

REVIEW

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Recent insights of PROTAC developments in inflammation-mediated and autoimmune targets: a critical review

Mary Sravani Galla, Nitika Sharma,[†] Priyanka Mishra[†] and Nagula Shankaraiah *

According to the mounting evidence in the literature, pro-inflammatory mediators/targets activate multiple signalling pathways to trigger illnesses that are ultimately responsible for acute pain, chronic inflammatory diseases, and several auto-immune disorders. Conventional drugs have been ruled out since proteolysis-targeting chimeras (PROTACs) are poised to overcome the limitations of traditional therapies. These heterobifunctional molecules help to degrade the targeted proteins of interest through ubiquitination. This review encompasses current and future aspects of PROTACs in inflammation-mediated and autoimmune targets. Different key points are highlighted and discussed, such as why PROTACs are preferred in this disease area, drawbacks and lessons learnt from the past, the role of linkers in establishing crucial degradation, *in vitro* findings, pharmacokinetics, *in silico* parameters, limitations of PROTACs in clinical settings, and future outcomes.

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1. Introduction

The dynamic interplay of the synthesis and degradation of the cellular components in living organisms serves as a cornerstone in maintaining homeostasis.¹ The strategy of leveraging the degradation mechanism has surfaced a novel paradigm for treating multiple disease conditions, representing a substantial milestone in modern drug discovery. In inhibitor-based strategies, there is always a setback when it comes to undruggable proteins, their accessible binding sites, or the specific quantifiable functions of these target proteins.² Driving these anomalies, intracellular targeted protein degradation (TPD) encompasses autophagy-lysosome systems [ALS], ubiquitin–proteasome systems [UPS], molecular glues and chaperone-mediated autophagy, with all of them adhering to the disposable mechanism.³ The advent of these bivalent degradations increases the scope for rendering the undruggable events.⁴ TPD could be the next frontier in treating inflammatory and autoimmune diseases.⁵ Unfortunately, drugs such as corticosteroids are still being used in the treatment of many inflammatory conditions for which they are not specifically intended. This is one of the main reasons for the downfall of treatment options in this area. For example, TNF inhibitors

were often used to treat various diseases such as psoriasis, IBD, and arthritis.⁶ However, they failed as a strategy upon identification of the distinct mechanism of these diseases. Consequently, the small molecule inhibitors or biologics, which are targeted in specific areas, are also limited by low organ penetration, attributed to high molecular weights, complicated routes of administration, toxicity, and off-target effects (Fig. 1).^{7,8}

According to the WHO assessment, “chronic inflammatory illness is considered the most significant health hazard to the human population”. To date, many diseases or disorders have no cure and only management is being perpetuated. Emphasizing this, we have identified a list of potential targets, with their diseases highlighted in Table 1. It has been indisputably acknowledged that treating immune-related disorders requires multifaceted approaches since their mechanisms are quite complex and clinically heterogeneous.⁹ In the pathophysiology of various immune checkpoints or endosomal receptors, when activated by pathogens or external/internal stimuli, they undergo an interconnected downstream pathway to awaken the pro-inflammatory genes and cytokines responsible for triggering the disease. Some of the well-known targets and receptors responsible for inflammatory signalling pathways are mentioned in Fig. 2.^{10–12}

PROTACs (proteolysis targeting chimeras) represent a revolutionary breakthrough advancement in TPD. They are also called heterobifunctional molecules meant to hijack the ubiquitin–proteasome system, thus overcoming all the glitches in traditional chemotherapy in terms of selectivity,

Department of Medicinal Chemistry, National Institute of Pharmaceutical Education and Research (NIPER), Hyderabad, 500037, India.
E-mail: shankar@niperhyd.ac.in, shankarnbs@gmail.com

[†] Contributed equally.

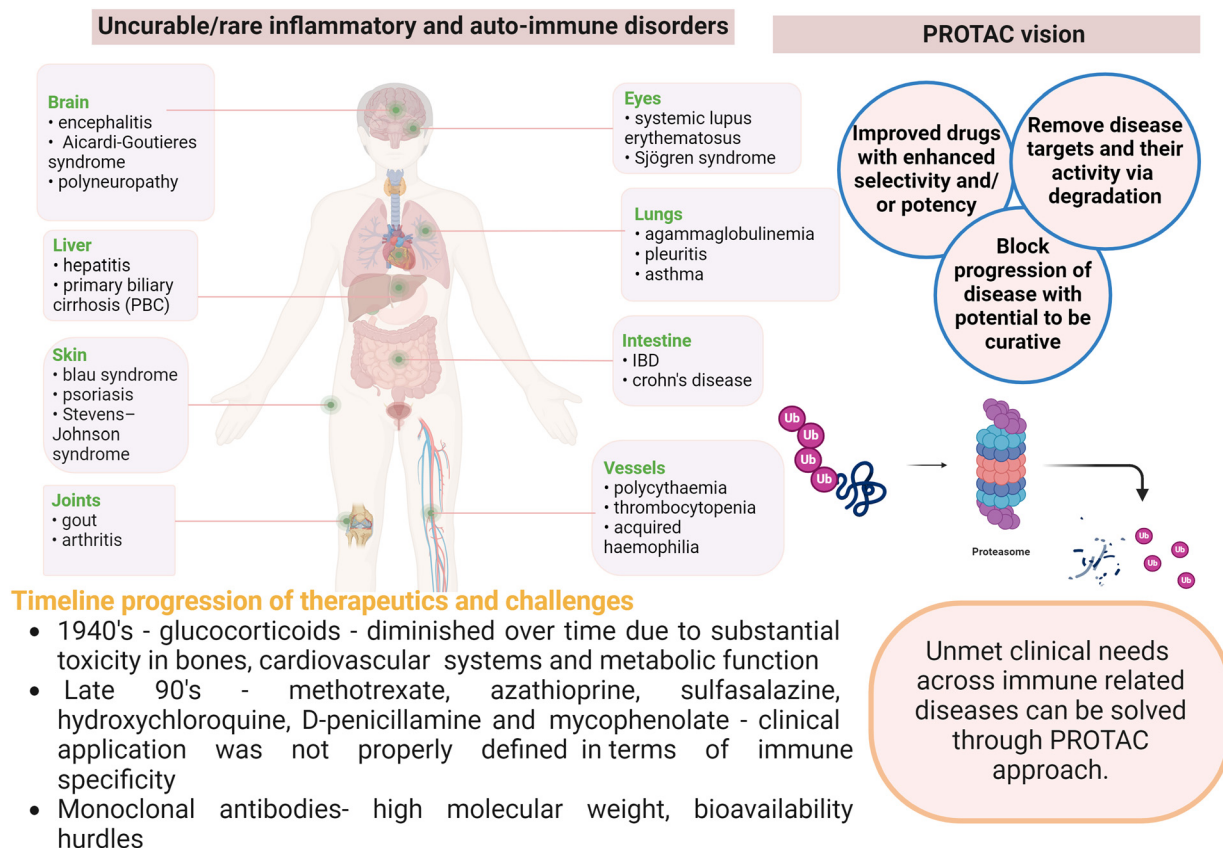


Fig. 1 Illustration of inflammatory/autoimmune disorders that need to be addressed by targeting through PROTACs.

Table 1 A list of important diseases along with their targets in inflammation/autoimmune-mediated diseases

S. no	Disease	Target	Disadvantages of conventional therapy
1.	Asthma and COPD	HDAC-VIII	Toxicities are related to gastrointestinal or constitutional symptoms and haematological abnormalities
2	Neuroinflammation	HDAC-VI	Inhibitors failed to achieve acute and reversible changes in genes
3	Diabetes-induced inflammation	HDAC-III	Limited isoenzyme selectivity, underprivileged pharmacokinetics properties
4	Asthma, rheumatoid arthritis, hidradenitis suppurativa	IRAK3/4	Traditional small-molecule inhibitors are unable to block kinase activity and scaffold signalling of IRAK
5	Rheumatoid arthritis, Alzheimer's neuroinflammation	JAK/STAT	Lipid profile disturbances, thromboembolic complications
6	Asthma, atopic dermatitis	IKT	Poor pharmacokinetics, insufficient target inhibition
7	Crohn's disease, IBD, multiple sclerosis	RIPK2	Toxicity and safety issues during clinical trials; intolerance to therapy over time
8	IBD, autoimmune uveitis	BRD4	Short half-life, off-target effects, development of resistance
9	Systemic lupus erythematosus, Aicardi-Goutières, psoriasis	STING	Poor solubility profile, limited selectivity
10	Inflammatory bowel syndrome, hepatitis	MIF	Low water solubility, pulmonary interstitial fibrosis
11	Asthma, arthritis	MAFF	Limited oral bioavailability
12	Acute lung injury, allergy	H-PGDS	Differences in pharmacokinetics and pharmacodynamics between preclinical animals and humans
13	Alzheimer's and Parkinson's diseases	GSK-3 β	Small therapeutic window, low selectivity
14	Neurodegenerative diseases	Keap 1	Off-target effects unpredicted reactions
15	Neuroinflammation, Alzheimer's disease	SHP 2	Higher dose
	Insulin-induced endothelial inflammation		Drug resistance, less efficacy
	COPD, asthma		

catalytic nature,¹³ and potency.^{14,15} Currently, the creation of tailored protein degradation techniques for treating multiple malignancies has been gaining much attention.^{16,17} This golden gateway has diminished nearly

all the major drawbacks of drug discovery using autophagy as a pivotal tool, *i.e.*, a slight modification of the body's destruction machinery, causing the target approach and thus facilitating its degradation through

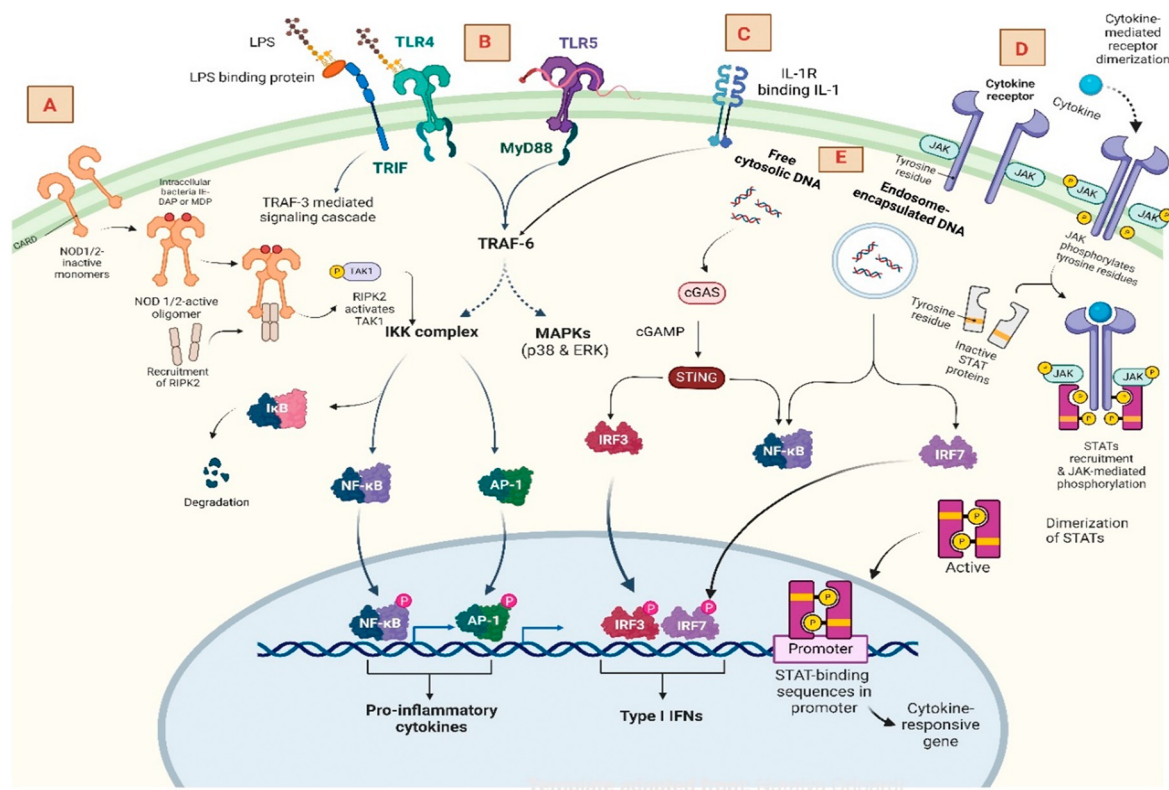


Fig. 2 The inflammatory signalling pathway and key components: A) NOD1 and NOD2 receptors involving RIPK1&2 to activate NF- κ B for gene translation. B) Toll-like receptors activate the MyD88 complex, followed by IKK and MAPK induction to produce an inflammatory response. C) IL-1R signalling pathway to TRAF-6 and NF- κ B. D) The JAK-STAT pathway activates the tyrosine kinase receptor recruiting JAK-mediated pathway to produce cytokine-responsive genes. E) Cytosolic DNA pathway in activating cGAS STING generating interferon regulating factor.

ubiquitination. Perhaps, implying this technique in inflammatory-mediated diseases would herald a new era in drug innovation.

1.1 Timeline of PROTAC

The Crews and Deshaies group pioneered the groundbreaking idea of PROTAC in 2001.¹⁸ In 2008, Schneekloth *et al.* introduced the first small-molecule PROTAC.¹⁹ The timeline of two decades of PROTACs demonstrates how briskly the technology has progressed from academia to industry to clinical candidates.²⁰

1.2 Mechanism of action

Since the mechanism of PROTACs has been described in numerous reports, its pathway is elucidated in short as follows.²¹ The highly conserved ubiquitin-proteasome system (UPS) is meant for degrading misfolded, non-functional, and other undesirable proteins in the body. It also reported that it is crucially engaged in a diverse array of cellular processes, such as gene transcription,²² protein transport,²³ synaptic plasticity and memory functions.²⁴ The degradation pathway is initiated with ubiquitin-activating enzyme (E1) activating the free ubiquitin, concurrently transferring the ubiquitin to the conjugating enzyme E2.²⁵

The E3 ligase enzyme then catalyses the final step of ubiquitination, which allows the transfer of the ubiquitin molecule from E2 to the target protein *via* forming an isopeptide bond between the lysine residue of the targeted protein and glycine of ubiquitin. Subsequent ubiquitination leads the target protein to be polyubiquitinated.²⁶ Finally, the 26S proteasomal degradation pathway easily recognizes this polyubiquitinated protein for destruction. Ubiquitin molecules are utilised for the subsequent cycle of degradation in this overall process.²⁷

The E3 ligases/degrons are designated as crucial elements in PROTACs, which are responsible for the ubiquitination. Although more than 600 E3 ligases are known to exist in the human body, only a few are being utilized in the development of PROTACs. Von Hippel-Lindau (VHL) and cereblon (CRBN) are the most used E3 ligases. Inhibitors of apoptosis protein (IAP), Kelch-like ECH-associated protein (KEAP1) and mouse double minute 2 (MDM2) are also being explored.^{28,29} In this review, we briefly summarize the status of anti-inflammatory targets being explored as PROTACs and their key findings. In the discussion section, we focus on the crucial topics relevant to the growth needed for anti-inflammatory PROTACs from the perspective of achieving success in bringing the molecule from the bench to the clinic.

2. HDAC

Histone and non-histone proteins present in DNA *via* acetylation and deacetylation govern various functions at the cellular level, such as gene expression and cellular proliferation.³⁰ Blocking of HDAC can lead to cell death and anti-inflammatory activities.^{31,32} A non-selective HDAC inhibitor, trichostatin A (TSA), showed elevation in the GM-CSF (granulocyte-macrophage colony-stimulating factor) and interleukin-8 (ref. 33). Eventually, SAHA (suberoylanilide hydroxamic acid) and TSA, well-known potent HDAC inhibitors used in cancer treatment, were proven to reduce swelling, pannus formation and bone erosion in rats with induced arthritis.³⁴ Hendrix *et al.* reported HDAC VIII inhibition with reduced pro-inflammatory and anti-inflammatory mediators.^{35,36} Likewise, HDAC VI is accountable for the regulation of inflammation and is used in the treatment of rheumatoid arthritis for the improvement of synovial inflammation.³⁷ NLRP3 (NLR family pyrin domain-containing protein 3 inflammasome) is expressed predominantly in macrophages, and the mutation of this gene is associated with the pathophysiology of many

immune-related diseases such as rheumatoid arthritis, gout, diabetes mellitus, *etc.*³⁸

Dekker and co-workers detailed the PROTACs approach over HDAC inhibitors in controlling inflammatory diseases like asthma, COPD (chronic obstructive pulmonary disorder), *etc.* With *o*-amino anilide CI994 (tacedinaline, ligand for class-1 HDAC I, II and III) as a warhead and cereblon as the E3 ligase, a series of HD-TACs (HDAC-PROTACs) has been developed by varying linker lengths. Compound 1 (Fig. 3), having *o*-aminoanilide substituted with the fluorine atom, showed improved selectivity resulting in an IC₅₀ value of 1.1 μM ± SD against HDAC III degradation in RAW 264.7 cells, and a DC₅₀ value of 0.32 μM, using western blot analysis.³⁹

To address a variety of inflammatory conditions linked to the NLRP3 inflammasome, Cao *et al.* investigated the HDAC VI protein. They introduced the concept of 'click' chemistry to synthesize PROTAC molecules comprised of different linker lengths by joining the alkyne moiety at the 1-position of indirubin with pomalidomide. Compound 2 exhibited a DC₅₀ of 108.9 nM against K562 cell lines, along with a significant reduction in the release of IL-1β, as well as nigericin-induced caspase-1 processing (p20). *In vivo* studies of compound 2 revealed that it leads to a decrease in IL-1β levels and has no role in affecting tumor necrosis factor-α (TNF-α) levels, indicating the inhibition of the NLRP3 inflammasome pathway activation.⁴⁰

Yang *et al.* optimized a cell-based assay method to evaluate the binding affinity for modified CRBN ligands with thalidomide, pomalidomide and lenalidomide. Triazoles were incorporated as linkers, and among them, lenalidomide exhibited maximum binding affinity, proving its promising ability to be incorporated as an E3 ligase for PROTAC molecules. Further analysis of compound 3 resulted in the significant HDAC VI degradation of 1.9 nM, which is comparable to PROTACs synthesised based on nexttrastat-A.⁴¹

Chotitumnavee and co-workers performed *in silico* studies upon obtaining the X-ray crystal structure of HDAC VIII with NCC149 (*N*-hydroxy-3-[1-(phenylthio)methyl-1*H*-1,2,3-triazol-4-yl]benzamide) to determine a suitable docking position for the inhibition of HDAC VIII. Among the synthesized PROTACs, compound 4 showed dose-dependent HDAC VIII degradation with 11 carbon linker length and exhibited a DC₅₀ of 0.702 μM. *In vitro* HDAC assay studies revealed that the compound has an IC₅₀ of 0.372 μM against HDAC VIII, irrespective of other isoforms of HDACs.⁴²

3. IRAK3 and 4 (interleukin-1 receptor-associated kinase)

Interleukin-1, interleukin-18, and toll-like receptors (TLRs) are critical immune factors involved in the pathway of interleukin-1-associated kinases. IRAK4 protein is currently the subject of intensive research for treating immunological disorders as it is involved in rheumatoid arthritis and hidradenitis suppurativa (HS) or atopic dermatitis (AD).

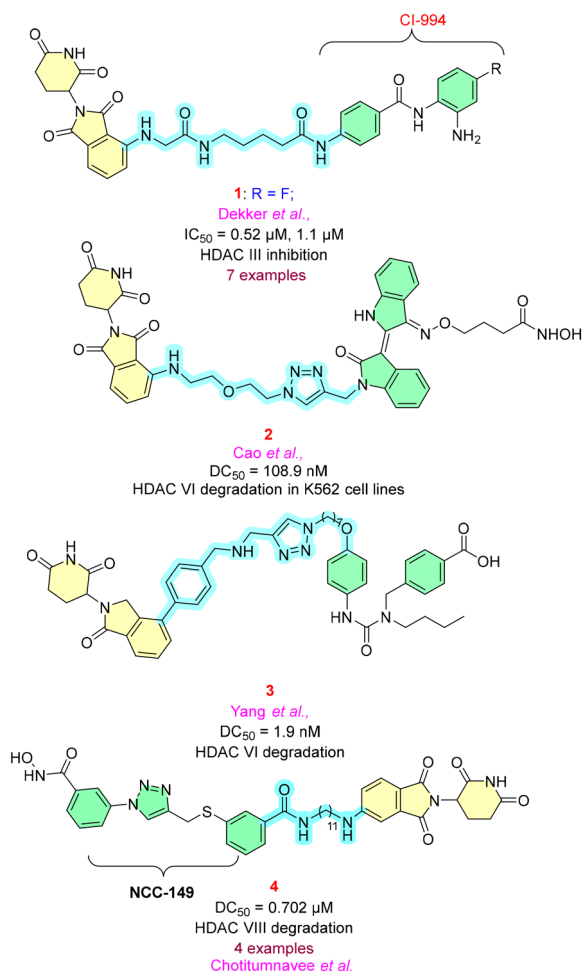


Fig. 3 HDAC-targeted PROTACs.

Presently one lead molecule, KT-474, has entered the phase II clinical trials targeting IRAK4 through TLR/IL-1R mediated pathways.⁴³

Due to the high degree of similarity between the ATP binding sites of IRAK1 and IRAK4, selective inhibitors of IRAK4 could be valuable for therapeutic strategy in inflammation-mediated diseases like systemic lupus erythematosus. Kargbo *et al.* reasoned that it is particularly challenging to study the catalytic domain of IRAK4's protein since it was highly conserved where the scope of exploration in IRAK4 was delayed. Since the PROTAC could overcome all these limitations, the team developed compounds **5** and **6** (Fig. 4) to provide potent degradation for IRAK4 resulting in IC₅₀ and DC₅₀ values below 30 nM.⁴⁴ Later, many groups focused on achieving a suitable linker for IRAK4 degradation, employing known inhibitors as warheads. Firstly, Nunes *et al.* employed isoquinoline warheads (IRAK4 inhibitor) and various E3 ligands, namely VHL (Von Hippel-Lindau), CRBN (cereblon) or IAP (inhibitor of apoptosis protein). Among them, compound **7**, possessing a fluoro-group, revealed a DC₅₀ of 259 nM through western blotting analysis. Further structural refinement of **7** into **8** exhibited a DC₅₀ value of 151 nM, which includes a spirocyclic pyrimidine linker, more rigid and polar than a flexible linear carbon chain.⁴⁵ Similarly, Chen and co-workers also investigated shorter

linker length PROTACs for IRAK4 observing no significant protein degradation, whereas compound **9** with a polyethylene glycol linker was effective in lowering IRAK4 levels.⁴⁶

Zhang *et al.* explained the inhibitory action of compound **10** by decreased IL-6 production; docking studies revealed that the molecule's cyclopropyl group in the solvent-exposed region is a convenient site for linker conjugation. By structurally altering the cyclopropyl group to a linker, a set of linkers was synthesized and evaluated, of which **10c** and **10d** showed DC₅₀ of 190 nM and 405 nM by immunoblotting.⁴⁷

We know that IRAK is a large group of families, involving IRAK1, IRAK2, and IRAK4 stimulating myeloid differentiation primary response 88 (MyD88)-dependent activation of nuclear factor kappa-light-chain enhancer of activated B cells (NF-κB) and thus pro-inflammatory cytokine production. Degorce *et al.* encountered serendipitous results in the discovery of potent IRAK3 inhibitors while studying the IRAK4 protein. The *cis*-isomer of pyrrolotriazine has been employed as an efficient POI ligand with VHL or CRBN as an E3 ligase. CRBN E3 ligases containing PEG linkers showed better degradation than VHL ligands. It was noted that the length of the linker is directly proportional to degradation in the case of CRBN. Compounds **11** and **12** showed selective degradation profiles

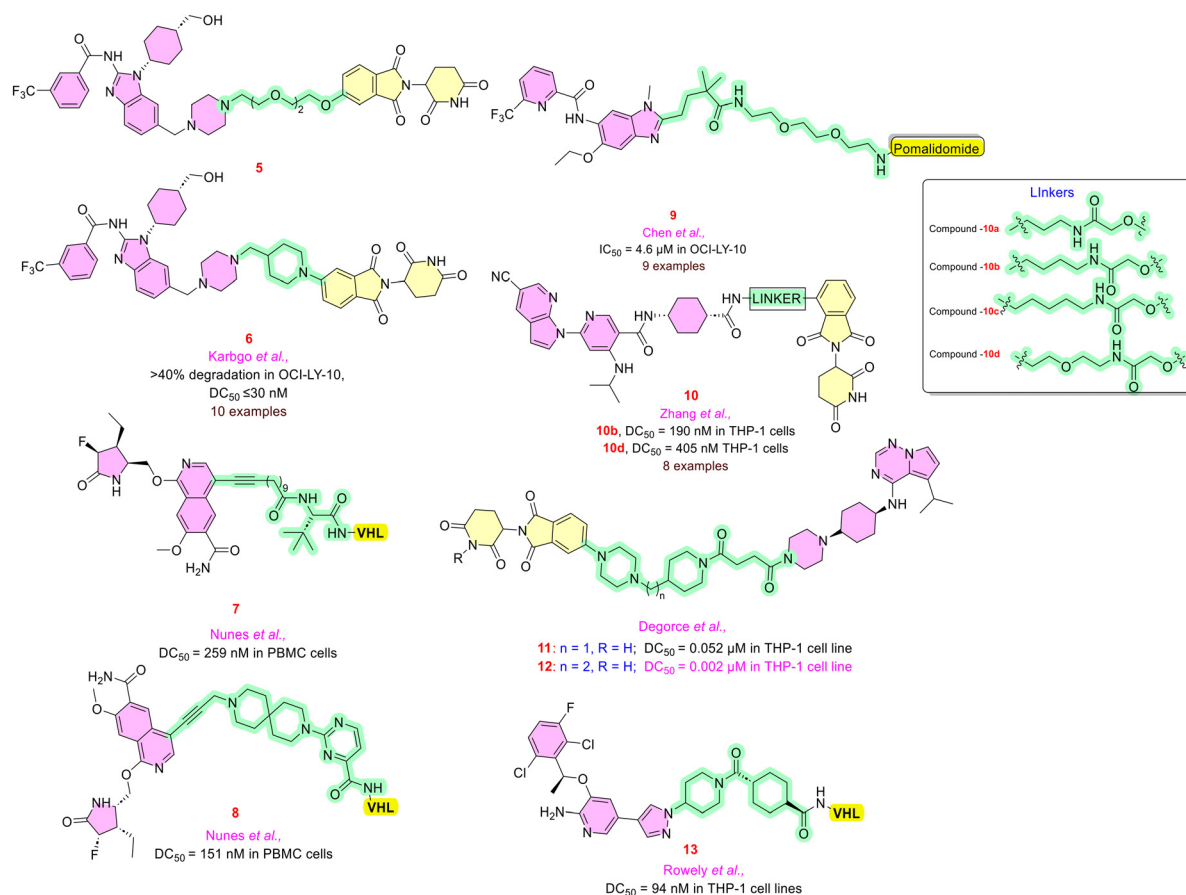


Fig. 4 IRAK3 and 4-targeted PROTACs.

with DC_{50} values of 0.052 μM and 0.002 μM for IRAK3 by 74% and 98% D_{max} , respectively.⁴⁸

Similar efforts were made by Rowley *et al.*, who studied the IRAK3 protein for controlling the dendritic cell-mediated production of interleukin-12 (IL-12). Crizotinib as the warhead, VHL as the E3 ligase ligand, and compound **13** with *trans*-cyclohexylamide linker resulted in the preferential degradation of IRAK3. Treatment of **13** in THP-1 cell lines resulted in a DC_{50} value of 94 nM and 95% D_{max} . Treatment of dendritic cells with **13** resulted in IRAK3 degradation both in the presence and absence of LPS, along with the significant elevation in IL-12p levels with LPS stimulation.⁴⁹

4. PCAF and GCN5

PCAF (p300/CBP-associated factor) and GCN5 (general control nonderepressible 5) are closely related epigenetic proteins that regulate immune functions in various cellular pathways. According to Zheng *et al.* and Zhuang *et al.*, PCAF and GCN5 are potential targets for treating autoimmune disorders as they cause a reduction in H3K18, H3K9 acetylation, accompanied by NF- κ B, IFN- β (interferon β), TBK1 (TANK binding kinase-1), which are overexpressed in injury, regulating inflammatory action, and modulating immune signaling.^{50–52}

Bassi *et al.* focused on the PCAF/GCN5 PROTACs and employed GSK4027, an active inhibitor of PCAF and GCN5, with CRBN as an E3 ligase for enhancing the activity. Compound **14** (Fig. 5) showed a DC_{50} value of 1.5 nM for PCAF degradation and 3 nM DC_{50} for GCN5 degradation in THP-1 cell lines, and 80% degradation of proteins was observed within 10 min at a concentration of 30 nM. The *cis* (*R,R*) enantiomer of the parent compound GSK4027 is more active than the other enantiomer.⁵³

5. JAK–STAT (Janus kinase-signal transducers and activators of transcription)

Another leading potential target in cytokine signaling is the JAK/STAT pathway involving interleukin-2 and other inflammatory factors, such as γ -interferons. Cytokines binding to JAK receptors initiate STAT transphosphorylation, leading to gene expression for activities like proliferation and differentiation,⁵⁴ which are responsible for numerous disorders, such as rheumatoid arthritis (RA), Alzheimer's disease (AD), inflammatory bowel disease (IBD), and multiple

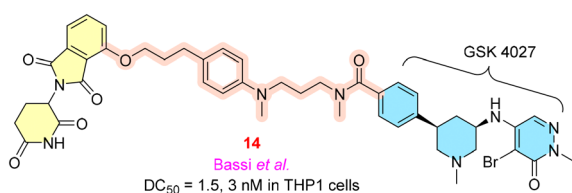


Fig. 5 PCAF-GCN5 targeted PROTACs.

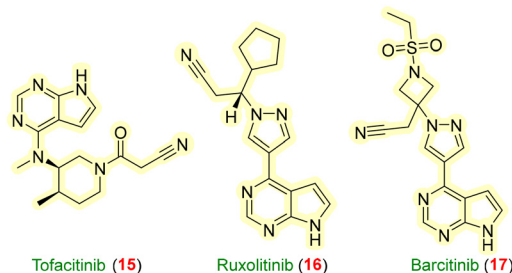


Fig. 6 Drugs used as JAK–STAT inhibitors.

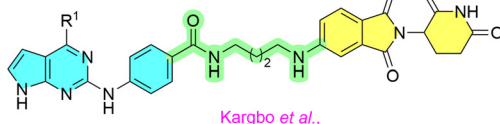
sclerosis (MS). JAK inhibitors such as tofacitinib **15**, ruxolitinib **16**, baricitinib **17**, (Fig. 6), *etc.*, provided benefits for the improvement of rheumatoid arthritis by altering T-cell activation and natural killer (NK) cell activity.⁵⁵ Although JAK/STAT targets were disclosed decades ago, there are still challenges such as the bulky dose of the drug with a short half-life, side effects like thromboembolism and cross-talk between the JAK/STAT pathway and non-canonical signal transductions.⁵⁶ PROTACs could be a simple answer to the limitations.

Shah *et al.* disclosed PROTACs with pyrimidine and quinoxaline cores as warheads. The primary aim was to develop PROTAC molecules with suitable physicochemical properties to permeate the cells. Among them, the quinoxaline motif showed better permeability than pyrimidine motifs and exhibited more selectivity towards JAK 2. The PBMC (peripheral blood mononuclear cells) assay has been performed to identify the trends regarding cell permeability. The molecules were tested against THP-1 cell lines bearing JAK 1 and 2 proteins, out of which six PROTACs (**21–26**, Fig. 7) were proven to show appropriate degradation.⁵⁷

In 2021, Kargbo filed a patent unravelling PROTAC-mediated degradation targeting Janus kinase (JAK). Pyrrolo-pyrimidines were evaluated for the Caco-2 cell permeability experiment, CRISPER-Cas9 editing, and other biological assays, which revealed that compound **28** has the lowest inhibitory concentration of $EC_{50} = 0.0002 \mu\text{M}$ for JAK protein degradation. A simple modification of the linker pattern resulted in a drastic increase in EC_{50} values. The team evaluated PROTACs **29** and **30** containing ruxolitinib and baricitinib (potent JAK inhibitors) as POI ligands with benzamide-containing linkers providing EC_{50} values of 0.00054 μM and 0.0039 μM , respectively.⁵⁸

6. ITK (interleukin (IL)-2-inducible T cell kinase)

Jiang *et al.* designed a potent, target-selective ITK (IL-2-inducible T-cell kinase) PROTAC **31** (Fig. 8) to help in degrading inflammatory responses with the inhibition of STAT3 and STAT5 signaling. Compound **31** showed >90% degradation of ITK selectively at a 1 μM concentration. To



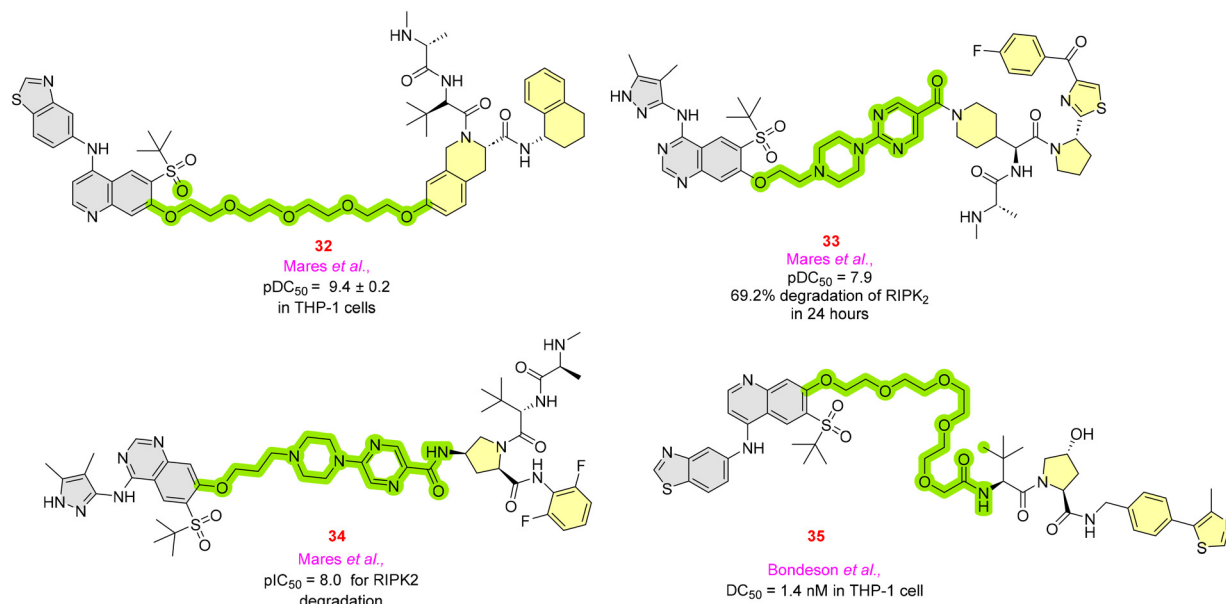


Fig. 9 RIPK2-targeted PROTACs.

8. BRD4 (bromodomain-containing proteins)

Bromodomain-containing proteins expressed by the BET (bromodomain and extra-terminal domain) family have been proven to significantly affect the inflammatory genes through NF-κB or Nrf2 (nuclear factor erythroid 2-related factor 2) pathways.⁶⁶ JQ1 and I-BET are two potent and selective BET inhibitors that have shown their capability in anti-inflammatory activities. As the literature suggests, JQ1 binds with greater affinity to BRD4 than other BET proteins and modulates histone acetylation function, which is responsible

for producing inflammatory genes.⁶⁷ In PROTACs for BRD4, the initial efforts of employing JQ1 as warheads, focusing on the optimal E3 ligases, have been discussed.

Tong *et al.* employed a terpene-based derivative such as bardoxolone, an E3 ligase recruiter, as an electrophilic moiety to establish reversible covalent bonds with the cysteine residue of the KEAP1 (Kelch-like ECH-associated protein) and JQ1 (BRD4 inhibitor), which was employed as a POI to explore the activity of bardoxolone as an E3 ligase. The paper highlights the critical need to explore diverse E3 ligases to withstand the emerging resistance of PROTACs. The descyano derivative **37** (Fig. 10) resulted in the loss of activity to degrade BRD4 in 231 MFP cells, indicating the significance of the α-cyanoenone moiety for the inhibitory action towards BRD4. In general, it has been found that any alteration to the bardoxolone methyl group would significantly impair both its anti-inflammatory efficacy and the activation of the Nrf2 pathway.⁶⁸

iNOS (inducible nitric oxide synthase), COX-2 (cyclooxygenase-2), IL-1β, TNF-α, IL-6, CCL2 and MMP-9 (matrix metalloproteinases) are pro-inflammatory factors that are modulated with BET proteins. A significant reduction in the expression of these proteins was observed with the degradation of BRD2 and BRD4. DeMars *et al.* studied dBET1 in the treatment of neuroinflammatory diseases. JQ1 was a warhead with CRBN E3 ligases. In SIM-A9 microglia cell lines, time-dependent degradation of BRD2 and BRD4 of **38** was observed at the concentration of 1 μM, blocking the production of pro-inflammatory genes.⁶⁹

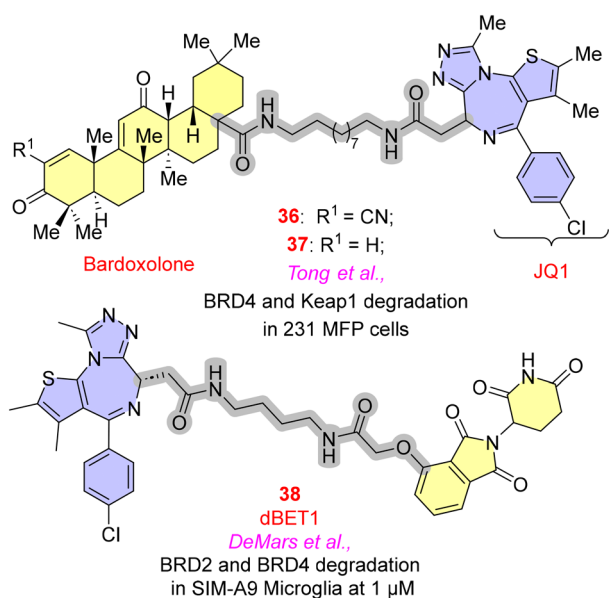


Fig. 10 BRD4-targeted PROTACs.

9. BTK (Bruton's tyrosine kinase)

Many pro-inflammatory factor modulations were correlated with Bruton's tyrosine kinase (BTK) to maintain immune

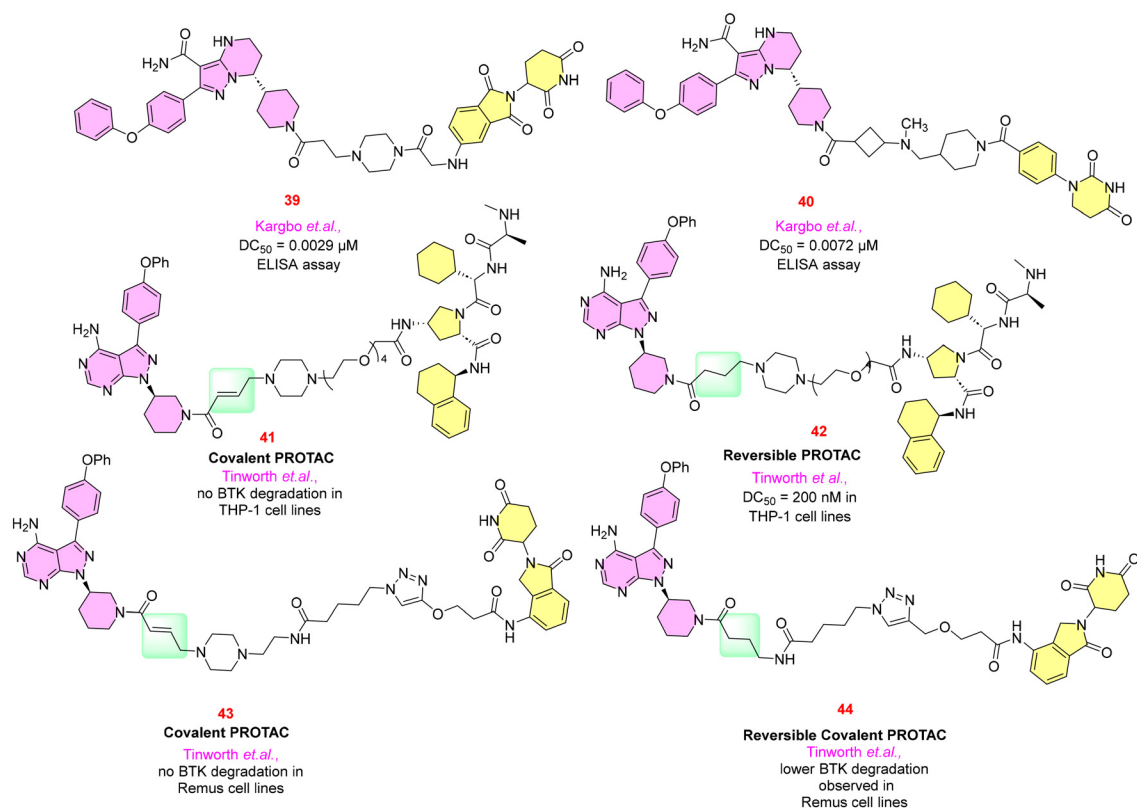
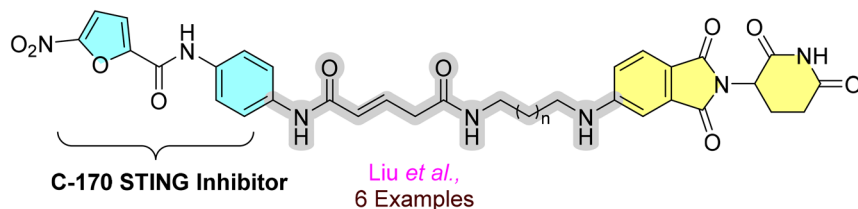


Fig. 11 BTK-targeted PROTACs.

haemostasis. X-linked agammaglobulinemia (XLA) is an immunodeficiency disorder caused by the suppression of BTK signalling. Targeting BTK protein would reduce various inflammatory factors such as IL-6, IL-1, TNF- α , type 1-IFN, *etc.* Robert B. Kargbo and co-workers designed and patented BTK degraders; compound **39** (Fig. 11), containing a piperazine linker with CRBN E3 ligase, and compound **40**, containing constrained butane and piperidine rings with

MDM2 E3 ligase, showed minimum DC₅₀ values of 0.0029 μM and 0.0072 μM, respectively, resulting in excellent activity.⁷⁰

Given the linker modifications, Tinworth *et al.* compared the degradation profile of covalent binding and reversible binding PROTAC molecules against the BTK target. Halo-PROTACs are a suitable example of covalent binding PROTACs with profound degradation. Compound **42** resulted



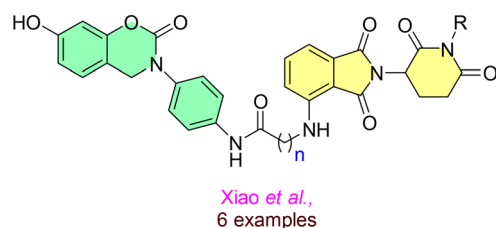
Compound	n	DC ₅₀ (μM)
45	4	10
46	5	9.2
47	6	3.2
48	2	no degradation
49	3	no degradation
50	1	no degradation

Fig. 12 STING-targeted PROTACs.

in concentration-dependent BTK degradation with a DC_{50} of 200 nM for recruiting the VHL E3 ligase. Covalent PROTACs **41**, **43** resulted in no BTK degradation. In contrast, **44** was slightly active towards the degradation. This suggests that non-covalent interactions primarily mediated the degradation as compared to the covalent binding PROTAC. The researchers proposed the reason for this as the blockade of the site by the covalently bound ligand for ubiquitin transfer or the inability of the proteasome to recognize the modified region required for degradation.⁷¹

10. STING (stimulator of interferon genes)

Stimulators of interferon genes (STING) present in the human body aid in the detection of bacterial nucleotides synthesized by the cyclic GMP-AMP synthase enzyme (cGAS) present in bacteria. Under normal circumstances, the human body has a cGAS-STING pathway to regulate innate immune responses. In abnormal conditions, this STING pathway activates pro-inflammatory mediators like cytokines and the release of IFNs to fight against bacterial infections. Liu and co-workers designed PROTACs to target the STING pathway to treat various autoimmune and inflammatory conditions like Aicardi-Goutieres syndrome and systemic lupus erythematosus. Compounds **45–50** (Fig. 12) were synthesized using C-170 as a warhead that interacts with the STING protein through its nitro and furan moieties. In contrast, the benzene ring plays a crucial role by providing a tethering site for the CRBN E3 ligase through a suitable linker. It was observed that the compounds exhibiting α , β -unsaturated carbonyl linkers are biologically active towards degrading the STING protein; compound **47** showed the most satisfactory results with a DC_{50} of 3.2 μ M with time-dependent degradation. *In vivo* studies by ELISA in the kidney tissues of mice showed that **47** could reduce the level of IFN- β , IL-6, and CXCL-10 in a dose-dependent manner.⁷²



Compound	n	R	Ki [nM]	Degradation (%) in 2 μ M
51	3	H	55 $\pm$ 3	not significant
52	4	H	86 $\pm$ 5	21 $\pm$ 7
53	6	H	51 $\pm$ 3	55 $\pm$ 4
54	7	H	71 $\pm$ 5	91 $\pm$ 5
55	9	H	65 $\pm$ 4	64 $\pm$ 2
56	7	CH ₃	55 $\pm$ 12	55 $\pm$ 12

Fig. 13 MIF-targeted PROTACs.

11. MIF (macrophage migration inhibitory factor)

Although MIF is known for its anti-cancer potential, it is also associated with immune responses. In 2021, Xiao *et al.* studied the effects of macrophage migration inhibitory factor (MIF) degradation by employing 7-hydroxycoumarin and 7-hydroxy-3,4-dihydrobenzoxazin-2-one cores as warheads, with cereblon as the E3 ligase ligand. Compound **54** (Fig. 13) was found to be the most potent, with 91% degradation at 2 μ M and 71% degradation against A549 cells. HEK293 cell lines had also been tested with **54**, which resulted in 90% degradation at 200 nM. The cellular uptake of **54** was also confirmed using intrinsic fluorescence microscopy studies.⁷³

12. IKZF (Ikaros zinc-finger transcriptional factors)

Ikaros zinc-finger transcriptional factors (encoded by gene IKZF1) and Aiolos (encoded by gene IKZF3) are critical for immunomodulatory activity and their polymorphism can lead to systemic lupus erythematosus. Two degronimers, **57** and **58** (Fig. 14), have been developed to target the E3 ligase cereblon, which may cause the potential degradation of Aiolos or Ikaros through the ubiquitin–proteasome pathway. The investigation of these compounds for binding to the CRBN-DDB1 (DNA damage binding protein-1) complex using fluorescence polarization assay resulted in values less than 10 μ M K_d .⁷⁴

13. MAFF (v-maf musculoaponeurotic fibrosarcoma oncogene family)

To date, there have been many reports on the anti-inflammatory activity of lathyrane diterpenoids. Wu *et al.* studied PROTACs as a potential strategy for identifying the targets for natural products. ZCY-001 was employed as the

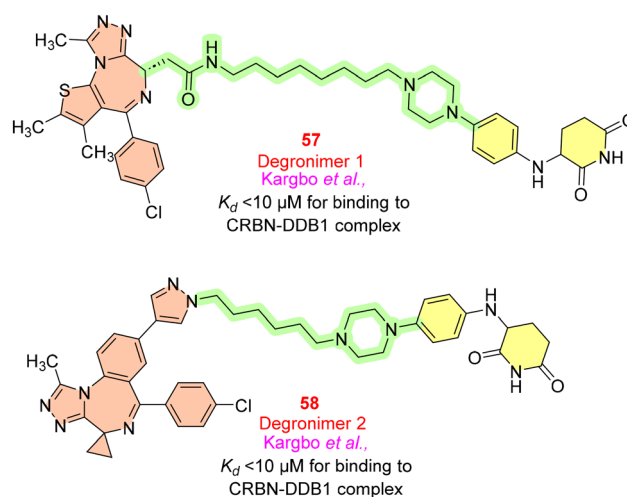


Fig. 14 IKZF-targeted PROTACs.

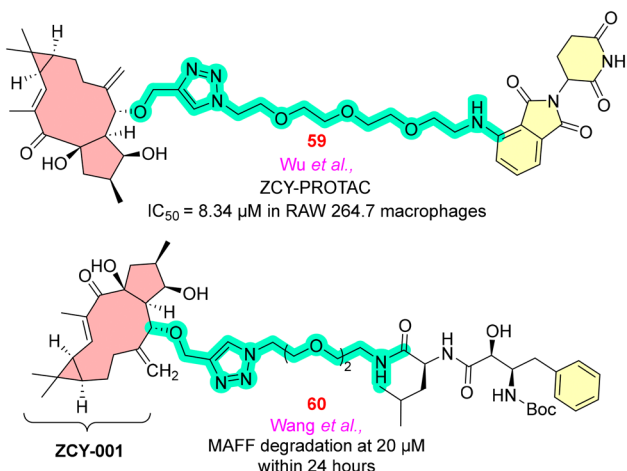


Fig. 15 MAFF-targeted PROTACs.

warhead for the development of ZCY-PROTAC **59** (Fig. 15) with thalidomide as the E3 ligase linked by a 3-PEG linker, which resulted in the profound inhibition of LPS-stimulated NO (nitric oxide) release. It also showed that MAFF degradation is responsible for inflammatory gene expression. However, individually, lathyrol and thalidomide treatment resulted in no degradation of MAFF, thus confirming that the degradation is induced through the ternary complex formation by PROTAC.⁷⁵

As an extension of this work, the same group designed 15 new PROTACs based on **59** to establish the anti-inflammatory action of lathyrol PROTACs. Different E3 ligase ligands (VHL, cIAP and MDM2) were employed with variations in the linker chains. Compound **60**, having a PEG linker chain and cIAP ubiquitin ligase ligand, showed maximum inhibitory activity with an IC_{50} value of $5.30 \pm 1.23 \mu M$ against LPS-induced NO production in RAW264.7 cells.⁷⁶

14. H-PGDS (hematopoietic prostaglandin D synthase)

For the treatment of allergic disorders, targeting H-PGDS (hematopoietic prostaglandin D synthase) would be

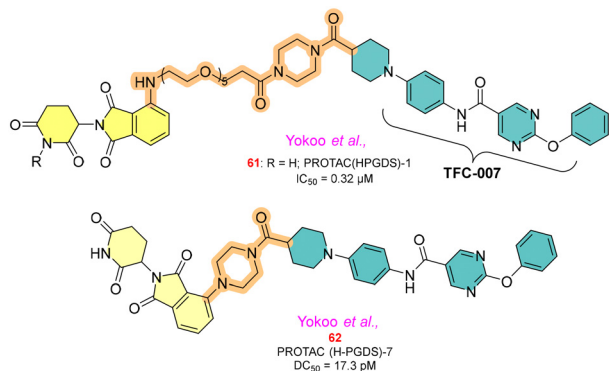


Fig. 16 H-PGDS-PROTACs molecules.

beneficial because some allergy disorders are linked to excessive prostaglandin D2 production. However, H-PGDS inhibitors have not shown adequate clinical efficacy. With the advancing research, Yokoo *et al.* approached PROTACs and SNIPERS (specific and nongenetic IAP-dependent protein erasers) as potential treatments (Fig. 16). TFC-007 (a potent inhibitor of H-PGDS with IC_{50} of 83 nM) was employed as a warhead and PROTAC (H-PGDS)-1 **61** was synthesized by employing pomalidomide as a E3 ligase. Significant reduction in H-PGDS was observed with maximum degradation between 100–1000 nM in Human KU812 and MEG-01 cell lines. For better outcomes in degradation and to improve pharmacokinetics, *in silico* simulations were conducted. A library of compounds was designed to dock and understand the binding pattern of the stable ternary complex of PROTAC (H-PGDS). A deeper analysis revealed that the length of the linker is inversely proportional to the stability of the ternary complex. Astonishingly, compound **62** without any linker showed maximum protein degradation with a DC_{50} value of 17.3 pM against KU812 cell lines. The co-crystal structure of the H-PGDS protein indicates that the *N*-carbonyl piperazine–piperidine moiety of TFC-007 serves as a linker rather than a warhead because it does not bind to the protein. This probably reduced its molecular weight and number of rotatable bonds and rendered compound **62** more druggable with the rule of five chemical spaces.⁷⁷

15. IDO1 (indoleamine 2,3-dioxygenase 1)

IDO1 has been identified as a possible candidate for cancer immunotherapy due to its substantial role in the tumor immune escape mechanism through T-cell regulation. Considering this, Hu *et al.* developed a PROTAC molecule based on Epa (highly selective IDO1 inhibitor) that was employed as a warhead with E3 ligases like lenalidomide and pomalidomide. Compound **63** (Fig. 17), containing seven PEG units, resulted in the degradation of IDO1 with a DC_{50} value and D_{max} of $2.84 \mu M L^{-1}$ and 93%, respectively.⁷⁸

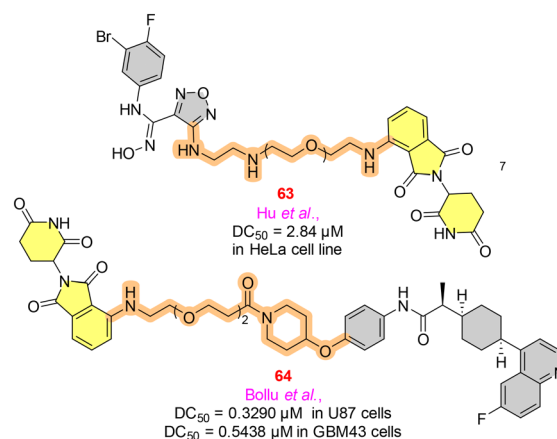


Fig. 17 IDO1-PROTACs molecules.

Later, Bollu *et al.* reported a PROTAC molecule-degrading IDO1 protein. A library of PROTACs was synthesized based on CRBN and VHL-based E3 ligase ligands and with the employment of linrodostat. CRBN-based PROTAC molecules with PEG linkers (<10 carbon atoms) showed a favourable pharmacokinetic profile. Compound **64** with a *para*-attachment to the linker was the most potent, with 94% degradation of IDO1 with a DC_{50} of 0.3290 μ M in U87 cell lines and 0.5438 μ M in GBM43 cell lines.⁷⁹

16. GSK-3 β (glycogen synthase kinase 3 β)

GSK-3 β is regarded as a critical target for treating several neurodegenerative disorders, most notably Alzheimer's disease. GSK-3 β demonstrates its pronounced pro-inflammatory effect, resulting in the loss of neurons. Jiang *et al.* conjugated the GSK-3 β inhibitor with E3 ligases with variable chain length linkers. Through click chemistry, compound **65** (Fig. 18) showed 65.8% degradation of the GSK-3 β at 10 μ M. To examine the inhibitory activity of compound **65** towards the TNF- α , neuroinflammation was induced in the microglia BV2 cells of the mouse. As anticipated, **65** reduced the TNF- α to its basal value, attenuating several inflammatory responses.⁸⁰

17. Keap 1 (Kelch-like ECH associated protein 1)

Lu *et al.* designed a peptide-based PROTAC that acts on Keap1 (Kelch-like ECH associated protein 1) that served as a substrate protein for the Cul3/Rbx1 (cullin-3/RING box protein-1)-dependent E3 ligase complex. The PROTAC facilitates the degradation of tau protein by binding with Keap1, which regulates the Nrf2 signalling pathway related to inflammatory diseases. Ac-LDPETGEYL-OH, a peptide-based PROTAC **66** (Fig. 19), was synthesized and it had a strong affinity for both Keap1 and Tau proteins, with K_d values of 22.8 nM and 763 nM, respectively. PROTAC **66** was crucial for tau degradation, as evidenced by CO-IP, flow cytometry, and western blotting assays in SH-SY5Y, Neuro-2a, and PC-12 cells. Targeting tau breakdown through Keap1 recruitment may be a better way to treat neurodegenerative diseases than suppressing the Nrf2 pathway.⁸¹

Keap1 protein is associated with Nrf2 pathway activation and mediates the oxidative response. Wang *et al.* designed

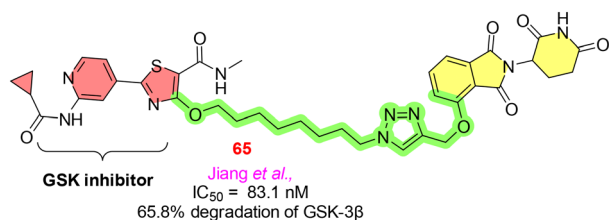


Fig. 18 GSK-3 β -PROTACs molecules.

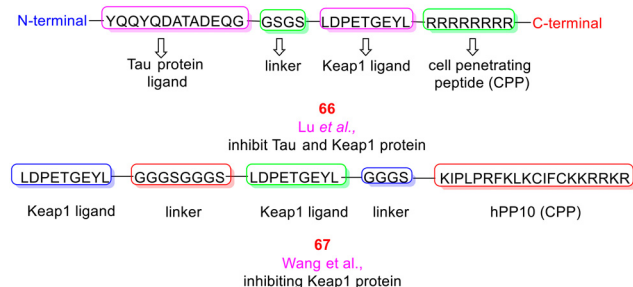


Fig. 19 Peptide PROTAC inhibiting Keap1 protein.

PROTAC **67**, which includes a chimeric Keap1-Keap1 peptide to recruit Keap1 target proteins and connect them to the E3 ligase. The peptide sequence was conjugated to the cell-penetrating peptide hPP10 through the C-terminal to obtain **67**, which resulted in 25% of Keap1 protein degradation when treated in HSC-T6 cells (hepatic stellate cells). The degradation of Keap1 protein was found to be associated with the activation of ARE (antioxidant response elements) through the Nrf2 pathway, ultimately leading to the downregulation of TNF- α , IL-1 β , and IL-6. The degradation was confirmed to be induced *via* the ubiquitin-proteasome pathway.⁸²

Ji *et al.* designed PROTACs based on the ARE (antioxidant response elements) sequence as the Nrf2 ligand. The double-stranded oligonucleotide sequence 5'-TCACAGTGACTCAGCA GAATC-3' from the NQO1 (NADPH quinone oxidoreductase 1) promoter was recruited as a warhead. Two series of ARE-PROTAC molecules were synthesized based on CRBN and VHL E3 ligands using a CuAAC (copper-catalyzed azide-alkyne cycloaddition) reaction for conjugation with the Nrf2-DNA ligand. CRBN-based ARE-PROTACs showed favourable depletion levels when tested in A549 cells. Among them, **68** (Fig. 20) is the most potent compound exhibiting a DC_{50} of 1.85 nM against Nrf2 protein and a DC_{50} value of 66 nM against MafG (musculoaponeurotic fibrosarcoma transcription factor G) in A549 cells. Antioxidant activity was also lowered, leading to oxidative damage to cancer cells.⁸³

18. SHP2 (Src homology 2 domain-containing phosphatase 2)

SHP2 is a protein tyrosine phosphatase encoded by the PTPN11 gene and was considered difficult to target because of its highly positively charged nature. In this regard, Wang *et al.*

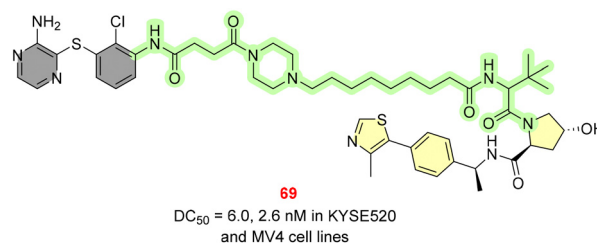


Fig. 20 Nrf2-targeted PROTACs.



A more intensive focused study on the anti-inflammatory targets will allow promising clinical candidates to be the next generation of therapeutics. Unlike small molecules, PROTACs offer unlimited benefits like binding, which is specifically a secondary concern for degradation, only relevant groups are mandatory for binding with the protein. This will reduce the molecular weight of the compounds and additional covalent interaction/irreversible bindings, leading to toxicity. These unnecessary groups can be modified as linkers to reduce the additional bulkiness for PROTACs. From the origin of PROTACs, it was majorly confined to anticancer drug development, while other targets received less attention in recognising their potential. In inflammatory/autoimmune diseases, the breach between the existing therapies like NSAIDs and the discovery rate of novel entities is very large. It was observed that the incidence rates of rare/uncurable disorders have been abruptly increasing irrespective of age, gender, species, *etc.* For example, international attention towards irritable bowel syndrome (IBS) is being carried forward as the condition depends on the individual genetic makeup enclosing minimal treatment options. Gratifyingly,

given the progress of BioPROTACs and hybrid PROTACs, the solutions to POI that lack the ligand binding site could be found. The footsteps of immuno-oncology will be an important modality for treating resistance-mediated cancers. All the concerns consolidate to highlight the immediate need for the development of novel PROTACs for future generations.

Author contributions

M. S. Galla: writing – review & editing – original draft. P. Mishra, N. Sharma: literature survey, writing N. Shankaraiah: conceptualization, investigation, writing – review & editing, supervision.

Conflicts of interest

There are no conflicts of interest to declare.

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References

- 1 Z. Yin, H. Popelka, Y. Lei, Y. Yang and D. J. Klionsky, *Cells*, 2020, **9**, 2025.
- 2 Y. Ding, Y. Fei and B. Lu, *Trends Pharmacol. Sci.*, 2020, **41**, 464–474.
- 3 S. J. Hughes, A. Testa, N. Thompson and I. Churcher, *Drug Discovery Today*, 2021, **26**, 2889–2897.
- 4 Q. Q. Zhou, H. T. Xiao, F. Yang, Y. D. Wang, P. Li and Z. G. Zheng, *Pharmacol. Res.*, 2023, **188**, 106627.
- 5 R. Ma, T. P. Ortiz Serrano, J. Davis, A. D. Prigge and K. M. Ridge, *FASEB J.*, 2020, **34**, 13156–13170.
- 6 I. B. McInnes and E. M. Gravallesse, *Nat. Rev. Immunol.*, 2021, **21**, 680–686.
- 7 J. Zheng, D. Chen, J. Xu, X. Ding, Y. Wu, H. C. Shen and X. Tan, *Bioorg. Med. Chem. Lett.*, 2021, **48**, 128229.
- 8 P. Li, Y. Zheng and X. Chen, *Front. Pharmacol.*, 2017, **8**, 460.
- 9 D. S. Pisetsky, *Nat. Rev. Nephrol.*, 2023, **19**, 509–524.
- 10 K. A. Zarembler and P. J. Godowski, *J. Immunol.*, 2002, **168**, 554–561.
- 11 K. Murphy, P. Travers and M. Walport, *Janeway's I.*, Garland Science, 8th edn, 2012.
- 12 G. George, G. L. Shyni and K. G. Raghu, *Inflammopharmacology*, 2020, **28**, 1457–1476.
- 13 D. P. Bondeson, B. E. Smith, G. M. Burslem, A. D. Buhimschi, J. Hines, S. Jaime-Figueroa, J. Wang, B. D. Hamman, A. Ishchenko and C. M. Crews, *Cell Chem. Biol.*, 2018, **25**, 78–87.
- 14 K. M. Riching, S. Mahan, C. R. Corona, M. McDougall, J. D. Vasta, M. B. Robers, M. Urh and D. L. Daniels, *ACS Chem. Biol.*, 2018, **13**, 2758–2770.
- 15 P. Ottis and C. M. Crews, *ACS Chem. Biol.*, 2017, **12**, 892–898.
- 16 M. Toure and C. M. Crews, *Angew. Chem., Int. Ed.*, 2016, **55**, 1966–1973.
- 17 G. E. Winter, D. L. Buckley, J. Paulk, J. M. Roberts, A. Souza, S. Dhe-Paganon and J. E. Bradner, *Science*, 2015, **348**, 1376–1381.
- 18 K. M. Sakamoto, K. B. Kim, A. Kumagai, F. Mercurio, C. M. Crews and R. J. Deshaies, *Proc. Natl. Acad. Sci. U. S. A.*, 2001, **98**, 8554–8559.
- 19 A. R. Schneekloth, M. Pucheault, H. S. Tae and C. M. Crews, *Bioorg. Med. Chem. Lett.*, 2008, **18**, 5904–5908.
- 20 M. Békés, D. R. Langley and C. M. Crews, *Nat. Rev. Drug Discovery*, 2022, **21**, 181–200.
- 21 (a) P. Ottis and C. M. Crews, *ACS Chem. Biol.*, 2017, **12**, 892–898; (b) M. Toure and C. M. Crews, *Angew. Chem., Int. Ed.*, 2016, **55**, 1966–1973; (c) J. Salami and C. M. Crews, *Science*, 2017, **355**, 1163–1167.
- 22 M. Muratani and W. P. Tansey, *Nat. Rev. Mol. Cell Biol.*, 2003, **4**, 192–201.
- 23 R. C. Aguilar and B. Wendland, *Curr. Opin. Cell Biol.*, 2003, **15**, 184–190.
- 24 T. J. Jarome and F. J. Helmstetter, *Neurobiol. Learn. Mem.*, 2013, **105**, 107–116.
- 25 A. Ocaña and A. Pandiella, *J. Exp. Clin. Cancer Res.*, 2020, **39**, 189.
- 26 D. Nandi, P. Tahiliani, A. Kumar and D. Chandu, *J. Biosci.*, 2006, **31**, 137–155.
- 27 N. Myeku, C. L. Clelland, S. Emrani, N. V. Kukushkin, W. H. Yu, A. L. Goldberg and K. E. Duff, *Nat. Med.*, 2016, **22**, 46–53.
- 28 A. Bricelj, C. Steinebach, R. Kuchta, M. Gütschow and I. Sosič, *Front. Chem.*, 2021, **9**, 707317.
- 29 X. Han and Y. Sun, *MedComm*, 2023, **4**, e290.
- 30 H. K. Namballa, P. Anchi, K. L. Mansa, J. P. Soni, C. Godugu, N. Shankaraiah and A. Kamal, *Bioorg. Chem.*, 2021, **117**, 105461.
- 31 I. M. Adcock, *Br. J. Pharmacol.*, 2007, **150**, 829–831.
- 32 P. J. Barnes, I. M. Adcock and K. Ito, *Eur. Respir. J.*, 2005, **25**, 552–563.
- 33 K. Tomita, P. J. Barnes and I. M. Adcock, *Biochem. Biophys. Res. Commun.*, 2003, **301**, 572–577.
- 34 Y. L. Chung, M. Y. Lee, A. J. Wang and L. F. Yao, *Mol. Ther.*, 2003, **8**, 707–717.
- 35 S. Hendrix, S. Sanchez, E. Ventriglia and S. Lemmens, *Int. J. Mol. Sci.*, 2020, **21**, 4539.
- 36 J. Ran and J. Zhou, *Thorac. Cancer*, 2019, **10**, 405–412.
- 37 B. R. Oh, D. H. Suh, D. Bae, B. Lee and Y. W. Song, *Arthritis Res. Ther.*, 2017, **19**, 154.
- 38 H. M. Blevins, Y. Xu, S. Biby and S. Zhang, *Front. Aging Neurosci.*, 2022, **14**, 879021.
- 39 F. Cao, S. de Weerd, D. Chen, M. R. H. Zwinderman, P. E. van der Wouden and F. J. Dekker, *Eur. J. Med. Chem.*, 2020, **208**, 112800.
- 40 Z. Cao, Z. Gu, S. Lin, D. Chen, J. Wang, Y. Zhao, Y. Li, T. Liu, Y. Li, Y. Wang, H. Lin and B. He, *ACS Chem. Biol.*, 2021, **16**, 2746–2751.

- 41 K. Yang, Y. Zhao, X. Nie, H. Wu, B. Wang, C. M. Almodovar-Rivera, H. Xie and W. Tang, *Cell Chem. Biol.*, 2020, **27**, 866–876.
- 42 J. Chotitumnavee, Y. Yamashita, Y. Takahashi, Y. Takada, T. Iida, M. Oba, Y. Itoh and T. Suzuki, *Chem. Commun.*, 2022, **58**, 4635–4638.
- 43 L. Ackerman, G. Acloque, S. Bacchelli, H. Schwartz, B. J. Feinstein, P. La Stella, A. Alavi, A. Gollerkeri, J. Davis, V. Campbell, A. McDonald, S. Agarwal, R. Karnik, K. Shi, A. Mishkin, J. Culbertson, C. Klaus, B. Enerson, V. Massa, E. Kuhn, K. Sharma, E. Keaney, R. Barnes, D. Chen, X. Zheng, H. Rong, V. Sabesan, C. Ho, N. Mainolfi, A. Slavin and J. A. Gollob, *Nat. Med.*, 2023, **29**, 3127–3136.
- 44 R. B. Kargbo, *ACS Med. Chem. Lett.*, 2019, **10**, 1251–1252.
- 45 J. Nunes, G. A. McGonagle, J. Eden, G. Kiritharan, M. Touzet, X. Lewell, J. Emery, H. Eidam, J. D. Harling and N. A. Anderson, *ACS Med. Chem. Lett.*, 2019, **10**, 1081–1085.
- 46 Y. Chen, Y. Ning, G. Bai, L. Tong, T. Zhang, J. Zhou, H. Zhang, H. Xie, J. Ding and W. Duan, *ACS Med. Chem. Lett.*, 2021, **12**, 82–87.
- 47 J. Zhang, L. Fu, B. Shen, Y. Liu, W. Wang, X. Cai, L. Kong, Y. Yan, R. Meng, Z. Zhang, Y. N. P. Chen, Q. Liu, Z. K. Wan, T. Zhou, X. Wang, P. Gavine, A. del Rosario, K. Ahn, U. Philipp, R. Attar, J. Yang, Y. Xu, J. P. Edwards and X. Dai, *Cell Chem. Biol.*, 2020, **27**, 1500–1509.
- 48 S. L. Degorce, O. Tavana, E. Banks, C. Crafter, L. Gingipalli, D. Kouvchinov, Y. Mao, F. Pachi, A. Solanki, V. Valge-Archer, B. Yang and S. D. Edmondson, *J. Med. Chem.*, 2020, **63**, 10460–10473.
- 49 A. Rowley, B. S. Brown, M. Stofega, H. Hoh, R. Mathew, V. Marin, R. X. Ding, R. A. McClure, F. M. Bittencourt, J. Chen, T. Gururaja, T. Kinoshita, X. Wang, A. Rivkin and K. R. Woller, *ACS Chem. Biol.*, 2022, **17**, 1315–1320.
- 50 J. Huang, D. Wan, J. Li, H. Chen, K. Huang and L. Zheng, *Epigenetics*, 2015, **10**, 62–72.
- 51 T. Agaloti, G. Chen and D. Thanos, *Cell*, 2002, **111**, 381–392.
- 52 Q. Jin, L. Zhuang, B. Lai, C. Wang, W. Li, B. Dolan, Y. Lu, Z. Wang, K. Zhao, W. Peng, S. Y. Dent and K. Ge, *EMBO Rep.*, 2014, **15**, 92–1201.
- 53 Z. I. Bassi, M. C. Fillmore, A. H. Miah, T. D. Chapman, C. Maller, E. J. Roberts, L. C. Davis, D. E. Lewis, N. W. Galwey, K. E. Waddington, V. Parravicini, A. L. Macmillan-Jones, C. Gongora, P. G. Humphreys, I. Churcher, R. K. Prinjha and D. F. Tough, *ACS Chem. Biol.*, 2018, **13**, 2862–2867.
- 54 R. Morris, N. J. Kershaw and J. J. Babon, *Protein Sci.*, 2018, **27**, 1984–2009.
- 55 C. J. Malesud, *Ther. Adv. Musculoskeletal Dis.*, 2018, **10**, 117–127.
- 56 X. Hu, J. Li, M. Fu, X. Zhao and W. Wang, *Signal Transduction Targeted Ther.*, 2021, **6**, 402.
- 57 R. R. Shah, J. M. Redmond, A. Mihut, M. Menon, J. P. Evans, J. A. Murphy, M. A. Bartholomew and D. M. Coe, *Bioorg. Med. Chem.*, 2020, **28**, 115326.
- 58 R. B. Kargbo, *ACS Med. Chem. Lett.*, 2021, **12**, 945–946.
- 59 B. Jiang, D. M. Weinstock, K. A. Donovan, H. W. Sun, A. Wolfe, S. Amaka, N. L. Donaldson, G. Wu, Y. Jiang, R. A. Wilcox, E. S. Fischer, N. S. Gray and W. Wu, *Cell Chem. Biol.*, 2023, **30**, 383–393.
- 60 D. J. Philpott, M. T. Sorbara, S. J. Robertson, K. Croitoru and S. E. Girardin, *Nat. Rev. Immunol.*, 2014, **14**, 9–23.
- 61 G. M. Argast, N. Fausto and J. S. Campbell, *Mol. Cell. Biochem.*, 2005, **268**, 129–140.
- 62 P. Canning, Q. Ruan, T. Schwerd, M. Hrdinka, J. L. Maki, D. Saleh, C. Suebsuwong, S. Ray, P. E. Brennan, G. D. Cuny, H. H. Uhlig, M. Gyrd-Hansen, A. Degterev and A. N. Bullock, *Chem. Biol.*, 2015, **22**, 1174–1184.
- 63 Y. Lai, X. Wang, X. Sun, S. Wu, X. Chen, C. Yang, W. Zhang, X. Yu, Y. Tong, F. Ma, H. Zheng, X. Zhang and S. He, *Biochem. Pharmacol.*, 2023, **214**, 115647.
- 64 A. Mares, A. H. Miah, I. E. D. Smith, M. Rackham, A. R. Thawani, J. Cryan, P. A. Haile, B. J. Votta, A. M. Beal, C. Capriotti, M. A. Reilly, D. T. Fisher, N. Zinn, M. Bantscheff, T. T. MacDonald, A. Vossenkamper, P. Dace, I. Churcher, A. B. Benowitz, G. Watt, J. Denyer, P. Scott-Stevens and J. D. Harling, *Commun. Biol.*, 2020, **3**, 140.
- 65 D. P. Bondeson, A. Mares, I. E. D. Smith, E. Ko, S. Campos, A. H. Miah, K. E. Mulholland, N. Routly, D. L. Buckley, J. L. Gustafson, N. Zinn, P. Grandi, S. Shimamura, G. Bergamini, M. Faelt-Savitski, M. Bantscheff, C. Cox, D. A. Gordon, R. R. Willard, J. J. Flanagan, L. N. Casillas, B. J. Votta, B. W. Den, K. Famm, L. Kruidenier, P. S. Carter, J. D. Harling, I. Churcher and C. M. Crews, *Nat. Chem. Biol.*, 2015, **11**, 611–617.
- 66 A. C. Belkina and G. V. Denis, *Nat. Rev. Cancer*, 2012, **12**, 465–477.
- 67 P. Filippakopoulos, J. Qi, S. Picaud, Y. Shen, W. B. Smith, O. Fedorov, E. M. Morse, T. Keates, T. T. Hickman, I. Felletar, M. Philpott, S. Munro, M. R. McKeown, Y. Wang, A. L. Christie, N. West, M. J. Cameron, B. Schwartz, T. D. Heightman, N. La Thangue, C. A. French, O. Wiest, A. L. Kung, S. Knapp and J. E. Bradner, *Nature*, 2010, **468**, 1067–1073.
- 68 B. Tong, M. Luo, Y. Xie, J. N. Spradlin, J. A. Tallarico, J. M. McKenna, M. Schirle, T. J. Maimone and D. K. Nomura, *Sci. Rep.*, 2020, **10**, 1–8.
- 69 K. M. DeMars, C. Yang, C. I. Castro-Rivera and E. Candelario-Jalil, *Biochem. Biophys. Res. Commun.*, 2018, **497**, 410–415.
- 70 R. B. Kargbo, *ACS Med. Chem. Lett.*, 2021, **12**, 688–689.
- 71 C. P. Tinworth, H. Lithgow, L. Dittus, Z. I. Bassi, S. E. Hughes, M. Muelbaier, H. Dai, I. E. D. Smith, W. J. Kerr, G. A. Burley, M. Bantscheff and J. D. Harling, *ACS Chem. Biol.*, 2019, **14**, 342–347.
- 72 J. Liu, L. Yuan, Y. Ruan, B. Deng, Z. Yang, Y. Ren, L. Li, T. Liu, H. Zhao, R. Mai and J. Chen, *J. Med. Chem.*, 2022, **65**, 6593–6611.
- 73 Z. Xiao, S. Song, D. Chen, R. van Merkerk, P. E. van der Wouden, R. H. Cool, W. J. Quax, G. J. Poelarends, B. N. Melgert and F. J. Dekker, *Angew. Chem., Int. Ed.*, 2021, **60**, 17514–17521.
- 74 R. B. Kargbo, *ACS Med. Chem. Lett.*, 2019, **10**, 276–277.

- 75 Y. Wu, Y. Yang, W. Wang, D. Sun, J. Liang, M. Zhu, H. Li and L. Chen, *Acta Pharm. Sin. B*, 2022, **12**, 4262–4265.
- 76 W. Wang, Y. Liu, L. Xiong, D. Sun, H. Wang, Z. Song, Y. Li, H. Li and L. Chen, *J. Nat. Prod.*, 2023, **86**, 67–781.
- 77 H. Yokoo, N. Shibata, M. Naganuma, Y. Murakami, K. Fujii, T. Ito, K. Aritake, M. Naito and Y. Demizu, *ACS Med. Chem. Lett.*, 2021, **12**, 236–241.
- 78 M. Hu, W. Zhou, Y. Wang, D. Yao, T. Ye, Y. Yao, B. Chen, G. Liu, X. Yang, W. Wang and Y. Xie, *Acta Pharm. Sin. B*, 2020, **10**, 1943–1953.
- 79 L. R. Bollu, P. V. Bommi, P. J. Monsen, L. Zhai, K. L. Lauing, A. Bell, M. Kim, E. Ladomersky, X. Yang, L. C. Platanias, D. E. Matei, M. G. Bonini, H. G. Munshi, R. Hashizume, J. D. Wu, B. Zhang, C. D. James, P. Chen, M. Kocherginsky, C. Horbinski, M. D. Cameron, A. A. Grigorescu, B. Yamini, R. V. Lukas, G. E. Schiltz and D. A. Wainwright, *J. Med. Chem.*, 2022, **65**, 15642–15662.
- 80 X. Jiang, J. Zhou, Y. Wang, X. Liu, K. Xu, J. Xu, F. Feng and H. Sun, *Eur. J. Med. Chem.*, 2021, **210**, 112949.
- 81 M. Lu, T. Liu, Q. Jiao, J. Ji, M. Tao, Y. Liu, Q. You and Z. Jiang, *Eur. J. Med. Chem.*, 2018, **146**, 251–259.
- 82 F. Wang, Y. Zhan, L. Manman, L. Wang, A. Zheng, C. Liu, H. Wang and T. Wang, *ACS Pharmacol. Transl. Sci.*, 2022, **6**, 76–87.
- 83 J. Ji, S. Ma, Y. Zhu, J. Zhao, Y. Tong, Q. You and Z. Jiang, *J. Med. Chem.*, 2023, **66**, 6070–6081.
- 84 M. Wang, J. Lu, M. Wang, C. Y. Yang and S. Wang, *J. Med. Chem.*, 2020, **63**, 7510–7528.
- 85 D. W. Robbins, M. A. Noviski, Y. S. Tan, Z. A. Konst, A. Kelly, P. Auger, N. Brathaban, R. Cass, M. L. Chan, G. Cherala, M. C. Clifton, S. Gajewski, T. G. Ingallinera, D. Karr, D. Kato, J. Ma, J. McKinnell, J. McIntosh, J. Mihalic, B. Murphy, J. R. Panga, G. Peng, J. Powers, L. Perez, R. Rountree, A. Tenn-McClellan, A. T. Sands, D. R. Weiss, J. Wu, J. Ye, C. Guiducci, G. Hansen and F. Cohen, *J. Med. Chem.*, 2024, **67**(4), 2321–2336.