

Cite this: *J. Mater. Chem. B*, 2022,
10, 7183

Supramolecular biomaterials for enhanced cancer immunotherapy

Han Zhang,^a Jiafei Zhu,^a Tianxu Fang,^b Meng Li,^{*c} Guojun Chen^{*b} and Qian Chen^{id *a}

Cancer immunotherapy has achieved promising clinical results. However, many limitations associated with current cancer immunotherapy still exist, including low response rates and severe adverse effects in patients. Engineering biomaterials for the delivery of immunotherapeutic reagents has been suggested to be an effective strategy to improve cancer immunotherapy. Among different biomaterials, supramolecular biomaterials with flexible and versatile structures and functions have exhibited unparalleled advantages in promoting cancer immunotherapy. In recent years, various supramolecular formulations have been extensively explored as immunotherapeutic delivery platforms due to their high cargo-loading capacity/feasibility, facile immunization function, and excellent biocompatibility, which make them possible candidates for modular and personalized cancer immunotherapy. These nanoarchitectures with unique topologies possess distinguishing advantages in cancer immunotherapy, incarnating a structure–property relationship. Based on extensive state-of-the-art research, this minireview highlights recent advances in supramolecular biomaterials for cancer immunotherapy and discusses the possible mechanisms underlying how supramolecular biomaterials promote the development of cancer immunotherapy together with their potential for clinical translation.

Received 8th January 2022,
Accepted 6th March 2022

DOI: 10.1039/d2tb00048b

rsc.li/materials-b

1. Introduction

Cancer immunotherapy is a revolutionary cancer treatment aiming to activate or boost the inherent immune system to recognize and kill cancer cells and has achieved promising therapeutic responses in the clinic.^{1–3} Adaptive immune responses are essential for killing tumor cells, which are predominantly cell-mediated with the following sequential steps: release of antigens from cancer cells, delivery of cancer antigens to antigen-presenting cells (APCs), presentation of antigens to T cells, priming and activation of T cells, trafficking and infiltrating of cytotoxic T lymphocytes (CTLs) into tumors, recognizing and killing of tumors by CTLs with cytokine secretion, and overcoming immunosuppression in the tumor microenvironment.^{4,5} In recent years, various cancer immunotherapies, including cancer vaccines,^{6,7} cytokine therapy,⁸ immune checkpoint blockade and adoptive T cell therapy,^{9–11} have shown promising clinical outcomes. Despite great

promise in certain scenarios, immune reaction cascades are often thwarted by inefficient and unendurable immune responses, which result in immune escape of tumor cells.^{12–14} Furthermore, these immunotherapies induce severe and sometimes lethal side effects in cancer patients, such as colitis, hepatitis, and endocrinopathies.^{15–17} To tackle these limitations of immunotherapy, novel materials spurred by the rapid development of materials science in past decades have been investigated.¹⁸ Nano/microscale biomaterials have been developed to encapsulate various immunotherapeutic agents and effectively deliver them into tumor tissue, which could boost multiple stages of immune responses and reduce off-target side effects.^{19–22} However, these delivery systems also bring additional challenges to clinical translation. Most of these biomaterials are often hindered by tedious organic synthesis, complicated fabrication and inefficient cargo loading.^{23,24} Biomaterials with uncontrollable physiochemical properties, such as size, charge and morphology, may induce uncertainty regarding the efficacy of cancer immunotherapy.^{25,26} Thus, it is of great importance to develop facile, flexible and versatile strategies to fabricate intelligent drug delivery systems to achieve efficient and controllable cancer immunotherapy.

Nature guides rational methods of supramolecular material construction, as exemplified by protein folding, the DNA double helix and phospholipid bilayer membrane formation, in harnessing the power of molecular self-assembly.^{27,28}

^a Institute of Functional Nano & Soft Materials (FUNSOM), Jiangsu Key Laboratory for Carbon-Based Functional Materials & Devices, Soochow University, Suzhou 215123, China. E-mail: chenqian@suda.edu.cn

^b Department of Biomedical Engineering, and the Rosalind & Morris Goodman Cancer Institute, McGill University, Montreal, QC H3G 0B1, Canada. E-mail: guojun.chen@mcgill.ca

^c Department of Dermatology, Shanghai 9th Peoples' Hospital, Shanghai Jiaotong University, Shanghai, China. E-mail: lemonlives_dr@163.com

Supramolecular self-assembly is a “bottom-up” organization tactic to spontaneously fabricate well-ordered nano/microscale architectures, which are driven by noncovalent interactions, including hydrophobic interactions, π - π stacking, hydrogen bonding, electrostatic attractions, and coordination interactions.^{29,30} Over the past few decades, self-assembly of building blocks, such as small molecules, biomacromolecules, and polymers, to form multitudinous nanoarchitectures with various topologies and dimensions from 0 dimensional (0D) to 3D in physiological environments has received increasing attention.^{31–33} It was found that the distinctive structures have induced significant influences on many properties of these nanoarchitectures, including specific surface area,³⁴ stability,³⁵ softness,³⁶ responsiveness,³⁷ encapsulation capability,³⁸ cellular uptake,³⁹ and pharmacokinetics.⁴⁰ Recently, supramolecular materials have been investigated as delivery platforms for various immunotherapeutic agents, including small molecule adjuvants,⁴¹ macromolecular antibodies,⁴² cytokines and immune cells.^{43,44} They have exhibited a high cargo-loading capacity/feasibility, controllable payload release behaviors, simple immune functionalization and excellent biocompatibility.^{45,46} It is worth noting that by modulating the structural properties, supramolecular materials exert significant influences on the activities of various immune cells.^{47,48} Specifically, supramolecular materials with suitable size, shape, charge, stiffness and surface pattern have been

proven essential in stimulating immune responses, including activation of APCs and T cells and intratumoral infiltration of cytotoxic T lymphocytes (CTLs) to recognize and kill cancer cells.^{49,50} Additionally, “self-delivery” supramolecular biomaterials based on immune stimulants have attracted wide attention due to their minimized excipient contents, high drug loading capacity, and synergistic immune functions with cargoes.^{51,52} Thus, supramolecular biomaterials present great advantages in promoting cancer immunotherapy.

In this minireview, we focus on the state-of-the-art developments of supramolecular biomaterials for enhanced cancer immunotherapy. This review is composed of two main sections (Scheme 1). First, noncovalent driving forces are introduced for the fabrication and physicochemical property modulation of supramolecular biomaterials. Second, we will discuss the supramolecular architectures with different topological morphologies and dimensions to improve cancer immunotherapy. In this section, self-assembly to control the biophysical properties of supramolecular architectures provides a better understanding of the relationship between biophysical properties and immune responses. The main goal of this review is to unveil the emerging opportunities and challenges of supramolecular biomaterials for cancer immunotherapy. Given the currently extensive investigations on supramolecular biomaterials for cancer immunotherapy, this review is expected to motivate and inspire further exploration in this field.



Han Zhang

Dr Han Zhang obtained her PhD in chemistry from Shandong University in 2019. After graduation, she started her postdoctoral career at Soochow University under the supervision of Dr Qian Chen and Dr Zhuang Liu. Her research interests are mainly focused on the chemistry and materials of biomolecule self-assembly and functional architectonics for biomedicine.



Qian Chen

Dr Qian Chen received her BS degree and PhD degree in 2013 and 2017, respectively, both from Soochow University. She then worked at the University of North Carolina at Chapel Hill as a postdoctoral scholar from 2017 to 2018. After that, she worked at the University of California, Los Angeles (UCLA) as a post-doc from 2018 to 2019. In 2019, Dr Chen joined the Institute of Functional Nano & Soft Materials (FUNSOM) at

Soochow University as a principal investigator. Dr Chen's research focuses on leveraging biomaterials and biomedical engineering strategies for applications in multimodal biomedical imaging, cancer immunotherapy and tissue engineering. She has published over 90 peer-reviewed papers in leading journals, including Nature Nanotechnology, Nature Communications, PNAS, Advanced Materials, etc., which have been cited by others over 10 000 times (H-index = 49). She was listed as a 'highly cited researcher' by Clarivate Analytics in 2019 and 2021. She has served as an editorial board member for Matter and Nano research since 2021. She won the Biomaterials Diversity Award for Young Investigator in 2021.



Scheme 1 Supramolecular self-assembly offers the possibility for modulating nanostructures and reprogramming immunological properties, paving the way for enhanced cancer immunotherapy.

2. Modulating self-assembly of supramolecular biomaterials

Supramolecular biomaterials are usually prepared through a bottom-up process, which is the self-assembly of building blocks *via* noncovalent interactions, including hydrophobic interactions, π - π stacking, hydrogen bonding, electrostatic interactions, and coordination interactions.⁵³ Supramolecular biomaterials with a tunable structure and composition and formed under environmental cues often exhibit exquisitely tailored physicochemical properties.⁵⁴ Importantly, understanding the self-assembly process of supramolecular biomaterials is crucial for the design and construction of biomaterials with desired biophysical properties and biofunctions.

The hydrophobic effect plays a vital role in regulating the self-aggregation behaviors of apolar molecules, especially amphiphiles.⁵⁵ According to the principle of least entropy production, hydrophobic groups tend to aggregate to form hydrophobic cores, while hydrophilic segments are arranged on the outside to contact water. Driven by hydrophobic interactions, amphiphiles tend to form ordered spherical aggregates.⁵⁶ The most representative example is liposomes, which exhibit amphiphilic phospholipid packing into vesicles.⁵⁷ Considering their unique architecture, liposomes are able to entrap hydrophobic substances in their membranes and load hydrophilic cargoes into the central aqueous hollow. π - π stacking interactions usually coexist with hydrophobic interactions, which can also induce directional growth of assemblies. These interactions are also robust in water because of the limited solubility of molecules containing aromatic groups.⁵⁸ Zheng and coworkers fabricated vesicles self-assembled by single phospholipid-conjugated porphyrin derivatives *via* hydrophobic and π - π stacking interactions, which have been proven to be ultrastable *in vivo* (Fig. 1a).⁵⁹

In addition to hydrophobic and π - π stacking interactions, hydrogen bonding with high selectivity and directionality is also a commonly used force for supramolecular biomaterial

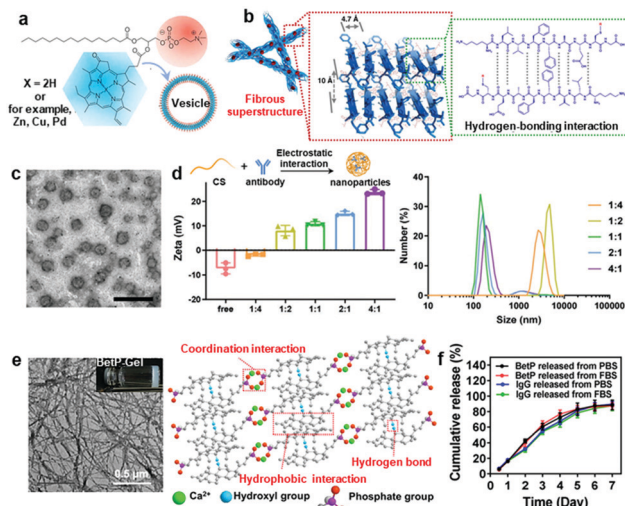


Fig. 1 (a) Schematic representation of pyropheophorbide-lipid self-assembly into nanovesicles driven by hydrophobic and π - π stacking interactions.⁵⁹ Copyright 2011, Springer Nature. (b) The fibrous superstructure self-assembled by the β -sheet-forming peptide with multiple hydrogen bonds forming between amide bonds.⁶³ Copyright 2019, Springer Nature. (c) Morphology of CS/antibody nanoparticles. (d) Zeta potential and size distribution of electrostatic assemblies with different feeding ratios of CS and antibody ranging from 1 : 4 to 4 : 1 (mass : mass).⁶⁵ Copyright 2021, John Wiley and Sons. (e) Morphology of BetP-Gel self-assembled by hydrogen-bonding, hydrophobic and coordination interactions. (f) Cumulative release curves of BetP and IgG from BetP-Gel.⁷³ Copyright 2020, American Chemical Society.

fabrication.⁶⁰ Typically, many biomolecules, such as nucleic acids, peptides and proteins, have multiple hydrogen bond formation sites, including amide groups, amino groups, hydroxyl groups, and carboxyl groups, which can facilitate hydrogen bond-modulated nanostructures.⁶⁰⁻⁶² Wang and Zhao designed a variety of peptide-assembled nanoparticles and nanofibers based on β -sheet-forming peptides (Fig. 1b).⁶³ These nanostructures with different morphologies showed different tumor permeability and retention behaviors. Electrostatic interactions between opposite charges are another non-directional noncovalent interaction used for self-assembly. Electrostatic interactions are largely dependent on the ratio of opposite charges, pH value, concentration and ionic strength.⁶⁴ Recently, our group fabricated chitosan (CS)-antibody self-assembled nanoparticles, which were driven by electrostatic interactions. The size and charge of the nanoparticles varied with the ratios of the two components, demonstrating that electrostatic interactions played a crucial role during self-assembly (Fig. 1c and d).⁶⁵ Coordination bonds are formed spontaneously between metal ions and organic ligands mainly through Lewis acid/base interactions, which is a special intermolecular force with an intermediate strength that is equivalent to weak interactions and exhibits a dynamic nature in certain circumstances.⁶⁶ For instance, Tezcan *et al.* engineered a monomeric protein with Zn^{2+} -binding sites, which could self-assemble to form 1D, 2D or 3D nanostructures by adjusting the metal ion/protein ratios or pH values.⁶⁷

Due to the adaptive or dynamic nature of noncovalent interactions, supramolecular biomaterials are more sensitive to external stimuli, indicating that structural destruction or deformation could occur in response to external stimuli, including pH, enzymes, solvents, and temperature, which could be leveraged for on-demand payload release.^{68,69} Stupp and coworkers reported a pH-responsive self-assembled nanofiber using histidine peptide amphiphiles, which can encapsulate camptothecin (CPT) with an encapsulation efficiency of up to 60%. In the acidic tumor microenvironment, most histidine residues are protonated, leading to electrostatic repulsion, disassembly of nanoparticles, and release of CPT.⁷⁰ In another work, a nanoplatfom formed by self-assembly of amphiphilic amino acids (Fmoc-L-Lys) and chlorin e6 (Ce6) was obtained *via* multiple weak intermolecular interactions, including electrostatic forces, π - π stacking, and hydrophobic interactions. These self-assembled nanodrugs exhibited multiple favorable therapeutic features, including tunable size, high loading efficiency, controllable drug release in response to pH, surfactant, and enzyme stimuli, as well as preferable cellular uptake and biodistribution.⁷¹ Ding *et al.* fabricated an acid-sensitive PEG-decorated calcium carbonate (CaCO₃) nanoparticle incorporating curcumin (CUR; a Ca²⁺ enhancer) (PEGCaCUR). PEGCaCUR released Ca²⁺ and CUR in an acid tumor microenvironment (TME) inducing mitochondrial Ca²⁺ overload and immunogenic cell death (ICD) for improved cancer therapy.⁷² Recently, our group engineered an injectable anti-inflammatory steroid drug-based supramolecular hydrogel (BetP-Gel) for the local delivery of antibodies. The nanofiber hydrogel was formed by hydrogen-bonding, hydrophobic and coordination interactions (Fig. 1e).⁷³ The multiple noncovalent interactions permitted its injectable properties (quick gel-sol phase transition) for minimally invasive administration. Due to the competitive interaction between phosphate under physiological conditions and calcium ions in BetP-Gel, this hydrogel was gradually degraded to release the encapsulated drugs (Fig. 1f).

In addition to the modulation of the disassembly process, the *in situ* self-assembly process could also be modulated by these noncovalent interactions.⁷⁴ Yin *et al.* prepared a size-reducible nanodrug using dye-chemodrug conjugates, which were synthesized by covalently attaching pentamethine indocyanine (ICy5) dye with cyclic Arg-Gly-Asp (RGD) peptide and camptothecin (CPT), *via* molecular self-assembly. Upon red light irradiation, the degradation of ICy5 through the C-C cleavage of polyene chains reduced the size of the nanodrug from 90 to 10 nm, which facilitated deep tumor penetration of the nanodrug and release of the chemodrug.⁷⁵ Wang and coworkers reported *in situ*-formed nanofibers of enzyme-responsive purpurin18-peptide conjugates.⁷⁶ The peptide precursors could be cleaved by gelatinase overexpressed in tumors to increase the hydrophobicity and reduce the steric hindrance of peptide molecules, resulting in the self-assembly of peptides into nanofibers *in situ* and enhancing the retention of peptides in tumors. Moreover, pH, reactive oxygen species (ROS) and light can also induce the morphological transformation of

self-assembled structures.⁷⁷ Therefore, supramolecular biomaterials exhibit many unique advantages for drug delivery.

3. Supramolecular biomaterials for enhanced cancer immunotherapy

Supramolecular biomaterials with flexible and accurate tailored physicochemical properties have shown versatility in modulating their biological performance related to drug encapsulation efficiency, immunogenicity, multivalency, immune cell behaviors and immunotherapy responses, which could contribute to efficient and safe cancer immunotherapy.^{77,78} Here, we will classify the supramolecular system for immunotherapy according to the dimensions of topological structures, which mainly include 0D, 1D and 3D structures, and introduce their application for cancer immunotherapy enhancement.

3.1 0D nanoparticles for cancer immunotherapy

0D nanobiomaterials are defined as nanoparticles with three dimensions confined to the nanoscale.⁷⁹ Supramolecular 0D nanoparticles exhibit many unique properties, such as tunable surface properties, versatile loading capacity, fast internalization rate, prone to deformability, and flexible administration routes.⁴⁷ Therefore, supramolecular 0D nanoparticles have been extensively explored for cancer immunotherapy. Here, we will mainly introduce two main types of 0D architectures, namely, nanocapsules with core-shell structures and nanoparticles with solid structures.

Micelles, liposomes or some polymeric particles are formed by self-assembly of amphiphilic materials.⁸⁰ These nanocapsules can be used to encapsulate water-soluble immunotherapeutic agents in aqueous cores and hydrophobic drugs within hydrophobic interiors.⁸¹ Furthermore, the surface of nanocapsules can be engineered with suitable charge, softness and multivalency to interact with immune cells to boost immune responses.⁸² Seder *et al.* developed a vaccine platform (SNP-7/8a) based on charge-modified peptide-TLR-7/8a conjugates that were chemically programmed to self-assemble into micelles with a uniform size of approximately 20 nm (Fig. 2a).⁸³ This approach realized precise loading of diverse peptide neoantigens after linking to the TLR-7/8a adjuvant, which exhibited increased APC uptake and T cell mediated immune responses (Fig. 2b). Li *et al.* constructed nanovesicles using an oxaliplatin (OXA) prodrug and PEGylated photosensitizer (PS) through hydrophobic interactions.⁸⁴ This nanovesicle with a size of 80 nm showed high tumor accumulation after intravenous (i.v.) injection into mice, which could further elicit antitumor immune responses by inducing immunogenic cell death (ICD) of tumor cells. Recently, Mooney and coworkers also developed a cationic liposome based on 1,2-dioleoyl-3-trimethylammonium-propane (DOTAP)/cholesterol encapsulated with hydrophilic STING agonists for treating lung metastasis of melanoma.⁸⁵ The cationic liposome could bind to the anionic cell membrane, leading to enhanced cell association and cytosolic delivery of cGAMP, effectively activating the

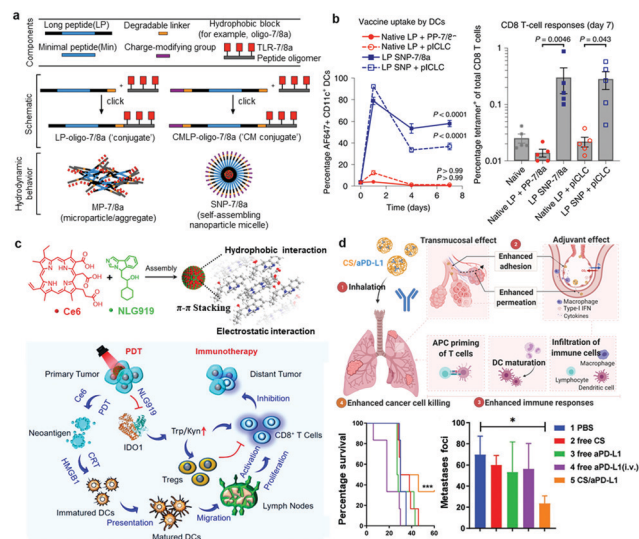


Fig. 2 (a) A nanovaccine platform (SNP-7/8a) based on peptide–TLR-7/8a conjugates that are chemically programmed to self-assemble into micelles. (b) The percentage of total CD11+ DCs that had taken up vaccine (left) and CD8⁺ T cell responses from blood assessed by tetramer staining (right).⁸² Copyright 2020, Springer Nature. (c) Ce6 and NLG919 self-assembled into uniform nanosized particles through hydrophobic, π - π stacking, and electrostatic interactions and the proposed mechanism for photodynamic sensitized immunotherapy.⁸⁸ Copyright 2020, American Chemical Society. (d) Schematic depiction of inhaled CS/aPD-L1 nanoparticles to suppress lung metastases (top), survival curves and numbers of lung metastatic foci after different treatments (bottom).⁶⁵ Copyright 2021, John Wiley and Sons.

STING pathway and inhibiting the growth of tumors. In another work, Chen *et al.* developed size-transformable artificial antigen-presenting cells (aAPCs), which were self-assembled by copolymer biotin-PEG-PHPMA(-SH)-PDMA, loaded with IL-2 in the inner core and decorated with a peptide-loaded major histocompatibility complex (MHC) monomer and CD28 on the surface.⁸⁶ When aAPCs encountered preactivated antigen-specific T cells, they transformed from nanosized to micro-sized with disulfide bond cleavage into thiols. aAPCs with micro-sizes exhibited obviously prolonged retention in tumors, achieving potent T cell-mediated immune responses.

In addition to the above-mentioned vesicles composed of amphiphiles, nanoparticles with solid structures that are made from substances such as drugs, pigments, polysaccharides, proteins, and even cell membranes have also been investigated for enhanced cancer immunotherapy.⁸⁷ These substances can not only act as building blocks of nanoparticles but are also able to activate antitumor immune responses. Li and coworkers constructed self-assembled nanoparticles by optimizing the noncovalent interactions between chlorine e6 (Ce6) and an inhibitor of indoleamine 2,3-dioxygenase (NLG919) for photodynamic immunotherapy (Fig. 2c).⁸⁸ These self-assembled nanodrugs exhibited improved solubility and stability, achieving relatively high drug loading capability and evading the risk of possible immunogenicity induced by accessory structures in core-shell nanoparticle compositions. In another work, Liu and

Wang prepared a cancer vaccine using Mn²⁺ ions and the nucleotide oligomerization binding domain 1 (Nod1) agonist *meso*-2,6-diaminopimelic acid (DAP) *via* coordination interactions with encapsulated ovalbumin (OVA).⁸⁹ The formed OVA@Mn-DAP nanoparticles exhibited a strong protective effect against cancer cells. In addition, Seder *et al.* attached hydrophobic Toll-like receptor agonists (TLR-7/8a) to hydrophilic HEMA-based polymers (polymer-TLR-7/8a) and evaluated the size influences of assemblies of polymer-TLR-7/8a on the location, magnitude and duration of the innate immune system.⁹⁰ More recently, our group reported self-assembled nanoparticles of chitosan (CS) and anti-programmed cell death protein ligand 1 (aPD-L1) *via* electrostatic interactions to treat lung metastasis of melanoma tumors (Fig. 2d).⁶⁵ CS not only temporarily opened the tight junctions of epithelial cells to promote the pulmonary delivery of aPD-L1 but also exhibited adjuvant effects by activating the STING pathway. Interestingly, noninvasive aerosol inhalation of CS/aPD-L1 nanoparticles could effectively activate different kinds of immune cells, especially cytotoxic T lymphocytes (CTLs), and prolonged the survival of mice (Fig. 2d). Very recently, Ding *et al.* reviewed the role of nanoparticle-mediated ICD in cancer immunotherapy. These nanoparticles delivered ICD-inducing drugs and antibodies to the tumors and improved the activity of reagents, regulating TME and boosting the immune response.⁹¹

3.2 1D fibrous biomaterials for cancer immunotherapy

1D nanomaterials, including filamentous micelles and nanofibers, with elongated structures, usually exhibit a number of unique physicochemical properties in terms of stability, tolerability and multivalency, which are closely associated with many biological processes, including phagocytosis, bio-distribution, and bioavailability.⁹² Thus, 1D fibrous biomaterials also possess some unique advantages in enhancing cancer immunotherapy.

Filamentous micelles with monolayers are usually formed by single amphiphilic components such as diblock copolymers and peptide amphiphiles through hydrophobic and/or hydrogen interactions.⁹³ However, while filamentous micelles have been studied for a long time, their foray into immunotherapy has only been reported in recent years.⁴⁷ Collier and coworkers developed a series of filomicelles formed by self-assembly of β -sheet peptide Q11-linked T cell epitopes and/or B cell epitopes (Q11 epitopes), which exhibited many advantageous immune effects.^{94–96} For instance, Q11-OVA_{323–339} (O-Q11) peptide self-assembled filomicelles with high-density antigens displayed on the surface could effectively deliver antigens to APCs and elicit strong antibody responses without any additional adjuvants (Fig. 3a).⁹⁴ In another work, they further demonstrated that the length of fibers could be optimized to improve the internalization, processing, and presentation of antigens.⁹⁷ Moreover, Rudra *et al.* linked the model antigenic peptide OVA to L- or D-amino acids to produce enantiomeric filamentous micelles through a self-assembly strategy.⁹⁸ Compared to filamentous micelles based on L-amino acids, D-amino acid peptide nanofibers elicited stronger antibody responses and long-term

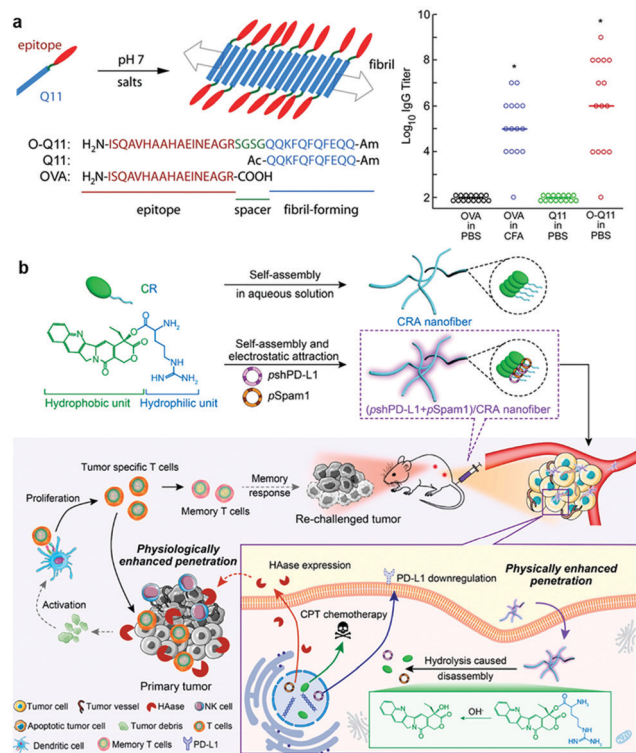


Fig. 3 (a) Schematic depiction of the assembly process and molecular composition of the Q11 epitope peptide (left) and IgG titer results suggesting that O-Q11 elicited high IgG titers without the addition of adjuvant (right).⁹⁴ Copyright 2010, PNAS National Academy of Sciences, American Institute of Physics. (b) Cationic spiral nanofibers assembled with a CPT prodrug for plasmid (pshPD-L1 and pSpam1) delivery and the immune response mechanism triggered by chemoimmunotherapy.¹⁰⁵ Copyright 2021, American Chemical Society.

antigen presentation *in vivo*, indicating that the stereochemistry of biomaterials could be used to program adaptive immune responses. Filomicelles assembled by polymers rather than peptides have also been investigated.⁹⁹ For example, Scott and coworkers developed filomicelles assembled from PEG-PPS block copolymers, which were capable of targeting dendritic cells (DCs) *in vivo*.¹⁰⁰ Recently, they further demonstrated that filomicelles can reassemble into micelles with hydrophilic group-modified propylene sulfide under oxidative conditions.¹⁰¹ This cylinder-to-sphere transition under physiological oxidative conditions allowed the sustained delivery of immunotherapeutic agents for one month without inflammatory bioresorption, providing a new tool for efficient and safe immunotherapy.

Apart from the monolayer micelles self-assembled from the amphiphilic building block, self-assembled nanofibers also exhibited diverse advantages for immunotherapy. In a recent work, dual functional coordination polymer nanofibers based on zoledronic acid and gadolinium were constructed *via* the ordered self-assembly process.¹⁰² Notably, compared with the coordination nanoparticles, nanofibers were more conducive to endocytosis by macrophages. Moreover, these nanofibers could deposit X-rays for improved reactive oxygen species production

to induce potent immunogenic cell death (ICD), synergistically improving DC maturation, promoting T cell infiltration, and inhibiting the growth of primary, distant, and metastatic tumors.¹⁰² In another work, Guler and coworkers investigated the ability of 0D and 1D self-assembled peptide nanostructures encapsulating unmethylated CpG motifs to activate the immune response.¹⁰³ The nanofibrous structures were found to directly induce Th1 immune responses and obviously promote uptake by DCs, whereas the nanospheres mainly induced the Th2-associated immune response. Furthermore, Yan *et al.* fabricated supramolecular nanofibrils through coassembly of clinically approved immunomodulatory thymopentin (TPS) and near-infrared indocyanine green (ICG) for localized photothermal immunotherapy of pancreatic tumors.¹⁰⁴ It was found that nanofibrils with long-range ordered structures show improved photophysical capabilities for photothermal conversion. More interestingly, compared to nanospheres, fibrous nanodrugs showed obviously improved retention in tumor tissue, which could significantly promote the proliferation and differentiation of both CD8⁺ T cells and CD4⁺ T cells to kill pancreatic tumors. In another work, Chen *et al.* constructed camptothecin (CPT) prodrug-assembled nanofibers to deliver two plasmids, pshPD-L1 and pSpam1, to enhance cancer chemoimmunotherapy (Fig. 3b).¹⁰⁵ Compared with the spherical form, the nanofibers exhibited better blood circulation ability and enhanced tumor penetration, which could effectively inhibit the growth of both primary and distant tumors while working together with immune checkpoint inhibitors. Hence, the integration of supramolecular nanofibers for immunotherapy is promising to inhibit tumor growth, metastasis and recurrence.

3.3 3D hydrogel for cancer immunotherapy

Supramolecular hydrogels, including injectable hydrogels and *in situ* formed hydrogels, with 3D networks are also ideal local delivery systems for cancer immunotherapy.¹⁰⁶ Moreover, supramolecular hydrogels assembled by dynamic and reversible noncovalent interactions could usually act as an “intelligent” drug delivery system with stimulus responsiveness, desirable biodegradability and high biosafety.¹⁰⁷ Owing to the simple preparation and administration process, supramolecular hydrogels have attracted extensive attention in encapsulating immune therapeutics, including small molecules, macromolecules or cells. More interestingly, hydrogels can achieve excellent spatial and temporal control of drug release by precisely adjusting the pore and mesh sizes of hydrogel scaffolds, together with the interactions between drugs and networks or the degradation behavior of hydrogels.¹⁰⁸ Thus, supramolecular hydrogels may play an important role in cancer immunotherapy.

Injectable supramolecular hydrogels with quick gel-sol phase transition properties could usually maintain their geometry and architecture at the injection site, achieving high local drug concentration, prolonged drug retention, and minimal invasiveness.¹⁰⁹ Wang and coworkers designed a self-assembled supramolecular hydrogel encapsulating DCs based

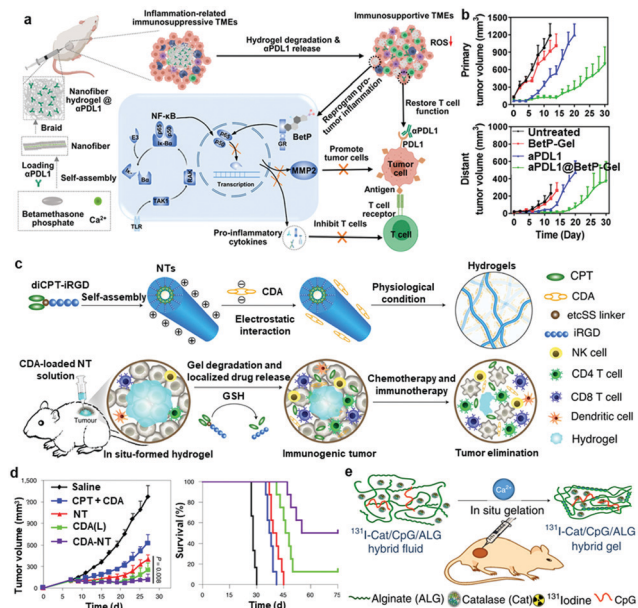


Fig. 4 (a) Schematic showing the formation of nanofiber hydrogels by cross-linking filamentous assemblies via physical interaction between betamethasone phosphate and calcium ions. This nanofiber hydrogel could reprogram the protumoral immunosuppressive TME by inhibiting the NF- κ B signaling pathway and sustainably release aPD1 to activate T cells. (b) Growth kinetics of primary and distant CT26 tumors in different groups.⁷³ Copyright 2021, American Chemical Society. (c) Schematics of localized CPT and CDA delivery using a bioresponsive CPT-based nanotube hydrogel for TME regulation and chemoimmunotherapy. (d) Tumour-growth kinetics and survival curve of GL-261 tumour-bearing mice in different groups.¹¹⁶ Copyright 2020, Springer Nature. (e) Scheme illustrating *in situ* gelation of a ¹³¹I-Cat/ALG hybrid fluid after local injection into tumours.¹¹⁷ Copyright 2018, Springer Nature.

on the RADA16 peptide to prepare DC-based vaccines.¹¹⁰ The injectable RADA16 peptide hydrogel could effectively deliver exogenous DCs, antigens, and aPD-L1 antibody simultaneously in a minimally invasive manner, significantly enhancing the proliferation of antigen-specific T cells and inducing potent cellular immune responses. In another example, Song *et al.* engineered an injectable PEG-*b*-poly(L-alanine) hydrogel for sustained local codelivery of tumor cell lysate, granulocyte-macrophage colony stimulating factor, and immune checkpoint inhibitors, achieving significantly enhanced tumor-specific immune responses.¹¹¹ More recently, self-delivery hydrogels that are directly self-assembled by bioactive gelators for delivery to target sites have become popular. In a recent work by our group, an anti-inflammatory nanofiber hydrogel self-assembled by steroid drugs was developed for the local delivery of aPD1 to achieve systemic cancer immunotherapy (Fig. 4a).⁷³ Interestingly, such a carrier-free system based on steroid drugs could not only reprogram the immunosuppressive TME to an antitumoral microenvironment but also serve as a reservoir for sustained release of aPD1, effectively inhibiting the growth of both local and abscopal tumors (Fig. 4b). In another study, Yang *et al.* developed a D- or L-peptide self-assembled supramolecular hydrogel with OVA entrapped in the cavity or physically adsorbed

on the surface of the nanofibers.¹¹² Compared with L-gel, D-gel was capable of serving as a promising vaccine adjuvant to evoke both humoral and cellular immune responses. Jiang *et al.* also demonstrated that right-handed fiber hydrogels could induce stronger humoral and cellular immune responses than left-handed hydrogels.¹¹³

Apart from injectable supramolecular hydrogels, supramolecular hydrogels formed *in situ* are also a useful delivery system for cancer immunotherapy, which could entrap bioactive molecules or cells by simple injection at the targeted sites.¹¹⁴ These supramolecular hydrogels usually have the ability to immediately undergo morphological changes to external stimuli, realizing sustained and controlled release of encapsulated therapeutics.¹¹⁵ Cui *et al.* developed a supramolecular hydrogel based on peptide-drug conjugates to intratumorally deliver a STING agonist to activate cancer immunotherapy (Fig. 4c).¹¹⁶ In aqueous solution, the synthesized drug amphiphile (diCPT-iRGD), consisting of a peptide moiety iRGD and camptothecin (CPT), could spontaneously assemble into supramolecular nanotubes. The negatively charged STING agonist (CDA) could be absorbed on the surface of positively charged nanotubes through electrostatic complexations. After injection into the tumor site, the nanotubes immediately formed hydrogels upon response to counterions, functioning as the local reservoir for extended local release of CDA and CPT to awake both innate and adaptive immune systems (Fig. 4d). They also demonstrated that aPD1 could be effectively delivered into tumors using such matrix metalloproteinase-responsive supramolecular hydrogels.⁴² In another study, Liu and coworkers designed an *in situ* formed sodium alginate (ALG) gel containing radioisotope-labeled catalase (¹³¹I-Cat) and CpG oligonucleotides.¹¹⁷ Upon injection, the ALG fluid containing different drugs rapidly transformed into a gel by coordination with endogenous Ca²⁺ ions within the tumor (Fig. 4e). When combined with an immune checkpoint blockade, this gel could induce strong antitumor immune responses to attack distant cancer cells and strong immunological memory effects to inhibit cancer recurrence. In addition, Ma and coworkers developed thermosensitive hydrogels based on triblock copolymers (PLGA-PEG-PLGA) for the sustained release of IL2.¹¹⁸ With the appropriate gelation temperature at 29.5–30 °C, thermosensitive hydrogels containing IL2 could be injected into any site of the body in a minimally invasive and highly efficient manner.

4. Challenges and future outlook

In this review, we summarized the recent significant research advancements of supramolecular biomaterials for cancer immunotherapy. By modulating multiple noncovalent interactions (hydrophobic, hydrogen bonding, electrostatic and coordination interactions), the physicochemical properties (size, morphology, charge, and specific surface area) and responsiveness of supramolecular biomaterials could be elaborately refined, facilitating the construction of functional adjustable

platforms for cancer immunotherapy. The dynamic and adaptive nature of self-assembled nanoarchitectures affords enhanced sensitivity to changes in environmental conditions, favoring the spatiotemporal modulation of payload encapsulation and liberation. These nanoarchitectures with various dimensions and different topologies, mainly including nanocapsules, nanoparticles, filamentous micelles, nanofibers and hydrogels, possess distinguishing advantages in applications for cancer immunotherapy, incarnating a structure–property relationship. Thus, supramolecular biomaterials could target multiple vulnerabilities of cancer to boost the antitumor immune response effectively and safely.

Compared with the existing immunotherapy systems, supramolecular biomaterials exhibit many unique advantages for cancer immunotherapy. First, self-assembly is a simple, flexible and green fabrication strategy. Supramolecular biomaterials integrate multifunctionality into immunotherapy systems without time consumption, expensive synthesis processes and toxic reagents, making these products more clinically translatable. Second, supramolecular biomaterials could achieve high drug encapsulation efficiency by coassembly *via* noncovalent interactions, and the physical interaction could finely retain the activity of immunotherapeutics. Third, due to their dynamic and adaptive nature, supramolecular biomaterials afford enhanced sensitivity to circumstance cues, favoring more rational and controllable therapeutic encapsulation or liberation. Finally, physiochemical properties, such as topological structure, surface charge and antigen density of supramolecular biomaterials, are flexibly modulated, which could exert synergistic functions with immune therapeutics to amplify immune responses and improve cancer immunotherapy.

Despite these unique advantages of supramolecular biomaterials for cancer immunotherapy, successful clinical translations of these nanoformulations remain challenging. First of all, nanomedicines administered intravenously face many extracellular and intracellular barriers *in vivo*. Due to the weak noncovalent properties, attention should be paid to the stability of supramolecular biomaterials during blood circulation in cancer immunotherapy. Synergism and cooperativity of various non-covalent interactions could be considered in the design and fabrication of supramolecular nanomedicines to improve their stability and immunotherapy performance. Second, although supramolecular immunotherapeutics could co-deliver different immunotherapeutic agents to achieve combination therapy, controllable release of multiple therapeutic agents spatiotemporally remains challenging and needs improvements. Multiple responsive release modalities and noncovalent bonds with different strengths could be involved in the supramolecular nanomedicines to construct elegant responsive nanoplatfoms. Last but not least, the long-term safety and toxicity profiles of supramolecular nanomedicine still remain obstacles for clinical cancer immunotherapy. The non-biocompatible carriers and untargeted drug delivery are the two main reasons for adverse effects caused in immunotherapy. On the one hand, employment of non-immunogenic constituents and the use of a drug itself as a building block may

reduce many adverse interactions with immune systems. Additionally, judicious investigation of the surface physiochemical properties of supramolecular nanoplatfoms and modification of targeting groups could be valuable for achieving precise and safe immunotherapy.

As stated, engineering elegant nanoplatfoms in the past decade provides potential strategies to improve cancer immunotherapy. It is hoped that continuous advances in this field will soon overcome the existing difficulties for further development of supramolecular immunotherapeutics. As interest in this research field continues to evolve, it is also anticipated that supramolecular nanotechnology will inspire the development of many novel and powerful approaches for cancer immunotherapy and act as one of the key drivers for successful clinical transformation in the near future.

Conflicts of interest

The authors declare that they have no competing interests.

Acknowledgements

This work was partially supported by the National Natural Science Foundation of China (91959104, 21927803, 51903182, 51525203 and 52103347), the Natural Science Foundation of Jiangsu Province (BK20190826), the China Postdoctoral Science Foundation (2020M671583), the Collaborative Innovation Center of Suzhou Nano Science and Technology, the 111 Program from the Ministry of Education of China, the Start-Up Package of McGill University (G. C.), NSERC Discovery Grant (G. C.), and GCI studentship (T. F.).

Notes and references

- 1 R. D. Schreiber, L. J. Old and M. J. Smyth, *Science*, 2011, **331**, 1565–1570.
- 2 I. Mellman, G. Coukos and G. Dranoff, *Nature*, 2011, **480**, 480–489.
- 3 D. F. Quail and J. A. Joyce, *Nat. Med.*, 2013, **19**, 1423–1437.
- 4 D. S. Chen and I. Mellman, *Immunity*, 2013, **39**, 1–10.
- 5 A. Stoddart, *Nat. Rev. Mater.*, 2017, **2**, 17027.
- 6 O. J. Finn, *Nat. Rev. Immunol.*, 2003, **3**, 630–641.
- 7 M. S. Gebre, L. A. Brito, L. H. Tostanoski, D. K. Edwards, A. Carfi and D. H. Barouch, *Cell*, 2021, **184**, 1589–1603.
- 8 M. B. Howren, D. M. Lamkin and J. Suls, *Psychosom. Med.*, 2009, **71**, 171–186.
- 9 A. Ribas and J. D. Wolchok, *Science*, 2018, **359**, 1350–1355.
- 10 P. C. Tume, C. L. Harview, J. H. Yearley, I. P. Shintaku, E. J. M. Taylor, L. Robert, B. Chmielowski, M. Spasic, G. Henry, V. Ciobanu, A. N. West, M. Carmona, C. Kivork, E. Seja, G. Cherry, A. J. Gutierrez, T. R. Grogan, C. Mateus, G. Tomasic, J. A. Glaspy, R. O. Emerson, H. Robins, R. H. Pierce, D. A. Elashoff, C. Robert and A. Ribas, *Nature*, 2014, **515**, 568–571.
- 11 C. H. June, *J. Clin. Invest.*, 2007, **117**, 1466–1476.

- 12 S. A. Rosenberg, J. C. Yang and N. P. Restifo, *Nat. Med.*, 2004, **10**, 909–915.
- 13 A. York, *Nat. Rev. Microbiol.*, 2021, **19**, 222–223.
- 14 R. Kuai, L. J. Ochyl, K. S. Bahjat, A. Schwendeman and J. J. Moon, *Nat. Mater.*, 2017, **16**, 489–496.
- 15 Y. Jing, J. Liu, Y. Ye, L. Pan, H. Deng, Y. Wang, Y. Yang, L. Diao, S. H. Lin, G. B. Mills, G. Zhuang, X. Xue and L. Han, *Nat. Commun.*, 2020, **11**, 4946.
- 16 M. S. Goldberg, *Nat. Rev. Cancer*, 2019, **19**, 587–602.
- 17 G. Dranoff, *Nat. Rev. Cancer*, 2004, **4**, 11–22.
- 18 R. S. Riley, C. H. June, R. Langer and M. J. Mitchell, *Nat. Rev. Drug Discovery*, 2019, **18**, 175–196.
- 19 G. Liu, M. Zhu, X. Zhao and G. Nie, *Adv. Drug Delivery Rev.*, 2021, **176**, 113889.
- 20 N. Gong, Y. Zhang, X. Teng, Y. Wang, S. Huo, G. Qing, Q. Ni, X. Li, J. Wang, X. Ye, T. Zhang, S. Chen, Y. Wang, J. Yu, P. C. Wang, Y. Gan, J. Zhang, M. J. Mitchell, J. Li and X.-J. Liang, *Nat. Nanotechnol.*, 2020, **15**, 1053–1064.
- 21 C.-T. Jiang, K.-G. Chen, A. Liu, H. Huang, Y.-N. Fan, D.-K. Zhao, Q.-N. Ye, H.-B. Zhang, C.-F. Xu, S. Shen, M.-H. Xiong, J.-Z. Du, X.-Z. Yang and J. Wang, *Nat. Commun.*, 2021, **12**, 1359.
- 22 Y. Xia, T. Song, Y. Hu and G. Ma, *Acc. Chem. Res.*, 2020, **53**, 2068–2080.
- 23 J. B. A. G. Haanen, F. Carbone, C. Robert, K. M. Kerr, S. Peters, J. Larkin and K. Jordan, *Ann. Oncol.*, 2017, **28**, 119–142.
- 24 P. Xing and Y. Zhao, *Small Methods*, 2018, **2**, 1700364.
- 25 J. Wang, Y. Li and G. Nie, *Nat. Rev. Mater.*, 2021, **6**, 766–783.
- 26 Q. Chen, M. Chen and Z. Liu, *Chem. Soc. Rev.*, 2019, **48**, 5506–5526.
- 27 J. H. van Esch, *Nature*, 2010, **466**, 193–194.
- 28 T. Aida, E. W. Meijer and S. I. Stupp, *Science*, 2012, **335**, 813–817.
- 29 T. Fukino, H. Joo, Y. Hisada, M. Obana, H. Yamagishi, T. Hikima, M. Takata, N. Fujita and T. Aida, *Science*, 2014, **344**, 499–504.
- 30 J. Wang, K. Liu, R. Xing and X. Yan, *Chem. Soc. Rev.*, 2016, **45**, 5589–5604.
- 31 S. Zhang, *Nat. Biotechnol.*, 2003, **21**, 1171–1178.
- 32 Q. Chen, C. Wang, X. Zhang, G. Chen, Q. Hu, H. Li, J. Wang, D. Wen, Y. Zhang, Y. Lu, G. Yang, C. Jiang, J. Wang, G. Dotti and Z. Gu, *Nat. Nanotechnol.*, 2019, **14**, 89–97.
- 33 Y. Zhang, S. Ma, X. Liu, Y. Xu, J. Zhao, X. Si, H. Li, Z. Huang, Z. Wang, Z. Tang, W. Song and X. Chen, *Adv. Mater.*, 2021, **33**, 2007293.
- 34 D. P. Patterson, A. Rynda-Applé, A. L. Harmsen, A. G. Harmsen and T. Douglas, *ACS Nano*, 2013, **7**, 3036–3044.
- 35 Z. Zhou, C. Du, Q. Zhang, G. Yu, F. Zhang and X. Chen, *Angew. Chem., Int. Ed.*, 2021, **60**, 21033–21039.
- 36 Y. Xia, J. Wu, W. Wei, Y. Du, T. Wan, X. Ma, W. An, A. Guo, C. Miao, H. Yue, S. Li, X. Cao, Z. Su and G. Ma, *Nat. Mater.*, 2018, **17**, 187–194.
- 37 H. Zhang, K. Liu, S. Li, X. Xin, S. Yuan, G. Ma and X. Yan, *ACS Nano*, 2018, **12**, 8266–8276.
- 38 J. Zheng, R. Fan, H. Wu, H. Yao, Y. Yan, J. Liu, L. Ran, Z. Sun, L. Yi, L. Dang, P. Gan, P. Zheng, T. Yang, Y. Zhang, T. Tang and Y. Wang, *Nat. Commun.*, 2019, **10**, 1604.
- 39 E. N. Chin, C. Yu, V. F. Vartabedian, Y. Jia, M. Kumar, A. M. Gamo, W. Vernier, S. H. Ali, M. Kissai, D. C. Lazar, N. Nguyen, L. E. Pereira, B. Benish, A. K. Woods, S. B. Joseph, A. Chu, K. A. Johnson, P. N. Sander, F. Martínez-Peña, E. N. Hampton, T. S. Young, D. W. Wolan, A. K. Chatterjee, P. G. Schultz, H. M. Pettrassi, J. R. Teijaro and L. L. Lairson, *Science*, 2020, **369**, 993–999.
- 40 L. Zhang, D. Jing, N. Jiang, T. Rojalin, C. M. Baehr, D. Zhang, W. Xiao, Y. Wu, Z. Cong, J. J. Li, Y. Li, L. Wang and K. S. Lam, *Nat. Nanotechnol.*, 2020, **15**, 145–153.
- 41 X. Li, Y. Wang, Y. Zhang, C. Liang, Z. Zhang, Y. Chen, Z. Hu and Z. Yang, *Adv. Funct. Mater.*, 2021, **31**, 2100729.
- 42 F. Wang, D. Xu, H. Su, W. Zhang, X. Sun, M. K. Monroe, R. W. Chakroun, Z. Wang, W. Dai, R. Oh, H. Wang, Q. Fan, F. Wan and H. Cui, *Sci. Adv.*, 2020, **6**, eaaz8985.
- 43 N. Guzewicz, A. Best, B. Perez-Ramirez and D. L. Kaplan, *Biomaterials*, 2011, **32**, 2642–2650.
- 44 S. B. Stephan, A. M. Taber, I. Jileeva, E. P. Pegues, C. L. Sentman and M. T. Stephan, *Nat. Biotechnol.*, 2015, **33**, 97–101.
- 45 J. Zhou, L. Rao, G. Yu, T. R. Cook, X. Chen and F. Huang, *Chem. Soc. Rev.*, 2021, **50**, 2839–2891.
- 46 Z. Shen, H. Ye, X. Yi and Y. Li, *ACS Nano*, 2019, **13**, 215–228.
- 47 C. W. Shields, L. L. Wang, M. A. Evans and S. Mitragotri, *Adv. Mater.*, 2020, **32**, 1901633.
- 48 X. Feng, W. Xu, Z. Li, W. Song, J. Ding and X. Chen, *Adv. Sci.*, 2019, **6**, 1900101.
- 49 E. Froimchuk, S. T. Carey, C. Edwards and C. M. Jewell, *Acc. Chem. Res.*, 2020, **53**, 2534–2545.
- 50 H. B. Eppler and C. M. Jewell, *Adv. Mater.*, 2020, **32**, 1903367.
- 51 T. Bhattacharyya, P. Saha and J. Dash, *ACS Omega*, 2018, **3**, 2230–2241.
- 52 X. Xu, Y. Jian, Y. Li, X. Zhang, Z. Tu and Z. Gu, *ACS Nano*, 2014, **8**, 9255–9264.
- 53 M. J. Webber, E. A. Appel, E. W. Meijer and R. Langer, *Nat. Mater.*, 2016, **15**, 13–26.
- 54 M. J. Webber and R. Langer, *Chem. Soc. Rev.*, 2017, **46**, 6600–6620.
- 55 S. Toksoz, H. Acar and M. O. Guler, *Soft Matter*, 2010, **6**, 5839.
- 56 J. Voskuhl and B. J. Ravoo, *Chem. Soc. Rev.*, 2009, **38**, 495–505.
- 57 H. Pick, A. C. Alves and H. Vogel, *Chem. Rev.*, 2018, **118**, 8598–8654.
- 58 W. Song, J. Kuang, C.-X. Li, M. Zhang, D. Zheng, X. Zeng, C. Liu and X.-Z. Zhang, *ACS Nano*, 2018, **12**, 1978–1989.
- 59 J. F. Lovell, C. S. Jin, E. Huynh, H. Jin, C. Kim, J. L. Rubinstein, W. C. W. Chan, W. Cao, L. V. Wang and G. Zheng, *Nat. Mater.*, 2011, **10**, 324–332.
- 60 M. J. Sailor and J.-H. Park, *Adv. Mater.*, 2012, **24**, 3779–3802.

- 61 T. P. Knowles, A. W. Fitzpatrick, S. Meehan, H. R. Mott, M. Vendruscolo, C. M. Dobson and M. E. Welland, *Science*, 2007, **318**, 1900–1903.
- 62 L. Zhou, T. Qiu, F. Lv, L. Liu, J. Ying and S. Wang, *Adv. Healthcare Mater.*, 2018, **7**, 1800670.
- 63 H.-W. An, L.-L. Li, Y. Wang, Z. Wang, D. Hou, Y.-X. Lin, S.-L. Qiao, M.-D. Wang, C. Yang, Y. Cong, Y. Ma, X.-X. Zhao, Q. Cai, W.-T. Chen, C.-Q. Lu, W. Xu, H. Wang and Y. Zhao, *Nat. Commun.*, 2019, **10**, 1–15.
- 64 J. J. Richardson, M. Björnmalm and F. Caruso, *Science*, 2015, **348**, aaa2491.
- 65 Q. Jin, W. Zhu, J. Zhu, J. Shen, Z. Liu, Y. Yang and Q. Chen, *Adv. Mater.*, 2021, **33**, 2007557.
- 66 H. Zhang, L. Kang, Q. Zou, X. Xin and X. Yan, *Curr. Opin. Biotechnol.*, 2019, **58**, 45–52.
- 67 J. D. Brodin, X. I. Ambroggio, C. Tang, K. N. Parent, T. S. Baker and F. A. Tezcan, *Nat. Chem.*, 2012, **4**, 375–382.
- 68 S. Mura, J. Nicolas and P. Couvreur, *Nat. Mater.*, 2013, **12**, 991–1003.
- 69 M. Grzelczak, L. M. Liz-Marzán and R. Klajn, *Chem. Soc. Rev.*, 2019, **48**, 1342–1361.
- 70 T. J. Moyer, J. A. Finbloom, F. Chen, D. J. Toft, V. L. Cryns and S. I. Stupp, *J. Am. Chem. Soc.*, 2014, **136**, 14746–14752.
- 71 K. Liu, R. Xing, Q. Zou, G. Ma, H. Möhwald and X. Yan, *Angew. Chem., Int. Ed.*, 2016, **55**, 3036–3039.
- 72 P. Zheng, B. B. Ding, Z. Y. Jiang, W. G. Xu, G. Li, J. X. Ding and X. Chen, *Nano Lett.*, 2021, **21**, 2088–2093.
- 73 M. Chen, Y. Tan, Z. Dong, J. Lu, X. Han, Q. Jin, W. Zhu, J. Shen, L. Cheng, Z. Liu and Q. Chen, *Nano Lett.*, 2020, **20**, 6763–6773.
- 74 G.-B. Qi, Y.-J. Gao, L. Wang and H. Wang, *Adv. Mater.*, 2018, **30**, 1703444.
- 75 C. Ji, Q. Gao, X. Dong, W. Yin, Z. Gu, Z. Gan, Y. Zhao and M. Yin, *Angew. Chem., Int. Ed.*, 2018, **57**, 11384–11388.
- 76 D. Zhang, G.-B. Qi, Y.-X. Zhao, S.-L. Qiao, C. Yang and H. Wang, *Adv. Mater.*, 2015, **27**, 6125–6130.
- 77 J. Yang, H.-W. An and H. Wang, *ACS Appl. Bio Mater.*, 2021, **4**, 24–46.
- 78 Z. Huang, W. Song and X. Chen, *Front. Chem.*, 2020, **8**(380), 1.
- 79 K. Panneerselvam, M. E. Lynge, C. F. Riber, S. Mena-Hernando, A. A. A. Smith, K. N. Goldie, A. N. Zelikin and B. Städler, *Biomicrofluidics*, 2015, **9**, 052610.
- 80 M. Ramanathan, L. K. Shrestha, T. Mori, Q. Ji, J. P. Hill and K. Ariga, *Phys. Chem. Chem. Phys.*, 2013, **15**, 10580.
- 81 S. Wu, Y. Xia, Y. Hu and G. Ma, *Adv. Drug Delivery Rev.*, 2021, **176**, 113871.
- 82 G. M. Lynn, C. Sedlik, F. Baharom, Y. Zhu, R. A. Ramirez-Valdez, V. L. Coble, K. Tobin, S. R. Nichols, Y. Itzkowitz, N. Zaidi, J. M. Gammon, N. J. Blobel, J. Denizeau, P. de la Rochere, B. J. Francica, B. Decker, M. Maciejewski, J. Cheung, H. Yamane, M. G. Smelkinson, J. R. Francica, R. Laga, J. D. Bernstock, L. W. Seymour, C. G. Drake, C. M. Jewell, O. Lantz, E. Piaggio, A. S. Ishizuka and R. A. Seder, *Nat. Biotechnol.*, 2020, **38**, 320–332.
- 83 Y. Wang, H. Xu and X. Zhang, *Adv. Mater.*, 2009, **21**, 2849–2864.
- 84 F. Zhou, B. Feng, H. Yu, D. Wang, T. Wang, Y. Ma, S. Wang and Y. Li, *Adv. Mater.*, 2019, **31**, 1805888.
- 85 S. T. Koshy, A. S. Cheung, L. Gu, A. R. Graveline and D. J. Mooney, *Adv. Biosyst.*, 2017, **1**, 1600013.
- 86 W. Yang, H. Deng, S. Zhu, J. Lau, R. Tian, S. Wang, Z. Zhou, G. Yu, L. Rao, L. He, Y. Ma and X. Chen, *Sci. Adv.*, 2020, **6**, eabd1631.
- 87 S. Naahidi, M. Jafari, F. Edalat, K. Raymond, A. Khademhosseini and P. Chen, *J. Controlled Release*, 2013, **166**, 182–194.
- 88 L.-P. Zhao, R.-R. Zheng, J.-Q. Huang, X.-Y. Chen, F.-A. Deng, Y.-B. Liu, C.-Y. Huang, X.-Y. Yu, H. Cheng and S.-Y. Li, *ACS Nano*, 2020, **14**, 17100–17113.
- 89 H. Zhao, J. Xu, Y. Li, X. Guan, X. Han, Y. Xu, H. Zhou, R. Peng, J. Wang and Z. Liu, *ACS Nano*, 2019, **13**, 13127–13135.
- 90 G. M. Lynn, R. Laga, P. A. Darrah, A. S. Ishizuka, A. J. Balaci, A. E. Dulcey, M. Pechar, R. Pola, M. Y. Gerner, A. Yamamoto, C. R. Buechler, K. M. Quinn, M. G. Smelkinson, O. Vanek, R. Cawood, T. Hills, O. Vasalatiy, K. Kastenmüller, J. R. Francica, L. Stutts, J. K. Tom, K. A. Ryu, A. P. Esser-Kahn, T. Etrych, K. D. Fisher, L. W. Seymour and R. A. Seder, *Nat. Biotechnol.*, 2015, **33**, 1201–1210.
- 91 Y. Sun, X. Feng, C. Wan, J. F. Lovell, H. Jin and J. Ding, *Asian J. Pharm.*, 2021, **16**, 129–132.
- 92 Q. Sun, M. Barz, B. G. De Geest, M. Diken, W. E. Hennink, F. Kiessling, T. Lammers and Y. Shi, *Chem. Soc. Rev.*, 2019, **48**, 351–381.
- 93 K. Ariga, M. Nishikawa, T. Mori, J. Takeya, L. K. Shrestha and J. P. Hill, *Sci. Technol. Adv. Mater.*, 2019, **20**, 51–95.
- 94 J. S. Rudra, Y. F. Tian, J. P. Jung and J. H. Collier, *Proc. Natl. Acad. Sci. U. S. A.*, 2010, **107**, 622–627.
- 95 C. Mora-Solano, Y. Wen, H. Han, J. Chen, A. S. Chong, M. L. Miller, R. R. Pompano and J. H. Collier, *Biomaterials*, 2017, **149**, 1–11.
- 96 Y. Si, Y. Wen, S. H. Kelly, A. S. Chong and J. H. Collier, *J. Controlled Release*, 2018, **282**, 120–130.
- 97 Y. Si, Q. Tian, F. Zhao, S. H. Kelly, L. S. Shores, D. F. Camacho, A. I. Sperling, M. S. Andrade, J. H. Collier and A. S. Chong, *Sci. Adv.*, 2020, **6**, eaba0995.
- 98 R. Appavu, C. B. Chesson, A. Y. Koyfman, J. D. Snook, F. J. Kohlhapp, A. Zloza and J. S. Rudra, *ACS Biomater. Sci. Eng.*, 2015, **1**, 601–609.
- 99 J. Mougín, C. Bourgaux and P. Couvreur, *Adv. Drug Delivery Rev.*, 2021, **172**, 127–147.
- 100 S. Yi, S. D. Allen, Y.-G. Liu, B. Z. Ouyang, X. Li, P. Augsornworawat, E. B. Thorp and E. A. Scott, *ACS Nano*, 2016, **10**, 11290–11303.
- 101 N. B. Karabin, S. Allen, H.-K. Kwon, S. Bobbala, E. Firlar, T. Shokuhfar, K. R. Shull and E. A. Scott, *Nat. Commun.*, 2018, **9**, 1–13.
- 102 Z. Huang, D. Yao, Q. Ye, H. Jiang, R. Gu, C. Ji, J. Wu, Y. Hu and A. Yuan, *ACS Nano*, 2021, **15**, 8450–8465.
- 103 R. Mammadov, G. Cinar, N. Gunduz, M. Goktas, H. Kayhan, S. Tohumeken, A. E. Topal, I. Orujalipoor,

- T. Delibasi, A. Dana, S. Ide, A. B. Tekinay and M. O. Guler, *Sci. Rep.*, 2015, **5**, 16728.
- 104 S. Li, W. Zhang, R. Xing, C. Yuan, H. Xue and X. Yan, *Adv. Mater.*, 2021, **33**, 2100595.
- 105 Z. Guo, Y. Hu, M. Zhao, K. Hao, P. He, H. Tian, X. Chen and M. Chen, *Nano Lett.*, 2021, **21**, 3721–3730.
- 106 M. Norouzi, B. Nazari and D. W. Miller, *Drug Discovery Today*, 2016, **21**, 1835–1849.
- 107 Z. Li, N. Song and Y.-W. Yang, *Matter*, 2019, **1**, 345–368.
- 108 R. Narayanaswamy and V. P. Torchilin, *Molecules*, 2019, **24**, 603.
- 109 D. Fan, Y. Tian and Z. Liu, *Front. Chem.*, 2019, **675**, 1–11.
- 110 P. Yang, H. Song, Y. Qin, P. Huang, C. Zhang, D. Kong and W. Wang, *Nano Lett.*, 2018, **18**, 4377–4385.
- 111 H. Song, P. Yang, P. Huang, C. Zhang, D. Kong and W. Wang, *Theranostics*, 2019, **9**, 2299–2314.
- 112 Z. Luo, Q. Wu, C. Yang, H. Wang, T. He, Y. Wang, Z. Wang, H. Chen, X. Li, C. Gong and Z. Yang, *Adv. Mater.*, 2017, **29**, 1601776.
- 113 Y. Tian, H. Wang, Y. Liu, L. Mao, W. Chen, Z. Zhu, W. Liu, W. Zheng, Y. Zhao, D. Kong, Z. Yang, W. Zhang, Y. Shao and X. Jiang, *Nano Lett.*, 2014, **14**, 1439–1445.
- 114 Y. Chao, Q. Chen and Z. Liu, *Adv. Funct. Mater.*, 2020, **30**, 1902785.
- 115 R. Dong, Y. Zhou, X. Huang, X. Zhu, Y. Lu and J. Shen, *Adv. Mater.*, 2015, **27**, 498–526.
- 116 F. Wang, H. Su, D. Xu, W. Dai, W. Zhang, Z. Wang, C. F. Anderson, M. Zheng, R. Oh, F. Wan and H. Cui, *Nat. Biomed. Eng.*, 2020, **4**, 1090–1101.
- 117 Y. Chao, L. Xu, C. Liang, L. Feng, J. Xu, Z. Dong, L. Tian, X. Yi, K. Yang and Z. Liu, *Nat. Biomed. Eng.*, 2018, **2**, 611–621.
- 118 Q. Lv, C. He, F. Quan, S. Yu and X. Chen, *Bioact. Mater.*, 2018, **3**, 118–128.