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Review

Gallic acid: A versatile antioxidant with promising therapeutic and industrial applications

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Gallic acid: A versatile antioxidant with promising therapeutic and industrial applications

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Abstract: Oxidative stress, a result of an overproduction and accumulation of free radicals, is the leading cause of several degenerative diseases such as cancer, atherosclerosis, cardiovascular diseases, ageing and inflammatory diseases. Polyphenols form an important class of naturally occurring antioxidants, having innumerable biological activities such as anticancer, antifungal, antibacterial, antiviral, antiulcer and anticholesterol, to name a few. Among various polyphenols, gallic acid (3,4,5-trihydroxybenzoic acid), a naturally occurring low molecular weight triphenolic compound, has emerged as a strong antioxidant and an efficient apoptosis inducing agent. Starting from the bioavailability and the biosynthetic pathway of gallic acid, this review includes various in vitro, in vivo and in silico studies providing the mode of action, radical scavenging activity, ability to inhibit lipid peroxidation, maintenance of endogenous defense system and metal ion chelation by this triphenolic molecule, along with a comprehensive overview of factors responsible for its high antioxidant activity. Gallic acid derivatives have also been found in a number of phytomedicines with diverse biological and pharmacological activities, including radical scavenging, interfering the cell signaling pathways and apoptosis of cancer cells. The diverse range of applications of this simple polyphenol is due to a fine amalgam between its antioxidant and prooxidant potential. The existing literature on this dual behavior of gallic acid and its derivatives is reviewed here. This is followed by an account of their potential clinical and industrial applications.

Keywords: Antioxidant, prooxidant, free radical, oxidative stress, gallic acid, polyphenol, metal chelator, phytomedicine, preventive therapy, signaling pathways, apoptosis, anticancer.

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1. Introduction

There is a rapidly growing literature on potential applications of antioxidants from natural sources. Naturally occurring antioxidants have the potential to protect cells from oxidative stress *via* a number of pathways. Since oxidative stress is responsible for a variety of degenerative diseases, these antioxidants can contribute significantly towards human welfare.¹ Determining and optimizing the antioxidant properties of a compound is therefore one of the most important areas of research.

Free radical generation, an inevitable and continuous process, is a part of our normal metabolism. Nevertheless, an overproduction and accumulation of reactive oxygen species

(ROS), due to internal and/or external factors, may create an imbalance resulting in oxidative stress. In general, oxidative stress increases the production of superoxide radicals (O_2^{-}) and hydrogen peroxide (H_2O_2). In the presence of a suitable transition metal catalyst, these reactive species can further interact to form highly toxic hydroxyl radicals (HO^{-}). Hence, an excess of H_2O_2 can be detrimental to cells. Under normal circumstances, these free radicals are counteracted by the endogenous antioxidant defense system, involving enzymatic systems such as superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase (GPx), glutathione reductase (GRx) and glutathione-S-transferase (GST).² However, an unchecked rampant free radical production can alter cell viability by disrupting biological macromolecules such as DNA, proteins and lipids. Oxidative damage to such macromolecules accumulates with age and has been postulated as a major type of endogenous damage leading to aging, degenerative diseases, brain dysfunction, atherosclerosis, cardiovascular diseases, inflammatory diseases and cancer.³⁻⁵

A number of synthetic drugs may provide protection against the deleterious effects of oxidative stress but these are also associated with adverse side effects. The focus has thus shifted towards identifying antioxidant compounds that are clinically potent with low or no side effects, to be used in preventive therapies. There is ample support for the claim that a higher intake of fresh fruits, vegetables or plants rich in natural antioxidants is associated with lower incidences of free radical induced diseases such as atherosclerosis, cardiovascular, neurodegenerative diseases and certain types of cancer.⁶⁻¹⁰ Consequently, special attention is being given to antioxidants of natural origin.¹¹⁻¹³

These preventive properties of plants are attributed to the presence of flavonoids, anthocyanins and phenolic compounds.¹⁴ A large number of phenolic compounds possess the ability to scavenge reactive species such as superoxide radicals and hydroxyl radicals; reduce lipid peroxyl radicals, and inhibit lipid peroxidation. These phenolic compounds emerge as strong antiradical agents largely due to their redox properties, which make them efficient hydrogen donors, reducing agents and metal chelators.¹⁵

Among various polyphenols, gallic acid (GA) (3,4,5-trihydroxybenzoic acid), a naturally occurring low molecular weight triphenolic compound, has been suggested to possess strong antioxidant activities in many studies.¹⁶⁻²³ It provides efficient protection against oxidative

damage caused by reactive species often encountered in biological systems including, hydroxyl (HO[•]), superoxide (O₂^{•-}), and peroxyl (ROO[•]) and the non-radicals, hydrogen peroxide (H₂O₂) and hypochlorous acid (HOCl). Furthermore, GA has been demonstrated as the chief antioxidant component responsible for the efficient antiradical and anticancer properties of a number of plant extracts.²⁴⁻²⁸ Similarly, gallic acid derivatives (GADs) have also been found in a number of phytomedicines with diverse biological and pharmacological activities, such as ROS scavenging, interfering the cell signaling pathways, and apoptosis of cancer cells.²⁹⁻³² This latter property *i.e.* induction of apoptosis, is mainly associated with its prooxidant, rather than antioxidant behavior. GA can thus exhibit both antioxidant as well as prooxidant characteristics,³³ displaying a dualedge sword behavior.

There is a diverse variety of bioactive natural molecules possessing the GA moiety, imparting several industrial and pharmacological applications such as being used as a vital building block for various pharmaceutical leads, including Combretastatin A-4, Podophyllotoxin and Colchicine.³⁴⁻³⁶ In analytical research, GA is used as a standard for determining the phenol content of various analytes.³⁷ Further, some of its ester derivatives (propyl gallate, octyl gallate and lauryl gallate) (Fig. 1) are widely used in cosmetics, processed food and food packing materials to prevent oxidative rancidity and spoilage.³⁸

In this review, the emphasis would lie on the dual behavior of gallic acid and its derivatives. The resulting biological and industrial applications would then be discussed, focusing majorly on antitumor activities.

1.1 Bioavailability

Gallic acid is widely present in the plant kingdom and largely found in free form or as a derivative in different food sources such as nuts, tea, grapes and sumac (*Rhus coriaria L.*).³⁹⁻⁴¹ Other sources include gallnuts, oak bark, honey, different berries, pomegranate, mango and other fruits, vegetables and beverages.^{37,42-47} Gallic acid is found in plant tissues in ester form and diverse esters with sugars glycosides, polyols and phenols have been reported.⁴⁸

Apart from the plant species involved, the amount of GA in plant tissues may also get affected due to some external stimuli such as UV radiation, chemical stressors and microbial infections.

On similar lines, the phenolic composition in grape juices and wines gets affected by factors such as grape variety, processing practices, and storage.⁴⁹ In particular, red wine has a high content of this phenolic acid. Burns *et al.* analyzed a number of red wines and found the total phenolic content as 1100 to 3165 mg L⁻¹, out of which 35 to 70 mg L⁻¹ and 120 to 360 mg L⁻¹ is allocated to GA and the epicatechin gallate derivatives, respectively.⁵⁰ GA content is higher in green tea⁵¹ but the total gallate content of cocoa is shown to be even higher than that of green tea or red wine.⁵²

The absorption and metabolism information of polyphenols is useful for the design and interpretation of numerous studies investigating their health effects.⁵³ It is known that among the hundreds of dietary polyphenols which are better absorbed and which lead to the formation of active metabolites, GA is extremely well absorbed, compared with other polyphenols.^{41,54-56} It most likely gets absorbed by the degradation of tannic acid (TA) and gallates in the gastrointestinal tract. Hydrolysable tannins, which are almost invariably present in every food plant, are naturally occurring polymers of galloyl moieties and a glucose molecule. Tannic acid exhibited greater antioxidant activity than α -tocopherol in multiple in vitro assays such as reducing power, DPPH radical, ABTS radical, superoxide anion radical and hydrogen peroxide scavenging and metal chelating activities.⁵⁷ It was thus proposed to be of use in the preservation of the shelf life and nutritional quality of food products. These hydrolysable tannins may get hydrolyzed in the acidic pH of the stomach, with the release of ten potentially reactive GA residues.⁵⁸ The major metabolites of GA are products of methylation (unconjugated and conjugated 4-O-methylgallic acid, 2-O-methylgallic acid), decarboxylation (unconjugated and conjugated pyrogallol, 4-O-methylpyrogallol) and dehydroxylation (resorcinol).⁵⁶ Additionally, some more methyl ether derivatives (3-O-methylgallic acid and 3,4-O-methylgallic acid) have also been reported.⁵⁹ Accordingly, gallates are first hydrolyzed to GA and then methylation vields 4-O-methylgallic acid as a biotransformation product.⁶⁰ Among the various derivatives of GA, the metabolism of ester derivatives shows that the octyl gallate (OG) and lauryl gallate (LG) are absorbed and hydrolyzed to a lesser extent than propyl gallate (PG), which is readily absorbed in the gastrointestinal tract.⁶⁰

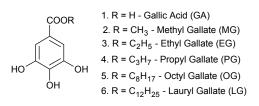


Fig. 1 Chemical structure of gallic acid and some of its *n*-alkyl ester derivatives.

1.2 Biosynthesis

Gallic acid is formed in substantial amounts by plants and fungi.⁶¹ Because of the importance of GA as an antioxidant in foods, controlling its production and accumulation in plants could significantly increase the nutritional value of many crop species.⁶² Thus, a number of studies have been reported regarding the elucidation of the route of biosynthesis of gallic acid.

Multiple studies have postulated the existence of two potential pathways for GA synthesis (Fig. 2). It could either be formed from phenylalanine (4) or from an early shikimate intermediate *i.e.* 3-dehydroshikimate (3-DHS) (2).⁶³⁻⁶⁵ However, the carboxylic group of GA has been found to be biosynthetically equivalent to the carboxylic group of shikimate, rather than to the phenylalanine side chain.⁶⁶⁻⁶⁸ It thus entails that GA is formed from 3-DSH^{61,68,69} either by direct dehydrogenation or *via* protocatechuic acid (3) as an intermediate.^{70,71} Though Werner *et al.*, using retrobiosynthetic NMR studies with ¹³C labeled glucose and oxygen isotope ratio mass spectrometry, showed that GA is primarily synthesized by direct dehydrogenation of 3-DHS.^{61,68}

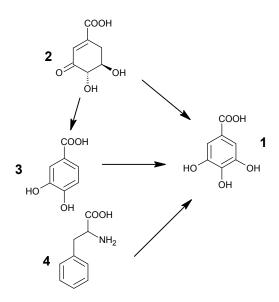


Fig. 2 Possible pathways for biosynthesis of gallic acid (1: GA; 2: 3-dehydroshikimic acid; 3: protocatechuic acid; 4: phenylalanine).

Two different pathways could also be effective in the same organism *i.e.*, *via* early shikimate intermediate and *via* one of the aromatic amino acid.^{64,72,73} It has been asserted that the young leaves of *Rhus succedanea* synthesize gallic acid predominantly by the dehydrogenation of shikimate, whereas the older ones adopt the phenylalanine route.⁷²

Following this, a key enzyme of the shikimate pathway *i.e.* shikimate dehydrogenase (SDH) has been isolated.^{68,69} In the presence of NADPH, SDH catalyzes the reduction of 3-DSH to shikimic acid, while in the presence of NADP+ it catalyzes its oxidation to 3,5-didehydroshikimate, an unstable compound that undergoes a spontaneous rearrangement to gallate.⁶²

Hence, the enzyme (SDH) responsible for the reduction of 3-DHS to shikimic acid, which is primarily used in the formation of aromatic amino acids L-phenylalanine, L-tryptophan and L-tyrosine,⁷⁴ is also required in the biosynthesis of GA.

1.3 Structural facet

Gallic acid is a planar molecule, consisting of an aromatic ring, three phenolic hydroxyl groups and a carboxylic acid group (Fig. 3). The three hydroxyl groups are bonded to the aromatic ring in an *ortho* position with respect to each other. It is this order of arrangement which is the chief determinant for the strong antioxidant capacity of phenolic compounds.⁷⁵ According to quantum chemical calculations with the Becke3P86 DFT functional and the 6-311G(d,p) basis set, the hydroxyl groups are coplanar and in a bent configuration, whereas the carboxylic group turns out of the plane of the benzene ring.⁷⁶ However, structure-activity relationship (SAR) studies on ester derivatives of GA revealed that the most stable conformers exhibit a planar geometry.⁷⁷ This is possibly due to the stabilizing effect of π -electron delocalization between the benzene ring and the carbonyl group, which is favored when they lie in the same plane. Unlike the neutral and the anionic forms, the carboxylate group of the anion radical is slightly turned out of the plane.⁷⁸ Depending on the orientation of the attached carboxylic and hydroxyl groups, GA may have a number of conformations. Different studies involving crystal structure^{38,79,80} and conformational^{81,82} analysis of GA have been reported. Four stable conformers (Fig. 3) were obtained, differing in the orientation of the hydroxyl groups around the ring. GA(I) conformer, having all the three hydrogen atoms of the hydroxyl groups oriented in the same direction, emerged as the most stable conformer.^{81,82}

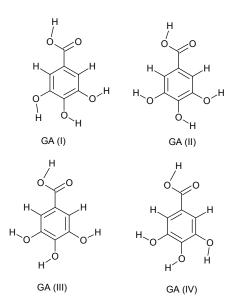


Fig. 3 Conformers of gallic acid.

Gallic acid can form both intra (between hydroxyl groups) as well as intermolecular hydrogen bonds.⁸² GA(I), GA(II), GA(III) and GA(IV) (Fig. 3) can accordingly have two, one, one and two intramolecular hydrogen bonds, respectively. According to a DFT study at the B3LYP/6-311++G(3df,2p) level,⁸³ the hydroxyl groups are involved in two hydrogen bonds (2.196 Å each). The two intramolecular hydrogen bonds between the three hydroxyl groups contribute towards the stability of the molecule.⁸⁴ For neutral GA, the intramolecular hydrogen bonds are of the moderate type (2.1–2.2 Å).⁷⁸ With the formation of the anionic or anion radical structure, the negative charge is mainly redistributed over oxygen atoms, resulting in shorter and stronger hydrogen bonds.

The phenolic aromatic arrangement markedly influences the antioxidant activity of phenolic acids. Accordingly, several factors such as the number and the position of hydroxyl group, the presence of other functional groups and their position with respect to hydroxyl groups have been shown to affect the antioxidant and antiradical activity.

Galato *et al.* have demonstrated, by studying eight phenolic and analogous compounds, that the antioxidant activity of a molecule increases with increase in the number of hydroxyl groups attached to the aromatic ring.⁸⁵ A positive correlation was further obtained between the number of –OH groups attached to the aromatic ring and the antioxidant as well as the antiradical activity of phenolic acids.^{27,75} Thus, gallic acid was found to exhibit the highest antioxidant capacity among various polyphenols.²⁷ On similar lines, all the pyrogallol (1,2,3-trihydroxybenzene) derivatives displayed a higher antioxidant activity compared to the catechol (1,2-dihydroxybenzene) analogues and even with the antioxidant used as reference (tocopherol or trolox).⁸⁶

SAR studies have shown that the radical scavenging efficiency of gallic acid derivatives is dependent on the presence of hydroxyl groups as well as steric freedom.⁸⁷ Especially the *para* substituted –OH group was found to be highly efficient in radical scavenging. Previously, the role of the *ortho* hydroxyl group in the antioxidant activity of phenolic acids has also been described.^{88,89} The extra hydroxyl group in trihydroxy phenolic acid provides greater stability

along with higher antioxidant activity. Hydroxyl groups not only affect antioxidant ability by intramolecular hydrogen bonding (Fig. 4)⁸⁹⁻⁹² but they also tend to stabilize the antioxidant radical formed.⁸⁹ According to Wright *et al.*, the –OH group *ortho* to phenol tends to stabilize the radical formed, resulting in a lower hydrogen bond dissociation enthalpy and hence an increased antioxidant capacity.⁸⁹

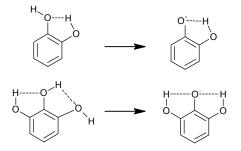


Fig. 4 Intramolecular hydrogen bonding in the catechol (top) and pyrogallol (bottom) moieties and radical stabilization.⁹²

The nature and the position of the substituents with respect to the hydroxyl group also affect the activity of polyphenols. For instance, the easily ionizable carboxylic group contributes toward the efficient hydrogen donation tendency of phenolic acids.⁸³ Gallic acid showed a higher antioxidant activity than pyrogallol,⁸⁸ thus proving a beneficial influence of carboxylate on the antioxidant activity of phenolic acids. However, Sroka and Cisowski did not observe any influence of carboxylate on the antioxidant or antiradical activities of gallic acid.⁷⁵

Thus, utilizing the findings of various studies, we can attribute the high antioxidant activity of gallic acid to a blend of multiple factors, as illustrated in Fig. 5 below.

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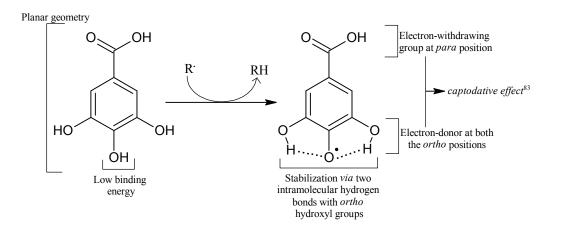


Fig. 5 Factors responsible for an efficient antioxidant activity of the gallic acid molecule.

Multiple studies have obtained experimental and simulated IR spectra of GA.^{76,82,93} The experimental and calculated frequencies differ quite significantly due to strong intermolecular H-bonding (Fig. 6).

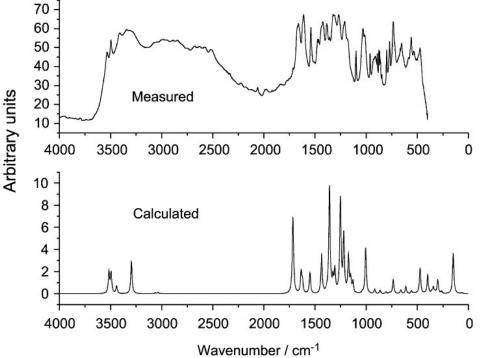


Fig. 6 Experimental (top) and calculated (bottom) IR spectra of gallic acid.⁷⁶

Gallic acid possesses four acidic protons with pK_a values of 4.0 (-COOH group), 8.7, 11.4, and >13 (-OH groups).⁹⁴ The first three pK_a values for gallate free radical, derived from gallic acid, are ~ 4 (-COOH group), 5 and 10 (-OH groups).^{95,96}

The three hydroxyl groups attached to the aromatic ring are prone to oxidation, resulting in the formation of hydrogen peroxide, quinones, and semiquinones.^{96,97} In a spectrophotometric study,⁹⁸ gallic acid and its analogues were shown to be rapidly oxidized by atmospheric oxygen at pH >7. Previously, at a pH range of 2-7, the oxidation was found to be irreversible, and the authors proposed a two-electron oxidation scheme leading to the production of quinoid structures in acidic media.⁹⁹

2. Dual edge sword behavior

2.1 Antioxidant activity

2.1.1 General mode of action

Several studies have been performed to investigate the mechanism followed by phenolic antioxidants.^{1,89,100-103} In a broad sense, phenolic acids can act as antioxidants either by donating a hydrogen atom (Hydrogen Atom Transfer or HAT) (1) or acting as electron donors (Single Electron Transfer or SET) (2).

$$R' + ArOH \rightarrow RH + ArO'$$
(1)

$$R^{\bullet} + ArOH \rightarrow R^{-} + ArOH^{+\bullet} \rightarrow RH + ArO^{\bullet}$$
⁽²⁾

The two specific molecular properties responsible for deciding which mechanism would be most favored are bond dissociation enthalpy (BDE) for the O-H bond and ionization potential (IP). According to Wright *et al.*, both the mechanisms should always occur in parallel, simply with different rates.⁸⁹ Analysis of several phenolic antioxidants using density functional theory suggested the prevalence of HAT mechanism.⁸⁹

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Using DFT calculations, the dependence of the mode of action on the nature of the reacting free radical and the polarity of the environment was demonstrated by considering the involvement of five different mechanisms *viz*. HAT, SET, sequential proton loss electron transfer (SPLET), sequential double proton loss electron transfer (SdPLET) and radical adduct formation (RAF).¹⁰⁴ In aqueous medium, the SPLET and SdPLET prevail for the HO[•] and HOO[•] radical scavenging reactions of gallic acid, respectively. While in non-polar medium, the major mechanism is HAT for HOO[•] radical and both HAT and RAF for HO[•] radical, a prior *in silico* study on propyl gallate supported the dominance of the HAT mechanism, regardless of the polarity of the solvent or the nature of the free radical.¹⁰⁵

Theoretical analysis of four phenolic acids *i.e.* gallic acid (GA), methyl gallate (MG), ethyl gallate (EG), and 4,5-dihydroxy-3-methoxy benzoic acid (OME) yielded the following trend in BDE and IP values: GA > OME > MG > EG (BDE) and GA > MG > EG > OME (IP),¹⁰⁰ suggesting a higher antioxidant potential of EG. However, the conversion to carboxylate (by proton dissociation) results in a significant lowering of the BDE value of GA due to electron donating effect of the carboxylate group,⁹⁴ thus resulting in greater antioxidant efficiency of gallic acid as compared to its ester derivatives.

Radicalization of GA yields gallic acid radical in which the unpaired electron spin density is delocalized over the entire molecule with the largest portion centered on the three carbon atoms bound to the hydroxyl groups.^{78,83} Two radicals, 3-OH (5-OH) and 4-OH, are possible by the abstraction of a hydrogen atom from the corresponding *meta* or *para* hydroxyl groups. The *para* O-H bond is found to be the weakest of all, with the lowest binding energy value.^{83,100,106} It is believed that the presence of an electron withdrawing group (-COOH) at the *para* position and an electron donating group (-OH) at the *ortho* position augments the stability of the free radical by resonance.⁸³ Moreover, the resulting radical (4-OH radical) is stabilized by two hydrogen bonds (2.152 Å) on either sides. On the contrary, DFT calculations at the hybrid HF/B3LYP/6-31G(d,p) level gave almost the same BDE for *meta* and *para* groups.¹⁰⁷

2.1.2 Free radical scavenging activity

Antiradical potential of antioxidants is mainly evaluated by 2.2-diphenyl-l-picrylhydrazyl (DPPH) assay. DPPH is a stable free radical with an absorbance band at 517 nm, which disappears upon reduction (either by accepting an electron or hydrogen radical) by an antiradical compound.¹⁰⁸ Gallic acid was shown to possess the highest antiradical potential, having reduced 6 DPPH radicals per molecule.¹⁰⁹ Abdelwahed et al. demonstrated that GA and 1,2,3,4,6pentagalloylglucose (PGA) adopt a hydrogen donating mechanism to scavenge the DPPH radical and the two are even more effective than Vitamin E.²⁰ According to Nenadis et al. as well, the order of antioxidant activity is: gallic acid > caffeic acid \sim ascorbic acid \sim Trolox > sinapinic acid > isoeugenol.¹¹⁰ Apart from DPPH assay, GA emerged as the strongest antioxidant in other well-known assays as well, such as Trolox equivalent antioxidant capacity (TEAC I-III) assay, Total radical-trapping antioxidant parameter (TRAP) assay, Photochemiluminescence (PCL) assay and Ferric reducing ability of plasma (FRAP) assay. It turned out to be a better antioxidant than popular antioxidants like ascorbic acid, Trolox and uric acid.¹⁹ Multiple computational studies also confirm a better antioxidant efficiency of GA and its derivatives, as compared to caffeine, uric acid, Trolox, sesamol, protocatechuic acid, sinapinic acid, capsaicin, melatonin etc.^{104,105}

At a concentration of 4.17 mM, GA exhibited 43.9% and 60% scavenging effects on DPPH and H_2O_2 radicals, respectively.¹⁰⁸ Moreover, out of various water soluble phenolic acids, the strongest H_2O_2 and DPPH radical scavenging activity was exhibited by gallic acid and pyrogallol.⁷⁵ Although GA exhibited a strong H_2O_2 and HO[•] radical scavenging power,^{108,75} it showed rather no $O_2^{\bullet-}$ scavenging activity.²⁰ Gallic acid and *n*-alkyl gallates were shown to scavenge another reactive species, HOCl, at low concentrations.¹¹¹ Thus, GA is able to protect α_1 -antiproteinase against inactivation by HOCl molecule. It also decreases the peroxidation of ox brain phospholipids.¹¹²

To further support the efficiency of GA in preventing the detrimental effect of hydroxyl radicals, a rate constant of the order of 10¹⁰ M⁻¹ s⁻¹ has been obtained for the diffusion-limited reaction of GA with the HO[•] radical, both in polar and non-polar media.^{104,113,114} Antioxidant capacity of GA and its ester derivatives has also been reported against a number of reactive oxygen and nitrogen species, including peroxyl, azide, and hydroperoxyl radicals.^{104,105,115}

2.1.3 Maintenance of endogenous antioxidant defense system

Another possible mode of action of antioxidants is through their ability to restore the depleted levels of endogenous antioxidant defense system. Antioxidant enzymes such as SOD, CAT, GPx, GRx and GST serve as the primary line of defense against injuries caused by free radicals. Excessive free radical concentration can inhibit the activity of these endogenous antioxidants, leading to oxidative stress. For example, superoxide radicals may temper with SOD and CAT, resulting in their reduced activities whereas GRx inactivation results in excess GSSG (glutathione disulfide) which may in turn inhibit protein synthesis.²² Gallic acid and propyl gallate were shown to elevate the hepatic levels of GPx and CAT, although they had no effect on SOD activity and hence may provide protection against deleterious effects of lipid peroxides and hydrogen peroxide.¹¹⁶ In another study on the effect of D-galactose in senescence accelerated mice (SAM) of different ages, a substantial decrease in the enzyme activities was observed. However, gallic acid treatment restored the activities of CAT and GPx, markedly decreased lipid peroxidation and reduced the amount of malondialdehyde (MDA) in the brain, kidney and liver.¹¹⁷ An increased activity of SOD, CAT, GPx, GRx and GST was also shown by Priscilla and Prince in gallic acid pre-treated ISO (isoproterenol)-injected rats.²² It significantly increased the levels of glutathione (GSH), Vitamin C and E, while it decreased the levels of uric acid in plasma, proving its antioxidant potential against free radicals induced damage.

2.1.4 Prevention of lipid peroxidation

Lipid peroxidation is another route for the propagation of reactive species and their deleterious effects. It is an uncontrolled reaction that elevates the level of lipid hydroperoxides in cellular and subcellular membranes.⁹² These highly reactive cytotoxic species can then disrupt a number of cellular components.

GA and PGA protect peroxidation of lipids, using free radical scavenging activity and lipid peroxidation inhibitory activity. This characteristic has been supported by a study in which the

two compounds scavenge the DPPH radical by a hydrogen donating mechanism and are more effective than Vitamin E itself.²⁰ Even though they exhibit strong inhibition against lipid peroxidation, they do not scavenge O_2^{-} radical. Similar anti-lipid-peroxidative effect with no antiradical potential has been reported for a flavonol compound as well.²⁰

The shelf life of processed food is of prime importance, both for the producer as well as the consumer. It is dramatically affected by the action of oxidative rancidity. Oxidative rancidity involves the autoxidation of fatty acids in the oxygenated environment, releasing off-flavors and undesirable compounds that can alter the sensory and nutritional characteristics of food, proving injuries to health.¹¹⁸ Since the preservation of shelf life as well as nutritional qualities is of vital importance, this deterioration is a major problem. Autoxidation, a slow, radical process, begins *via* a chain reaction with induction, propagation and termination steps. The alkyl radicals, formed in the induction period, start the propagation by reacting with oxygen to form hydroperoxides and peroxide radicals. Ultimately, the association of two radicals results in a stable adduct formation, marking the termination step.¹⁰⁹

The use of antioxidants as food additives can arrest the oxidative deterioration of lipids,¹¹⁸ which in turn may lead to prevention of certain diseases such as cancer, hypertension and vascular diseases.^{8,118} These antioxidants include both natural antioxidants such as Vitamin E, rosemary extracts, flavonoids, as well as synthetic antioxidants such as butylated hydroxytoluene (BHT), butylated hydroxyanisole (BHA) and propyl gallate. However, the adverse side effects of synthetic antioxidant food additives, often used by food manufacturers, raise some health concerns. Thus, limitations over the use of synthetic antioxidants in food and the protective behavior of natural antioxidants together give rise to the emerging interest in using natural antioxidants.^{119,120}

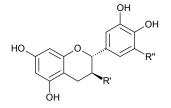
In a study by Jacobsen *et al.*,¹²¹ lipid hydroperoxide level in mayonnaises and effect on flavor in the presence and absence of gallic acid was investigated. As compared to the reference mayonnaises, the amount of free radicals and lipid hydroperoxides is much less in the gallic acid containing mayonnaises. These lower levels of free radicals, along with a rather increased intensity of fishy, rancid, and metallic off-flavor, implies an elevated production of secondary oxidation products due to a rapid decay of lipid hydroperoxides.¹²¹ In a previous study, Halliwell

et al. proposed that because of the ability of GA to reduce metal ions, it is capable of catalyzing the decomposition of lipid hydroperoxide.¹²² The binding of the gallates to the lipid membrane has also been proposed to be the prime factor affecting their antioxidant activity.⁴¹

Zhang *et al.* proposed that solubility, a parameter that affects the mobility of antioxidants between membranes, plays a vital role in determining the antioxidant properties of phenolic acids with respect to lipid peroxidation.⁹⁰ Phenolic acids, being more hydrophilic, are thus less effective than Vitamin E (lipid soluble). At the physiological pH, the hydrophilic carboxylic group is easily ionized, hence phenolic acids are believed to exhibit their antioxidant effect in the aqueous phase.¹²³ But since biomembranes suffer a continuous attack by free radicals generated in the aqueous phase of cellular and subcellular fractions, it is essential to prevent the initial reaction between these aqueous radicals and membrane phospholipids. Thus, the aqueous phase antiradical activity of phenolic acids must be considered as a fruitful method for retarding the peroxidation of membrane phospholipids.⁹²

2.1.5 Gallic acid derivatives

Naturally occurring complex derivatives of gallic acid have attracted much attention due to their ubiquitous nature.⁸⁸ Gallocatechins (Fig. 7), the major constituents of green tea, are persistently emerging as remarkable antioxidant and anticancer agents both *in vivo* and *in vitro*.¹²⁴⁻¹³⁰ Apart from being highly efficient in scavenging biologically damaging oxyl species (superoxide radical and singlet oxygen), these polyphenols have the unique ability to repair the Vitamin E radical.⁹⁵ These antioxidant properties of gallocatechins lay the foundation for the efficient chemoprevention of oxyl radical mediated degenerative diseases.¹³¹⁻¹³⁴



R' = OH; R" = H - Epicatechin R' = OH; R" = OH - Epigallocatechin R' = gallate ester; R" = H - Epicatechin gallate R' = gallate ester; R" = OH - Epigallocatechin gallate

Fig. 7 Naturally occurring derivatives of gallic acid.

The widely used food antioxidant additives *i.e.* methyl gallate (MG), lauryl gallate (LG), and propyl gallate (PG), are manufactured using gallic acid. GA and its derivatives are therefore present in the diet. PG, MG and LG were shown to scavenge hypochlorous acid (HOCl) and trichloromethyl peroxyl radical (CC1₃O₂[•]). All three could also decrease the peroxidation of ox brain phospholipids, better than GA.¹¹²

Gallate esters are efficient scavengers of the DPPH radical both in the ethanol solution and in the liposome.⁸⁷ Along with the importance of the presence of free hydroxyl groups, DPPH scavenging assay suggested that the optimal chain length for gallic acid esters to act as strong antioxidants is four carbon atoms in the aliphatic chain.^{135,136} In another study,⁸⁷ a theoretical analysis was able to explain the activity difference between molecules **1** to **4** (Fig. 8) in terms of O-H bond dissociation energy (BDE), but the lower activity of ester derivatives (MG, PG, OG and LG), in spite of having lower BDEs, could not be explained. Furthermore, Kasture *et al.* demonstrated the poorest ability of triacetoxy gallic acid to scavenge free radicals.¹³⁵

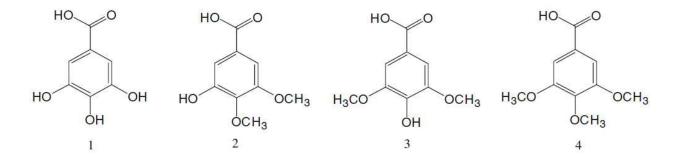


Fig. 8 Structures of gallic acid derivatives.

Even though phenolic acids have been proven to possess strong antiradical and antioxidant activities, a finite therapeutic success of natural antioxidants might be associated with their limited distribution throughout the body and their inherent difficulties to attain the target sites.⁸⁶ Thus, the poor bioavailability and solubility of phenolic compounds in the cell membrane render

their antioxidant properties less evident *in vivo* as compared to *in vitro*.¹³⁷ Hence, increasing the lipophilicity of water soluble antioxidants holds great therapeutic potential.¹¹¹ Lauryl gallate is one such example which exhibits antioxidant activity in the prevention of mitochondrial lipid peroxidation.¹³⁸ Methyl gallate is another naturally occurring antioxidant having anticarcinogenic and antimicrobial properties.

2.1.6 Synergism

A promising solution for a finite therapeutic success of natural antioxidants is taking advantage of the synergy effect. Synergism is obtaining a favorable effect by the use of two antioxidants simultaneously, one reinforcing the effect of the other. A mixture¹³⁹ of two different polyphenols as well as a hybrid^{86,139,140} of the two can have a synergistic effect, resulting in a greater efficiency than individual antioxidants. For instance, the four major phenolic compounds present in 'Ataulfo' mango pulp are chlorogenic, gallic, protocatechuic and vanillic acid. Apart from hydrogen atom donation, these acids augment the overall antioxidant capacity by regenerating other prooxidant phenols *via* electron donation.²⁷ Ester derivatives (**3-5**) (Fig. 9), as well as lipophilic hybrid compounds (**6-8**) of gallic (**1**) and caffeic (**2**) acid were synthesized and analyzed for their antioxidant properties. While esterification had no significant effect on the net antioxidant capacity, the galloyl-cinnamic acid hybrids (**6-8**) emerged as the best antioxidants.⁸⁶

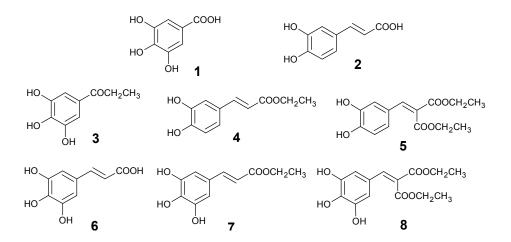


Fig. 9 Structure of phenolic antioxidants derived from gallic acid (1) and caffeic acid (2).⁸⁶

Another hybrid compound (GA-LA) was prepared using gallic acid and linoleic acid (LA).¹⁴¹ GA-LA exhibited a higher tyrosinase inhibition than either of the two parent molecules.¹⁴² It was found to be very effective against human and mouse melanoma and lung cancer cell, much efficient than either GA or LA.

2.2 Prooxidant behavior: Fenton chemistry

The simultaneous presence of H_2O_2 and a transition metal in the reduced form (Fenton type systems) may result in the formation of a strong oxidant, hydroxyl radical, according to the following reaction,

$$Fe^{2+} + H_2O_2 \rightarrow Fe^{3+} + HO^- + HO^{\bullet}$$
(3)

Hydroxyl radical scavenging efficiency can thus be used as an indicator of antioxidant activity.

In general, depending on the stability constants with Fe(II) and Fe(III), the iron chelators may either boost or inhibit the generation of HO[•] radicals.³³ The antioxidant activity of GA is thus considerably influenced by the presence of transition metals. Although it is a free radical scavenger, GA may either inhibit or promote free radical production due to metal chelation.³³ It was shown to exhibit a slight prooxidative effect in the presence of Fe(II) or Fe(III).¹⁴³ In general, several *in vitro* studies have shown that GA promotes HO[•] radical production in iron containing systems, whereas it acts as an HO[•] radical scavenger in the absence of iron.^{33,108,144,145}

A steady state concentration of Fe(II) has been observed in the Fe(III)/ H₂O₂ system as well.¹⁴⁶

$$Fe^{3+} + H_2O_2 \rightarrow Fe^{2+} + HO_2^{\bullet} + H^+$$
(4)

$$Fe^{3+} + HO_2 \cdot \rightarrow Fe^{2+} + O_2 + H^+$$
(5)

Another determinant of the antioxidative activity of GA in Fenton-type systems containing $Fe(III)/H_2O_2$ is GA concentration. At low concentration (GA:Fe < 2:1), prooxidative activity

dominates due to reduction of Fe^{3+} to Fe^{2+} , leading to HO' radical production, whereas, at higher concentrations (GA:Fe > 2:1), the overall effect is antioxidative due to marked scavenging of HO' radicals by GA.^{108,33} The prooxidative action of GA at lower concentration was ascribed to a weak metal-chelating effect and a strong reducing ability. However, the reducing power increases with increasing concentration and hence it inhibits the oxidation process by scavenging the hydroxyl radical.¹⁰⁸

Direct hydroxyl radical scavenging assay also shows a similar trend *i.e.* excess gallic acid with respect to iron results in significant radical scavenging, whereas below GA:Fe::2:1, iron chelation becomes the dominating factor leading to prooxidant behavior.³³

With peroxidases, GA again shows a dual behavior depending on the presence or absence of H_2O_2 . In the absence of H_2O_2 , GA causes cell injury *via* the prooxidative effect, whereas in its presence, GA inhibits the formation of oxidative species in the peroxidase cycle of peroxidases.¹⁴⁷

Although multiple *in vitro* studies have demonstrated the behavior of gallic acid in Fenton-type systems, it is necessary to note that an *in vitro* analysis alone cannot establish an overall *in vivo* response.

3. Chelation with metal ions

Gallic acid and tannic acid affect the bioavailability of certain key minerals such as iron, zinc and calcium, by forming insoluble complexes with them.^{58,148,149} Among the various metal ions present in the body, iron is the most abundant transition metal, with the dietary intake being in the ferric form.¹⁵⁰ Thus, the complex forming tendency of polyphenols may lead to antinutritional effects of iron, causing its deficiency.⁴⁵ This has led to a number of ongoing *in vitro* and *in vivo* studies involving iron biofortification, an approach to counterbalance the inhibitory effect of polyphenols on iron absorption.¹⁵¹⁻¹⁵⁴ In an attempt to distinguish the polyphenols that inhibit iron uptake, Hart and Glahn found that gallic acid is among those

polyphenols that enhance iron uptake in Caco-2 cells.¹⁵⁵ Nevertheless, extensive data is required to fully decipher the net effect of such polyphenols on iron bioavailability.

Accumulation of iron in the pre-frontal cortex and variations in the distribution of ferritin, resulting in enhanced ROS production, evolves into degenerative changes leading to Alzheimer's and Parkinson's disease.^{156,157} Utilizing the chelation potential of hydroxyl groups, polyphenols can inhibit ferrous ion catalyzed DNA damage either by binding with the Fe(II) ion or by stabilizing the Fe(III)-polyphenol complex formed by autooxidation in the presence of oxygen. Hence, the ferrous ion is either unable to catalyze HO' radical generation or is oxidized to ferric which does not promote Fenton reaction. Kinetic study of the autooxidation of Fe(II) complexed with some galloyl containing polyphenols yielded a positive correlation between the rate of oxidation and the antioxidant property of the polyphenol.¹⁵⁸ The rate of autooxidation of gallate-Fe(II) was ten times faster than that of the catecholate analogues and is attributed to a stronger binding affinity of gallates. A high chelating potential of galloyl moiety tends to inhibit the ability of Fe to promote ROS formation through catalysis of hydrogen peroxide decomposition and Fenton type reactions,^{159,160} hence preventing the deleterious effects of oxidative stress. The use of iron chelators, such as gallocatechins, to control the concentration of free iron, can thus help in eliminating the Fenton chemistry contribution to neurodegenerative diseases.¹⁵⁶ Hence, it is of vital importance to study the interaction between iron and GA.

Gallic acid, being a strong chelating agent, forms highly stable complexes with Fe(III).¹⁶¹⁻¹⁶³ It has also been claimed that the complex formation with Fe(III) leads to the oxidation of GA to hydroxyquinone with a simultaneous reduction of Fe(III) to Fe(II).¹⁶⁴ Multiple studies^{84,165} have shown that the complexation process is pH dependent, proceeding from pH = 3 and continuing till pH = 9 with increasing degree of chelation. In a 1:1 complex, iron is bound to the two adjacent hydroxyl groups of the GA molecule. Additionally, GA and its azo derivatives were found to form stable complexes with ratios varying from 1:1 to 1:4.¹⁶³ A prior study stated that at a pH range of 4-6, a 1:3 complex is formed with ferric ions.¹⁶⁶

In the presence of ferrous sulfate, a bathochromic shift in the UV-Vis spectra of gallic acid has been observed.^{167,168} The shift is accompanied by the appearance of a new peak in the visible

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region, indicating the formation of a GA-Fe(II) complex. Kinetic study further indicated the formation of a 1:1 complex following second order kinetics.¹⁶⁸

The complexation of Fe(III) with gallocatechins was reported by Jovanovic *et al.*¹⁵⁶ A 1:2 stoichiometry for Fe(III) complexes with gallocatechins and with methyl gallate, and 1:3 for that of Fe(III) and catechin (Table 1) was observed. Further, the value log K \approx 27 was obtained for the formation of Fe(III)-bis(gallocatechin) at pH 7. Analogously, a molar ratio of 1:2 was shown for Fe(III) complex with gallic acid at pH 7, with the value log K = 34.³³ It is because of the high stability of these complexes that intake of tea may hinder the absorption and bioavailability of iron.¹⁶⁹ However, a net effect on iron absorption is determined by a number of dietary elements such as ascorbic acid, which can reverse the inhibitory effect of polyphenols on heme iron absorption.¹⁷⁰

Complex	п	
Fe (III) (catechin)	3	
Fe (III) (epigallocatechin gallate)	2	
Fe (III) (epigallocatechin)	2	
Fe (III) (epicatechin gallate)	2	
Fe (III) (methyl gallate)	2	
Fe (III) (catechol)	3	

Table 1 Stoichiometric composition of metal complexes of gallocatechins determined by the Job's method at pH 7, $20^{\circ}C^{156}$

Other than Fe, GA forms complexes with various other metal ions too.^{171,172} A complex with neodymium, having a stoichiometry of 1:2, has been obtained as a solid.¹⁷³ Lanthanide complexes with gallic acid have been obtained with the general formula $Ln(C_7H_5O_5)(C_7H_4O_5).nH_2O$ (n = 2 for La-Ho; n = 0 for Er-Lu).⁸⁴ Moreover, GA and its azo derivatives form stable complexes with transition metal ions such as Fe(III), Mn(II), Co(II), Ni(II), Cu(II), Zn(II) and Cd(II).^{84,163} Some of these complexes showed notable antimicrobial activity as well.⁸⁴ Complexation of Cu(II), both with gallic acid and epigallocatechin gallate, has

been demonstrated to be pH dependent with respect to the stoichiometry of the resultant complex.¹⁷⁴ Utilizing the efficient chelating potential of gallic acid, GA-capped nanoassemblies have been successfully prepared and tested for the detection of Pb(II) in a green tea sample.¹⁷⁵

Recently, Nath *et al.* synthesized some potential metal based anticancer drugs *viz.* triorganotin (IV) hydroxycarboxylates and screened them against various cancer cell lines.¹⁷⁶ The gallic acid complexes, *n*-Bu₃Sn(GA) and Ph₃Sn(GA), emerged as the most potent anticancer and anti-inflammatory agents, respectively. The anticancer activity was primarily imparted to ROS generation by triorganotin (IV) hydroxycarboxylates, ultimately leading to apoptosis in cancer cells.¹⁷⁶

The ability to form complexes with transition metals with the advent of intense blue color (with iron) has been utilized in the determination of the gallocatechin content of beverages, like beer and tea.¹⁵⁶ Additionally, gallic acid, because of its blue colored complex with iron, proved to be of prime importance in the production of writing inks (iron gall inks), which primarily contain a mixture of gallic acid, ferrous sulfate and gum arabic.^{167,177} A rich blue color is obtained in the presence of oxygen and an excess of ferrous ions.¹⁷⁸

In water systems, including ground waters, surface waters and municipal wastewaters, high concentrations of iron are often associated with organic matter of natural origin. The humic substances in natural waters retard the oxidation of Fe(II) ions,¹⁷⁹⁻¹⁸¹ due to complexation. In addition, it has been observed that the humic materials stabilize this iron and enhance its nutritive availability for the growth of aquatic species.^{182,183} Similarly, a number of model organic compounds having a similar structural features as these humic substances, can also modify the rate of oxidation of Fe(II) ions.¹⁸¹ In this context, gallic acid and pyrogallol were shown to completely inhibit Fe(II) oxidation, while vanillic acid, vanillin, phenol, resorcinol, synergic acid, and histidine had no effect on its oxidation rate.¹⁸⁴ Especially tannic acid could stabilize high levels of Fe(II) for several days. Furthermore, galloyl moiety containing compounds were able to rapidly reduce Fe(III). Thus, the organic species of natural origin such as tannic acid, gallic acid and pyrogallol are capable of maintaining sufficiently high levels of dissolved Fe(II) concentration under aerobic aquatic conditions, due to the formation of oxidation resistant complex with Fe(II).

However on the downside, the presence of humic substances in groundwater can interrupt the water quality management processes. In order to remove iron from groundwater, water treatment plants employ oxidation of Fe(II) ions followed by sedimentation and filtration of ferric hydroxide formed. The existence of oxidation resistant complexes of Fe(II) can thus obstruct these techniques.¹⁸⁴

4. Potential clinical applications

4.1 As anticancer agent

A number of studies have demonstrated the potential anticancer activity of gallic acid and its derivatives, both *in vivo* and *in vitro*. It is not the antioxidant, but rather the prooxidant action of gallate compounds, which is responsible for their potent anticancer and apoptosis inducing properties. Reactive oxygen species generated by gallates have been asserted to be responsible for the apoptotic and necrotic cell death.^{185,186} There has been ample evidence claiming that gallates induce apoptosis selectively in fast growing tumor cells, leaving the healthy cells intact.^{25,29,108,185-192} This selective cytotoxicity towards cancer cells was proposed to be due to the ability of normal cells to resist GA induced apoptosis by the release of some inhibitors, unlike tumor cells which could not produce sufficient amounts of these inhibitors.¹⁸⁹ Thus, cancer cells are more sensitive towards GA induced apoptosis than normal healthy cells.

Anticancer activity of GA has been reported in a variety of cancer cells, such as, leukemia,^{29,30,32,188,192-197} prostate cancer,^{24,190,198-202} lung cancer,²⁰³⁻²⁰⁷ stomach and colon cancer,^{191,192,23} lymphocyte proliferation,^{31,208} breast, cervical and esophageal cancer.^{187,209}

Several studies have provided evidence that the inhibitory effect of GA on carcinogenesis is mediated through the regulation of multiple signaling pathways.¹⁸⁷ Depending on the cell lines used, GA may induce cell death or cell cycle arrest in different manners.²¹⁰ But still a common mechanism is associated with oxidative stress, derived from reactive oxygen species, mitochondrial dysfunction and an increase in intracellular Ca²⁺ level. Considering that GA induced cell death is mediated by ROS and intracellular Ca²⁺, Isuzugawa *et al.*¹⁸⁹ found that the

ROS generated by GA is hydrogen peroxide, which induced cell death in dRLh-84 (rat hepatoma) cells. Since catalase is responsible for H_2O_2 degradation, the normal cells are insensitive to the mentioned H_2O_2 generation which can be dealt by the catalase enzyme. However, due to lack of proper protection system in tumor cells, they are incapable of defending themselves from the deleterious effects of H_2O_2 resulting in cell death.¹⁸⁹

Fenton chemistry, involving HO' radical production by H₂O₂ and Cu(II), is generally considered to be the cause of oxidative DNA lesion in tumor cells. This copper mediated DNA damage is initiated by the preferential coordination of the Cu(II) ion to N7 of the guanine base of DNA,²¹¹ followed by the reduction of Cu(II) to Cu(I) by gallates.¹⁸⁶ Cu(I) ion then reduces molecular O₂ to superoxide anion, and then to hydrogen peroxide with the formation of Cu(II). The interaction between hydrogen peroxide and DNA bound copper then results in the production of hazardous hydroxyl radicals.²¹² which cause DNA strand breaks and pronounced degradation of guanine bases along with the formation of 8-hydroxy-2'-deoxyguanosine (8-OHdG), a guanine adduct at the C-8 position, as a marker of oxidative DNA damage.¹⁸⁶ Propyl gallate resulted in an increased oxidative DNA lesion in human leukemia cell line HL-60, but not in HP100, which is a hydrogen peroxide resistant cell line derived from HL-60.²¹³ The HP100 cells exhibit 18 times higher catalase activity as compared to that in HL-60 cells.²¹⁴ Apart from Cu(II), GA has also been shown to induce DNA damage in the presence of Fe(III) complexes such as Fe(III)EDTA or Fe(III)ADP.²¹³ Fe(III) complex-mediated DNA damage is also related to Fenton chemistry. Since serum, tissue and cellular copper levels in cancer cells are much elevated in various malignancies, cancer cells are perhaps more subjected to electron transfer between copper ions and polyphenols, resulting in ROS formation.²¹⁵

In addition to ROS generation, these compounds can also induce apoptosis in cancer cells by inhibiting protein tyrosinase kinases (PTKs), one of the proteases vital for normal cell processes. Gallates have been shown to have excellent inhibitory activity against PTKs.²¹⁶ GA derivatives displayed much higher inhibiting activity than widely used PTK inhibitors.²¹⁷ Lauryl gallate for example proved to be a good inhibitor of human spleen PTK.

GA administration could also inhibit metastasis formation and growth.²¹⁸ P-selectin is a protein which is expressed by several metastatic pancreatic tumor cells²¹⁹ and has a role in metastasis by

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promoting tumor growth. P-selectin deficiency is thus accompanied by reduced implanted carcinoma cells growth and reduced metastasis formation.²²⁰ GA has been reported to bind and antagonizing P-selectin under static and dynamic conditions, resulting in beneficial effects in conditions such as coronary artery disease, thrombosis, and cancer. Moreover, the required concentration is easily attainable by moderate wine, green tea, or cocoa consumption.²¹⁹

The dependence of the action of gallic acid on cell cycle has been argued upon in numerous studies. Sakaguchi *et al.*,³⁰ using flow cytometric analysis and agarose gel electrophoresis, observed an inter nucleosomal breakdown of chromatin DNA in HL-60RG cells but not in dRLh-84, HeLa, and PLC/PRF/5 cells; and the mechanism was found to be independent of the cell cycle. Whereas in colon adenocarcinoma cells,¹⁹¹ GA was found to hinder with the G2/M phase. Similarly in human bladder carcinoma cells, GA prompted G2/M cell cycle arrest involving molecular alterations in cell cycle regulatory proteins.²⁰⁵ On the contrary, in the studies by Inoue *et al.*,¹⁸⁵ Faried *et al.*¹⁸⁷ and Hsu *et al.*,²²¹ no such phenomena was observed. Although GA had no effect on the cell cycle profile, it increased the number of apoptotic cells in a time dependent manner.¹⁸⁷ According to still another study,¹⁹⁵ GA induces cell cycle perturbation in the G1 phase. Meanwhile, multiple studies^{205,222,223} have demonstrated the effect of GA on cancer cell proliferation by arresting cells at the G2/M phase. GA downregulated the activities of cyclin B1, Cdc2 and Cdc25C, which are essential for G2/M transition. Thus, it is being increasing implied that GA can block proliferation of cancer cells in culture *via* apoptosis and/or cell cycle arrest.^{24,198}

GA inhibits cell growth of prostate cancer (PCa) cell lines. Highly significant cytotoxicity was shown in PCa DU145 cells by the activation of pre-existing apoptotic processes and cell cycle arrest machinery.^{199,202} With respect to its potency against prostate cancer, both anticancer and cancer chemopreventive effects in human PCa DU145 cells in culture and TRAMP model, respectively, have been demonstrated.^{198,201}

GA dose dependently decreased the growth and induced cell death in Calu-6 and A549 lung cancer cells.²⁰⁶ Lung cancer cells were much more susceptible to growth inhibition by gallic acid as compared to normal human pulmonary fibroblast (HPF) cells. GA-induced lung cancer cell death has been related to ROS increase as well as GSH depletion.^{206,207} Similarly, GA-induced

HeLa cell death was accompanied by a significant increase in ROS and GSH depletion.²⁰⁹ Introduction of gallic acid into human colon adenocarcinoma COLO 205 cells and stomach cancer KATO III cells resulted in growth inhibition and induction of apoptosis.¹⁹²

Gallic acid-based indanone derivatives have also been shown to exhibit substantial anticancer activity against various human cancer cell lines, such as KB403 (oral and mouth cancer cells), WRL68 (liver cancer cells), Caco2 (colon cancer cells), HepG2 (liver cells) and MCF7 (hormone dependent breast cancer cells).³⁶

Besides, GA is one of the major phenolic acids present in grape seed extract (GSE). Due to the strong antioxidant activity shown by GSE, it is considered to be highly beneficial for the treatment of various diseased conditions.^{28,24,225} A number of *in vivo* and *in vitro* models have been utilized by numerous studies in order to demonstrate the anticancer effects of GSE.^{190,201,226-235} Being one of the main phenolic compounds in GSE, gallic acid may be a major contributory factor responsible for these therapeutically beneficial activities. Accordingly, Veluri *et al.* identified gallic acid as one of the most active GSE constituents contributing significantly in growth inhibition, death and apoptosis in human PCa DU145 cells.²⁴ Following this, Agarwal *et al.* further investigated the efficacy and associated mechanism of GA.¹⁹⁸ Gallic acid treatment resulted in a strong cell growth inhibition, cell cycle arrest, and apoptotic death in DU145 cells. Caspases are a group of proteases which are responsible for programmed cell death. Activation of caspases is thus associated with the process of apoptosis.²³⁶ Both caspase-dependent and caspase-independent pathways have been suggested for the antitumor activity of GSE derived gallic acid.^{24,198,237}

Furthermore, GA purified from pomegranate peel extract, was found to induce apoptosis in the human alveolar epithelial A549 cell line, through the elevation of ROS and by the activation of Caspase 9.²³⁸

GA is also abundantly present in *Toona sinensis*, one of the most popular vegetarian cuisines in Taiwan.²³⁹ It has been demonstrated that *T. sinensis* exerts antiproliferative effects on cultured human HL-60 cells *in vitro* and *in vivo* through the induction of cell-cycle arrest and apoptosis. Later on, using the *in vivo* bioluminescent imaging technique, Hsiang *et al.* affirmed the anti-

inflammatory potential of both *T. sinensis* and GA in mice.²⁴⁰ Such potent biological activities of *T. sinensis* have been attributed to the abundance of gallic acid which acts as its chief bioactive molecule.^{25,199,239,240}

4.2 Other therapeutically beneficial activities

Gallic acid has been recognized to display diverse therapeutically beneficial activities. It has been shown to possess antifungal,²⁴¹⁻²⁴⁴ antibacterial,^{244,245} antiviral,²⁴⁶⁻²⁵⁰ antiallergic,²⁵¹ antiinflammatory,^{252,253} antimelanogenic,^{21,254-256} antitubercular,²⁵⁷⁻²⁵⁹ antimutagenic,²⁰ antiulcer,^{196,260} anticholesterol,²⁶¹ antiobesity²⁶² and immunomodulatory activities.²⁶³ Additionally, it displays neuroprotective,^{87,135,264-269} cardioprotective,²² hepatoprotective²⁷⁰⁻²⁷² and nephroprotective²⁷²⁻²⁷⁴ potential.

Anti-inflammatory

Myeloperoxidase (MO), a key enzyme in infection and inflammation processes, converts hydrogen peroxide and chloride to HOCl. It is capable of reducing H_2O_2 to water and oxygen, but a small portion is also utilized in the production of tyrosyl radicals.¹¹¹ Phenolic acids, including gallates, exhibit a strong MO inhibition.^{111,275} Gallic acid and its *n*-alkyl ester derivatives, differing in lipophilicity, demonstrated anti-inflammatory activity by two mechanisms: (1) Strong inhibition of MO activity, hence directly inhibiting the production of HOCl, and (2) scavenging the hazardous reactive species produced by the enzyme.¹¹¹ Although the former mechanism is influenced by the lipophilicity of gallates (inhibition increased by 50% with increase in lipophilicity), the latter one remains unaffected.¹¹¹

Antimicrobial

Numerous studies have shown the antibacterial and antifungal potential of gallates.^{276,277} For instance GA showed antifungal activity against *F. fusiformis, Fusarium semitectum* and *Alternaria altternata*.²⁴² Díaz-Gómez *et al*.²⁷⁸ demonstrated the antimicrobial effect of GA on *Helicobacter pylori*, a gram negative bacterium which is one of the leading causes of gastric cancer. It was found to significantly inhibit the growth of *H. pylori*. Derivatives of GA have also

been shown to display fungicidal and bactericidal properties.^{241,244,245} Oxadiazole derivatives for example are known to possess antimicrobial,^{257,279} antiparasitic²⁸⁰ and antitubercular activities.²⁵⁷⁻²⁵⁹

Antimelanogenic

The overproduction of melanin, the chief determinant of skin color, leads to various dermatological disorders.^{281,282} Potential of GA as a skin protecting agent has been demonstrated by studying its antimelanogenic and antityrosinase activities. GA has been shown to act against hyperpigmentation activity *via* the inhibition of tyrosinase activity. It is a much stronger inhibitor of tyrosinase than kojic acid, a standard antityrosinase agent.²¹ Meanwhile, Panich *et al.* demonstrated the protective effect of GA against UVA mediated melanogenesis.²⁵⁶

Antiviral

GA²⁴⁶ and MG²⁴⁷ showed inhibition of Human Immunodeficiency Virus Type 1 (HIV-1) and Herpes Simplex Virus Type 1 and 2 (HSV) replication, respectively. Moreover, octyl gallate exhibited marked anti-HSV-1 activity and inhibitory effect against RNA viruses.^{248,249} Kratz *et al.* investigated the inhibitory effect of gallic acid and its *n*-alkyl derivatives on HSV-1 and HIV-1 replication.²⁵⁰ GA and PG emerged as the most active compounds against HSV-1 replication and an anti-HIV-1 activity was also found. Additionally, an antiviral activity of GA against human rhinoviruses has also been reported.²⁸³

Antiallergic

Gallic acid has been shown to act as an antiallergic agent by blocking the release of histamine, which would otherwise result in immediate hypersensitivity.²⁵¹

Neuroprotective

Excess ROS production due to the oxidation of dopamine can result in neurodegerative diseases such as Parkinson's and Alzheimer's disease.²⁸⁴⁻²⁸⁶ GA and its derivatives provide neuroprotection from this oxidative stress.^{87,100,135,286} The ability of gallates to scavenge DPPH in liposomes along with molecular polarity accounts for their neuroprotective effects.⁸⁷ Thus, apart

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from the antioxidant capacity, the *in vivo* neuroprotection also depends on hydrophobicity. Hydrophobicity is an essential parameter as the compounds need to be transported to the reaction site effectively. It is due to this latter factor that MG, PG and BG turn out to be more active as anti-Parkinson's agents than the parent compound (GA) itself.¹³⁵ Hence a balance between antioxidant ability and hydrophobicity can lead to effective neuroprotection. Further, Nabavi *et al.* demonstrated their effective neuroprotection against sodium fluoride induced oxidative stress in rat brain, possibly by the inhibition of initiation and propagation of lipid peroxidation.²⁶⁴

Nephroprotective and hepatoprotective

GA exhibited nephroprotective behavior against lindane, ferric nitriloacetic acid and sodium fluoride induced nephrotoxicity.^{272,274} Intoxication can deplete the levels of our antioxidant defense system, resulting in cell damage *via* oxidative stress. Several studies have demonstrated the ability of GA to restore the activity of these endogenous antioxidants.^{271-273,287,288} Additionally, GA isolated from *Peltiphyllum peltatum* showed protective effect against sodium fluoride induced nephrotoxicity, hepatotoxicity and oxidative stress.^{271,273} Pre-treatment with gallic acid neutralized the deleterious effects of fluoride intoxication possibly by its nitric oxide and hydroxyl radical scavenging activity^{33,289} as well as by the ability to restore the depleted levels of endogenous antioxidants.

Miscellaneous

Squalene epoxidase (SE), an enzyme responsible for the catalysis of a rate limiting step in cholesterol biosynthesis, is a potential target for cholesterol lowering drugs.²⁶¹ The inhibitory potential of naturally occurring esters of gallic acid against SE has been reported.²⁹⁰ Abe *et al.*²⁶¹ further evaluated the analogous inhibitory activities of *n*-alkyl esters of GA, showing lauryl ester as a much more potent inhibitor of SE than its natural derivatives. GA has also been shown to protect pancreas from streptozotocin induced toxicity *via* its antihyperglycemic, anti-lipid-peroxidative and antioxidant effects.²⁹¹ It promotes hyperglycemia by elevating insulin secretion. Recently, Abdelwahab²⁶⁰ presented the inhibitory role of GA and its novel derivatives on ethanol induced gastric lesions in rats. The prime mode of action proposed for the antiulcer property

includes, effects on gastric acid secretion, promotion of mucosal protection by endogenous factors and inhibition of oxidative stress-induced apoptosis.

5. Industrial applications

GA and its esters have a diverse range of applications in industrial uses as well, such as additives in food and in cosmetics. Its ester derivatives (PG, OG and LG) are widely used in cosmetics, processed food and food packing materials to prevent oxidative rancidity and spoilage.³⁸

GA is extensively used in tanning as well as in the manufacturing of paper.²⁹² It is used for the stabilization of collagen in the process of leather making.²⁹³ Additionally, it is employed as a source material for inks, paints, color developers and pharmaceuticals.¹⁷⁷ The iron gall inks were one of the key writing materials used in the preparation of documents, drawings, and other written materials.^{294,295}

In analytical research, gallic acid is being persistently used as a standard antioxidant in order to determine the phenolic content of plant extracts or any other analyte and the results are expressed in Gallic Acid Equivalents (GAE).^{37,296,297} GA can also act as a non-toxic corrosion inhibitor by adsorbing on the metal surfaces. It has been suggested that it can cover the steel surface by forming an insoluble complex with ferric ions, thus inhibiting corrosion.²⁹⁸ Additionally, propyl gallate has been shown to be an effective stabilizing agent for the protection of biodiesel oxidation.²⁹⁹

6. Conclusions and perspective

Considering the increasing occurrence of oxidative stress mediated degenerative diseases, there is a pressing need to boost the body's defense system with minimal side effects. This latter requirement has popularized the use of naturally occurring compounds as potent antioxidants. In this regard, hydroxybenzoic acid derivatives have shown favorable activities. Among them, gallic acid displays the highest antioxidant potential and wide bioavailability. Moreover, the

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antioxidant potential can be further improved by forming a hybrid between gallic acid and another phenolic antioxidant like caffeic acid. Gallic acid and its derivatives have been gaining attention, not only because of their antiradical, but also due to their selective apoptosis inducing potential. However, it is the prooxidant aspect of this molecule that gives rise to this latter effect. The feasibility of both anti- and pro-oxidant behavior gives rise to a diverse range of biological activities, including antitumor, antimicrobial, antimelanogenic and anticholesterol. The manifold properties of gallic acid and its derivatives discussed in this review make it a highly promising therapeutic agent in preventive therapies. However, a thorough mechanistic investigation at the molecular level is of prime importance in order to overcome the limited clinical success of natural compounds.

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