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Electrostatic [FeFe]-hydrogenase–carbon nitride assemblies for efficient solar hydrogen production†

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The assembly of semiconductors as light absorbers and enzymes as redox catalysts offers a promising approach for sustainable chemical synthesis driven by light. However, achieving the rational design of such semi-artificial systems requires a comprehensive understanding of the abiotic–biotic interface, which poses significant challenges. In this study, we demonstrate an electrostatic interaction strategy to interface negatively charged cyanamide modified graphitic carbon nitride (^{NCN}CN_x) with an [FeFe]-hydrogenase possessing a positive surface charge around the distal FeS cluster responsible for electron uptake into the enzyme. The strong electrostatic attraction enables efficient solar hydrogen (H₂) production *via* direct interfacial electron transfer (DET), achieving a turnover frequency (TOF) of 18 669 h⁻¹ (4 h) and a turnover number (TON) of 198 125 (24 h). Interfacial characterizations, including quartz crystal microbalance (QCM), photoelectrochemical impedance spectroscopy (PEIS), intensity-modulated photovoltage spectroscopy (IMVS), and transient photocurrent spectroscopy (TPC) have been conducted on the semi-artificial carbon nitride–enzyme system to provide a comprehensive understanding for the future development of photocatalytic hybrid assemblies.

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Introduction

Converting solar energy into clean chemical fuels, such as molecular hydrogen (H₂), holds promise for advancing the concept of a circular economy.¹ Among various photocatalysts, carbon nitride (CN_x) has emerged as a particularly attractive candidate due to its unique advantages, including visible light absorption, cost-effective fabrication, scalability, and low toxicity.² To further enhance the photocatalytic performance of CN_x, significant efforts have been devoted to chemical modifications and the incorporation of co-catalysts.³ The introduction of ionic cyanamide functional groups into CN_x (^{NCN}CN_x) has demonstrated substantial improvements in charge separation and photocatalytic activity, attributed to the prolonged lifetimes of photogenerated electrons.^{4,5} Moreover, the negatively

charged cyanamide group provides a versatile platform for potential electrostatic interactions with co-catalysts.

Nature has evolved enzymes as highly specific biological catalysts to facilitate essential processes in living organisms. Among these enzymes, hydrogenases (H₂ases) stand out for their remarkable ability to catalyze the interconversion of protons and H₂ with high efficiency at near-zero overpotential under mild conditions,⁶ surpassing the capabilities of synthetic catalysts.⁷ H₂ases can be classified into three main types based on their metal cofactors: [NiFe]-H₂ase, [FeFe]-H₂ase, and [Fe]-H₂ase, with [FeFe]-H₂ase being generally the most active for the hydrogen evolution reaction (HER).⁶ The extensive investigation of H₂ases as model biocatalysts⁸ has not only inspired the design of artificial systems such as synthetic Fe₂S₂(CO)₆ catalysts that mimic the active site of the Fe₂S₂ subunit of the [FeFe]-H₂ase (Fig. S1†),^{9,10} but also paves the way for developing bio-hybrid assemblies in semi-artificial photosynthesis systems.^{11,12}

By interfacing CN_x with H₂ase, we combine the strengths of both artificial and biological approaches, resulting in unique properties that neither system can achieve individually.^{11,12} This integration opens up new avenues for exploring synergistic effects and unlocking unprecedented possibilities in solar energy conversion and catalysis. The activation of [FeFe]-H₂ase by light can be considered as a model for the development of efficient bio-hybrid systems. Such a photocatalytic system has thus far been demonstrated using either toxic and expensive CdTe nanocrystals¹³ or carbon dots with a low turnover number

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(TON) of 20 000 over 24 h.¹⁴ Direct electron transfer (DET) between graphitic carbon nitride (g-C₃N₄) and [NiFeSe]-H₂ase has been established with non-specific interactions, resulting in a turnover frequency (TOF) of 4117 h⁻¹ over 4 h.¹⁵ Subsequent improvements involved the incorporation of a non-diffusional electron mediator, TiO₂, between g-C₃N₄ and [NiFeSe]-H₂ase, leading to an enhanced TON (4 h) of 80 000.¹⁶ In addition to its application in H₂ase systems, CN_x has predominantly been utilized for the regeneration of NADH in mediated electron transfer (MET) processes involving formate dehydrogenase¹⁷ or alcohol dehydrogenase.¹⁸

In this work, we present an approach for biological integration with CN_x by demonstrating the electrostatic interaction with enzymes to form a functional biohybrid assembly. This method establishes a benchmark for solar H₂ production, complemented by comprehensive interfacial characterizations utilizing a quartz crystal microbalance (QCM), photoelectrochemical impedance spectroscopy (PEIS), intensity-modulated photovoltage spectroscopy (IMVS), and transient photocurrent spectroscopy (TPC). Specifically, we coupled negatively charged ^{NCN}CN_x with H₂ases containing different surface charges for *in vitro* photocatalytic H₂ production without an external electron relay. The adsorption process of H₂ases on ^{NCN}CN_x is quantified by QCM, whereas PEIS provides insights into the charge carrier dynamics at the biomaterial interface.

Results and discussion

The ^{NCN}CN_x photocatalysts were synthesized using melamine and potassium thiocyanate following previously published methods.^{4,19} Detailed synthesis procedures and characterizations, including scanning electron microscopy (SEM), attenuated total reflectance Fourier-transform infrared (ATR-FTIR) spectroscopy, fluorescence spectroscopy, and ultraviolet-visible spectroscopy are provided in the ESI (Fig. S2–S5).[†] [FeFe]-H₂ase from *Clostridium pasteurianum* (CpI, heterologously produced in *Escherichia coli*) and [NiFeSe]-H₂ase from *Desulfovibrio vulgaris* Hildenborough (DvH) were expressed and purified under anaerobic conditions.^{20,21}



Fig. 1 Schematic of photocatalytic H₂ evolution coupled with alcohol oxidation to aldehyde using an electrostatic ^{NCN}CN_x||[FeFe]-H₂ase (CpI, PDB: 4XDC) assembly. Scale bar refers to enzyme and CN_x is not shown to scale.

Fig. 1 illustrates the hypothesis that the negatively charged ^{NCN}CN_x possesses the ability to activate enzymes with positively charged electron entry points, such as CpI [FeFe]-H₂ase.²² Additionally, the unique property of ^{NCN}CN_x in converting alcohols selectively into aldehydes offers an opportunity to monitor the clean oxidation reaction of 4-methylbenzyl alcohol (4-MBA) to 4-methylbenzaldehyde (*p*-tolualdehyde), allowing the quantification of stoichiometry from the products resulting from oxidation and reduction.^{5,23}

To construct a photocatalytic system for solar H₂ production coupled with selective alcohol oxidation, we interfaced [FeFe]-H₂ase with ^{NCN}CN_x in the presence of 50 mM 4-MBA in 1 mL aqueous MOPS buffer solution (0.1 M, pH 7). To determine the surface charge of ^{NCN}CN_x, zeta potential measurements were performed. Notably, the presence of light-induced blue radicals (absorption band from 500–750 nm, Fig. S5[†])^{5,24} did not impact the surface charge of ^{NCN}CN_x (Fig. S6[†]), indicating that these radicals are long-lived and deeply trapped photoelectrons.^{23,25} The negatively charged surface is primarily attributed to the cyanamide group, which maintains a negative zeta potential even at a pH below 2 (Fig. S7[†]). CpI [FeFe]-H₂ase was selected due to its distal FeS cluster ([4Fe-4S]) being surrounded by a positively charged region containing surface arginine and lysine residues. *In vivo*, this distal FeS cluster region is thought to interact with the negatively charged region of ferredoxin for electron transfer.²² By constructing electrostatic ^{NCN}CN_x||[FeFe]-H₂ase assemblies, we achieve efficient solar H₂ production, with ^{NCN}CN_x mimicking the role of ferredoxin to deliver electrons directly into [FeFe]-H₂ase (Fig. 1).

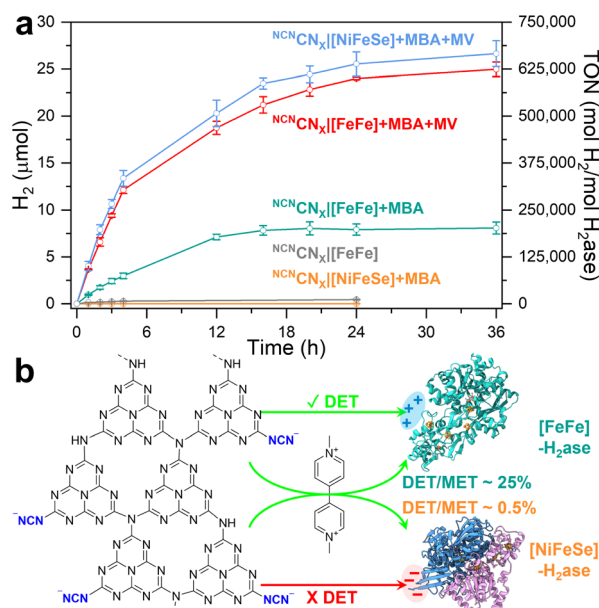


Fig. 2 (a) Time-dependent photocatalytic H₂ evolution of ^{NCN}CN_x with H₂ases for direct electron transfer (DET) and mediated electron transfer (MET). (b) Schematic of electron transfer process. Conditions: 1 mL anaerobic MOPS buffer (0.1 M, pH 7) containing 50 mM 4-MBA, 2 mg ^{NCN}CN_x, 40 pmol H₂ase (either CpI [FeFe] (PDB: 4XDC) or DvH [NiFeSe] (PDB: 5JSH)), AM 1.5G irradiation, 600 rpm stirring, 25 °C. For MET experiments, 2 mM methyl viologen (MV) was used. Error bars represent the standard deviation for a sample size of 3.



Time-dependent photocatalytic H₂ evolution using ^{NCN}CN_x[[FeFe]-H₂ase complexes is illustrated in Fig. 2a. The TON is determined by the ratio between the number of moles of product (H₂) and the number of moles of catalyst (H₂ase) and the TOF is calculated by the TON per hour. Notably, a nearly linear increase in H₂ yield is observed during the initial 4 h, reaching 3.0 ± 0.3 μmol with a TOF of 18 669 h⁻¹. This TOF value is approximately 4.5 times higher than the previous benchmark (4117 h⁻¹)¹⁵ and is even comparable to systems utilizing MET such as g-C₃N₄/TiO₂ (TOF = 20 000 h⁻¹)¹⁶ (note that previous systems used *Desulfomicrobium baculatum* (*Dmb*) [NiFeSe]-H₂ases). Continuous irradiation of the ^{NCN}CN_x[[FeFe]-H₂ase assemblies for 24 h yielded 7.9 ± 0.6 μmol of H₂ (TON = 198 125). The efficient DET between ^{NCN}CN_x and [FeFe]-H₂ase can be attributed to the specific electrostatic interaction at the interface, which will be further evaluated by QCM and PEIS. Furthermore, 4-MBA was selectively oxidized to *p*-tolualdehyde (Fig. S8–S10),[†] with a H₂:*p*-tolualdehyde ratio of 0.77. The observed ratio indicates the deep trapping of some photo-generated electrons within the CN_x polymeric structure in addition to some buffer (MOPS) oxidation (see below).

Recent transient spectroscopic and electron paramagnetic resonance (EPR) studies extensively characterized the deep traps stored in ^{NCN}CN_x on the time scale from ps to s.^{23–26} EPR analysis showed that these long-lived and deeply trapped photoelectrons emerged as blue radicals, processing a symmetric Gaussian line near the free electron *g* value at X-band frequency of ~9.6 GHz.²⁴ These blue radicals can also be visualized by UV-vis spectroscopy (Fig. S5[†]) and qualitatively by the eye (Fig. S11[†]). The oxidation ability of ^{NCN}CN_x has been further evaluated using glycerol (a waste product from the biodiesel industry) as the reductant on a model ^{NCN}CN_x/Pt (2 wt%) system. The oxidation products are quantified as glyceraldehyde at 134.2 ± 5.7 μmol h⁻¹ g⁻¹ and dihydroxyacetone at 54.9 ± 6.8 μmol h⁻¹ g⁻¹ (Fig. S12–S15)[†] and the H₂ yield is 94 ± 9 μmol h⁻¹ g⁻¹. Replacing 4-MBA with ethylenediaminetetraacetic acid (EDTA) led to a ~16% enhancement of H₂ yield (4 h) with ^{NCN}CN_x[[FeFe]-H₂ase assemblies (Table S1[†]). This indicates that, under DET conditions, the rate of 4-MBA oxidation is limiting compared to HER. Despite this observed higher activity, it is worth noting that EDTA is considered as a sacrificial electron donor, and its oxidation results in a range of products that cannot be easily characterized. As a result, the primary focus of this study is the conversion of 4-MBA to *p*-tolualdehyde, serving as a model reaction for the selective oxidation of alcohols to aldehydes.

Exclusion controls were conducted by removing individual components from the photocatalytic system (Table S1[†]). As depicted in Fig. 2a and S16,[†] the system exhibited significantly reduced efficiency in the absence of any component, with H₂ yields below 0.3 μmol over 4 h. Minor H₂ evolution activity was observed with ^{NCN}CN_x[[FeFe]-H₂ase assemblies even in the absence of 4-MBA, yielding 285 ± 24 nmol H₂ in 4 h. This observation suggests that MOPS serves as a much less efficient electron donor in the photocatalytic reaction. NMR analysis (Fig. S17–S20[†]) provides evidence of MOPS oxidation during photocatalysis. However, it is noteworthy that alcohol oxidation

on ^{NCN}CN_x is so efficient and selective that the photocatalytic activity of MOPS oxidation is only ~5% compared to 4-MBA oxidation in 24 h (Fig. 2a). Upon replacing MOPS buffer with pH 7 phosphate buffer, the reaction is drastically reduced in the absence of 4-MBA, accompanied by a ~27% decrease in photocatalytic activity in the presence of 4-MBA (Fig. S21, Table S2[†]). This observation is in line with literature that MOPS as a standard Good's buffer can maintain high *in vitro* biochemical and biological activities.²⁷ Notably, control experiments were also performed to validate the proposed electrostatic interactions using a DvH [NiFeSe]-H₂ase, differing from the previously reported *Dmb* [NiFeSe]-H₂ase.^{15,16} The distal FeS cluster of both [NiFeSe]-H₂ases near its surface is surrounded by amino acids that lead to a local negative charge, serving as the electron entry point for interaction with the positively charged heme of cytochrome *c*₃ during electron transfer *in vivo*.²⁸ Interfacing DvH [NiFeSe]-H₂ase with ^{NCN}CN_x resulted in the production of 8.6 ± 0.33 nmol of H₂ in 4 h, with a significantly lower TON of 215 (Fig. 2a). These results indicate that electrostatic repulsion prevents DET in this system. A detailed comparison among state-of-the-art photocatalytic systems combining carbonaceous photocatalysts and H₂ase are listed in Table 1.

To determine the charge transfer efficiency of DET, methyl viologen (MV) as a soluble electron mediator is used to activate MET (Fig. 2a). Note that the presence of MV may suppress DET due to the kinetic and thermodynamic favorable one electron reduction of MV molecules to MV radicals.^{25,30} Upon the addition of MV (2 mM), the H₂ yield reaches 24.0 ± 0.1 μmol for *CpI* [FeFe]-H₂ase and 25.6 ± 1.3 μmol for DvH [NiFeSe]-H₂ase after 24 h of irradiation. The comparable H₂ yields, despite differences in specific activity, indicate that the rate limiting step during MET is not enzyme turnover but the photoactivity of ^{NCN}CN_x, as evidenced by control experiments (Table S1[†]). Specifically, by doubling the H₂ase loading from 40 pmol to 80 pmol, no significant changes in H₂ yields were observed over

Table 1 State-of-the-art photocatalytic systems combining carbonaceous light absorbers and H₂ases

Photocatalytic system (DET)	TOF ^a (h ⁻¹)	TON ^b	AQE	Ref.
^{NCN} CN _x [[FeFe] + MBA ^d	18 669	198 125	0.35%	This work
g-C ₃ N ₄ [[NiFeSe] + EDTA ^d	4117	36 000	0.07%	15
CDs[[NiFeSe] + EDTA ^d	3125	44 000	0.36%	29
CDs[[FeFe] + TEOA ^e	1500	19 000	1.7%	14
Photocatalytic system (MET)	TOF ^a (h ⁻¹)	TON ^b	AQE	Ref.
^{NCN} CN _x [[FeFe] + MBA + MV ^d	75 769	600 350	1.4%	This work
^{NCN} CN _x [[NiFeSe] + MBA + MV ^d	83 588	638 825	N.A.	This work
g-C ₃ N ₄ /TiO ₂ [[NiFeSe] + EDTA ^{c,d}	20 000	275 000	0.51%	16
CDs[[FeFe] + TEOA + MV ^e	2000	32 000	N.A.	14

^a TOF is calculated from 4 h photocatalysis. ^b TON is calculated from 24 h photocatalysis. ^c TiO₂ as a non-diffusional electron mediator. Illumination conditions. ^d AM 1.5G, 100 mW cm⁻², Xe lamp. ^e 50 mW cm⁻², LED lamp.



both 4 hour and 24 hour periods under MET conditions. The efficiency of DET is qualitatively determined by the DET:MET ratio, defined by the ratio of H₂ yield in the absence of MV (DET) and in the presence of MV (MET). In the case of [FeFe]-H₂ase and [NiFeSe]-H₂ase, DET:MET ratios of approximately 25% and 0.5% are observed, respectively (Fig. S22†). By comparing the results obtained from DET and MET, a schematic representation can be depicted in Fig. 2b. It highlights the establishment of efficient electron transfer directly between ^{NCN}CN_x and [FeFe]-H₂ase, with a DET:MET ratio of 25%. This finding emphasizes the effectiveness and benefits of electrostatic interactions in facilitating DET. Conversely, the electrostatic repulsion between [NiFeSe]-H₂ase and ^{NCN}CN_x prevents DET, resulting in a low DET:MET ratio of 0.5%. Notably, the apparent quantum efficiency (AQE) measured at 450 nm with ^{NCN}CN_x/[FeFe]-H₂ase assemblies under DET and MET conditions are 0.35% and 1.4% (Table S3†), respectively. In contrast, a model ^{NCN}CN_x/Pt (2 wt%) system yields 0.28 ± 0.05 mmol h⁻¹ g⁻¹ H₂ and 0.51 ± 0.09 mmol h⁻¹ g⁻¹ *p*-tolualdehyde with an AQE of 1.92% at 450 nm. In terms of the overall stability of the designed systems, the DET system exhibited a rather linear photocatalytic activity up to 12 h. While MET systems are fully inactive only after 20 h. Long-term experiments up to 36 h confirmed these trends, with no further H₂ production in DET after 24 h and minimal H₂ yield increases in MET (24.0 to 25.0 μmol for [FeFe] and 25.6 to 26.7 μmol for [NiFeSe]). These findings align with recent observations on carbon dot/[FeFe]-H₂ase photocatalytic systems.¹⁴

To gain deep insights into the interaction between ^{NCN}CN_x and H₂ase, QCM analysis was conducted. As shown in the schematic in Fig. 3a, the Au-coated quartz chip was functionalized with a thin layer of ^{NCN}CN_x by drop casting 0.5 mL of an ultrasonicated suspension (0.1 mg mL⁻¹) of ^{NCN}CN_x, to mimic the operando conditions during photocatalysis. By flowing a buffer solution containing enzymes on the chip, the adsorption process of H₂ase at the surface of ^{NCN}CN_x can be monitored as a function of time and quantified based on the Sauerbrey equation.³¹ Fig. 3b shows the QCM analysis of *Cpl* [FeFe]-H₂ase and *DvH* [NiFeSe]-H₂ase on the ^{NCN}CN_x-modified chip. After establishing a stable baseline by circulating 0.1 M MOPS pH 7 buffer with 50 mM 4-MBA, the enzymes were introduced separately at the same concentration as in the photocatalysis

experiments. The adsorption of both H₂ases on ^{NCN}CN_x exhibits in two distinct stages, a fast adsorption process before 1.5 h and slow adsorption after 1.5 h. Interestingly, a higher amount of [NiFeSe]-H₂ase (39.5 pmol cm⁻²) is adsorbed onto ^{NCN}CN_x compared to [FeFe]-H₂ase (16.6 pmol cm⁻²) over 10 h. The observed adsorption profiles can be explained by the proposed electrostatic interactions in Fig. 3c. Based on the electrostatic potential maps (Fig. 3c), both H₂ases exhibit distinct surface charge distribution. By indexing the specific protein structures, *Cpl* [FeFe]-H₂ase (PDB: 4XDC), and *DvH* [NiFeSe]-H₂ase (PDB: 5JSH), within the protein dipole moment database,³² it is found that [NiFeSe]-H₂ase possesses a larger dipole moment of 1972 D compared to [FeFe]-H₂ase (1707 D), resulting in a stronger association (Fig. 3b). However, for [NiFeSe]-H₂ase, DET can only be established *via* the negatively charged patch near the distal FeS cluster, which is unfavorable for the negatively charged ^{NCN}CN_x and thus dramatically reduces DET to the enzyme active site for catalysis. Consequently, the strong association observed at the ^{NCN}CN_x/[NiFeSe]-H₂ase interface is non-specific and results mainly in inactive biohybrid assemblies. In contrast, adsorbed [FeFe]-H₂ase has positively charged distal FeS cluster that can specifically interact with negatively charged ^{NCN}CN_x for DET.^{22,28} The presence of other positively charged regions in the [FeFe]-H₂ase (Fig. 3c) might also attract ^{NCN}CN_x. However, due to the rigidity of the heptazine-based ^{NCN}CN_x (A-B' stacking), proper orientation for DET near the distal FeS cluster could be hindered. This might explain the observed relatively low 25% DET/MET ratio. Therefore, QCM analysis provides valuable insights into the significance of specific interactions in facilitating DET.³³

To investigate the charge carrier dynamics between H₂ases and ^{NCN}CN_x, PEIS was performed using a three-electrode configuration. By applying a sinusoidal potential modulation to the ^{NCN}CN_x-modified working electrode, which was made by depositing a ^{NCN}CN_x suspension (5 μL, 24 mg mL⁻¹) on FTO-coated glass,²³ the impedance was recorded as the ratio of the complex-valued potential and current.³⁴ A Randles circuit consisting of a series resistance (R_s) in parallel with a combination of bulk capacitance (C_{bulk}) and charge transfer resistance (R_{ct}), was proposed to fit the impedance response (Fig. 4a).³⁵ The Nyquist plot (Fig. 4a) of the impedance response measured at -0.1 V vs. the reversible hydrogen electrode (RHE), is

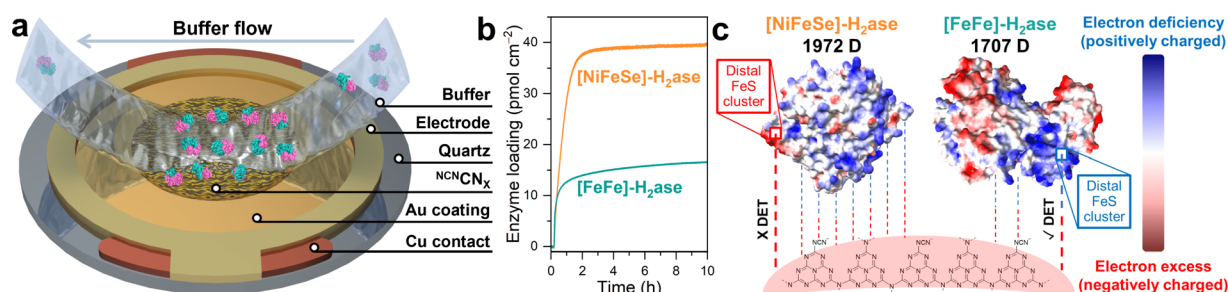


Fig. 3 (a) Schematic illustration of a ^{NCN}CN_x-coated quartz chip. Buffer containing [FeFe]-H₂ase (PDB: 4XDC) is flowing towards the chip and [FeFe]-H₂ase is adsorbed onto the surface. (b) QCM analysis of the adsorption process of H₂ase on a ^{NCN}CN_x-coated quartz chip. Conditions: 2 mL anaerobic MOPS buffer (0.1 M, pH 7) containing 50 mM 4-MBA, 80 pmol H₂ase (either *Cpl* [FeFe] or *DvH* [NiFeSe]), 25 °C. (c) Electrostatic potential maps of *Cpl* [FeFe] (PDB: 4XDC) and *DvH* [NiFeSe] (PDB: 5JSH), and their interactions with ^{NCN}CN_x.



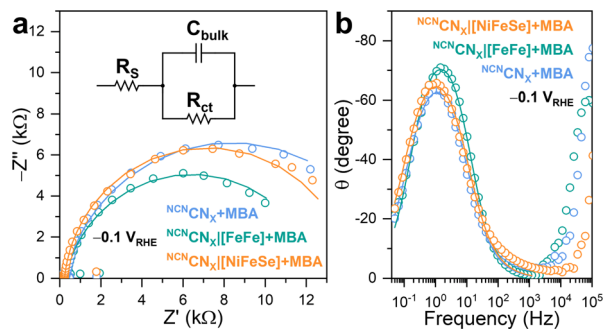


Fig. 4 (a) Nyquist plots and (b) Bode phase plots of PEIS signal (open circles) with corresponding fitting curves (solid lines). Inset: proposed equivalent circuit to fit the impedance response. Conditions: 20 mL anaerobic MOPS buffer (0.1 M, pH 7) containing 50 mM 4-MBA, Ag/AgCl (3 M NaCl) reference electrode, Pt mesh counter electrode, AM 1.5G irradiation, 25 °C.

dominated by a single semicircle with no indication of a Warburg diffusion element. The proposed equivalent circuit provided a good fit ($r^2 > 0.95$) to the impedance response, enabling quantitative analysis of the charge transfer process.

Upon introduction of [FeFe]-H₂ase on the working electrode, a decrease in the semicircle diameter is observed, corresponding to a decrease in R_{ct} from 15 975 Ω to 12 317 Ω. This indicates that [FeFe]-H₂ase, as a biocatalyst, facilitates the charge transfer from ^{N^{CN}}CN_x to the electrolyte for HER. Likewise, ^{N^{CN}}CN_x[[NiFeSe]-H₂ase shows a R_{ct} of 13 950 Ω, similar to the bare ^{N^{CN}}CN_x. Such behaviors have been widely observed when incorporating synthetic co-catalysts onto semiconductors.³⁶ The fitting results allowed determination of the pseudo first-order rate constant for charge transfer (k_{ct}), based on the phenomenological model developed for an illuminated photoelectrode.^{37,38} Specifically, the angular frequency at the maximum imaginary component of the semicircle in Nyquist plot (Fig. 4a) is equal to k_{ct} . The addition of [FeFe]-H₂ase significantly enhances k_{ct} from 6.85 s⁻¹ to 11.99 s⁻¹, whereas [NiFeSe]-H₂ase shows a negative impact on k_{ct} with a value of 5.02 s⁻¹, further confirming the importance of specific interactions in facilitating the charge transfer process. The Bode phase plots (Fig. 4b) revealed that the charge transfer process occurred within the frequency range of 0.1 Hz to 1 kHz, consistent with the reported timeframe for photocatalytic HER using CN_x.³⁶ Within this range, the characteristic frequency at the maximum phase shift of ^{N^{CN}}CN_x[[FeFe]-H₂ase are higher than ^{N^{CN}}CN_x[[NiFeSe]-H₂ase and pristine ^{N^{CN}}CN_x, indicating [FeFe]-H₂ase can initiate a faster charge transfer process for HER. Likewise, the characteristic frequencies of ^{N^{CN}}CN_x[[NiFeSe]-H₂ase and pristine ^{N^{CN}}CN_x remain the same, meaning that DET cannot be established between ^{N^{CN}}CN_x and [NiFeSe]-H₂ase. The voltage-dependent impedance response is illustrated in Fig. S23.† A more negative applied potential yields a diminished semicircular feature in the Nyquist plots, indicating reduced charge transfer resistance. This observation arises from the introduction of a larger band bending, resulting in improved separation of photogenerated charges.^{39,40} Consequently, a greater population of free charge carriers is localized

within the semiconductor, increasing the conductivity of ^{N^{CN}}CN_x. Notably, the RC response of the conductive substrate forms a semicircle with a diameter of approximately 200 Ω in the Nyquist plots (Fig. S24a†) at high frequency region (10 kHz to 1 MHz, Fig. S24b†) in the Bode phase plots.²³ This impedance study on CN_x with H₂ase demonstrates that a specific interaction enables efficient DET by decreasing in R_{ct} and increasing in k_{ct} .

The charge carrier dynamics between H₂ases and ^{N^{CN}}CN_x were further examined using IMVS and TPC techniques in a three-electrode setup. IMVS is a spectroelectrochemical method widely employed in assessing electron recombination processes in photovoltaics. It monitors the open circuit voltage response to the sinusoidally modulated incident light intensity. The characteristic frequency observed at the minimum point of the Nyquist plot (f_{min}) directly correlates to the time constant of electron recombination. This parameter can be calculated using the following equation, providing the first-order electron lifetime τ_n :⁴¹

$$\tau_n = (2\pi f_{min})^{-1}$$

This model has been recently expanded to photoanodes for solar water oxidation.^{42,43} Although IMVS is not *operando* due to the distinct differences between a photoelectrode and a photocatalyst, photoelectrochemical techniques have been widely employed to gain insights into charge carrier behaviors in photochemical systems. This equation can be applied to a ^{N^{CN}}CN_x-based photoelectrode due to it functions as a photoanode in the presence of 4-MBA and under open circuit conditions.²³ As depicted in Fig. 5a, the IMVS response exhibits a distinct semicircle in quadrant IV of the Nyquist plot, suggesting rapid kinetics in 4-MBA oxidation, similar to the cases of sacrificial Na₂SO₃ and H₂O₂ oxidation on a hematite photoanode.^{42,43}

The non-specific interaction between [NiFeSe]-H₂ase and ^{N^{CN}}CN_x yields τ_n of 57.0 μs, whereas 55.1 μs is observed for pristine ^{N^{CN}}CN_x. This consistency indicates that the charge recombination process remains unaffected, matching with our

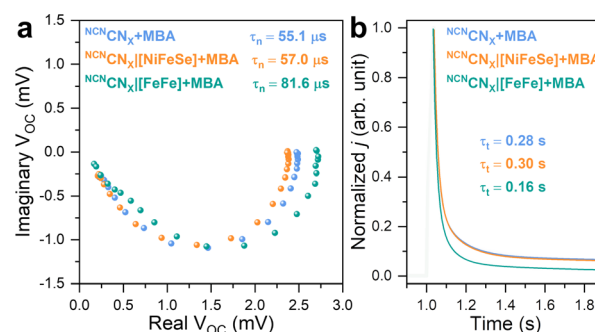


Fig. 5 (a) Nyquist plots of IMVS response. (b) Normalized TPC response with corresponding exponential fitting curves. Conditions: 20 mL anaerobic MOPS buffer (0.1 M, pH 7) containing 50 mM 4-MBA, Ag/AgCl (3 M NaCl) reference electrode, Pt mesh counter electrode, AM 1.5G irradiation, 25 °C.



previous observations of the absence of DET between [NiFeSe]-H₂ase and ^{NCN}CN_x. Upon the introduction of [FeFe]-H₂ase onto ^{NCN}CN_x, we observe a prolonged τ_n of 81.6 μ s that can be explained as follows: Even under open circuit conditions, where no net electron exchange occurs at the ^{NCN}CN_x|electrolyte interface, photogenerated electrons theoretically have the potential to react with protons *via* H₂ases. Note that a TOF of 18 669 h⁻¹ (^{NCN}CN_x|[FeFe]-H₂ase) corresponds to a frequency of 5.2 Hz. Therefore, all losses of photogenerated electrons within the measured frequency range (0.5 MHz to 0.5 Hz) are a combination of bulk recombination and catalytic reaction. The influence of a catalytic overlayer on IMVS response remains a topic of debate. Recent studies indicate that a co-catalyst overlayer can delocalize photogenerated charge carriers from the bulk photoelectrode, promoting charge separation and prolonging electron lifetime.⁴² Here, despite the absence of net exchange current, we hypothesize that photogenerated electrons can be stored in H₂ase in the form of metal hydrides and reduced FeS clusters,⁴⁴ *i.e.*, reversible intermediates for H₂ evolution reaction. This storage mechanism reduces the probability of charge recombination with holes, resulting in an extended electron lifetime. IMVS observations are in line with the recent transient spectroscopic study on the impact of electron accumulation to charge recombination in ^{NCN}CN_x.²⁵ Thus, we demonstrate the use of IMVS on carbon nitride materials and on studying bio-hybrids.

Having gained insights into charge recombination, we conducted TPC measurements to assess the influence of H₂ases on the electron extraction process of ^{NCN}CN_x. The normalized TPC response, illustrated in Fig. 5b, reveals that the integration of [FeFe]-H₂ase with ^{NCN}CN_x leads to a significant reduction in electron transit time (τ_e) from 0.28 s to 0.16 s. This indicates that [FeFe]-H₂ase, acting as a co-catalyst for ^{NCN}CN_x, can effectively collect photoelectrons for HER, thereby facilitating electron transport within ^{NCN}CN_x.^{45,46} Likewise, ^{NCN}CN_x|[NiFeSe]-H₂ase assemblies are non-specific, resulting in an unchanged τ_e of 0.30 s. The combined results from IMVS and TPC highlight the favorable effects of DET between [FeFe]-H₂ase and ^{NCN}CN_x on both charge recombination and transport. In contrast, the non-specifically interacted ^{NCN}CN_x|[NiFeSe]-H₂ase assemblies demonstrate minimal alterations in both τ_n and τ_e .

Conclusions

We present an electrostatic strategy for linking enzymes with carbon nitrides, demonstrating a benchmark for DET between ^{NCN}CN_x and [FeFe]-H₂ase for solar H₂ production with a TON of 2×10^5 and a DET/MET ratio of 25% over 24 h. In contrast, the electrostatic repulsion between [NiFeSe]-H₂ase and ^{NCN}CN_x drastically reduced DET, leading to a DET/MET ratio of 0.5%. QCM analysis demonstrates that specific interactions play a pivotal role in enabling DET, irrespective of the observed differences in the adsorption profiles. Complementary spectroelectrochemical analysis using PEIS, IMVS, and TPC show that interfacing [FeFe]-H₂ase with ^{NCN}CN_x facilitates charge transfer and suppresses charge recombination, as evidenced by a 23% less resistive R_{ct} , a 75% faster k_{ct} , a 48% longer τ_n , and

a 43% shorter τ_e than bare ^{NCN}CN_x. This study provides a promising and straightforward approach for achieving efficient electron transfer between carbon nitride and enzymes and serves as a reference for studying the charge carrier behavior of enzyme-photocatalyst assemblies using interfacial characterizations.

Data availability

Data supporting the findings of this study are available from the Cambridge data repository: <https://doi.org/10.17863/CAM.106936>.

Author contributions

Yongpeng Liu conceptualization, data curation, software, formal analysis, funding acquisition, investigation, visualization, methodology, writing – original draft, project administration, writing – review & editing; Carolina Pulignani conceptualization, resources, data curation, formal analysis, investigation, visualization, methodology, writing – original draft, writing – review & editing; Sophie Webb resources, investigation, methodology, writing – review & editing; Samuel J. Cobb data curation, investigation, methodology, writing – review & editing; Santiago Rodríguez-Jiménez formal analysis, investigation, methodology, writing – review & editing; Dongseok Kim investigation, methodology; Ross D. Milton conceptualization, resources, supervision, funding acquisition, validation, writing – original draft, project administration, writing – review & editing; Erwin Reisner conceptualization, resources, formal analysis, supervision, funding acquisition, validation, investigation, visualization, writing – original draft, project administration, writing – review & editing.

Conflicts of interest

There are no conflicts to declare.

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References

- S. Nandy, S. A. Savant and S. Haussener, *Chem. Sci.*, 2021, **12**, 9866–9884.
- X. Wang, K. Maeda, A. Thomas, K. Takane, G. Xin, J. M. Carlsson, K. Domen and M. Antonietti, *Nat. Mater.*, 2009, **8**, 76–80.
- M. Kwak, J. Bok, B.-H. Lee, J. Kim, Y. Seo, S. Kim, H. Choi, W. Ko, W. H. Antink, C. W. Lee, G. H. Yim, H. Seung, C. Park, K.-S. Lee, D.-H. Kim, T. Hyeon and D. Yoo, *Chem. Sci.*, 2022, **13**, 8536–8542.
- V. W. Lau, I. Moudrakovski, T. Botari, S. Weinberger, M. B. Mesch, V. Duppel, J. Senker, V. Blum and B. V. Lotsch, *Nat. Commun.*, 2016, **7**, 12165.
- H. Kasap, C. A. Caputo, B. C. M. Martindale, R. Godin, V. W. Lau, B. V. Lotsch, J. R. Durrant and E. Reisner, *J. Am. Chem. Soc.*, 2016, **138**, 9183–9192.
- W. Lubitz, H. Ogata, O. Rüdiger and E. Reijerse, *Chem. Rev.*, 2014, **114**, 4081–4148.
- A. Le Goff, V. Artero, B. Jusselme, P. D. Tran, N. Guillet, R. Métayé, A. Fihri, S. Palacin and M. Fontecave, *Science*, 2009, **326**, 1384–1387.
- W. Lubitz, E. Reijerse and M. van Gastel, *Chem. Rev.*, 2007, **107**, 4331–4365.
- G. Si, W.-G. Wang, H.-Y. Wang, C.-H. Tung and L.-Z. Wu, *Inorg. Chem.*, 2008, **47**, 8101–8111.
- J.-X. Jian, C. Ye, X.-Z. Wang, M. Wen, Z.-J. Li, X.-B. Li, B. Chen, C.-H. Tung and L.-Z. Wu, *Energy Environ. Sci.*, 2016, **9**, 2083–2089.
- X. Fang, S. Kalathil and E. Reisner, *Chem. Soc. Rev.*, 2020, **49**, 4926–4952.
- N. Kornienko, J. Z. Zhang, K. K. Sakimoto, P. Yang and E. Reisner, *Nat. Nanotechnol.*, 2018, **13**, 890–899.
- K. A. Brown, S. Dayal, X. Ai, G. Rumbles and P. W. King, *J. Am. Chem. Soc.*, 2010, **132**, 9672–9680.
- K. Holá, M. V. Pavliuk, B. Németh, P. Huang, L. Zdražil, H. Land, G. Berggren and H. Tian, *ACS Catal.*, 2020, **10**, 9943–9952.
- C. A. Caputo, M. A. Gross, V. W. Lau, C. Cavazza, B. V. Lotsch and E. Reisner, *Angew. Chem., Int. Ed.*, 2014, **53**, 11538–11542.
- C. A. Caputo, L. Wang, R. Beranek and E. Reisner, *Chem. Sci.*, 2015, **6**, 5690–5694.
- Y. Zhang and J. Liu, *Chem.–Eur. J.*, 2022, **28**, e202201430.
- S. Zhang, Y. Zhang, Y. Chen, D. Yang, S. Li, Y. Wu, Y. Sun, Y. Cheng, J. Shi and Z. Jiang, *ACS Catal.*, 2021, **11**, 476–483.
- J. Liu, Y. Liu, N. Liu, Y. Han, X. Zhang, H. Huang, Y. Lifshitz, S.-T. Lee, J. Zhong and Z. Kang, *Science*, 2015, **347**, 970–974.
- Y. Liu, S. Webb, P. Moreno-García, A. Kulkarni, P. Maroni, P. Broekmann and R. D. Milton, *JACS Au*, 2023, **3**, 124–130.
- M. C. Marques, C. Tapia, O. Gutiérrez-Sanz, A. R. Ramos, K. L. Keller, J. D. Wall, A. L. De Lacey, P. M. Matias and I. A. C. Pereira, *Nat. Chem. Biol.*, 2017, **13**, 544–550.
- J. W. Peters, W. N. Lanzilotta, B. J. Lemon and L. C. Seefeldt, *Science*, 1998, **282**, 1853–1858.
- C. Pulignani, C. A. Mesa, S. A. J. Hillman, T. Uekert, S. Giménez, J. R. Durrant and E. Reisner, *Angew. Chem., Int. Ed.*, 2022, **61**, e202211587.
- V. W. Lau, D. Klose, H. Kasap, F. Podjaski, M.-C. Pignié, E. Reisner, G. Jeschke and B. V. Lotsch, *Angew. Chem., Int. Ed.*, 2017, **56**, 510–514.
- W. Yang, R. Godin, H. Kasap, B. Moss, Y. Dong, S. A. J. Hillman, L. Steier, E. Reisner and J. R. Durrant, *J. Am. Chem. Soc.*, 2019, **141**, 11219–11229.
- R. Godin, Y. Wang, M. A. Zwijnenburg, J. Tang and J. R. Durrant, *J. Am. Chem. Soc.*, 2017, **139**, 5216–5224.
- N. E. Good, G. D. Winget, W. Winter, T. N. Connolly, S. Izawa and R. M. M. Singh, *Biochemistry*, 1966, **5**, 467–477.
- P. M. Matias, C. M. Soares, L. M. Saraiva, R. Coelho, J. Morais, J. Le Gall and M. A. Carrondo, *JBIC, J. Biol. Inorg. Chem.*, 2001, **6**, 63–81.
- G. A. M. Hutton, B. Reuillard, B. C. M. Martindale, C. A. Caputo, C. W. J. Lockwood, J. N. Butt and E. Reisner, *J. Am. Chem. Soc.*, 2016, **138**, 16722–16730.
- M. Heyrovský, *J. Chem. Soc. Chem. Commun.*, 1987, 1856–1857.
- G. Sauerbrey, *Z. Med. Phys.*, 1959, **155**, 206–222.
- C. E. Felder, J. Prilusky, I. Silman and J. L. Sussman, *Nucleic Acids Res.*, 2007, **35**, W512–W521.
- V. M. Badiani, C. Casadevall, M. Miller, S. J. Cobb, R. R. Manuel, I. A. C. Pereira and E. Reisner, *J. Am. Chem. Soc.*, 2022, **144**, 14207–14216.
- A. J. Bard and L. R. Faulkner, *Electrochemical Methods: Fundamentals and Applications*, John Wiley & Sons, Incorporated, 2nd edn, 2000.
- J. E. B. Randles, *Discuss. Faraday Soc.*, 1947, **1**, 11–19.
- W. Jiang, Y. Zhao, X. Zong, H. Nie, L. Niu, L. An, D. Qu, X. Wang, Z. Kang and Z. Sun, *Angew. Chem., Int. Ed.*, 2021, **60**, 6124–6129.
- K. G. U. Wijayantha, S. Saremi-Yarahmadi and L. M. Peter, *Phys. Chem. Chem. Phys.*, 2011, **13**, 5264–5270.
- P. Xu, C. L. Gray, L. Xiao and T. E. Mallouk, *J. Am. Chem. Soc.*, 2018, **140**, 11647–11654.
- J. Bisquert, *J. Phys. Chem. B*, 2002, **106**, 325–333.
- Y. Liu, J. Quiñero, L. Yao, X. D. C. Pereira, M. Mensi, R. Gómez, K. Sivula and N. Guijarro, *J. Mater. Chem. A*, 2021, **9**, 2888–2898.
- J. Krüger, R. Plass, M. Grätzel, P. J. Cameron and L. M. Peter, *J. Phys. Chem. B*, 2003, **107**, 7536–7539.
- Y. Liu, N. Guijarro and K. Sivula, *Helv. Chim. Acta*, 2020, **103**, e2000064.
- J. E. Thorne, J.-W. Jang, E. Y. Liu and D. Wang, *Chem. Sci.*, 2016, **7**, 3347–3354.
- D. Schilter, J. M. Camara, M. T. Huynh, S. Hammes-Schiffer and T. B. Rauchfuss, *Chem. Rev.*, 2016, **116**, 8693–8749.
- F. Le Formal, K. Sivula and M. Grätzel, *J. Phys. Chem. C*, 2012, **116**, 26707–26720.
- Y. Liu, M. Xia, L. Yao, M. Mensi, D. Ren, M. Grätzel, K. Sivula and N. Guijarro, *Adv. Funct. Mater.*, 2021, **31**, 2010081.

