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Brain grants permission of access to Zika virus but denies entry to drugs: a molecular modeling perspective to infiltrate the boundary†

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The magnetism of the Zika virus to neuronal cells proves to be one of the major concerns in the development of effective inhibitors. Although the blood–brain barrier limits the entry of most drugs, tailored small molecule inhibitors and drug delivery systems are currently being designed to overcome this obstacle. We have identified the core challenge to be addressed – blood–brain barrier permeability – and provided insight into strategies that can be used to improve drug delivery to the brain. We have compiled drugs that have previously been proposed as potential Zika virus inhibitors and classified chemical features of those drugs, which influence blood–brain barrier permeability. Thereafter, we created a route map to design drugs with improved blood–brain barrier permeability. An alternative approach using drug delivery systems to transport membrane-impermeable Zika virus inhibitors to the brain is also proposed, along with descriptions of known drug carriers. This review provides information for further research toward inhibitors of Zika virus.

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The tale of Zika virus

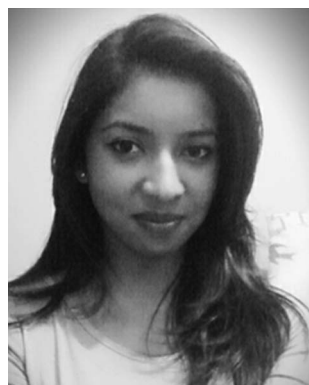
The Zika virus (ZIKV), a mosquito-borne virus, belongs to the Flaviviridae family and has similar characteristics to other flaviviruses such as Dengue virus, West Nile virus and Japanese Encephalitis virus.¹ The rapid disseminating potential and repercussion in humans are attributed to the various modes of

transmission, primarily through the bite of an infected *Aedes aegypti* mosquito.² The ZIKV is also transmitted through sexual intercourse,³ blood transfusions⁴ and from a mother to child perinatally.⁵

The ZIKV was originally isolated in Uganda in the Zika forest in 1947.⁶ For nearly 7 decades thenceforth, sporadic infections caused by ZIKV were reported in several countries worldwide. These include more equatorial countries of Africa such as Tanzania, Egypt, Kenya, Nigeria, Central African Republic, and Gabon; some Asian countries including Pakistan, India, Malaysia, Thailand, the Philippines and Indonesia; and many islands in the Pacific Ocean.^{7–10} The most devastating and

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highly publicized outbreak that captured the world's attention occurred in Brazil in 2015, which triggered global panic as it rapidly spread across America.¹¹ In 2016, the ZIKV broadened its geographic spectrum to North American Florida and Texas where the infection was locally transmitted.¹⁰

There has been prior ambiguity regarding the diagnosis of ZIKV, as its infection manifests similarly to common colds and other *flavivirus* infections.¹² These symptoms include fever, headaches, conjunctivitis, joint pain, muscle pain and skin rash.^{13,14} The speculation of ZIKV infection depends on its manifestation and history of mosquito bites.¹² The ZIKV is detectable in bodily fluids such as saliva,¹⁵ semen,^{16,17} urine¹⁸ and amniotic fluid,¹⁹ which can be verified in the laboratory.²⁰ The various modes of transmission of ZIKV make the human body highly susceptible to infection. When ZIKV enters the body through an infected female mosquito bite, the infection manifests as a rash in the vicinity of the bite.⁹ This occurs due to the release of virions into dermal and epidermal layers of the skin, where ZIKV is introduced to the bloodstream and advances to the lymph nodes to replicate and cause viremia.²¹

The ZIKV is an enveloped icosahedral virus that is made up of a single-stranded, positive-sense genome. The enveloped virion comprises of an 11 kilobase genome consisting of 10 794 nucleotides encoding 3419 amino acids.²² The open reading frame (ORF) of the 5' and 3' untranslated region (UTR) encodes a polyprotein that is cleaved into three structural proteins, being the capsid (C), precursor membrane (prM), and envelope (E).²³ Seven non-structural (NS) proteins are also found in this assembly, namely, NS1, NS2a, NS2b, NS3, NS4a, NS4b, and NS5 (largest viral protein).²⁴ The genomic protein organization is 5'-C-prM-E-NS1-NS2a-NS2b-NS3-NS4a-NS4b-NS5-3'²⁵ and contains

an m⁷gpppAmpN₂ at the 5' end and lacks a poly-A tail at the 3' end.²⁶ A highly conserved 90–120-nucleotide strand is situated close to the 3' end that develops into a hairpin loop and is vital for replication.^{26,27} Of the non-structural proteins, NS1, NS3 and NS5 are highly conserved whereas the NS2a, NS2b, NS4a and NS4b are small and hydrophobic.²⁴ Of critical importance is the proteolytic cleavage of prM to produce the pr and M protein by furin-like protease located in the *trans*-Golgi network during the egress of the particles as this promotes maturation of virions.²⁸

The ramifications of ZIKV infections have heavily impacted thousands worldwide, particularly in newborns, since ZIKV-infected pregnant women have given birth to babies with congenital brain abnormalities, predominantly microcephaly and intracranial calcification.¹³ The ZIKV infection has also been shown to elicit Guillain-Barré Syndrome, which ultimately advances to paralysis and death.²⁹

Studies have shown tropism of the ZIKV for cells of the nervous system, whereby entry into neuronal and endothelial cells occur *via* AXL receptors situated on the cell surface.^{30–32} The ZIKV has also affected retinal cells that line the blood-retinal barrier (*i.e.* retinal pigment epithelium and retinal endothelium) in mice, which also express AXL receptors. This presents as conjunctivitis.³³

Blood–brain barrier permeability as a core challenge in ZIKV therapy

Treating the symptoms of ZIKV will not yield permanent results; hence nipping the cause at the bud may be the best route to a solution. The blood–brain barrier (BBB) and placental barrier are surrounded by lipophilic membranes and junctions, through which only certain compounds can permeate.³⁴ It has been shown that the placenta is permissive to most drugs as it serves to allow for the exchange of nutrients for its biological purpose.³⁵ The ZIKV can penetrate these membranes, which is evident by its downstream pathogenic effects in fetal nervous systems.^{13,26,29,30}

The specific characteristics of a compound govern the method by which it is transported across the BBB, or whether or not it is transported at all. Compounds that have surface hydrogen bonds (hydrophilic compounds) are only permissive through tight junctions of the BBB, which ultimately serve to prevent the passage of molecules between cells of the endothelium. These hydrophilic compounds are impermeable through the lipophilic endothelium and require lipid-mediated transport in order to permeate transcellularly. Large molecules, such as insulin and transferrin, require receptors, whereas small molecules require carrier-mediated transport to move across the barrier. There is also active efflux transport for endogenous BBB transporters (Fig. 1 – BBB).^{34,36–38}

Bioavailability features of screened drugs as prospective ZIKV inhibitors

There are FDA-approved drugs that have been proposed as anti-ZIKV drugs based on their diverse antiviral/antimicrobial/antibacterial activities in diseases other than ZIKV,³⁹ however,



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Fig. 1 A schematic representation of the blood–brain barrier and pathways across this barrier.

the ability of those drugs to pass the BBB, their properties as hydrophilic/lipophilic compounds, and their ability to be absorbed by the gastrointestinal tract (GIT) have not yet been clearly elucidated. To this effect, we have taken a step further and utilized a chemical data base, PubChem, along with ADME prediction tools, Swiss ADME⁴⁰ to predict specific characteristics of these candidate anti-ZIKV drugs and to verify whether or not the proposed compounds by Barrows *et al.* (2016) could be potential anti-ZIKV drugs with BBB-permeable profiles. SWISS ADME is a “website allows you to compute physicochemical descriptors as well as to predict ADME parameters, pharmacokinetic properties, druglike nature and medicinal chemistry friendliness of one or multiple small molecules to support drug discovery”.⁴⁰ The proposed compounds are listed in Table 1.

The partition coefficient ($\log P$) measures how hydrophilic or hydrophobic a molecule is. The desired $\log P$ value of a molecule likely to permeate lipophilic membranes should lie between 0–5.⁴¹ The ability of a drug to pass through the BBB is influenced by their unique but varying properties.³⁶ In Table 1, ~80% of the drugs described have the ability to pass through lipophilic membranes, however, less than 20% of those drugs can penetrate the BBB.⁴⁰ Efficient GIT absorption of orally administered drugs is required for entry into the bloodstream and sufficient drug delivery,⁴² although, just half the drugs mentioned are highly absorbed *via* the GIT. The results of this

table suggest that the ability of a compound to pass through the BBB depends on factors additional to lipophilicity. From this exercise we highlight 5 compounds that are predicted to pass the BBB and the hydrophobic spots of each compound are shadowed in yellow in Fig. 2.

Hydrophobic groups of compounds are required for hydrophobic interactions with target molecules. Hydrophobic interactions are comparably stronger than some weak intermolecular forces, such as hydrogen bonds or van der Waals interactions, and ensure protein–ligand complexes remain stable and biologically active.⁴³ As depicted in Fig. 2, all five compounds from Table 1 that have the ability to pass the BBB bear hydrophobic groups and therefore, possess the potential to form hydrophobic interactions with target molecules. Although, fingomolid and sertraline have more hydrophobic spots than the other compounds in Fig. 2, and therefore, are more likely to form stronger hydrophobic interactions.

Systematic approach to tackle the challenge

To overcome the hurdle faced by most drug therapies, we are presenting two main strategies that could potentially assist with the design and bioavailability of compounds with an improved BBB permeability profile. Furthermore, we include an approach



Table 1 Predicted bioavailability features of prospective anti-ZIKV inhibitors

Drug name	Lipid solubility (log <i>P</i>)	BBB permeation	GIT absorption
Auranofin	0.00	No	High
Azathioprine	0.72	No	Low
Bortezomib	0.00	No	High
Clofazimine	4.72	No	Low
Cyclosporine A	6.16	No	Low
Dactinomycin	5.33	No	Low
Daptomycin	0.79	No	Low
Deferasirox	2.48	No	High
Digoxin	4.69	No	Low
Fingolimod	3.76	Yes	High
Gemcitabine HCl	0.00	No	High
Ivermectin	6.31	No	Low
Mebendazole	1.27	No	High
Mefloquine HCl	0.00	No	Low
Mercaptopurine hydrate	0.47	No	High
Methoxsalen	2.22	Yes	High
Micafungin	-0.72	No	Low
Mycophenolate mofetil	3.67	No	High
Mycophenolic acid	2.38	No	High
NITD008	1.30	No	Low
Palonosetron HCl	0.00	Yes	High
Pyrimethamine	2.15	Yes	High
Sertraline	3.40	Yes	High
Sofosbuvir	3.05	No	Low
Sorafenib tosylate	3.84	No	Low
Thioguanine	0.14	No	High

that relies on the pre-existing BBB-impermeable drugs conjugated to drug delivery systems. These strategies are: (1) improve the inhibitor and (2) carry the cargo.

Improve the inhibitor

In this approach, *in silico* tools may be used to model and optimize potential compounds (Fig. 3), followed by compound

synthesis and biological testing. Phase 1 includes targeted selection of potential anti-ZIKV compounds. This incorporates screening for potential compounds with specific physicochemical properties and antiviral activities using chemical databases, and the use of absorption, distribution, metabolism, excretion (ADME) prediction tools such as Swiss ADME, in order to filter out compounds that encompass the ability to pass the BBB. The ability of a compound to pass through the BBB is governed by a function of lipophilicity, the molecular characteristics of charged and hydrophobic residues of the compound as well as molecular weight.³⁷ The main lipophilic properties that must be considered include the Hansch constant (π), hydrophobic fragmental constant (*f*), log *P*, capacity factor values from RP-HPLC (log *k*_w), calculated log *P* values (CLOGP) and molecular lipophilic potential (MLP).⁴⁴ With regard to the charge of the compound, only uncharged molecules can diffuse across the membrane to become reprotonated once it leaves the membrane and enters the brain fluid.³⁷ As the size of a compound gets larger, its ability to permeate the BBB decreases.⁴⁵

Phase 2 involves the prediction of lipid permeability of the potential anti-ZIKV compounds using molecular dynamic simulations and 3D-modelled lipid bilayer simulations. Due to the surrounding lipid membrane in the BBB, it is necessary to assess compound interactions with the target enzyme within lipid membrane.⁴⁶

Phase 3 involves the estimation of binding affinities between potential compounds, which pass through the BBB, and viral enzymes. This may be achieved *via* binding free energy calculations or molecular docking of the compound of interest into the active site of the target enzyme.⁴⁷

Following the design process, the compounds must be synthesized for further testing. Synthesis of the compound involves construction of the carbon framework and the addition/deletion/transformation of functional groups for functionality of compound. Validation of synthesis is carried out by ligand-binding assays which involves placing the target

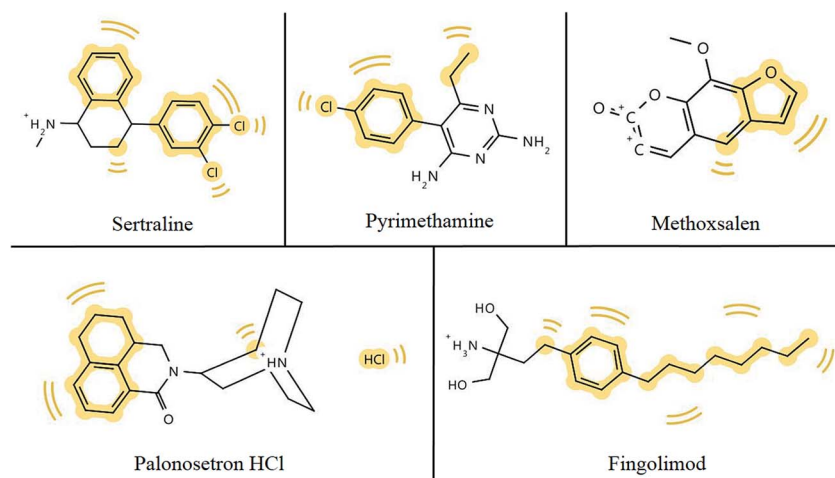


Fig. 2 Hydrophobic footprints (highlighted in yellow) in chemical structures of potential anti-ZIKV compounds which have the ability to permeate the BBB.



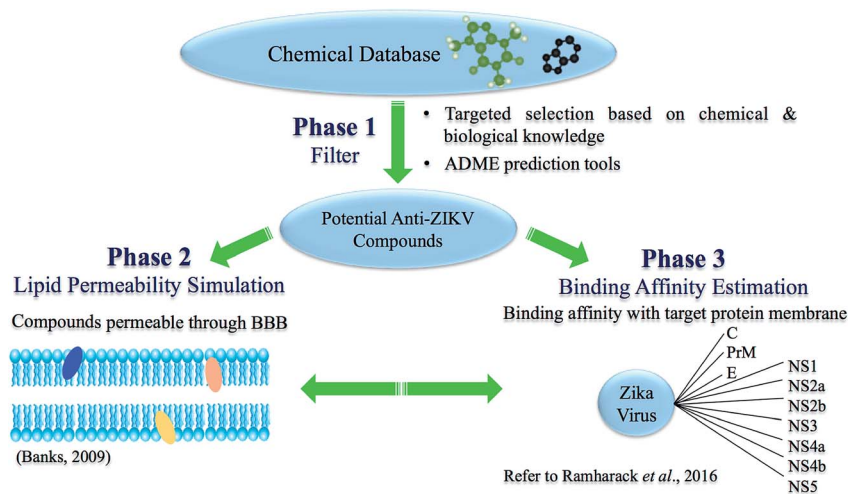


Fig. 3 Phases involved in the suggested approach to improve the BBB-permeability profile of the inhibitor.

enzyme and compound of interest into a solvent (*e.g.* water) to allow the compound to interact with the active site residues of the enzyme.⁴⁸ Binding studies provide reliable assessments of binding affinities, errors and binding mode.⁴⁸ Further studies including biological testing will investigate toxicity and efficacy of the compound, including *in vitro* studies (cellular level), *in vivo* studies (organism level) and ultimately clinical trials. Microscale thermophoresis (MST) may also be used in order to analyze the interaction between the inhibitors and receptors, experimentally, which is established on the controlled movement of particles along a temperature gradient.⁴⁹

Carry the cargo

An alternate to creating new BBB-permeable drugs will involve the utilization of drug carriers that have already been successful in delivering drugs to the brain. This approach eliminates the additional time and expense required to design and formulate new drugs and drug delivery systems. A drug carrier could be used to transport a BBB-impermeable drug to the brain and allow for the drug to carry out its function against ZIKV.

There are existing drugs that have been proven to inhibit ZIKV replication in isolated ZIKV infected cells, such as NITD008 (ref. 50) and sofosbuvir,⁵¹ however the drugs cannot pass the BBB to counteract the virus (Table 1). With half the battle won due to their ability to inhibit ZIKV, ultimate triumph over ZIKV would entail overcoming impermeability, which requires the potential ZIKV inhibitors to be transported across the BBB.

Previously utilized membrane-permeable drug delivery approaches have been successful in transporting membrane-impermeable drugs across the BBB for other infections and diseases, such as Parkinson's disease and Alzheimer's disease.^{52–55} Some known drug delivery systems used in various disease cases include polymers and polymeric nanoparticles such as micro/nanospheres, micro/nanocapsules, dendrimers, liposomes, hydrogels, gold nanoparticles, micelles; others include lipoproteins and aptamers (Table 2).^{54,56}

The inhibitor-carrier approach will involve techniques similar to the previous suggested approach. The first step would be to create an inhibitor-carrier complex by binding the known ZIKV inhibitor to the carrier. The complex must then be simulated to analyze the trajectory of the complex as well as the potential energy of the entire complex through a lipid bilayer. Once there is computational evidence to show that the inhibitor can theoretically bind to the carrier and move through a lipid membrane, the inhibitor-carrier complex must be synthesized to test it biologically (*e.g.* *in vivo* testing), which will be used to confirm drug delivery through the BBB to the brain. Examples of computationally docking a compound-polymer complex are provided in the ESI.†

Technical guidance

A number of tools are available which can be utilized to screen for compounds on chemical databases based on a set of criteria. Structure-based virtual screening will allow searching through combinatorial chemistry libraries for compounds that may be potential inhibitors of a target protein and will rapidly dock them into the 3D target's active pocket.^{47,78} Screening for potential compounds can be carried out on ZINC Database⁷⁹ or ZincPharmer.⁸⁰ Several molecules may have the potential to bind to the active site of the protein; therefore, the free binding energy of every pose is calculated. Binding affinity estimations may be carried out using molecular docking approaches and free binding energy calculations. This will generate a scoring function to rank the ligands that best suit the target protein.⁴⁷ Computational software that can be used to calculate binding affinities include UCSF Chimera⁸¹ and AutoDock Vina.⁸² Protein-ligand complexes of lowest free binding energy may be used as inhibitor candidates, which may subsequently be validated *via* molecular dynamic simulations, as binding affinity predictions may not be one hundred percent accurate.^{78,83–86}

Molecular dynamic simulations calculate the potential energy of a system and analyze free binding energies and binding modes between compounds and enzymes.⁸⁷ Force



Table 2 Pre-existing drug delivery systems and their principle roles in disease and virus therapies

Drug delivery system	Description	Disease/viral target	References
Dendrimer	<ul style="list-style-type: none"> • Hyperbranched, monodispersed, water soluble (1–100 nm) macromolecule • Encapsulated drug in its interior or adsorbs drug on and conjugates to its surface groups • Releases free 5-fluorouracil upon hydrolysis • Intracellular delivery of poorly soluble drugs • Carriers in gene therapy 	<ul style="list-style-type: none"> • Inhibitors of haemagglutinin of human erythrocytes by Influenza virus • Amino groups of dendrimer react with nucleic acid phosphate groups to form transfection complexes 	56 and 57
Microsphere & nanocapsule	<ul style="list-style-type: none"> • Microsphere – drug is dispersed within polymer throughout particle • Nanocapsule-cavity contains drug surrounded by polymer membrane • Drug release through diffusion through polymer or degradation of polymer 	<ul style="list-style-type: none"> • Entraps luteinizing hormone-releasing hormone in prostate cancer 	57 and 58
Liposomes	<ul style="list-style-type: none"> • Amphiphilic vesicular structures made up of cholesterol & phospholipids • Core suited for hydrophilic drug delivery • Phospholipid membrane encapsulate hydrophobic drugs 	<ul style="list-style-type: none"> • Facultative intracellular bacteria-mediated infections • Parasites (<i>e.g.</i> Leishmania) • Viruses • Systemic fungal diseases in cancer • Melanomas 	56, 59 and 60
Micelles	<ul style="list-style-type: none"> • Core comprised of hydrophobic polymers • Shell comprised of hydrophilic polymers • Suitable for drugs with poor solubility • Nanosize; <i>in vivo</i> endurance; remains stable in plasma • Delivery of drugs & small interfering RNA 	<ul style="list-style-type: none"> • Targets tumour sites in cancer by active/passive mechanisms 	56, 61 and 62
Hydrogel	<ul style="list-style-type: none"> • Network of natural/synthetic hydrophilic polymers that are cross-linked • Highly absorbent, biodegradable, high porosity, biocompatible • Swell rapidly in aqueous solution • Used in oral & topical drug delivery 	<ul style="list-style-type: none"> • Local & systemic diseases 	56, 63 and 64
Gold/silver nanoparticle	<ul style="list-style-type: none"> • Low toxicity; high specificity & selectivity • Easily controlled & modifiable • High ratio of surface area:amount • Conjugation of proteins on colloidal gold nanoparticles occurs <i>via</i> electrostatic interactions between citrate (–) on surfaces of gold nanoparticles & groups (+) on proteins • Effortless cellular penetration 	<ul style="list-style-type: none"> • Oral delivery of insulin in diabetes; salmon calcitonin for postmenopausal osteoporosis; growth hormone for decelerated/stunted growth-associated diseases • Selective damage of tumour cells • Certain infectious & dermal diseases • H1N1, H3N2, H5N1 Influenza A virus • Herpes simplex virus • Human immunodeficiency virus • Hepatitis B virus • Metapneumovirus • Respiratory syncytial virus 	65–68
Lipid nanoparticle	<ul style="list-style-type: none"> • 10–1000 nm • Carriers with dispersed melted lipid in surfactant • Colloidal system with hydrophobic core that encloses drug & surface coated with hydrophilic polymers 	<ul style="list-style-type: none"> • Humoral immunity against Ebola infection • Silencing of hepatitis C virus replication • Gene therapy 	56 and 69–72
Aptamer	<ul style="list-style-type: none"> • Short, single-stranded (ss) DNA or RNA that have definitive 2° & 3° structures that strongly bind to specific target proteins • Low immunogenicity & toxicity • Variety of targets & modifiable chemical structure 	<ul style="list-style-type: none"> • α-Thrombin in thrombosis • PTK7 and nucleolin in cancer • IGHM in lymphoma • VEGF in age-related macular degeneration • A1 domain of vWF in thrombotic microangiopathies & carotid artery disease • Neutralizes r5 strains of HIV-1 • Blocks gp120-CCR5 interaction 	73–77

fields are used to calculate potential energies of particles and electrostatic forces that occur between atoms in a system.⁸⁸ Some force fields that can be used for molecular dynamic simulations include NAMD,⁸⁹ Gromacs,⁹⁰ Amber⁹¹ and Charm.⁹² Complexes can also be simulated through lipid bilayer, in cases where potential compounds need to enter

certain target tissues, which are surrounded by lipophilic membranes. This can give a prediction as to whether or not the potential compound will be able to pass through the lipid membrane or not. Softwares that can be used to generate a 3D lipid bilayer model include CHARRM-GUI and Visual Molecular Dynamics (MEMBPLUGIN).^{46,93} A brief background and the



Table 3 Various molecular modeling approaches in drug discovery

	Description/application	Reference
Molecular mechanics (MM) calculations	Molecular systems are modeled using MM, which employs force fields to calculate a system's potential energy. MM may be used to analyze molecular systems of various sizes and complexity, and is the preferred approach for the simulation of proteins	25, 94 and 95
Quantum mechanics (QM/MM) calculations	The hybrid QM/MM method is a molecular dynamic simulation approach that integrates the accuracy of QM and speed of MM, to allow for chemical processes within proteins and solutions to be studied. QM/MM provides enhanced mechanistic aspects and binding energies of the interaction between a ligand and receptor	66, 94 and 96
Virtual screening	Virtual screening involves screening through databases for compounds with similar features. There are two types of approaches involved in virtual screening: (1) ligand or pharmacophore-based virtual screening which requires knowledge of an active ligand, and (2) structure-based virtual screening which relies on a known 3D structure of the protein	79, 83, 84, 86 and 97
Molecular docking	Molecular docking is utilized for: (1) studying potential ligand conformations within a protein's binding site and (2) approximation of the force of ligand–protein interactions. During docking, Cartesian coordinates of a separate ligand and receptor are utilized to anticipate the conformation of the ligand that fits into the binding site of the protein the best, producing a ligand–protein complex. Molecular docking uses a scoring function, which ranks each docking “pose” according to its binding energy	47, 78, 87 and 97
Molecular dynamics	Influential tool for analyzing energetics, mechanisms and dynamics of proteins and ligand–protein complexes. Molecular dynamics provide a perspective of the interaction between a ligand and protein at a molecular level, presenting an improved understanding of the experimental conclusions. These are effective in the identification of specific biological properties, e.g., binding mode, per-residue protonation state, and protein flexibility	87–89, 92, 97 and 98

applications of the methods that have been mentioned above are shown in Table 3.

A full expansion of the methods explained in Table 3 may be found in Honarparvar *et al.*, 2014.⁹⁷

Conclusion

The various challenges associated with ZIKV treatment has led to the ongoing search for a cure – one of the major problems being drug delivery across the BBB. The approaches described in this review serve to provide information that can be used for further research into the design of drugs with improved BBB-permeability profile that may have a greater ability to inhibit ZIKV. Though experimental validation is necessary, this is not the scope of the current study. Instead, this study serves as a cornerstone that will open doors to further experimental and molecular validation regarding ZIKV therapy.

Conflicts of interest

Authors declare no potential conflicts of interest.

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References

- I. Briguglio, S. Piras, P. Corona and A. Carta, *Int. J. Med. Chem.*, 2011, **2011**, 1–22.
- R. K. Singh, K. Dhama, Y. S. Malik, M. A. Ramakrishnan, K. Karthik, R. Tiwari, S. Saurabh, S. Sachan and S. K. Joshi, *Vet. Q.*, 2016, **36**, 150–175.
- D. Musso, C. Roche, E. Robin, T. Nhan, A. Teissier and V. M. Cao-Lormeau, *Emerging Infect. Dis.*, 2015, **21**, 359–361.
- D. Musso, T. Nhan, E. Robin, C. Roche, D. Bierlaire, K. Zisou, A. Shan Yan, V. M. Cao-Lormeau and J. Broult, *Eurosurveillance*, 2014, **19**, 14–16.
- M. Besnard, S. Lastère, A. Teissier, V. M. Cao-Lormeau and D. Musso, *Eurosurveillance*, 2014, **19**, 8–11.
- G. W. A. Dick, S. F. Kitchen and A. J. Haddow, *Trans. R. Soc. Trop. Med. Hyg.*, 1952, **55**, 509–520.
- M. R. Duffy, C. Tai-Ho, T. Hancock, A. M. Powers, J. L. Kool, R. S. Lanciotti, M. Pretrick, C. Dubray, L. Guillaumot, A. Griggs, M. Bel, A. J. Lambert, J. Laven, O. Kosoy, A. Panella, B. J. Biggerstaff, M. Fischer and E. B. Hayes, *N. Engl. J. Med.*, 2009, **360**, 2536–2543.
- D. Musso, E. J. Nilles and V. M. Cao-Lormeau, *Clin. Microbiol. Infect.*, 2014, **20**, O595–O596.
- V. Sikka, V. K. Chattu, R. K. Popli, T. J. Galwankar, C. Sagar, D. Kelkar, S. G. Sawicki, S. P. Stawicki and T. J. Papadimos, *J. Global Infect. Dis.*, 2016, **8**, 3–15.
- S.-I. Yun and Y.-M. Lee, *J. Microbiol.*, 2017, **55**, 204–219.
- G. S. Campos, A. C. Bandeira and S. I. Sardi, *Emerging Infect. Dis.*, 2015, **21**, 1885–1886.
- D. Musso and D. J. Gubler, *Nature*, 2016, **11**, 10–20.
- F. Krauer, M. Riesen, L. Reveiz, O. T. Oladapo, R. Martínez-Vega, T. V. Porgo, A. Haefliger, N. J. Broutet and N. Low, *PLoS Med.*, 2017, **14**, 1–27.
- P. Ramharack and M. E. S. Soliman, *RSC Adv.*, 2016, **6**, 68719–68731.



- 15 D. Musso, C. Roche, T. X. Nhan, E. Robin, A. Teissier and V. M. Cao-Lormeau, *J. Clin. Virol.*, 2015, **68**, 53–55.
- 16 J. M. Mansuy, M. Dutertre, C. Mengelle, C. Fourcade, B. Marchou, P. Delobel, J. Izopet and G. Martin-Blondel, *Lancet Infect. Dis.*, 2016, **16**, 405.
- 17 J. M. Turmel, P. Abgueguen, B. Hubert, Y. M. Vandamme, M. Maquart, H. Le Guillou-Guillemette and I. Leparcoffart, *Lancet*, 2016, **387**, 2501.
- 18 A. C. Gourinat, O. O'Connor, E. Calvez, C. Goarant and M. Dupont-Rouzeyrol, *Emerging Infect. Dis.*, 2015, **21**, 84–86.
- 19 G. Calvet, R. S. Aguiar, A. S. O. Melo, S. A. Sampaio, I. de Filippis, A. Fabri, E. S. M. Araujo, P. C. de Sequeira, M. C. L. de Mendonça, L. de Oliveira, D. A. Tschoeke, C. G. Schrago, F. L. Thompson, P. Brasil, F. B. dos Santos, R. M. R. Nogueira, A. Tanuri and A. M. B. de Filippis, *Lancet Infect. Dis.*, 2016, **16**, 653–660.
- 20 O. Faye, O. Faye, A. Dupressoir, M. Weidmann, M. Ndiaye and A. Alpha Sall, *J. Clin. Virol.*, 2008, **43**, 96–101.
- 21 R. Hamel, O. Dejarnac, S. Wichit, P. Ekcharyawat, A. Neyret, N. Luplertlop, M. Perera-Lecoin, P. Surasombattana, L. Talignani, F. Thomas, V. M. Cao-Lormeau, V. Choumet, L. Briant, P. Despres, A. Amara, H. Yssel and D. Misse, *J. Virol.*, 2015, **89**, 8880–8896.
- 22 O. Faye, C. C. M. Freire, A. Iamarino, O. Faye, J. Velasco, C. De Oliveira, M. Diallo, P. M. A. Zanotto and A. A. Sall, *PLoS Neglected Trop. Dis.*, 2014, **8**, 1–10.
- 23 H. Tian, X. Ji, X. Yang, W. Xie, K. Yang, C. Chen, C. Wu, H. Chi, Z. Mu, Z. Wang and H. Yang, *Protein Cell*, 2016, **7**, 450–454.
- 24 H. Tian, X. Ji, X. Yang, Z. Zhang, Z. Lu, K. Yang, C. Chen, Q. Zhao, H. Chi, Z. Mu, W. Xie, Z. Wang, H. Lou, H. Yang and Z. Rao, *Protein Cell*, 2016, **7**, 562–570.
- 25 B. Ganguly and E. Ganguly, *bioRxiv*, 2016, p. 54486.
- 26 M. K. White, H. S. Wollebo, J. David Beckham, K. L. Tyler and K. Khalili, *Ann. Neurol.*, 2016, **80**, 479–489.
- 27 N. J. da Fonseca Jr, *et al.*, *Biochem. Biophys. Res. Commun.*, 2017, DOI: 10.1016/j.bbrc.2017.01.041.
- 28 G. Li, M. Poulsen, C. Fenyvuesvolgyi, Y. Yashiroda, M. Yoshida and J. M. Simard, *Proc. Natl. Acad. Sci. U. S. A.*, 2016, **114**, 201619735.
- 29 P. Brasil, P. C. Sequeira, A. D. Freitas, H. E. Zogbi, G. A. Calvet, R. V. de Souza, A. M. Siqueira, M. C. de Mendonca, R. M. Nogueira, A. M. de Filippis and T. Solomon, *Lancet*, 2016, **387**, 1482.
- 30 J. J. Miner and M. S. Diamond, *Cell Stem Cell*, 2016, **18**, 559–560.
- 31 T. J. Nowakowski, A. A. Pollen, E. Di Lullo, C. Sandoval-Espinosa, M. Bershteyn and A. R. Kriegstein, *Cell Stem Cell*, 2016, **18**, 591–596.
- 32 H. Retallack, E. Di Lullo, C. Arias, K. A. Knopp, M. T. Laurie, C. Sandoval-Espinosa, W. R. M. Leon, R. Krencik, E. M. Ullian, J. Spatzza, A. A. Pollen, C. Mandel-Brehm, T. J. Nowakowski, A. R. Kriegstein, J. L. Derisi, N. Sestan and P.-Y. Shi, *Proc. Natl. Acad. Sci. U. S. A.*, 2016, **113**, 14408–14413.
- 33 P. K. Singh, J.-M. Guest, M. Kanwar, J. Boss, N. Gao, M. S. Juzych, G. W. Abrams, F.-S. Yu and A. Kumar, *JCI Insight*, 2017, **2**(4), DOI: 10.1172/jci.insight.92340.
- 34 N. R. Saunders, M. D. Habgood, K. Møllgård and K. M. Dziegielewska, *F1000Research*, 2016, **5**, 1–15.
- 35 M. R. Syme, J. W. Paxton and J. a. Keelan, *Clin. Pharmacokinet.*, 2004, **43**, 487–514.
- 36 W. A. Banks, *BMC Neurol.*, 2009, **9**, S3.
- 37 A. Seelig, R. Gottschlich and R. M. Devant, *Proc. Natl. Acad. Sci. U. S. A.*, 2016, **91**, 68–72.
- 38 J. M. Tarbell and M. Y. Pahakis, *J. Intern. Med.*, 2006, **259**, 339–350.
- 39 N. J. Barrows, R. K. Campos, S. T. Powell, K. R. Prasanth, G. Schott-Lerner, R. Soto-Acosta, G. Galarza-Muñoz, E. L. McGrath, R. Urrabaz-Garza, J. Gao, P. Wu, R. Menon, G. Saade, I. Fernandez-Salas, S. L. Rossi, N. Vasilakis, A. Routh, S. S. Bradrick and M. A. Garcia-Blanco, *Cell Host Microbe*, 2016, **20**, 259–270.
- 40 A. Daina, O. Michielin and V. Zoete, *Sci. Rep.*, 2017, **7**, 1–13.
- 41 A. Daina, O. Michielin and V. Zoete, *J. Chem. Inf. Model.*, 2014, **54**, 3284–3301.
- 42 Y. N. Gavhane and A. V. Yadav, *Saudi Pharm. J.*, 2012, **20**, 331–344.
- 43 P. Atkins and J. De Paula, *Physical chemistry for the life sciences*, 2011.
- 44 P. P. Kore, M. M. Mutha, R. V. Antre, R. J. Oswal and S. S. Kshirsagar, *J. Med. Chem.*, 2012, **2**, 139–148.
- 45 J. A. Arnott and S. L. Planey, *Expert Opin. Drug Discovery*, 2012, **7**, 863–875.
- 46 R. Guixà-González, I. Rodríguez-Espigares, J. M. Ramírez-Angueta, P. Carrió-Gaspar, H. Martínez-Seara, T. Giorgino and J. Selent, *Bioinformatics*, 2014, **30**, 1478–1480.
- 47 X.-Y. Meng, H.-X. Zhang, M. Mezei and M. Cui, *Curr. Comput.-Aided Drug Des.*, 2011, **7**, 146–157.
- 48 E. C. Hulme and M. A. Trevethick, *Br. J. Pharmacol.*, 2010, **161**, 1219–1237.
- 49 T. H. Scheuermann, S. B. Padrick, K. H. Gardner and C. A. Brautigam, *Anal. Biochem.*, 2016, **496**, 79–93.
- 50 Y. Q. Deng, N. N. Zhang, C. F. Li, M. Tian, J. N. Hao, X. P. Xie, P. Y. Shi and C. F. Qin, *Open Forum Infect. Dis.*, 2016, **3**, 6–9.
- 51 S. E. Reznik and C. R. Ashby, *Int. J. Infect. Dis.*, 2017, **55**, 29–30.
- 52 C. Roney, P. Kulkarni, V. Arora, P. Antich, F. Bonte, A. Wu, N. N. Mallikarjuana, S. Manohar, H.-F. Liang, A. R. Kulkarni, H.-W. Sung, M. Sairam and T. M. Aminabhavi, *J. Controlled Release*, 2005, **108**, 193–214.
- 53 C. Spuch and C. Navarro, *J. Drug Delivery*, 2011, **2011**, 1–12.
- 54 M. S. Gunay, A. Y. Ozer and S. Chalou, *Curr. Neuropharmacol.*, 2016, **14**, 376–391.
- 55 E. Blanco, H. Shen and M. Ferrari, *Nat. Biotechnol.*, 2015, **33**, 941–951.
- 56 A. Srivastava, T. Yadav, S. Sharma, A. Nayak, A. Kumari and N. Mishra, *J. Biosci. Med.*, 2016, **4**, 69–84.
- 57 W. B. Liechty, D. R. Kryscio, B. V. Slaughter and N. A. Peppas, *Annu. Rev. Chem. Biomol. Eng.*, 2010, **1**, 149–173.
- 58 Z. Abouelfadel and D. Crawford, *Ther. Clin. Risk Manage.*, 2008, **4**, 513–526.
- 59 A. Coune, *Infection*, 1988, **16**, 141–147.
- 60 M. A. Tran, R. J. Watts and G. P. Robertson, *Pigm. Cell Melanoma Res.*, 2009, **22**, 388–399.



- 61 U. Kedar, P. Phutane, S. Shidhaye and V. Kadam, *Nanomedicine*, 2010, **6**, 714–729.
- 62 X.-B. Xiong, H. Uludağ and A. Lavasanifar, *Biomaterials*, 2010, **31**, 5886–5893.
- 63 D. Bhowmik, H. Gopinath, B. Pragati Kumar, S. Duraivel and K. P. Sampath Kumar, *Pharma Innovation*, 2012, **1**, 12–31.
- 64 L. A. Sharpe, A. M. Daily, S. D. Horava and N. A. Peppas, *Expert Opin. Drug Delivery*, 2014, **11**, 901–915.
- 65 X. Yu, I. Trase, M. Ren, K. Duval, X. Guo and Z. Chen, *J. Nanomater.*, 2016, DOI: 10.1155/2016/1087250.Design.
- 66 H. Daraee, A. Eatemadi, E. Abbasi, S. Fekri Aval, M. Kouhi and A. Akbarzadeh, *Artif. Cells, Nanomed., Biotechnol.*, 2014, **1401**, 1–13.
- 67 W. Tao, B. L. Hurst, A. K. Shakya, M. J. Uddin, R. S. J. Ingrole, M. Hernandez-Sanabria, R. P. Arya, L. Bimler, S. Paust, E. B. Tarbet and H. S. Gill, *Antiviral Res.*, 2017, **141**, 62–72.
- 68 M. Rai, S. D. Deshmukh, A. P. Ingle, I. R. Gupta, M. Galdiero and S. Galdiero, *Crit. Rev. Microbiol.*, 2014, **42**, 46–56.
- 69 S. Mukherjee, S. Ray and R. S. Thakur, *Indian J. Pharm. Sci.*, 2009, **71**, 349–358.
- 70 J. D. Bazzill, C. L. Cooper, Y. Fan, S. Bavari and J. J. Moon, *J. Immunol.*, 2016, **196**(1 Suppl.), 76.
- 71 J. Torrecilla, A. del Pozo-Rodríguez, M. yñgeles Solinís, P. S. Apaolaza, B. Berzal-Herranz, C. Romero-López, A. Berzal-Herranz and A. Rodríguez-Gascón, *Colloids Surf., B*, 2016, **146**, 808–817.
- 72 A. del Pozo-Rodríguez, M. Á. Solinís and A. Rodríguez-Gascón, *Eur. J. Pharm. Biopharm.*, 2016, **109**, 184–193.
- 73 A. K. Dey, M. Khati, M. Tang, R. Wyatt, S. M. Lea and W. James, *J. Virol.*, 2005, **79**, 13806–13810.
- 74 F. Jiang, B. Liu, J. Lu, F. Li, D. Li, C. Liang, L. Dang, J. Liu, B. He, S. A. Badshah, C. Lu, X. He, B. Guo, X. B. Zhang, W. Tan, A. Lu and G. Zhang, *Int. J. Mol. Sci.*, 2015, **16**, 23784–23822.
- 75 S. D. Jayasena, *Clin. Chem.*, 1999, **45**, 1628–1650.
- 76 P. Ray and R. R. White, *Pharmaceuticals*, 2010, **3**, 1761–1778.
- 77 M. Blind and M. Blank, *Mol. Ther.–Nucleic Acids*, 2015, **4**, e223.
- 78 E. Lionta, G. Spyrou, D. K. Vassilatis and Z. Cournia, *Curr. Top. Med. Chem.*, 2014, **14**, 1923–1938.
- 79 J. J. Irwin and B. K. Shoichet, *J. Chem. Inf. Model.*, 2005, **45**, 177–182.
- 80 D. R. Koes and C. J. Camacho, *Nucleic Acids Res.*, 2012, **40**, 409–414.
- 81 E. F. Pettersen, T. D. Goddard, C. C. Huang, G. S. Couch, D. M. Greenblatt, E. C. Meng and T. E. Ferrin, *J. Comput. Chem.*, 2004, **25**, 1605–1612.
- 82 O. Trott and A. J. Olson, *J. Comput. Chem.*, 2010, **31**, 445–461.
- 83 N. Mhlongo and M. E. Soliman, *Lett. Drug Des. Discovery*, 2016, **13**, 1033–1046.
- 84 F. N. Cele, R. Muthusamy and M. E. Soliman, *Drug Des., Dev. Ther.*, 2016, **10**, 1365–1377.
- 85 A. Skelton, Y. Maharaj and M. E. Soliman, *Cell. Mol. Bioeng.*, 2014, **7**, 45–57.
- 86 E. O. Pyzer-Knapp, C. Suh, R. Gómez-Bombarelli, J. Aguilera-Iparraguirre and A. Aspuru-Guzik, *Annu. Rev. Mater. Res.*, 2015, **45**, 195–216.
- 87 H. Alonso, A. A. Bliznyuk and J. E. Gready, *Med. Res. Rev.*, 2006, **26**, 531–568.
- 88 M. A. González, *Collection SFN*, 2011, **12**, 169–200.
- 89 M. Nelson, W. Humphrey, A. Gursoy, A. Dalke, L. Kal, R. D. Skeel and K. Schulten, *Int. J. High Perform. Comput. Appl.*, 1996, **251–268**, 251–268.
- 90 B. Hess, C. Kutzner, D. Van Der Spoel and E. Lindahl, *J. Chem. Theory Comput.*, 2008, **4**, 435–447.
- 91 D. A. Case, T. E. Cheatham, T. Darden, H. Gohlke, R. Luo, K. M. Merz, A. Onufriev, C. Simmerling, B. Wang and R. J. Woods, *J. Comput. Chem.*, 2005, **26**, 1668–1688.
- 92 B. R. Brooks, R. E. Bruccoleri, B. D. Olafson, D. J. States, S. Swaminathan and M. Karplus, *J. Comput. Chem.*, 1983, **4**, 187–217.
- 93 Y. Qi, X. Cheng, J. Lee, J. V. Vermaas, T. V. Pogorelov, E. Tajkhorshid, S. Park, J. B. Klauda and W. Im, *Biophys. J.*, 2015, **109**, 2012–2022.
- 94 A. W. Duster, C.-H. Wang, C. M. Garza, D. E. Miller and H. Lin, *WIREs Computational Molecular Science*, 2017, DOI: 10.1002/wcms.1310.
- 95 K. Vanommeslaeghe, O. Guvench and A. D. Mackerell, *Curr. Pharm. Des.*, 2014, **20**, 3281–3292.
- 96 S. B. Vepuri, H. C. Devarajgowda and M. E. Soliman, *J. Mol. Struct.*, 2016, **1105**, 194–204.
- 97 B. Honarparvar, T. Govender, G. E. M. Maguire, M. E. S. Soliman and H. G. Kruger, *Chem. Rev.*, 2013, **114**(1), 493–537.
- 98 A. Ganesan, M. L. Coote and K. Barakat, *Drug Discovery Today*, 2017, **22**, 249–269.

