

PAPER

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Iron–copper complexes have been extensively studied in the search for efficient cytochrome c oxidase models. Whereas most dinuclear materials usually focus on fine-tuning the coordination of heme-Fe, this work shows that the coordination of copper in cytochrome c oxidase models should be carefully taken into consideration. A β -cyclodextrin dimer was built around a bipyridine linker and combined with Fe-tetraphenylsulfonatoporphyrinate (**FeTPPS**) to generate a self-assembled hydrosoluble cytochrome c oxidase model. Cyclic voltammetry and rotating ring disk electrode experiments showed that this model with a tetrahedral coordination of copper(I) is efficient for the reduction of molecular oxygen with an average of 3.6 electrons indicating a preference and efficiency for the four-electron reduction to water.

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Introduction

The reduction of molecular dioxygen by cytochrome c oxidase (CcO) is the terminal step of the respiratory process. The active center responsible for the 4-electron reduction of oxygen comprises an iron (Fe_{a3}) and a copper (Cu_B) in various oxidation states.¹ As shown by the original solid-state structure reported by Yoshikawa, the iron is bound to a heme topped by a binding site for copper that contains three histidines, one of which is coupled to a tyrosine.^{2–4} Although several models have successfully reproduced either the function or the structure of various states of the enzyme, non-systematic variation of the molecular architectures used to position a copper ion above a iron porphyrinate hampers the rationalization of the parameters governing the efficiency of the models.^{5–8} Our groups previously reported a porphyrin-cyclodextrin scaffold [Fe^{III}TPPS/Cu^{II}TerpyCD₂][–] (Fig. 1) which, after reduction to its

Fe^{II}Cu^I derivative, is able to reduce oxygen in aqueous medium with a 50% preference for the 4-electron pathway.⁹ At that time, a terpyridine was chosen as the ligand for copper at the distal site. This ligand was introduced as a linker between two permethylated β -cyclodextrins (CDs) that encapsulate an iron tetraphenylsulfonatoporphyrin (**FeTPPS**). Although terpyridine forms stable complexes with Cu^{II}, its Cu^I complexes are less stable because the tridentate, planar terpyridine ligand does not allow Cu^I to adopt its preferred tetrahedral geometry, rendering it more susceptible to disproportionation¹⁰ or oxidation. We suspected that the poor stability of the Cu^I complex might influence O₂ reduction. Hence, the terpyridine bridge between two CDs has now been replaced with a 2,2'-bipyridine, which is capable of forming stable complexes with both oxidation states of copper. Once combined with an **FeTPPS** core, the resulting supramolecular assembly [Fe^{III}TPPS/BipyCD₂]^{3–} allows a direct comparison between bi- and tridentate ligands for copper in CcO models.

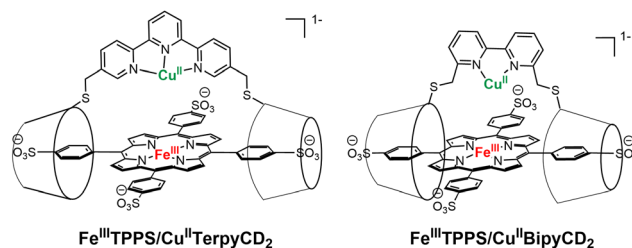


Fig. 1 Porphyrin- β -CD₂ assemblies previously reported (left) and studied (right).

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Results and discussion

The permethylated β -cyclodextrin dimer (**BipyCD₂**) was prepared by reacting dithiol derivative^{11,12} **1** with the epoxy derivative^{13,14} **2** of the permethylated β -cyclodextrin to yield the cyclodextrin dimer **BipyCD₂** (Scheme 1). This dimer was purified by column chromatography and then by steric exclusion chromatography and isolated in 43% yield.

The ability of the **BipyCD₂** to bind **FeTPPS**^{3−}, as schematically illustrated in Fig. 1, was probed by titration in an aqueous phosphate buffer (pH = 7). As shown in the UV-visible spectra in Fig. 2, addition of the **BipyCD₂** dimer to **FeTPPS**^{3−} induces a significant red shift of the Soret band of the porphyrin, from 406 to 419 nm. This change is concomitant with the appearance of an absorption at 296 nm, which is typical of the CD-dimer, and with a red shift of the Q bands from 566 to 574 nm and from 608 to 620 nm. A hyperchromic effect is also observed for the Q bands. These changes continue until the equivalence point is reached. Further addition of **BipyCD₂** resulted only in the growth of the absorption band of **BipyCD₂** at 296 nm.

All of the changes in the absorption spectrum of **FeTPPS** are consistent with the reported formation of an inclusion complex in CD-dimers.¹⁴ The association constant for the formation of the inclusion complex [**FeTPPS/BipyCD₂**]^{3−} was too high to be determined from the UV-vis titration. A rough estimation based on reported methods¹⁵ yields a value in the 10⁷ M^{−1} range. This inclusion complex is extremely stable in aqueous solutions. In the negative electrospray ionization (ESI[−]) spectrum of the inclusion complex, the isotopic profile of the major peak at m/z = 1004.0844 is consistent with a species in which an axial hydroxyl ligand is bound to the iron [**Fe(OH)TPPS/BipyCD₂**]^{4−} (see ESI[†]). Additionally, a peak at m/z = 1333.1348 corresponding to the same compound without the

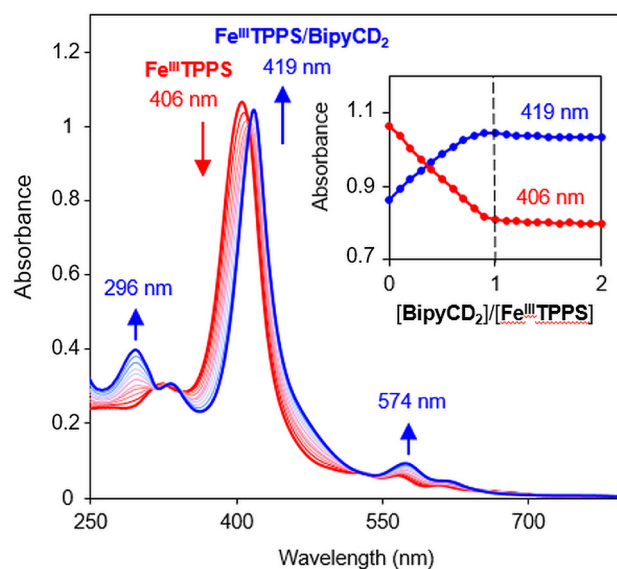
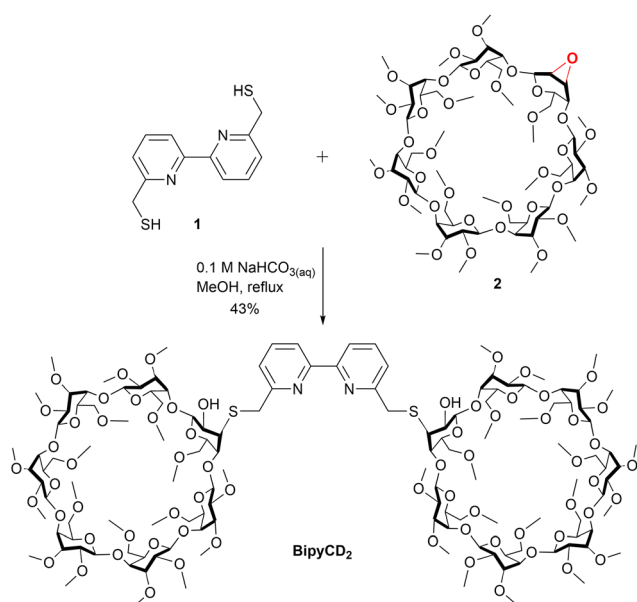


Fig. 2 UV-visible titration of **Fe^{III}TPPS** (10 μ M) in 0.05 M phosphate buffer with the stepwise addition of **BipyCD₂** in phosphate buffer. The inset shows changes in absorbance as a function of [**BipyCD₂**]/[**Fe^{III}TPPS**] concentrations.

axial ligand [**FeTPPS/BipyCD₂**]^{3−} was observed. These results suggest that, in aqueous medium, the iron core in [**Fe(OH)TPPS/BipyCD₂**]^{4−} is probably in a pentacoordinated high-spin state with an axial hydroxyl ligand. X-band electron paramagnetic resonance (EPR) confirmed this hypothesis, showing effective g values of $g_{\perp}$ = 5.68 and $g_{\parallel}$ = 1.99 at 25 K at pH = 7 in a phosphate buffer (*vide infra*). These results fully agree with literature reports.^{16–18} The ligands in the coordination sphere for the iron core of the porphyrin can be pH dependent. Thus, UV-visible monitored pH titration studies were performed on [**FeTPPS/BipyCD₂**]^{3−} in a sodium perchlorate buffer to explore a wider range of pH values (2 to 12) with non-coordinating counter ions (Fig. 3). Initially, the pH of the solution of [**FeTPPS/BipyCD₂**]^{3−} was adjusted to 2 by the addition of 0.1 M perchloric acid. Then, the pH was varied by stepwise addition of a NaOH solution. At pH higher than 5.2, a spectrum identical to that of [**Fe(OH)TPPS/BipyCD₂**]^{4−} in Fig. 2 (blue line) was observed and is consistent with literature data.¹⁸ According to the same reports, the species present in solution at pH < 5.2 display a Soret band at 400 nm that is characteristic of the low spin, hexacoordinated bis-aquo complex [**Fe(OH₂)₂TPPS/BipyCD₂**]^{3−}. In contrast to the spectrum of the complex in phosphate buffer (Fig. 2, blue line), at pH 2, only one absorption band at 328 nm is observed in the UV region. This band was attributed to a protonated bipyridine, for which a pK_a of 2.7 was estimated. The reversible nature of this spectral change in the UV upon addition of base (comparison of the spectra in blue in Fig. 2 and 3) indicates that the thioether bipyridine-CD linkage remains intact at low pH.

Having confirmed the formation of the inclusion complex with the porphyrin heme model, the insertion of copper in the



Scheme 1 Synthesis of the bipyridine-bridged CDs.



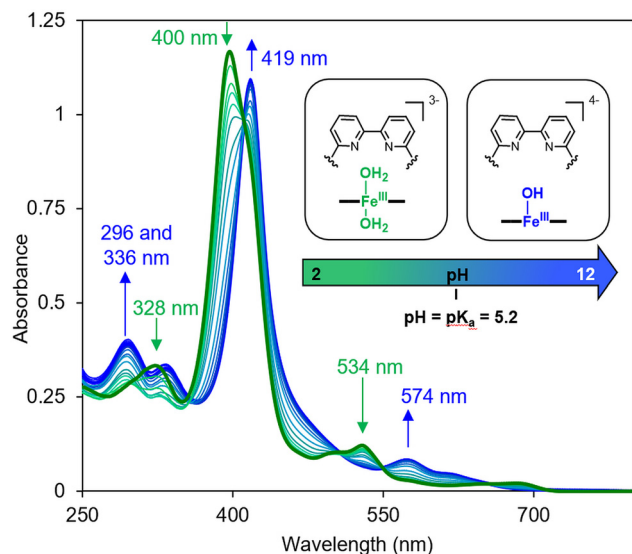


Fig. 3 UV-visible spectral changes of $\text{Fe}^{\text{III}}\text{TPPS}/\text{BipyCD}_2$ (10 μM) in aqueous 0.1 M NaClO_4 as a function of the pH of the solution from 2 to 12. The pH was adjusted by addition HClO_4 and NaOH .

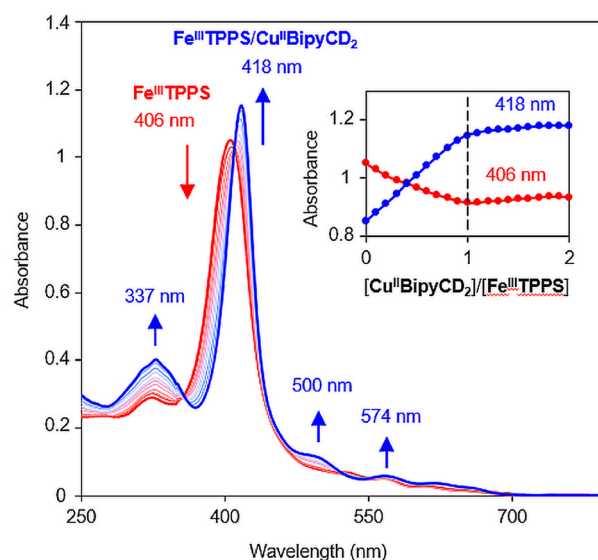


Fig. 4 UV-visible titration of $\text{Fe}^{\text{III}}\text{TPPS}$ (10 μM) in 0.05 M phosphate buffer with the stepwise addition of $\text{Cu}^{\text{II}}\text{BipyCD}_2$ in water. The inset shows changes in absorbance as a function of ratio of the concentrations $[\text{Cu}^{\text{II}}\text{BipyCD}_2]/[\text{Fe}^{\text{III}}\text{TPPS}]$.

CuO model was then studied. Two approaches were possible to generate iron-copper complexes. Unfortunately, the first strategy consisting of insertion of copper(II or I) in the previously described inclusion complex $[\text{Fe}^{\text{III}}\text{TPPS}/\text{BipyCD}_2]^{3-}$ was unsuccessful. On one hand, the instability of Cu^{I} in aqueous media makes experiments with Cu^{I} difficult to reproduce. On the other hand, although the addition of Cu^{II} to the inclusion complex $[\text{Fe}^{\text{III}}\text{TPPS}/\text{BipyCD}_2]^{3-}$ seemed appropriate, the use of a phosphate buffer was precluded because interactions between phosphate anions and Cu^{II} gave a water-insoluble ion pair that precipitated. Although no precipitation occurs in the absence of phosphate buffer, the approach described in the next paragraph was preferred.

A second method was thus carried out in which $\text{Fe}^{\text{III}}\text{TPPS}$ in phosphate buffer was added in small aliquots to a solution of pre-formed $[\text{Cu}^{\text{II}}\text{BipyCD}_2]^{2+}$ in pure water. First, the $[\text{Cu}^{\text{II}}\text{BipyCD}_2]^{2+}$ complex was formed by adding one equivalent of a solution of $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$ in Milli-Q water to a solution of BipyCD_2 in Milli-Q water. The formation of $[\text{Cu}^{\text{II}}\text{BipyCD}_2]^{2+}$ was monitored by absorption spectroscopy in the UV region (see ESI†), where spectral changes were consistent with the stoichiometric binding of Cu^{II} to the bipyridyl bidentate with an approximate binding constant of $3 \times 10^6 \text{ M}^{-1}$ in water at pH 7. Mass spectrometry of a stoichiometric mixture of BipyCD_2 and $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$ in Milli-Q water also confirmed the coordination of copper to the bipyridine (see ESI†).

With the copper bipyridine complex in hand, the $[\text{Fe}^{\text{III}}\text{TPPS}/\text{Cu}^{\text{II}}\text{BipyCD}_2]^-$ inclusion complex could be formed in phosphate buffer, and no precipitation occurred. From the titration curves in Fig. 4, a binding constant greater than 10^7 M^{-1} was determined, thus confirming the stability of the supramolecular scaffold. In addition to confirming the formation of a heme-copper complex, mass spectrometry pro-

vided complementary information regarding the coordination spheres of both metals. ESI in negative mode showed two major peaks that correspond to the unbound porphyrin $[\text{Fe}^{\text{III}}\text{TPPS}]^{3-}$ and to the porphyrin bearing an axial hydroxyl ligand $[\text{Fe}^{\text{III}}(\text{OH})\text{TPPS}]^{4-}$ (see ESI†). Several other peaks suggested the formation of a μ -oxo $\text{Fe}^{\text{III}}\text{Cu}^{\text{II}}$ derivative. This type of bridged species has been observed in organic solvents by many authors in the past.^{19–22}

Because this work aimed at O_2 reduction under physiological conditions, it was necessary to determine the predominant species at neutral pH. The existence of μ -oxo species is pH dependent; therefore, a pH titration of the $\text{Fe}^{\text{III}}\text{Cu}^{\text{II}}$ species was performed. A solution of the $[\text{Fe}^{\text{III}}\text{TPPS}/\text{Cu}^{\text{II}}\text{BipyCD}_2]^-$ complex in a 0.1 M perchlorate buffer was prepared and the pH was adjusted to 2 with a dilute perchloric acid solution. Then, the pH was varied by addition of an aqueous solution of NaOH . Four species were identified by comparison with literature data^{9,18,21–23} (Fig. 5).

At acidic pH, a first species, with a Soret band at 398 nm, is assigned to the low spin inclusion complex $[\mu\text{-(H}_2\text{O)-Fe}^{\text{III}}\text{TPPS}/\text{Cu}^{\text{II}}(\text{H}_2\text{O})\text{-BipyCD}_2]^-$ bearing two molecules of water in axial positions. This assignment is in agreement with a recent report describing a μ -aquo $\text{Fe}^{\text{III}}\text{Cu}^{\text{II}}$ species in organic solvent formed in the presence of $[\text{H}(\text{OEt}_2)_2][\text{B}(\text{C}_6\text{F}_5)_4]$.²³ This species is dominant over the pH range of 2 to 5.3 but could not be characterized by paramagnetic NMR due to the complexity of the spectrum. Above pH 5.3, a new species appears (blue curves, Fig. 5) that is very similar to that of the $[\text{HO-Fe}^{\text{III}}\text{TPPS}/\text{BipyCD}_2]^{4-}$ and can be assigned to the formation of $[\mu\text{-(HO)-Fe}^{\text{III}}\text{TPPS}/\text{Cu}^{\text{II}}(\text{H}_2\text{O})\text{-BipyCD}_2]^{2-}$ by the loss of one proton from the bridging water molecule. This species dominates from pH 5.3–7.6. Above pH 7.6, a new species



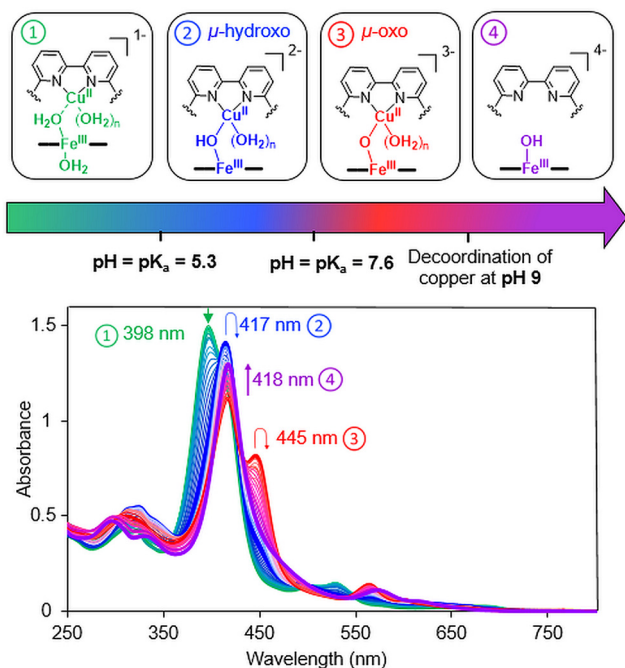


Fig. 5 UV-visible spectral changes of $[\text{Fe}^{\text{III}}\text{TPPS}/\text{Cu}^{\text{II}}\text{BipyCD}_2]$ (14 μM) in aqueous 0.1 M NaClO_4 as a function of the pH of the solution from 2 to 12. The pH was adjusted by addition HClO_4 and NaOH .

appears (red curves, Fig. 5) and shows a Soret band at 445 nm, which, according to the work of Karlin and coworkers, is typical of μ -oxo species.^{21,22} This species is dominant in solution in the range from $7.6 < \text{pH} < 9$. At pH higher than 9, a species is observed with a Soret band at 418 nm and two other absorption bands at 296 and 336 nm (purple curves, Fig. 5). The two bands in the UV region are typical of the bipyridine ligand without copper, confirming that at $\text{pH} > 9$ the bipyridine strap releases the copper ion. In conclusion, the dominant species at pH 7 is the $[\mu(\text{HO})\text{-Fe}^{\text{III}}\text{TPPS}/\text{Cu}^{\text{II}}(\text{H}_2\text{O})\text{-BipyCD}_2]^{2-}$ and at lower pH the bridging hydroxyl may be protonated.

The paramagnetic nature of both the ferric and cupric ions prevented structural investigations by routine NMR, therefore EPR experiments were performed. To characterize the coordination sphere of Cu^{II} in our supramolecular assembly, two samples of $[\text{Cu}^{\text{II}}\text{BipyCD}_2]^{2+}$, either in pure Milli-Q water or in buffer solution were studied by X-band EPR spectroscopy at 25 K (Fig. 6a). Each sample was prepared by mixing stoichiometric amounts of $[\text{Fe}^{\text{III}}\text{TPPS}/\text{BipyCD}_2]^{3-}$ and $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$ in water in the absence of phosphate buffer. In Milli-Q water, the pH was adjusted to 7 with dilute acidic and alkaline solutions, whereas for the buffered solution, the stoichiometric mixture was evaporated to dryness and redissolved in a 0.025 M phosphate buffer.

In pure water, a Cu^{II} species with a rhombic symmetry is in very good agreement with a simulated spectrum for $g_x = 2.18$, $g_y = 2.06$ and $g_z = 2.03$. The fact that $g_z > g_x$ and g_y indicates that the lone electron is located in the $d_{x^2-y^2}$ orbital.

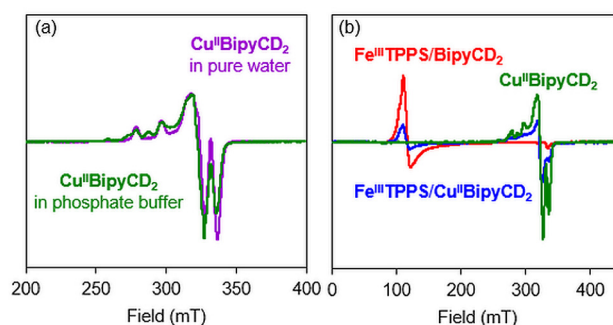


Fig. 6 EPR spectra at 25 K of (a) $[\text{Cu}^{\text{II}}\text{BipyCD}_2]^{2+}$ (0.5 μM) in water (purple line) or in 0.025 μM phosphate buffer at pH 7 (green line) and (b) $[\text{Fe}^{\text{III}}\text{TPPS}/\text{BipyCD}_2]^{3-}$ (red line), $[\text{Cu}^{\text{II}}\text{BipyCD}_2]^{2+}$ (green line) and $[\text{Fe}^{\text{III}}\text{TPPS}/\text{Cu}^{\text{II}}\text{BipyCD}_2]^{2-}$ (blue line) in 0.025 M phosphate buffer at pH 7.

According to the literature, these parameters are in agreement with elongated octahedral (Jahn–Teller distortion), square pyramidal or square planar geometries.²⁴ Considering that the bipyridine is a bidentate ligand, we can deduce that two to four molecules of water surround the cupric ion in the complex. In buffered solution, the two species observed correspond to the previously observed species of rhombic symmetry and a second species of axial or rhombic symmetry. The latter species must correspond to a species with a phosphate coordinated to Cu^{II} . No hyperfine coupling constants were extracted from the data.

Similar experiments were then recorded for the $\text{Fe}^{\text{III}}\text{-Cu}^{\text{II}}$ species $[\mu(\text{HO})\text{-Fe}^{\text{III}}\text{TPPS}/\text{Cu}^{\text{II}}(\text{H}_2\text{O})\text{-BipyCD}_2]^{2-}$ in phosphate buffer (Fig. 6b). The EPR spectrum differs strikingly from the EPR-silent behavior of cytochrome *c* oxidase in its oxidized state.²⁵ Indeed, two signals corresponding to the ferric and the cupric ions are observed in the spectrum of $[\mu(\text{HO})\text{-Fe}^{\text{III}}\text{TPPS}/\text{Cu}^{\text{II}}(\text{H}_2\text{O})\text{-BipyCD}_2]^{2-}$. However, the superposition of the separate Cu and Fe complexes with that of the binuclear species at identical concentrations (Fig. 6b) clearly shows the weakening of both the Cu^{II} and the Fe^{III} signals, suggesting that these two ions undergo an antiferromagnetic coupling through a μ -hydroxo ligand. Such a coupling was also observed for the terpyridine analogue.⁹ Thus, the formation of the supramolecular CcO model $[\text{Fe}^{\text{III}}\text{TPPS}/\text{Cu}^{\text{II}}\text{BipyCD}_2]^{2-}$ has been established and, at $\text{pH} = 7$, the dominant species is a μ -hydroxo bridged dinuclear species $[\mu(\text{HO})\text{-Fe}^{\text{III}}\text{TPPS}/\text{Cu}^{\text{II}}(\text{H}_2\text{O})\text{-BipyCD}_2]^{2-}$.

Once the nature of the resting state present at physiological pH was established, the reactivity of the reduced species was investigated. First, the copper-free model $[\text{Fe}^{\text{III}}\text{TPPS}/\text{BipyCD}_2]^{3-}$ was reduced with an excess of sodium dithionite in a degassed 0.05 M phosphate buffer under argon. The UV-visible spectrum of $[\text{Fe}^{\text{II}}\text{TPPS}/\text{BipyCD}_2]^{4-}$ (black spectrum, Fig. 7) showed a hypsochromic shift of the Q band from 573 nm to 555 nm characteristic of the formation of a reduced, iron(II) complex.²⁶ In addition, a strong hyperchromic effect and bathochromic shift of the Soret band, from 420 to 432 nm, are consistent with the formation of a pentacoordinated, high spin iron(II) complex, as seen in previous por-



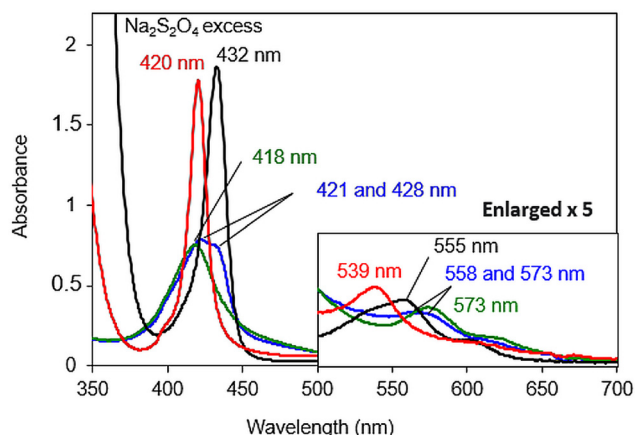


Fig. 7 UV-visible spectra of 8 μM solutions of the [Fe^{III}TPPS/BipyCD₂][−] complex (green line), its reduced Fe^{II} (black line), oxygenated Fe^{III}–O₂^{•−} (blue line) and carbonylated Fe^{II}–CO (red line) products in 0.05 M phosphate buffer at pH 7. The inset shows an enlargement of the Q-band region.

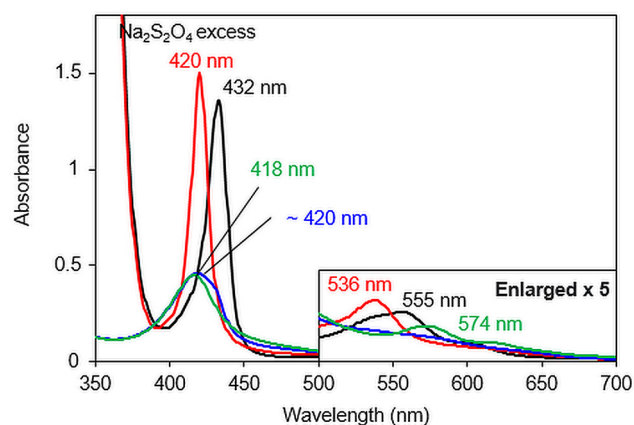


Fig. 8 UV-visible spectra of 6 μM solutions of the [Fe^{III}TPPS/Cu^IBipyCD₂][−] complex (green line), its reduced Fe^{II}/Cu^I (black line), oxygenated Fe^{III}–O₂^{•−}/Cu^I (blue line) and carboxylated Fe^{II}–CO/Cu^I (red line) products in 0.05 M phosphate buffer at pH 7. The inset shows an enlargement of the Q-band region.

phyrin/cyclodextrin dimer inclusion complexes.¹⁴ Overall, the UV-vis spectrum is very similar to that of deoxymyoglobin, which indicates that the Fe^{II} center in the model without copper is a pentacoordinated, high-spin complex, for which the fifth ligand is probably a water molecule.

The carbonmonoxy species was generated by bubbling CO into a solution of the iron(II) complex [Fe^{II}TPPS/BipyCD₂]^{4−}. A spectrum similar to that of carboxymyoglobin is observed, with a blue-shifted Soret band at 420 nm and a blue-shifted Q band at 539 nm (red spectrum, Fig. 7). These spectral changes correspond to a hexacoordinated low spin Fe^{II} in which a CO as well as a hydroxyl ligand are coordinated to the iron center.^{9,14}

To form an oxygenated species, the excess sodium dithionite was first removed from a solution of [Fe^{II}TPPS/BipyCD₂]^{4−} by chromatography of the solution over Sephadex G-25^{9,27} under argon. The reduced sample was then mixed in a UV-visible cell with an oxygen-saturated buffer at 10 °C. Under these conditions, the spectrum obtained shows the existence of two species with Soret bands at 421 (major) and 430 nm (minor), (blue spectrum, Fig. 7 top), suggesting that the reduced species (Soret at 430 nm) is only partially oxygenated. The presence of two Q bands at 558 and 573 nm provides further confirmation of a mixture of these two species.

The influence of copper at the distal site of the heme was also studied by UV-visible spectroscopy. The spectra of reduced species [Fe^{II}TPPS/Cu^IBipyCD₂]^{3−} (Fig. 8 black line) and its CO adduct (Fig. 8 red line) are similar to those observed in the copper-free complex (Fig. 7) indicating the same coordination numbers and spins of the Fe^{II} centers in the absence or presence of copper. For the CO adduct [Fe^{II}TPPS-CO/Cu^IBipyCD₂]^{3−}, a 3 nm blue shift of its highest energy Q band (536 nm) compared to that of the copper-free complex (539 nm, shown in Fig. 7) suggests a distortion of the CO coordination to the iron, possibly due to a hindered distal

site²⁸ in the presence of copper. The oxygenated species, prepared as described above, displays a Soret band at 420 nm but no distinct Q bands. The absence of a distinct Q band was attributed to the presence of a mixture of several species including the reduced complex [Fe^{II}TPPS/Cu^IBipyCD₂]^{3−}, the superoxo species [Fe^{III}TPPS-O₂^{•−}/Cu^IBipyCD₂]^{3−} and/or the peroxo species [Fe^{III}TPPS-O₂/Cu^IBipyCD₂]^{3−}. Nevertheless, the Soret band at 420 nm appears at a wavelength similar to that of the superoxo terpyridine-bridged complex⁹ [Fe^{III}TPPS-O₂^{•−}/Cu^ITerpyCD₂]^{3−}.

The electrochemical properties of the complexes were investigated by cyclic voltammetry (CV) in homogeneous, neutral aqueous solution with a phosphate buffer at pH 7 with Na₂SO₄ as supporting electrolyte and deposited on a rotating ring disk electrode (RRDE) and rotating disk electrode (RDE). In solution, as previously observed with an inclusion complex of Mn (TPPS) bound within two permethylated-β-CDs,²⁹ no signal was observed for the FeTPPS once inserted in the cyclodextrin dimer BipyCD₂. With the iron/copper complex [Fe^{III}TPPS/Cu^IBipyCD₂][−], cyclic voltammetry showed a weak reduction signal for the Cu^{III/I} couple and the Fe^{III/II} couple. A broad reduction peak (*E*_p = −0.06 V vs. SCE) of the copper complex indicates an ill-defined coordination sphere around the Cu^{II}, as seen earlier with the coordination of multiple water molecules for example. A sharper signal for the re-oxidation of Cu^I to Cu^{II} suggests a better-defined coordination sphere for Cu^I in [Fe^{II}TPPS/Cu^IBipyCD₂]^{3−}. Nevertheless, the voltammograms were reproducible and showed no change after 10 cycles. This reproducibility confirms that the supramolecular model withstands the redox processes without decomposition or precipitation at the electrode.

In an O₂-saturated electrolyte, the cyclic voltammetry shows a catalytic wave that is facilitated for [Fe^{II}TPPS/Cu^IBipyCD₂]^{3−} (*E*_p = −0.55 V vs. SCE) compared to [Fe^{II}TPPS/BipyCD₂] (*E*_p = −0.75 V vs. SCE) (see ESI†). In the presence of copper, this



Table 1 Electrocatalytic properties of [Fe^{III}TPPS/BipyCD₂] and [Fe^{III}TPPS/Cu^{II}BipyCD₂] compared to those of [Fe^{III}TPPS/TerpyCD₂] and [Fe^{III}TPPS/Cu^{II}TerpyCD₂]

Catalyst	E_{pc} (V vs. SCE)	E_{ponset} (V vs. SCE)	n_{RRDE}^a	n_{KL}^b	K_{app}^c (M s) ⁻¹
[Fe ^{III} TPPS/BipyCD ₂]	-0.75	-0.20	3.5	3.47	2.1×10^{-5}
[Fe ^{III} TPPS/Cu ^{II} BipyCD ₂]	-0.55	-0.10	3.6	3.55	3.0×10^{-5}
[Fe ^{III} TPPS/TerpyCD ₂]	—	-0.24 ^d	—	1.63 ^d	—
[Fe ^{III} TPPS/Cu ^{II} TerpyCD ₂]	—	-0.20 ^d	—	3.03 ^d	—

^a Number of electrons exchange during the reduction of O₂ by the catalyst at -1 V determined from eqn (1). ^b Number of electrons exchange during the reduction of O₂ by the catalyst at -0.6 V determined from eqn (2). ^c Determined from eqn (3). ^d Taken from ref. 9; calculated at -1.2 V.

gain of potential clearly demonstrates the contribution of copper in the reduction of O₂. Compared to the previous [Fe^{III}TPPS/Cu^{II}TerpyCD₂]³⁻ inclusion complex (see Table 1), the new iron/copper [Fe^{III}TPPS/Cu^{II}BipyCD₂]³⁻ model shows a lower onset potential ($E_{\text{ponset}} = -0.1$ V vs. SCE), suggesting that O₂ reduction is facilitated because of a better adapted coordination sphere for copper in the latter model.⁴ As this work aimed at studying the effects of the copper coordination sphere on the efficiency of the catalysis, RRDE experiments were performed to determine the number of electrons involved in the catalytic reduction of O₂.

In RRDE experiments, the 4-electron reduction of oxygen is performed by the catalyst deposited on the central, edge oriented pyrolytic graphite disk while the potential of the peripheral ring, towards which the reduction products diffuse due to the rotation of the electrode, is set at 0.8 V per SCE to re-oxidize H₂O₂ should it be produced by a competing 2-electron process. To prevent desorption and dissolution of the water-soluble catalysts, the catalysts were fixed on the pyrolytic graphite disk with Nafion® as in our previous work.⁹ Using the RRDE setup, the number of electrons exchanged can be calculated from eqn (1):³⁰

$$n = \frac{4I_d N_c}{I_d N_c + I_r} = \frac{4 - 2\text{H}_2\text{O}_2\%}{100} \quad (1)$$

where n = the average number of electrons exchanged (comprised between 2 and 4), I_d = the maximum current intensity for the disk, I_r = the maximum current intensity for the ring, and N_c = the collection factor (characteristic of the electrode and determined experimentally). The collection factor (N_c) can be determined experimentally by calculating the ratio of the plateau currents of the ring (I_r) over the disk (I_d) for the non-catalyzed reduction of oxygen on the disk, using a pyrolytic graphite electrode where oxygen is exclusively reduced to H₂O₂ via a 2-electron pathway. The ratio between I_r and I_d was experimentally determined to be 0.21, meaning that 21% of the H₂O₂ that is produced is detected at the ring.

All RRDE experiments were performed by depositing 2.5 nmol of the supramolecular models [Fe^{III}TPPS/BipyCD₂]³⁻ and [Fe^{III}TPPS/Cu^{II}BipyCD₂]³⁻ as their sodium salts on the disk and covering them with a 10 μ L Nafion membrane in an oxygen-saturated phosphate buffer containing 0.5 M Na₂SO₄ as supporting electrolyte. The currents recorded for O₂ as a reference, [Fe^{III}TPPS/BipyCD₂]³⁻ and [Fe^{III}TPPS/Cu^{II}BipyCD₂]³⁻ are

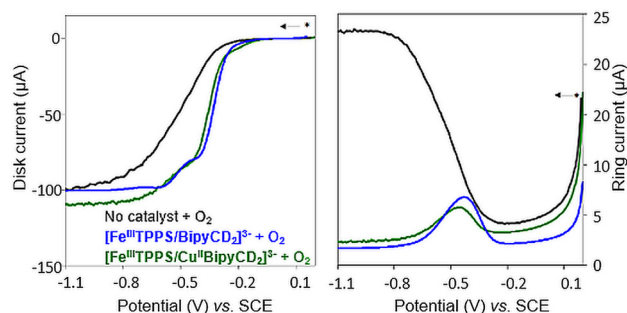


Fig. 9 Voltammograms measured by RRDE experiments of the various complexes (10 μ L of a 2.5 mM solution) coated with Nafion (10 μ L) on the disk electrode in O₂-saturated solution, measured at pH 7 in 0.05 M phosphate buffer containing 0.5 M Na₂SO₄ at a scan rate of 0.02 V s⁻¹ and 100 rpm.

plotted in Fig. 9 for both the disk (left traces) and the ring (right traces) currents. The RRDE experiments clearly show that, despite the minor gain in terms of potential, the reduction of oxygen by the catalysts immobilized on the disk produces a minimum amount of hydrogen peroxide. From the ratio of ring and disk currents, the number of electrons (n_{RRDE} in Table 1) involved in the reduction of oxygen is respectively 3.5 with the iron-only complex and 3.6 with the iron-copper complex. These numbers compare favorably with the 3.03 average number of electrons measured with the reported terpyridine analogue (Table 1).⁹ This first observation suggests that the reduction of oxygen occurs through a preferred 4-electron pathway but complementary RDE experiments were necessary to confirm the number of electrons exchanged at the electrode.

In these complementary experiments, for a given amount of catalyst deposited on the disk, the rotation speed of the electrode was systematically varied, and the reciprocal intensity of the corresponding current I_m was plotted against a function of the angular rotation rate of the electrode: $I_m^{-1} = f(\omega^{-1/2})$. The resulting electrochemical response and the associated Koutecký-Levich plots are represented in Fig. 10. For comparison, a 2-electron reference plotted from the experimental reduction of oxygen on the graphite electrode without catalyst, and a 4-electron reference plot simulated from the previous reference are also shown. For both catalysts, the average number of electrons (n_{KL}) involved in the electrocatalytic



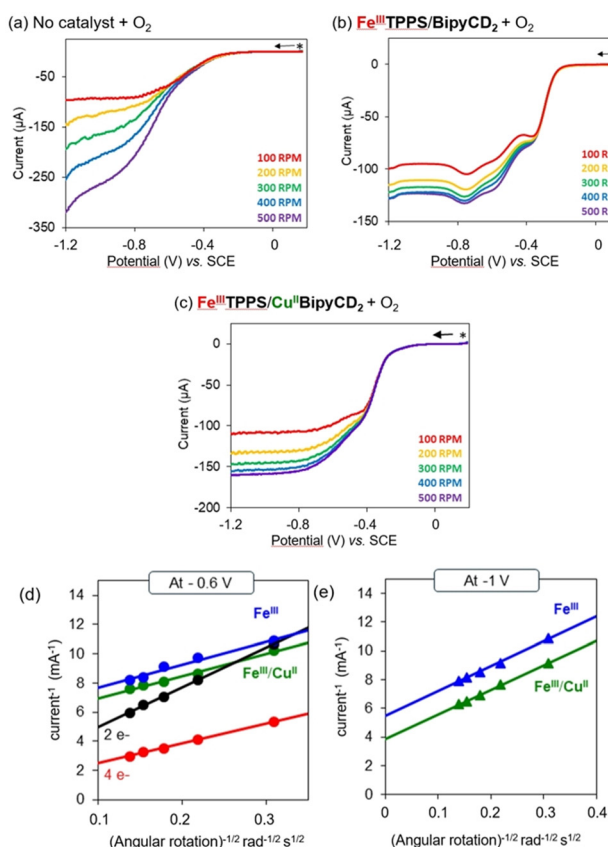


Fig. 10 (a–c) Stationary voltammograms at various rotation speeds on a RDE modified with (a) O_2 reference with no catalyst, (b) $[\text{Fe}^{\text{III}}\text{TPPS}/\text{BipyCD}_2]^{3-}$ and coated with Nafion or (c) $[\text{Fe}^{\text{III}}\text{TPPS}/\text{Cu}^{\text{II}}\text{BipyCD}_2]^-$ and coated with Nafion. Medium: phosphate buffer pH 7 + 0.5 M Na_2SO_4 . (d and e) Koutecký–Levich plots for $[\text{Fe}^{\text{III}}\text{TPPS}/\text{Cu}^{\text{II}}\text{BipyCD}_2]^-$. Values observed at -0.6 V vs. SCE were used to determine the average number of electrons used for the reduction of O_2 . Values measured at -1 V vs. SCE were used to determine kinetic parameters.

process was determined from the slope of Koutecký–Levich plots at -0.6 V (see Table 1 and ESI†). These averages are consistent with the number of electrons (n_{RDDE}) previously determined by RRDE experiments. In addition, Koutecký–Levich can provide information regarding the kinetics of the process at the electrode. In the Koutecký–Levich equation (eqn (2)):³¹

$$\frac{1}{I_{\text{m}}} = \frac{1}{I_{\text{k}}} + \frac{1}{B_{\text{L}}} \omega^{-\frac{1}{2}} \quad (2)$$

the term $1/I_{\text{k}}$ is the intercept of the plotted line with the current axis and represents the reduction current measured on a non-rotating electrode. Thus, the current values are related to the apparent kinetic constant of the reduction reaction by eqn (3):³²

$$I_{\text{k}} = nFA\Gamma_{\text{cat}}[\text{O}_2]k_{\text{app}} \quad (3)$$

where I_{k} is the kinetic current (A), n is the number of electrons, F is the Faraday constant ($96.487 \text{ C mol}^{-1}$), A is the electrode surface (0.25 cm^2), Γ_{cat} is the catalyst surface coverage ($10 \times 10^{-9} \text{ mol cm}^{-2}$), $[\text{O}_2]$ is the O_2 volumetric concentration ($\approx 1 \times$

10^{-3} M)³³ and K_{app} is the apparent rate constant ($(\text{M s})^{-1}$). Using these values and the experimental results extracted from values at -1 V of Koutecký–Levich plots (in the region of plateau), it was possible to determine the maximum K_{app} for the two electrocatalysts to be $2.1 \times 10^5 (\text{M s})^{-1}$ and $3.0 \times 10^5 (\text{M s})^{-1}$ for $[\text{Fe}^{\text{III}}\text{TPPS}/\text{BipyCD}_2]^{3-}$ and $[\text{Fe}^{\text{III}}\text{TPPS}/\text{Cu}^{\text{II}}\text{BipyCD}_2]^-$, respectively. These values are comparable to those reported for a previous cytochrome *c* oxidase model describe by Collman and collaborators.³⁴

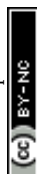
The difference in these values is significant whereas the number of electrons exchanged is almost identical, which suggests that the presence of copper in these models mostly influences the kinetics but does not play a significant role in the reduction mechanism. The number of electrons exchanged is significantly higher than with the terpyridine-based model, which also suggests that the efficiency of the electrocatalysts strongly depends on the stability and/or denticity of the ligand for the cuprous ion *versus* the cupric ion. Thus, the synergy between the Fe^{II} and Cu^{I} in models seems to reside in the orientation of the bound oxygen molecule and the affinity of oxygen for the distal site of the models, as suggested by the work of Karlin^{21,22,35,36} or Collman^{37,38} in which the copper-iron scaffold is organized around a μ -peroxide or superoxide, respectively. These results also suggest that to augment the preference for the 4-electron pathway, a tetrahedral cuprous ion is ideal.

Conclusions

A water-soluble supramolecular cytochrome *c* oxidase model consisting of a FeTPPS bound within a copper-bipyridine-bridged cyclodextrin dimer was prepared and characterized. The electrocatalytic properties of this supramolecular CcO model were investigated and compared with a terpyridine analogue. This work is a rare example of the direct comparison of iron–copper electrocatalysts built on a similar supramolecular inclusion complex in the 4-electron reduction of oxygen. Previous results from our group and others demonstrated that three N-ligands provide an ideal coordination sphere around Cu^{I} but, as demonstrated by the comparison of $[\text{Fe}^{\text{III}}\text{TPPS}/\text{Cu}^{\text{II}}\text{Terpy-CD}_2]^-$ with $[\text{Fe}^{\text{III}}\text{TPPS}/\text{Cu}^{\text{II}}\text{BipyCD}_2]^-$, co-planarity of the three nitrogen atoms is prejudicial to the 4-electron efficiency of the resulting electrocatalysts. Future work will tackle the preparation of CD-dimers organized around TREN caps to provide a perfect fit in order to stabilize Cu^{I} intermediates.

Author contributions

JW, JAW and HK designed and supervised the project. MB performed the synthesis, characterization and all titrations. CB and MB designed, performed and analyzed the electrochemical experiments. NLB performed and analyzed the EPR experiments. TH and KO designed and supervised the gas binding experiments performed by MB. MB, JW and JAW wrote the manuscript and all authors contributed to its preparation.



Data availability

The data supporting this article have been included as part of the ESI.†

Conflicts of interest

There are no conflicts to declare.

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