



Cite this: *J. Anal. At. Spectrom.*, 2020, 35, 1011

'Blind time' – current limitations on laser ablation multi-collector inductively coupled plasma mass spectrometry (LA-MC-ICP-MS) for ultra-transient signal isotope ratio analysis and application to individual sub-micron sized uranium particles

Grant Craig, *^{ab} Matthew S. A. Horstwood, ^c Helen J. Reid^a and Barry L. Sharp ^a

The application of laser ablation multi-collector inductively coupled plasma mass spectrometry (LA-MC-ICP-MS) to the isotope ratio analysis of UO_x particles has the potential to improve the isotopic determination of these particles when compared to currently utilised ICP-MS techniques. To investigate this a high-speed, integrated ablation cell and dual concentric injector design was tested in the expectation that the resulting increase in signal to noise ratio and sample ion yield would improve the determination of $^{234}\text{U}/^{238}\text{U}$, $^{235}\text{U}/^{238}\text{U}$ and $^{236}\text{U}/^{238}\text{U}$ for such materials. However, when compared to a slower washout, more established low-volume cell design, the highly transient signals of the new design proved challenging for the mixed detector array of the multi-collector mass spectrometer, introducing a new bias. We describe a major component of this bias, referred to as 'blind time', and model its impact on UO_x particle analysis. After accounting for blind time, average precisions for the uranium isotopic composition of sub-micron sized UO_x particles using LA-MC-ICP-MS were 3% 1RSD for $^{235}\text{U}/^{238}\text{U}$ and 8% 1RSD for $^{234}\text{U}/^{238}\text{U}$. When ablating a glass rather than a UO_x particle, uncertainties of 1.3% 1RSD for $^{235}\text{U}/^{238}\text{U}$ were achieved for 150 nm equivalent particle sizes using LA-MC-ICP-MS.

Received 20th February 2020
Accepted 16th April 2020

DOI: 10.1039/d0ja00066c

rsc.li/jaas

Introduction

International governmental agencies have introduced safeguards designed to ensure the compliance of nuclear facilities (e.g. nuclear reactors, enrichment facilities or test sites) to stated declarations and to detect undeclared, unauthorised activities.¹ A prime concern of these safeguards are microparticles (down to a few hundred nanometres in diameter) containing perhaps only picograms of actinide material. Such actinide-bearing particles are also vital to nuclear forensics^{2,3} (the study of intercepted nuclear material) and to environmental studies of previously contaminated locations.^{4,5} For all three areas of interest the isotopic composition can give a wealth of information including but not limited to the sample age, intended use and place of manufacture.^{3,6–8} Routine inspections for safeguards analysis use cotton swipes to collect particles which are then extracted to determine the uranium (U) or plutonium (Pu) isotopic composition by mass spectrometry.⁹

Dissolution of the collected particles for analysis by inductively coupled plasma mass spectrometry (ICP-MS) is routine,¹⁰ however the isotope ratios recovered are for the bulk of the material: analysing each particle individually could reveal a strong deviation from the bulk.¹¹ Separation and dissolution of individual U particles has been successfully implemented for ICP-MS,^{12–14} and successfully compared to secondary ion mass spectrometry (SIMS) analysis of the same material.¹⁵ For routine analysis thermal ionisation mass spectrometry (TIMS) and SIMS are commonly used for uranium particles, with SIMS predominant.¹ However other forms of mass spectrometry, including laser ablation (LA-)ICP-MS, have been investigated as complementary techniques.

LA-ICP-MS is a promising technique for nuclear safeguards, requiring less sample preparation than either TIMS or SIMS. The primary advantages of LA-ICP-MS are speed of analysis, more widespread instrumentation and therefore accessibility and ease of use. Over time, sample size for LA-ICP-MS has varied from pressed powder pellets¹⁶ to 10–30 μm particles^{11,17,18} and smaller.¹⁹ Recently the scale has shifted from the micron to submicron,^{20–24} attention being focussed by recent inter-laboratory comparisons.^{25–27} To date, several studies have analysed micron and sub-micron uranium particles by LA-ICP-MS. Pointurier *et al.*²¹ compared a 213 nm laser system coupled with a quadrupole-based ICP-MS, to TIMS and SIMS for $^{235}\text{U}/^{238}\text{U}$

^aCentre for Analytical Science, Department of Chemistry, Loughborough University, Loughborough, Leicestershire, LE11 3TU, UK. E-mail: grant.craig@thermofisher.com

^bThermo Fisher Scientific Bremen GmbH, Hanna-Kunath Str. 11, 28359 Bremen, Germany

^cNERC Isotope Geosciences Laboratory, British Geological Survey, Nicker Hill, Keyworth, Nottinghamshire, NG12 5GG, UK

isotope ratio analysis. Kappel *et al.*²² analysed $^{235}\text{U}/^{238}\text{U}$ by LA-MC-ICP-MS as part of an evaluation of data handling strategies for single particles and Claverie *et al.*²⁸ applied LA-MC-ICP-MS to micron-sized uranium particles, measuring both $^{235}\text{U}/^{238}\text{U}$ and $^{234}\text{U}/^{238}\text{U}$ to a precision of 2.0% 1RSD using a multiple Faraday configuration. Critically, the flexibility of ICP-MS allows modification of signal to noise ratios (SNR) by adapting conventional analysis approaches. Increasing the SNR improves the limit of detection and limit of quantification, important when considering the lack of material in sub-micron uranium particles. The SNR can be improved by compressing the transient signals generated into a shorter time frame. Short transient signals are challenging in ICP-MS, in terms of data acquisition and handling by both the ICP-MS instrument and the user,²⁹ requiring changes in methodology. One such change has been calculating isotope ratios from entire integrated transient signals rather than by each mass spectrometer cycle.^{18,30,31} Isotope ratio analysis of transient signals has already been pioneered for single shot laser ablation of larger materials^{30,32} as well as single particles. Spectral skew is a known problem for transient signal analysis with single collector ICP-MS, whereby inaccuracies are introduced due to differential delays in signal acquisition.³³ Adopting pseudo-simultaneous detection to overcome spectral skew with LA-ICP-TOF-MS for uranium particle analysis has been investigated but was limited in detector linearity and sample ion yield.²³ Multi-collector inductively coupled plasma mass spectrometers (MC-ICP-MS) achieves full simultaneous measurement by collecting each isotope on separate detectors as well as high ion yield (>7%, Craig *et al.*³⁴) and as such should be ideally suited for highly transient signals. However, although isotopes signals are detected simultaneously on MC-ICP-MS, the isotope signals need not be output simultaneously. For Faraday cup detectors the signal output is reliant on the first order tau constant of the resistor in the associated amplifier.^{35,36} As such, differential decay of the signal on different Faraday detectors can lead to variation in transient signal isotope ratio analysis. For highly transient signals, this can introduce bias in isotope ratio measurements *e.g.* Krupp and Donard³⁷ This differential detector response is at its most extreme when comparing output signals from transient pulses recorded on mixed detector arrays (*e.g.* Faradays and ion counters).

We set out to investigate the utility of LA-MC-ICP-MS for analysis of individual submicron UO_x particles for U isotope ratio determination. Given the small quantity of material to be analysed and the low abundance of ^{234}U and ^{236}U in many samples it was hypothesised that a high detection efficiency and signal to noise ratio would be required. To this end we determined to apply an ultrafast combined ablation cell and torch injector design, the prototype DCI,³⁸ to the LA-MC-ICP-MS system. This system built on a key area of development in laser ablation cell design, “two-volume” cells, where a smaller cell is housed inside a larger casing. These cells have better flow dynamics than previous generations of ‘single volume’ ablation cells and are now standard issue from most manufacturers.³⁹ Ultrafast laser ablation sample introduction systems aim to reduce elemental fractionation, improve signal to noise, remove

memory effects and achieve higher transport efficiency with sub-100 ms washout times.⁴⁰ As well as the DCI, similar systems first described by Wang *et al.*⁴¹ and van Malderen *et al.*⁴² have become commercially available. Applications for these ultrafast laser ablation cells have primarily focussed on elemental analyses and their potential utility for isotope ratio analysis by MC-ICP-MS has not yet been fully investigated.³⁴

Experimental

An ESI® (New Wave Research™) UP193FX 193 nm excimer laser system was coupled to a Thermo Scientific™ Neptune Plus™ MC-ICP-MS. The MC-ICP-MS was equipped with a multiple ion counting (MIC) array to simultaneously measure ^{234}U , ^{235}U and ^{236}U . ^{238}U was collected on a Faraday cup (Table 1) coupled to a $10^{11} \Omega$ amplifier.

Two different laser ablation configurations were used to transport the laser plume from the ablation site to the plasma. The first, more established configuration used a low-volume single-volume cell, the Zircon Cell, developed by Horstwood *et al.*⁴³ which followed the work of Bleiner and Günther⁴⁴ by restricting the internal volume to a flat-sided teardrop shape of *ca.* 3 cm³. The helium ablation gas exited the cell along 1/8" ID Teflon-lined Tygon tubing. Ar make-up gas and N₂ sourced from the outlet of an Aridus™ desolvating nebuliser system (CETAC Technologies, Omaha, Nebraska) was mixed with the ablation gas *via* a Y-piece halfway between the cell and the injector. The total length of tubing used was 1.5 m and as a result the total internal volume of the configuration was *ca.* 15 cm³.

The second configuration, incorporating a high speed, integrated ablation cell and dual concentric injector (DCI) developed at Loughborough University⁴⁵ (Fig. 1) was first described in Douglas *et al.*³⁸ It consisted of three main components, an inner cell, an outer cell and the DCI. The inner cell, the micro “Sniffer” cell, sat about 100 μm above the sample surface within the larger holding “Enterprise” outer cell. The inner cell had to remain stationary relative to the laser head and therefore the sample moves within the outer cell, and under the floating inner cell, by magnetic coupling. From the inner cell, a 250 μm i.d. thin fused silica tube transported the ablated material to the injector. In the DCI the fused silica line was passed down the centre of a 2.0 mm quartz injector, hence acting as a secondary internal injector. Around the fused silica line was added the Ar make-up gas and N₂, again sourced from an Aridus, creating a “sheath” around the He ablation gas. The DCI system was intended to work in a close-coupled configuration in which the fused silica tubing was kept straight and did not exceed 30 cm in length, restricting the total volume to 0.015 cm³. Due to the size of the MC-ICP-MS such a close coupling

Table 1 Cup configuration on the Thermo Scientific™ Neptune Plus™ MC-ICP-MS. ^{234}U was collected on a CDD rather than a full-size SEM

Cup	IC 5	IC 3	IC 2	IC 1	L4	C
Detector	CDD	SEM	SEM	SEM	FAR ($10^{11} \Omega$)	FAR ($10^{11} \Omega$)
Mass	^{234}U	^{235}U	^{236}U	237.05	^{238}U	255.5

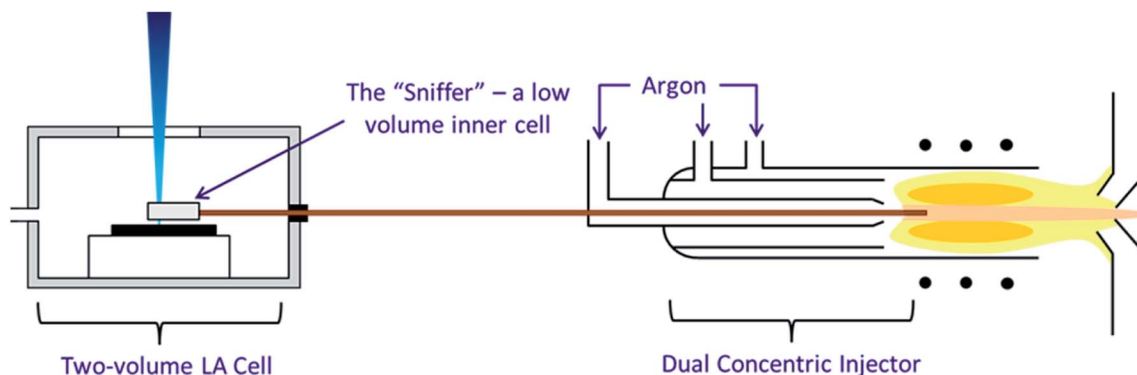


Fig. 1 The three components of the high speed, integrated ablation cell and DCI, reproduction from Douglas D. N., Managh A. J., Reid H. J., Sharp B. L., *Anal. Chem.*, 2015, **87**(22), 11285–11294.

was impossible and as a result in our configuration the fused silica tubing was both curved and increased in length to 150 cm, resulting in a total volume of 0.075 cm³.

Correction factors were determined from solutions of known uranium isotopic composition introduced *via* the Aridus desolvating nebulizer prior to LA analysis. Certified reference material (CRM) 112A (New Brunswick Laboratory, US

Department of Energy, Washington DC, USA)⁴⁶ with concentration <15 ppb in no more than 2% (m m⁻¹) HNO₃, was used to calculate the UH⁺/U⁺ and abundance sensitivity. CRM U010 (New Brunswick Laboratory, US Department of Energy, Washington DC, USA)⁴⁷ was used to determine correction factors for mass bias and relative detector gain.

Laser ablation of a single solid reference material, NIST SRM@611 (National Institute of Standards and Technology, Gaithersburg, Maryland, USA, 461.5 ± 1.1 ppm U)⁴⁸ was used to compare the different instrumental configurations.

The European Joint Research Center Geel (Belgium, formerly IRMM) have so far carried out eight Nuclear Signatures Inter-Laboratory Measurement Evaluation Programme (NUSIMEP) inter-laboratory comparisons (ILC). Two of these, NUSIMEP-6 and NUSIMEP-7, took the form of sub-micron uranium oxide particles of known composition coated onto carbon planchets^{25,27} and as such are useful reference materials to evaluate the ability of LA-ICP-MS to analyse such samples. The particles analysed in this study, composed of U₃O₈, were from both NUSIMEP-6 and NUSIMEP-7. NUSIMEP-6 consisted of a single graphite planchet with particles of a single isotopic composition condensed onto it. NUSIMEP-7 used two planchets, one containing particles of a single isotopic composition and another containing particles of two different compositions, the 'double composition' planchet. Reference uranium isotope ratios for the particles on each planchet are given in Table 3.

Due to the dynamic range required to detect and appropriately measure all uranium isotopes, a mixed detector array of Faraday cups and ion counters was required. Across the acquisition period of a transient signal (generated by either laser ablation or gas chromatography) using MC-ICP-MS, systematic

Table 2 Operating parameters for LA-MC-ICP-MS, both laser ablation configurations

	Zircon Cell	DCI
ESI® New Wave Research™ UP-193FX excimer laser		
Ablation mode	Single spot	Single spot
Fluence	11 J cm ⁻²	11 J cm ⁻²
Repetition rate	1 Hz	1 Hz
Spot size	5 μm (UO _x)	5 μm (UO _x)
	20 μm (SRM611)	20 μm (SRM611)
He carrier gas	0.8 L min ⁻¹	9 mL min ⁻¹
Cell pressure	≈ 0 kPa	77 kPa
CETAC Aridus™ I		
Nebulizer	PFA 50	PFA 50
Sweep gas	3.5 L min ⁻¹	5.3 L min ⁻¹
N ₂ add gas	9 mL min ⁻¹	9 mL min ⁻¹
Thermo Scientific™ Neptune Plus™ MC-ICP-MS		
RF power	1170 W	1170 W
Cool gas	15 L min ⁻¹	15 L min ⁻¹
Aux. gas	0.9 L min ⁻¹	0.9 L min ⁻¹
Skimmer cone	X	X
Sampler cone	Jet	Jet
Sample gas	0.82 L min ⁻¹	0.82 L min ⁻¹

Table 3 Reference uranium isotope ratios of NIST SRM 611 and NUSIMEP particles. Expanded uncertainty, $k = 2$, in brackets

	²³⁴ U/ ²³⁸ U	²³⁵ U/ ²³⁸ U	²³⁶ U/ ²³⁸ U
SRM611 (ref. 54)	9.45(05) × 10 ⁻⁶	2.3855(47) × 10 ⁻³	4.314(04) × 10 ⁻⁵
NUSIMEP-6 (ref. 25)	4.9817(48) × 10 ⁻⁵	7.0439(35) × 10 ⁻³	5.2048(86) × 10 ⁻⁷
NUSIMEP-7 (1) ²⁶	7.4365(60) × 10 ⁻⁵	9.0726(45) × 10 ⁻³	8.0205(71) × 10 ⁻⁶
NUSIMEP-7 (2) ²⁶	7.4365(60) × 10 ⁻⁵	9.0726(45) × 10 ⁻³	8.0205(71) × 10 ⁻⁶
	3.4514(24) × 10 ⁻⁴	3.4148(17) × 10 ⁻²	1.03268(70) × 10 ⁻⁴

isotope ratio drift has been observed.^{36,37,49,50} The cause of the isotope ratio drift in such cases has been identified as different tau response rates on the associated Faraday detector amplifiers,⁴⁹ and approaches to correct for the different amplifier tau response rates have been developed.^{31,50–53} For our analysis the difference in response rate was expected to be more pronounced; without a high resistance amplification circuit, the ion counter response is near instantaneous (Table 2).

We adopted the approach of Cottle *et al.*³⁰ who measured a response delay of *ca.* 0.2 s between ion counter and Faraday cup signals during single shot LA-MC-ICP-MS. This response delay strongly affected the accuracy of the isotope ratio when calculated on a time-slice (cycle-by-cycle) basis. As a consequence, the integrated area for each individual isotope was used to calculate the ratios for each pulse; that is total signal integration (TSI).

Results

Uranium isotope ratio measurement of SRM 611

Uranium isotope ratios of SRM 611 were calculated for each of 300 single shot ablations. For the ion counters total peak width averaged 510 ms for the Zircon Cell and 110 ms for the Enterprise Cell and DCI (Fig. 2). The reduction in peak width using

the DCI increased the signal-to-noise ratio by a factor of 25 and improved the definition of the minor (²³⁴U and ²³⁶U) isotopes. However, due to the slow Faraday amplifier response an average peak width significantly greater than 110 ms was measured for ²³⁸U (Fig. 4). To calculate an isotope ratio from such divergent peak profiles proved the necessity of the TSI data evaluation strategy. The three uranium isotope ratios, ²³⁴U/²³⁸U, ²³⁵U/²³⁸U and ²³⁶U/²³⁸U, were determined for the 300 ablation shots collected with both laser ablation systems (Table 4).

For the Zircon Cell the three uranium isotope ratios were accurate, within uncertainty, to the reference values and the precision closely tracked the values predicted from counting statistics. The increased detection efficiency and signal-to-noise ratio of the DCI and Enterprise Cell did not however, translate into a comparative improvement in precision. The ²³⁵U/²³⁸U SD of 2.9–3.0% was three times larger than counting statistics (0.99%). Plots of ²³⁵U/²³⁸U for each pulse (Fig. 3) confirmed the wider distribution of ratios with the DCI, with outliers biased towards a more depleted ²³⁵U composition. Given the broadly similar count rates with both laser ablation systems, results suggested the introduction of the DCI had introduced a new source of imprecision. A possible source of the additional uncertainty ('blind time') was investigated.

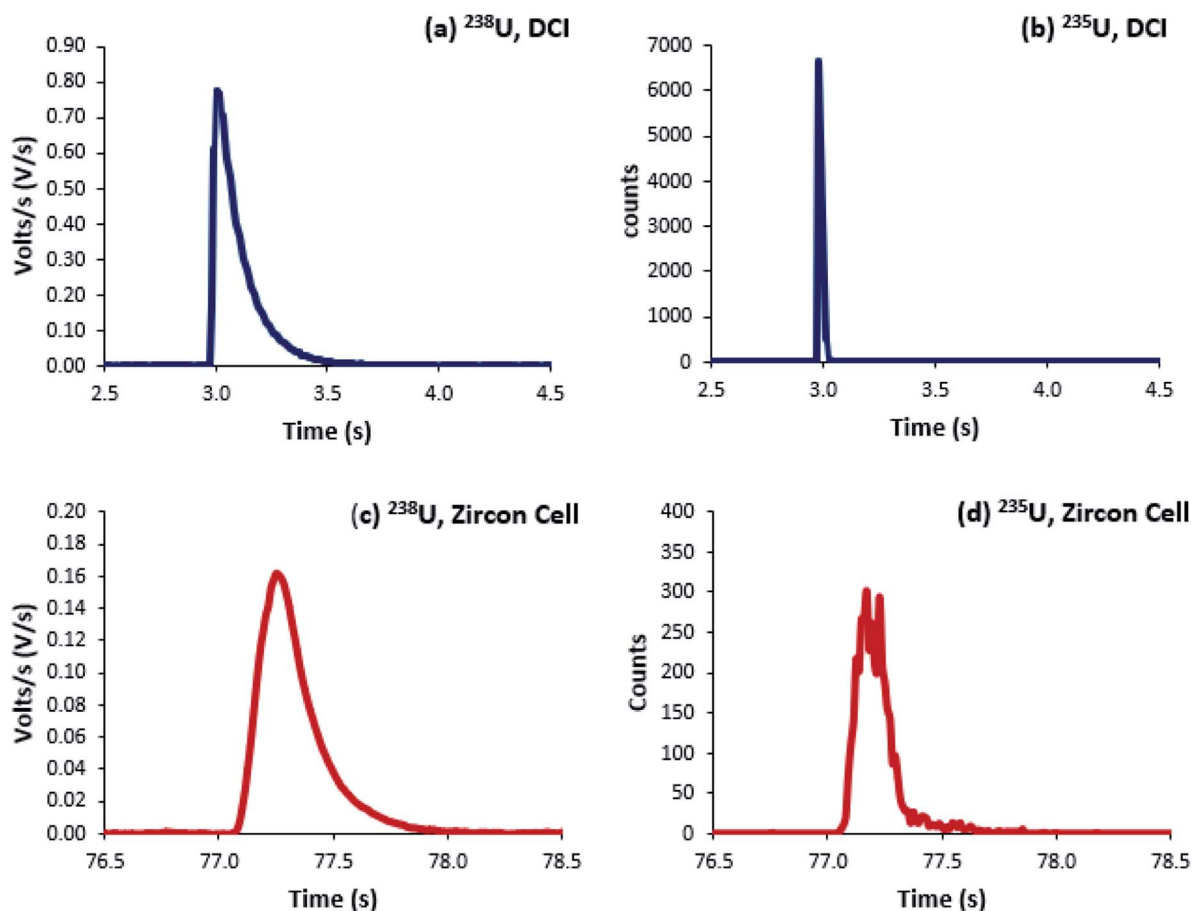


Fig. 2 Four single-shot laser ablation profiles of uranium isotopes in SRM 611, integration time 8 ms. (a and b) are ablations from the high speed, integrated cell and DCI; (c and d) the Zircon Cell. (a and c) were collected on a Faraday detector; (b and d) on an ion counter.

Table 4 Mean uranium isotope ratios of 300 single shots on SRM611 for both laser ablation configurations. Integration time = 66 ms

	Zircon Cell			DCI		
	$^{234}\text{U}/^{238}\text{U}$	$^{235}\text{U}/^{238}\text{U}$	$^{236}\text{U}/^{238}\text{U}$	$^{234}\text{U}/^{238}\text{U}$	$^{235}\text{U}/^{238}\text{U}$	$^{236}\text{U}/^{238}\text{U}$
Mean	1.04×10^{-5}	2.35×10^{-3}	4.55×10^{-5}	1.05×10^{-5}	2.34×10^{-3}	4.56×10^{-5}
–SD (%) ^a	15.7	1.33	7.38	13.2	2.89	7.62
+SD (%) ^a	18.6	1.35	7.96	15.2	2.97	8.24
Relative difference (%) ^b	9.64	–1.50	5.29	10.4	–1.93	5.53
RSD _{Poisson} , counting statistics (%)	16.8	1.11	8.18	15.1	0.99	7.28

^a Geometric mean: non-symmetric log-normal distribution, confidence intervals either side of the mean are not identical. ^b Relative difference = (geometric mean/reference value) – 1.

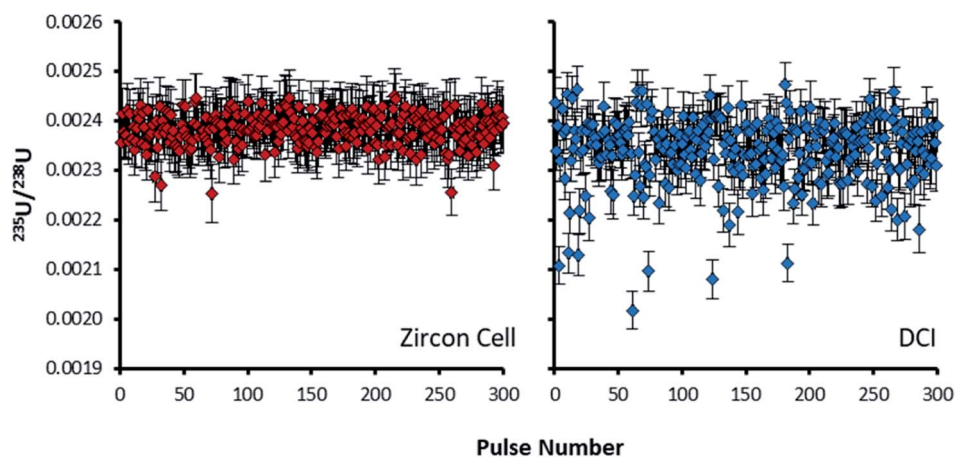


Fig. 3 $^{235}\text{U}/^{238}\text{U}$ isotope ratios of 300 pulses on SRM611 glass, calculated by total signal integration. Plot on the left (red) is for the Zircon Cell, plot on the right (blue) the DCI. Error bars on each ratio are 2SE, determined from counting statistics. All of the pulses collected with the Zircon Cell describe a single population within 2SD. Some pulses collected with the DCI had depleted $^{235}\text{U}/^{238}\text{U}$ values, out with uncertainty, relative to the main population.

'Blind time' – undetected ions between integrations

In the Neptune MC-ICP-MS software there is a discrepancy between how different integration times are identified, e.g. '66 ms', and the integration time of the data output, e.g. '68–69 ms'. For the example 66 ms integration time, the output of each integration is divided into three sections; a main 59 ms section; a secondary 7 ms section and the final 2 or 3 ms (average 2.37 ms over 10 000 integrations) residual time. The three sections are a legacy of the data acquisition system for the Faraday cups, which was developed with the collection of continuous signals in mind.³⁷ During operation the Faraday cups generate an analogue current. The analogue current is subsequently digitized using voltage to frequency conversion (by an analogue-to-digital converter). In this application rather than encoding the size of the signal, the change in the signal is encoded instead (removing any quantisation error). The current from the Faraday is therefore converted into a stream of digital pulses. For volt level signals on the Faraday the pulses occur at a frequency in the kHz range. If measured within a fixed time window in the kHz (ms) range the next pulse could occur either inside or outside of the window, limiting the precision to $\approx 1\%$. To achieve precision levels in the ppm range, requires the timing to a subsequent pulse to be measured against a fast,

MHz, clock. It is this fast clock measurement which occurs in the secondary section (7 ms) of the integration. At the end of the integration the system resets to the slower, kHz, clock. Once reset, the detector system waits until the next digital pulse is detected before beginning the next integration. It is the delay in waiting for the next pulse on each channel which is responsible for the extra time, known here as "blind time".⁵⁵ During blind time the mass spectrometer is effectively dead; no signal is recorded, on either the Faraday cups or ion counters. With the 66 ms integration time, only 96.4% of a continuous signal is

Table 5 Calculation of percentage of time lost due to blind time per integration. All values are an average of 2000 cycles. % lost = blind time/total time \times 100

Selected integration time (ms)	Measured total time (ms)	Blind time (ms)	% lost
8	11.50	3.50	30.4%
66	68.47	2.47	3.61%
131	134.3	3.28	2.44%
262	265.3	3.31	1.25%
524	527.8	3.82	0.72%
1049	1051	2.44	0.23%

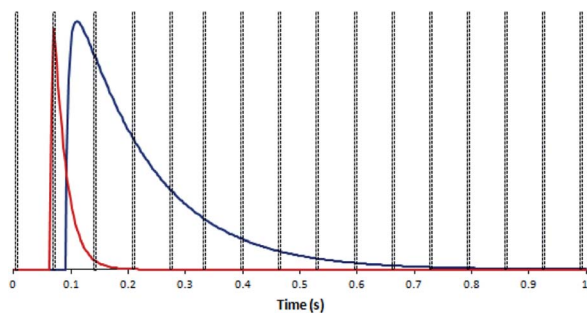


Fig. 4 Model ^{235}U IC signal response (red) and ^{238}U Faraday detector signal response (blue) for the DCI: created with the determined exponential Gaussian hybrid functions, values every 3 ms. Blind time events (black dotted box) denote an example 66 ms integration time. The proportion of total counts lost to blind time differs between the IC and Faraday signal responses.

being detected (Table 5). Interestingly, this indicates that due to the tau decay constant being longer than the integration time and therefore, encompassing multiple blind time events, 100% of the signal is never detected using Faraday detection and maximum precision is degraded using shorter integration times where proportionately more of the signal is lost during the 'blind time'.

The 8 ms integration time apart, it can be assumed that such short interruptions in data collection will have a negligible

effect on either continuous or semi-continuous signal responses. However, for transient signals collected on a mixed detector array, 'blind time' may have a significant impact on isotope ratio analysis.

In order to test the effect of blind time on isotope ratio measurement with the DCI, a model was created. 30 single shot laser ablation pulses on SRM 611, collected with both laser ablation configurations, were merged to produce an 'average' single shot ablation profile for each type of detector. To these average response profiles exponential modified Gaussian functions were fitted using the software package Igor Pro™ (version 6.36, Wavemetrics, Oregon, USA) and the Multipeak Fitting function.³⁸ The parameters generated were used in conjunction with the exponential-Gaussian hybrid function⁵⁶ to produce four model laser ablation pulses, one for each combination of laser ablation configuration and detector type (Fig. 4). Now that model curves had been constructed based on real data, some modelling calculations could now be carried out to demonstrate the effect of a 'blind time' component on the total amount of signal recorded within each peak area using different integration times. The 'blind time' used in the model curves was 3 ms, a value close to the average measured blind time of the detector system. By removing 3 ms integrations every 66 ms for each of the possible full integer start times (65 in this case), the range of signal which could potentially be lost on each detector was calculated for each integration period option in the Neptune software.

Table 6 Estimated percentage range of total signal which could be lost due to blind time on each detector, integration time = 66 ms

	Faraday – ^{238}U			SEM – ^{235}U			$^{235}\text{U}/^{238}\text{U}$
	Min signal lost (%)	Max signal lost (%)	Range (%)	Min signal lost (%)	Max signal lost (%)	Range (%)	Estimated RSD (%)
Zircon Cell	4.21	4.44	0.23	4.13	4.55	0.42	0.15
DCI	3.50	5.00	1.50	0.67	11.20	10.53	2.76

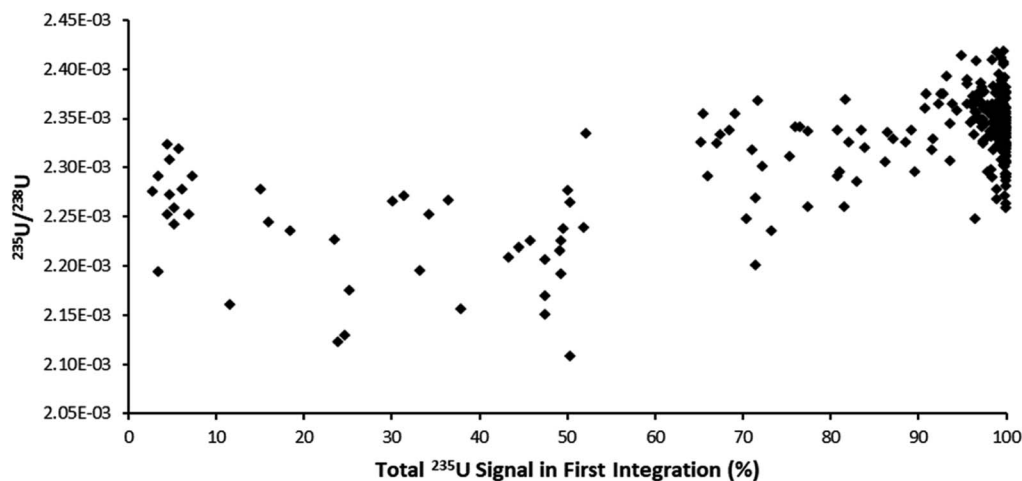


Fig. 5 Plot of $^{235}\text{U}/^{238}\text{U}$ for single-shot laser ablation pulses on SRM611, integration time 262 ms, against the percentage of ^{235}U signal contained within the first integration. Pulses with 100% of the ^{235}U signal in a single integration were biased higher relative to pulses split between two integrations.

In the plot of a model DCI pulse (Fig. 4), the signal collected on the IC is interrupted by two blind time events, in contrast to the 9 blind time events which interrupt the output of the Faraday signal. If the proportion of signal lost is different between the two detectors this would introduce a bias to the determined isotope ratio.

The ultrafast single shot profiles of the DCI more strongly affected the percentage of total signal which could be lost to blind time on the ion counters compared to the Faraday detector. Using the model, it was estimated, with 66 ms integration (Table 6), blind time could be responsible for at most 0.15% RSD $^{235}\text{U}/^{238}\text{U}$ on the mean single shot ablation pulses collected with the Zircon Cell, but up to 2.76% RSD with the DCI. The model supported the hypothesis that blind time was responsible for the additional $^{235}\text{U}/^{238}\text{U}$ uncertainty with the DCI. To further bolster the blind time hypothesis more single shot ablation pulses were collected on NIST 611, this time using a 262 ms integration time instead of 66 ms. As the average peak width on the SEM was only 110 ms, the ^{235}U signal collected could either be contained within a single integration or split between two integrations with blind time in between. The model suggested for a 262 ms integration time ^{238}U , collected on a Faraday detector, would lose between 0.34 to 2.16% of signal due to blind time events. ^{235}U would either lose no counts, resulting in a slightly enriched $^{235}\text{U}/^{238}\text{U}$ ratio, or lose up to 10.7% and give a depleted $^{235}\text{U}/^{238}\text{U}$ ratio. For our experimental particle data, the $^{235}\text{U}/^{238}\text{U}$ ratio was plotted against the percentage of the total ^{235}U signal captured in the first integration (Fig. 5). According to our model, particle data with approximately 50% of the total signal in the first integration would be most likely to exhibit blind time effects. This is indeed what Fig. 5 appears to show. In the plot most pulses with 100% of the ^{235}U signal in a single integration also have a higher

$^{235}\text{U}/^{238}\text{U}$ ratio, than those pulses where the ^{235}U signal was interrupted by blind time, consistent with the theoretical model.

Strategies to reduce 'blind time' for ultrafast transient signal analysis

Increasing the integration time from 66 ms to 262 ms reduced the proportion of pulses with significantly depleted $^{235}\text{U}/^{238}\text{U}$ ratios. By reducing the number of blind time events the likelihood of blind time occurring at the maxima of ^{235}U counts was also reduced. However, increasing the integration time did not eliminate blind time completely and furthermore opened up the possibility of enriched $^{235}\text{U}/^{238}\text{U}$ ratios by capturing the entire SEM output within a single integration. For particle analysis, where a single particle could represent the entire population of one composition, any bias due to blind time could seriously bias data interpretation.

The existence of blind time in some models of MC-ICP-MS may prevent the successful analysis of ultrafast transient signals. Until such time as next-generation instrumentation is developed, it is important to understand the minimum transient signal duration required in order for every ratio to be unaffected by blind time in LA-MC-ICP-MS set-ups. To estimate this value the parameters of the blind time model were modified to create results for pulses of approximately 50, 100, 200 and 300 ms duration. Each increase in pulse duration reduced the impact of blind time on the isotope ratio precision (Fig. 6). From this figure we can conclude that reducing the transient signal duration much below the 0.5 s achieved by the Zircon Cell will lead to blind time effects on ultrafast transient isotope ratio analyses. As such the reported washout times of most commercially available two-volume laser ablation cells (≤ 1 s)³⁷

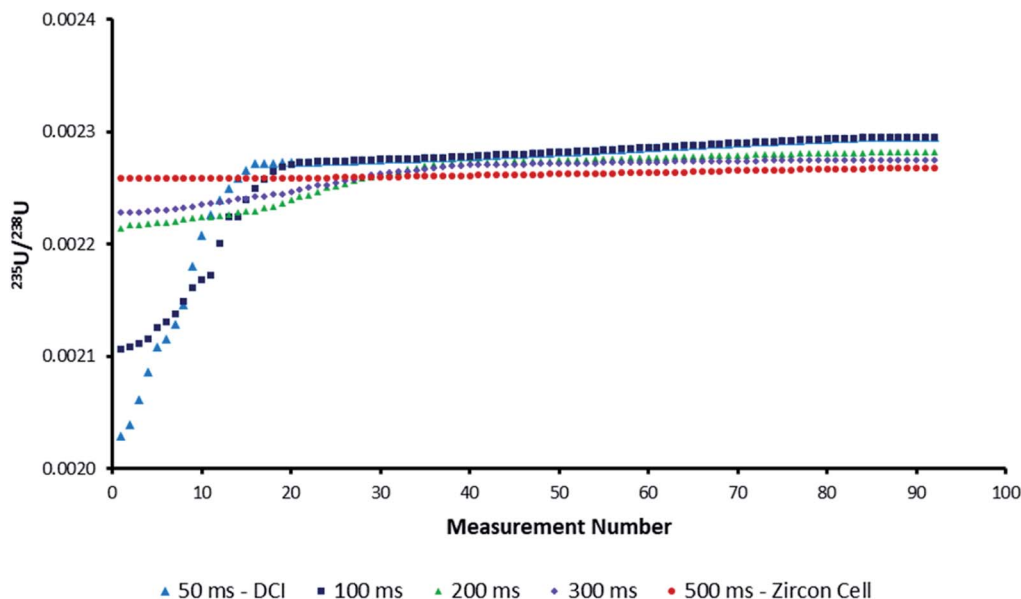


Fig. 6 Plot of possible uncorrected $^{235}\text{U}/^{238}\text{U}$ isotope ratios for the average model pulse due to the influence of blind time. The 100–300 ms datasets were created by widening the model IC pulse *via* changes to the parameters in the exponential-Gaussian hybrid function. As the pulse duration increased, the impact of blind time on isotope ratio accuracy was reduced.

may represent the best compromise for particle analysis with current MC-ICP-MS systems.

Effect of blind time on isotope ratio measurement of sub-micron uranium particles

NUSIMEP-6. A large number (Zircon Cell = 300 shots, DCI = 500 shots) of single-shot laser ablation pulses, spot size 5 μm , were conducted on the NUSIMEP-6 planchet. The 5 μm spot size was much less than the average spacing between particles determined by SEM-EDX, reducing the risk of simultaneously ablating more than one particle. However, the NUSIMEP-6 planchet has been described elsewhere as possessing a “smear” of uranium material across its surface,²⁷ alongside discrete particles. Here almost every laser shot resulted in some uranium signal. The mean ^{238}U counts for each single-shot ablation of the NUSIMEP-6 planchet was double that for SRM 611. From the density of U_2O_3 and the concentration of uranium in SRM 611, it was calculated that the mean SRM 611 signal would be equivalent to a ~ 150 nm diameter U_2O_3 particle. Due to the extremely low abundance of ^{236}U in the NUSIMEP-6 particles the average ^{236}U counts detected was less than 15 counts. Consequently, the $^{236}\text{U}/^{238}\text{U}$ isotope ratio values are not reported.

From the $^{235}\text{U}/^{238}\text{U}$ ratios obtained it was observed that particles, regardless of laser ablation configuration, seemed to be affected by blind time effects. This had been expected for the ultrafast DCI, but not the Zircon Cell. Those particles affected with the Zircon Cell were those single-shot ablation peak profiles which were interrupted with spikes in signal, of single integration duration (Fig. 7). The occurrence of such spikes was very strongly correlated with bias towards depleted uranium isotope ratios. Each spike was thought to represent the entrainment of a whole particle, or large fragment, into the plasma. Unlike the rest of the signal profile, these highly transient spikes would be susceptible to blind time effects.

To filter out the biased signals the application Flexmix, a software package in R, described by Kappel *et al.*²² was used to assign strongly biased signals to a separate cluster. The particles isolated in the separate cluster were not used to determine the average uranium isotope ratios (Table 7).

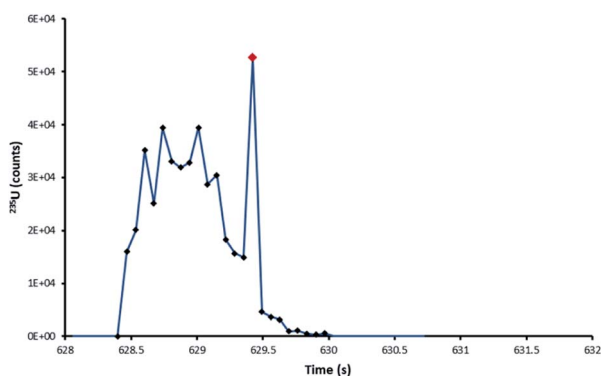


Fig. 7 ^{235}U pulse profile of the ablation of a NUSIMEP-6 particle, Zircon Cell. The standard profile is interrupted by a large single integration spike in the signal.

Table 7 Mean isotope ratio results for NUSIMEP-6. Zircon Cell, $n = 253$. DCI and Enterprise Cell, $n = 480$. Integration time = 66 ms

	Zircon Cell		DCI	
	$^{234}\text{U}/^{238}\text{U}$	$^{235}\text{U}/^{238}\text{U}$	$^{234}\text{U}/^{238}\text{U}$	$^{235}\text{U}/^{238}\text{U}$
Mean	4.92×10^{-5}	6.76×10^{-3}	4.76×10^{-5}	6.80×10^{-3}
–RSD (%)	7.66	2.93	9.54	2.79
+RSD (%)	8.29	3.01	10.6	2.87
RD (%)	–1.21	–4.05	–4.49	–2.84

NUSIMEP-7 (single composition). As expected the single composition NUSIMEP-7 planchet did not return a uranium signal with every ablation pulse. Out of 1000 shots at 9 different locations across the planchet only 277 (27.7%) detected uranium material. Another 500 shots focused on the centre of the planchet had a higher success rate of 82%. The Flexmix package was used to separate outliers such as those shown in Fig. 7. The theoretical RSE determined from counting statistics were 21.8% and 61.4% for $^{234}\text{U}/^{238}\text{U}$ and $^{236}\text{U}/^{238}\text{U}$ respectively. The high uncertainties (Table 8) reported for the minor isotope ratios, $^{234}\text{U}/^{238}\text{U}$ and $^{236}\text{U}/^{238}\text{U}$, were a consequence of the low number of detected ions. The precision on the $^{235}\text{U}/^{238}\text{U}$ ratio was not count rate limited (2.2% 1SD); the additional uncertainty determined for both ablation cells was attributable to blind time.

NUSIMEP-7 (double composition). The dual composition NUSIMEP-7 planchet was analysed with the same conditions as the single composition planchet. Particles of both expected compositions were detected in roughly equal quantities. A significant proportion of the measured ablation pulses had uranium isotopic compositions of neither expected composition, but instead fitted onto a ‘mixing line’ between the two expected compositions. The Flexmix software package was again used to group the particles into clusters, with two of the clusters representing each expected composition (Table 9).

Discussion

Sub-micron and micron-sized uranium particles are a challenging sample material for LA-MC-ICP-MS. A possible way forward for $^{235}\text{U}/^{238}\text{U}$ ratio measurements is measuring both isotopes on Faraday detectors.^{22,24,28} In doing so three of the limitations imposed by the ion counter are removed: the automatic protection system which deflects away ion beams greater than 2000 cpms, the limited dynamic range and now, significant blind time effects which are minor on Faraday detector systems due to their long and largely equivalent tau decay times. In our study the data from many of the largest, most valuable, particles had to be rejected due to at least one of these effects. Although the automatic protection could potentially be altered, the limited dynamic range cannot, and it would seem for LA-MC-ICP-MS analysis of uranium particles both ^{235}U and ^{238}U should be measured on Faraday detectors. However, for small or depleted uranium particles, the resulting small ion beam would be close to the Johnson noise of the $10^{11} \Omega$ amplifier. A

Table 8 Mean isotope ratio results for NUSIMEP-7 (single composition). Zircon Cell, $n = 349$. DCI and Enterprise Cell, $n = 202$. Integration time = 66 ms

	Zircon Cell			DCI		
	$^{234}\text{U}/^{238}\text{U}$	$^{235}\text{U}/^{238}\text{U}$	$^{236}\text{U}/^{238}\text{U}$	$^{234}\text{U}/^{238}\text{U}$	$^{235}\text{U}/^{238}\text{U}$	$^{236}\text{U}/^{238}\text{U}$
Mean	7.97×10^{-5}	8.97×10^{-3}	1.18×10^{-5}	7.78×10^{-5}	8.86×10^{-3}	9.59×10^{-6}
–RSD (%)	21.5	3.63	44.0	21.6	4.77	43.8
+RSD (%)	27.4	3.76	78.6	27.6	5.01	78.0
RD (%)	6.91	–1.19	38.3	4.47	–2.36	17.8

Table 9 Mean isotope ratio results for NUSIMEP-7 (double composition). Integration time = 66 ms

1 st composition	Zircon Cell				DCI			
	$n = 73$	$^{234}\text{U}/^{238}\text{U}$	$^{235}\text{U}/^{238}\text{U}$	$^{236}\text{U}/^{238}\text{U}$	$n = 104$	$^{234}\text{U}/^{238}\text{U}$	$^{235}\text{U}/^{238}\text{U}$	$^{236}\text{U}/^{238}\text{U}$
Mean		7.43×10^{-5}	8.72×10^{-3}	1.12×10^{-5}		7.75×10^{-5}	9.05×10^{-3}	1.40×10^{-5}
–RSD (%)		8.49	3.27	30.0		12.8	6.80	35.6
+RSD (%)		9.27	3.38	42.1		14.7	7.30	55.3
RD (%)		–0.04	–3.97	33.10		4.14	–0.23	55.83

2 nd composition	Zircon Cell				DCI			
	$n = 119$	$^{234}\text{U}/^{238}\text{U}$	$^{235}\text{U}/^{238}\text{U}$	$^{236}\text{U}/^{238}\text{U}$	$n = 70$	$^{234}\text{U}/^{238}\text{U}$	$^{235}\text{U}/^{238}\text{U}$	$^{236}\text{U}/^{238}\text{U}$
Mean		3.35×10^{-4}	3.12×10^{-2}	1.10×10^{-4}		3.57×10^{-4}	3.10×10^{-2}	1.20×10^{-4}
–RSD (%)		16.7	6.76	26.9		22.4	7.73	37.9
+RSD (%)		20.1	7.25	36.9		28.9	8.37	60.9
RD (%)		–2.96	–8.99	6.02		3.25	–9.52	14.72

recent instrumental development available for MC-ICP-MS, high gain $10^{13} \Omega$ amplifiers, have been applied to the LA-MC-ICP-MS analysis of similar small ion beams,⁵³ and have been used for uranium isotope ratio analysis by TIMS.⁵⁸ Unless TSI data processing is adopted, using high gain amplifiers for LA-MC-ICP-MS of uranium particles does require correcting for the different amplifier tau.⁵⁹ Claverie *et al.*²⁸ used the time-shift method developed by Gourgiotis *et al.*⁵⁰ to correct for differences in detector response when analysing 1–3.5 μm uranium particles by LA-MC-ICP-MS. For the population of particles analysed RSD's were 2.1% $^{235}\text{U}/^{238}\text{U}$ (expected composition 7.248×10^{-3}) and 2.5% $^{234}\text{U}/^{238}\text{U}$ (expected composition 6.1×10^{-5}). Furthermore, by using the time-shift correction they were able to use a point-by-point data reduction strategy to assign an uncertainty to each individual particle.

In our study the NUSIMEP-6 population, with a similar isotopic composition, was determined to $\approx 3\%$ RSD $^{235}\text{U}/^{238}\text{U}$ and $\approx 8\%$ RSD $^{234}\text{U}/^{238}\text{U}$, for a particle size distribution of $0.64 \pm 0.43 \mu\text{m}$. An RSD of 1.3% was achieved with the Zircon Cell on SRM 611, for an amount of uranium material equivalent to a 150 nm uranium particle. The solid glass matrix of SRM 611 was significantly less likely to produce large discrete particulate and hence was less susceptible to blind time.

For mixed detector arrays the results obtained from the ultrafast DCI suggest the washout time which will give the best signal to noise ratio, without introducing compromising blind time effects, is *ca.* 0.5 s. As ^{234}U and ^{236}U will still need to be measured on ion counters, using the ultrafast DCI or other rapid

washout systems for LA-MC-ICP-MS of uranium particles cannot be recommended, unless either the precision and accuracy required is less than the uncertainty due to blind time, or blind time is removed in MC-ICP-MS instrumentation. Such an improvement would be welcome outside uranium particle analysis as currently the efficiency of any analysis with a small integration time is reduced by the multiple blind time events experienced. For an 8 ms integration time 30% of the introduced ions are currently undetected and ironically, the only time when 100% of a signal is detected is on an ion counter when the input signal pulse is shorter than the integration time. Even with the $>5\%$ detection efficiency of the ultrafast laser ablation configuration,³⁴ the average number of counts of ^{234}U and ^{236}U detected for each sub-micron particle was still too low to reach the precision required by the nuclear safeguards community. This also rules out the use of a pseudo-simultaneous LA-ICP-TOF-MS (where blind time would not be a factor) for particle analysis as the sensitivity of ICP-TOF-MS is orders of magnitude less than MC-ICP-MS.²³ For LA-MC-ICP-MS to be a viable tool for minor isotope ratio determination in uranium particle analysis, the particles analysed must either be larger, micron sized and above, or be enriched in the minor isotopes.

Conclusions

Despite significantly improving the signal to noise ratio of the sample introduction set-up for LA-MC-ICP-MS using an ultrafast washout laser ablation cell and torch design, expected

improvements in isotope ratio precision were not realized. The highly transient data resulting from the adoption of the ultrafast LA-MC-ICP-MS has introduced an additional source of bias and scatter, here termed 'blind time' which recognizes a fundamental time limit in some MC-ICP-MS systems where data are not recorded. For highly transient data significant components of time and therefore data can be missed, indicating that accurate determination of such analyses may not be achievable with some MC-ICP-MS instruments equipped with mixed detector arrays. Possible exceptions, such as the $^{234}\text{U}/^{238}\text{U}$ and $^{236}\text{U}/^{238}\text{U}$ compositions investigated in this study, are where counting statistic limitations dominate the uncertainty budget, masking the blind time effects on the isotope ratios. The improvement in signal to noise ratio provided by ultrafast washout laser ablation cell systems was of great benefit in resolving the minor uranium isotopes from the baseline and may yet have a role in future analytical methods if the observed biases and scatter due to blind time could be eliminated. For the Neptune Series MC-ICP-MS, modelling of blind time suggested a minimum signal duration of 500 ms is required to eliminate blind time effects when using a mixed detection array. We would therefore recommend that commercially available two-volume cells with washout times of 0.5–1 s are currently most appropriate to use in conjunction with MC-ICP-MS instruments for uranium particle analysis. The Zircon Cell (average pulse duration 0.5 s) measured an uncertainty of 1.3% 1RSD for $^{235}\text{U}/^{238}\text{U}$ for a population of single-shot ablations of SRM 611: the ablated material from each shot was equivalent to a 150 nm uranium particle.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This research was made possible through the Natural Environment Research Council-Royal Society of Chemistry Analytical Chemistry Trust Fund (NERC-ACTF), Analytical Science & Technology research programme, funding NE/I019786/1. Work at the NERC Isotope Geosciences Laboratory (NIGL), British Geological Survey was funded through IP-1315-0512. The authors would also like to thank the two anonymous reviewers for their helpful and constructive reviews.

References

- M. Betti, L. A. de las Heras and G. Tamborini, *Appl. Spectrosc. Rev.*, 2006, **41**, 491–514.
- M. May, R. Abedin-Zadeh, D. Barr and A. Carnesale, *Nuclear Forensics. Role, State of the Art, Program Needs*, 2008.
- K. Mayer, M. Wallenius and Z. Varga, *Chem. Rev.*, 2013, **113**, 884–900.
- R. R. Parrish, M. Horstwood, J. G. Arnason, S. Chenery, T. Brewer, N. S. Lloyd and D. O. Carpenter, *Sci. Total Environ.*, 2008, **390**, 58–68.
- N. S. Lloyd, S. R. N. Chenery and R. R. Parrish, *Sci. Total Environ.*, 2009, **408**, 397–407.
- Z. Varga, M. Wallenius and K. Mayer, *J. Anal. At. Spectrom.*, 2010, **25**, 1958–1962.
- F. E. Stanley, *J. Anal. At. Spectrom.*, 2012, **27**, 1821–1830.
- G. R. Eppich, R. W. Williams, A. M. Gaffney and K. C. Schorzman, *J. Anal. At. Spectrom.*, 2013, **28**, 666–674.
- E. Kuhn, D. Fischer and M. Ryjinski, *Environmental Sampling for IAEA Safeguards: A Five Year Review*, 2001.
- R. C. B. Pestana, J. E. S. Sarkis, R. C. Marin, C. H. Abreu-Junior and E. F. U. Carvalho, *J. Radioanal. Nucl. Chem.*, 2013, **298**, 621–625.
- T. M. Witte, PhD thesis, Iowa State, 2011.
- X. Z. Zhang, F. Esaka, K. T. Esaka, M. Magara, S. Sakurai, S. Usuda and K. Watanabe, *Spectrochim. Acta, Part B*, 2007, **62**, 1130–1134.
- T. Shinonaga, F. Esaka, M. Magara, D. Klose and D. Donohue, *Spectrochim. Acta, Part B*, 2008, **63**, 1324–1328.
- F. Esaka, M. Magara and T. Kimura, *J. Anal. At. Spectrom.*, 2013, **28**, 682–688.
- F. Esaka, M. Magara, C. G. Lee, S. Sakurai, S. Usuda and N. Shinohara, *Talanta*, 2009, **78**, 290–294.
- Z. Stefánka, R. Katona and Z. Varga, *J. Anal. At. Spectrom.*, 2008, **23**, 1030–1033.
- N. S. Lloyd, R. R. Parrish, M. Horstwood and S. R. N. Chenery, *J. Anal. At. Spectrom.*, 2009, **24**, 752–758.
- Z. Varga, *Anal. Chim. Acta*, 2008, **625**, 1–7.
- J. S. Becker, H. Sela, J. Dobrowolska, M. Zoriy and J. S. Becker, *Int. J. Mass Spectrom.*, 2008, **270**, 1–7.
- F. Pointurier, A. Pottin and A. Hubert, *Anal. Chem.*, 2011, **83**, 7841–7848.
- F. Pointurier, A. Hubert and A.-C. Pottin, *J. Radioanal. Nucl. Chem.*, 2012, **296**, 609–616.
- S. Kappel, S. F. Boulyga, L. Dorta, D. Günther, B. Hattendorf, D. Koffler, G. Laaha, F. Leisch and T. Prohaska, *Anal. Bioanal. Chem.*, 2013, **405**, 2943–2955.
- A.-L. Ronzani, F. Pointurier, M. Rittner, O. Borovinskaya, M. Tanner, A. Hubert, A.-C. Humbert, J. Aupiais and N. Dacheux, *J. Anal. At. Spectrom.*, 2018, **33**, 1892–1902.
- Z. Varga, M. Krachler, A. Nicholl, M. Ernstberger, T. Wiss, M. Wallenius and K. Mayer, *J. Anal. At. Spectrom.*, 2018, **33**, 1076–1080.
- Y. Aregbe, J. Truyens, R. Kips, S. Richter, E. Stefaniak, H. Kühn and M. Kraiem, *NUSIMEP-6: uranium isotope amount ratios in uranium particles*, 2008.
- J. Truyens, E. Stefaniak, S. Mialle and Y. Aregbe, *NUSIMEP-7: uranium isotope amount ratios in uranium particles*, 2011.
- J. Truyens, E. Stefaniak and Y. Aregbe, *J. Environ. Radioact.*, 2013, **125**, 50–55.
- F. Claverie, A. Hubert, S. Beraïl, A. Donard, F. Pointurier and C. Pécheyran, *Anal. Chem.*, 2016, **88**, 4375–4382.
- M. Tanner and D. Günther, *Anal. Chim. Acta*, 2009, **633**, 19–28.
- J. M. Cottle, M. S. A. Horstwood and R. R. Parrish, *J. Anal. At. Spectrom.*, 2009, **24**, 1355–1363.
- T. Pettke, F. Oberli, A. Audétat, U. Wiechert, C. R. Harris and C. a. Heinrich, *J. Anal. At. Spectrom.*, 2011, **26**, 475–492.

- 32 L. Allen, H. Pang and A. Warren, *J. Anal. At. Spectrom.*, 1995, **10**, 267–271.
- 33 M. Wiedenbeck, R. Bugoi, M. J. M. Duke, T. Dunai, J. Enzweiler, M. Horan, K. P. Jochum, K. Linge, J. Košler, S. Merchel, L. F. G. Morales, L. Nasdala, R. Stalder, P. Sylvester, U. Weis and A. Zoubir, *Geostand. Geoanal. Res.*, 2012, **36**, 337–398.
- 34 G. Craig, A. J. Managh, C. Stremtan, N. S. Lloyd and M. S. A. Horstwood, *Anal. Chem.*, 2018, **90**, 11564–11571.
- 35 A. Gourgiotis, G. Manhès, P. Louvat, J. Moureau and J. Gaillardet, *J. Anal. At. Spectrom.*, 2015, **30**, 1582–1589.
- 36 A. Gourgiotis, G. Manhès, P. Louvat, J. Moureau and J. Gaillardet, *Rapid Commun. Mass Spectrom.*, 2015, **29**, 1617–1622.
- 37 E. M. Krupp and O. F. X. Donard, *Int. J. Mass Spectrom.*, 2005, **242**, 233–242.
- 38 D. N. Douglas, A. J. Managh, H. J. Reid and B. L. Sharp, *Anal. Chem.*, 2015, **87**, 11285–11294.
- 39 W. Müller, M. Shelley, P. Miller and S. Broude, *J. Anal. At. Spectrom.*, 2009, **24**, 209–214.
- 40 S. J. M. van Malderen, A. J. Managh, B. L. Sharp and F. Vanhaecke, *J. Anal. At. Spectrom.*, 2016, **31**, 423–439.
- 41 H. A. O. Wang, D. Grolimund, C. Giesen, C. N. Borca, J. R. H. Shaw-Stewart, B. Bodenmiller and D. Günther, *Anal. Chem.*, 2013, **85**, 10107–10116.
- 42 S. J. M. Van Malderen, J. T. van Elteren and F. Vanhaecke, *J. Anal. At. Spectrom.*, 2015, **30**, 119–125.
- 43 M. S. A. Horstwood, G. L. Foster, R. R. Parrish, S. R. Noble and G. M. Nowell, *J. Anal. At. Spectrom.*, 2003, **18**, 837–846.
- 44 D. Bleiner and D. Günther, *J. Anal. At. Spectrom.*, 2001, **16**, 449–456.
- 45 D. N. Douglas, PhD thesis, Loughborough University, 2013.
- 46 J. Neuhoff, *Certificate of Analysis CRM 112-A Uranium (normal) Metal Assay and Isotopic Standard*, New Brunswick Laboratory, Argonne IL, 2010.
- 47 J. Neuhoff, *Certificate of Analysis CRM U010 Uranium Isotopic Standard (5 mg Uranium as U₃O₈)*, New Brunswick Laboratory, Argonne IL, 2008.
- 48 S. A. Wise and R. L. Watters, *National Institute of Standards & Technology Certificate of Analysis Standard Reference Material® 611 Trace Elements in Glass*, NIST, Gaithersburg MD, 2012.
- 49 M. Dzurko, D. Foucher and H. Hintelmann, *Anal. Bioanal. Chem.*, 2009, **393**, 345–355.
- 50 A. Gourgiotis, S. Bérail, P. Louvat, H. Isnard, J. Moureau, A. Nonell, G. Manhès, J.-L. Birck, J. Gaillardet, C. Pecheyran, F. Chartier and O. F. X. Donard, *J. Anal. At. Spectrom.*, 2014, **29**, 1607–1617.
- 51 T. Hirata, Y. Hayano and T. Ohno, *J. Anal. At. Spectrom.*, 2003, **18**, 1283–1288.
- 52 T. Iizuka and T. Hirata, *Chem. Geol.*, 2005, **220**, 121–137.
- 53 J.-I. Kimura, Q. Chang, N. Kanazawa, S. Sasaki and B. S. Vaglarov, *J. Anal. At. Spectrom.*, 2016, **31**, 790–800.
- 54 M. Zimmer, W. Kinman, A. Kara and R. Steiner, *Minerals*, 2014, **4**, 541–552.
- 55 A. J. Managh, D. N. Douglas, K. Makella Cowen, H. J. Reid and B. L. Sharp, *J. Anal. At. Spectrom.*, 2016, **31**, 1688–1692.
- 56 K. Lan and J. W. Jorgenson, *J. Chromatogr. A*, 2001, **915**, 1–13.
- 57 J. M. Cottle, A. R. Kylander-Clark and J. C. Vrijmoed, *Chem. Geol.*, 2012, **332–333**, 136–147.
- 58 A. Trinquier and P. Komander, *J. Radioanal. Nucl. Chem.*, 2016, **307**, 1927–1932.
- 59 G. Craig, Z. Hu, A. Zhang, N. S. Lloyd, C. Bouman and J. Schwieters, *Thermo Fish. Sci. Tech. Note 30396*, 2017.