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Identification of sulfonylated indolo[1,2-*a*]quinolines as EGFR tyrosine kinase inhibitors†

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Two series of indolo[1,2-*a*]quinolines (IQs), comprising six 6-trifluoromethylthio indolo[1,2-*a*]quinolines and nine 6-arenesulfonyl indolo[1,2-*a*]quinolines, were screened for their inhibitory activity against EGFR tyrosine kinase (EGFR-TK) using the ADP-Glo™ kinase assay. Among the 15 IQs screened, four compounds exhibited cytotoxic activity against a lung cancer cell line (A549) that was as potent as the known drug afatinib with lower cytotoxicity in Vero cells. In addition, while they displayed cytotoxic activity against a head and neck squamous cell carcinoma cell line (SCC cells), they were inactive against a colorectal cancer cell line (LS174T cells). Molecular dynamics (MD) simulations revealed that IQSO₂R-I (IC₅₀: 0.28 ± 0.05 μM) formed a stable complex with wild-type EGFR through hydrophobic interactions and hydrogen bonding with the K745 residue. Additionally, the compound complied with the extended rule of five. This class of compounds represents a novel class of EGFR-TK inhibitors, which may serve as a novel scaffold for the development of anticancer therapeutics targeting EGFR-TK.

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Introduction

Cancer is a major disease that causes high mortality worldwide. Among the various types of cancer, lung cancer is the second most common cancer and has the highest mortality rates.¹ Treatment options for patients with lung cancer include surgery, radiotherapy (radiation), chemotherapy, and immunotherapy.² Nowadays, targeted chemotherapy for cancer treatment has received much attention globally owing to its specificity for cancer cells.³ A number of targeted anticancer drugs are now available for many common cancers, including breast cancer, colorectal, pancreatic cancer, lung cancer, leukemia, lymphoma, and multiple myeloma.⁴ However, even though targeted therapies are typically better tolerated than traditional chemotherapy, they are still associated with several adverse effects.

Epidermal growth factor receptor (EGFR) is a transmembrane protein and a member of the ErbB family, a subfamily of four closely related receptor tyrosine kinases (TKs).⁵ Mutations leading to EGFR overexpression are closely associated with the occurrence of several types of cancer, including non-small cell lung cancer (NSCLC), head cancer, breast cancer, ovarian cancer, and bladder carcinoma.⁶ As a result, targeting the EGFR protein has been recognized as a promising strategy for the targeted therapy of cancer.

Afatinib, a second-generation EGFR-TK inhibitor, is an FDA-approved anticancer drug used for the treatment of EGFR

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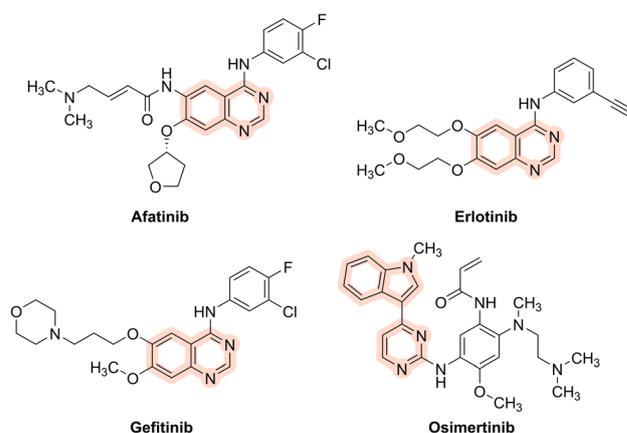


Fig. 1 Structures of afatinib, erlotinib, gefitinib, and osimertinib.



mutation-driven NSCLC.⁷ Afatinib is used in cancer therapy owing to its inhibitory activity against exon 19 deletion and exon 21 (L858R) substitution mutations. Although afatinib is recommended as a first-line therapy for EGFR-mutated metastatic lung cancer, acquired drug resistance, caused by the T790M mutation of the EGFR-TK domain, unavoidably develops after a median duration of treatment. Additionally, common side effects of tyrosine kinase inhibitors include rash, diarrhea, and liver toxicity. Due to these reasons, new therapeutics that can reduce risks and exhibit fewer adverse side effects are needed.

Nitrogen heterocycles are an important class of compounds found in a number of natural products, drugs and functional materials.⁸ Among them, indolo[1,2-*a*]quinolines (IQs) have a unique nitrogen-containing tetracyclic scaffold.⁹ Afatinib and other approved drugs targeting EGFR, including erlotinib, osimertinib and gefitinib, contain a nitrogen heterocyclic scaffold (Fig. 1). On the basis of our previously reported work¹⁰ and the fact that 6-arenesulfonyl indolo[1,2-*a*]quinolines (IQSO₂R_s) and

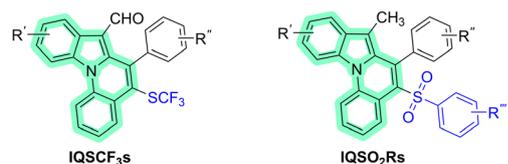
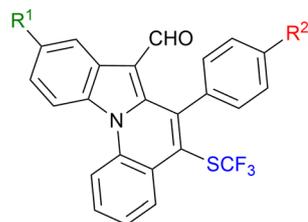
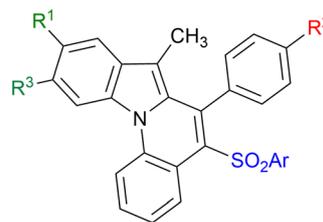


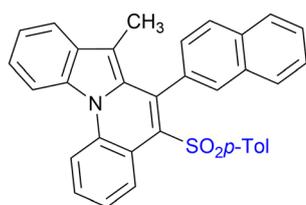
Fig. 2 Indolo[1,2-*a*]quinoline (IQ) core (highlighted in green) of 6-trifluoromethylthio indolo[1,2-*a*]quinolines (IQSCF₃s) and 6-arenesulfonyl indolo[1,2-*a*]quinolines (IQSO₂R_s).



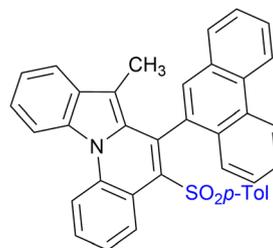
- IQSCF₃-I:** R¹ = R² = H
IQSCF₃-II: R¹ = CH₃, R² = H
IQSCF₃-III: R¹ = OCH₃, R² = H
IQSCF₃-IV: R¹ = F, R² = H
IQSCF₃-V: R¹ = Cl, R² = H
IQSCF₃-VI: R¹ = H, R² = NO₂



- IQSO₂R-I:** R¹ = R² = R³ = H, Ar = *p*-MeC₆H₄
IQSO₂R-II: R¹ = R² = R³ = H, Ar = C₆H₅
IQSO₂R-III: R¹ = R² = R³ = H, Ar = *p*-MeOC₆H₄
IQSO₂R-IV: R¹ = H, R² = CH₃, R³ = H, Ar = *p*-MeC₆H₄
IQSO₂R-V: R¹ = F, R² = H, R³ = H, Ar = *p*-MeC₆H₄
IQSO₂R-VI: R¹ = OCH₃, R² = H, R³ = H, Ar = *p*-MeC₆H₄
IQSO₂R-VII: R¹ = H, R² = H, R³ = CH₃, Ar = *p*-MeC₆H₄



IQSO₂R-VIII



IQSO₂R-IX

Fig. 3 Chemical structures of the six IQSCF₃s (IQSCF₃-I–IQSCF₃-VI) and nine IQSO₂R_s (IQSO₂R-I–IQSO₂R-IX) screened toward EGFR-TK inhibitory activity.

6-trifluoromethylthio indolo[1,2-*a*]quinolines (IQSCF₃s) bear both indole and quinoline rings in their structures, we hypothesized that IQSO₂R_s and IQSCF₃s (Fig. 2) might exhibit potential EGFR-TK inhibitory activity. Therefore, our objectives in the present study were to investigate the EGFR-TK inhibitory activity of some selected IQs, focusing on those bearing either a arenesulfonyl group (SO₂R) or trifluoromethylthio group (SCF₃) in their structures, and to evaluate their effect on the cell viability of EGFR wild-type non-small-cell lung cancer (A549), EGFR wild-type head and neck squamous cell carcinoma (SCC), and EGFR-KRAS^{G12D}-mutated colorectal cancer cells (LS174T) and normal cells (Vero cells).

Results and discussion

Synthesis of IQSCF₃s and IQSO₂R_s

The syntheses of six 6-trifluoromethylthio indolo[1,2-*a*]quinolines (IQSCF₃-I–IQSCF₃-VI) and nine 6-arenesulfonyl indolo[1,2-*a*]quinolines (IQSO₂R-I–IQSO₂R-IX) were efficiently achieved by following previously reported protocols (Fig. 3) (see the ESI†). All the synthesized compounds in this study were purified and characterized (see the ESI†).

Inhibition of EGFR-TK by IQs (IQSCF₃s and IQSO₂R_s)

Primarily, the EGFR-TK inhibitory activity of the six synthesized IQSCF₃ series (IQSCF₃-I–IQSCF₃-VI) and nine synthesized IQSO₂R series (IQSO₂R-I–IQSO₂R-IX) was evaluated. Afatinib, a medication used to treat non-small cell lung carcinoma



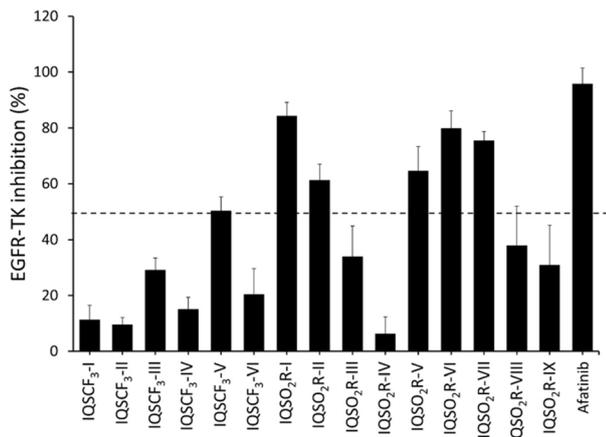


Fig. 4 Kinase inhibitory activity screening of six synthesized IQSCF₃ series (IQSCF₃-I–IQSCF₃-VI) and nine synthesized IQSO₂R series (IQSO₂R-I–IQSO₂R-IX) toward EGFR-TK at 1 μM as determined by EGFR kinase activity assays. The data are presented as the mean ± standard error of mean (S.E.M.) from three independent experiments (*n* = 3).

(NSCLC), was employed as a benchmark. Thus, indolo[1,2-*a*]quinoline derivatives (IQs) and afatinib were screened at 1 μM using EGFR kinase inhibition assay kits. The results are shown in Fig. 4, and it revealed that all the compounds showed inhibitory activity toward EGFR-TK. Among the 15 compounds screened, six compounds, namely IQSCF₃-V, IQSO₂R-I, IQSO₂R-II, IQSO₂R-V, IQSO₂R-VI, and IQSO₂R-VII showed EGFR-TK inhibitory activity greater than 50% inhibition at 1 μM (96.96% inhibition for afatinib). The preliminary screening results suggested that the IQs could serve as EGFR-TK inhibitors. These six compounds that exhibited greater than 50% inhibition at 1 μM were subjected to further investigation.

Cell viability assay of EGFR-overexpressing cancer cell lines in a lung adenocarcinoma cell line (A549), head and neck squamous cell carcinoma (SCC), colorectal cancer cell (LS174T), and normal kidney epithelial cell (Vero) lines

The six selected compounds, *i.e.*, IQSCF₃-V, IQSO₂R-I, IQSO₂R-II, IQSO₂R-V, IQSO₂R-VI, and IQSO₂R-VII, were subjected to an

Table 1 IC₅₀ values of IQSCF₃-V, IQSO₂R-I, IQSO₂R-II, IQSO₂R-V, IQSO₂R-VI, IQSO₂R-VII, and afatinib against A549, SCC, LS174T, and Vero cell lines^a

Compounds	IC ₅₀ (μM)			
	A549	SCC	LS174T	Vero
IQSCF ₃ -V	50.07 ± 1.83	46.35 ± 1.31	>100	17.80 ± 0.31
IQSO ₂ R-I	14.55 ± 3.14	19.80 ± 4.07	>100	22.64 ± 3.59
IQSO ₂ R-II	41.13 ± 5.10	46.81 ± 5.48	>100	18.44 ± 0.14
IQSO ₂ R-V	16.39 ± 2.40	36.71 ± 3.50	>100	18.84 ± 4.31
IQSO ₂ R-VI	14.59 ± 0.87	22.58 ± 1.29	>100	21.67 ± 1.40
IQSO ₂ R-VII	13.71 ± 1.20	21.80 ± 2.42	>100	19.43 ± 2.75
Afatinib	13.09 ± 0.80	0.98 ± 0.10	4.97 ± 1.09	5.77 ± 1.59

^a The data are presented as the mean ± standard error of mean (S.E.M.) from triplicate independent experiments (*n* = 3).

in vitro cytotoxicity assay against EGFR wild-type non-small-cell lung cancer (A549), EGFR wild-type head and neck squamous cell carcinoma (SCC), and EGFR-KRAS^{G12D}-mutated colorectal cancer (LS174T) cells, in comparison with Vero kidney epithelium cells using MTT assays. The cell viability of A549, SCC, LS174T, and Vero cells treated with various concentrations of the six selected compounds or afatinib was evaluated to obtain the half-maximal inhibitory concentration (IC₅₀) values (Table 1). The results revealed that IQSO₂R-I, IQSO₂R-V, IQSO₂R-VI, IQSO₂R-VII, and afatinib exhibited cytotoxicity activity against EGFR wild-type A549 cells, with IC₅₀ values of 14.55 ± 3.14, 16.39 ± 2.40, 14.59 ± 0.87, 13.71 ± 1.20, and 13.09 ± 0.80 μM, respectively (Fig. 5A). IQSO₂R-I, IQSO₂R-V, IQSO₂R-VI, and IQSO₂R-VII exhibited cytotoxicity effects with IC₅₀ values of 19.80 ± 4.07, 36.71 ± 3.50, 22.58 ± 1.29, and 21.80 ± 2.42 μM in EGFR wild-type SCC cells, respectively (Fig. 5B). Notably, IQSO₂R-I, IQSO₂R-V, IQSO₂R-VI, and IQSO₂R-VII displayed slightly lower toxicity in Vero cells with IC₅₀ values in a range of 18–23 μM, whereas the IC₅₀ of afatinib was 6 μM. The six selected compounds did not exhibit cytotoxicity effects against EGFR-KRAS^{G12D} LS174T cells.

Structure–activity relationship (SAR)

To better understand the structure–activity relationship, the inhibitory activity of EGFR-TK and the chemical structure of the IQs were analyzed and revealed that three different locations of substitution on the IQ scaffold, namely (1) the sulfur moiety and the substituent on the arene sulfonyl group (blue color), (2) the substituent on the aryl ring (red color), and (3) the substituents on the indole core (green color), affected the inhibitory potency (Fig. 6). For the sulfur moiety, the IQSCF₃ series (IQSCF₃-I–IQSCF₃-VI) were relatively less active than the IQSO₂R series. Among the IQSCF₃ series, IQSCF₃-V exhibited the highest inhibitory activity (50% inhibition). Among the IQSO₂R series, IQSO₂R-I, IQSO₂R-II, IQSO₂R-V, IQSO₂R-VI, and IQSO₂R-VII, displayed greater than 50% inhibition, with IQSO₂R-I exhibiting the highest inhibitory activity (84% inhibition). It should be noted that the IQSO₂R series that exhibited inhibitory activity greater than 50% inhibition at 1 μM possessed a *para*-Me substituent at the arenesulfonyl moiety, except for IQSO₂R-II. Thus, the presence of a *para*-Me moiety at the arenesulfonyl group may play an important role in the EGFR-TK inhibitory activity. For the substituent on the aryl ring, the presence of a *para*-Me substituent (R² = Me) at aryl ring had a detrimental effect on the inhibitory activity, with IQSO₂R-IV being the least active compound in the IQSO₂R series. The electronically different groups (R¹ and R³) on the indole scaffold of compounds IQSO₂R-V, IQSO₂R-VI, and IQSO₂R-VII did not show any significant effect.

Kinase inhibitory activity

The results for the EGFR-TK activity obtained from both the EGFR-TK inhibition assay and cell viability analyses against A549 cell lines by MTT assay demonstrated that IQSO₂R-I, IQSO₂R-V, IQSO₂R-VI, and IQSO₂R-VII exhibited cytotoxic effects similar to those of afatinib. Therefore, the IC₅₀ values of



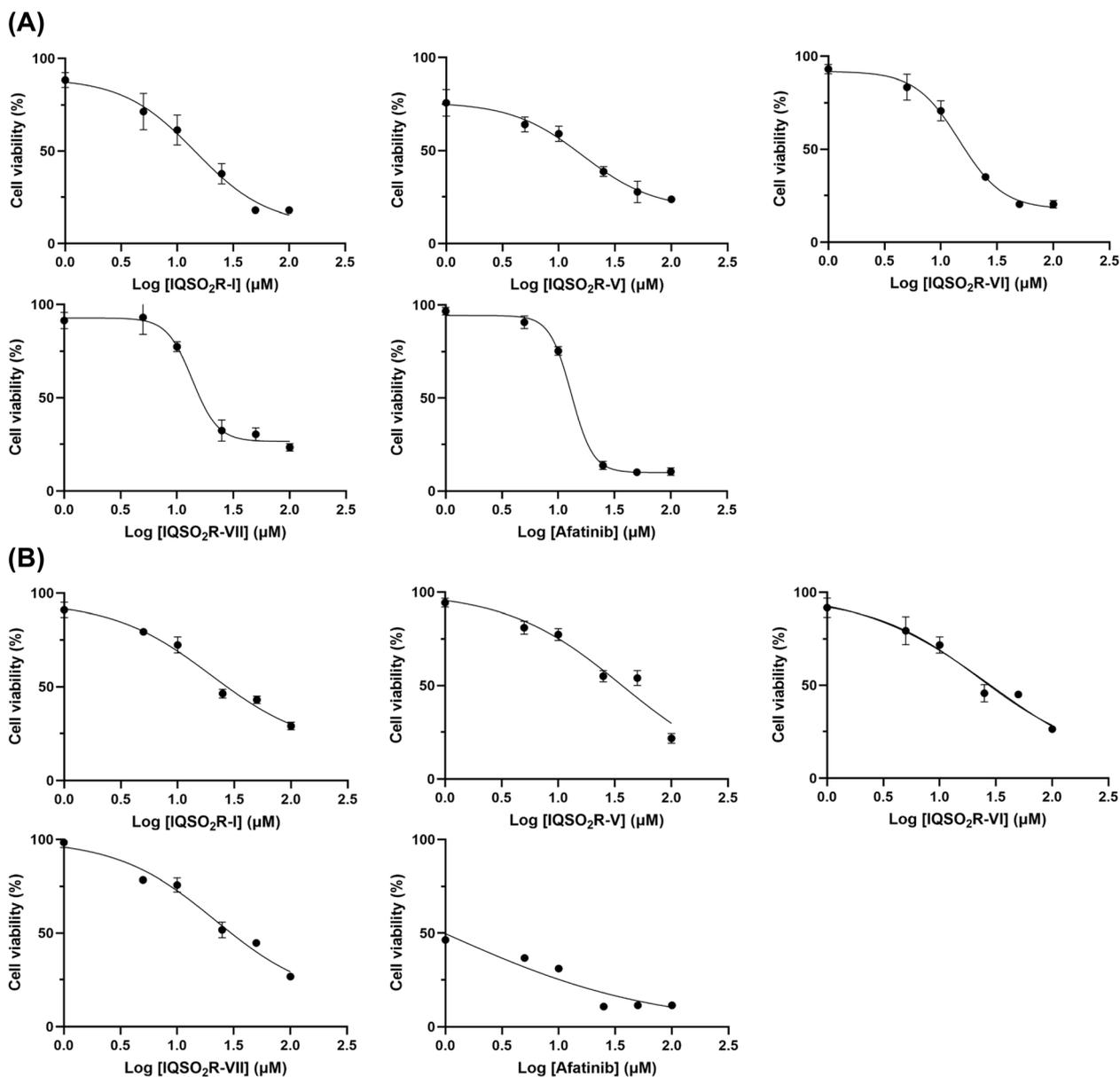


Fig. 5 Dose–response studies of IQSO₂R-I, IQSO₂R-V, IQSO₂R-VI, and IQSO₂R-VII and afatinib in EGFR wild-type non-small-cell lung cancer (A549) (A) and EGFR wild-type head and neck squamous cell carcinoma (SCC) (B). The data are presented as the mean \pm standard error of mean (S.E.M.) from triplicate independent experiments ($n = 3$).

IQSO₂R-I, IQSO₂R-V, IQSO₂R-VI, and IQSO₂R-VII on the kinase inhibitory activity were determined using EGFR-TK inhibition assays to investigate the ligand–protein binding between these compounds and EGFR-TK (Table 2). The IC₅₀ values of IQSO₂R-I, IQSO₂R-V, IQSO₂R-VI, and IQSO₂R-VII were 0.28 ± 0.05 , 0.38 ± 0.12 , 0.44 ± 0.07 , and 0.48 ± 0.19 μM , respectively.

Prediction of the physicochemical properties

The drug-likeness of the four potent IQSO₂R derivatives (IQSO₂R-I, IQSO₂R-V, IQSO₂R-VI, and IQSO₂R-VII) obtained from the EGFR kinase assay was predicted using the SwissADME web server, based on the extended rule of five and their physicochemical properties.¹¹ This analysis involved assessing various characteristics, such as the molecular weight (MW),

number of hydrogen bond acceptors (HBA), number of hydrogen bond donors (HBD), log of octanol-to-water partition coefficient or lipophilicity (LogP), polar surface area (PSA), and the number of rotatable bonds (RB). The results are shown in Table 3. Four IQ derivatives met the acceptable criteria. For validation of the drug-likeness, investigation of the pharmacokinetics properties of these compounds warrants further investigation.

Molecular dynamics study of the IQSO₂R-I/EGFR-TK complex

Considering the experimental data and predicted physicochemical properties of IQSO₂R derivatives, IQSO₂R-I binding susceptibility to the EGFR-TK domain was examined using 500 ns MD simulations with three independent replicates (Fig. 7).



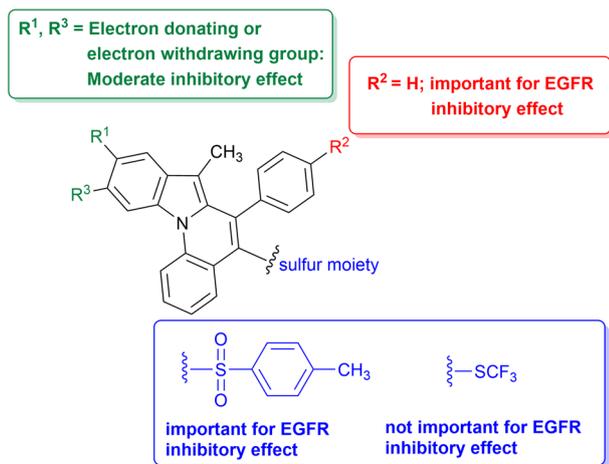


Fig. 6 Analysis of the structure–activity relationship.

Table 2 Kinase inhibitory activity of IQSO₂R-I, IQSO₂R-V, IQSO₂R-VI, and IQSO₂R-VII^a

Compounds	IC ₅₀ (μM)
IQSO₂R-I	0.28 ± 0.05
IQSO₂R-V	0.38 ± 0.12
IQSO₂R-VI	0.44 ± 0.07
IQSO₂R-VII	0.48 ± 0.19

^a The data are presented as the mean ± standard error of mean (S.E.M.) from triplicate independent experiments ($n = 3$).

Table 3 Predicted molecular and drug-likeness properties for IQSO₂R-I, IQSO₂R-V, IQSO₂R-VI, and IQSO₂R-VII^a

Compounds	MW	HBA	HBD	log <i>P</i>	PSA	RB	Drug-likeness
IQSO₂R-I	461.57	2	0	6.40	46.93	3	Yes
IQSO₂R-V	465.54	3	0	6.38	46.93	3	Yes
IQSO₂R-VI	481.99	2	0	6.59	46.93	3	Yes
IQSO₂R-VII	526.44	2	0	6.68	46.93	3	Yes

^a Extended rule of five:¹² molecular weight (MW) ≤ 700 Da; hydrogen bond acceptors (HBAs) ≤ 10; hydrogen bond donors (HBDs) ≤ 5; log *P* (octanol–water partition coefficient) ≤ 7.5; polar surface area (PSA) ≤ 200 Å²; and rotatable bonds (RBs) ≤ 20.

The stability of the **IQSO₂R-I**/EGFR complex was assessed by evaluating the all-atom RMSD, the radius of gyration (Rg) of the complex, the number of intermolecular hydrogen bonds (#Hbonds), and the number of atomic contacts (#Atom contacts), as illustrated in Fig. 7A. The RMSD values of the complex only fluctuated during the initial simulations (till ~100 ns for Run1, ~250 ns for Run2, and ~200 ns for Run3), while the Rg (19.9–20.3 Å), #Hbonds (1–2 bonds), and #Atom contacts (10–30 contacts) values remained stable throughout the simulations. The results suggested a high level of system stability and ligand/protein binding.

The MM/GBSA per-residue binding free energy (Δ*G*_{residue}) was obtained based on 100 snapshots taken from the last 100 ns

of all the simulations. It can be seen from Fig. 7B that there were 9 amino acids L718, F723, V726, K745, A743, T790, M793, G796, and L844 associated with the **IQSO₂R-I** binding. Notably, the contribution of M793 (hinge region) has also been found in the other reported EGFR-TK inhibitors.¹³ Additionally, the sulfonyl moiety of **IQSO₂R-I** formed a strong hydrogen bond with K745 (~72% occupation). As shown in Fig. 8, the presence of the methyl (CH₃) substituent at the SO₂Ar position in **IQSO₂R-I** facilitated strong hydrophobic interactions with EGFR-TK residues I789 (74%), I744 (41%), and I788 (99%). These interactions likely contribute to the enhanced anticancer efficacy of **IQSO₂R-I**, as evidenced by its IC₅₀ value of 14.55 ± 3.14 μM. In contrast, the absence of the CH₃ group at the SO₂Ar position in compounds **IQSCF₃-V** and **IQSO₂R-II** may have resulted in the loss of these hydrophobic interactions, potentially leading to reduced anticancer activity. Furthermore, the sulfur moiety in the **IQSCF₃** series demonstrated lower activity in both the EGFR kinase assay and anticancer evaluation compared to in the **IQSO₂R** series. The replacement of the SO₂Ar group with SCF₃ may eliminate the critical hydrogen bond with K745 and essential hydrophobic interactions, resulting in the reduced activity for the **IQSCF₃** series. For example, **IQSCF₃-V** exhibited less than 50% EGFR-TK inhibition and an IC₅₀ of 50.07 ± 1.83 μM against A549. These findings highlight the importance of both hydrophobic interactions and hydrogen bonding in playing a crucial role in enhancing the EGFR-TK and anticancer activity.¹⁴

Experimental section

Preparation of 6-trifluoromethylthio indolo[1,2-*a*]quinolines (IQSCF₃s) and 6-arenesulfonyl indolo[1,2-*a*]quinolines (IQSO₂Rs)

The investigated 6-trifluoromethylthio indolo[1,2-*a*]quinolines (**IQSCF₃-I**–**IQSCF₃-VI**) and 6-arenesulfonyl indolo[1,2-*a*]quinolines (**IQSO₂R-I**–**IQSO₂R-IX**) were prepared by following a previously reported protocol (see ESI†).^{9d,f}

EGFR-TK inhibition assay

The ADP-Glo™ kinase assay kit was obtained from Promega (Madison, WI, USA). The series of **IQSCF₃s** and **IQSO₂Rs** were screened to determine their EGFR tyrosine kinase activity using the ADP-Glo™ kinase assay, as previously described.⁸ After kinase buffer was added to a well plate, EGFR enzyme at a concentration of 1.25 ng μL⁻¹ and inhibitors were added, followed by a mixture of 25 μM ATP and 12.5 μM poly (Glu–Tyr) and then 1 h incubation. Then, the ADP-Glo reagent and the kinase detection reagent were added and incubated for 40 and 30 min, respectively. The ATP was measured by measuring the luminescence using a microplate reader (BioTek Instruments, VT, USA). The relative inhibition (%) of the inhibitors was calculated compared with the control (no inhibitor), as shown in eqn (1).

$$\% \text{ Relative inhibition} = \frac{[(\text{positive} - \text{negative}) \times (\text{sample} - \text{negative})]}{(\text{positive} - \text{negative})} \times 100 \quad (1)$$



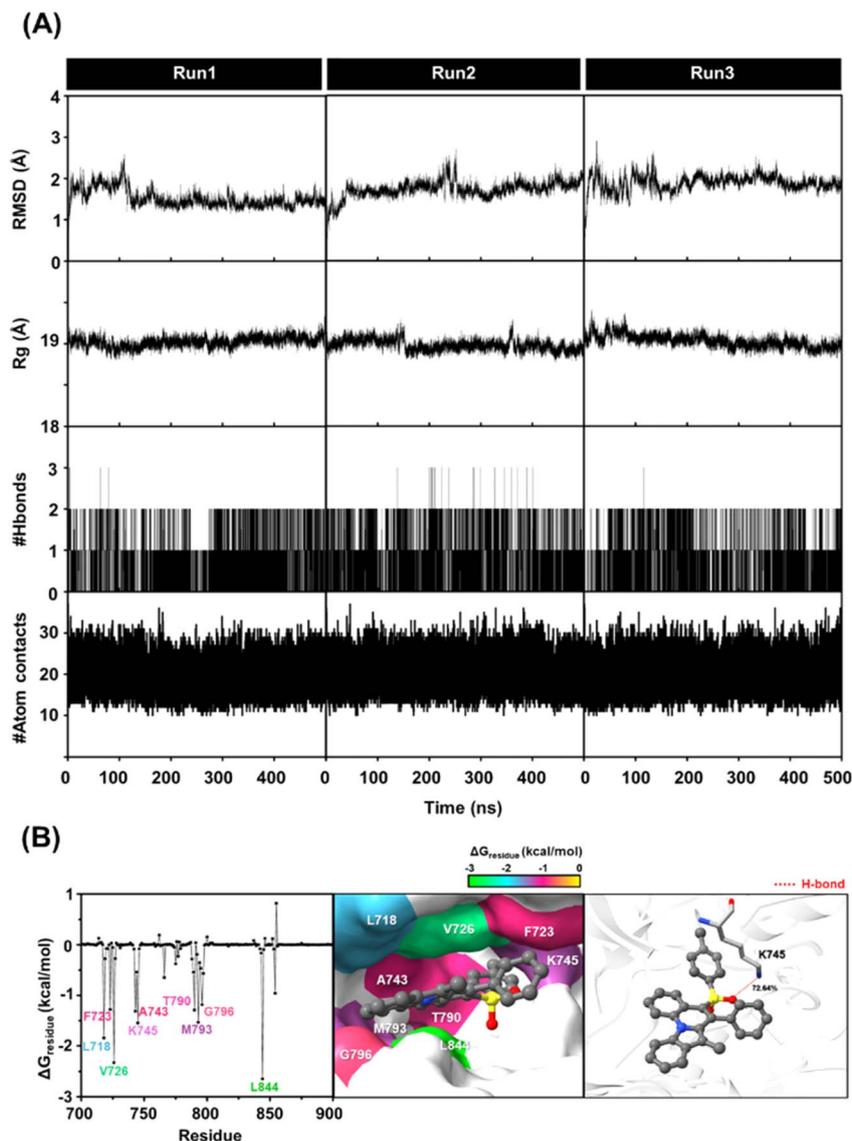


Fig. 7 (A) Structural and dynamic analysis of IQSO₂R-I binding to wild-type EGFR-TK over 500 ns MD simulations in terms of the all-atom RMSD, radius of gyration (Rg) of complex, and the number of hydrogen bonds and atomic contacts. (B) Binding pattern and hydrogen bonding of the IQSO₂R-I/EGFR-TK complex.

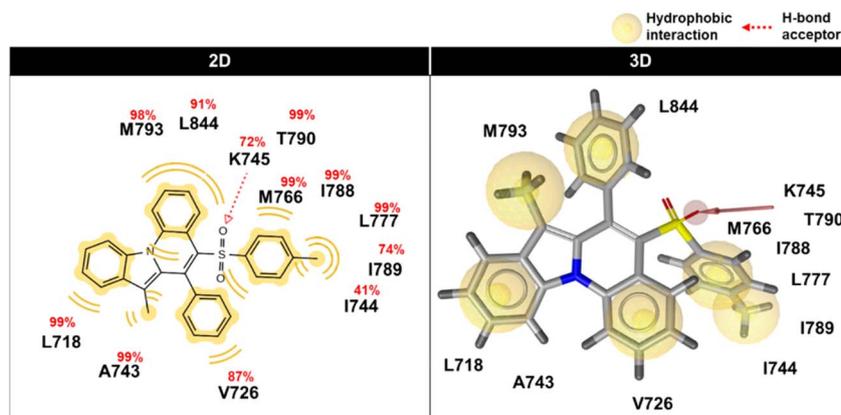


Fig. 8 Key interactions of IQSO₂R-I with wild-type EGFR-TK depicted in 2D and 3D pharmacophore models derived from the final 100 ns of MD simulations. The red arrow denotes the hydrogen bond acceptor (HBA), while the yellow circles indicate the hydrophobic interactions.



Evaluation of cell cytotoxicity in cancer cell lines

A549 Lung adenocarcinoma, SCC head and neck squamous cell, LS174T carcinoma colorectal cancer cell, and Vero cell lines were obtained from American Type Culture Collection (Manassas, VA, USA). The A549 cells were maintained in a Kaighn's modification of Ham's F-12 medium (F-12k Medium) together with 10% FBS and 100 U mL⁻¹ penicillin (Life Technologies, Carlsbad, CA, USA). The SCC cells were maintained in Dulbecco's modified Eagle's medium/Ham's F-12 (DMEMF-12) with 10% FBS and 100 U mL⁻¹ penicillin. The LS174T cells were maintained in RPMI-1640 contained with 10% FBS and 100 U mL⁻¹ penicillin. The Vero cells were maintained in eagle's minimum essential medium (EMEM) together with 10% FBS and 100 U mL⁻¹ penicillin. All cells were kept at 37 °C under humidified 95% O₂ with 5% CO₂ atmosphere.

The cell viability of the A549, SCC, LS174T, and Vero cells was assessed using MTT assays. Briefly, A549 cells (5000 cells per well), SCC cells (10 000 cells per well), LS174T cells (7000 cells per well), and Vero cells (4000 cells per well) were seeded in 96-well plates and incubated at 37 °C for 24 h. After 24 h incubation, the cells were then treated with different concentrations of the second-generation EGFR-TKIs drug (afatinib), **IQSCF₃s**, and **IQSO₂R_s** at 37 °C for 72 h incubation. MTT solution (5 mg mL⁻¹) was added to the cells and kept for 2 h incubation at 37 °C. The reaction was stopped by adding DMSO (100 μL per well). Colorimetric quantification was measured the absorbance at 570 nm using a microplate reader (BioTek Instruments, VT, USA).

Physicochemical property predictions

The physicochemical features, including the number of hydrogen bond donors, hydrogen bond acceptors, and drug-likeness, play a crucial role in the drug discovery and development process. In this study, we calculated these properties for four compounds (**IQSO₂R-I**, **IQSO₂R-V**, **IQSO₂R-VI**, and **IQSO₂R-VII**) using the web-based application SwissADME (<http://www.swissadme.ch/>).

Molecular dynamic simulations

The crystal structure of erlotinib in complex with the wild-type EGFR-TK was obtained from the Protein Data Bank (PDB ID: 1M17). The **IQSO₂R-I**/EGFR-TK complex was derived by molecular docking using the GOLD program¹⁵ according to our previous study on this system.¹⁶ The best docking pose (highest GOLD fitness score) of **IQSO₂R-I** binding to EGFR-TK was considered as the starting structure for the all-atom molecular dynamics simulations with three independent runs using different initial velocities. The system was simulated under periodic boundary conditions using the isothermal–isobaric (NPT) ensemble, with a temperature of 310 K and a pressure of 1 atm as per previous studies.¹⁷ The AMBER ff14SB force field and the generalized AMBER force field version 2 (GAFF2) were employed to handle the bonded and non-bonded interaction parameters for the protein and ligand, respectively. The system was solvated in the TIP3P water model. Chloride ions were randomly introduced to neutralize the overall charge of the

system. The hydrogen atoms and water molecules were subjected to energy minimization using 500 steps of the steepest descent followed by 1500 steps of conjugated gradient methods, while the remaining molecules were kept fixed. The protein–ligand complex (constrained solvents) and the entire complex system were then subjected to further minimization, following the same procedure. Electrostatic interactions were handled using the particle mesh Ewald summation approach, and hydrogen atoms were constrained using the SHAKE algorithm. The temperature was gradually increased from 10 to 310 K using a Langevin thermostat with a collision frequency of 2 ps⁻¹, while the pressure was controlled using the Berendsen barostat. MD simulations were conducted for 500 ns with a time step increment of 2 fs. The MD outputs were analyzed using the cpptraj module, and the per-residue decomposition energy ($\Delta G_{\text{bind, residue}}$) was computed using MM/PBSA.py in AMBER20. The interactions of the potent EGFR inhibitor complexed with EGFR-TK were visualized using LigandScout 4.4.9 software.¹⁸

Conclusions

In the present work, for the first time, experimental and computational methods were employed to identify potential candidates for EGFR inhibitors based on compounds bearing indolo[1,2-*a*]quinoline as a core structure. Six 6-trifluoromethylthio indolo[1,2-*a*]quinolines and nine 6-arene-sulfonyl indolo[1,2-*a*]quinolines, were experimentally screened for their inhibitory activity against EGFR-TK; six compounds (**IQSCF₃-V**, **IQSO₂R-I**, **IQSO₂R-II**, **IQSO₂R-V**, **IQSO₂R-VI**, and **IQSO₂R-VII**) were found to inhibit EGFR-TK activity greater than 50% inhibition at 1 μM (~90% inhibition for afatinib). From a cell-based assay in human cancer cell lines with EGFR wild-type overexpression (A549 and SCC cell lines), four compounds (**IQSO₂R-I**, **IQSO₂R-V**, **IQSO₂R-VI**, and **IQSO₂R-VII**) exhibited cytotoxic activity against A549 and SCC cells as potent as afatinib, with a slightly lower toxicity in Vero cells. The MD simulation study on the most potent compound, *i.e.*, **IQSO₂R-I** with an IC₅₀ of 0.28 ± 0.05 μM, revealed nine amino acids (L718, F723, V726, K745, A743, T790, M793, G796, and L844) contributed to the formation of the **IQSO₂R-I**/EGFR complex. Therefore, **IQSO₂R-I** represents a novel type of EGFR-TK inhibitor, which may be useful as a novel scaffold for the development of anticancer therapeutics targeting EGFR-TK.

Data availability

The data supporting this article have been included as part of the ESI.†

Author contributions

CK, CM, and TR conceived and designed the experiments. PS, DT and TR conducted theoretical and experimental studies. JP, TU, and NS synthesized all compounds. CK, PS, CM, DT and TR analyzed the data. CK, PS, CM, DT and TR wrote the original manuscript. All authors reviewed and edited the manuscript. All



authors have read and agreed to the published version of the manuscript.

Conflicts of interest

Authors declare no conflicts of interest.

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References

- 1 WHO factsheet, <https://www.who.int/news-room/fact-sheets/detail/lung-cancer>, 2023.
- 2 H. Lemjabbar-Alaoui, O. U. Hassan, Y.-W. Yang and P. Buchanan, *Biochim. Biophys. Acta*, 2015, **1856**, 189.
- 3 X. Ke and L. Shen, *Front. Lab. Med.*, 2017, **1**, 69.
- 4 (a) S. Di Martino, A. Rainone, A. Troise, M. Di Paolo, S. Pugliese, S. Zappavigna, A. Grimaldi and D. Valente, Overview of FDA-approved anticancer drugs used for targeted therapy, *World Cancer Res. J.*, 2015, **2**, e553; (b) F. R. Hirsch, G. V. Scagliotti, J. L. Mulshine, R. Kwon, W. J. Curran, Y.-L. Wu and L. Paz-Ares, *Lancet*, 2017, **389**, 299.
- 5 R. Roskoski Jr, *Pharmacol. Res.*, 2014, **79**, 34.
- 6 (a) H. Ogiso, R. Ishitani, O. Nureki, S. Fukai, M. Yamanaka, J. H. Kim, K. Saito, A. Sakamoto, M. Inoue, M. Shirouzu and S. Yokoyama, *Cell*, 2002, **110**, 775; (b) C. H. Yang, H. C. Chou, Y. N. Fu, C. L. Yeh, H. W. Cheng, I. C. Chang, K. J. Liu, G. C. Chang, T. F. Tsai, S. F. Tsai, H. P. Liu, Y. C. Wu, Y. T. Chen, S. F. Huang and Y. R. Chen, *Biochim. Biophys. Acta*, 2015, **1852**, 1540; (c) F. R. Hirsch, M. Varella-Garcia and F. Cappuzzo, *F. Oncogene*, 2009, **28**, S32.
- 7 V. P. Reddy, Organofluorine Compounds as Anticancer Agents, in *Organofluorine Compounds in Biology and Medicine*, Elsevier, 2015.
- 8 (a) J. P. Michael, *Nat. Prod. Rep.*, 2008, **25**, 139; (b) A. F. Pozharskii, A. T. Soldatenkov and A. R. Katritzky, *Heterocycles in Life and Society: an Introduction to Heterocyclic Chemistry, Biochemistry and Applications*, John Wiley & Sons, 2011; (c) T. Eicher, S. Hauptmann and A. Speicher, *The Chemistry of Heterocycles: Structure, Reactions, Syntheses, and Applications*, Wiley-VCH, 2013; (d) C. T. Walsh, *Tetrahedron Lett.*, 2015, **56**, 3075; (e) C. Cabrele and O. Reiser, *J. Org. Chem.*, 2016, **81**, 10109; (f) A. P. Taylor, R. P. Robinson, Y. M. Fobian, D. C. Blakemore, L. H. Jones and O. Fadeyi, *Org. Biomol. Chem.*, 2016, **14**, 6611; (g) M. M. Heravi and V. Zadsirjan, *RSC Adv.*, 2020, **10**, 44247.
- 9 (a) D. G. Hulcoop and M. Lautens, *Org. Lett.*, 2007, **9**, 1761; (b) A. K. Verma, S. P. Shukla, J. Singh and V. Rustagi, *J. Org. Chem.*, 2011, **76**, 5670; (c) S. P. Shukla, R. K. Tiwari and A. K. Verma, *J. Org. Chem.*, 2012, **77**, 10382; (d) K. Sun, X. L. Chen, Y. L. Zhang, K. Li, X. Q. Huang, Y. Y. Peng, L. B. Qu and B. Yu, *Chem. Commun.*, 2019, **55**, 12615; (e) R. Heckershoff, G. May, J. Däumer, L. Eberle, P. Krämer, F. Rominger, M. Rudolph, F. F. Mulks and A. S. K. Hashmi, *Chem.-Eur. J.*, 2022, **28**, e202201816; (f) T. Uppalabat, N. Hassa, N. Sawektreeratana, P. Leowanawat, P. Janthakit, P. Nalaoh, V. Promarak, D. Soorukram, V. Reutrakul and C. Kuhakarn, *J. Org. Chem.*, 2023, **88**, 5403; (g) S. Zhang, J. Yuan, G. Huang, C. Ma, J. Yang, L. Yang, Y. Xiao and L. Qu, *J. Org. Chem.*, 2023, **88**, 11712.
- 10 K. Hengphasatporn, T. Aiebchun, P. Mahalapbutr, A. Auepattanapong, O. Khaikate, K. Choowongkamon, C. Kuhakarn, J. Meesin, Y. Shigeta and T. Rungrotmongkol, *ACS Omega*, 2023, **8**, 19645.
- 11 A. Daina, O. Michielin and V. Zoete, *Sci. Rep.*, 2017, **7**, 42717.
- 12 C. A. Lipinski, *Drug Discovery Today: Technol.*, 2004, **1**, 337.
- 13 (a) G. P. Doss, B. Rajith, C. Chakraborty, N. NagaSundaram, K. Ali and H. Zhu, *Sci. Rep.*, 2014, **4**, 5868; (b) F. Martínez-Jiménez, J. P. Overington, B. Al-Lazikani and M. A. Marti-Renom, *Sci. Rep.*, 2017, **7**, 46632; (c) J. Stamos, M. X. Sliwkowski and C. Eigenbrot, *J. Biol. Chem.*, 2002, **277**, 46265.
- 14 W. Nie, L. Tang, H. Zhang, J. Shao, Y. Wang, L. Chen, D. Li and X. Guan, *Int. J. Oncol.*, 2012, **40**, 1763.
- 15 M. L. Verdonk, J. C. Cole, M. J. Hartshorn, C. W. Murray and R. D. Taylor, *Proteins*, 2003, **52**, 609.
- 16 (a) D. Todsaporn, A. Zubenko, V. G. Kartsev, P. Mahalapbutr, A. Geronikaki, S. N. Sirakanyan, L. N. Divaeva, V. Chekrisheva, I. Yildiz, K. Choowongkamon and T. Rungrotmongkol, *J. Phys. Chem. B*, 2024, **128**, 12389; (b) T. Aiebchun, P. Mahalapbutr, A. Auepattanapong, O. Khaikate, S. Seetaha, L. Tabtimmai, C. Kuhakarn, K. Choowongkamon and T. Rungrotmongkol, *Molecules*, 2021, **26**, 2211.
- 17 (a) D. Todsaporn, A. Zubenko, V. Kartsev, T. Aiebchun, P. Mahalapbutr, A. Petrou, A. Geronikaki, L. Divaeva, V. Chekrisheva, I. Yildiz, K. Choowongkamon and T. Rungrotmongkol, *Molecules*, 2023, **28**, 3014; (b) D. Todsaporn, P. Mahalapbutr, R. P. Poo-arporn, K. Choowongkamon and T. Rungrotmongkol, *Comput. Biol. Med.*, 2022, **147**, 105787.
- 18 G. Wolber and T. Langer, *J. Chem. Inf. Model.*, 2005, **45**, 160.

