

ChemComm

Chemical Communications

rsc.li/chemcomm



ISSN 1359-7345

COMMUNICATION

Lena J. Daumann *et al.*

Learning from nature: recovery of rare earth elements by the
extremophilic bacterium *Methylophilum furax*



Cite this: *Chem. Commun.*, 2023, 59, 9066

Received 18th March 2023,
Accepted 31st May 2023

DOI: 10.1039/d3cc01341c

rsc.li/chemcomm

Learning from nature: recovery of rare earth elements by the extremophilic bacterium *Methylophilum fumariolicum*[†]

Helena Singer,^a Robin Steudtner,^{ib} Ignacio Sottorff,^a Björn Drobot,^{ib} Arjan Pol,^c Huub J. M. Op den Camp^{ib} and Lena J. Daumann^{ib} [★]

We present the extremophilic bacterium *Methylophilum fumariolicum* SolV as a platform for the recovery of rare earth elements (REE). Strain SolV is able to selectively extract the light REE from artificial industrial waste sources, natural REE-containing and post-mining waters. Upscaling, different media composition and accumulation over several cycles were successfully implemented, underlining the potential for bio-recovery of REE.

Modern life without rare earth elements (REE) can hardly be imagined today. The elements Sc, Y, La and the following 14 lanthanides (Ln) Ce to Lu have made their way into every aspect of our lives. REE are mainly obtained from primary sources such as the ore bastnaesite which also releases the radioactive elements thorium and uranium as by-products.¹ Secondary sources are hardly used to recycle REE (less than 2%) despite their large potential,^{2,3} focusing end-of-life products (EOL)^{4,5} such as fluorescent lamps,⁶ or industrial by-products and residues like mining waters.^{7,8} REE are crucial for new energy technologies, however their mining and refining poses environmental problems, since these processes are energy and material consuming. So, it appears indispensable to develop ecologically and economically compatible methods to recycle REE to extend the life cycle and reduce the environmental footprint of these elements.⁹ Nowadays, recovery of REE from EOL is based on hydro- and pyrometallurgical strategies, electrochemical treatments, biometallurgical technologies, nanomaterials, siderophores as chelators, and physical methods.^{4,10} All of them have in common that they have to face complex mixtures with non-REE in combination with low REE amounts.¹⁰

Not only mankind relies on REE, but in nature bacteria can be found that use these elements for their metabolism such as the methanotroph *Methylophilum fumariolicum* SolV¹¹ or *Methylobacterium extorquens* AM1.¹² While strain SolV is strictly dependent on Ln, strain AM1 is able to switch between a Ln and Ca-based pathway.¹² Ongoing, proteins are identified which include the essential lanthanides (Ln) as Lewis acid in the active site (methanol dehydrogenase XoxF)^{13,14} or can bind them such as LanModulin^{15–17} or LanPepsy.¹⁸ As a consequence, these biological systems have evolved a highly-efficient, selective Ln-uptake machinery that can be used to develop alternative approaches for bio-recovery of REE.¹⁹ Connecting hydrometallurgy and biochemistry was already shown for various metals, mainly focusing on d-block elements.^{20,21} Recently, copper-binding peptides were functionalized onto the surface of fungal mycelia,²² and microorganisms can be genetically engineered to bioaccumulate heavy metals.²⁰ In addition, the field of biosorption and accumulation for REE, using bacteria, fungi or algae is progressing.^{10,23,24} High adsorption capacities of REE were lately discovered for 12 cyanobacterial strains.²⁵ The Ln-utilizing bacterium strain AM1 was genetically modified for a selective uptake of heavier Ln, such as Gd.²⁶

For our studies, we used the native Ln-dependent strain SolV¹¹ to accumulate and enrich certain Ln from a variety of natural and artificial REE sources. As an extremophile, strain SolV needs high temperatures up to 60 °C and an acidic environment of pH 2–3 for optimal growth.¹¹ Since metal ions display a good solubility in acids (H₂SO₄, HNO₃ or HCl), this is also used by various hydrometallurgical metal recovery strategies.^{10,27} Consequentially, many industrial waste waters are in an acidic matrix, with a high concentration of other elements e.g. Mg, Ca, Sr, Se, As, Fe, Al, Hg and U.⁵ Since strain SolV is strictly Ln-dependent, and can tolerate these conditions, it is an ideal candidate for recovering REE from different low-grade sources. It has been previously shown that especially larger Ln such as La enhance growth of strain SolV, and in this regard, it is also interesting to examine the system for potential selectivity.^{11,14} To ensure a complex mixture of elements with

^a Department of Chemistry, Ludwig-Maximilians-University Munich, 81377 München, Germany. E-mail: lena.daumann@lmu.de

^b Institute of Resource Ecology, Helmholtz-Zentrum Dresden-Rossendorf e.V., 01328 Dresden, Germany

^c Department of Microbiology, Research Institute for Biological and Environmental Sciences, Radboud University Nijmegen, 6525 AJ Nijmegen, The Netherlands

[†] Electronic supplementary information (ESI) available. See DOI: <https://doi.org/10.1039/d3cc01341c>



high composition variability, we tested various REE sources – ranging from flooding waters, mining leachates, primary sources and EOL industrial waste (ESI† 1.1). Königstein and Gessenhalde water are redevelopment surface flooding waters from former uranium ore mining by sulfuric acid leaching. Other low-grade sources discussed and tested are post-mining waters, more specifically industrial decomposition waters from leaching of different ores (zinc blende, Li mica, quartz) which contain REE in low concentrations as by-products. To include a primary source with a high REE concentration in contrast, two digestion samples of bastnaesite were added. Finally, an industrial sample of an EOL product was screened. To analyse the potential of the bacterial strain SolV to selectively deplete REE from the different sources, ICP-MS analysis of the supernatants as well as of the bacterial cells were performed. Growth was monitored through optical density measurement at 600 nm (OD_{600}). The amount of REE depleted by strain SolV, growth curves and thereof calculated doubling times helped us to understand how strain SolV can be used as bio-recovery system for Ln from a small-scale approach (Step I), to optimization (Step II and III) to larger-scale application (Step IV).

We initially investigated, whether SolV showed depletion of REE in the 10 different REE sources (ESI† Table S1, Fig. S20, S21, Step I). We analysed the REE concentration in the supernatant before (S) and after cultivation (E) with strain SolV as well as in the control (C) without added biomass to investigate precipitation formation that could hamper with the OD_{600} readout (Fig. 1a, b and Fig. S1, ESI†), in combination with

OD_{600} measurements to monitor the bacterial growth (Fig. S2, ESI†). Fig. 1a shows the amount of REE in waste water from an old gold mine, where the Ln concentration does not significantly decrease in the presence of strain SolV. The slight reduction observed is comparable to the control which it is due to precipitate formation by the complex media matrix. Similar observations were made for Lithium mica (with strong precipitate formation in the control), water of ore leaching, and quartz leaching. In these samples some metal ions may be in too high concentration and thus toxic (e.g. As, Cr, Table S1, ESI†). In contrast, a significant decrease of the Ln amount in the presence of strain SolV was observed in the water coming from Königstein, Gessenhalde (Fig. 1b) and EOL waste. Leaching of zinc blende also showed a decrease of Ln concentration in the medium, but strong precipitate formation was present in the strain SolV and control, which hampered OD_{600} readout for growth experiments. Bastnaesite samples did not show a clear reduction of REE during the course of a three-day cultivation, very likely due to the high REE concentration (around 10 μ M). As bacterial growth was observed, this REE source was investigated further in a diluted fashion. To confirm that depletion of Ln is selective within the series and over other elements and due to active uptake of Ln by SolV (and not a result of simple cell-wall adsorption), a control experiment without CH_4 and CO_2 was conducted (including a measurement of all elements present). Without these gases, SolV cells are alive and intact but not growing. Fig. S3–S6 (ESI†) show that from a bastnaesite solution only P, Mg, Fe, Cu, Zn, Mn and La/Ce concentrations

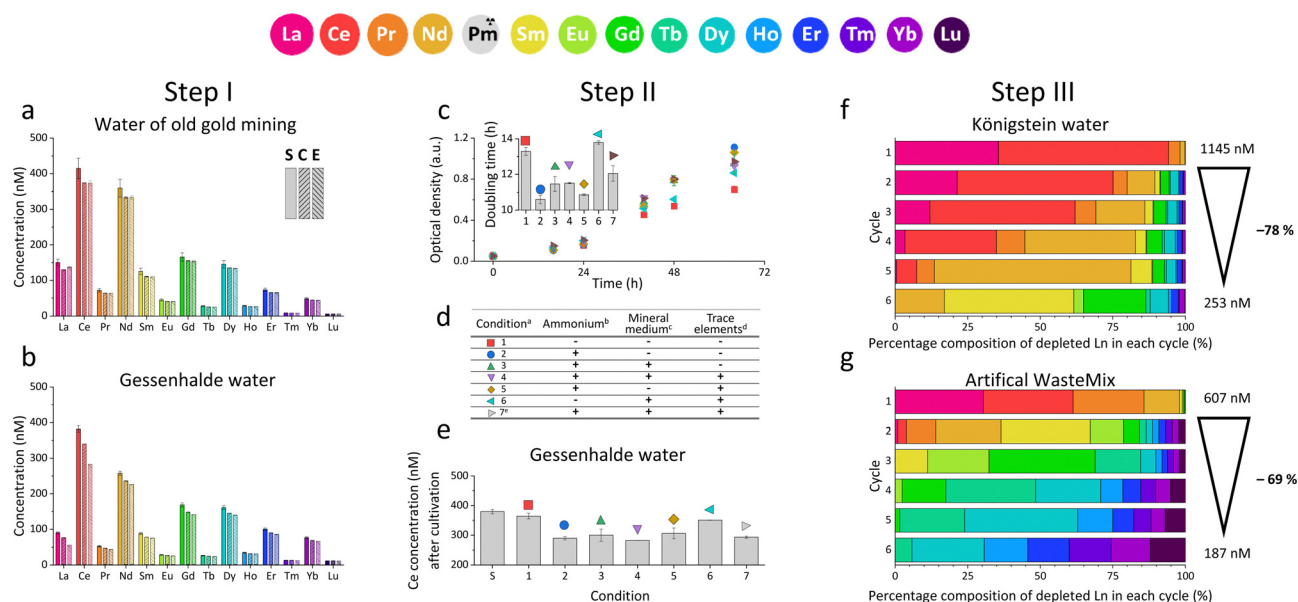


Fig. 1 ICP-MS analysis of the Ln concentration in (a) water of an old gold mine and (b) Gessenhalde water supernatant after strain SolV was removed. All tested REE-containing solutions before (S) and after (E) cultivation with strain SolV, including a control without bacteria (C). (c) Growth behaviour of strain SolV measured as increase in OD_{600} and calculated growth rates (small bar chart) with Gessenhalde water and (e) Ce concentration in the supernatant in the beginning (S) and after cultivation (1–7) under (d) different media compositions 1–7. + was added; – was not added; ^a pH adjusted to 2.7 with H_2SO_4 and 10% CH_4 and 5% CO_2 added; ^b $(NH_4)_2SO_4$; ^c $MgCl_2$, Na_2SO_4 , K_2SO_4 , NaH_2PO_4 , $CaCl_2$; ^d $FeSO_4$, $MnCl_2$, $CuSO_4$, $NiCl_2$, $ZnSO_4$, $CoCl_2$, Na_2MoO_4 ; ^e Medium autoclaved with all components. Depletion of different Ln in % by strain SolV in (f) Königstein water and (g) artificial WasteMix over 6 cycles of cultivation. Biomass was removed by centrifugation, and then fresh SolV biomass was added. Outline on the right shows the total Ln concentration after the 6 cycles, including total values and percentage. The bars display only the fraction of depleted Ln and their composition in each individual cycle.

are decreasing when SolV is actively growing. More importantly, other metals such as U or Pb are not taken up.

Based on the results of the first screening for potential candidates, we tested different media compositions (omitting/limiting N-source, salts for buffering, and metals for cofactors) to optimize the growth and Ln uptake by strain SolV (Step II). Ammonium (nitrogen source),²⁸ mineral medium (MM, minimal salts without additional nutritional supplements, mainly for buffering) and trace element solution (TE, providing cofactors for essential enzymes) were added to Gessenhalde and Königstein water in seven different combinations (Fig. 1d). The required gas supply, consistent of CH₄ and CO₂ was indispensable for growth of the methanotrophic strain SolV, and therefore couldn't be omitted. CO₂ is required for the carbon fixation pathway,²⁹ while CH₄ is an energy source, as it is first oxidized by methane monooxygenases to methanol, and finally to CO₂.³⁰ Fig. 1c shows the growth curves of strain SolV in Gessenhalde water under varying conditions, and calculated doubling times, which both indicated a reduced and limited growth behavior under condition 1 and 6 compared to the other tested medium compositions. This was also supported by the ICP-MS analysis (Fig. 1c), shown here with Ce (other elements in Fig. S7, ESI†). Under condition 1 and 6, almost no Ce was depleted by strain SolV within one incubation cycle whereas under better conditions 2–5 and 7 up to 25% of the initial Ce was removed. Similar observations were made for Königstein water (Fig. S8, S9, ESI†), EOL waste and bastnaesite samples (Fig. S10, condition 1,2,7, ESI†). Thus, ammonium, acting as a nitrogen source for growth, is the most crucial part of the media composition. It was shown that strain SolV is able to replace ammonium with atmospheric N₂ (under reduced oxygen concentrations <0.5%), however, a 2.8 times slower growth was observed,³¹ which could also explain the higher doubling times, and reduced Ln depletion from the growth medium observed, when no ammonium was added. The addition of the components TE and MM was not necessary, as the ICP-MS/OES analyses revealed a sufficiently high concentration of these elements in the waste waters themselves. These results are particularly important for a potential industrial application, where processes with the least amount of (potentially costly) additives are economically feasible. Thus, recovery in a process that can be repeated the same way several times to remove the maximum amount of Ln from a single sample would be beneficial. Consequentially, we tested whether SolV bacteria can be added for several cycles to REE sources until maximum OD₆₀₀ was reached. The experimental setup was upscaled from 10 mL in Step I and II to a volume of 50 mL and the accumulation process was run up to six 72 h cycles while the biomass was removed by centrifugation after every cycle, and fresh bacterial cells were added for standardization of all samples (Step III). However, it is also possible to use the harvested biomass for starting a new growth cycle. Independent of the used REE source, the multi-cycle upscaling experiment (Step III) revealed that about 70% of the initial Ln amount was removed from the sources by strain SolV constantly in the 6 cycles (Fig. 1e, f and Fig. S11–15, ESI†). By ongoing cycling, it seems achievable to

deplete 100% of the Ln. Matching, the depleted Ln can be found in the lyophilized and subsequently disrupted cell pellet (Fig. 2b). Particularly noteworthy is that Ln are not removed randomly by strain SolV, but a clear trend is present, showing a selective uptake of Ln. First, the light Ln are completely removed, then the heavier Ln are depleted from the source. We already showed that when strain SolV is presented to equal amounts of Ln and selected actinides, the strain will deplete preferentially the light Ln over the heavier ones.^{11,14} We proposed that this is mainly based on the size of these elements. While Ln have very similar coordination and chemical properties, they display a decreasing size with increasing atomic number along the series (Ln contraction). In addition, we assume that strain SolV evolved for a preferential uptake of the more abundant Ln, which are the larger ones in the series. Despite the versatile REE source composition, a trend for light Ln-preference is clearly observed. Following the up-scaling experiments, we performed a large-scale approach in a self-constructed batch reactor system with a continuous gas feeding to outline a possible industrial approach. 3.7 liters of Königstein water were mixed with medium, and bacterial cells of strain SolV were added to start the cultivation while a continuous supply of air, CH₄ and CO₂ was provided. Samples for OD₆₀₀ measurements were collected at different time intervals (Fig. 3a), and supernatant samples from selected time points were analysed with ICP-MS, as well as the final biomass. As expected, a clear decrease in the total Ln concentration (Fig. 3b) occurred as the cultivation progressed, and again the light Ln were depleted first. In total, 47.4% of present Ln were removed from Königstein water by strain SolV, however, here a maximum in bacterial growth seemed to be reached due to flattening of the OD₆₀₀ values around 20. For further uptake of Ln, biomass might be removed while the supernatant should be fed to the bioreactor again. In total, 15 g of lyophilized bacterial cells were obtained from 3.7 L of medium, with mainly incorporated La, Ce, Pr and Nd (Fig. S18 and S19, ESI†), while within the cycling experiments around 50–70 mg of lyophilized biomass were obtained over 6 cycles with 50 mL of medium. A direct comparison to the cyclic experiments (Step III) is not possible, since different limiting

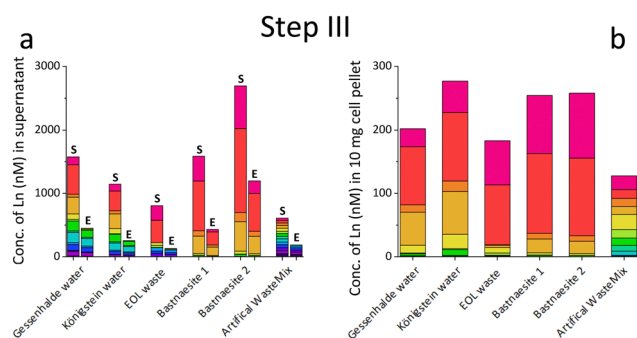


Fig. 2 (a) Concentration of Ln in different REE sources before (S) and after six cycles of cultivation (E) with strain SolV. (b) Concentration of Ln in 10 mg of the collected disrupted bacterial cell pellet (lyophilized and acidic microwave treatment), resuspended in 15 mL of MilliQ water, to enable a direct comparison between the different REE biomasses in the same unit (nM).



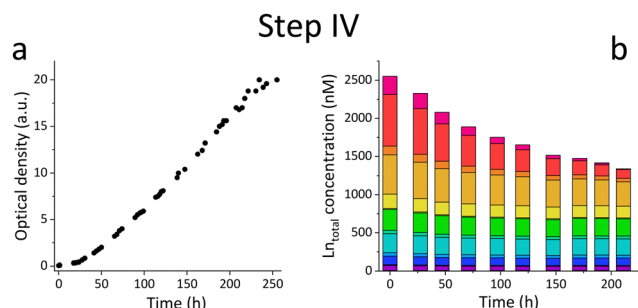


Fig. 3 (a) Growth of strain SolV in Königstein water performed in 3.7 L bioreactor displayed with OD₆₀₀ measurements. (b) ICP-MS analysis of the total Ln concentration in the supernatant with ongoing cultivation time.

factors apply to a closed system such as bottles compared to a continuous gas-flow system. However, it was shown that under both setups, there is a significant as well as a selective removal of Ln from the REE sources by strain SolV.

The reported results are promising, the extremophilic strain *Methylophilum fumariolicum* can be used to recover REE from a variety of low-grade sources. Other present elements play a subordinate role, since strain SolV shows sufficient growth even under minimal conditions, in combination with an acidic pH and high temperatures. Even the presence of toxic heavy metals such as Th and U in non-negligible concentrations did not impair the growth of strain SolV. Only the addition of ammonium, CH₄, O₂ and CO₂ are essential for sufficient bacterial growth and Ln uptake. Also, the flexible procedure, as small-scale-, up-scale cycling, or large-scale approach shows the promising potential application. By carefully adjusting the REE concentration in the system, it is possible to control which Ln are absorbed at which time. At high concentrations (μM range), mainly the light ones are depleted, whereas at low concentrations (up to 100 nM), the heavier ones can also be removed in selected fractions. This study shows that using bacteria for the recovery of Ln hold great potential, since already existing biological systems which evolved for a selective Ln uptake can be used to pave new ways for, ultimately, the recycling of critical elements.

The authors are grateful to the Wismut GmbH, G. E. O. S. GmbH, Amberger Kaolinwerke GmbH and to the Applied Geology and Microbiology groups of Friedrich Schiller University Jena for the post-mining waters. We thank Falk Lehmann and Dr Manja Vogel for kindly providing EOL and bastnaesite samples. LJD greatly acknowledges the ERC Starting Grant Lanthanophor (945846), BD thanks the PepTight Projekt (BMBF 031B1122).

Conflicts of interest

There are no conflicts to declare.

References

- 1 P. Fröhlich, T. Lorenz, G. Martin, B. Brett and M. Bertau, *Angew. Chem., Int. Ed.*, 2017, **56**, 2544–2580.
- 2 A. B. Patil, V. Paetzl, R. P. W. J. Struis and C. Ludwig, *Separations*, 2022, **9**, 56.
- 3 K. Binnemans, P. McGuinness and P. T. Jones, *Nat. Rev. Mater.*, 2021, **6**, 459–461.
- 4 C. Ramprasad, W. Gwenzi, N. Chaukura, N. Izyan Wan Azelee, A. Upamali Rajapaksha, M. Naushad and S. Rangabhashiyam, *Chem. Eng. J.*, 2022, **442**, 135992.
- 5 R. K. Jyothi, T. Thenepalli, J. W. Ahn, P. K. Parhi, K. W. Chung and J.-Y. Lee, *J. Clean. Prod.*, 2020, **267**, 122048.
- 6 C. Tunsu, M. Petranikova, C. Ekberg and T. Retegan, *Sep. Purif. Technol.*, 2016, **161**, 172–186.
- 7 M. Hermassi, M. Granados, C. Valderrama, C. Ayora and J. L. Cortina, *Sci. Total Environ.*, 2022, **810**, 152258.
- 8 S. Costis, K. K. Mueller, L. Coudert, C. M. Neculita, N. Reynier and J.-F. Blais, *J. Geochem. Explor.*, 2021, **221**, 106699.
- 9 P. S. Arshi, E. Vahidi and F. Zhao, *ACS Sustainable Chem. Eng.*, 2018, **6**, 3311–3320.
- 10 T. G. Ambaye, M. Vaccari, F. D. Castro, S. Prasad and S. Rtimi, *Environ. Sci. Pollut. R.*, 2020, **27**, 36052–36074.
- 11 A. Pol, T. R. M. Barends, A. Dietl, A. F. Khadem, J. Eygensteyn, M. S. M. Jetten and H. J. M. Op den Camp, *Environ. Microbiol.*, 2014, **16**, 255–264.
- 12 H. N. Vu, G. A. Subuyy, S. Vijayakumar, N. M. Good, N. C. Martinez-Gomez and E. Skovran, *J. Bacteriol.*, 2016, **198**, 1250–1259.
- 13 H. Lumpe, A. Pol, H. J. M. Op den Camp and L. J. Daumann, *Dalton Trans.*, 2018, **47**, 10463–10472.
- 14 H. Singer, R. Steudtner, A. S. Klein, C. Rulofs, C. Zeymer, B. Drobot, A. Pol, N. C. Martinez-Gomez, H. J. M. Op den Camp and L. J. Daumann, *Angew. Chem., Int. Ed.*, 2023, e202303669.
- 15 G. J. P. Deblonde, J. A. Mattocks, D. M. Park, D. W. Reed, J. A. Cotruvo and Y. Jiao, *Inorg. Chem.*, 2020, **59**, 11855–11867.
- 16 H. Singer, B. Drobot, C. Zeymer, R. Steudtner and L. J. Daumann, *Chem. Sci.*, 2021, **12**, 15581–15587.
- 17 C.-E. Wegner, M. Westermann, F. Steiniger, L. Gorniak, R. Budhraj, L. Adrian and K. Küsel, *Appl. Environ. Microbiol.*, 2021, **87**, e03144–03120.
- 18 J. L. Hemmann, P. Keller, L. Hemmerle, T. Vonderach, A. M. Ochsner, M. Bortfeld-Miller, D. Gunther and J. A. Vorholt, *J. Biol. Chem.*, 2023, **299**, 102940.
- 19 Z. Dong, J. A. Mattocks, G. J. Deblonde, D. Hu, Y. Jiao, J. A. Cotruvo, Jr. and D. M. Park, *ACS Cent. Sci.*, 2021, **7**, 1798–1808.
- 20 P. Diep, R. Mahadevan and A. F. Yakunin, *Front. Bioeng. Biotechnol.*, 2018, **6**, 157.
- 21 M. J. Capeness and L. E. Horsfall, *Biochem. Soc. Trans.*, 2020, **48**, 1367–1378.
- 22 J. Urbina, A. Patil, K. Fujishima, I. G. Paulino-Lima, C. Saltikov and L. J. Rothschild, *Sci. Rep.*, 2019, **9**, 16422.
- 23 F. Barmettler, C. Castelberg, C. Fabbri and H. Brandl, *AIMS Microbiol.*, 2016, **2**, 190–204.
- 24 R. M. Brown, A. Mirkouei, D. Reed and V. Thompson, *Renew. Sust. Energ. Rev.*, 2023, **173**, 113099.
- 25 M. Paper, M. Koch, P. Jung, M. Lakatos, T. Nilges and T. B. Brück, *Front. Bioeng. Biotechnol.*, 2023, **11**, 1130939.
- 26 N. M. Good, H. D. Lee, E. R. Hawker, M. Z. Su, A. A. Gilad and N. C. Martinez-Gomez, *Front. Microbiol.*, 2022, **13**, 820327.
- 27 E. O. Opare, E. Struhs and A. Mirkouei, *Renew. Sust. Energ. Rev.*, 2021, **143**, 110917.
- 28 S. S. Mohammadi, A. Pol, T. van Alen, M. S. M. Jetten and H. J. M. Op den Camp, *Front. Microbiol.*, 2017, **8**, 1901.
- 29 A. F. Khadem, A. Pol, A. Wiczorek, S. S. Mohammadi, K.-J. Francois, H. G. Stunnenberg, M. S. M. Jetten and H. J. M. Op den Camp, *J. Bacteriol.*, 2011, **193**, 4438–4446.
- 30 N. Picone, S. S. Mohammadi, A. C. Waajen, T. A. van Alen, M. S. M. Jetten, A. Pol and H. J. M. Op den Camp, *Front. Microbiol.*, 2020, **11**, 604485.
- 31 A. F. Khadem, A. Pol, M. S. M. Jetten and H. J. M. Op den Camp, *Microbiology*, 2010, **156**, 1052–1059.

