

## REVIEW

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# Advancements in the preparation methods of artificial cell membranes with lipids

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In order to better understand the structure and function of biological cell membranes, various artificial systems have been developed. Among them, unilamellar lipid vesicle (liposome)-based artificial cell membranes have attracted great interest due to their inherently similar structure and compositions to cellular membranes. Diverse methodologies, including film hydration methods, emulsion templated technologies and FGA strategies, have been well established for simulating the cell membrane structure. In this review, advancements in the preparation methods of unilamellar liposomes will be summarized, and the applications, advantages and challenges of each method will also be discussed.

## 1. Introduction

A cell membrane is primarily composed of a phospholipid bilayer membrane, which is an important component of cells to separate the intracellular and extracellular environments. While it is necessary to protect the metabolic activities inside the cell, the cell membrane provides support for various functional membrane proteins, which facilitate the cellular activities, such as the transport of molecules,<sup>1</sup> energy exchange,<sup>2</sup> and signal transduction.<sup>3</sup> Therefore, it is of great significance to investigate the functions of the cellular membrane and its molecular mechanism in order to

understand the fundamental biological phenomena. However, the living cells are too complex to be studied directly, so artificial cell systems have been widely developed and utilized to simplify the relevant investigation due to their similar physicochemical properties to natural cell membranes.<sup>4–6</sup>

Artificial cells are usually defined as ordered structures enclosed by a membrane, which could integrate complex biological reactions in living cells.<sup>7</sup> Nowadays, several typically artificial membrane systems, including polymersomes,<sup>8</sup> proteinosomes,<sup>9</sup> colloidosomes,<sup>10</sup> as well as liposomes,<sup>11</sup> have been developed. Polymersomes are vesicles assembled from block copolymers, which could expand the properties of membranes. Proteinosomes are usually formed by chemically cross-linked protein-polymer nanoconjugates, providing an alternative method for membrane activation. Colloidosomes are microcapsules whose shells are composed of colloidal particles, and are widely applied in microencapsulation, such

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as the controlled release of cargoes. Liposomes are vesicles composed of lipids, which are most frequently used as a frame for artificial cells. Owing to their compositions and structures similar to native cell membranes, liposomes have attracted increasing attention and been widely investigated in biology-related fields.<sup>12</sup>

To prepare liposomes with similar properties to natural membrane systems, constructing liposomes with customized size and composition and a unilamellar structure is the essential target. During the last few decades, many methods have been developed, *e.g.*, film-based hydration methodologies and emulsion templated methods. Taking advantage of different technologies, unilamellar liposomes could be prepared with sizes ranging from nanometers to hundreds of micrometers, allowing the construction of artificial systems to mimic the organelle or cell structures. Here, in this review, advancements in the preparation methods of liposomes will be summarized.

## 2. Cell membrane structure

Cell is the fundamental structural unit of biological systems, which perform the basic functions including autopoiesis, metabolism, genetics, and interactions with the environment. Cell membrane is the boundary of the cell, which plays

important roles in protecting metabolic activities inside the cell.<sup>13</sup> Since the cell structures were discovered, the composition and structure of the natural membrane system have been widely investigated and revealed during the last few decades.<sup>14,15</sup> The phospholipid bilayer is the basic structural support of the cell membrane, in which the phospholipids arrange themselves with their polar heads to the outside, and their hydrophobic tails inside.<sup>16</sup> Many proteins are integrated into the lipid bilayer or simply associated with the membrane. In particular, the membranes of cells are distributed in an asymmetric manner with different phospholipids, proteins, or carbohydrates on each leaflet.<sup>17</sup> This asymmetry serves certain functions, such as the formation of a procoagulant interface upon cell disruption to trigger recognition events, the regulation of membrane budding, or the structural stability.<sup>18</sup> The importance of the cell membrane has been widely demonstrated and it is of great significance to study the formation and functions of cell membranes. However, it is immensely complex to directly study biological membranes with a lot of unexpected or unknown influences. Therefore, there is an urgent need for simplified and controllable models to understand these fundamental biological functions.

## 3. Methods for the preparation of unilamellar liposomes

Lipid vesicles (liposomes) have been developed for studying and emulating biological membranes.<sup>4,19</sup> While the composition of artificial liposomes is similar to or the same as those of natural membranes, the size and structure of vesicles are also comparable to those of native cell membranes. Furthermore, asymmetric lipid membranes can also be constructed using special technologies, which further facilitate the investigation of the structure and function of natural cell membranes.

Since the gentle hydration strategy was proposed by Bangham *et al.*,<sup>20</sup> many strategies have been developed to construct liposome-based bio-simulated systems with similar properties to natural membrane systems. Here, we will summarize the recent progress of some strategies to prepare customized liposomes with controlled features (*e.g.*, desired size, low polydispersity, tailored shapes, high yield production). Since the investigation of liposomes is such a broad field, it is impossible to cover all the technologies and here we will focus on the development of film hydration methods, emulsion templated strategies and frame-guided assembly.

### 3.1 Film hydration methodologies

Film hydration-based strategies are the earliest and most classical technologies for liposome fabrication.<sup>20,21</sup> It should be noted that film hydration-based strategies can not only be employed to construct liposome structures, but can also be widely applied for other amphiphiles or block polymers.<sup>12,22</sup> In the film hydration-based strategy, lipids or amphiphiles need to be dissolved in a volatile organic solvent (*e.g.*, chloroform), which will be then removed by evaporation under a flow of argon



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*the frame-guided assembly (FGA) strategy and its application in the structural analysis of protein complexes by Cryo-EM. In his research, a customized morphology of the amphiphilic assembly, *e.g.*, cuboid/rod-shaped vesicles and 2D nanopores, can be constructed by introducing Leading Hydrophobic Groups (LHG) onto the designed frame, which will subsequently guide the assembly of the amphiphiles. The FGA has also been demonstrated as a general strategy for different types of amphiphiles, including small-molecule surfactants, polymers, and natural lipids. Furthermore, combined with DNA nano-technology, a single-membrane protein has been guided into a nano-frame in the mono-disperse form, which has provided a promising tool to analyze low contrast protein structures with strong orientation preference under cryo-EM through the single-particle analysis method.*



Fig. 1 Schematic diagrams of the film hydration methods including (a) gentle hydration, (b) electroformation, and (c) gel-assisted swelling.<sup>12</sup> © 2019 Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim.

or nitrogen, as shown in Fig. 1. It has been demonstrated that, during the evaporation step, phospholipid molecules will self-assemble into a stack of bilayers on a solid substrate.<sup>23</sup> With the addition of an aqueous solution, the multi-lamellar membranes could be ruptured and separated from each other, which can be accelerated by extra forces, freezing, osmotic pressure, electric field, *etc.*<sup>24</sup> Therefore, subsequent swelling would result in the re-fusion of the separated lipid lamellas, finally forming unilamellar vesicle structures. According to the swelling process, the film hydration-based strategy can be divided into (1) gentle hydration, (2) electroformation and (3) gel-assisted swelling. In this section, we will introduce these strategies and their recent progress below.

**3.1.1 Gentle hydration.** A swelling-based gentle hydration method, generally referred to as natural swelling, was first described by Reeves and Dowben to prepare thin-walled vesicles.<sup>21</sup> In their work, a lipid film was first deposited on the surface of a flask and then carefully dried using nitrogen gas. Consequently, after the swelling process for two hours or more, a large number of liposome vesicles were generated. However, it was inevitable to produce multilamellar vesicles by this natural swelling process because the fusion process induced quick growth of liposome formation.<sup>21</sup> Later, freeze-thaw cycles were introduced to the hydration procedure and it was observed that the multilamellar vesicles could be further broken into unilamellar vesicles.<sup>25</sup> While freeze-thaw improved the encapsulation efficiency of the biomolecules (*e.g.*, protein), it should be noted that it would result in an unfavorable denaturation of the protein, which limits its application in certain biological investigations.<sup>24</sup>

It has been demonstrated that, during the swelling process, the electrostatic repulsion and steric effects of lipid head groups play an important role in the separation of lamellae, so it is difficult to prepare liposomes from electrostatically neutral lipids with a simply gentle hydration strategy.<sup>12</sup> Furthermore, electrostatic screening under high ionic strength could also hinder repulsion, which causes strong influences of salt concentration on the preparation of liposomes.<sup>26</sup> In order to address these problems, Yoshimura *et al.*<sup>27</sup> presented a method to enhance the formation of oligolamellar liposomes by doping the lipids with sugar (glucose, mannose, and fructose) (Fig. 2A). Upon hydration, water would permeate through lipid lamellae to dissolve the sugar and the difference in osmotic pressure will gradually increase from the film to the glass



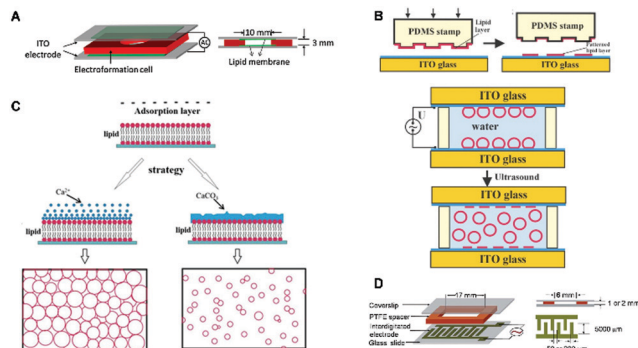
Fig. 2 Gentle hydration-based methods for generating unilamellar liposomes. (A) Swelling of lipid bilayer membranes assisted by sugar.<sup>27</sup> Copyright © 2009, American Chemical Society. (B) PEGylated lipid-and-sugar-doped gentle hydration.<sup>31</sup> Copyright © 2015 The Authors. Published by Elsevier B.V. (C) Swelling of the lipid film on the glass beads.<sup>33</sup> Copyright © 2012 Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim. (D) Formation of hybrid lipid compositions with giant vesicles (GVs).<sup>34</sup> Copyright © 2018, MDPI.

surface. Consequently, the osmotic pressure could increase the interlamellar distance and improve the formation of unilamellar liposomes. Besides doping with sugar, other kinds of molecules, such as polymers,<sup>28</sup> proteins<sup>29</sup> and DNA,<sup>30</sup> could also be employed to tune the osmotic pressure for enhanced liposome formation. Later, a similar technology was further developed to construct PEGylated liposomes, where the large molecular volume of PEG could increase the steric effects of the headgroup (Fig. 2B).<sup>31</sup> It is also found that the simultaneous usage of PEGylated lipids and sugar could facilitate the formation of unilamellar liposomes synergistically even at low temperatures or physiological concentrations of metals. Therefore, unstable bio-molecules, *e.g.*, transcription of RNA, could also be successfully accomplished within the GUVs by carrying out the gentle swelling process at low temperatures.

In addition to the typically used planar solid supports, sphere glass bead substrates possessing a larger surface area can also be utilized to improve the formation of liposomes. It is found that the increased surface area of the substrates exhibits a higher yield of unilamellar liposomes, and, taking this into account, glass beads have become attractive to be used as substrates for liposome preparation.<sup>32</sup> In 2012, a liposome encapsulating a minimal gene expression system was successfully created by using the porous bead matrix-assisted gentle hydration method, as shown in Fig. 2C.<sup>33</sup> Assisted by a freeze-thaw procedure, a higher population of GUVs and more efficient encapsulation of biomolecules were achieved. While liposomes with hybrid lipid compositions could be produced from the beads when a hybrid lipid film is deposited, it could also be prepared by combining two sets of beads coated with different lipids with shaking under hydration (Fig. 2D).<sup>34</sup>

Overall, gentle hydration is a common and straightforward way for unilamellar liposome preparation. Owing to its facile procedure, a large amount of unilamellar lipid vesicles can be produced without specific equipment. However, there are also some shortcomings to be solved: the preparation is always





**Fig. 3** Electroformation-based methods for unilamellar liposome preparation. (A) Electroformation on ITO electrodes.<sup>36</sup> © 2016 Elsevier B.V. All Rights Reserved. (B) Micro-contact printing technology involved an electroformation system for fabricating monodisperse liposomes.<sup>44</sup> Reproduced from ref. 44 with permission from The Royal Society of Chemistry. (C) Ca-mediated electroformation platforms for liposome generation.<sup>46</sup> Copyright © 2015, The Author(s). (D) The interdigitated ITO electrode-based electroformation platform for producing liposomes.<sup>47</sup> Reproduced from ref. 47 with permission from The Royal Society of Chemistry.

time-consuming and the yield under physiological ion conditions is usually low; it is inevitable to produce both single- and multi-layer liposomes at the same time with high polydispersity. Furthermore, the encapsulation efficiency of the biomolecules (*e.g.*, proteins and nucleic acids) is relatively poor.<sup>26</sup>

**3.1.2 Electroformation.** Electroformation is an aided film rehydration method to construct liposomes with more unilamellar membranes, compared to gentle film hydration. Different from the gentle hydration method, the amphiphiles/lipids should be deposited on special electrodes (Pt or indium titanium oxide (ITO) plates) rather than normal glass solid substrates. It is believed that the electric current can cause fluctuations in the bilayers, thus facilitating the detachment of lamellae and resulting in the formation of vesicles.<sup>35,36</sup> It is noted that the experimental conditions in the sample preparation, such as ionic strength, lipid composition, the substrate used to dry the lipid film and additional sugars, are critical to obtain giant vesicles.

The electroformation technology was invented by Angelova and Dimitrov, and, in the first study, a direct current (DC) electric field was applied to generate lipid vesicles.<sup>37</sup> As the DC electric field could result in bubbling due to water electrolysis, later, an alternating current (AC) electric field was introduced in the swelling process, which has become a common strategy.<sup>38</sup> Compared with the gentle hydration methods, it is observed that the electroformation method could produce a higher population of unilamellar vesicles without significant defects. This could be explained by the fact that the electric field can generate mechanical stress (visualized as gentle agitation of the lipid layers by the alternate low-frequency field) to rupture the lipid bilayers and the formation of pieces, which could further bend and form unilamellar liposomes.

In the early electroformation method, the unilamellar liposomes are usually prepared under low ionic strength conditions because the swelling process will be hindered by the screening

of the electric field induced by high ionic strength.<sup>39</sup> In order to construct liposomes under high ionic strength or physiological conditions, different strategies have been developed. Han *et al.*<sup>36</sup> applied a high-frequency AC field on the plasma-cleaned ITO electrode to generate giant liposomes (Fig. 3A). Under physiological conditions, it is possible for the high-concentration counterions to form an electric double layer near the electrode surface, which can hinder the formation of liposomes. However, the high-frequency AC could disrupt the electric double layer and induce abundant vesicles formed on the ITO electrode. Besides, the hydrophilic surface after plasma cleaning may also facilitate the hydration of the solid lipid film and the formation of lipid bilayers that subsequently bend and form vesicles. Therefore, the giant vesicles from various compositions of charged lipids or lipid mixtures could be electroformed under physiological conditions and even higher concentrations of NaCl. More interestingly, it was found that both the NaCl concentration and the AC field were associated with the diameter and formation of liposomes, respectively. Moreover, unilamellar liposomes with a high yield could also be generated under biorelevant conditions (*e.g.*, PBS buffer and cell culture medium), which provides a promising way to prepare GUVs on a large scale under physiologically relevant conditions. Unilamellar liposomes could also be generated by using a specific sequence of increasing AC field in phosphate-buffered saline.<sup>40</sup> Using a similar strategy, Bassereau and co-workers successfully reconstituted functional voltage-gated ion channels into cell-sized giant unilamellar vesicles (GUVs) by using buffers containing low (5 mM) or near-physiological salt concentrations (100 mM KCl, 4 mM HEPES (pH 7.2), 200 mM glucose).<sup>41</sup> With lots of effort, to date, liposomes can be generated using various buffers with the development of the electroformation method.<sup>12,36,42</sup>

Although it is believed that the applied AC field can cause fluctuations in the bilayers facilitating the detachment of lamellae, the fusion process includes quick growth of vesicles, and thus the size of the resultant GUV population is often heterogeneous.<sup>43</sup> To address this problem, in 2003, the Paunov group<sup>44</sup> developed a technique for the generation of monodisperse giant liposomes based on a combination of micro-patterning of ITO glass slides with lipid solution and electroformation, as shown in Fig. 3B. Since the micro-patterned lipid films provide a sufficient distance between the lipid patches, the coalescence of neighbouring liposomes is prevented, which is an essential process affecting the final size and distribution of the liposomes. Hence, in this way, liposomes with narrow distribution could be generated. More importantly, the average size of the produced liposomes could be tuned by designing the features of the micro-pattern. Similarly, Misawa and colleagues<sup>45</sup> utilized patterned supported lipid bilayers (SLBs) to prepare solvent-free giant vesicles (GVs). The SLBs were first formed on a patterned SiO<sub>2</sub> surface through vesicle fusion, then the electroformation was carried out to obtain the liposomes. Through this method, the produced GVVs possessed a mean size of 8.53 μm. For the same purpose, the Yang group<sup>46</sup> demonstrated a new strategy by applying calcium ions (Ca<sup>2+</sup>) or a calcium carbonate (CaCO<sub>3</sub>) mineral layer on the dry film of lipids during the electro-

formation process. Under an AC field, the homogeneous  $\text{Ca}^{2+}$  adsorption layer was transformed into a patterned  $\text{Ca}^{2+}$  or  $\text{CaCO}_3$  “barrier”, and therefore liposomes could be generated only from the interspaces among the formed “barrier” when the electroosmotic flow passes over the lipid film surface. The “barrier” subsequently confined the growth and fusion of the GUVs, leading to the formation of high-quality and narrow-dispersed GUVs (Fig. 3C). From these results, it could be found that the film pattern would be useful for generating more uniform liposomes under electroformation.

In addition to conventional electrodes (e.g., ITO and Pt plates), new types of electrodes have also been developed to obtain unilamellar lipid vesicles. In 2013, the Han group<sup>47</sup> developed a coplanar interdigitated electrode as a novel platform for the preparation of unilamellar liposomes (Fig. 3D). The interdigitated electrode itself provided a patterned substrate for the uniform deposition of lipid films, which broke the limitations of the opposite electrode layout. It was found that larger unilamellar liposomes could be generated when interdigitated electrodes with smaller widths were used under the same conditions. The size of the GUVs decreased with an elevated frequency at a constant amplitude. At a constant frequency of 10 Hz, the size of the GUVs increased with an increase in amplitude from 1 to 5 V and then decreased from 5 to 10 V. Recently, another novel type of electrode has also been utilized to readily generate giant unilamellar liposomes. In this study, readily available syringe needles were selected as stainless-steel electrodes, which could produce relatively monodisperse vesicles without observed lipid oxidation or hydrolysis.<sup>48</sup> Besides syringe needles, the electroformation on copper electrodes has also been employed for stable giant liposome preparation, which proved to be easy, inexpensive, and convenient.<sup>49</sup>

In general, the electroformation method is a simple and relatively fast way to obtain unilamellar and pure liposomes with a relatively high yield, which is summarized in Table 1. Furthermore, different parameters, such as voltage, frequency, and time, can be easily and precisely adapted to match the types of lipids or the ionic strength. However, the size of the unilamellar liposomes prepared through electroformation is always heterogeneous and the oxidation of the encapsulated

biomolecules also hinders its application. Additionally, special equipment is required to obtain liposomes.

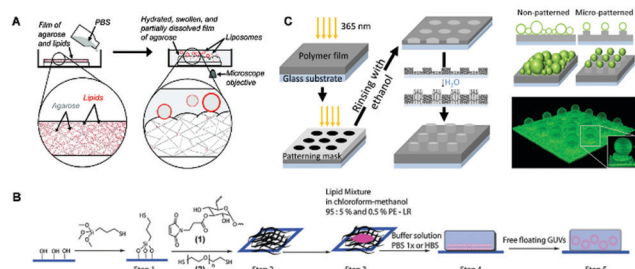
**3.1.3 Gel-assisted swelling.** Gel-assisted swelling is also a useful and common way for the preparation of liposomes. In this method, the solid surface is first pre-coated with a thin gel (e.g., agarose or poly(vinyl alcohol) (PVA)) layer, followed by the deposition of the lipid film on it. The porous properties of the hydrogel could increase the contact area between the hydrogel and the lipid molecules, which will promote the formation and separation of the lipid lamellae when the aqueous solution is added. Both the size and dispersity of the generated liposomes are affected by the physicochemical properties of the hydrogel, so most research studies focus on tuning the hydrogel substrates.

The gel-assisted swelling method was first described by the Mayer group<sup>50</sup> and agarose gel was used for enhanced swelling to form giant liposomes at physiological ionic strength (Fig. 4A). Due to the efficient swelling, the separation of the hybrid film were promoted, rendering the facile formation of unilamellar liposomes. More importantly, this method was much faster compared with the gentle hydration and electroformation methods which did not require any specialized equipment or lipids. Furthermore, this gel-assisted swelling was gentle enough to encapsulate bio-molecules and proteins including the active cytoskeletal actin-myosin networks, which have been demonstrated to be encapsulated with high efficiency.<sup>50</sup> Later, based on a similar concept, Marques *et al.*<sup>51</sup> developed a poly(vinyl alcohol) (PVA)-based swelling method for preparing liposomes. With the assistance of the PVA network, a variety of lipids or lipid hybrids were assembled into GUVs, which could also encapsulate biomolecules (proteins) efficiently.

Since the agarose and PVA hydrogel are cross-linked physically, these macro-molecules may be incorporated into the vesicles during hydration, which could change the physical properties of the membranes. To solve this problem, Kros *et al.*<sup>52</sup> reported a chemically cross-linked dextran-poly(ethylene glycol) (Dex-PEG) hydrogel substrate to prepare vesicles with various compositions (Fig. 4B). In this study, it was observed that the size distribution of the liposomes could be controlled by tuning the degree of

Table 1 Scope of electroformation conditions

Composition	Electrode	Frequency	Voltage	Time	Ionic strength	Yield	Ref.
EggL, DMPC	Pt	—	<3 V	—	Distilled water	Low	37
EPC, Chol/PC, PE/PC	ITO	10 Hz	1 V	2 h	Milli-Q water	Low	39
DOPC, POPC, DMPC, DPPC, DOPS or DOTAP	ITO	1 kHz	2.5 V	2 h	100 mM NaCl	High	36
egg-PC	ITO	10 Hz	0.1 V	25 min	1 mM $\text{CaCl}_2$	High	46
EggPC, CHS	ITO Pt	500 Hz	260–1670 $\text{V m}^{-1}$	5 min–overnight	PBS (10 mM, pH 7.4)	High	40
(DPhPC, EPG, EPA) SUVs	Pt	10 Hz	0.7 V	About 2 h	100 mM KCl, 4 mM HEPES (pH 7.2), 200 mM glucose	High	41
DMPC	ITO	10 Hz	0.5 V	1–1.5 h	0.1865 M glucose solution	High	44
DOPC/BODIPY-HPC	Au	10 Hz	10 V	1 h	100 mM KCl, 25 mM HEPES/NaOH pH 7.4	High	45
Egg PC, NBD-PE, TR-DHPE, DOPC	ITO	1 Hz to $10^4$ Hz	1–10 V	1–60 min	200 mM of sucrose solution	High	47
DOPC/Chol	Stainless steel	10 Hz, then 5 Hz	5 V	2 h + 30 min	200 mM sucrose in deionized (DI) water	High	48



**Fig. 4** Various hydrogel-assisted swelling methods for the formation of unilamellar liposomes. (A) Liposome formation from hybrid films of agarose and lipids.<sup>50</sup> Copyright © 2009, American Chemical Society. (B) Vesicle generation from dextran(ethylene glycol) hydrogels.<sup>52</sup> Reproduced from Ref. 52 with permission from The Royal Society of Chemistry. (C) Fabrication of liposomes from the patterned PNIPAAm hydrogel.<sup>55</sup> Copyright © 2019, American Chemical Society.

cross-linking within the gel network. Since then, the various types of chemically hydrogel substrates, including polyacrylamide (PAA),<sup>53</sup> polydimethylsiloxane (PDMS),<sup>54</sup> as well as poly(*N*-isopropylacrylamide) (PNIPAAm),<sup>55</sup> have also been developed for preparing giant unilamellar vesicles.

It is found that the characteristics of the hydrogel substrate, including crosslinking density, thickness, surface chemistry, as well as morphology, affect the formation of unilamellar liposomes. In order to achieve a hydrogel substrate with controllable characteristics, the Malmstadt group<sup>56</sup> reported an initiated chemical vapor deposition (iCVD) technique to generate porous charged polymer membranes with distinct morphologies and thicknesses, which could be further controlled to fabricate liposome low-polydispersity vesicles with higher yields. Later, an accessible method based on a patterned PNIPAAm hydrogel substrate for generating liposomes with controllable size was developed (Fig. 4C).<sup>55</sup> For obtaining the patterned hydrogel membrane, a photolithography technology was used, which resulted in shallow circular holes with controllable diameters ordered in a precise square pattern. By performing the swelling process in these defined holes, unilamellar liposomes with narrow distribution were prepared within a few minutes.

To sum up, the gel-assisted method provides a fast, robust, and simple way to prepare liposomes with a high yield as well as controllable size. Importantly, it is suitable for the preparation of liposomes under physiological buffer conditions as well as with the incorporation of different types of charged lipids. However, the incorporation of hydrogels (e.g., PVA or agarose) into the membrane may affect the properties of the bilayer of the liposomes (e.g., interfacial tension and permeability), and there were also difficulties in detaching the vesicles from the surface of the hydrogel substrates. As a novel method, the gel-assisted method has shown great promising potential for the preparation of liposomes that can be further utilized to serve as an artificial cell model.

### 3.2 Emulsion templated methods for liposome construction

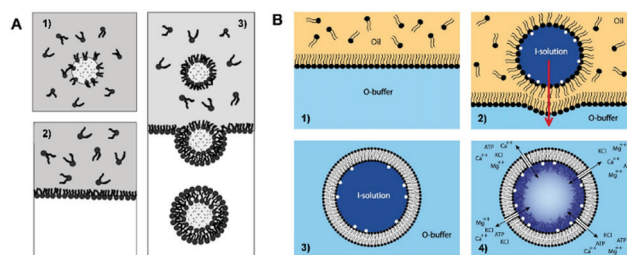
Although film hydration methods provide a simple way to prepare unilamellar liposomes, there are some limitations

including the poor size control, low encapsulation efficiency of the biomolecules, as well as time consumption.

Besides the simple film hydration methods, the emulsion templated technologies are also powerful tools to generate liposomes with the desired and controllable properties. In contrast to the hydration methods for generating liposomes through a process free of organic solvents, emulsion templated methods involve the use of organic solvents in the formation of liposomes. Because the emulsion involves both oil (organic) and aqueous phases, the methods can be generally classified into water-in-oil (W/O) emulsion templated and water-in-oil-in-water (W/O/W) double emulsion templated methods according to the number of involved phases. By controlling the properties of droplet emulsion, such as size, encapsulation, composition and symmetry, the desired liposomes possessing customized features can be achieved.

**3.2.1 Water-in-oil (W/O) emulsion templated method.** The water-in-oil (W/O) droplet emulsion templated method was first developed by Weitz *et al.* to prepare unilamellar vesicles from various amphiphilic compositions (e.g., lipids, mixed lipids, polymers, and surfactants).<sup>57</sup> The steps of the water-in-oil (W/O) emulsion templated method are as follows (Fig. 5A): initially, the aqueous droplets are emulsified in oil with an amphiphilic lipid, generating a lipid-stabilized inverted emulsion. Then, the lipid-saturated oil is poured over the second aqueous droplets to form a fully covered lipid monolayer at the water/oil interface. Subsequently, the emulsion is gently transferred into a lipid-saturated oil containing the monolayer, and gradually sinks down to pass through the lipid monolayer. Consequently, the emulsion picks up the second lipid layer to form the final vesicles.

Pontani and co-workers<sup>58</sup> successfully prepared symmetric unilamellar liposomes combined with actin, where the actin meshwork grew specifically from the inner monolayer of the liposome membrane (Fig. 5B). For this purpose, the first monolayer-stabilized droplets of I-solution containing actin were fabricated, followed by picking the second monolayer to form unilamellar vesicles surrounded by an O-buffer. The actin polymerization was accomplished by replacing the inner solution (I-solution) with a higher salt concentration solution (P-buffer).



**Fig. 5** Water-in-oil (W/O) emulsion templated method for generating unilamellar symmetric liposomes. (A) Schematic representation of the synthesis of liposomes from the emulsion.<sup>57</sup> Copyright © 2003, American Chemical Society. (B) Actin meshwork-containing symmetric liposome prepared through emulsion templated method.<sup>58</sup> Copyright © 2009 Biophysical Society. Published by Elsevier Inc. All rights reserved.



In a similar way, Elani *et al.*<sup>59</sup> successfully prepared ternary and thermally-responsive GUVs (DOPC/DPPC/cholesterol) for the first time by using a mixed solution of lipids. This kind of liposome could be further used to study the phase behaviors, and is also beneficial for incorporating content release function into individual vesicles, which may find applications in smart synthetic biosystems. For improving the yield or efficiency of unilamellar liposome production, liposomes possessing high polydispersity were further purified by second centrifugation.<sup>60</sup> So far, using the emulsion templated method, uniform liposomes with various compositions and encapsulation have been prepared.<sup>61–63</sup>

Since the inner and outer monolayers of liposomes are formed independently, Weitz *et al.*<sup>64</sup> developed a symmetric method for constructing vesicles with different compositions in each leaflet, which is realized by creating chemically different lipid monolayers at the droplet and oil/water interface. Based on this method, Oiki *et al.*<sup>65</sup> prepared asymmetric liposomes for protein insertion with controlled orientation. In this work, the KcsA potassium channels were added in the initial aqueous phase or the second aqueous solution, and they were successfully reconstituted with either an outside-out or inside-out orientation in the liposomes, which was due to the lipid-dependence of KcsA insertion (Fig. 6A). Similarly, the Lecommandoux group<sup>66</sup> reported a hybrid polymer-lipid vesicle possessing an asymmetric membrane containing an inner polymer (PBut-*b*-PEO) monolayer and outer lipid (POPC) monolayer (Fig. 6B), which provided a way to prepare a cell-like model system. Furthermore, asymmetrical lipid vesicles have been recently developed as useful nanocarriers for biomolecular molecules, *e.g.*, nucleic acids.<sup>67</sup> Taking advantage of the emulsion templated method, liposomes with asymmetric membrane composition could be readily prepared.

In order to obtain unilamellar liposomes with more controlled features, microfluidic devices have been combined with water-in-oil (W/O) droplet emulsions. In 2006, Lee *et al.*<sup>68</sup> demonstrated a shear-focusing-based emulsion generation system for preparing unilamellar liposomes, as shown in Fig. 7A. In this work, the droplet emulsions containing the target encapsulated species were first produced in the microfluidic channel, followed by injection into an aqueous mixture of ethanol and water. As the oil (oleic acid) rapidly dissolved in ethanol, the organic solvent could be removed immediately, which facilitated the separation of the phospholipids from the

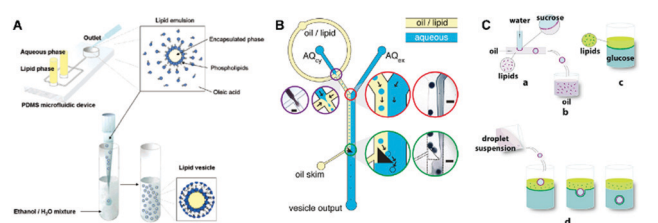


Fig. 7 Unilamellar liposomes prepared from the emulsion templated method with the assistance of microfluidic devices. (A) A shear-focusing-based emulsion generation system for preparing unilamellar liposomes.<sup>68</sup> Copyright © 2006, American Chemical Society. (B) Microfluidic assembly line for the stepwise synthesis of unilamellar liposomes.<sup>69</sup> Copyright © 2011, American Chemical Society. (C) Microfluidic channel for preparing asymmetric giant unilamellar vesicles.<sup>70</sup> Copyright © 2011, American Chemical Society.

liquid lipids, thus forcing the phospholipids to rearrange around the emulsion to obtain the vesicles. By using this method, various biological substances (*e.g.*, cancer cells, yeast cells, nanosized proteins) could be efficiently encapsulated in the liposomes. Later, Paegel *et al.*<sup>69</sup> developed a microfluidic assembly line for the stepwise synthesis of unilamellar liposomes (Fig. 7B). Droplet emulsions generated in a flow-focusing geometry flowed to a junction where a stable and reproducible flow-patterned interface was established from the oil and extracellular aqueous media. Continuous production of unilamellar liposomes was generated by a triangular post-mediated emulsion phase transfer process. In this way, unilamellar phospholipid vesicles with uniform and tunable size were realized, and the encapsulation efficiency of small-molecule cargoes was high (83%) compared to the bulk strategies such as gentle hydration and electroformation.

Microfluidic methods could also be applied to prepare asymmetric liposomes. In 2011, Malmstadt *et al.*<sup>70</sup> reported a microfluidic channel for preparing compositionally asymmetric giant unilamellar vesicles (Fig. 7C). The droplet emulsions of one aqueous flow were generated in a channel containing an immiscible oil stream, and the first lipid monolayer of the emulsions was formed. When these monolayer-coated emulsions pass through the second oil-water interface, the second monolayer comprising another lipid was formed. Finally, vesicles with desired encapsulated contents and controlled composition of lipids were formed with centrifugation, which may find potential applications in the investigation of membrane biophysics. Later, a novel microfluidic platform with a channel meander was also developed to generate compositionally controlled unilamellar liposomes.<sup>71</sup> The bending rigidity of these vesicles showed that asymmetry has a significant effect on the mechanical properties.

**3.2.2 Water-in-oil-in-water (W/O/W) double emulsion templated method.** The water-in-oil-in-water (W/O/W) double emulsion templated method is also a robust and efficient technology for the preparation of unilamellar liposomes. Different from the water-in-oil (W/O) emulsion, the double emulsion contains a middle oil (organic) phase between the two monolayers of lipids (Fig. 8A), and it is generated within microfluidic devices.<sup>72,73</sup> The encapsulation and lipid

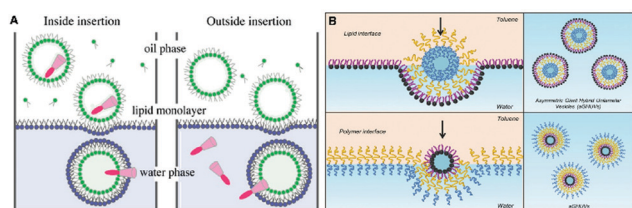
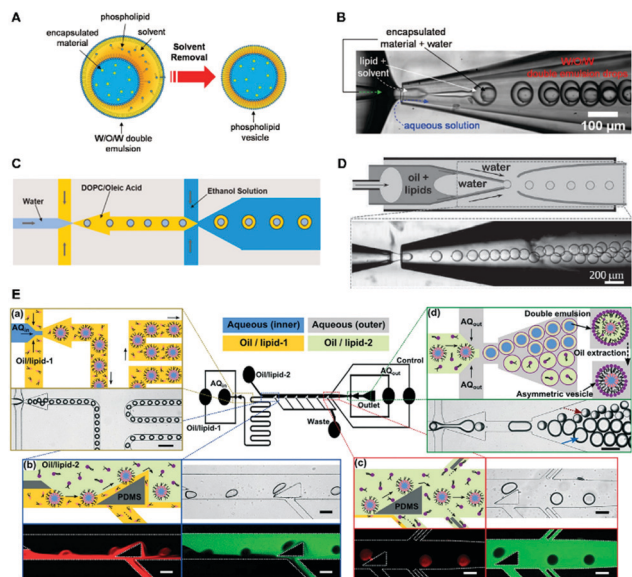


Fig. 6 (A) Schematic illustration of the experimental protocol used to prepare liposomes containing KcsA in either orientation.<sup>65</sup> Copyright © 2011, American Chemical Society. (B) Schematic representation of the generation of asymmetric giant hybrid unilamellar vesicles (aGHUV).<sup>66</sup> © 2017 The Authors. Published by Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim.



**Fig. 8** (A) Formation of liposomes through the double emulsion templated method.<sup>73</sup> Copyright © 2008, American Chemical Society. (B–D) Double emulsion generated from various microfluidic devices. (B) Double emulsion generated in a microfluidic device that combined a co-flow and a flow-focusing geometry.<sup>73</sup> Copyright © 2008, American Chemical Society. (C) The double emulsion prepared by consecutive flow-focusing channel geometries.<sup>74</sup> Rights managed by AIP Publishing. (D) Monodisperse ultrathin shell double emulsion.<sup>78</sup> © 2013 Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim. (E) Asymmetric unilamellar vesicles from an integrated microfluidic device.<sup>81</sup> Reproduced from ref. 81 with permission from The Royal Society of Chemistry.

composition of the double emulsion are determined by the inner phase and middle liquid, respectively. The construction of defined liposomes is achieved by using the double emulsion as the template after a de-wetting procedure to remove the oil from the middle phase. The double emulsion templated method is also a robust way to construct the desired unilamellar liposomes with customized features, such as size, shape, and encapsulation efficiency.

In 2008, Weitz *et al.*<sup>73</sup> reported a microfluidic device that combined a co-flow and a flow-focusing geometry to prepare liposomes (Fig. 8B). The high uniformity in both the size and shape of the double emulsion could be tuned by adjusting the flow rate of each fluid phase and the diameter of each capillary in the device. A little later, a stable and biocompatible double emulsion was also prepared by using consecutive flow-focusing channel geometries (Fig. 8C), and lipid vesicles were then formed through a solvent extraction process.<sup>74</sup> Since no toxic organic solvent was used, the double emulsion templated method enabled safe encapsulation and manipulation of the assortment of biological entities, such as cells, proteins, as well as nucleic acids. Recently,  $\beta$ -carotene-loaded double emulsion droplets have also been readily prepared for the formation of GUVs, which have potential applications in the protection and delivery of active compounds.<sup>75</sup>

In the double emulsion templated method, it is necessary to reduce the thickness of the oil shell to remove the oil, especially

for the quality of GUVs, because the oil trapped in the formed liposomes may cause the exposure of sensitive biorelevant materials (*e.g.*, nucleic acids, protein) to the organic solvent, which might either affect the activity of the biological macromolecules or make the giant liposomes extremely unstable.<sup>76,77</sup> To address the issue of residual organic solvent, Weitz *et al.*<sup>78</sup> developed a different microfluidic device to continuously produce monodisperse ultrathin shell double emulsion as a template to generate GUVs. As shown in Fig. 8D, by specially treating the surface of the tapered capillaries of the device, the double emulsion was forced to generate. The ultrathin oil shell could minimize the residual solvent that may be trapped within the bilayer membrane. Using the ultrathin shell double emulsion as the template, HeLa-based cell-free expression has been achieved within the engineered artificial cells. In order to deal with the problem of the time-consuming removal process of the organic solvent in the middle oil phase, rapid oil removal methods have been developed by using octanol or a copolymer surfactant (F-68).<sup>79,80</sup> When using octanol or F-68, the interfacial tension between the two lipid monolayers decreased sharply. As a result, the oil was expelled from the shell, and the two lipid monolayers stuck together, forming a lipid bilayer membrane.

The advantages of the double emulsion templated method make it a powerful tool to prepare asymmetrical structures. By using the double emulsion templated method, Chiarot *et al.*<sup>81</sup> successfully prepared asymmetric unilamellar vesicles with customized membrane composition, size, and luminal content. During fabrication, one special step was the replacement of the inner-leaflet-lipid solution with an outer-leaflet-lipid solution inside the channel (Fig. 8E), which enabled the produced vesicles to possess asymmetric membrane compositions. Most vesicles remained stable for several weeks, the membrane asymmetry was maintained for over 30 hours, and the trans-membrane protein ( $\alpha$ -hemolysin,  $\alpha$ HL) can be reconstituted into the bilayer, which hold the potential to be employed as model systems in membrane biology or as vehicles for drug delivery. For a similar purpose, this method has been further utilized to construct vesicles possessing lipid composition of Gram-negative bacterial outer membrane (OM).<sup>82</sup> It was observed that this technique generated monodisperse water-in-oil-in-water double emulsions at high-throughput. It was also found that the bending and expansion moduli of synthetic asymmetric membranes is physiologically relevant to the OM in *Pseudomonas aeruginosa* bacteria.

By the droplet emulsion templated methods, the vesicles with low polydispersity and high yield can be readily fabricated, which has also been summarized in Table 2, and the composition of the amphiphiles in the inner and outer layers of the vesicles is controllable. Moreover, the encapsulation of the desired hydrophilic cargoes can be achieved by doping with the inner water droplets. However, the oil is possibly trapped in the layers of the vesicles, which is likely to have undesired effects on the function of the vesicles when the biomolecules are embedded. Besides, the formation of droplet emulsions requires special microfluidic devices. In addition, the stability of droplet emulsions is also an annoying issue. For future preparation of biomimetic-stabilized



Table 2 Comparison of various emulsion-based methods

Methods	Size range, distribution	Encapsulation efficiency	Lamellarity	Asymmetric membrane	Yield	Residue of the organic solvent
W/O emulsion templated method	SUVs–GUVs, monodisperse	High	Unilamellar	Easy	High	Large
W/O/W double emulsion templated method	GUVs, monodisperse	High	Unilamellar	Relatively difficult	High	Small

liposomes, these concerns should be taken into consideration and addressed by integrating the methods.

### 3.3 SUVs for mimicking the cell membrane system

**3.3.1 Traditional strategies for SUV preparation.** While the large and giant unilamellar liposome vesicles (usually 100 nm to 200  $\mu\text{m}$ ) provide significant membrane models for mimicking cell membranes, the small unilamellar vesicles (SUVs, usually less than 100 nm) are also important to simulate and investigate the structure and functions of exosomes, viruses, as well as organelles.<sup>83–85</sup> It should be noted that, during the last few decades, many other methods have also been developed to prepare SUVs, including heating, sonication, extrusion, freeze-thawed, ethanol injection, detergent removal, as well as reverse-phase evaporation method.<sup>24,76</sup> In this section, we will focus on the typical methods based on the film hydration methodologies, where extra processes should be conducted at the hydration stage, including classical heating, sonication, and extrusion methods.

To obtain sterilized liposomes, a heating method was proposed by Kikuchi *et al.*<sup>86</sup> Interestingly, liposomes with a complete structure and high encapsulation efficiency were achieved after heating. Using a similar method, Mozafari *et al.*<sup>87</sup> introduced nontoxic glycerol to the system and conducted the preparation process at 120  $^{\circ}\text{C}$ , which successfully prepared SUVs rapidly without harmful chemicals. Later, sterol was also demonstrated to be a useful liposome ingredient for generating SUVs at high temperatures.<sup>88</sup> Besides the heating method, sonication is also one of the most widely employed technologies for preparing SUVs.<sup>24</sup> There are two main sonication methods: bath sonication and probe sonication. Owing to the high energy of the sonicator probe tip, the former method involves immersion in liposome dispersion in a water/ice bath.<sup>62</sup> Meanwhile, titanium may drop off the probe tip and pollute the solution. Therefore, the second sonication method is frequently used, which makes it easier to control the temperature of liposome dispersion, compared to probe sonication. Extrusion is also an important method to prepare nanosized SUVs, in which the liposome suspension is passed through a film filter with controlled pore size.<sup>89</sup> An extruder carrying a pump that pushes suspensions through the films is always needed to accomplish the extrusion process. Extrusion is widely utilized in sizing the liposomes.<sup>90</sup>

**3.3.2 Frame-guided assembly (FGA).** While most artificial cell systems are spherical liposomes, the natural systems exhibit much more complex morphologies. It is known that cytoskeletal proteins play an important role in maintaining the eukaryote cell morphology, which can bind to the membrane

proteins embedded in the lipid bilayer by either dynamic interactions or covalent bonds.<sup>91,92</sup> Therefore, many separated points of the bilayer are anchored onto the cytoskeleton, so the final morphology of the cell is influenced by the cytoskeleton. Inspired by the mechanism of the cytoskeleton-membrane protein-lipid bilayer structures, frame-guided assembly has been recently proposed by Liu and co-workers.<sup>93,94</sup> As shown in Fig. 9A, to conduct the frame-guided assembly process, three basic components are needed: a scaffold, leading hydrophobic groups (LHGs), as well as assembly amphiphiles. The LHGs are first anchored onto the well-designed scaffold (*e.g.*, gold nanoparticles, AuNPs) to form the initial frame, which would guide the assembly of the subsequently added assembly amphiphiles. Both the spherical and rod-like hetero-vesicles have been successfully constructed with gold nanoparticles as the scaffold.<sup>93</sup> Since then, a diversity of scaffolds and amphiphiles have been demonstrated to construct customized vesicle structures.<sup>95–97</sup> For example, cuboid vesicles or dumbbell-shaped assemblies were successfully constructed by using the 3D DNA nanostructure as the scaffold.<sup>97</sup>

In principle, liposomes could also be prepared through frame-guided assembly, which is powerful to construct membranes



Fig. 9 Various lipid-involved membrane mimicking systems from different scaffolds through the FGA method. (A) Shape-controlled vesicles constructed from the scaffold of AuNPs.<sup>93</sup> Copyright © 2014, Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim. (B) Virus-inspired enveloped DNA nanostructure from the scaffold of DNA nano-octahedrons.<sup>98</sup> Copyright © 2014, American Chemical Society. (C) Liposome mimicking cell membrane structure obtained from the scaffold of the AuNPs.<sup>99</sup> Copyright © 2020, Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim. (D) Uniform liposomes prepared from the scaffold of the DNA origami ring.<sup>100</sup> Copyright © 2017, Nature Publishing Group. (E) Liposomes in various shapes generated using DNA nanocages as outer templates.<sup>101</sup> Copyright © 2017, Springer Nature.

with similar physicochemical properties to the natural membranes. In 2014, Shih *et al.*<sup>98</sup> developed a virus-inspired enveloped DNA nanostructure using a similar method, which is illustrated in Fig. 9B. In their design, the lipid molecules were introduced to the DNA nano-octahedron (DNO) to preassemble onto outer handles. The initial solution was mixed with the lipids and surfactant (N-octyl- $\beta$ -D-glucopyranoside), and after the surfactant was removed through a dialysis procedure, the lipid was guided around the DNO by the handles to form the bilayer structures. It has also been observed that the envelopment of the DNA nanostructures within the lipid bilayers conferred protection against nuclease digestion and the immune activation was decreased 2 orders of magnitude below control. Recently, using a similar dialysis procedure, a liposome mimicking cell membrane structure has been successfully constructed under physiological conditions (Fig. 9C).<sup>99</sup> In this work, transmembrane  $\alpha$ -helices peptides, which play dominant roles in the interactions between membrane proteins and the cell membrane, were selected as LHGs to form the initial frame. Owing to their well-matched hydrophobic interactions between the transmembrane peptides and lipids, cell structure-like vesicles were readily prepared. Both the  $\alpha$ -helix conformation and transmembrane lengths of the peptides were demonstrated to benefit the formation as well as the stability of hetero-liposomes, and the FGA liposomes also exhibited enhanced stability compared to the liposomes prepared by conventional ways. Taking advantage of the biocompatibility and stability, the FGA liposomes were also employed to prepare novel drug delivery vehicles, which are promising in diagnostic imaging and cancer therapy applications.

Liposomes with controlled size can also be realized using DNA-origami rings as outer nano-templates. As illustrated in Fig. 9D, the lipid molecules were grafted onto the inner surface of the DNA rings to confine the liposome self-assembly during detergent removal.<sup>100</sup> Both the lipid grafting density and the template size were important factors for generating the size-defined vesicles. By designing the template sizes and using various anchored lipid densities, uniform liposomes with predefined sizes were successfully prepared. Besides the predictable size, the method can also be utilized with diverse lipid compositions, which represents a general way to guide the formation of liposomes. For fully mimicking the complexity of biological membranes, the assembly, arrangement and remodeling of liposomes with designed geometries were exquisitely controlled by a set of reconfigurable DNA nanocages (Fig. 9E).<sup>101</sup> Liposomes with high fidelity in tubular, toroid and other shapes were successfully prepared *in vitro*. Moreover, driven by the conformational changes of DNA nanocages, predictable membrane fusion and bending were also realized, which provides ideas for the systematic study of membrane mechanics.

In general, FGA provides a new, facile, and universal method for amphiphilic assembly, and exhibits promising application in the construction of liposomes with predictable shapes and sizes. The potential scaffold variability and assembly of amphiphiles make it possible in developing diverse amphiphilic assemblies. Combined with DNA nanotechnology, liposomes

with higher complexity and curvature can also be generated. Nevertheless, for the preparation of liposomes, frames are needed to anchor the leading hydrophobic groups and a proper LHG density is also required because it cannot sustain the assembly of the amphiphiles if too low. Moreover, the diameter of liposomes constructed from this method is always of nano-scale dimensions, and the development of this technology to prepare large-sized liposomes remains to be explored.

## 4. Summary and perspective

Owing to their similar composition and bilayer structure to biological membranes, liposomes, such as giant unilamellar liposomes, have become ideal artificial cellular membrane models for studying and exploring the intrinsic formation and functions of cell membranes. Diverse methods, including film hydration methodologies, emulsion templated strategies, as well as frame-guided assembly (FGA) technologies, have been developed to construct desired liposomes. Generally, film hydration methods provide common and straightforward techniques for preparing high-quality unilamellar liposomes. The emulsion templated technologies offer powerful tools to fabricate vesicles with low polydispersity, a high yield, and controllable asymmetry. FGA provides a reliable means to construct structures with predictable shapes and sizes.

While unilamellar liposomes with desired features, including size, uniformity, yield, symmetry, and composition, can be generated through these powerful methods, it should be noted that efficient encapsulation of biomolecules (*e.g.*, nucleic acids, proteins) is important to render artificial cell models with certain functions. However, the current film hydration strategies still suffer from poor encapsulation, which was described previously, as the difficult trans-bilayer movements of large molecules restrict efficient incorporation.<sup>26</sup> The nanosized liposomes generated from FGA limited the encapsulation of large-sized biomolecules. The emulsion templated methods may suffer from oil residues, which may affect the properties of the liposomes when the biomolecules are embedded. Therefore, developing novel or integrated strategies is necessary to increase encapsulation efficiency.

Meanwhile, although the construction of asymmetric liposomes possessing certain proteins has been demonstrated, the components of natural cellular membranes are far more complex than those of artificial membranes prepared using the present methods. In nature, cell membranes involved with many special lipids, membrane proteins or carbohydrates are still seldom incorporated in artificial systems. Furthermore, the orientation of membrane proteins is also important for the function of the cell membrane, which is still difficult to control by the currently reported strategies since the knowledge of interactions between the lipids and proteins is still scarce. Moreover, though the present artificial membrane systems with definite functions have been achieved, how to prepare more diverse and intricate systems is still in urgent need, which still requires the development of novel preparation methods.

In conclusion, based on the diverse well-established methods, controlled features of well-defined giant liposomes, including the encapsulation of biological molecules and asymmetric lipid membranes, have been achieved. It can be anticipated that, in the future, unilamellar liposomes prepared using these methods will keep providing powerful platforms to better understand complex natural biological phenomena.

## Author contributions

DYC conceived the study and participated in its design and coordination. YW participated in the sequence alignment and drafted the manuscript. PJF helped in drafting the manuscript. All authors read and approved the final manuscript.

## Conflicts of interest

There are no conflicts to declare.

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## Notes and references

- 1 J. Kulbacka, A. Choromańska, J. Rossowska, J. Weźgowiec, J. Saczko and M.-P. Rols, Cell membrane transport mechanisms: Ion channels and electrical properties of cell membranes, *Transport across natural and modified biological membranes and its implications in physiology and therapy*, 2017, 39.
- 2 R. B. Hamanaka and N. S. Chandel, Mitochondrial reactive oxygen species regulate cellular signaling and dictate biological outcomes, *Trends Biochem. Sci.*, 2010, **35**, 505.
- 3 P. Liu, H. Cheng, T. M. Roberts and J. J. Zhao, Targeting the phosphoinositide 3-kinase pathway in cancer, *Nat. Rev. Drug Discovery*, 2009, **8**, 627.
- 4 X. Wang, H. Du, Z. Wang, W. Mu and X. Han, Versatile Phospholipid Assemblies for Functional Synthetic Cells and Artificial Tissues, *Adv. Mater.*, 2021, **33**, 2002635.
- 5 Y. Zhu, X. Guo, J. Liu, F. Li and D. Yang, Emerging Advances of Cell-Free Systems toward Artificial Cells, *Small Methods*, 2020, **4**, 2000406.
- 6 M. Komiya, M. Kato, D. Tadaki, T. Ma, H. Yamamoto, R. Tero, Y. Tozawa, M. Niwano and A. Hirano-Iwata, Advances in Artificial Cell Membrane Systems as a Platform for Reconstituting Ion Channels, *Chem. Rec.*, 2020, **20**, 730.
- 7 Y. Ding, F. Wu and C. Tan, Synthetic biology: A bridge between artificial and natural cells, *Life*, 2014, **4**, 1092.
- 8 M. Marguet, C. Bonduelle and S. Lecommandoux, Multicompartmentalized polymeric systems: towards biomimetic cellular structure and function, *Chem. Soc. Rev.*, 2013, **42**, 512.
- 9 X. Huang, A. J. Patil, M. Li and S. Mann, Design and construction of higher-order structure and function in proteinosome-based protocells, *J. Am. Chem. Soc.*, 2014, **136**, 9225.
- 10 M. Li, R. L. Harbron, J. V. Weaver, B. P. Binks and S. Mann, Electrostatically gated membrane permeability in inorganic protocells, *Nat. Chem.*, 2013, **5**, 529.
- 11 R. Chandrawati and F. Caruso, Biomimetic liposome- and polymersome-based multicompartmentalized assemblies, *Langmuir*, 2012, **28**, 13798.
- 12 E. Rideau, F. R. Wurm and K. Landfester, Self-Assembly of Giant Unilamellar Vesicles by Film Hydration Methodologies, *Adv. Biosyst.*, 2019, **3**, 1800324.
- 13 X. Cheng and J. C. Smith, Biological membrane organization and cellular signaling, *Chem. Rev.*, 2019, **119**, 5849.
- 14 P. L. Yeagle, Lipid regulation of cell membrane structure and function, *FASEB J.*, 1989, **3**, 1833.
- 15 H. T. McMahon and J. L. Gallop, Membrane curvature and mechanisms of dynamic cell membrane remodelling, *Nature*, 2005, **438**, 590.
- 16 R. Mezzenga, J. M. Seddon, C. J. Drummond, B. J. Boyd, G. E. Schröder-Turk and L. Sagalowicz, Nature-Inspired design and application of lipidic lyotropic liquid crystals, *Adv. Mater.*, 2019, **31**, 1900818.
- 17 J. E. Rothman and J. Lenard, Membrane asymmetry, *Science*, 1977, **195**, 743.
- 18 G. Di Paolo and P. De Camilli, Phosphoinositides in cell regulation and membrane dynamics, *Nature*, 2006, **443**, 651.
- 19 X. Zhang, X. Shao, Z. Cai, X. Yan and W. Zong, The fabrication of phospholipid vesicle-based artificial cells and their functions, *New J. Chem.*, 2021, **45**, 3364.
- 20 A. D. Bangham and R. Horne, Negative staining of phospholipids and their structural modification by surface-active agents as observed in the electron microscope, *J. Mol. Biol.*, 1964, **8**, 660.
- 21 J. P. Reeves and R. M. Dowben, Formation and Properties of Thin-walled Phospholipid Vesicles, *J. Cell. Physiol.*, 1969, **73**, 49.
- 22 E. Rideau, R. Dimova, P. Schwill, F. R. Wurm and K. Landfester, Liposomes and polymersomes: a comparative review towards cell mimicking, *Chem. Soc. Rev.*, 2018, **47**, 8572.
- 23 Y. P. Patil and S. Jadhav, Novel methods for liposome preparation, *Chem. Phys. Lipids*, 2014, **177**, 8.
- 24 A. Jesorka and O. Orwar, Liposomes: technologies and analytical applications, *Annu. Rev. Anal. Chem.*, 2008, **1**, 801.
- 25 J. P. Colletier, B. Chaize, M. Winterhalter and D. Fournier, Protein encapsulation in liposomes: efficiency depends on interactions between protein and phospholipid bilayer, *BMC Biotechnol.*, 2002, **2**, 1.
- 26 P. Walde, K. Cosentino, H. Engel and P. Stano, Giant vesicles: preparations and applications, *ChemBioChem*, 2010, **11**, 848.



- 27 K. Tsumoto, H. Matsuo, M. Tomita and T. Yoshimura, Efficient formation of giant liposomes through the gentle hydration of phosphatidylcholine films doped with sugar, *Colloids Surf., B*, 2009, **68**, 98.
- 28 L. M. Dominak and C. D. Keating, Polymer encapsulation within giant lipid vesicles, *Langmuir*, 2007, **23**, 7148.
- 29 F. C. Tsai, B. Stuhmann and G. H. Koenderink, Encapsulation of active cytoskeletal protein networks in cell-sized liposomes, *Langmuir*, 2011, **27**, 10061.
- 30 S. F. Shimobayashi and M. Ichikawa, Emergence of DNA-encapsulating liposomes from a DNA-lipid blend film, *J. Phys. Chem. B*, 2014, **118**, 10688.
- 31 K. Shohda, K. Takahashi and A. Suyama, A method of gentle hydration to prepare oil-free giant unilamellar vesicles that can confine enzymatic reactions, *Biochem. Biophys. Rep.*, 2015, **3**, 76.
- 32 D. S. Dimitrov, J. Li, M. Angelova and R. K. Jain, Surface Effects in Preparation of Cell-Size Liposomes, *FEBS Lett.*, 1984, **176**, 398.
- 33 Z. Nourian, W. Roelofsen and C. Danelon, Triggered gene expression in fed-vesicle microreactors with a multifunctional membrane, *Angew. Chem., Int. Ed.*, 2012, **51**, 3114.
- 34 R. Tanasescu, U. Mettal, A. Colom, A. Roux and A. Zumbuehl, Facile and Rapid Formation of Giant Vesicles from Glass Beads, *Polymers*, 2018, **10**, 54.
- 35 P. Méléard, L. A. Bagatolli and T. Pott, Giant Unilamellar Vesicle Electroformation, *Methods Enzymol.*, 2009, **465**, 161.
- 36 Q. Li, X. Wang, S. Ma, Y. Zhang and X. Han, Electroformation of giant unilamellar vesicles in saline solution, *Colloids Surf., B*, 2016, **147**, 368.
- 37 M. I. Angelova and D. S. Dimitrov, Liposome Electroformation, *Faraday Discuss. Chem. Soc.*, 1986, **81**, 303.
- 38 D. S. Dimitrov and M. I. Angelova, Lipid swelling and liposome formation mediated by electric fields, *J. Electroanal. Chem. Interfacial Electrochem.*, 1988, **253**, 323.
- 39 M. I. Angelova, S. Soléau, P. Méléard, F. Faucon and P. Bothorel, Preparation-of-giant-vesicles-by exexternal AC electric fields. Kinetics and applications, *Trends in colloid and interface science*, 1992, **89**, 127.
- 40 P. Lefrancois, B. Goudeau and S. Arbault, Electroformation of phospholipid giant unilamellar vesicles in physiological phosphate buffer, *Integr. Biol.*, 2018, **10**, 429.
- 41 S. Aimon, J. Manzi, D. Schmidt, J. A. Poveda Larrosa, P. Bassereau and G. E. Toombes, Functional reconstitution of a voltage-gated potassium channel in giant unilamellar vesicles, *PLoS One*, 2011, **6**, 25529.
- 42 L. R. Montes, A. Alonso, F. M. Goni and L. A. Bagatolli, Giant unilamellar vesicles electroformed from native membranes and organic lipid mixtures under physiological conditions, *Biophys. J.*, 2007, **93**, 3548.
- 43 J. R. Howse, R. A. Jones, G. Battaglia, R. E. Ducker, G. J. Leggett and A. J. Ryan, Templated formation of giant polymer vesicles with controlled size distributions, *Nat. Mater.*, 2009, **8**, 507.
- 44 P. Taylor, C. Xu, P. D. Fletcher and V. N. Paunov, A novel technique for preparation of monodisperse giant liposomes, *Chem. Commun.*, 2003, 1732.
- 45 N. Misawa, T. Motegi and R. Tero, Electroformation from patterned single-layered supported lipid bilayers for formation of giant vesicles with narrow size distribution, *Appl. Phys. Express*, 2014, **7**, 117001.
- 46 F. Tao and P. Yang, Ca-mediated electroformation of cell-sized lipid vesicles, *Sci. Rep.*, 2015, **5**, 9839.
- 47 H. Bi, B. Yang, L. Wang, W. Cao and X. Han, Electroformation of giant unilamellar vesicles using interdigitated ITO electrodes, *J. Mater. Chem. A*, 2013, **1**, 7125.
- 48 V. Pereno, D. Carugo, L. Bau, E. Sezgin, J. Bernardino de la Serna, C. Eggeling and E. Stride, Electroformation of Giant Unilamellar Vesicles on Stainless Steel Electrodes, *ACS Omega*, 2017, **2**, 994.
- 49 H. G. Behuria, B. K. Biswal and S. K. Sahu, Electroformation of liposomes and phytosomes using copper electrode, *J. Liposome Res.*, 2020, **1**.
- 50 K. S. Horger, D. J. Estes, R. Capone and M. Mayer, Films of agarose enable rapid formation of giant liposomes in solutions of physiologic ionic strength, *J. Am. Chem. Soc.*, 2009, **131**, 1810.
- 51 A. Weinberger, F. C. Tsai, G. H. Koenderink, T. F. Schmidt, R. Itri, W. Meier, T. Schmatko, A. Schroder and C. Marques, Gel-assisted formation of giant unilamellar vesicles, *Biophys. J.*, 2013, **105**, 154.
- 52 N. Lopez Mora, J. S. Hansen, Y. Gao, A. A. Ronald, R. Kieleyka, N. Malmstadt and A. Kros, Preparation of size tunable giant vesicles from cross-linked dextran(ethylene glycol) hydrogels, *Chem. Commun.*, 2014, **50**, 1953.
- 53 E. Parigoris, D. L. Dunkelmann, A. Murphy, N. Wili, A. Kaeck, C. Dumrese, N. Jimenez-Rojo and U. Silvan, Facile generation of giant unilamellar vesicles using polyacrylamide gels, *Sci. Rep.*, 2020, **10**, 4824.
- 54 T. Fan, Q. Wang, N. Hu, Y. Liao, X. Chen, Z. Wang, Z. Yang, J. Yang and S. Qian, Preparation of giant lipid vesicles with controllable sizes by a modified hydrophilic polydimethylsiloxane microarray chip, *J. Colloid Interface Sci.*, 2019, **536**, 53.
- 55 J. Schultze, A. Vagias, L. Ye, E. Prantl, V. Breising, A. Best, K. Koynov, C. M. Marques and H. J. Butt, Preparation of Monodisperse Giant Unilamellar Anchored Vesicles Using Micropatterned Hydrogel Substrates, *ACS Omega*, 2019, **4**, 9393.
- 56 N. Movsesian, M. Tittensor, G. Dianat, M. Gupta and N. Malmstadt, Giant Lipid Vesicle Formation Using Vapor-Deposited Charged Porous Polymers, *Langmuir*, 2018, **34**, 9025.
- 57 S. Pautot, B. J. Frisken and D. A. Weitz, Production of Unilamellar Vesicles Using an Inverted Emulsion, *Langmuir*, 2003, **19**, 2870.
- 58 L. L. Pontani, J. van der Gucht, G. Salbreux, J. Heuvingh, J. F. Joanny and C. Sykes, Reconstitution of an actin cortex inside a liposome, *Biophys. J.*, 2009, **96**, 192.
- 59 K. Karamdad, J. W. Hindley, G. Bolognesi, M. S. Friddin, R. V. Law, N. J. Brooks, O. Ces and Y. Elani, Engineering thermoresponsive phase separated vesicles formed via emulsion phase transfer as a content-release platform, *Chem. Sci.*, 2018, **9**, 4851.

- 60 K. Tsumoto, Y. Hayashi, J. Tabata and M. Tomita, A reverse-phase method revisited: Rapid high-yield preparation of giant unilamellar vesicles (GUVs) using emulsification followed by centrifugation, *Colloids Surf., A*, 2018, **546**, 74.
- 61 Y. Ai, R. Xie, J. Xiong and Q. Liang, Microfluidics for Biosynthesizing: from Droplets and Vesicles to Artificial Cells, *Small*, 2020, **16**, 1903940.
- 62 C. Has and P. Sunthar, A comprehensive review on recent preparation techniques of liposomes, *J. Liposome Res.*, 2020, **30**, 336.
- 63 A. Moga, N. Yandrapalli, R. Dimova and T. Robinson, Optimization of the Inverted Emulsion Method for High-Yield Production of Biomimetic Giant Unilamellar Vesicles, *ChemBioChem*, 2019, **20**, 2674.
- 64 S. Pautot, B. J. Frisken and D. A. Weitz, Engineering asymmetric vesicles, *Proc. Natl. Acad. Sci. U. S. A.*, 2003, **100**, 10718.
- 65 M. Yanagisawa, M. Iwamoto, A. Kato, K. Yoshikawa and S. Oiki, Oriented reconstitution of a membrane protein in a giant unilamellar vesicle: experimental verification with the potassium channel KcsA, *J. Am. Chem. Soc.*, 2011, **133**, 11774.
- 66 A. Peyret, E. Ibarboure, J. F. Le Meins and S. Lecommandoux, Asymmetric Hybrid Polymer-Lipid Giant Vesicles as Cell Membrane Mimics, *Adv. Sci.*, 2018, **5**, 1700453.
- 67 M. B. C. de Matos, B. S. Miranda, Y. Rizky Nuari, G. Storm, G. Leneweit, R. M. Schiffelers and R. J. Kok, Liposomes with asymmetric bilayers produced from inverse emulsions for nucleic acid delivery, *J. Drug Targeting*, 2019, **27**, 681.
- 68 Y. C. Tan, K. Hettiarachchi, M. Siu, Y. R. Pan and A. P. Lee, Controlled microfluidic encapsulation of cells, proteins, and microbeads in lipid vesicles, *J. Am. Chem. Soc.*, 2006, **128**, 5656.
- 69 S. Matosevic and B. M. Paegel, Stepwise synthesis of giant unilamellar vesicles on a microfluidic assembly line, *J. Am. Chem. Soc.*, 2011, **133**, 2798.
- 70 P. C. Hu, S. Li and N. Malmstadt, Microfluidic fabrication of asymmetric giant lipid vesicles, *ACS Appl. Mater. Interfaces*, 2011, **3**, 1434.
- 71 K. Karamdad, R. V. Law, J. M. Seddon, N. J. Brooks and O. Ces, Preparation and mechanical characterisation of giant unilamellar vesicles by a microfluidic method, *Lab Chip*, 2015, **15**, 557.
- 72 A. S. Utada, E. Lorenceau, D. R. Link, P. D. Kaplan, H. A. Stone and D. A. Weitz, Monodisperse double emulsions generated from a microcapillary device, *Science*, 2005, **308**, 537.
- 73 H. C. Shum, D. Lee, I. Yoon, T. Kodger and D. A. Weitz, Double emulsion templated monodisperse phospholipid vesicles, *Langmuir*, 2008, **24**, 7651.
- 74 S. Deshpande, A. Birnie and C. Dekker, Stable, biocompatible lipid vesicle generation by solvent extraction-based droplet microfluidics, *Biomicrofluidics*, 2011, **5**, 044113.
- 75 M. Michelon, Y. Huang, L. G. de la Torre, D. A. Weitz and R. L. Cunha, Single-step microfluidic production of W/O/W double emulsions as templates for  $\beta$ -carotene-loaded giant liposomes formation, *Chem. Eng. J.*, 2019, **366**, 27.
- 76 A. Akbarzadeh, R. Rezaei-Sadabady, S. Davaran, S. W. Joo, N. Zarghami, Y. Hanifehpour, M. Samiei, M. Kouhi and K. Nejati-Koshki, Liposome: classification, preparation, and applications, *Nanoscale Res. Lett.*, 2013, **8**, 102.
- 77 K. Funakoshi, H. Suzuki and S. Takeuchi, Formation of giant lipid vesiclelike compartments from a planar lipid membrane by a pulsed jet flow, *J. Am. Chem. Soc.*, 2007, **129**, 12608.
- 78 L. R. Arriaga, S. S. Datta, S. H. Kim, E. Amstad, T. E. Kodger, F. Monroy and D. A. Weitz, Ultrathin shell double emulsion templated giant unilamellar lipid vesicles with controlled microdomain formation, *Small*, 2014, **10**, 950.
- 79 S. Deshpande, Y. Caspi, A. E. Meijering and C. Dekker, Octanol-assisted liposome assembly on chip, *Nat. Commun.*, 2016, **7**, 10447.
- 80 N. N. Deng, M. Yelleswarapu and W. T. Huck, Monodisperse Uni- and Multicompartment Liposomes, *J. Am. Chem. Soc.*, 2016, **138**, 7584.
- 81 L. Lu, J. W. Schertzer and P. R. Chiarot, Continuous microfluidic fabrication of synthetic asymmetric vesicles, *Lab Chip*, 2015, **15**, 3591.
- 82 S. Maktabi, J. W. Schertzer and P. R. Chiarot, Dewetting-induced formation and mechanical properties of synthetic bacterial outer membrane models (GUVs) with controlled inner-leaflet lipid composition, *Soft Matter*, 2019, **15**, 3938.
- 83 S. G. Antimisariis, S. Mourtas and A. Marazioti, Exosomes and Exosome-Inspired Vesicles for Targeted Drug Delivery, *Pharmaceutics*, 2018, **10**, 218.
- 84 L. Schoonen and J. C. van Hest, Compartmentalization Approaches in Soft Matter Science: From Nanoreactor Development to Organelle Mimics, *Adv. Mater.*, 2016, **28**, 1109.
- 85 L. Xu, P. Frederik, K. F. Pirolo, W. H. Tang, A. Rait, L. M. Xiang, W. Q. Huang, I. Cruz, Y. Z. Yin and E. H. Chang, Self-Assembly of a Virus-Mimicking Nanostructure System for Efficient Tumor-Targeted Gene Delivery, *Hum. Gene Ther.*, 2002, **13**, 469.
- 86 H. Kikuchi, A. Carlsson, K. Yachi and S. Hirota, Possibility of heat sterilization of liposomes, *Chem. Pharm. Bull.*, 1991, **39**, 1018.
- 87 M. R. Mozafari, C. J. Reed, C. Rostron, C. Kocum and E. Piskin, Construction of stable anionic liposome-plasmid particles using the heating method: a preliminary investigation, *Cell. Mol. Biol. Lett.*, 2002, **7**, 923.
- 88 M. R. Mozafari, Liposomes: an overview of manufacturing techniques, *Cell. Mol. Biol. Lett.*, 2005, **10**, 711.
- 89 N. Berger, A. Sachse, J. Bender, R. Schubert and M. Brandl, Filter extrusion of liposomes using different devices: comparison of liposome size, encapsulation efficiency, and process characteristics, *Int. J. Pharm.*, 2001, **223**, 55.
- 90 G. Liu, S. Hou, P. Tong and J. Li, Liposomes: Preparation, Characteristics, and Application Strategies in Analytical Chemistry, *Crit. Rev. Anal. Chem.*, 2020, **1**.
- 91 M. P. Sheetz, Cell control by membrane-cytoskeleton adhesion, *Nat. Rev. Mol. Cell Biol.*, 2001, **2**, 392.
- 92 E. Luna and A. Hitt, Cytoskeleton-plasma membrane interactions, *Science*, 1992, **258**, 955.

- 93 Y. Dong, Y. Sun, L. Wang, D. Wang, T. Zhou, Z. Yang, Z. Chen, Q. Wang, Q. Fan and D. Liu, Frame-guided assembly of vesicles with programmed geometry and dimensions, *Angew. Chem., Int. Ed.*, 2014, **53**, 2607.
- 94 Y. Dong and D. Liu, Frame-Guided Assembly of Amphiphiles, *Chem. – Eur. J.*, 2015, **21**, 18018.
- 95 Y. Dong, Z. Yang and D. Liu, Using Small Molecules to Prepare Vesicles with Designable Shapes and Sizes via Frame-Guided Assembly Strategy, *Small*, 2015, **11**, 3768.
- 96 C. Zhou, Y. Zhang, Y. Dong, F. Wu, D. Wang, L. Xin and D. Liu, Precisely Controlled 2D Free-Floating Nanosheets of Amphiphilic Molecules through Frame-Guided Assembly, *Adv. Mater.*, 2016, **28**, 9819.
- 97 Y. Dong, Y. R. Yang, Y. Zhang, D. Wang, X. Wei, S. Banerjee, Y. Liu, Z. Yang, H. Yan and D. Liu, Cuboid Vesicles Formed by Frame-Guided Assembly on DNA Origami Scaffolds, *Angew. Chem., Int. Ed.*, 2017, **56**, 1586.
- 98 S. D. Perrault and W. M. Shih, Virus-inspired membrane encapsulation of DNA nanostructures to achieve in vivo stability, *ACS Nano*, 2014, **8**, 5132.
- 99 C. Wang, J. Piao, Y. Li, X. Tian, Y. Dong and D. Liu, Construction of Liposomes Mimicking Cell Membrane Structure through Frame-Guided Assembly, *Angew. Chem., Int. Ed.*, 2020, **59**, 15176.
- 100 Y. Yang, J. Wang, H. Shigematsu, W. Xu, W. M. Shih, J. E. Rothman and C. Lin, Self-assembly of size-controlled liposomes on DNA nanotemplates, *Nat. Chem.*, 2016, **8**, 476.
- 101 Z. Zhang, Y. Yang, F. Pincet, M. C. Llaguno and C. Lin, Placing and shaping liposomes with reconfigurable DNA nanocages, *Nat. Chem.*, 2017, **9**, 653.