



# Aquaporin-driven hydrogen peroxide transport: a case of molecular mimicry?<sup>†</sup>

Darren Wragg,<sup>a</sup> Stefano Leoni<sup>\*b</sup> and Angela Casini<sup>†a</sup>Cite this: *RSC Chem. Biol.*, 2020, 1, 390Received 30th August 2020,  
Accepted 18th November 2020

DOI: 10.1039/d0cb00160k

rsc.li/rsc-chembio

**Aquaporins (AQPs) are membrane proteins that have evolved to control cellular water uptake and efflux, and as such are amongst the most ancient biological “devices” in cellular organisms. Recently, using metadynamics, we have shown that water nano-confinement within aquaporin channels results into bidirectional water movement along single file chains, extending previous investigations. Here, the elusive mechanisms of H<sub>2</sub>O<sub>2</sub> facilitated transport by the human ‘peroxiporin’ AQP3 has been unravelled via a combination of atomistic simulations, showing that while hydrogen peroxide is able to mimic water during AQP3 permeation, this comes at a certain energy expense due to the required conformational changes within the channel. Furthermore, the intrinsic water dynamics allows for host H<sub>2</sub>O<sub>2</sub> molecule solvation and transport in both directions, highlighting the fundamental role of water nano-confinement for successful transduction and molecular selection. Overall, the bidirectional nature of the water flux under equilibrium conditions along with the mimicking behavior of hydrogen peroxide during a conductance event introduce a new chemical paradigm never reported so far in any theoretical paper involving any aquaporin isoform.**

## Introduction

The transport of H<sub>2</sub>O<sub>2</sub> across cellular membranes by specific transmembrane proteins, named aquaporins (AQPs), has been considered one of the last milestones in the timeline of hydrogen peroxide discoveries in biochemistry.<sup>1</sup> H<sub>2</sub>O<sub>2</sub> can be dangerous or acts as a signaling molecule in various cellular processes, including cell migration and stem cell proliferation.<sup>2</sup> In general, AQPs are responsible for water permeation across

biomembranes,<sup>1</sup> and are involved in a variety of important physiological processes. Therefore, they are also considered promising targets for therapeutic intervention.<sup>1,3,4</sup> To date, the mechanism of hydrogen peroxide conductance is not fully understood. Water and H<sub>2</sub>O<sub>2</sub> are highly related molecules, featuring similar dipole moment and hydrogen-bonding capability. Based on this similarity, it has been postulated that a permeating H<sub>2</sub>O<sub>2</sub> molecule replacing a H<sub>2</sub>O molecule within the AQP channel should be energetically costless. Beitz and co-workers suggested that in principle all water-permeable AQPs can be H<sub>2</sub>O<sub>2</sub> channels, and studying wild-type and mutated AQPs showed a positive correlation between the extent of water and H<sub>2</sub>O<sub>2</sub> permeability in different isoforms.<sup>5</sup> The few molecular dynamics (MD) studies reported so far investigated H<sub>2</sub>O<sub>2</sub> permeation across AQP1<sup>6,7</sup> – an orthodox aquaporin which has not yet been fully validated experimentally as a hydrogen peroxide channel<sup>5,8</sup> – or using plant AQPs isoforms,<sup>6</sup> which are functionally and structurally very different from the mammalian isoforms.

The functional unit of human AQPs is a tetramer with each monomer providing an independent channel consisting of three topological elements: an extracellular and a cytoplasmic vestibule connected by an extended narrow pore (Fig. 1A). Various structural and computational studies revealed water molecules passing through the AQPs channel in a single file.<sup>1,9,10</sup> Two main constriction sites have been identified within the channel, which are responsible for substrate selectivity,<sup>1,9,11,12</sup> namely the aromatic/arginine (ar/R) selectivity filter (SF), in proximity of the extracellular entrance, forming the narrowest part of the pore, and two conserved Asn-Pro-Ala (NPA) motifs in the middle of the channel, where the positive N-terminal ends of two half helices meet (Fig. 1A). Water that enters this region is re-oriented by the dipoles of the emanating half helices, such that hydrogen bonds between neighbouring water molecules in the chain are disrupted. Concerning other substrates, such as glycerol, the AQPs filtering can be to some extent explained by size selection due to steric hindrance, a venue that has been extensively explored in former approaches.<sup>1</sup>

<sup>a</sup> Department of Chemistry, Technical University of Munich, Lichtenbergstr. 4, 85748 Garching, Germany. E-mail: angela.casini@tum.de

<sup>b</sup> School of Chemistry, Cardiff University, Park Place, CF103AT Cardiff, UK. E-mail: leonis@cf.ac.uk

<sup>†</sup> Electronic supplementary information (ESI) available. See DOI: 10.1039/d0cb00160k





**Fig. 1** (A) hAQP3 monomer showing the ar/R (yellow) and NPA (green) selectivity filters. (a) Position of H<sub>2</sub>O<sub>2</sub> as it enters the extracellular pocket of hAQP3 with lipophilic/hydrophilic molecular surface (hydrophilic = blue, lipophilic = red). (b) H<sub>2</sub>O<sub>2</sub> orientation within the ar/R SF including H-bond network with residues Tyr212, Ala213 and Arg218. (c) H<sub>2</sub>O<sub>2</sub> orientation during translocation between ar/R and NPA selectivity filters, interacting with residues Ala213 and Asn215 of the NPA motif. (d) H<sub>2</sub>O<sub>2</sub> orientation when leaving the NPA selectivity filter, including H-bond network involving His81 and a bridging water molecule to Asn83. Figure generated using the MOE software.<sup>23</sup> (B) Average free-energy surface of H<sub>2</sub>O<sub>2</sub> uptake (solid line) and efflux (dashed line), calculated using metadynamics sampling. Averaged curve shown in red.

This framework however, fails to justify how a membrane protein, evolved for water transport and osmotic pressure equilibration, can also selectively conduct ‘small’ signalling molecules as H<sub>2</sub>O<sub>2</sub>.

To further expand our understanding of AQP3 conductance, a combination of theoretical atomistic methods is applied here to attempt the elucidation of the mechanism of hydrogen peroxide permeation *via* human aquaglyceroporin-3 (AQP3).<sup>1</sup> While this isoform is known to permeate both water and glycerol, it also facilitates uptake of H<sub>2</sub>O<sub>2</sub><sup>13,14</sup> and has an important role in cancer development.<sup>3</sup>

## Results

In details, a homology model of AQP3 was built following previously reported procedures.<sup>15,16</sup> Thus, both biased and unbiased *in silico* techniques were used to study the permeation of H<sub>2</sub>O<sub>2</sub> through the protein channel, namely metadynamics,<sup>17,18</sup> to provide both free energy and mechanistic trajectories of conductance events, and transition path sampling (TPS),<sup>19</sup> to achieve unbiased trajectory information.

Metadynamics is an atomistic simulation technique enabling, within the same framework, acceleration of rare events along selected reaction coordinates, so-called collective variables (CV), and estimation of the free energy of complex molecular systems.<sup>17,18</sup> This technique makes it possible to accelerate conformational transitions between metastable states, broadening the scope of MD simulations, and it has been already successfully applied to calculate the free-energy surface (FES) for the interactions of substrate/drugs with biomolecules, including DNA secondary structures.<sup>20</sup> Recently, using metadynamics simulations, we investigated the mechanisms of glycerol permeation by the human AQP3 isoform.<sup>21</sup> The obtained results showed that single-file water permeation

through AQP3 is always bidirectional at equilibrium conditions, and that individual water molecules are able to ‘hop’ over each other (‘leap-frog’ mechanism), whilst traveling in opposite directions in certain key areas of the pore. Such mechanism for water transport may be essential to understand aquaporins ability to switch from uptake to efflux quickly under changing physiological conditions.<sup>21</sup> Interestingly, it appeared that glycerol molecules exploit an existing water conduction mechanism in AQP3.

The herewith performed metadynamics calculations consisted of manifold 200 ns simulations to capture as many H<sub>2</sub>O<sub>2</sub> permeation events as possible (see Experimental for details, ESI†). Overall, 12 uptake and 10 efflux events of H<sub>2</sub>O<sub>2</sub> molecules were successfully observed from a total of 1.2 μs of combined simulation time. The averaged free-energy surfaces for both H<sub>2</sub>O<sub>2</sub> uptake and efflux are shown in Fig. 1B and indicate the NPA region of the AQP3 channel as the one with the highest energy barrier during hydrogen peroxide conductance. This result suggests the NPA region as selectivity site for uncharged solutes, at variance with previously reported classical MD studies,<sup>22</sup> whereby the ar/R SF was identified as the primary filter for other uncharged substrates. Specifically, the calculated absolute free-energy ΔG values for H<sub>2</sub>O<sub>2</sub> match the trend previously observed for water and glycerol permeation, with H<sub>2</sub>O<sub>2</sub> values fitting between those of the other two substrates<sup>21</sup> (Table 1 and Fig. 1B).

Previous studies reported the experimentally calculated permeability values for water and glycerol permeation *via* different AQPs, including AQP3, in different cells.<sup>1,24–26</sup> Moreover, a few studies also included hydrogen peroxide transport rates *via* AQP3.<sup>14,27</sup> In general, permeability values for water are in the order of 10<sup>-3</sup> cm s<sup>-1</sup>, while glycerol and H<sub>2</sub>O<sub>2</sub> are in the range of 10<sup>-6</sup> cm s<sup>-1</sup>. It should be noted that while experimental permeability values are usually assessed using an osmotic flux,





molecules within the extracellular pocket (EP, Fig. 2A), with little interplay with the protein residues until it approached the ar/R SF.

The preferred conformation of hydrogen peroxide is the nonplanar *gauche* or *skew* form;<sup>28</sup> however, during the entire permeation event, H<sub>2</sub>O<sub>2</sub> was observed to switch conformation whilst maintaining H-bond interactions with surrounding water molecules, as shown by the variation of the dihedral angle of the peroxide molecule (Fig. 2B and Fig. S2, Movie M2 in ESI†). Specifically, as H<sub>2</sub>O<sub>2</sub> approaches the ar/R SF, it adopts a close to *trans* orientation to H-bond residues Tyr 212, Ala 213 and Arg218 (Fig. 2B and Fig. S3A, ESI†). Within the ar/R SF, H<sub>2</sub>O<sub>2</sub> prefers a *cis*-like conformation (Fig. 1A(b), 2B and Fig. S3B, ESI†), which is maintained until approaching the NPA selectivity filter, with the oxygen atom facing downwards towards the NPA motif, mimicking water molecules (Fig. 1A(c), 2C and Fig. S3C, ESI†). Within the NPA region H<sub>2</sub>O<sub>2</sub> reverts back to the preferred *skew* orientation (Fig. 2B, C and Fig. S3D, ESI†).

Finally, as H<sub>2</sub>O<sub>2</sub> leaves the NPA region it again adopts a *cis*-like orientation with the oxygen atoms facing upwards, fully mimicking the water flipping mechanism (Fig. 1A(d), 2C and Fig. S3E, ESI†). In this region, H<sub>2</sub>O<sub>2</sub> interacts with the backbone oxygen of the highly conserved His81 (Fig. 1A(d) and 2B). This interaction is observed globally in metadynamics and specifically in TPS calculations, and has been previously reported for the glycerol transport mechanism in hAQP3<sup>21</sup> and GlpF,<sup>29</sup> respectively. TPS also confirms that water molecules coordinate to the backbone oxygen of the His81 residue located below the NPA selectivity filter (Movie M3 in ESI†). Whilst doing so, adjacent water molecules are able to hop over each other and re-join the water chain ahead of the coordinating molecule, or compete for binding to His81 (Video M3 in the ESI†). Interestingly, H<sub>2</sub>O<sub>2</sub> can also bind to His81 and, mimicking water movement, allow adjacent waters to ‘hop’ over itself, therefore, maintaining the bidirectional water mechanism (Movie M2 in ESI†).

Overall, the extended residence time and multiple orientation and conformational changes observed for H<sub>2</sub>O<sub>2</sub> passing through the NPA motif can explain the increase in free energy observed within this region (Fig. 1B). As H<sub>2</sub>O<sub>2</sub> enters and exits the NPA, to fully mimic the water movement it must acquire a *cis*-type configuration, which has been shown to be the higher energy conformation for the peroxide molecule.<sup>30</sup> These higher energy conformations, whilst essential for matching mechanistic rules of interaction with both the water chain and pore residues, are in line with the observed free energy increase. Therein, differences seen between calculated FES of individual trajectories, can therefore, be attributed to more or less strong fluctuations in the population of high-energy conformers. The constant switching of orientation to maintain optimal favorable interactions appears to be a justified energy expenditure for the conductance mechanism.

## Conclusions

Recently, metadynamics showed us that water nanoconfinement within aquaporin channels results into bidirectional water movement along single file chains.<sup>21</sup> This structure and

intrinsic water dynamics allows for host molecule solvation and transport in both directions, highlighting the fundamental role of water nanoconfinement for successful transduction and molecular selection. Here, we shed light onto mechanistic steps of hydrogen peroxide facilitated transport by the human AQP3 using a combination of metadynamics and TPS methods, unveiling how mimicking water is key to efficiently negotiate internal energy barriers, while turning bare water transport into a signalling gateway. Moreover, our study further pinpoints the existence of a basic mechanism of water transport, over which guest/solute molecules are translocated. This allows for a direct comparison of peroxide with glycerol and for an atomistic elucidation of the mechanistic basis of selectivity, which is rooted into specific protein–guest molecule–water interactions. In summary, by harvesting details of water-mediated small molecule transport, we are laying the foundation for a molecular understanding of selective permeation in aquaporins, which is key for drug (inhibitor) design. We hope our work will stimulate the use of such advanced atomistic simulations to compute relevant quantities such as substrate/transporter binding free energies, and to generate very detailed transition path ensembles for in-depth understanding of elusive steps of solute transports across biomembranes, as well as for the design of bioinspired synthetic water channels.<sup>31</sup>

## Conflicts of interest

Authors have no conflict of interest to declare.

## Acknowledgements

This work has been performed using resources provided by the ‘‘Cambridge Service for Data Driven Discovery’’ (CSD3, <http://csd3.cam.ac.uk>) system operated by the University of Cambridge Research Computing Service (<http://www.hpc.cam.ac.uk>) funded by EPSRC Tier-2 capital grant EP/P020259/1. We gratefully acknowledge the support of NVIDIA Corporation with the donation of a Quadro P5000 GPU used for this research. We thank ARCCA for access to computational facilities.

## References

- 1 G. Soveral, S. Nielsen and A. Casini, *Aquaporins in Health and Disease: New Molecular Targets for Drug Discovery*, CRC Press, Taylor & Francis Group, 2017.
- 2 G. P. Bienert and F. Chaumont, *Biochim. Biophys. Acta, Gen. Subj.*, 2014, **1840**, 1596–1604.
- 3 B. Aikman, A. De Almeida, S. M. Meier-Menches and A. Casini, *Metallomics*, 2018, **10**, 696–712.
- 4 C. Prata, S. Hrelia and D. Fiorentini, *Int. J. Mol. Sci.*, 2019, **20**, 1–15.
- 5 A. Almasalmeh, D. Krenc, B. Wu and E. Beitz, *FEBS J.*, 2014, **281**, 647–656.
- 6 R. M. Cordeiro, *Biochim. Biophys. Acta, Gen. Subj.*, 2015, **1850**, 1786–1794.
- 7 M. Yusupov, J. Razzokov, R. M. Cordeiro and A. Bogaerts, *Oxid. Med. Cell. Longevity*, 2019, **2019**, 2930504.



- 8 G. P. Bienert, A. L. B. Møller, K. A. Kristiansen, A. Schulz, I. M. Møller, J. K. Schjoerring and T. P. Jahn, *J. Biol. Chem.*, 2007, **282**, 1183–1192.
- 9 H. Sui, B. G. G. Han, J. K. J. K. K. Lee, P. Walian and B. K. B. K. K. Jap, *Nature*, 2001, **414**, 872–878.
- 10 L. Janosi and M. Ceccarelli, *PLoS One*, 2013, **8**, e59897.
- 11 K. Murata, K. Mitsuoka, T. Hirai, T. Walz, P. Agre, J. B. Heymann, A. Engel and Y. Fujiyoshi, *Nature*, 2000, **407**, 599–605.
- 12 D. Fu, A. Libson, L. J. W. Miercke, C. Weitzman, P. Nollert, J. Krucinski and R. M. Stroud, *Science*, 2000, **290**, 481–486.
- 13 E. W. Miller, B. C. Dickinson and C. J. Chang, *Proc. Natl. Acad. Sci. U. S. A.*, 2010, **107**, 15681–15686.
- 14 J. R. Thiagarajah, J. Chang, J. A. Goettel, A. S. Verkman and W. I. Lencer, *Proc. Natl. Acad. Sci. U. S. A.*, 2017, **114**, 568–573.
- 15 A. De Almeida, A. F. Mó, D. Wragg, M. Wenzel, P. Kavanagh, G. Barone, S. Leoni, G. Soveral and A. Casini, *Chem. Commun.*, 2017, **53**, 3830–3833.
- 16 A. De Almeida, A. P. Martins, A. F. Móscá, H. J. Wijma, C. Prista, G. Soveral and A. Casini, *Mol. BioSyst.*, 2016, **12**, 1564–1573.
- 17 G. Bussi and A. Laio, *Nat. Rev. Phys.*, 2020, **2**, 200–212.
- 18 A. Barducci, M. Bonomi and M. Parrinello, *Wiley Interdiscip. Rev.: Comput. Mol. Sci.*, 2011, **1**, 826–843.
- 19 P. G. Bolhuis and C. Dellago, in *Advanced Computer Simulation Approaches for Soft Matter Sciences III*, ed. K. Kremer and C. Holm, Springer, Berlin, Heidelberg, 2009, pp. 167–233.
- 20 D. Wragg, A. de Almeida, R. Bonsignore, F. E. Kühn, S. Leoni and A. Casini, *Angew. Chem.*, 2018, 14524–14528.
- 21 D. Wragg, A. de Almeida, A. Casini and S. Leoni, *Chem. – Eur. J.*, 2019, **25**, 8713–8718.
- 22 J. S. Hub and B. L. de Groot, *Proc. Natl. Acad. Sci. U. S. A.*, 2008, **105**, 1198–1203.
- 23 C. C. Group, Inc. Montr. Quebec, Canada., 2018.
- 24 I. V. Da Silva, M. Barroso, T. Moura, R. Castro and G. Soveral, *Int. J. Mol. Sci.*, 2018, **19**, 130–143.
- 25 A. Serna, A. Galán-Cobo, C. Rodrigues, I. Sánchez-Gomar, J. J. Toledo-Aral, T. F. Moura, A. Casini, G. Soveral and M. Echevarría, *J. Cell. Physiol.*, 2014, **229**, 1787–1801.
- 26 L. Méndez-Giménez, S. Becerril, S. P. Camões, I. V. Da Silva, C. Rodrigues, R. Moncada, V. Valentí, V. Catalán, J. Gómez-Ambrosi, J. P. Miranda, G. Soveral, G. Frühbeck and A. Rodríguez, *Int. J. Obes.*, 2017, **41**, 1394–1402.
- 27 C. Rodrigues, C. Pimpao, A. Coxixo, D. Lopes, P. A. Pedersen, F. Antunes and G. Soveral, *Cancers*, 2019, **11**, 1–17.
- 28 G. P. Bienert, J. K. Schjoerring and T. P. Jahn, *Biochim. Biophys. Acta, Biomembr.*, 2006, **1758**, 994–1003.
- 29 M. O. Jensen, S. Park, E. Tajkhorshid and K. Schulten, *Proc. Natl. Acad. Sci. U. S. A.*, 2002, **99**, 6731–6736.
- 30 L. Song, M. Liu, W. Wu, Q. Zhang and Y. Mo, *J. Chem. Theory Comput.*, 2005, **1**, 394–402.
- 31 I. Kocsis, Z. Sun, Y. M. Legrand and M. Barboiu, *npj Clean Water*, 2018, **1**, 1–11.

