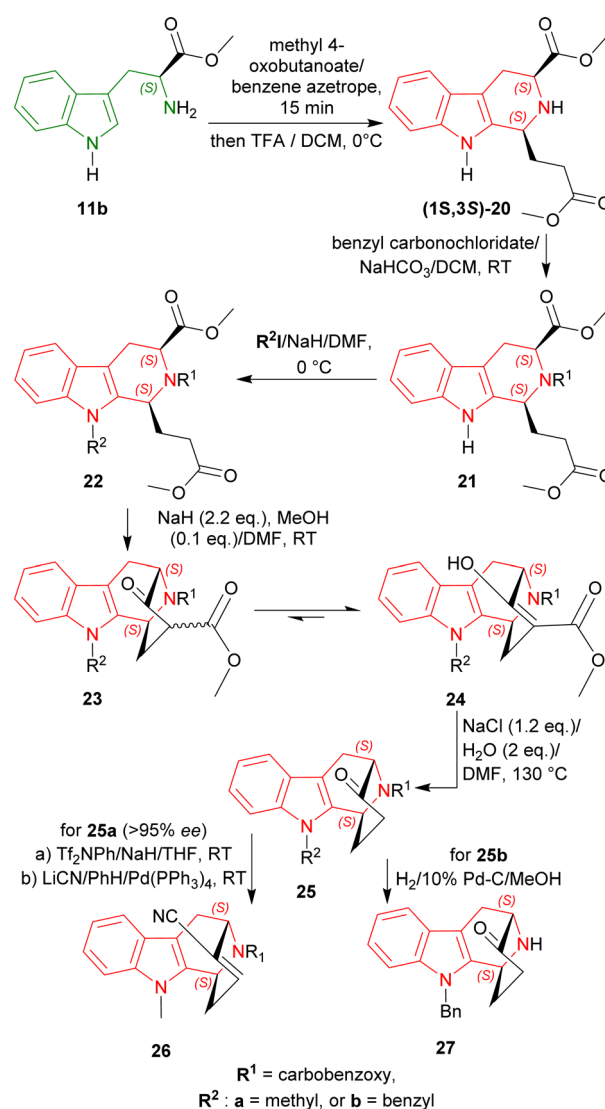
The N2 of (1*S*,3*S*)-**20** was then protected by benzyl carbonochloridate producing **21**, and N9 of **21** was protected by methyl iodide (CH₃I)/sodium hydride (NaH) or benzyl iodide/NaH respectively at 0 °C giving **22a** or **22b**.

With NaH and protic methanol (MeOH), Dieckmann cyclization of **22a** and **22b** gave the β-keto ester **23a** and **23b** and their enolic form **24a** and **24b**. These esters were hydrolyzed and decarboxylated by heating at 130 °C with NaCl and H₂O in *N,N*-

dimethylformamide (DMF)⁵⁰ producing the bridged ketone **25a** (>95% ee) and **25b**.

25a was then reacted with Tf₂NPh/NaH/THF, and LiCN/benzene (PhH)/Pd(PPh₃)₄ (ref. 51) to get benzyl (1*S*,12*S*)-13-hydroxy-3-methyl-3,16-diazatetracyclo[10.3.1.0^{2,10}.0^{4,9}]hexadeca-2(10),4,6,8,13-pentaene-16-carboxylate **26** which possesses an α,β-unsaturated nitrile for Michael addition of a C₄ fragment, giving access to the full carbon skeleton of N9-methylated alkaloids of the ajmaline–sarpagine group. This overall route is more efficient than that of the N2-benzyl derivative of **26**.⁵²

On the other hand, catalytic hydrogenation of **25b** with 10% Pd–C in MeOH produced (1*S*,12*S*)-3-benzyl-13-oxo-3,16-diazatetracyclo[10.3.1.0^{2,10}.0^{4,9}]hexadeca-2(10),4,6,8-tetraene **27** which is the antipode of the intermediate used in the syntheses of (+)-koumine, (+)-taberpsychine, and (+)-koumidine (Scheme 2).⁵³



Scheme 2 Asymmetric formal syntheses of (–)-koumine, (–)-taberpsychine, and (–)-koumidine intermediates from L-tryptophan methyl ester.



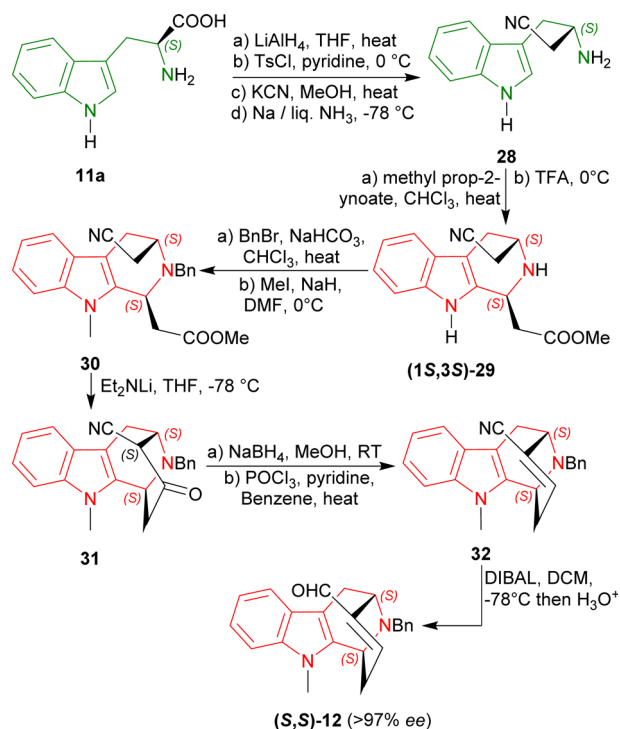
So, methyl 4-oxobutanoate predominantly produced (1*S*,3*S*)-diastereomer with L-tryptophan methyl ester by Pictet–Spengler reaction under kinetic control.

Example 2. Modified Pictet–Spengler reaction for formal syntheses of (–)-suaveoline, (–)-raumacline, and (–)-*N*^b-methylraumacline intermediates. Bailey *et al.* devised reaction pathways to produce intermediates of the ajmaline–sarpagine family alkaloids such as (–)-suaveoline, (–)-raumacline, and (–)-*N*^b-methylraumacline.⁴⁰

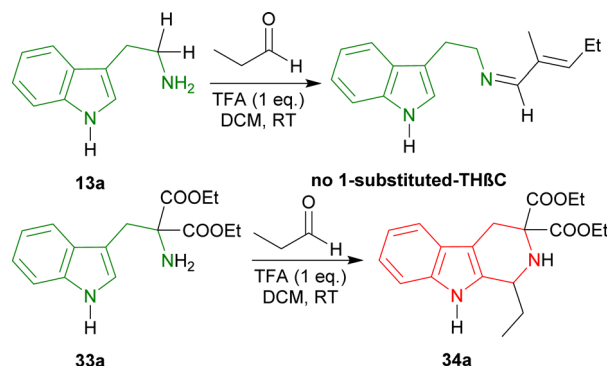
L-Tryptophan **11a** was converted to its homologated nitrile **28** in four steps in 50% overall yield.⁵⁴ Modified Pictet–Spengler reaction of **28** with methyl prop-2-ynoate followed by treatment with TFA gave rise to a 60% yield of the acetate (1*S*,3*S*)-**29** (54% de). In this reaction, Bailey *et al.* used a carbonyl-conjugated alkyne instead of the conventional aldehyde.^{52,55,56}

N2 benzylation and N9 methylation of (1*S*,3*S*)-**29** furnished the compound **30** in an overall 46% total yield. With lithium diethylamide at –78 °C, Dieckmann/Thorpe cyclisation⁵⁷ of **30** gave **31** in 90% yield. The reduction of **31** with sodium borohydride in MeOH at room temperature (RT) afforded the corresponding hydroxynitrile⁵⁸ and dehydration with POCl₃ produced **32** in 87% yield. Finally, reduction with bis(2-methylpropyl)alumane (DIBAL) gave a 99% yield of (*S,S*)-**12** (>97% ee) which was used in the synthesis of (–)-suaveoline, (–)-raumacline and (–)-*N*^b-methylraumacline (Scheme 3).⁵⁹

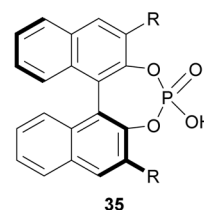
So, methyl prop-2-ynoate predominantly produced (1*S*,3*S*)-diastereomer with (3*S*)-3-amino-4-(1*H*-indol-3-yl)butanenitrile by modified Pictet–Spengler reaction.



Scheme 3 Asymmetric formal syntheses of (–)-suaveoline, (–)-raumacline, and (–)-*N*^b-methylraumacline intermediates from L-tryptophan.



Scheme 4 Tryptamine failed to produce any 1-substituted-THβC by Pictet–Spengler reaction but diethyl 2-amino-2-(1*H*-indol-3-ylmethyl)propanedioate was able to produce diethyl 1-ethyl-1,2,4,9-tetrahydropyrido[3,4-*b*]indole-3,3-dicarboxylate.



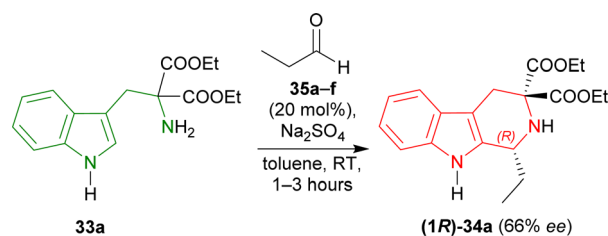
R: **a** = H, **b** = 4-nitrophenyl, **c** = 3,5-bis(trifluoromethyl)phenyl, **d** = 2,4,6-trimethylphenyl, **e** = naphthalen-2-yl, **f** = 2,4,6-tri(propan-2-yl)phenyl

Fig. 2 Chiral organic Brønsted acid for catalytic asymmetric Pictet–Spengler reaction.

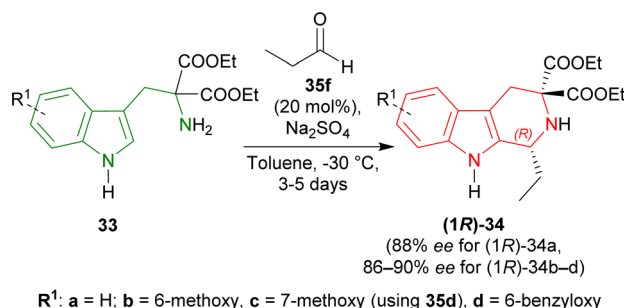
Example 3. Catalytic asymmetric Pictet–Spengler reaction with chiral organic Brønsted acid. Seayad *et al.* experimented on the acid catalysis of the Pictet–Spengler reaction. Still, they failed to cyclize tryptamine **13a** to produce any 1-substituted-THβC in the presence of propanal and TFA in DCM at RT, but diethyl 2-amino-2-(1*H*-indol-3-ylmethyl)propanedioate **33a** gave >90% yield of diethyl 1-ethyl-THβC-3,3-dicarboxylate **34a** (Scheme 4).⁶⁰

To find an appropriate chiral organic Brønsted acid, 20 mol% **35a–f** (Fig. 2) was then examined with Na₂SO₄ in toluene at RT for 1–3 hours. Among them, **35f** gave the highest 66% ee with a good yield of 90% of (1*R*)-**34a** (Scheme 5).

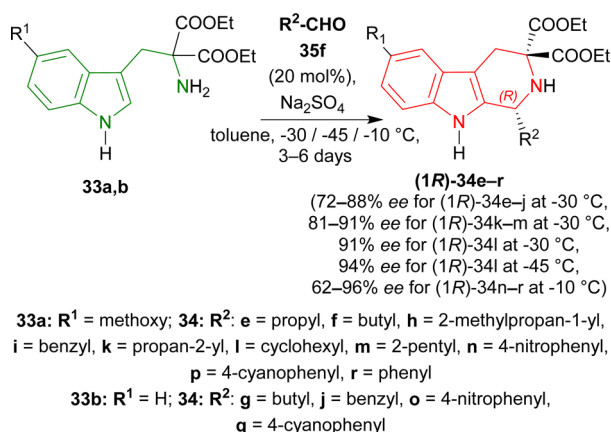
When the previous reaction was conducted with **35f** at –30 °C for 3–5 days, the yield of (1*R*)-**34a** decreased to 76% while ee increased to 88%. **33b–d** gave a similar ee of 86–90% with an excellent yield of 94–98% for (1*R*)-**34b–d** (Scheme 6).



Scheme 5 Screening of chiral organic Brønsted acid for catalytic asymmetric Pictet–Spengler reaction.



Scheme 6 Screening of diethyl 2-amino-2-(1H-indol-3-ylmethyl)propanedioate derivatives.



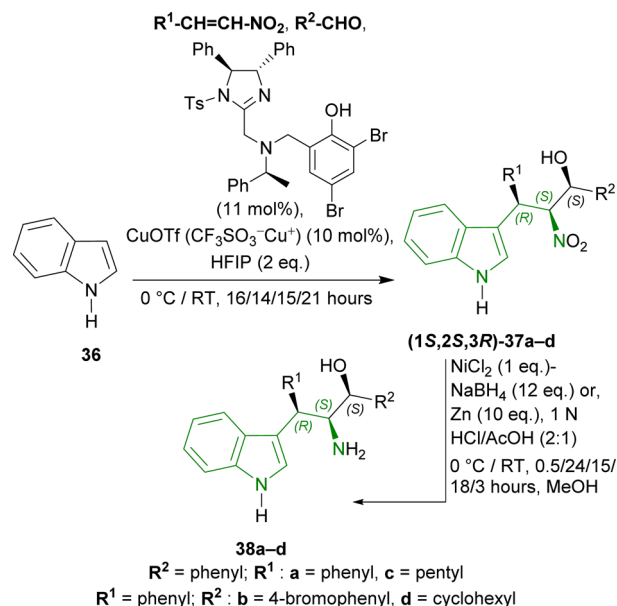
Scheme 7 Screening of aldehydes.

Various aldehydes were then reacted with **33a,b**. Aliphatic unbranched and branched aldehydes produced **(1R)-34e–j** (58–98% yield, 72–88% ee) and **(1R)-34k–m** (50–93% yield, 81–91% ee) respectively at -30°C in toluene for 3–6 days. When the temperature was decreased from -30°C to -45°C , ee of **(1R)-34l** increased slightly from 91% to 94% but yield decreased from 93% to 64%. Aromatic and electron-poor aromatic aldehydes also gave moderate to good yield (40–98%) and ee (62–96%) for **(1R)-34n–r** at -10°C in DCM (Scheme 7).

So, propionaldehyde and other aldehydes predominantly produced (1R)-enantiomer with diethyl 2-amino-2-(1H-indol-3-ylmethyl)propanedioate and its derivatives catalyzed by chiral organic Brønsted acid.

Example 4. Synthesis of (1S,3S,4R)-TH β Cs from 1H-indole through Friedel–Crafts/Henry adducts. Arai *et al.* developed a four-step synthetic pathway to produce chiral 1-substituted-TH β Cs from 1H-indole.⁶¹

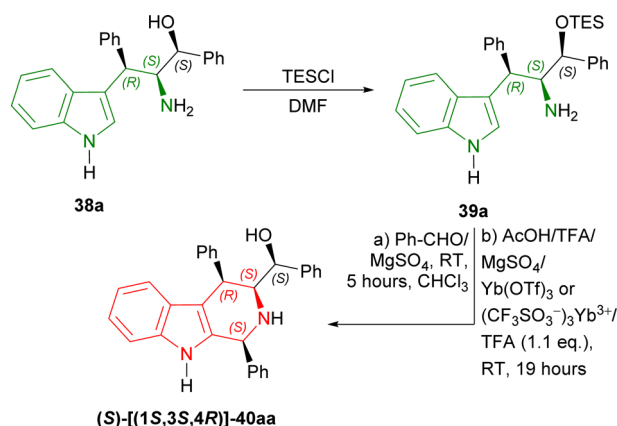
1H-indole **36** was reacted with different nitroalkenes and aldehydes in the presence of 2,4-dibromo-6-[[[(4S,5S)-1-(4-methylphenyl)sulfonyl-4,5-diphenyl-4,5-dihydroimidazol-2-yl]methyl-[(1S)-1-phenylethyl]amino]methyl]phenol (11 mol%) (Scheme 8), copper(i) trifluoromethanesulfonate (CuOTf or $\text{CF}_3\text{SO}_3^-\text{Cu}^+$, 10 mol%), fluoro(iodo)phosphane (HFIP, 2 equivalents or eq.)⁶² in toluene to produce **(1S,2S,3R)-37a–d** in 76–84% yield and 97–99% ee at 0°C or RT which are (R,S,S)-



Scheme 8 Synthesizing Friedel–Crafts/Henry adducts from 1H-indole and their reduction.

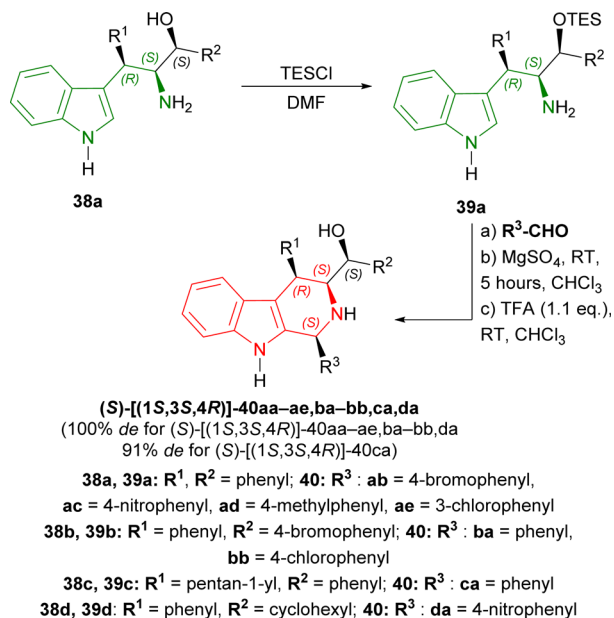
Friedel–Crafts/Henry adducts. Next **(1S,2S,3R)-37a**, reduced with nickel boride,^{63,64} gave **(1S,2S,3R)-3-(1H-indol-3-yl)-2-amino-1,3-diphenylpropan-1-ol 38a** (20% yield) at 0°C for 0.5 hour. But, Zn powder under acidic condition⁶⁵ at 0°C for 24 hours gave a 59% yield of **38a**. At RT for 15–18 hours, this condition gave a 99% yield of **38a–b** from **(1S,2S,3R)-37a–b**. Ultimately, Zn-nanopowder was used to reduce the reaction time to 3 hours at RT to give 98–99% yields of **38c–d** from **(1S,2S,3R)-37c–d** (Scheme 8).

Then, **38a** was experimented with to optimize Pictet–Spengler reaction. **38a** was converted into **39a** with triethylsilyl chloride (TESCl) in DMF. **39a** was cyclized with benzaldehyde and ethanoic acid, TFA, MgSO_4 , ytterbium(iii) trifluoromethanesulfonate ($\text{Yb}(\text{OTf})_3$ or $(\text{CF}_3\text{SO}_3^-)_3\text{Yb}^{3+}$); but TFA gave 30% yield of (S)-[(1S,3S,4R)-1,4-diphenyl-2,3,4,9-tetrahydro-1H-pyrido[3,4-b]indol-3-yl]-phenylmethanol **40aa**. After that, **39a**



Scheme 9 Protecting the OH group with TES group, Pictet–Spengler reaction, and removing the OH-protection.





Scheme 10 Screening of aldehydes.

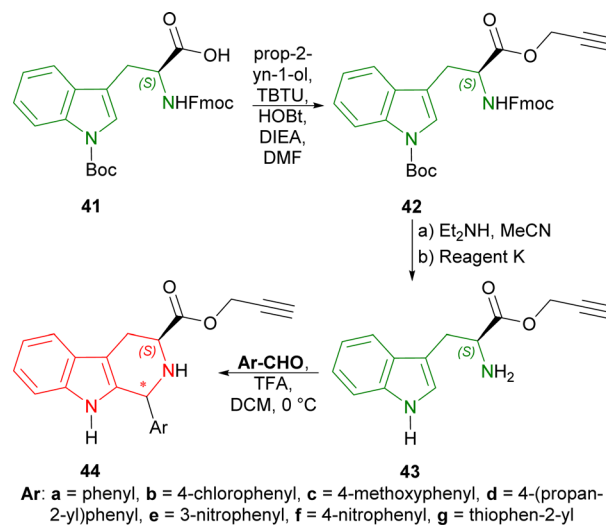
with TFA (1.1 eq.) at RT for 19 hours following without and with $MgSO_4$ at RT for 5 hours in $CHCl_3$ gave 47% and 67% yield of **40aa** respectively (Scheme 9).

38a–d was tested with different aromatic aldehydes for 19–25 hours to give (S)-[(1S,3S,4R)]-40ab–ae,ba–bb,ca,da in 38–72% yields. Every product had 100% de except (S)-[(1S,3S,4R)]-40ca of which de was 91% (Scheme 10).

So, phenylaldehyde and other aldehydes produced (1S,3S,4R)-TH β Cs predominantly with (1S,2S,3R)-1-(1H-indol-3-yl)-1,3-diphenyl-3-triethylsilyloxypropan-2-amine and its derivatives (made from Friedel–Crafts/Henry adducts) by Pictet–Spengler reaction.

Example 5. Two methods for TFA-catalyzed Pictet–Spengler reaction. Vavsari *et al.* developed two methods for TFA-catalyzed Pictet–Spengler reaction.⁶⁶

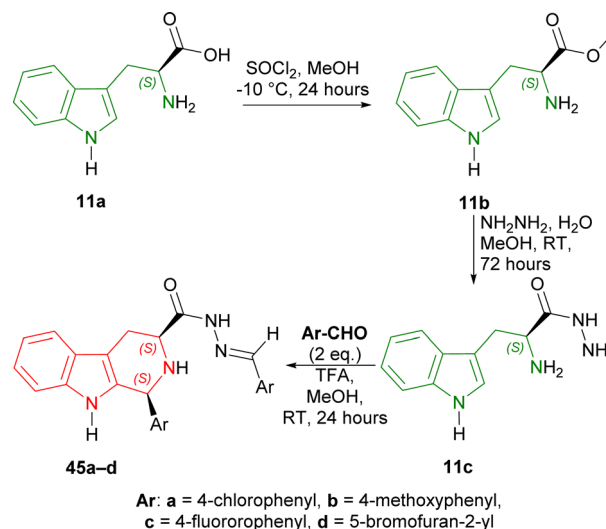
Firstly, (2S)-2-(9H-fluoren-9-ylmethoxycarbonylamino)-3-[1-[(2-methylpropan-2-yl)oxycarbonyl]indol-3-yl]propanoic acid **41** (10 mmol) was treated with prop-2-yn-1-ol (2 eq.), [benzotriazol-1-yloxy(dimethylamino)methylidene]-dimethylazanium;tetrafluoroborate (TBTU, 1.1 eq.), 1-hydroxybenzotriazole (HOBt·H₂O, 1.1 eq.), N-ethyl-N-propan-2-ylpropan-2-amine (DIEA, 2.2 eq.) and in DMF to get 87% yield of **42**. The Fmoc protection was then removed by diethylamine and acetonitrile and the Boc protection group was eliminated by cooled reagent K (TFA, water, phenol, ethanedithiol, triethylsilane, thioanisole) gaining a 66% yield of **43**. The compound **43** was then reacted with various aromatic aldehydes and TFA in DCM at 0 °C. Benzaldehyde gave higher yield (73% for prop-2-ynyl (1S,3S)-1-phenyl-TH β C-3-carboxylate (**1S,3S**)-44a) than 3- and 4-substituted-benzaldehydes (52–67% yields for prop-2-ynyl (1S,3S)-1-substituted-TH β C-3-carboxylate (**1S,3S**)-44b–f) and thiophene-2-carbaldehyde (57% yield for prop-2-ynyl (1R,3S)-1-thiophen-2-yl-TH β C-3-carboxylate (**1R,3S**)-44g) (Scheme 11).



Scheme 11 Synthesizing prop-2-ynyl ester of (2S)-2-(9H-fluoren-9-ylmethoxycarbonylamino)-3-[1-[(2-methylpropan-2-yl)oxycarbonyl]indol-3-yl]propanoic acid, removing Fmoc protection group, followed by Pictet–Spengler reaction.

Secondly, L-tryptophan **11a** is converted to **11b** (95% yield) with thionyl chloride in MeOH at –10 °C for 24 hours. With **11b** and hydrazine in MeOH at RT for 72 hours, **11c** was obtained in 95% yield. **11c** was then reacted with aromatic aldehydes in the presence of TFA as a catalyst in MeOH at RT for 24 hours. 4-Substituted-benzaldehydes gave similar yields (78–83% for (1S,3S)-1-(4-substituted-phenyl)-N-[(E)-(4-substituted-phenyl)methylideneamino]-TH β C-3-carboxamide **45a–c**); while 5-bromofuran-2-aldehyde had slightly better yield (85% for (1S,3S)-1-(5-bromofuran-2-yl)-N-[(E)-(5-bromofuran-2-yl)methylideneamino]-TH β C-3-carboxamide **45d**) (Scheme 12).

So, prop-2-ynyl (2S)-2-amino-3-(1H-indol-3-yl)propanoate and L-tryptophan hydrazide produced mainly (1S,3S)-TH β Cs



Scheme 12 Synthesizing methyl ester of L-tryptophan, then L-tryptophan hydrazide, and Pictet–Spengler reaction.

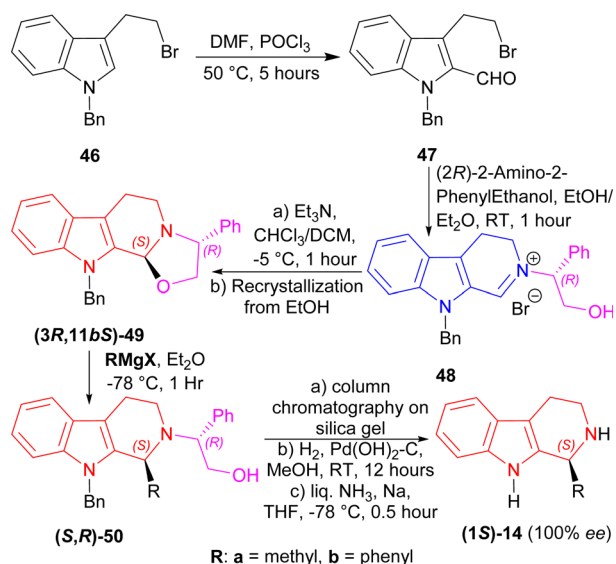
with aromatic aldehydes by TFA-catalyzed Pictet–Spengler reaction; the only exception being prop-2-ynyl (1*R*,3*S*)-1-thio-phen-2-yl-THβC-3-carboxylate.

Method 2. Chiral auxiliary

A chiral auxiliary is actually a pure enantiomeric organic chemical that is coupled with the starting material to generate a new product that may then undergo diastereoselective reactions using intramolecular asymmetric induction.^{67,68} At the end of the reaction, the auxiliary is removed under circumstances that ensures no racemization of the product. Then, it is often recovered and reused. Two of the widely used chiral auxiliaries are: Evans oxazolidinones,⁶⁹ and Oppolzer sultams.⁷⁰ There are many applications with the use of chiral auxiliaries.^{67,71}

Example 1. Asymmetric synthesis of (1*S*)-1-methyl-THβC and (1*S*)-1-phenyl-THβC with (2*R*)-2-amino-2-phenylethanol as a chiral auxiliary. Qais *et al.* synthesized (1*S*)-1-methyl-THβC and (1*S*)-1-phenyl-THβC with the help of (2*R*)-2-amino-2-phenylethanol as a chiral auxiliary.⁷²

1-Benzyl-3-(2-bromoethyl)indole **46** underwent Vilsmeier–Haack reaction to get **47** (50% yield). With (2*R*)-2-amino-2-phenylethanol at RT for 1 hour, **47** formed the iminium salt **48** after azeotropic distillation with benzene. Treatment with triethylamine (Et₃N) at –5 °C for 1 hour in chloroform/DCM then cyclized **48** into (3*R*,11*bS*)-**49** (85% de); after recrystallization from ethanol, it was found in 100% de with 62% yield. Then, (3*R*,11*bS*)-**49** was reacted with two Grignard reagents (MeMgI and PhMgI) at –78 °C for 1 hour to give 90% de of (3*R*)-**50a** and (3*R*)-**50b**. The purification process involved column chromatography on silica gel. Hydrogenolysis on Pd(OH)₂-carbon at RT for 12 hours will remove the chiral auxiliary, and sodium in liquid ammonia removed the *N*-benzyl group with 100% ee of (1*S*)-1-methyl-THβC (1*S*)-**14a** and (1*S*)-1-phenyl-THβC (1*S*)-**14b** (Scheme 13).



Scheme 13 Synthesizing (1*S*)-1-methyl-THβC and (1*S*)-1-phenyl-THβC with (2*R*)-2-amino-2-phenylethanol as chiral auxiliary.

So, the (*R*)-configured chiral auxiliary (2*R*)-2-amino-2-phenylethanol predominantly produced (1*S*)-1-substituted-THβCs.

Example 2. Enantioselective Synthesis of (*R*)-tetrahydroharman with chiral acetylenic sulfoxides as chiral auxiliaries. Lee *et al.* used two chiral acetylenic sulfoxides, 1-[(*R*)-ethynylsulfinyl]-2-nitrobenzene (**R**)-51a and 1-[(*R*)-ethynylsulfinyl]-4-methylbenzene (**R**)-51b, to enantioselectively synthesize (*R*)-tetrahydroharman by Michael addition and cyclization.⁴¹

At first, tryptamine **13a** was added with (**R**)-51a,b to form **52a** and **52b**. Then **52a,b** was cyclized with TFA or toluene-*p*-sulfonic acid (*p*-TsOH) to form **53a** and **53b** as a major compound. RANEY® nickel desulfurization of **53a,b** then resulted in 80% yield of optically pure (100% ee) (*R*)-tetrahydroharman (1*R*)-**14** (ref. 73 and 74) (overall 57% yield) (Scheme 14).

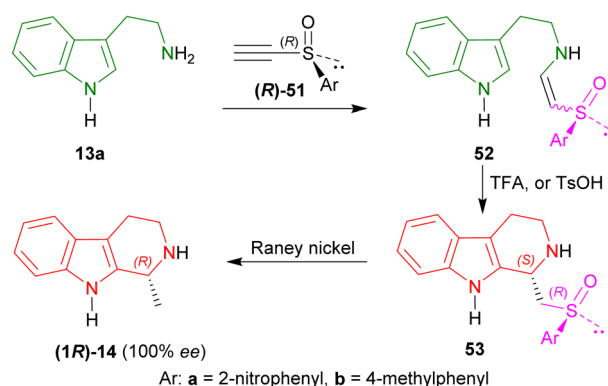
So, (*R*)-configured chiral acetylenic sulfoxides produced (*R*)-configured THβC named (*R*)-tetrahydroharman.

Example 3. Asymmetric synthesis of 1-substituted-THβC using pyroglutamic acid derivatives as chiral auxiliaries. Itoh *et al.* used (*S*)-pyroglutamic acid derivatives as chiral auxiliaries to synthesize 1-substituted-THβC.⁷⁵

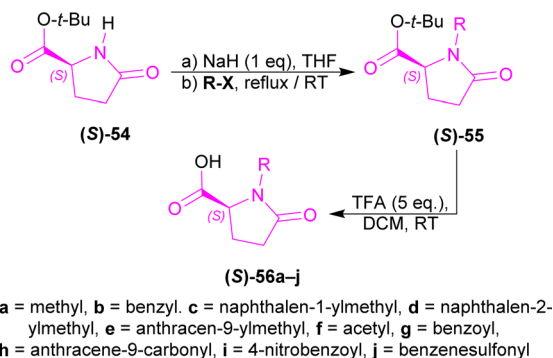
tert-Butyl (*S*)-pyroglutamate (**S**)-54, NaH with various *N*-protecting reagents (R-X) producing (**S**)-55a-j (highest 94% yield for (**S**)-55a); which was converted to (2*S*)-1-substituted-5-oxopyrrolidine-2-carboxylic acid (**S**)-56a-j (highest 97% yield for (2*S*)-1-(2-naphthylmethyl)-5-oxopyrrolidine-2-carboxylic acid (**S**)-56d) using TFA at RT (Scheme 15).

(**S**)-56a-j were reacted with β-carboline 57 and 1-(3-dimethylaminopropyl)-3-ethylcarbodiimide hydrochloride (EDCI) to get **58a-j**. Among these, **58g** reached 99% yield in 20 hours but, **58e** reached 95% yield in 4 hours.

58a-j was reacted with 2,2,2-trichloroethylcarbonyl chloride (2 eq.), and tributyl(prop-2-enyl)stannane (allyltributyltin, 3 eq.) as a nucleophile in DCM at –40 °C for 24 hours to produce **59a-j**. Among these, **59d,f,i** were found in quantitative yields; and **59e,b,g,c** in good yields of 98, 95, 92, and 87%. NaOH in THF-H₂O at RT for 1.5–2.5 hours were needed to remove the chiral auxiliary to give 2,2,2-trichloroethyl 1-prop-2-enyl-1,9-dihydropyrido[3,4-*b*]indole-2-carboxylate **60**.



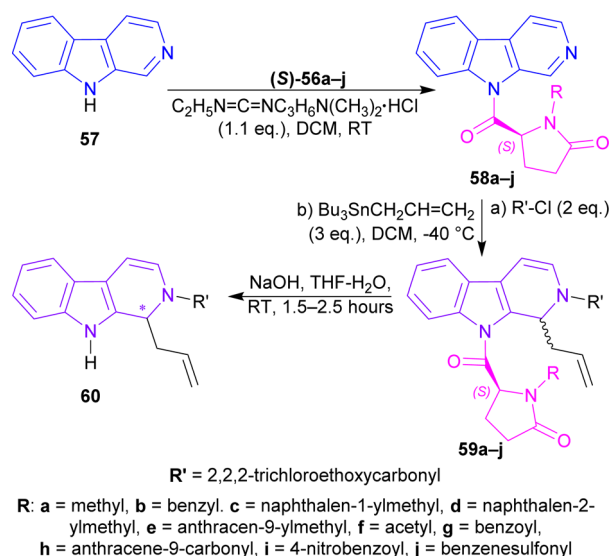
Scheme 14 Synthesizing (*R*)-tetrahydroharman with chiral acetylenic sulfoxides as chiral auxiliary.



Scheme 15 Synthesis of (S)-pyroglutamic acid derivatives.

For **59a–e**, which had alkyl *N*-protecting groups, (*S*)-configuration product (1*S*)-2,2,2-trichloroethyl 1-prop-2-enyl-1,9-dihydropyrido[3,4-*b*]indole-2-carboxylate (**1S**)-**60** were found, and for **59f–j**, which had acyl and sulfonyl *N*-protecting groups, (*R*)-configuration product (1*R*)-2,2,2-trichloroethyl 1-prop-2-enyl-1,9-dihydropyrido[3,4-*b*]indole-2-carboxylate (**1R**)-**60** were noticed. Among **59a–e**, the bulkier the *N*-protecting groups, the more ee was seen in (**1S**)-**60** (e.g., highest 91% ee in (**1S**)-**60** for **59e** having anthracene-9-ylmethyl substituent; lowest 7% ee in (**1S**)-**60** for **59a** having methyl substituent). Among **59f–j**, the bulkiness of substituents did not affect ee of (**1R**)-**60** that much, only lowered the % yields (Scheme 16).

After that, silyl enol ethers **61a–e**⁷⁶ were used as nucleophiles instead of allyltributyltin. At 0 °C, reaction with (5*S*)-1-(anthracene-9-ylmethyl)-5-(pyrido[3,4-*b*]indole-9-carbonyl)pyrrolidin-2-one **58e** and **61a** reached only 40% yield with 79% ee of 2,2,2-trichloroethyl (1*S*)-1-(2-oxopropyl)-1,9-dihydropyrido[3,4-*b*]indole-2-carboxylate (**1S**)-**62a** in 24 hours; **61b** needed 2.5 hours to reach 79% yield with 86% ee of 2,2,2-trichloroethyl



Scheme 16 N9 addition of chiral auxiliary to the β-carboline, C1 addition of allyltributyltin and N2 protecting, then ultimately removal of the chiral auxiliary.

(1*S*)-1-phenacyl-1,9-dihydropyrido[3,4-*b*]indole-2-carboxylate (**1S**)-**62b**; **61c** gained quantitative yield only at 30 minutes with 82% ee of 2,2,2-trichloroethyl (1*S*)-1-(1-methoxy-2-methyl-1-oxopropan-2-yl)-1,9-dihydropyrido[3,4-*b*]indole-2-carboxylate (**1S**)-**62c**. Reducing the temperature to −40 °C reduced yields to 81 and 83% with **61d** and **61e** respectively even at reaction times of 12 and 19 hours but, increased ee slightly to 88 and 87% of 2,2,2-trichloroethyl (1*S*)-1-(2-oxo-2-phenylmethoxyethyl)-1,9-dihydropyrido[3,4-*b*]indole-2-carboxylate (**1S**)-**62d** and 2,2,2-trichloroethyl (1*S*)-1-(2-benzylsulfanyl-2-oxoethyl)-1,9-dihydropyrido[3,4-*b*]indole-2-carboxylate (**1S**)-**62e** respectively.

61d was chosen to react with **58f,g,i** at −78 °C for 40 hours to produce 2,2,2-trichloroethyl (1*R*)-1-(2-oxo-2-phenylmethoxyethyl)-1,9-dihydropyrido[3,4-*b*]indole-2-carboxylate (**1R**)-**62d** (75–76% ee). Less steric hindered acetyl-substitution **58f** had a 93% yield of (**1R**)-**62d** but more steric hindered benzoyl-substitution **58g** and 4-nitrobenzoyl-substitution **58i** both had quantitative yields of (**1R**)-**62d** (Scheme 17).

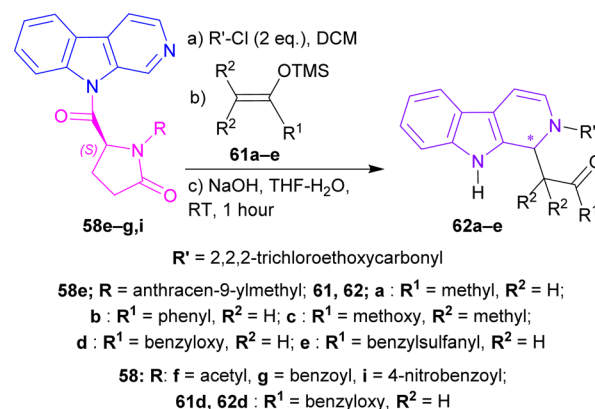
2,2,2-Trichloroethyl (1*S*)-1-(2-oxo-2-phenylmethoxyethyl)-THβC-2-carboxylate (**1S**)-**62d** was reduced with Et₃SiH in DCM at RT for 15 minutes gave rise to (**1S**)-**63** which was again reduced and *N*-2-deprotected with Zn-acetic acid (AcOH) to produce 92% yield of methyl 2-[(1*S*)-2,3,4,9-tetrahydro-1*H*-pyrido[3,4-*b*]indol-1-yl]acetate (**1S**)-**64** (88% ee calculated according to Tietze *et al.*⁷⁷) (Scheme 18).

So, (5*S*)-1-substituted-5-(β-carboline-9-carbonyl)pyrrolidin-2-one having alkyl *N*9-protecting groups as the chiral auxiliary ultimately produced (1*S*)-1-substituted-THβC.

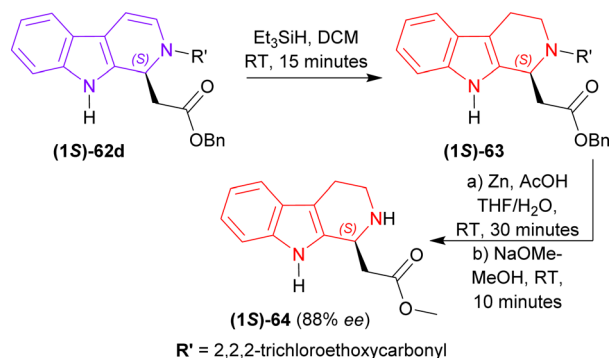
Example 4. (1*R*)-1-Aryl-ethanamines as chiral auxiliaries. Siwicka *et al.* used (1*R*)-1-aryl-ethanamines as chiral auxiliaries to produce 1-substituted-THβCs.⁷⁸

Tryptamine **13a** with diethyl oxalate produced **65**. (1*R*)-1-Phenylethanamine (**1R**)-**66a**⁷⁹ and (1*R*)-1-naphthalen-1-ylethanamine (**1R**)-**66b**⁸⁰ with **65**, produced (*R*)-**67a** and (*R*)-**67b**. Bischler-Napieralski cyclization of (*R*)-**67a,b** with POCl₃ in refluxing DCM gave (*R*)-**68a** and (*R*)-**68b**.

After that, several reducing agents were experimented with e.g., sodium borohydride (NaBH₄), sodium triacetoxyborohydride (NaBH(OAc)₃), sodium tris(2-methylpropanoyloxy)



Scheme 17 N2 protecting, C1 addition of silyl enol ether, and then ultimately removal of the chiral auxiliary.

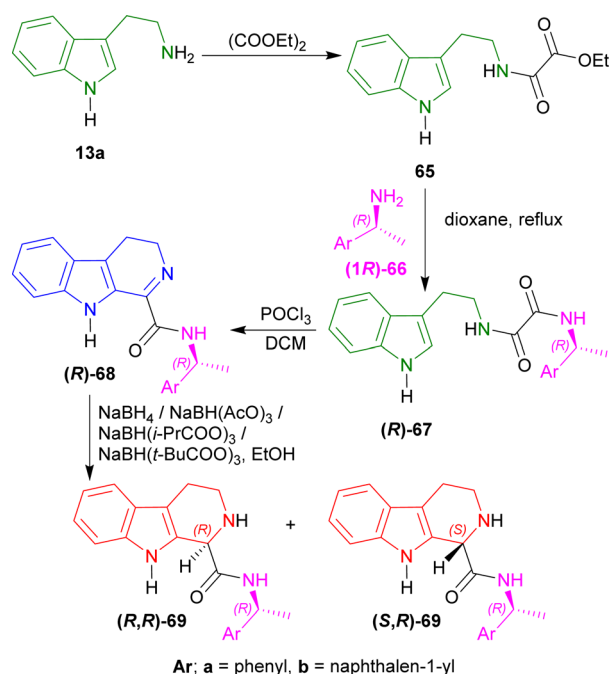


Scheme 18 Reducing to THβC and removing N9 protection.

borohydride ($\text{NaBH}(i\text{-PrCOO})_3$), sodium tris(2,2-dimethylpropanoyloxy)borohydride ($\text{NaBH}(t\text{-BuCOO})_3$) in ethanol to produce dr of 62:38–78:22 for (1*R*)-*N*-[(1*R*)-1-phenylethyl]-THβC-1-carboxamide (*R,R*)-69a and (*S,R*)-69a from (*R*)-68a; and dr of 64:36–83:17 for (1*R*)-*N*-[(1*R*)-1-naphthalen-1-ylethyl]-THβC-1-carboxamide (*R,R*)-69b and (*S,R*)-69b from (*R*)-68b (Scheme 19).

So, the (*R*)-configured chiral auxiliary (1*R*)-1-phenylethanamine and (1*R*)-1-naphthalen-1-ylethanamine predominantly produced (1*R*)-1-substituted-THβCs.

Example 5. Using Ellman's sulfinamide as a chiral auxiliary to synthesize (–)-tetrahydroharman, (–)-komaroidine, (+)-*N*-methyltetrahydroharman, (+)-*N*-acetylkomaroidine, and (–)-harmicine. Reddy *et al.* used Ellman's sulfinamide as a chiral auxiliary to synthesize various chiral 1-substituted-THβCs.⁸¹

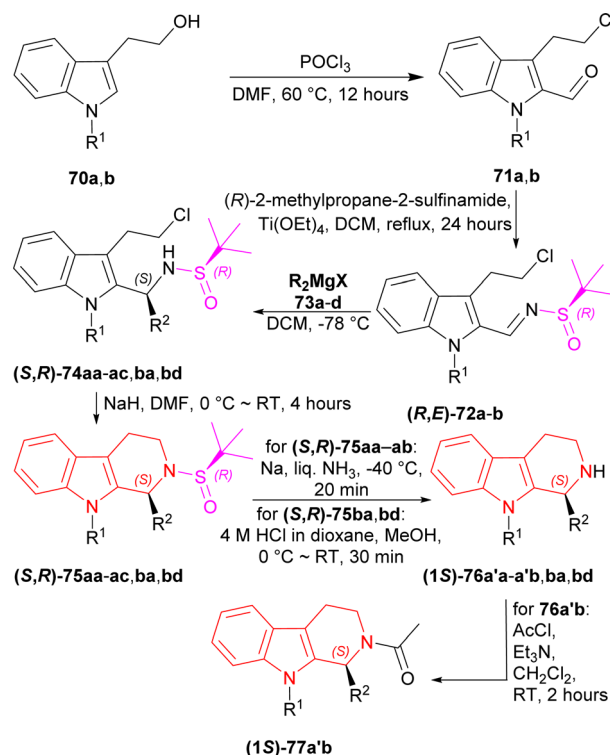


Scheme 19 Addition of chiral auxiliary, Bischler–Napieralski cyclization, and reduction to THβC.

2-(1-Benzylindol-3-yl)ethanol **70a** and 2-(1-methylindol-3-yl)ethanol **70b** was reacted with POCl_3 in DMF at 60 °C for 12 hours to produce **71a** and **71b**. Then **71a,b** was refluxed with Ellman's sulfinamide or (*R*)-2-methylpropane-2-sulfinamide as chiral auxiliary,^{82–85} and $\text{Ti}(\text{OEt})_4$ in DCM for 24 hours to have 78% yield of (*R,E*)-**72a** and 79% yield of (*R,E*)-**72b**.⁸⁶

Then it was experimented with various Grignard reagents **73a–d** *e.g.*, MeMgI , PrMgBr , allyl magnesium bromide, and EtMgCl in DCM at –78 °C to have 77–84% yield and 84 to >98% de of (*S,R*)-**74aa**, (*S,R*)-**74ab**, (*S,R*)-**74ac**, (*S,R*)-**74ba**, (*S,R*)-**74bd**; among which (*S,R*)-**74ac** had the highest de of >98%.⁸⁷ Base-catalyzed cyclization⁸⁸ of (*S,R*)-**74aa–ac,ba,bd** with NaH in DMF at 0 °C to RT for 4 hours gave rise to 72–85% yield of (*S,R*)-**75aa**, (*S,R*)-**75ab**, (*S,R*)-**75ac**, (*S,R*)-**75ba**, (*S,R*)-**75bd**; among which (*S,R*)-**75ac** had the highest yield of 85%.

(*S,R*)-**75aa,ab** with Na in liquid NH_3 (ref. 89) at –40 °C for 20 minutes removed the chiral auxiliary, N9-benzyl and produced 70% yield of (–)-tetrahydroharman (**1S**)-**76a'a** and 68% yield of (–)-komaroidine (**1S**)-**76a'b**. (*S,R*)-**75ba,bd** with 4 M HCl in dioxane in MeOH at 0 °C to RT for 30 minutes removed the chiral auxiliary and produced 87% yield of (+)-*N*-methyltetrahydroharman (**1S**)-**76ba** and 83% yield of (1*S*)-1-ethyl-9-methyl-1,2,3,4-tetrahydropyrido[3,4-*b*]indole (**1S**)-**76bd**.



70, 71, 72; **R¹** : **a** = benzyl, **b** = methyl.

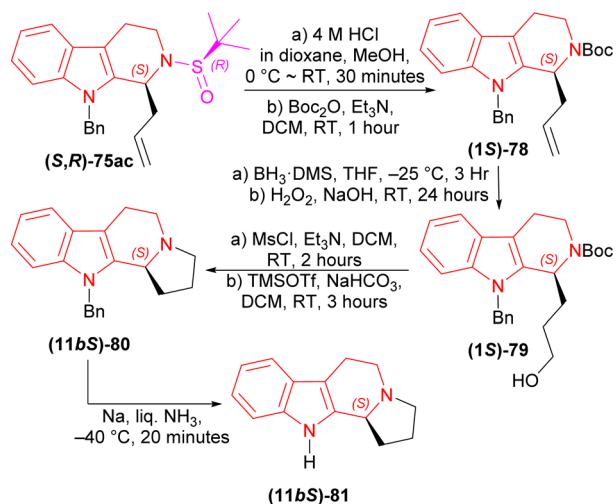
73; **R²** : **a** = methyl, **b** = propyl, **c** = but-3-en-1-yl, **d** = ethyl.

74, 75; **R¹** = benzyl; **R²** : **aa** = methyl, **ab** = propyl, **ac** = but-3-en-1-yl.

74, 75, 76; **R¹** = methyl; **R²** : **ba** = methyl, **bd** = ethyl.

76; **R¹** = H; **R²** : **a'a** = methyl, **a'b** = propyl. **77a'b;** **R¹** = H; **R²** = propyl.

Scheme 20 Addition of chiral auxiliary, base-catalyzed cyclization to THβC and removal of the chiral auxiliary.



Scheme 21 Removal of chiral auxiliary and protecting *N*-2, cyclization of the fourth ring, removal of *N*-9 protection.

(1S)-76a'b with acetyl chloride and Et₃N in DCM at RT for 2 hours produced an 87% yield of (+)-*N*-acetylkomaroidine (1S)-77a'b (Scheme 20).

(1S)-9-Benzyl-2-[(*R*)-*tert*-butylsulfinyl]-1-prop-2-enyl-THβC (S,R)-75ac with 4 M HCl in dioxane in MeOH solvent at 0 °C to RT for 30 minutes; *tert*-butyl (2-methylpropan-2-yl)oxycarbonyl carbonate (Boc₂O) and Et₃N in DCM at RT for 1 hour produced 88% yield of (1S)-78. Then (1S)-78 with BH₃·DMS in THF at −25 °C for 3 hours; H₂O₂ in NaOH at RT for 24 hours gave 83% yield of (1S)-79. After that, (1S)-79 with methanesulfonyl chloride (MsCl) and Et₃N in DCM at RT for 2 hours; TMSOTf and NaHCO₃ in DCM at RT for 3 hours had 70% yield of (11bS)-80. Lastly, (11bS)-80 with Na in liquid NH₃ at −40 °C for 20 minutes gave the ultimate product (−)-harmicine (11bS)-81 of 72% yield (Scheme 21).

So, Ellman's sulfinamide as chiral auxiliary produced (1S)-1-substituted-THβCs e.g., (−)-tetrahydroharman, (−)-komaroidine, (+)-*N*-methyltetrahydroharman, (+)-*N*-acetylkomaroidine, and (−)-harmicine which all have various important pharmacological activities.^{12,90}

Method 3. Asymmetric transfer hydrogenation reaction with chiral catalysts

The transfer hydrogenation reaction, which involves the addition of hydrogen to a molecule from a non-H₂ source, is a versatile and effective approach for producing various hydrogenated compounds. This method is gaining popularity in hydrogenation research as an appealing alternative to direct hydrogenation. The key reasons for its growing interest include: (i) it eliminates the need for potentially hazardous pressurized H₂ gas and complex experimental setups, (ii) the hydrogen donors used are typically affordable, easy to handle, and widely available, (iii) the main byproduct can often be recycled, and (iv) the catalysts involved are generally easy to obtain and not highly sensitive.^{91–101}

ATH emerged in the early 1980s. The first reports were of the Ru catalyzed ATH.^{93,102,103} ATH that used the late transition-

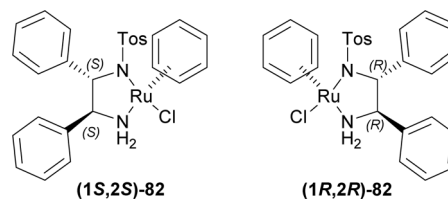


Fig. 3 Chiral catalysts for ATH.

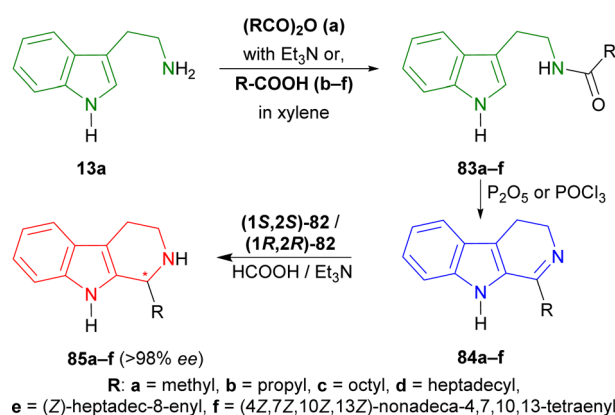
metal catalysts has shown to be one of the most potent strategies for asymmetric reduction of diverse unsaturated substrates to create chiral chemicals.^{94,95,97,98,104–107}

Example 1. ATH to synthesize 1-alkyl-1,2,3,4-tetrahydropyrrodo[3,4-*b*]indole. Roszkowski *et al.* used (1S,2S)-82 and (1R,2R)-82 as chiral catalysts for the ATH of 1-alkyl-3,4-dihydropyrrodo[3,4-*b*]indole (Fig. 3).¹⁰⁸

Tryptamine 13a was reacted with acetic anhydride with Et₃N, butyric acid in xylene, nonanoic acid in xylene, stearic acid in xylene, oleic acid, and arachidonic acid to produce the 83a-f.¹⁰⁹ With P₂O₅ or POCl₃, Bischler–Napieralski cyclization produces 84a-f which instantly underwent ATH³⁴ with (1S,2S)-82 and (1R,2R)-82.⁷⁷ All ATH products had >98% ee. For 84a-f, (1S,2S)-82 gave 70–85% yields of (1R)-1-substituted-THβCs (1R)-85a-f; and for 84a-e, (1R,2R)-82 gave 77–88% yields of (1S)-1-substituted-THβCs (1S)-85a-e. Highest yield of 88% was found for (1S)-1-propyl-THβC (1S)-85b and lowest yield of 70% for (1R)-1-[(4Z,7Z,10Z,13Z)-nonadeca-4,7,10,13-tetraenyl]-THβC (1R)-85f having highly sterically hindered substituents. Switching catalyst from (1S,2S)-82 to (1R,2R)-82 lowered the % yields of the products of (1S)-85a,c,d slightly by 2–4% from that of (1R)-85a,c,d. (1S)-85b had 9% more yield than (1R)-1-propyl-THβC (1R)-85b while (1S)-1-[(Z)-heptadec-8-enyl]-THβC (1S)-85e had 7% less yield than (1R)-1-[(Z)-heptadec-8-enyl]-THβC (1R)-85e (Scheme 22).

So, (1S,2S)-82 chiral catalyst produced (1R)-1-substituted-THβCs and (1R,2R)-82 chiral catalyst produced (1S)-1-substituted-THβCs predominantly.

Example 2. ATH to synthesize of (*R*)-harmicine and (*R*)-desbromoarborescine A. Szawka *et al.* used (1S,2S)-82



Scheme 22 Synthesizing *N*-[2-(1*H*-indol-3-yl)ethyl]amides from tryptamine, then 1-substituted-DHβCs, and ultimately ATH to get 1-substituted-THβCs with chiral catalysts.

(Fig. 3) for the ATH to synthesize of (*R*)-harmicine and (*R*)-desbromoarborescicine A.¹¹⁰

Oxolan-2-one (γ -butyrolactone) **86a**¹, and oxan-2-one (δ -valerolactone) **86a**² were treated with tryptamine **13a** produced **87aa**¹ (ref. 111) (78% yield), and **87aa**² (ref. 112) (87% yield). Then Bischler-Napieralski cyclization in POCl₃ gave iminium salts **88aa**¹ and **88aa**². Immediate ATH of **88aa**^{1,2} with (1*S*,2*S*)-**82** ultimately gave rise to (11*bR*)-2,3,5,6,11,11*b*-hexahydro-1*H*-indolizino[8,7-*b*]indole or (*R*)-harmicine (**R**)-**89aa**¹ (81% yield, 79% ee) and (12*bR*)-1,2,3,4,6,7,12,12*b*-octahydroindolo[2,3-*a*]quinolizine or (*R*)-desbromoarborescicine A (**R**)-**89aa**² (84% yield, 90.5% ee) (Scheme 23).

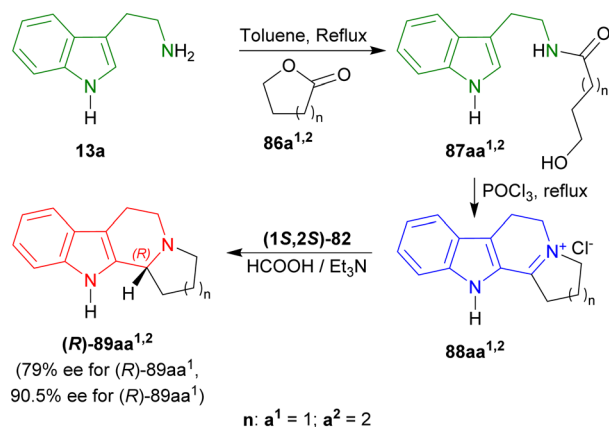
So, (1*S*,2*S*)-**82** chiral catalyst produced (1*R*)-1-substituted-TH β Cs predominantly.

Example 3. ATH to synthesize of (*R*)-tryptargine. Czarnocki *et al.* used (1*S*,2*S*)-**82** (Fig. 3) to synthesize (*R*)-tryptargine *via* ATH.¹¹³

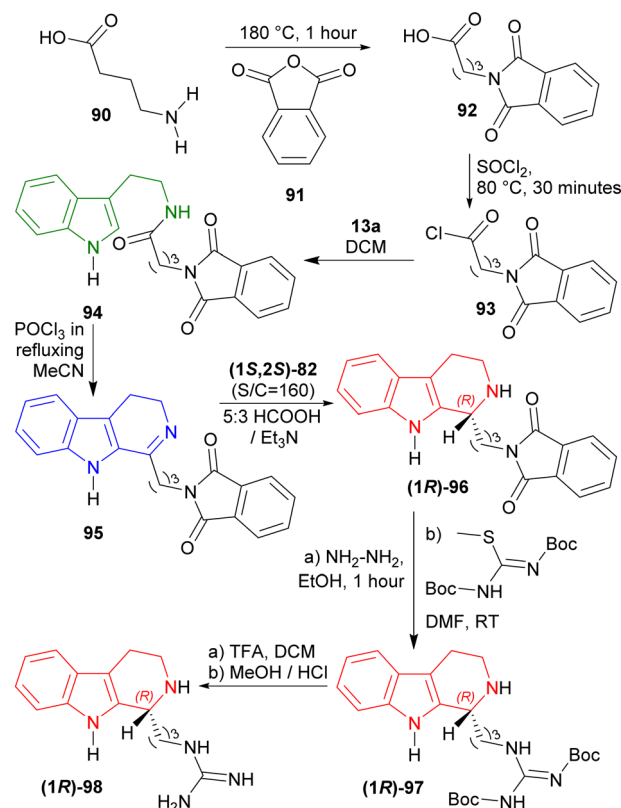
4-Aminobutanoic acid **90** was treated with 2-benzofuran-1,3-dione (phthalic anhydride) **91** at 180 °C for 1 hour¹¹⁴ to give **92** which was turned into **93** with sulfonyl chloride at 80 °C for 30 minutes. This was reacted with **13a** in DCM to get 82% yield of **94**. It was reacted with POCl₃ in refluxing acetone (MeCN) to give 85% yield of **95** *via* Bischler-Napieralski reaction. After that, ATH of **95** with (1*S*,2*S*)-**82** (S : C ratio of 160 : 1) in 5 : 3 azeotropic solution of formic acid (HCOOH) : Et₃N³⁴ afforded 92% yield of (**1R**)-**96** (>98% ee).

(**1R**)-**96** was reacted with hydrazine in ethanol at RT for 1 hour to remove the phthaloyl group which was readily subjected to *tert*-butyl *N*-[(2-methylpropan-2-yl)oxycarbonylamino]-methylsulfanylmethylidene]carbamate in DMF at RT to get 53% yield of (**1R**)-**97**. At the last step, Boc group of (**1R**)-**97** was removed by TFA in DCM at RT, and successive evaporation with methanolic HCl provided the final product HCl salt of (*R*)-tryptargine (**1R**)-**98** in quantitative yield. The isolated compound's analytical results were entirely consistent with what was previously published by Cesar *et al.*¹¹⁵ (Scheme 24).

So, (1*S*,2*S*)-**82** chiral catalyst produced HCl salt of (*R*)-tryptargine predominantly.



Scheme 23 Synthesizing *N*-[2-(1*H*-indol-3-yl)ethyl]hydroxamides from tryptamine, then 1-substituted-DH β Cs, and ultimately ATH to get 1-substituted-TH β Cs with chiral catalysts.



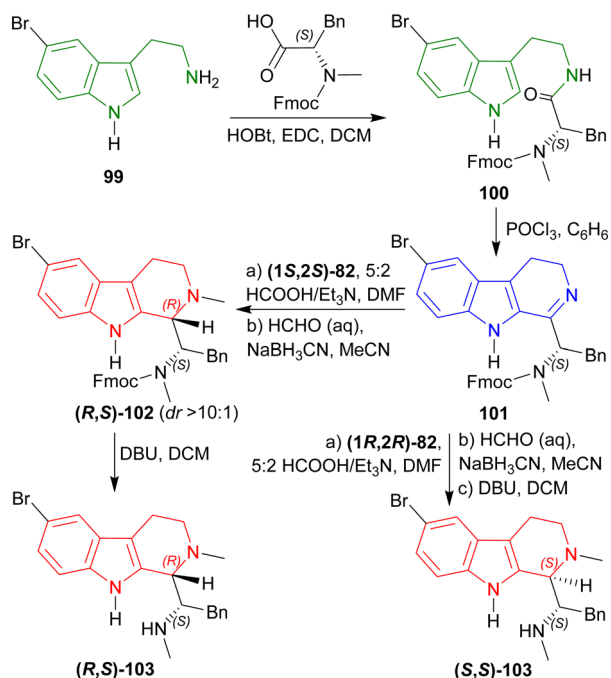
Scheme 24 ATH with chiral catalysts to synthesize HCl salt of (*R*)-tryptargine.

Example 4. Asymmetric hydrogen-transfer to synthesize eudistomidin B and its diastereomer. Takahashi *et al.* used (1*S*,2*S*)-**82** and (1*R*,2*R*)-**82** (Fig. 3) to synthesize eudistomidin B and its diastereomer *via* asymmetric hydrogen-transfer.¹¹⁶

2-(5-Bromo-1*H*-indol-3-yl)ethanamine **99** (ref. 117) with (2*S*)-2-[9*H*-fluoren-9-ylmethoxycarbonyl(methyl)amino]-3-phenylpropanoic acid,¹¹⁸ 1,2-dichloroethane (EDC), and HOBT in DCM produced **100**. Bischler-Napieralski cyclization¹¹⁹ of **100** in benzene gave rise to **101**. The compound **101** was then treated with (1*S*,2*S*)-**82** followed by 5 : 2 HCOOH/Et₃N in DMF and *N*-2 was methylated with aq. HCHO, NaBH₃CN, in CH₃CN into (*R*,*S*)-**102** with 89% yield (predominating (*S*,*S*)-**102** by >10 : 1 dr). The Fmoc group was removed by 2,3,4,6,7,8,9,10-octahydropyrimido [1,2-*a*]azepine (DBU) in DCM to produce (1*S*)-1-[(1*R*)-6-bromo-2-methyl-1,3,4,9-tetrahydropyrido[3,4-*b*]indol-1-yl]-*N*-methyl-2-phenylethanamine (*R*,*S*)-**103** (eudistomidin B) in 85% yield. But when (1*R*,2*R*)-**82** was used, (1*S*)-1-[(1*S*)-6-bromo-2-methyl-1,3,4,9-tetrahydropyrido[3,4-*b*]indol-1-yl]-*N*-methyl-2-phenylethanamine (*S*,*S*)-**103** (diastereomer of eudistomidin B) was found in 78% yield (Scheme 25).

So, (1*S*,2*S*)-**82** chiral catalyst produced (1*R*)-1-substituted-TH β C eudistomidin B; and (1*R*,2*R*)-**82** chiral catalyst produced (1*S*)-1-substituted-TH β C, the (*S*,*S*)-diastereomer of eudistomidin B.

Example 5. Transfer hydrogenation reaction of hydroxylactams catalyzed by chiral phosphoric acid. Yin *et al.* used (*S*)-



Scheme 25 Asymmetric synthesis of eudistomidin B and its diastereomer.

BINOL, VAPOL, and SPINOL-derived chiral phosphoric acid catalysts (Fig. 4) for ATH of hydroxylactams.¹²⁰

Tryptamine and its derivatives **13a–j** were refluxed with phthalic anhydride **91** in toluene, and trifluoromethanesulfonic acid (TfOH) in DCM to give 2-hydroxy-10,20-diazapentacyclo[11.7.0.0^{2,10}.0^{3,8}.0^{14,19}]icosa-1(**13**),3,5,7,14,16,18-heptaen-9-one and its derivatives **104a–j** in 39–63% yields (Scheme 26).^{121,122}

(S)-BINOL, VAPOL, and SPINOL-derived chiral phosphoric acid catalysts (**S**)-**35c**, **e–i**, (**S**)-**105a**, and (**R**)-**105b**^{123,124} (Fig. 4) respectively were tested for transfer hydrogenation reaction of 2-hydroxy-10,20-diazapentacyclo[11.7.0.0^{2,10}.0^{3,8}.0^{14,19}]icosa-1(**13**),3,5,7,14,16,18-heptaen-9-one **104a** with a Hantzsch ester (diethyl 2,6-dimethyl-1,4-dihydropyridine-3,5-dicarboxylate, 2 eq.) as the hydride source to give (2R)-10,20-diazapentacyclo

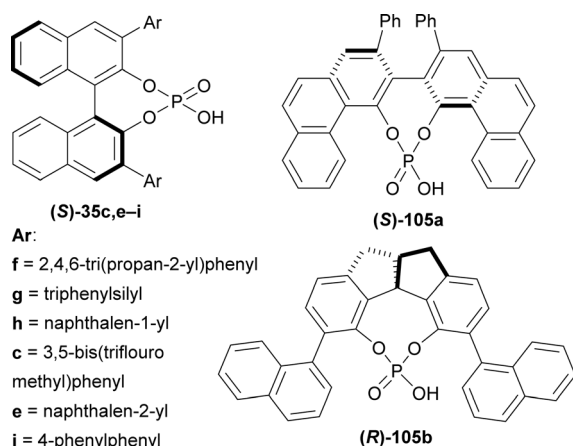
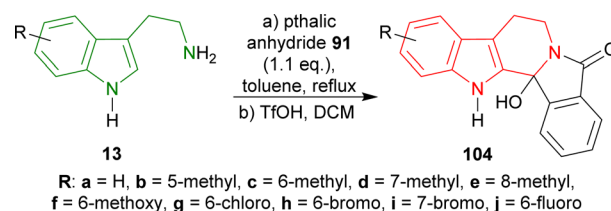


Fig. 4 (S)-BINOL, VAPOL, and SPINOL derived chiral phosphoric acid.



Scheme 26 Synthesis of hydroxylactams from tryptamine and its derivatives.

[11.7.0.0^{2,10}.0^{3,8}.0^{14,19}]icosa-1(**13**),3,5,7,14,16,18-heptaen-9-one (**R**)-**106a** in DCM at RT. Among the catalysts, (**S**)-**35g** provided with highest ee of 52% with 88% yield of the product. After that, different solvents were tested among which dioxane had highest ee of 75% with 84% yield (Scheme 27).

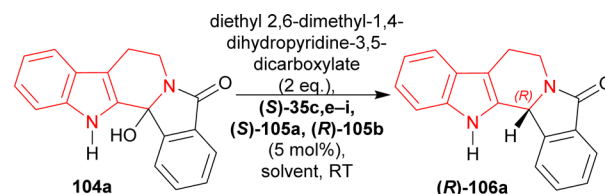
Then, a different Hantzsch ester (di-tert-butyl 2,6-dimethyl-1,4-dihydropyridine-3,5-dicarboxylate, 2 eq.) was tested as hydride donor with the presence of (**S**)-**35g** that resulted in 91% yield and 80% ee of (**R**)-**106a** from **104a** in 24 hours. But reactions using 3, 4, or 5 Å meshes and magnesium sulphate did not increase the yield or ee (Scheme 28).

Under the optimized conditions, **104b–j** were converted to (**R**)-**106b–j** with (**S**)-**35g**; among which (**R**)-**106b–f** containing an electron-rich group had 68–93% yields with 77–85% ee, and (**R**)-**106g–j** containing an electron-poor group had 90–94% yields with 82–90% ee (Scheme 29).

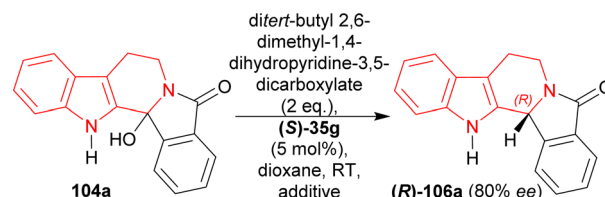
So, (**S**)-BINOL-derived chiral phosphoric acid produced (1R)-1-substituted-THβCs predominantly.

Example 6. Total synthesis of (+)-reserpine by primary amine catalysts and [Ir(COD)(PCy₃)(py)]BAR_F. Rajapaksa *et al.* synthesized (+)-reserpine with the help of primary amine catalysts and [Ir(COD)(PCy₃)(py)]BAR_F as a chiral catalyst for ATH.⁴²

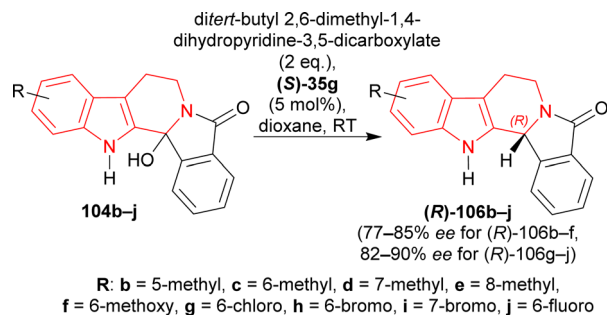
1° amine catalyst *e.g.*, hexan-1-amine **107**, (2S)-2-[[[(1R,2R)-2-aminocyclohexyl]carbamothioylamino]-N-benzhydryl-N,3,3-trimethylbutanamide (**S,R,R**)-**108** or (2R)-2-[[[(1S,2S)-2-aminocyclohexyl]carbamothioylamino]-N-benzhydryl-N,3,3-



Scheme 27 Screening of chiral phosphoric acid catalysts for the acid catalyzed ATH of hydroxylactam.



Scheme 28 Screening of additive for the acid catalyzed ATH of hydroxylactam.



Scheme 29 Acid catalyzed ATH reaction of hydroxylactams by (S)-35g under the optimized conditions.

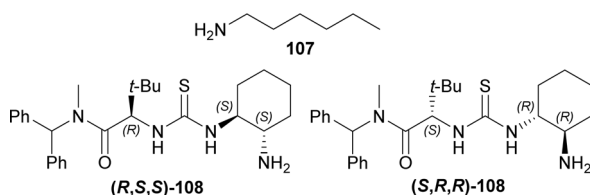


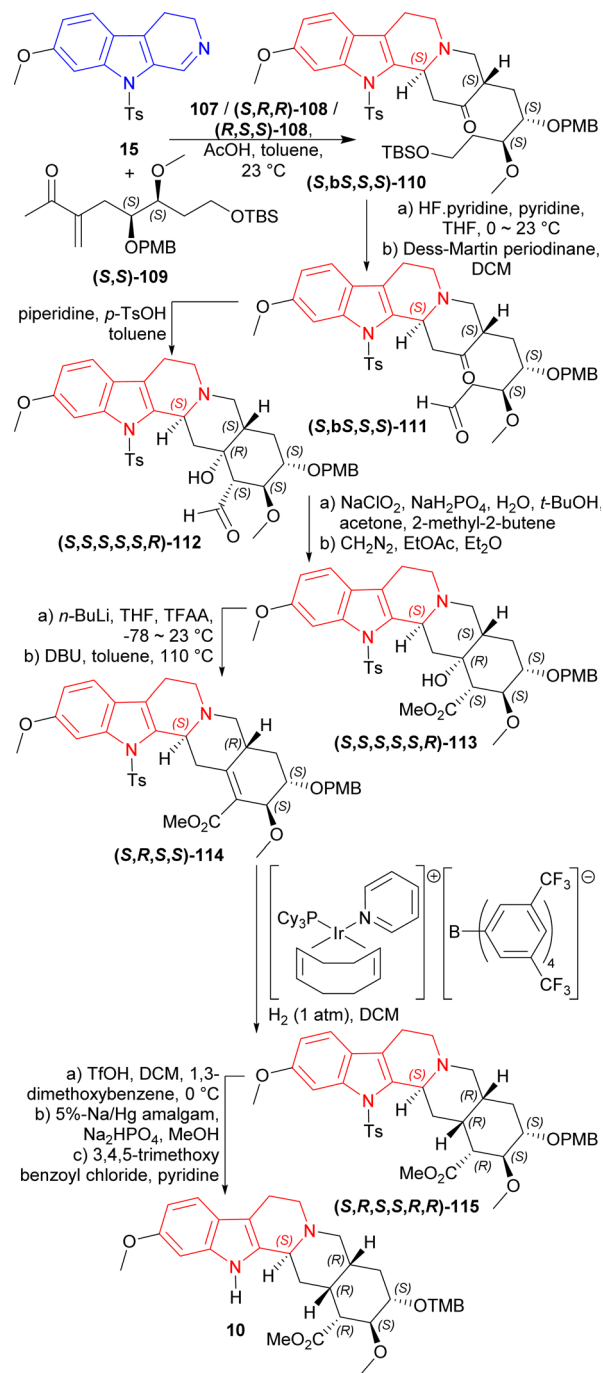
Fig. 5 1° amine catalysts for coupling of 7-methoxy-9-tosyl-DHβC.

trimethylbutanamide (*R,S,S*)-108 (Fig. 5) were used to couple 7-methoxy-9-(4-methylphenyl)sulfonyl-3,4-dihydropyrido[3,4-*b*]indole (7-methoxy-9-tosyl-DHβC) **15** with (*S,S*)-109 (ref. 125) (1.2 eq.)^{126,127} around acetic acid in toluene at 23 °C. 100 mol% of **107** had 90% conversion rate after 9 Days with 1 : 1 dr of the (*S,bS,S,S*)-110 and (*R,bR,S,S*)-110. 20 mol% of (*S,R,R*)-108 (ref. 125) produced >99% conversion after 6 days with 11.5 : 1 : 1.8 dr of the (*S,bS,S,S*)-110, (*R,bR,S,S*)-110, and (*R,bS,S,S*)-110; ultimately yielding 76% of the desired (*S,bS,S,S*)-110. Here, the use of 20 mol% (*R,S,S*)-108 produced greater dr for (*R,bR,S,S*)-110. The cleavage of the primary TBS ether of (*S,bS,S,S*)-110 was done in two steps by pyridine-buffered HF in pyridine at 0 °C to 23 °C; and then oxidation with the Dess–Martin periodinane in DCM producing (*S,bS,S,S*)-111. Piperidine and *p*-TsOH was treated with (*S,bS,S,S*)-111 to produce an intramolecular enamine aldol (*S,S,S,S,S,R*)-112. Pinnick oxidation followed by esterification with diazomethane of (*S,S,S,S,S,R*)-112 provided (*S,S,S,S,S,R*)-113. Trifluoroacetylation of (*S,S,S,S,S,R*)-113 and subsequent elimination by DBU gave (*S,R,S,S*)-114. Hydrogenation with H₂ (1 atm) in DCM and [Ir(COD)(PCy₃)(py)]BAR_F (ref. 128 and 129) gave 6 : 1 dr of (*S,R,S,S,R,R*)-115 (44% isolated yield) and (*S,R,S,S,R,S*)-115. Treating (*S,R,S,S,R,R*)-115 with TFOH, sodium–mercury amalgam, and 3,4,5-trimethoxy benzoyl chloride¹³⁰ resulted in cleavage of PMB ether (86% yield), cleavage of tosyl protective group (69% yield), and esterification (90% yield) respectively; which ultimately gave (+)-reserpine **10** (Scheme 30).

So, 1° amine catalysts and [Ir(COD)(PCy₃)(py)]BAR_F created (1*S*) and (19*R*,20*R*)-chiral centers predominantly to produce (+)-reserpine ultimately.

Method 4. Asymmetric addition reaction

(*S*)-Proline catalyzed asymmetric addition reactions^{131,132} are getting popularized day-by-day. In this method, unsaturated C1



Scheme 30 Total synthesis of (+)-reserpine by primary amine catalysts and [Ir(COD)(PCy₃)(py)]BAR_F.

and N2 get saturated and 1-substituted-THβCs are produced with the help of a ketone. Besides that, cycloaddition¹³³ is also being used highly in synthetic organic chemistry,^{134,135} specially [3 + 3] cycloaddition to produce heterocyclic compounds.^{136–140}

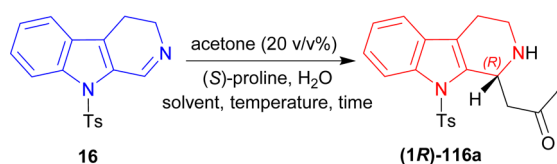
Example 1. Asymmetric addition reaction of 9-tosyl-DHβC to synthesize the precursor of yohimbine and deserpidine catalyzed by (*S*)-proline. Itoh *et al.* synthesized precursor of yohimbine and deserpidine by asymmetric addition reaction of 9-tosyl-DHβC with ketones and proline-catalyzation.⁴³

9-Tosyl-DH β C **16** (ref. 141) was reacted with 20% (v/v) of MeCN, (*S*)-proline (30 mol%) in DCM, MeCN, or DMSO at RT for 1.5–2 hours to produce 1-[(1*R*)-9-(4-methylphenyl)sulfonyl-1,2,3,4-tetrahydropyrido[3,4-*b*]indol-1-yl]propan-2-one (**1R**)-**116a** in good to quantitative yield but ee was very low (5–34%). After adding 10 eq. of water in each solvent, ee increased significantly (67–80%) but reaction time also increased (2–3.5 hours). For DCM, increasing water to 50 eq. did not help at all (trace yield). For MeCN, increasing water to 50 eq. increased ee only 2% (with quantitative yield), though 100 eq. of water decreased both the yield and ee. Lastly for DMSO, increasing water gradually to 50, 100, and 150 eq. increased ee to 80, 86, and 87%. So, DMSO was chosen as the solvent.

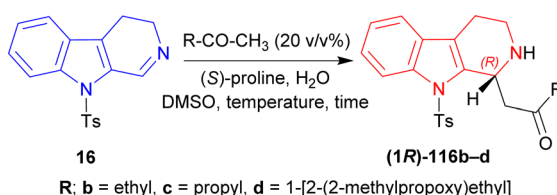
Then at -2°C , the lowest temperature at which the solvent remained liquid, 50 and 100 eq. of water produced similar ee (92–93%) with increasing yields (91 and 99% respectively). Decreasing (*S*)-Proline to 3 mol% did not decrease the yield, but increasing water from 2 to 10 eq. at RT increased ee from 4 to 60%; and increasing water to 50 eq. at -2°C required 23 hours to get 99% yield with 94% ee (Scheme 31).

Then butan-2-one, pentan-2-one, and 4-(2-methylpropoxy)butan-2-one was used instead of MeCN with 50 mol% of (*S*)-proline. Without water, the products had low ee of 7–28% with 57–78% yields at RT for reaction times of 4–26 hours. With 10 eq. of water, 1-[(1*R*)-9-(4-methylphenyl)sulfonyl-1,2,3,4-tetrahydropyrido[3,4-*b*]indol-1-yl]butan-2-one (**1R**)-**116b** and 1-[(1*R*)-9-(4-methylphenyl)sulfonyl-1,2,3,4-tetrahydropyrido[3,4-*b*]indol-1-yl]-4-(2-methylpropoxy)butan-2-one (**1R**)-**116d** had 51–80% ee with 65–81% yields at RT after 8–20 hours of reaction. For 50 eq. of water, products had 75–88% ee with 51–81% yields at RT after 7–20 hours reaction times; at -2°C , (**1R**)-**116b** and 1-[(1*R*)-9-(4-methylphenyl)sulfonyl-1,2,3,4-tetrahydropyrido[3,4-*b*]indol-1-yl]pentan-2-one (**1R**)-**116c** had 89–92% ee with 76–85% yields after reacting 30–48 hours.

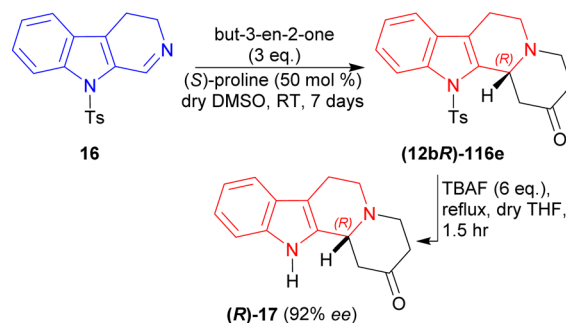
When 5 mol% of (*S*)-proline was used with 50 eq. of water at RT, (**1R**)-**116b** and (**1R**)-**116c** required 36–72 hours to reach 85% ee with 86–98% yield; at -2°C , (**1R**)-**116b** and (**1R**)-**116c** required 120 hours to reach 66–81% ee with 91–92% yield (Scheme 32).



Scheme 31 Screening of solvent, temperature, and time.



Scheme 32 Screening of ketones.



Scheme 33 Synthesis of the precursor of yohimbine and deserpidine.

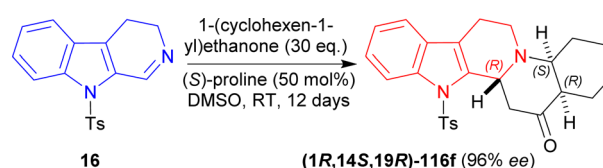
Then, 3 eq. of but-3-en-2-one was reacted with **16** in dry DMSO with (*S*)-proline at RT for 7 days to give (12*bR*)-**116e** (76% yield, 92% ee). Then (12*bR*)-**116e** was refluxed with 6 eq. of tetrabutylazanium;fluoride (TBAF) in dry THF for 1.5 hours to yield 74% (12*bR*)-3,4,6,7,12,12*b*-hexahydro-1*H*-indolo[2,3-*a*]quinolizin-2-one (**R**)-**17** (ref. 142) (92% ee) which has been used as a precursor for the synthesis of yohimbine¹⁴³ and deserpidine¹⁴⁴ (Scheme 33).

So, but-3-en-2-one was asymmetrically added to 9-tosyl-DH β C by (*S*)-proline catalysis to synthesize a (1*R*)-1-substituted-TH β C, the precursor of yohimbine and deserpidine.

Example 2. Synthesis of enantiomer of dihydrocorynantheol. Itoh *et al.* synthesized enantiomer of dihydrocorynantheol by asymmetric addition reaction of 9-tosyl-DH β C with ketones and proline-catalyzed.¹²⁶

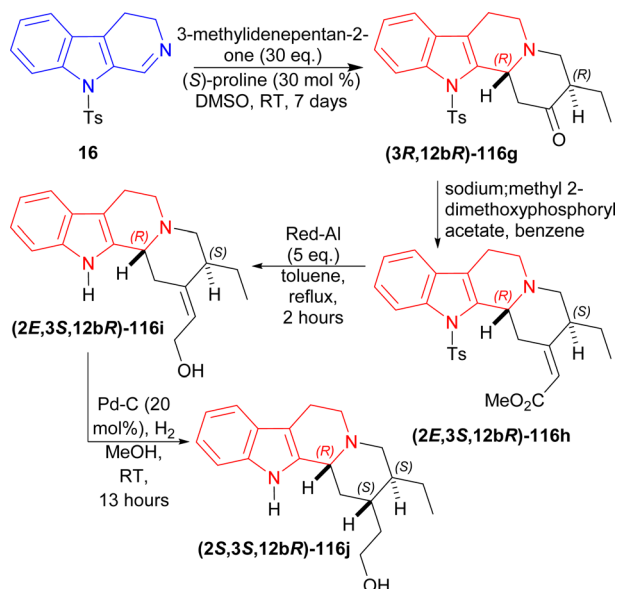
16 was reacted with 30 eq. of 1-(cyclohexen-1-yl)ethanone, 50 mol% of (*S*)-proline in DMSO at RT for 12 days to produce 91% yield and 96% ee of (1*R*,14*S*,19*R*)-3-(4-methylphenyl)sulfonyl-3,13-diazapentacyclo[11.8.0.0^{2,10}.0^{4,9}.0^{14,19}]henicosa-2(10),4,6,8-tetraen-20-one (**1R,14S,19R**)-**116f** (Scheme 34).

16 was reacted with 30 eq. of 3-methylidenepentan-2-one,¹⁴⁵ 50 mol% of (*S*)-proline in DMSO at RT for 7 days to produce (3*R*,12*bR*)-**116g** (85% yield, 99% ee). After that, (3*R*,12*bR*)-**116g** was reacted with a Wittig reagent (sodium;methyl 2-dimethoxyphosphorylacetate) in benzene to have (2*E*,3*S*,12*bR*)-**116h** in very much higher ratio than its *Z* isomer (*E*:*Z* = 20:1). Then reflux with 5 eq. of Red-Al in DCM for 2 hours reduced (2*E*,3*R*,12*bR*)-**116h** to (2*E*,3*S*,12*bR*)-**116i** (63% yield). Lastly hydrogenation of (2*E*,3*S*,12*bR*)-**116i** with H₂ in presence of 20 mol% Pd-C in methanol at RT for 13 hours yielded 74% (38% total yield) of 2-[(2*S*,3*S*,12*bR*)-3-ethyl-1,2,3,4,6,7,12,12*b*-octahydroindolo[2,3-*a*]quinolizin-2-yl]ethanol (2*S*,3*S*,12*bR*)-**116j** (enantiomer of dihydrocorynantheol) (Scheme 35).



Scheme 34 Generating three chiral centers in a single step (*S*)-proline catalyzed asymmetric addition reaction.





Scheme 35 Synthesizing enantiomer of dihydrocorynantheol by (S)-proline catalyzed asymmetric addition reaction in four steps.

So, 1-(cyclohexen-1-yl)ethenone and 3-methylidenepentan-2-one were asymmetrically added to 9-tosyl-DHβC by (S)-proline catalysis to synthesize (1R)-1-substituted-THβCs in one step and four steps respectively.

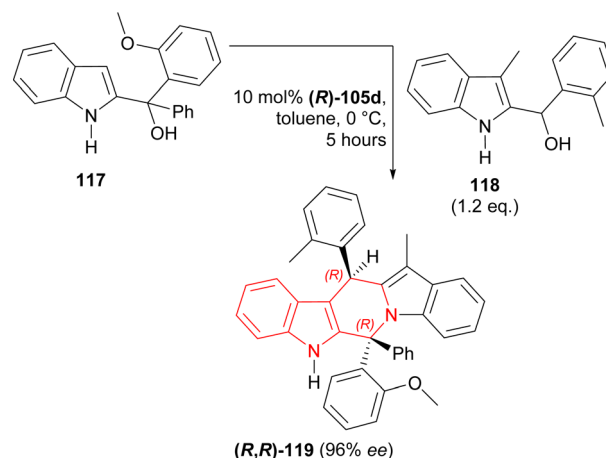
Example 3. Catalytic asymmetric (3 + 3) cycloaddition of different 2-indolylmethanols. Li *et al.* used (R)-H8-BINOL derived catalyst (R)-105c (Fig. 6) for the catalytic asymmetric (3 + 3) cycloaddition of (1H-indol-2-yl)(2-methoxyphenyl)(phenyl)methanol **117** and (3-methyl-1H-indol-2-yl)-(2-methylphenyl)methanol **118**.¹⁴⁶

117 was stirred with 1.2 eq. **118** and 10 mol% (R)-105c for 5 hours in toluene at 0 °C to give catalytic asymmetric (3 + 3) cycloaddition product (6R,13R)-6-(2-methoxyphenyl)-12-methyl-6-phenyl-13-(2-methylphenyl)-6,13-dihydro-5H-pyrido[1,2-a:5,4-b']diindole (R,R)-**119** in 57% yield and 96% ee (Scheme 36).

So, (R)-configured H8-BINOL derived catalyst gave (R,R)-configured catalytic asymmetric (3 + 3) cycloaddition product.

Method 5. Enzymatic catalysis

Biocatalysis have already been employed in a variety of synthesis methods throughout the last few decades.^{147,148} Enzymes are proteins that activate any reaction process by binding to



Scheme 36 Catalytic asymmetric (3 + 3) cycloaddition of two different 2-indolylmethanols.

a particular location on the substrate. They are active at mild reaction conditions (pH, temperature, and reaction media *e.g.*, water). These multifunctional catalysts enable several complex chemical processes to be performed in very mild conditions while maintaining excellent activity, selectivity, and specificity.^{149,150} For these reasons, enzymes are essential for the production of chiral building blocks, enantiopure medicines, and pharmaceuticals.^{150–154}

Example 1. Asymmetric reduction by imine reductase (IRED), freshly prepared or 24 hours old aliquot, from *Amycolatopsis orientalis*. Aleku *et al.* used imine reductase (IRED),^{155–157} freshly purified from *Amycolatopsis orientalis*, AoIRED (UniProt: R4SNK4) to reduce 1-methyl-DHβC **120a** and 1-cyclohexyl-DHβC **120b** to (1R)-1-methyl-THβC (1R)-**121a** and (1R)-1-cyclohexyl-THβC (1R)-**121b** respectively.¹⁵⁸ (1R)-**121a** had >99% ee but only 6% conversion while (1R)-**121b** had 71% ee with 66% conversion. But 24 h old aliquot of AoIRED reduced 7-methoxy-1-methyl-4,9-dihydro-3H-pyrido[3,4-b]indole **120c** to (1S)-7-methoxy-1-methyl-THβC (1S)-**121c** with the highest ee of 79% (15% conversion).

Then, six different variants of AoIRED were used to reduce **120c**. Only AoIRED N241A produced (1R)-7-methoxy-1-methyl-THβC (1R)-**121c** with 96% conversion and 60% ee. AoIRED N171D, AoIRED Y179F, and AoIRED Y179A produced 99% ee of (1S)-**121c** with low conversion of 6, 9, and 10% respectively. AoIRED WT and AoIRED N171A slightly increased % conversion

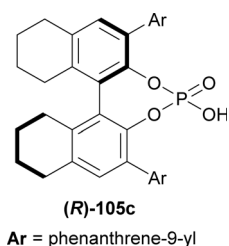
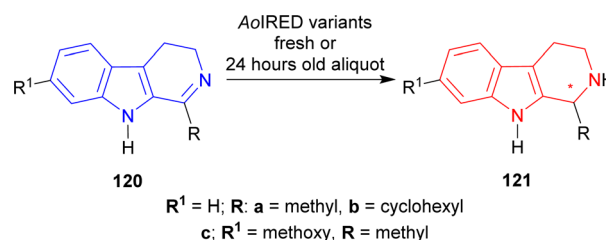


Fig. 6 (R)-H8-BINOL derived chiral phosphoric acid.



Scheme 37 Reduction of 1-substituted-DHβC with fresh or 24 hours old aliquot of AoIRED variants.

to 15 and 18 but ee decreased to 62 and 71 respectively (Scheme 37).

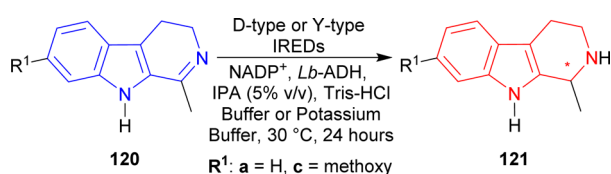
So, freshly purified AoIRED and AoIRED N241A produced (1*R*)-1-substituted-THβCs; 24 h old aliquot of AoIRED and AoIRED N171D, AoIRED Y179F, AoIRED Y179A, AoIRED WT, AoIRED N171A produced (1*S*)-1-substituted-THβCs predominantly.

Example 2. Asymmetric reduction by IREDs of D-type and Y-type. Velikogne *et al.* examined D-type IREDs such as IRED-A-IRED-H and Y-type IREDs such as IRED-I-IRED-N to reduce 1-methyl-4,9-dihydro-3*H*-pyrido[3,4-*b*]indole **120a** and 7-methoxy-1-methyl-4,9-dihydro-3*H*-pyrido[3,4-*b*]indole **120c** with NADP⁺, *Lactobacillus brevis* alcohol dehydrogenase (*Lb*-ADH), pH 7.5 Tris-HCl buffer or pH 6.0 potassium phosphate buffer, 5% (v/v) IPA, 30 °C, 24 hours.¹⁵⁹

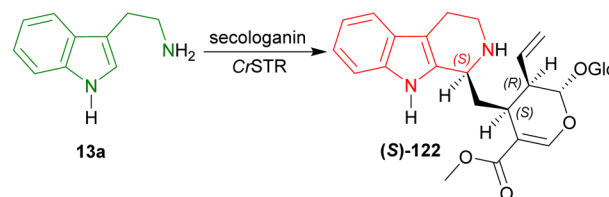
Among the Y-type IREDs, IRED-J (UniProt: D2PR38, collected from *Kribbella flavida* DSM 17836),¹⁶⁰ IRED-K (UniProt: D2AW14, collected from *Streptosporangium roseum* DSM 43201), IRED-L (UniProt: K0F8R0, collected from *Nocardia brasiliensis* ATCC 700358), and IRED-M (UniProt: K0K4C6, collected from *Saccharothrix espanaensis* ATCC 51144) produced (1*S*)-1-methyl-THβC (**1S**)-**121a** of 91–92% conversion rate (96 to >99% ee) and (1*S*)-7-methoxy-1-methyl-THβC (**1S**)-**121c** of 95–88% conversion rate (92 to >99% ee). IRED-I (from *Streptomyces* sp. GF3546, UniProt: M4ZS15)¹⁵⁷ produced >99% ee for both products but (**1S**)-**121a** had 90% conversion while (**1S**)-**121c** had only 9% conversion; and IRED-N (from *Bacillus cereus*, UniProt: J7YM26)¹⁶¹ showed very little conversion (5 and 2% respectively) for both.

The D-type IREDs did not show any good activity at all for **120c**. For **120a**, IRED-A (UniProt: M4ZRJ3, collected from *Streptomyces* sp. GF3587)¹⁵⁶ and IRED-G (UniProt: L8EIW6, collected from *Streptomyces rimosus* ATCC 10970) gave (1*S*)-1-methyl-THβC (**1S**)-**121a** in 8, 21% conversion and 93, >99% ee respectively; IRED-D (UniProt: V7GV82, collected from *Mesorhizobium* sp. L2C089B000) and IRED-F (UniProt: V6KA13, collected from *Streptomyces niveus* NCIMB 11891) gave (1*R*)-1-methyl-THβC (**1R**)-**121a** in 27, 13% conversion and 78, >99% ee respectively. IRED-B (UniProt: Q1EQE0, collected from *Streptomyces kanamyceticus*)¹⁶² did not give any product at all for both reactions while IRED-C (UniProt: W7VJL8, collected from *Micromonospora* sp. M42) did the same as above for **120a** and only had 1% conversion for its product. IRED-E (UniProt: J7LAY5, collected from *Nocardiopsis alba*) and IRED-H (UniProt: I8QLV7, collected from *Frankia* sp. QA3) gave only 1% conversion for both of their products (Scheme 38).

So, IRED-A, IRED-G, IRED-I-IRED-N produced (1*S*)-1-substituted-THβCs; and IRED-D, IRED-F produced (1*R*)-1-substituted-THβCs predominantly.



Scheme 38 Reduction of 1-substituted-DHβC with D-type IREDs e.g., IRED-A-IRED-H, and Y-type IREDs e.g., IRED-I-IRED-N.



Scheme 39 Stereoselective condensation of tryptamine and secologanin with CrSTR.

Example 3. Stereoselective condensation by strictosidine synthase from *Catharanthus roseus* (CrSTR), *Ophiorrhiza pumila* (OpSTR), *Rauwolfia serpentina* (RsSTR) and its V208A variant (RvSTR). In 1977, Stöckigt and Zenk used strictosidine synthase (STR, EC 4.3.3.2) from *Catharanthus roseus* for stereoselectively condensing tryptamine **13a** with secologanin to produce (*S*)-strictosidine (**S**)-**122** for the very first time (Scheme 39).¹⁶³

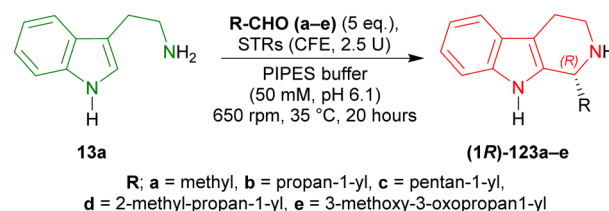
Pressnitz *et al.* tested CrSTR, OpSTR, RsSTR, and RvSTR for the stereoselective condensation of tryptamine **13a** and five small aliphatic aldehydes that gave (1*R*)-1-substituted-THβCs (**1R**)-**123a–e** as products.¹⁶⁴

14–38% conversion and 28–43% ee was seen for (**1R**)-**123a** in case of acetaldehyde as the aliphatic aldehyde. Even, racemic **123a** was found for CrSTR and RsSTR. The ee improved to 61–91% for (**1R**)-**123b** when butanal was used. Overall conversion also increased to 7–45%. Decreased conversion of 4–8% and ee of 46–82% was seen for (**1R**)-**123c** when hexanal was used. So, carbon number in the aliphatic aldehyde was not increased further.

After that, steric hindrance in the aliphatic chain of the aldehyde was increased by the use of 3-methylbutanal instead of butanal. Improved 12–77% conversion and 88 to >98% ee was seen for (**1R**)-**123d** compared to (**1R**)-**123b**.

Lastly, methyl 4-oxobutanoate was used to asymmetrically condense with **13a**. The product (**1R**)-**123e** showed >98% ee for all the STRs and overall increased conversion of 11–95% was seen compared to (**1R**)-**123b**.

CrSTR had generally much lower conversion and ee than any other STRs. While RvSTR showed highest ee in each product, RsSTR had highest conversion for only (**1R**)-**123a,c,e**. RsSTR and RvSTR had same or almost similar conversion and ee for (**1R**)-**123b,c**. RsSTR had higher conversion compared to OpSTR for each product except (**1R**)-**123c**. RvSTR had higher or same ee compared to OpSTR for each product (Scheme 40).



Scheme 40 Stereoselective condensation of tryptamine and small aliphatic aldehydes with CrSTR, OpSTR, RsSTR, and RvSTR.



So, *CrSTR* produced (*S*)-strictosidine with secologanin, while *CrSTR*, *OpSTR*, *RsSTR*, and *RvSTR* produced (1*R*)-1-substituted-THβCs predominantly with small aliphatic aldehydes.

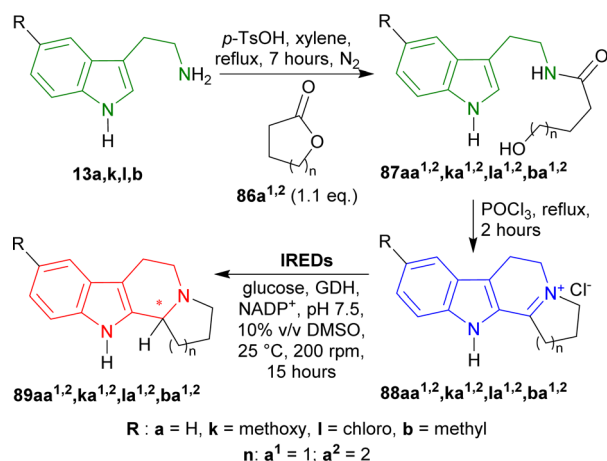
Example 4. Asymmetric synthesis of fused-ring THβCs by imine reductases. Yang *et al.* used four IREDs to enantioselectively reduce iminium salts, produced from the Bischler-Napieralski cyclization of hydroxamides, derived from tryptamine and 5-substituted tryptamines.¹⁶⁵

13a,b,k,l was stirred with 1.1 eq. of oxolan-2-one (γ -butyrolactone) **86a**¹ or oxan-2-one (δ -valerolactone) **86a**² and *p*-TsOH in toluene, refluxed for 7 hours to produce hydroxamides **87aa**^{1,2}, **ba**^{1,2}, **ka**^{1,2}, **la**^{1,2}. Then they were stirred with POCl₃ in toluene, refluxed for 2 hours to get iminium salts **88aa**^{1,2}, **ba**^{1,2}, **ka**^{1,2}, **la**^{1,2} in 58–72% yields by Bischler-Napieralski cyclization.^{110,166} After that, the iminium salts were asymmetrically reduced by IREDs named IR51 (from *Myxococcus fulvus*, Protein Identifier: WP_074958336.1), IR64 (from *Actinocorallia populi*, Protein Identifier: WP_106402132.1), IR86 (from *Paenibacillus lactis*, Protein Identifier: WP_007130043.1),¹⁶⁷ and IR88 (Metagenome (pIR23)).^{168,169}

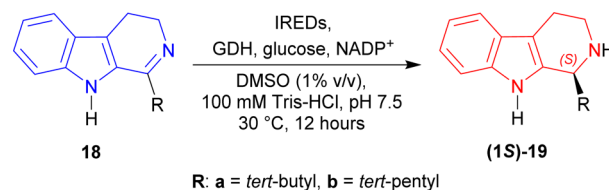
IR51 and IR88 produced only (*R*)-configured products. IR51 had highest yield of >98% and 99% ee for (*R*)-**89aa**¹; other products (*R*)-**89aa**², **ba**^{1,2}, **ka**^{1,2}, **la**^{1,2} had 80–98% yields and 95–99% ee. IR88 showed the best result having 99% ee for all products (*R*)-**89aa**^{1,2}, **ba**^{1,2}, **ka**^{1,2}, **la**^{1,2} with >98% yields except only (*R*)-**89ka**¹ (80–98% yield).

On the other hand, IR64 and IR86 produced only (*S*)-configured products. IR64 did not even react with **88aa**¹, **ka**¹. Among the other products, (*S*)-**89la**¹ had the lowest ee of 29% with 50–80% yield and (*S*)-**89aa**² had the highest ee of 99% with 50–80% yield. (*S*)-**89ba**², **ka**², **la**² had the lowest yields of 10–50% with 82–95% ee. IR86 had 99% ee for (*S*)-**89aa**² with 80–98% yield; 98–99% ee for (*S*)-**89ba**^{1,2}, **la**^{1,2} with 50–98% yields; 76 and 70% ee for (*S*)-**89ka**^{1,2} with 10–50% and 80–98% yields respectively; 80–98% yield for (*S*)-**89aa**¹ with no ee data reported.

Lastly, IR86 and IR88 were used for the preparative scale synthesis of chiral **89aa**^{1,2}, **ba**^{1,2}, **ka**^{1,2}, **la**^{1,2}. IR86 achieved >98 to >99% ee, 77–95% conversion, and 57–82% yields for (*S*)-



Scheme 41 Asymmetrically synthesizing fused-ring THβCs by imine reductases.



Scheme 42 Reduction of 1-substituted-DHβC with IRED-G, IRED-I–IRED-M.

89aa², **ba**^{1,2}, **la**² while, IR88 achieved >99% ee, 79–97% conversion, and 57–74% yields for (*R*)-**89aa**^{1,2}, **ba**^{1,2}, **ka**^{1,2}, **la**^{1,2} (Scheme 41).

So, IR86 produced (*S*)-configured products and IR88 produced (*R*)-configured products predominantly.

Example 5. Asymmetric synthesis of 1-substituted-THβCs by imine reductases. Li *et al.* tested the enzymes IRED-G, IRED-I–IRED-M to reduce 1-*tert*-butyl-4,9-dihydro-3*H*-pyrido[3,4-*b*]indole **18a**.⁴⁴

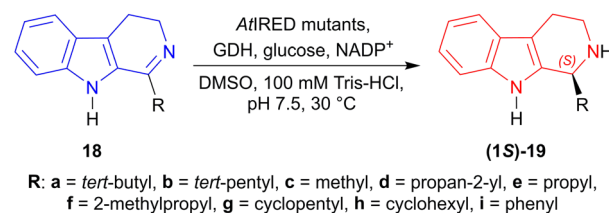
Except IRED-I, others had 97 to >99% conversion of (1*S*)-1-*tert*-butyl-THβC (**1S**)-**19a**; IRED-G (from *Streptomyces*, Accession No.: WP_003985113.1), IRED-I (from *Streptomyces* sp. GF3546, Accession No.: 4OQY) had 40–41% ee, IRED-J (from *Kribbella flavida*, Accession No.: WP_012921542.1) had 73% ee, IRED-M (from *Saccharothrix espanaensis*, Accession No.: WP_015105194.1) had 97% ee, and IRED-K (from *Streptosporangium roseum*, Accession No.: WP_012890722.1), IRED-L (from *Nocardia brasiliensis*, Accession No.: WP_014988976.1) had the highest 99% ee of (**1S**)-**19a**.

Then, IRED-K–IRED-M were tested on 1-*tert*-pentyl-4,9-dihydro-3*H*-pyrido[3,4-*b*]indole **18b**. They produced 79–96% ee but conversion was only 44–70% of (1*S*)-1-*tert*-pentyl-THβC (**1S**)-**19b** (Scheme 42).

Site-saturation mutagenesis on *AtIRED* (from *Amycolatopsis thermoflava*, Accession No.: WP_027931120.1) produced two single mutants named as M118'L and P120'G; one double mutant named as M118'L/P120'G which reduced **18a**–**i** to (**1S**)-**19a**–**i** in 98 to >99% ee.

M118'L reduced 1-propyl-4,9-dihydro-3*H*-pyrido[3,4-*b*]indole **18e** and 1-cyclopentyl-4,9-dihydro-3*H*-pyrido[3,4-*b*]indole **18g** to (1*S*)-1-propyl-THβC (**1S**)-**19e** and (1*S*)-1-cyclopentyl-THβC (**1S**)-**19g** respectively in 69% yields, while 1-(2-methylpropyl)-DHβC **18f** to 1-(2-methylpropyl)-THβC (**1S**)-**19f** in 51% yield.

P120'G reduced **18b** to (**1S**)-**19b** in 78% yield, 1-cyclohexyl-DHβC **18h** to (1*S*)-1-cyclohexyl-THβC (**1S**)-**19h** in 64% yield, and 1-phenyl-DHβC **18i** to (1*S*)-1-phenyl-THβC (**1S**)-**19i** in 30% yield.



Scheme 43 Reduction of 1-substituted-DHβC with *AtIRED* mutants.

M118'L/P120'G reduced **18a** to (**1S**)-**19a** in 87% yield, 1-propan-2-yl-DH β C **18d** to (1S)-propan-2-yl-TH β C (**1S**)-**19d** in 62% yield, and **18c** to (1S)-1-methyl-TH β C (**1S**)-**19c** in 50% yield (Scheme 43).

So, IRED-G, IRED-I-IRED-M and AtIRED mutants produced (1S)-1-substituted-TH β Cs predominantly.

3. Conclusion

Novel natural and synthetic TH β C products continued to be discovered, with ongoing exploration of their biological activity directly related to the C1 chiral center. 1-Substituted-TH β Cs and their derivatives have diverse biological actions, indicating that they are a promising drug scaffold for treating various diseases. We discussed five synthetic methods with the purpose for creating C1 chiral center. For Pictet–Spengler reaction, the highest yield 99% and >97% ee was found from modified Pictet–Spengler reaction for formal syntheses of (–)-suaveoline, (–)-raumacline, and (–)-N^b-methylraumacline intermediates; for chiral auxiliary, the highest 97% yield and highest 91% ee was reported from asymmetric synthesis of 1-substituted-TH β C using pyroglutamic acid derivatives; for ATH with chiral catalysts, the highest afforded 92% yield and highest >98% ee was observed from ATH to synthesize 1-alkyl-1,2,3,4-tetrahydropyrindo[3,4-*b*]indole; for asymmetric addition reaction, the highest 91% yield and 96% ee was recorded from synthesis of enantiomer of dihydrocorynantheol; for enzymatic catalysis, the highest conversion of 95% with >98% ee was obtained from stereoselective condensation by STR from *Rauwolfia serpentina*.

The methods that we have discussed here are the most used and widely found pathways for creating C1 chirality which is crucial for prominent biological activities. More efficient and economically feasible pathways should be revised so they could be applied for synthesizing new promising TH β Cs.

Data availability

No primary research results, software or code have been included and no new data were generated or analysed as part of this review.

Author contributions

M. M. A. Asif: writing – original draft, visualization, writing – review & editing. S. R. Lisa: writing – original draft, writing – review & editing. N. Qais: conceptualization, supervision, writing – review & editing.

Conflicts of interest

There are no conflicts to declare.

References

- 1 M. Hesse, *Alkaloids: Nature's Curse or Blessing?*, WILEY-VCH, Weinheim, 2002.

- 2 A. Daugan, P. Grondin, C. Ruault, A.-C. Le Monnier de Gouville, H. Coste, J. M. Linget, J. Kirilovsky, F. Hyafil and R. Labaudinière, *J. Med. Chem.*, 2003, **46**, 4533–4542.
- 3 R. Skouta, M. Hayano, K. Shimada and B. R. Stockwell, *Bioorg. Med. Chem. Lett.*, 2012, **22**, 5707–5713.
- 4 S. Hotha, J. C. Yarrow, J. G. Yang, S. Garrett, K. V. Renduchintala, T. U. Mayer and T. M. Kapoor, *Angew. Chem., Int. Ed.*, 2003, **42**, 2379–2382.
- 5 J. H. Van Maarseveen, P. H. H. Hermkens, E. De Clercq, J. Balzarini, H. W. Scheeren and C. G. Kruse, *J. Med. Chem.*, 1992, **35**, 3223–3230.
- 6 J. F. Miller, E. M. Turner, R. G. Sherrill, K. Gudmundsson, A. Spaltenstein, P. Sethna, K. W. Brown, R. Harvey, K. R. Romines and P. Golden, *Bioorg. Med. Chem. Lett.*, 2010, **20**, 256–259.
- 7 A. Gellis, A. Dumêtre, G. Lanzada, S. Hutter, E. Ollivier, P. Vanelle and N. Azas, *Biomed. Pharmacother.*, 2012, **66**, 339–347.
- 8 R. A. Davis, S. Duffy, V. M. Avery, D. Camp, J. N. A. Hooper and R. J. Quinn, *Tetrahedron Lett.*, 2010, **51**, 583–585.
- 9 M. Rottmann, C. McNamara, B. K. S. Yeung, M. C. S. Lee, B. Zou, B. Russell, P. Seitz, D. M. Plouffe, N. V. Dharia, J. Tan, S. B. Cohen, K. R. Spencer, G. E. González-Páez, S. B. Lakshminarayana, A. Goh, R. Suwanarusk, T. Jegla, E. K. Schmitt, H.-P. Beck, R. Brun, F. Nosten, L. Renia, V. Dartois, T. H. Keller, D. A. Fidock, E. A. Winzeler and T. T. Diagana, *Science*, 2010, **329**, 1175–1180.
- 10 A. Pictet and T. Spengler, *Ber. Dtsch. Chem. Ges.*, 1911, **44**, 2030–2036.
- 11 B. T. Ho, W. M. McIsaac, K. E. Walker and V. Estevez, *J. Pharm. Sci.*, 1968, **57**, 269–274.
- 12 R. Cao, W. Peng, Z. Wang and A. Xu, *Curr. Med. Chem.*, 2007, **14**, 479–500.
- 13 J. Wang, A. N. Pearce, S. T. S. Chan, R. B. Taylor, M. J. Page, A. Valentin, M.-L. Bourguet-Kondracki, J. P. Dalton, S. Wiles and B. R. Copp, *J. Nat. Prod.*, 2016, **79**, 607–610.
- 14 A. I. Marcus, U. Peters, S. L. Thomas, S. Garrett, A. Zelnak, T. M. Kapoor and P. Giannakakou, *J. Biol. Chem.*, 2005, **280**, 11569–11577.
- 15 T. Akizawa, K. Yamazaki, T. Yasuhara, T. Nakajima, M. Roseghini, G. F. Erspamer and V. Erspamer, *Biomed. Res.*, 1982, **3**, 232–234.
- 16 R. M. Van Wagoner, J. Jompa, A. Tahir and C. M. Ireland, *J. Nat. Prod.*, 1999, **62**, 794–797.
- 17 L. M. M. Cesar, M. A. Mendes, C. F. Tormena, M. R. Marques, B. M. De Souza, D. M. Saidemberg, J. C. Bittencourt and M. S. Palma, *Toxicon*, 2005, **46**, 786–796.
- 18 B. Elgoyhen, P. S. Lorenzo, M. T. Tellez-Iñón and E. Adler-Graschinsky, *J. Pharmacol. Exp. Ther.*, 1992, **261**, 534–539.
- 19 P. K. Fischhof, R. Möslinger-Gehmayer, W. M. Herrmann, A. Friedmann and D. L. Rußmann, *Neuropsychobiology*, 1996, **34**, 29–35.
- 20 R. v. d. Heijden, D. I. Jacobs, W. Snoeijs, D. Hallard and R. Verpoorte, *Curr. Med. Chem.*, 2004, **11**, 607–628.
- 21 S. E. O'Connor and J. J. Maresh, *Nat. Prod. Rep.*, 2006, **23**, 532–547.



- 22 J. Garnier, J. Mahuteau, M. Plat and C. Merienne, *Phytochemistry*, 1989, **28**, 308–309.
- 23 N. Neuss, *Indole and Biogenetically Related Alkaloids*, Academic Press, London, 1980.
- 24 J. M. Müller, E. Schlittler and H. J. Bein, *Experientia*, 1952, **8**, 338.
- 25 A. Itoh, T. Tanahashi, N. Nagakura and T. Nishi, *Phytochemistry*, 2003, **62**, 359–369.
- 26 J. H. van Maarseveen, H. W. Scheeren, E. De Clercq, J. Balzarini and C. G. Kruse, *Bioorg. Med. Chem.*, 1997, **5**, 955–970.
- 27 A. M. Deveau, M. A. Labroli, C. M. Dieckhaus, M. T. Barthen, K. S. Smith and T. L. Macdonald, *Bioorg. Med. Chem. Lett.*, 2001, **11**, 1251–1255.
- 28 H. Osada, C.-B. Cui, R. Onose and F. Hanaoka, *Bioorg. Med. Chem.*, 1997, **5**, 193–203.
- 29 A. M. P. Koskinen, *Asymmetric Synthesis of Natural Products*, Wiley, 1st edn, 2012.
- 30 P. D. Bailey, P. D. Clingan, T. J. Mills, R. A. Price and R. G. Pritchard, *Chem. Commun.*, 2003, 2800–2801.
- 31 H. Waldmann, G. Schmidt, H. Henke and M. Burkard, *Angew. Chem., Int. Ed. Engl.*, 1995, **34**, 2402–2403.
- 32 J. Li, T. Wang, P. Yu, A. Peterson, R. Weber, D. Soerens, D. Grubisha, D. Bennett and J. M. Cook, *J. Am. Chem. Soc.*, 1999, **121**, 6998–7010.
- 33 A. Bischler and B. Napieralski, *Ber. Dtsch. Chem. Ges.*, 1893, **26**, 1903–1908.
- 34 N. Uematsu, A. Fujii, S. Hashiguchi, T. Ikariya and R. Noyori, *J. Am. Chem. Soc.*, 1996, **118**, 4916–4917.
- 35 A. Laine, C. Lood and A. Koskinen, *Molecules*, 2014, **19**, 1544–1567.
- 36 P. Maity, D. Adhikari and A. K. Jana, *Tetrahedron*, 2019, **75**, 965–1028.
- 37 T. Szabó, B. Volk and M. Milen, *Molecules*, 2021, **26**, 663.
- 38 J. Wang, F. Gong, T. Liang, Z. Xie, Y. Yang, C. Cao, J. Gao, T. Lu and X. Chen, *Eur. J. Med. Chem.*, 2021, **225**, 113815.
- 39 Y. Du, A. Semghouli, H. Mei, L. Kiss and J. Han, *Adv. Synth. Catal.*, 2024, **366**, 3050–3084.
- 40 P. D. Bailey, I. D. Collier, S. P. Hollinshead, M. H. Moore, K. M. Morgan, D. I. Smith and J. M. Vernon, *J. Chem. Soc., Chem. Commun.*, 1994, 1559–1560.
- 41 A. W. M. Lee, W. H. Chan, Y. Tao and Y. K. Lee, *J. Chem. Soc., Perkin Trans. 1*, 1994, 477–481.
- 42 N. S. Rajapaksa, M. A. McGowan, M. Rienzo and E. N. Jacobsen, *Org. Lett.*, 2013, **15**, 706–709.
- 43 T. Itoh, M. Yokoya, K. Miyauchi, K. Nagata and A. Ohsawa, *Org. Lett.*, 2003, **5**, 4301–4304.
- 44 Y. Li, X. Yue, Z. Li, Z. Huang and F. Chen, *Org. Lett.*, 2023, **25**, 1285–1289.
- 45 G. Tatsui, *J. Pharm. Soc. Jpn*, 1928, **48**, 92–99.
- 46 Z.-J. Liu and R.-R. Lu, in *The Alkaloids: Chemistry and Pharmacology*, ed. A. Brossi, Academic Press, 1988, vol. 33, pp. 83–140.
- 47 P. R. Benoin, R. H. Burnell and J. D. Medina, *Tetrahedron Lett.*, 1968, **9**, 807–809.
- 48 P. D. Bailey and N. R. McLay, *Tetrahedron Lett.*, 1991, **32**, 3895–3898.
- 49 P. D. Bailey, S. P. Hollinshead and N. R. McLay, *Tetrahedron Lett.*, 1987, **28**, 5177–5180.
- 50 A. P. Krapcho, *Synthesis*, 1982, **1982**, 805–822.
- 51 E. Piers and F. F. Fleming, *J. Chem. Soc., Chem. Commun.*, 1989, 756–757.
- 52 P. D. Bailey and S. P. Hollinshead, *J. Chem. Soc., Perkin Trans. 1*, 1988, 739–745.
- 53 P. Magnus, B. Mugrage, M. R. DeLuca and G. A. Cain, *J. Am. Chem. Soc.*, 1990, **112**, 5220–5230.
- 54 J. P. Kutney, G. K. Eigendorf, H. Matsue, A. Murai, K. Tanaka, W. L. Sung, K. Wada and B. R. Worth, *J. Am. Chem. Soc.*, 1978, **100**, 938–943.
- 55 J. Vercauteren, C. Lavaud, J. Levy and G. Massiot, *J. Org. Chem.*, 1984, **49**, 2278–2279.
- 56 P. D. Bailey, S. P. Hollinshead and Z. Dauter, *J. Chem. Soc., Chem. Commun.*, 1985, 1507–1509.
- 57 P. D. Bailey and S. P. Hollinshead, *Tetrahedron Lett.*, 1987, **28**, 2879–2882.
- 58 P. D. Bailey, S. P. Hollinshead, M. H. Moore, K. M. Morgan, D. I. Smith and J. M. Vernon, *Tetrahedron Lett.*, 1994, **35**, 3585–3586.
- 59 X. Fu and J. M. Cook, *J. Org. Chem.*, 1993, **58**, 661–672.
- 60 J. Seayad, A. M. Seayad and B. List, *J. Am. Chem. Soc.*, 2006, **128**, 1086–1087.
- 61 T. Arai, M. Wasai and N. Yokoyama, *J. Org. Chem.*, 2011, **76**, 2909–2912.
- 62 T. Arai and N. Yokoyama, *Angew. Chem., Int. Ed.*, 2008, **47**, 4989–4992.
- 63 J. O. Osby and B. Ganem, *Tetrahedron Lett.*, 1985, **26**, 6413–6416.
- 64 S. Handa, V. Gnanadesikan, S. Matsunaga and M. Shibasaki, *J. Am. Chem. Soc.*, 2007, **129**, 4900–4901.
- 65 W. Oppolzer, O. Tamura, G. Sundarababu and M. Signer, *J. Am. Chem. Soc.*, 1992, **114**, 5900–5902.
- 66 V. Vavsari, V. Dianati, S. Ramezanzpour and S. Balalaie, *Synlett*, 2015, **26**, 1955–1960.
- 67 *Compendium of Chiral Auxiliary Applications*, ed. G. Roos, Academic Press, New York, 2002.
- 68 Y. Gnass and F. Glorius, *Synthesis*, 2006, **2006**, 1899–1930.
- 69 D. A. Evans, J. Bartroli and T. L. Shih, *J. Am. Chem. Soc.*, 1981, **103**, 2127–2129.
- 70 W. Oppolzer, C. Chapuis and G. Bernardinelli, *Helv. Chim. Acta*, 1984, **67**, 1397–1401.
- 71 *Handbook of Reagents for Organic Synthesis: Chiral Reagents for Asymmetric Synthesis*, ed. L. A. Paquette, Wiley, Chichester, 2003.
- 72 N. Qais, N. Nakao, K. Hashigaki, Y. Takeuchi and M. Yamato, *Chem. Pharm. Bull.*, 1991, **39**, 3338–3340.
- 73 H. Akimoto, K. Okamura, M. Yui, T. Shioiri, M. Kuramoto, Y. Kikugawa and S. Yamada, *Chem. Pharm. Bull.*, 1974, **22**, 2614–2623.
- 74 K. Yamada, M. Takeda and T. Iwakuma, *J. Chem. Soc., Perkin Trans. 1*, 1983, 265–270.
- 75 T. Itoh, M. Miyazaki, S. Ikeda, K. Nagata, M. Yokoya, Y. Matsuya, Y. Enomoto and A. Ohsawa, *Tetrahedron*, 2003, **59**, 3527–3536.



- 76 E. W. Colvin, *Silicon Reagents in Organic Synthesis*, Academic Press, San Diego, 1988.
- 77 L. F. Tietze, Y. Zhou and E. Töpken, *Eur. J. Org. Chem.*, 2000, **2000**, 2247–2252.
- 78 A. Siwicka, K. Wojtasiewicz, A. Leniewski, J. K. Maurin, A. Zawadzka and Z. Czarnocki, *Can. J. Chem.*, 2007, **85**, 1033–1036.
- 79 F. M. Cordero, F. Pisaneschi, K. Meschini Batista, S. Valenza, F. Machetti and A. Brandi, *J. Org. Chem.*, 2005, **70**, 856–867.
- 80 J. Biała, Z. Czarnocki and J. K. Maurin, *Tetrahedron: Asymmetry*, 2002, **13**, 1021–1023.
- 81 N. S. S. Reddy, R. A. Babu and B. V. S. Reddy, *Synthesis*, 2016, **48**, 1079–1086.
- 82 J. A. Ellman, T. D. Owens and T. P. Tang, *Acc. Chem. Res.*, 2002, **35**, 984–995.
- 83 X.-W. Sun, M.-H. Xu and G.-Q. Lin, *Org. Lett.*, 2006, **8**, 4979–4982.
- 84 G.-Q. Lin, M.-H. Xu, Y.-W. Zhong and X.-W. Sun, *Acc. Chem. Res.*, 2008, **41**, 831–840.
- 85 F. Ferreira, C. Botuha, F. Chemla and A. Pérez-Luna, *Chem. Soc. Rev.*, 2009, **38**, 1162–1186.
- 86 F. A. Davis, J. Y. Melamed and S. S. Sharik, *J. Org. Chem.*, 2006, **71**, 8761–8766.
- 87 D. A. Cogan, G. Liu and J. Ellman, *Tetrahedron*, 1999, **55**, 8883–8904.
- 88 J. L. García Ruano, J. Alemán and M. B. Cid, *Synthesis*, 2006, **2006**, 687–691.
- 89 J. Fujiwara, Y. Fukutani, H. Sano, K. Maruoka and H. Yamamoto, *J. Am. Chem. Soc.*, 1983, **105**, 7177–7179.
- 90 K. Pulka, *Curr. Opin. Drug Discovery Dev.*, 2010, **13**, 669–684.
- 91 G. Brieger and T. J. Nestrick, *Chem. Rev.*, 1974, **74**, 567–580.
- 92 R. A. W. Johnstone, A. H. Wilby and I. D. Entwistle, *Chem. Rev.*, 1985, **85**, 129–170.
- 93 G. Zassinovich, G. Mestroni and S. Gladiali, *Chem. Rev.*, 1992, **92**, 1051–1069.
- 94 R. Noyori and S. Hashiguchi, *Acc. Chem. Res.*, 1997, **30**, 97–102.
- 95 M. J. Palmer and M. Wills, *Tetrahedron: Asymmetry*, 1999, **10**, 2045–2061.
- 96 K. Everaere, A. Mortreux and J. Carpentier, *Adv. Synth. Catal.*, 2003, **345**, 67–77.
- 97 S. Gladiali and E. Alberico, *Chem. Soc. Rev.*, 2006, **35**, 226–236.
- 98 T. Ikariya and A. J. Blacker, *Acc. Chem. Res.*, 2007, **40**, 1300–1308.
- 99 C. Wang, X. Wu and J. Xiao, *Chem.-Asian J.*, 2008, **3**, 1750–1770.
- 100 X. Wu and J. Xiao, *Chem. Commun.*, 2007, 2449–2466.
- 101 S. E. Clapham, A. Hadzovic and R. H. Morris, *Coord. Chem. Rev.*, 2004, **248**, 2201–2237.
- 102 M. Bianchi, U. Matteol, G. Menchi, P. Frediani, U. Pratesi, F. Piacenti and C. Botteghi, *J. Organomet. Chem.*, 1980, **198**, 73–80.
- 103 U. Matteoli, P. Frediani, M. Bianchi, C. Botteghi and S. Gladiali, *J. Mol. Catal.*, 1981, **12**, 265–319.
- 104 J. S. M. Samec, J.-E. Backvall, P. G. Andersson and P. Brandt, *Chem. Soc. Rev.*, 2006, **35**, 237–248.
- 105 T. R. Ward, *Acc. Chem. Res.*, 2011, **44**, 47–57.
- 106 M. Bartok, *Chem. Rev.*, 2010, **110**, 1663–1705.
- 107 H.-U. Blaser, C. Malan, B. Pugin, F. Spindler, H. Steiner and M. Studer, *Adv. Synth. Catal.*, 2003, **345**, 103–151.
- 108 P. Roszkowski, K. Wojtasiewicz, A. Leniewski, J. K. Maurin, T. Lis and Z. Czarnocki, *J. Mol. Catal. A: Chem.*, 2005, **232**, 143–149.
- 109 Z. Czarnocki, M. P. Matuszewska and I. Matuszewska, *Org. Prep. Proced. Int.*, 1998, **30**, 699–702.
- 110 J. Szawkało, S. J. Czarnocki, A. Zawadzka, K. Wojtasiewicz, A. Leniewski, J. K. Maurin, Z. Czarnocki and J. Drabowicz, *Tetrahedron: Asymmetry*, 2007, **18**, 406–413.
- 111 S. McLean, G. I. Dmitrienko and A. Szokolcai, *Can. J. Chem.*, 1976, **54**, 1262–1277.
- 112 M. Nakagawa, M. Kiuchi, M. Obi, M. Tonoizuka, K. Kobayashi, T. Hino and Y. Ban, *Chem. Pharm. Bull.*, 1975, **23**, 304–312.
- 113 S. J. Czarnocki, K. Wojtasiewicz, A. P. Jóźwiak, J. K. Maurin, Z. Czarnocki and J. Drabowicz, *Tetrahedron*, 2008, **64**, 3176–3182.
- 114 L. K. Vinograd and N. N. Suvorov, *Chem. Heterocycl. Compd.*, 1984, **20**, 984–988.
- 115 L. M. M. Cesar, C. F. Tormena, M. R. Marques, G. V. J. Silva, M. A. Mendes, R. Rittner and M. S. Palma, *Helv. Chim. Acta*, 2005, **88**, 796–801.
- 116 Y. Takahashi, H. Ishiyama, T. Kubota and J. Kobayashi, *Bioorg. Med. Chem. Lett.*, 2010, **20**, 4100–4103.
- 117 T. Ito, M. Kitajima and H. Takayama, *Tetrahedron Lett.*, 2009, **50**, 4506–4508.
- 118 E. Biron, J. Chatterjee, O. Ovadia, D. Langenegger, J. Brueggen, D. Hoyer, H. A. Schmid, R. Jelinek, C. Gilon, A. Hoffman and H. Kessler, *Angew. Chem., Int. Ed.*, 2008, **47**, 2595–2599.
- 119 A. Bischler and F. J. Howell, *Ber. Dtsch. Chem. Ges.*, 1893, **26**, 1384–1399.
- 120 Q. Yin, S.-G. Wang and S.-L. You, *Org. Lett.*, 2013, **15**, 2688–2691.
- 121 J. Selvakumar, A. Makriyannis and C. R. Ramanathan, *Org. Biomol. Chem.*, 2010, **8**, 4056–4058.
- 122 J. Selvakumar and C. R. Ramanathan, *Org. Biomol. Chem.*, 2011, **9**, 7643–7646.
- 123 F. Xu, D. Huang, C. Han, W. Shen, X. Lin and Y. Wang, *J. Org. Chem.*, 2010, **75**, 8677–8680.
- 124 I. Čorić, S. Müller and B. List, *J. Am. Chem. Soc.*, 2010, **132**, 17370–17373.
- 125 M. P. Lalonde, M. A. McGowan, N. S. Rajapaksa and E. N. Jacobsen, *J. Am. Chem. Soc.*, 2013, **135**, 1891–1894.
- 126 T. Itoh, M. Yokoya, K. Miyauchi, K. Nagata and A. Ohsawa, *Org. Lett.*, 2006, **8**, 1533–1535.
- 127 K. Nagata, H. Ishikawa, A. Tanaka, M. Miyazaki, T. Kanemitsu and T. Itoh, *Heterocycles*, 2010, **81**, 1791–1798.
- 128 L. D. Vazquez-Serrano, B. T. Owens and J. M. Buriak, *Inorg. Chim. Acta*, 2006, **359**, 2786–2797.



- 129 B. Wüstenberg and A. Pfaltz, *Adv. Synth. Catal.*, 2008, **350**, 174–178.
- 130 G. Stork, *Pure Appl. Chem.*, 1989, **61**, 439–442.
- 131 B. List, *Synlett*, 2001, **2001**, 1675–1686.
- 132 B. List, *Tetrahedron*, 2002, **58**, 5573–5590.
- 133 *The IUPAC Compendium of Chemical Terminology: The Gold Book*, ed. V. Gold, International Union of Pure and Applied Chemistry (IUPAC), Research Triangle Park, NC, 4th edn, 2019.
- 134 W. Carruthers, *Cycloaddition Reactions in Organic Synthesis*, Pergamon Press, Oxford, England, New York, 1st edn, 1990.
- 135 *Cycloaddition Reactions in Organic Synthesis*, ed. S. Kobayashi and K. A. Jørgensen, Wiley, 1st edn, 2001.
- 136 G. S. Buchanan, J. B. Feltenberger and R. P. Hsung, *Curr. Org. Synth.*, 2010, **7**, 363–401.
- 137 A. Moyano and R. Rios, *Chem. Rev.*, 2011, **111**, 4703–4832.
- 138 J. Adrio and J. C. Carretero, *Chem. Commun.*, 2014, **50**, 12434–12446.
- 139 J. Deng, X. Wang and R. P. Hsung, in *Methods and Applications of Cycloaddition Reactions in Organic Syntheses*, ed. N. Nishiwaki, Wiley, 1st edn, 2014, pp. 283–354.
- 140 R. Narayan, M. Potowski, Z.-J. Jia, A. P. Antonchick and H. Waldmann, *Acc. Chem. Res.*, 2014, **47**, 1296–1310.
- 141 A. W. Rey, W. A. Szarek and D. B. MacLean, *Can. J. Chem.*, 1992, **70**, 2922–2928.
- 142 H. Waldmann, M. Braun, M. Weymann and M. Gewehr, *Tetrahedron*, 1993, **49**, 397–416.
- 143 T. Kametani, Y. Hirai, M. Kajiwara, T. Takahashi and K. Fukumoto, *Chem. Pharm. Bull.*, 1975, **23**, 2634–2642.
- 144 C. Szántay, G. Blaskó, K. Honty, L. Szabó and L. Töke, *Heterocycles*, 1977, **7**, 155–160.
- 145 D. J. Faulkner and M. R. Petersen, *J. Am. Chem. Soc.*, 1973, **95**, 553–563.
- 146 T. Li, S. Liu, S. Wu, Q. Cheng, Q. Chen, Y. Jiao, Y. Zhang and F. Shi, *Sci. China: Chem.*, 2024, **67**, 2629–2636.
- 147 E.-L. Teo, G.-K. Chuah, A. R. J. Huguet, S. Jaenicke, G. Pande and Y. Zhu, *Catal. Today*, 2004, **97**, 263–270.
- 148 K. Koch, R. Vandenberg, P. Nieuwland, R. Wijtmans, M. Wubbolts, H. Schoemaker, F. Rutjes and J. Vanhest, *Chem. Eng. J.*, 2008, **135**, S89–S92.
- 149 C. Mateo, J. M. Palomo, G. Fernandez-Lorente, J. M. Guisan and R. Fernandez-Lafuente, *Enzyme Microb. Technol.*, 2007, **40**, 1451–1463.
- 150 D. Muñoz Solano, P. Hoyos, M. J. Hernáiz, A. R. Alcántara and J. M. Sánchez-Montero, *Bioresour. Technol.*, 2012, **115**, 196–207.
- 151 J. Ogawa and S. Shimizu, *Trends Biotechnol.*, 1999, **17**, 13–21.
- 152 E. Quiroga, N. Priolo, D. Obregón, J. Marchese and S. Barberis, *Biochem. Eng. J.*, 2008, **39**, 115–120.
- 153 M. G. Moghaddam, F. B. H. Ahmad, M. Basri and M. B. Abdul Rahman, *J. Appl. Sci.*, 2010, **10**, 337–342.
- 154 M. G. Moghaddam, F. B. H. Ahmad, M. Basri and M. B. Abdul Rahman, *Electron. J. Biotechnol.*, 2010, **13**(3), DOI: [10.2225/vol13-issue3-fulltext-9](https://doi.org/10.2225/vol13-issue3-fulltext-9).
- 155 K. Mitsukura, M. Suzuki, K. Tada, T. Yoshida and T. Nagasawa, *Org. Biomol. Chem.*, 2010, **8**, 4533–4535.
- 156 K. Mitsukura, M. Suzuki, S. Shinoda, T. Kuramoto, T. Yoshida and T. Nagasawa, *Biosci., Biotechnol., Biochem.*, 2011, **75**, 1778–1782.
- 157 K. Mitsukura, T. Kuramoto, T. Yoshida, N. Kimoto, H. Yamamoto and T. Nagasawa, *Appl. Microbiol. Biotechnol.*, 2013, **97**, 8079–8086.
- 158 G. A. Aleku, H. Man, S. P. France, F. Leipold, S. Hussain, L. Toca-Gonzalez, R. Marchington, S. Hart, J. P. Turkenburg, G. Grogan and N. J. Turner, *ACS Catal.*, 2016, **6**, 3880–3889.
- 159 S. Velikogne, V. Resch, C. Dertnig, J. H. Schrittwieser and W. Kroutil, *ChemCatChem*, 2018, **10**, 3236–3246.
- 160 D. Wetzl, M. Berrera, N. Sandon, D. Fishlock, M. Ebeling, M. Müller, S. Hanlon, B. Wirz and H. Iding, *ChemBioChem*, 2015, **16**, 1749–1756.
- 161 H. Man, E. Wells, S. Hussain, F. Leipold, S. Hart, J. P. Turkenburg, N. J. Turner and G. Grogan, *ChemBioChem*, 2015, **16**, 1052–1059.
- 162 M. Rodríguez-Mata, A. Frank, E. Wells, F. Leipold, N. J. Turner, S. Hart, J. P. Turkenburg and G. Grogan, *ChemBioChem*, 2013, **14**, 1372–1379.
- 163 J. Stöckigt and M. H. Zenk, *J. Chem. Soc., Chem. Commun.*, 1977, 646–648.
- 164 D. Pressnitz, E. Fischereider, J. Pletz, C. Kofler, L. Hammerer, K. Hiebler, H. Lechner, N. Richter, E. Eger and W. Kroutil, *Angew. Chem.*, 2018, **130**, 10843–10847.
- 165 L. Yang, J. Li, Z. Xu, P. Yao, Q. Wu, D. Zhu and Y. Ma, *Org. Lett.*, 2022, **24**, 6531–6536.
- 166 G.-H. Hou, J.-H. Xie, P.-C. Yan and Q.-L. Zhou, *J. Am. Chem. Soc.*, 2009, **131**, 1366–1367.
- 167 M. Li, Y. Cui, Z. Xu, X. Chen, J. Feng, M. Wang, P. Yao, Q. Wu and D. Zhu, *Adv. Synth. Catal.*, 2022, **364**, 372–379.
- 168 J. R. Marshall, P. Yao, S. L. Montgomery, J. D. Finnigan, T. W. Thorpe, R. B. Palmer, J. Mangas-Sanchez, R. A. M. Duncan, R. S. Heath, K. M. Graham, D. J. Cook, S. J. Charnock and N. J. Turner, *Nat. Chem.*, 2021, **13**, 140–148.
- 169 P. Yao, J. R. Marshall, Z. Xu, J. Lim, S. J. Charnock, D. Zhu and N. J. Turner, *Angew. Chem., Int. Ed.*, 2021, **60**, 8717–8721.

