



Cite this: *J. Anal. At. Spectrom.*, 2025, **40**, 3192

LA-ICP-TOF-MS for quantitative mapping of biogenic carbonate samples using matrix-matched nanoparticulate pressed powder pellets

Ana Lores-Padín, ^a Thibaut Van Acker, ^a Niels J. de Winter,^b Martin Wiech, ^c Simon Nordstad,^d Yannic Hallier^d and Frank Vanhaecke ^a

This study evaluated the micro-homogeneity of seven different commercially available nanoparticulate pressed pellets based on a CaCO_3 matrix and their utility for quantitative elemental mapping of biogenic carbonates using laser ablation-inductively coupled plasma-time-of-flight-mass spectrometry (LA-ICP-TOF-MS). The analytical performance of matrix-matched calibration (using the aforementioned nano-pellets) was compared against that of non-matrix-matched calibration using a silicate glass (NIST SRM 610) reference material for quantification. Calibration with nano-pellets, combined with the use of Ca as an internal standard, significantly improved the quantification accuracy, providing recoveries between 80–120% for the majority of the 18 elements selected and spread across a wide concentration range (from $\text{sub-}\mu\text{g g}^{-1}$ to tens of wt%). However, some nano-pellets (e.g., BPLM-NP and BAM-RS3-NP) exhibited higher heterogeneity, leading to biased recoveries. Also, an inverse correlation between the mass fraction and the relative standard deviation (RSD) was observed. Throughout the work, elemental mapping was conducted with a laser beam size of $20 \times 20 \mu\text{m}^2$ as a compromise between spatial resolution, sensitivity ($\text{sub-}\mu\text{g g}^{-1}$ limits of detection required for trace elements), linear dynamic range, total analysis time and size of the region-of-interest. The quantitative mapping approach enabled the generation of high-resolution, multi-elemental 2D-maps of various CaCO_3 -based natural chronological archives, including fish otoliths and bivalve shells, revealing detailed elemental distribution patterns for both trace (e.g., Mn, Ba) and major elements (e.g., Sr). This LA-ICP-TOF-MS methodology provides

Received 18th July 2025
Accepted 9th September 2025

DOI: 10.1039/d5ja00280j

rsc.li/jaas

^aAtomic & Mass Spectrometry-A&MS Research Group, Department of Chemistry, Ghent University,

a powerful tool for resolving intricate microstructures and thus, for chronologically tracking bioaccumulation of environmentally relevant metals, offering significant advantages over traditional 1D (line scanning) analysis as the latter may lead to misinterpretation of elemental distributions.

Introduction

Biogenic carbonates, such as bivalve shells and otoliths, serve as natural chronological archives of past environmental conditions. Their incremental growth allows for the incorporation of major, minor, and trace elements from the surrounding environment into their calcium carbonate (CaCO_3) matrix, preserving chemical signatures that can remain unaltered over long (geological) timescales.¹ This makes them valuable archives as several proxies measured in carbonates can be relied on for reconstructing seasonal and long-term climate and environmental changes, biological processes, and even anthropogenic influences such as pollution episodes.² Utilizing the full potential of these records requires the use of a highly sensitive analytical technique with high spatial resolution (*i.e.*, $\leq 20\ \mu\text{m}$) capable of distinguishing micro-scale elemental and isotopic variations.³

Laser ablation – inductively coupled plasma-mass spectrometry (LA-ICP-MS) has been commonly used in the study of biogenic carbonates⁴ due to its excellent limits of detection (LoDs, typically in the ng g^{-1} range), a linear dynamic range of up to 10 orders of magnitude, minimal sample preparation requirements and high sample throughput.⁵ The technique has been used for both bulk (using large beam diameters, typically $\geq 100\ \mu\text{m}$) and microanalysis (bulk spot analysis and 1D-line scan analysis using spots typically ranging from 20 to 80 μm diameter).⁶ Recent advances in LA technology, including the introduction of low-dispersion ablation cells^{7,8} and transfer devices with improved aerosol transport efficiency,⁹ have significantly enhanced the technique's capabilities. These developments now enable mapping with a spatial resolution down to approximately 1 μm and yield shorter single pulse response (SPR) profiles of $\leq 1\ \text{ms}$ duration (defined as the width of the transient signal observed upon firing a single laser pulse at 10% of its maximum height). Furthermore, the integration of time-of-flight analyzers in ICP-MS instrumentation (ICP-TOF-MS), allowing quasi-simultaneous monitoring of nearly the entire periodic table (typically $^{23}\text{Na}^+ - ^{238}\text{U}^{16}\text{O}^+$) within just a few microseconds ($< 50\ \mu\text{s}$), has further expanded the applicability of LA-ICP-MS.¹⁰ Together, these advances allow for rapid data acquisition at up to 1000 pixels per second, significantly boosting the technique's potential for ultra-fast, high-resolution, multi-elemental mapping.¹¹

Despite significant advances in LA and ICP-MS instrumentation, accurate elemental quantification in carbonate matrices remains inherently challenging. This is primarily due to the matrix-dependent nature of the laser-sample interaction: with the commonly used nanosecond pulse lasers, the occurrence of thermal effects during the ablation process results in non-stoichiometric sampling and heterogeneous particle plumes, leading to elemental fractionation during ablation, aerosol transport and plasma processes such as particle vaporization

and ion generation. In this context, apart from optimally tuning the LA parameters and ICP settings to minimize elemental fractionation ($^{238}\text{U}^+/^{232}\text{Th}^+$ signal ratio ~ 1 upon ablation of a NIST SRM 610 or 612 glass from the National Institute of Standards and Technology),^{12,13} previous studies on carbonates have shown that also the laser wavelength exerts an important effect on the quantification. The use of a shorter laser wavelength, such as 193 nm, instead of 213 or 266 nm, enables more effective ablation of carbonate matrices due to the higher absorption efficiency, the formation of smaller particles (which significantly reduces vaporization-induced elemental fractionation), and reduced thermal

that using MACS-3 for obtaining quantitative high-resolution maps (*i.e.*, 5 μm pixel size) of small otoliths led to high relative standard deviations (RSDs) of 30–80% due to trace element heterogeneity as previously discussed.

A significant advancement in the field was the development of a novel carbonate pellet fabrication method by Garbe-Schönberg and Müller,³⁰ which enabled a reduction of the grain size to the sub-micron scale by implementing a 2-step milling procedure that uses a planetary ball mill for further grinding the material into a nanoscale powder after a freeze-drying step. The nanoparticulate powder, previously homogenized using a mixer mill to ensure uniform particle distribution, is pressed into pellets using a hydraulic press without the need of any binder. Jochum *et al.* demonstrated the potential of three novel “nanopellets”: JCP-1-NP, JCT-1-NP (natural matrix source), and MACS-3NP (synthetic matrix source) showing a 2- to 3-fold higher uniformity in trace element distribution, leading to lower variability and higher accuracy in LA-ICP-MS spot analysis quantification.³⁰ JCT-1-NP and SPLT-NP (speleothem, natural matrix) have been recently used as external standard for 1D-elemental analysis (line profiling) of speleothems with 50 μm laser beam spots¹⁹ as well as to enable accurate quantitative high spatial-resolution 2D-mapping of marine carbonates, using a laser spot size of $\sim 2\ \mu\text{m}$.³¹

Typically, LA-ICP-MS analysis of carbonate archives relies on 1D analysis (line scanning) across a specimen's cross-section.^{6,32} Such approach assumes homogeneous deposition of CaCO_3 and the other elements within well-defined growth layers. However, recent research has shown that elemental distributions within biogenic carbonates are often more heterogeneous than previously assumed, influenced by both environmental and biological factors that introduce structural discontinuities and variations in crystallization.³³ Such complexities can lead to misinterpretations, especially in areas lacking a clear chronological structure. In this context, two-dimensional (2D) elemental mapping offers a more comprehensive analytical approach, capturing the full spatial distribution of elements across the specimen. Although different 2D-mapping applications in carbonates using techniques like synchrotron X-ray fluorescence microscopy (SXFM) and wavelength-dispersive X-ray electron probe microanalysis (EPMA) have been employed for otolith analysis, they require long measurement times (often exceeding 8 hours for areas $>4\ \text{mm}^2$) and access to large-scale synchrotron facilities.³⁴ EPMA provides high spatial resolution ($<3\ \mu\text{m}$) but suffers from relatively poor detection limits ($>10\ \mu\text{g g}^{-1}$).³³ Synchrotron- and lab-based micro-X-ray fluorescence analysis techniques have also aided in the 2D-characterization and speciation analysis of mollusc shell compositions.^{35–38} While the resolution of these techniques (typically 20–50 μm spot size) approaches that of LA-ICP-MS, quantification limits (especially of lab-based systems) tend to be higher ($\sim 1\ \mu\text{g g}^{-1}$). Additionally, the long attenuation length of X-rays ($>100\ \mu\text{m}$ for energies needed to excite and measure Sr) has a tendency to integrate results below the sample surface, which can lead to time-averaging in incremental carbonate archives which often contain curved growth increments in three-dimensional space (see, *e.g.*, discussion in de Winter and Claeys, 2017).³⁵

More recently, LA-ICP-TOF-MS has been applied to the study of biogenic structures, enabling fast multi-elemental 2D-mapping at high spatial resolution. For instance, otolith specimens were mapped using laser spot size of 4 μm at an acquisition rate of up to 25 pixels per s.²⁸ Coral samples have also been analysed using similar setups, employing even smaller spot sizes (1–2 μm) and faster acquisition rates (up to 200 pixels per s).³¹ These advances demonstrate the capabilities of LA coupled to ICP-TOF-

Table 1 Instrument settings and data acquisition conditions used in this work: icpTOF 2R and Iridia 193 nm LA-unit

LA unit		ICP-TOF-MS	
Carrier gas flow, He (L min ⁻¹)	1.05	RF power (W)	1550
Laser beam size (μm)	20 × 20 (2D LA-mapping), 10 × 10, 5 × 5	Sampling depth (mm)	5.2
Fluence (J cm ⁻²)	4.0	Neb. Flow, Ar (mL min ⁻¹)	0.92
Repetition rate (Hz)	50 (2D LA-mapping), 2 (SPR & ODG opt)	CCT-mode, H ₂ (mL min ⁻¹)	2.0
Dosage	1 (2D LA-mapping)	Notched masses (<i>m/z</i>)	40
Scan speed (mm s ⁻¹)	1.0	Notch amplitude (V)	1.0

aiming at high sensitivity across the elemental mass range (⁵⁹Co⁺, ¹¹⁵In⁺ and ²³⁸U⁺), low oxide ion formation (²³²Th¹⁶O⁺/²³²Th⁺ < 2%) and minimizing elemental fractionation (²³⁸U⁺/²³²Th⁺ ~ 1). The optimized instrument settings and data acquisition conditions used in this work are summarized in the Table 1. Note that for assessing of the single pulse response (SPR) profiles and setting the offset delay generator (ODG) time, sometimes referred to as aerosol transfer delay time, short integration times of <500 μs were required to define the signal peak in sufficient detail. The ODG is the total time it takes between firing an individual laser pulse and the arrival of ions at the Multi-Channel Plate (MCP). This value is required for adequate timing of the trigger signal of the laser system towards the ICP-TOF-MS unit. This offset delay is influenced not only by the matrix of the sample but also by instrumental parameters such as gas flow rates and torch position.

Standards and samples

In this study, several nanoparticulate (NP) powder-pressed pellets, manufactured by myStandards GmbH (Kiel, Germany), were examined. These pellets are manufactured according to a protocol first described by Garbe-Schönberg and Müller in 2014.³⁰ The resulting carbonate nanopowder is pressed into pellets with a diameter of 10 mm using a programmable hydraulic press. Notably, these pellets are pressed without binders to avoid potential contamination and matrix effects during laser ablation. For this work, the following nano-pellets were evaluated as matrix-matched external calibration standards: SPLT-NP-B01 (speleothem powder), NFHS-2-NP (foraminifera powder), ECRM-752-1*-NP (limestone powder), CRMS-NP-B01 (coral powder), BAM-RS3-NP ("RS3" powder, synthetic calcite), Apatite-NP-B01 (crystal apatite powder, note that this is a calcium phosphate matrix while the other nano-pellets have a calcium carbonate matrix), and BPLM-NP-B01 (Bivalve shell powder). The elemental compositions of Apatite-NP, BPLM-NP, and SPLT-NP have been certified in accordance with ISO

Results & discussion

LA-ICP-TOF-MS method optimization for the CaCO₃ matrix

For successful analysis of CaCO₃ sample matrices using LA-ICP-TOF-MS, it is essential to achieve sufficient sensitivity for the reliable detection of major, minor, and trace elements across the entire elemental mass range, while paying attention to the linear dynamic range avoiding detector saturation due to the high signal intensities originating from major elements (*e.g.*, Ca). Thus, the use of Collision/Reaction Cell Technology (CCT) with hydrogen (H₂) gas was evaluated to suppress Ar-based polyatomic interferences and boost the sensitivity as a result of collisional focusing. The sensitivity was monitored at different H₂ gas flow rates (0–6 mL min^{−1}) while ablating a NIST SRM 610 glass and in Fig. 1A, the sensitivities obtained for the target nuclides ²⁴Mg, ⁵²Cr, ⁵⁶Fe, ⁸⁸Sr, ¹³⁷Ba, ²³²Th and ²³⁸U are displayed. It was found that pressurizing the cell with pure H₂ at a flow rate of 2 mL min^{−1} increased sensitivity by approximately 1.7-fold for the high-mass range, *e.g.*, ²³²Th, and ²³⁸U, and 1.5-fold for the mid-mass range, *e.g.*, ⁵²Cr, ⁵⁶Fe, and ⁸⁸Sr and ¹³⁷Ba, while slightly reducing the sensitivity for lighter elements such as ²⁴Mg which is in accordance with observations previously reported by Burger *et al.*,⁴⁵ when using a mix of H₂/He. Moreover, the use of H₂ also attenuated the signal intensity of polyatomic ions, such as ArO⁺ interfering with the monitoring of ⁵⁶Fe⁺ and other Ar-containing ions,³⁹ including ⁴⁰Ar⁺. In general, by using a low flow rate (*i.e.*, ~2 mL min^{−1} of H₂) the signal-to-

noise ratio (SNR) was improved by decreasing the background levels (Fig. S1A). Therefore, finding a balance between reaching high sensitivity for detecting minor and trace elements and avoiding overloading the ICP-TOF-MS detector with intense signals from major elements like Ca (which represents ~40% of the total mass) is crucial for both nano-pellets and samples. To mitigate the latter risk, the most abundant isotope, ⁴⁰Ca⁺, was attenuated using the notch filter to prevent detector saturation (see Table 1 for notching filter details), though even less abundant isotopes such as ⁴³Ca⁺ and ⁴⁴Ca⁺ still produced highly intense signals as shown in the mass spectrum below observed upon ablation of NFHS-2-NP (Fig. 1B). Additionally in Fig. 1C, a part of the mass spectrum is displayed, zoomed in on the low mass range between 36–47 amu. Removing ⁴⁰Ca⁺ using the notch filter also affected the signal of neighbouring masses such as that for ⁴²Ca⁺, as a result of which its signal intensity was lower than that of ⁴³Ca⁺, despite the higher natural relative abundance (0.647% *vs.* 0.135% for ⁴²Ca and ⁴³Ca, respectively). It is also important to consider hydride formation when H₂ is used as a reaction gas, for instance, while the ³⁶Ar⁺ peak is almost negligible, a significant ³⁶ArH⁺ signal can be observed at *m/z* 37. Although ThO⁺/Th⁺ ratios increased from <1% (0 mL min^{−1}, standard mode) to 2.8% (6 mL min^{−1}), they remained below 1.5% at 2 mL min^{−1} H

At the LA-side, it was evaluated whether for a laser beam size of $20 \times 20 \mu\text{m}^2$ (square mask) different laser fluences of 2, 3, 4 and 5 J cm^{-2} were capable of effectively ablating the CaCO_3 matrix. Using the NFHS-2-NP nano-pellet as a CaCO_3 matrix reference, increasing the fluence from 2 to 4 J cm^{-2} resulted in improved sensitivity across all examined elements (by a factor of 1.5), including those present at low ($<1 \mu\text{g g}^{-1}$; e.g., Co, Cr, Pr, U, Th), mid ($1\text{--}20 \mu\text{g g}^{-1}$; e.g., Ce, Pb, Cu, Zn), and high concentrations ($>20 \mu\text{g g}^{-1}$; e.g., Na, Mn, Mg, Al, Fe, Sr), as shown in Fig. 2. However, increasing the fluence beyond 4 J cm^{-2} to 5 J cm^{-2} resulted in a slight enhancement of $<5\%$ only, thus 4 J cm^{-2} was selected as the optimal value ensuring effective carbonate ablation.

Evaluation of the micro-homogeneity of nano-pellets using LA-ICP-TOF-MS

To evaluate the elemental micro-homogeneity of the different nano-pellets, firstly, a qualitative study was carried out by performing 2D-mapping for each individual nano-pellet from the set. Therefore, the duration of the SPR profiles and the ODG had to be assessed for this specific matrix type. Thus, by using the parameters described above (i.e., $20 \times 20 \mu\text{m}^2$ beam size and 4 J cm^{-2} fluence and the gas flows summarised in Table 1), the SPR duration defined as FW0.1 M was found to be approximately 20 ms, which restricted the pixel acquisition rate to 50 pixels/s. The laser repetition rate was set to 50 Hz and only a single shot was fired per pixel position (dosage of 1). Note that apart from the matrix type and the parameters described above, also the PEEK transport tubing length, positioning of the ARIS with respect to the torch injector, as well as the He flow rates play a role in achieving well-shaped and constant SPRs.

Simultaneous 2D multi-element mapping of $3 \times 1 \text{ mm}^2$ regions of interest (ROIs) were performed for each nano-pellet (corresponding to 3.82% of the total nano-pellet surface) using LA-ICP-TOF-MS, generating a total of 7500 data points (i.e., pixels in the 2D-map), corresponding to 50 lines of 3 mm

length per pellet (1 line = 150 data points). Visual inspection of the qualitative ^{44}Ca normalized maps for selected target elements present in the nano-pellets reveals the degree of homogeneity. For instance, Fig. 3 allows visual comparison between NFHS-2-NP and BPLM-NP nano-pellets. While NFHS-2-NP shows a high degree of micro-homogeneity for all the target elements, even those prone to surface contamination such as Zn, in BPLM-NP elements such as Al, Mn, Fe, Cu, Zn and Pb show a less homogeneous distribution. This is also the case for other nano-pellets (Fig. S2), with BAM-RS3-NP exhibiting more heterogeneity compared to the more uniform distribution observed in CRMS-NP or ECRM-752-1-NP.

In addition, also the repeatability based on the relative standard deviation (RSD) of the average signal intensity for independent line scan measurements was evaluated (total lines, $n = 50$ & sub-sets $n = 25, 10, 5$ and 3) and compared to values obtained for NIST SRM 610 glass (RSDs $<2\%$, Table S1). For example, while NFHS-2-NP exhibited RSD values between 1% and 7% across all elements (Table 2, top

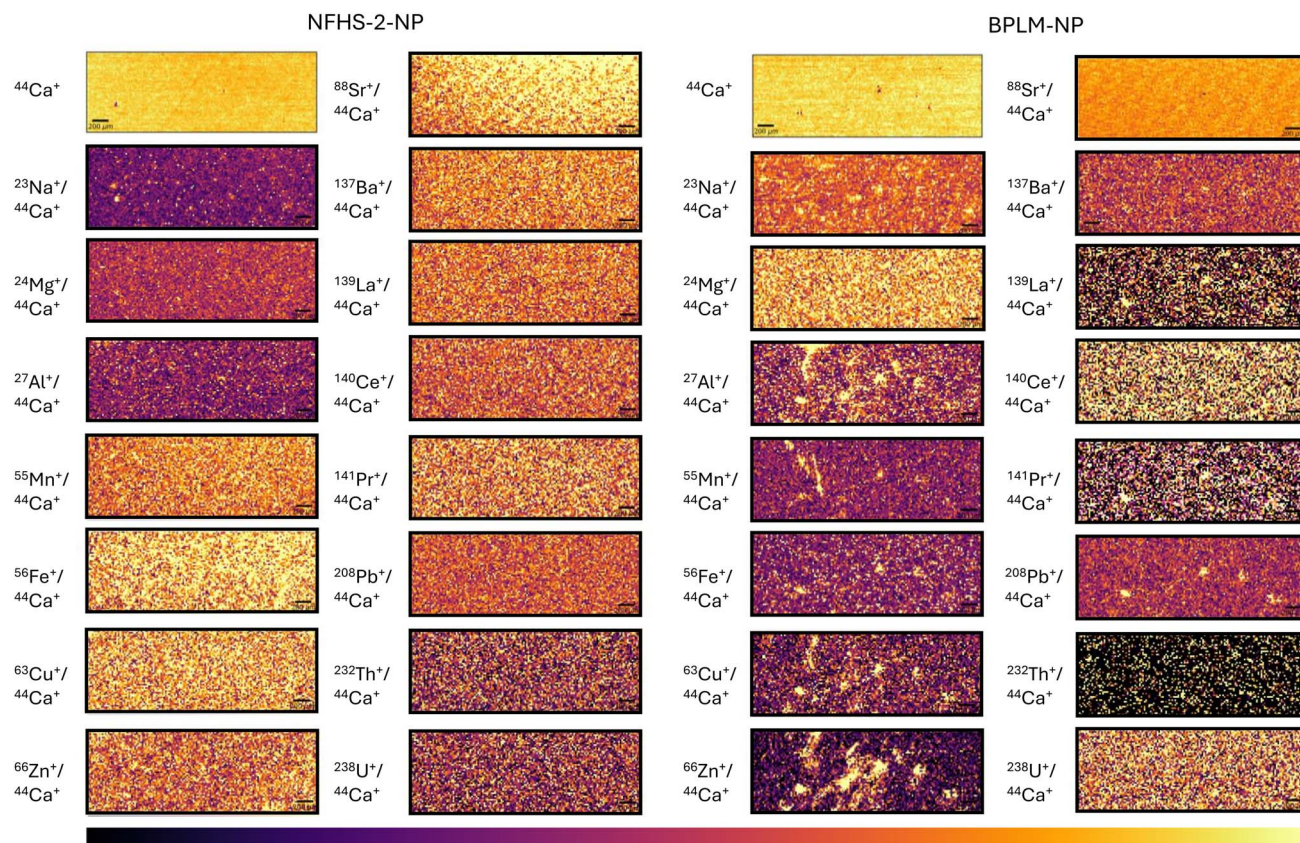


Fig. 3 Qualitative 2D elemental maps normalized to ^{44}Ca (3 mm $\times$ 1 mm, with 20 $\times$ 20 μm^2 laser beam) for ^{24}Mg , ^{27}Al , ^{55}Mn , ^{56}Fe , ^{63}Cu , ^{66}Zn , ^{88}Sr , ^{137}Ba , ^{139}La , ^{140}Ce , ^{141}Pr , ^{208}Pb , ^{232}Th and ^{238}U in NFHS-2-NP and BPLM-NP pressed powder pellets. The scale bar indicates 200 μm and the colour bar indicates the intensity based on the nuclide/ $^{44}\text{Ca}^+$ ratio from min to max (always from 0 to each max value).

while the average signal intensities may vary depending on the number of lines (n) selected—up to 20% for Zn in BPLM—most elements showed a variation $< 5\%$ across all nano-pellets (Table S1), supporting the high micro-homogeneity of the nano-pellets.

Higher uncertainties tend to correlate with lower elemental concentrations, especially for elements present at or below the $\mu\text{g g}^{-1}$ level. In Fig. 4, plotting the RSD% (based on 10-line measurements) against the element mass fraction ($\mu\text{g g}^{-1}$) across all nano-pellets reveals a general trend: elements at higher concentrations exhibit lower RSDs. However, this trend is not consistent across all materials. In the case of NFHS-2-NP, RSDs remained below 5% for elements above $1 \mu\text{g g}^{-1}$ with an increase to 20% for concentrations below the $\mu\text{g g}^{-1}$ level. However, in other nano-pellets, like CRMS-NP or Apatite-NP, elements around $5\text{--}100 \mu\text{g g}^{-1}$ displayed higher RSDs than some sub- $\mu\text{g g}^{-1}$ elements, suggesting that concentration alone does not fully explain the variability. Overall, except for elements at sub- $\mu\text{g g}^{-1}$ levels in BPLM-NP and BAM-RS3-NP (RSDs in the range of 50–100%), RSD values remained within acceptable limits—mostly below 30%, and in some pellets (e.g., NFHS-2-NP) under 10%. This result matches the RSDs found by Jochum *et al.*¹⁸ when evaluating the first ever nanoparticulate pellets (MACS-3NP, JcP-1NP and JcT-1-NP) using larger spot

Table 2 Calculated RSDs based on the average signal intensities of individual lines from the elemental 2D-maps for nano-pellets: NFHS-2-NP (top) and BPLM-NP (bottom). The average signal intensities and the corresponding RSDs are shown for sets of 50, 25, 10, 5, and 3 lines. Information for the entire range of target elements as well as for the rest of the nano-pellets is available in the SI (Table S1)

n	55Mn		56Fe		63Cu		66Zn		88Sr		137Ba		208Pb		238U	
	Mean (cts)	RSD	Mean (cts)	RSD	Mean (cts)	RSD	Mean (cts)	RSD	Mean (cts)	RSD	Mean (cts)	RSD	Mean (cts)	RSD	Mean (cts)	RSD
50	2.14×10^2	1%	6.83×10^1	2%	6.58×10^0	4%	6.68×10^0	7%	6.26×10^3	1%	1.11×10^2	2%	2.15×10^1	2%	2.35×10^0	8%
25	2.13×10^2	1%	6.82×10^1	2%	6.66×10^0	4%	6.55×10^0	4%	6.26×10^3	1%	1.12×10^2	2%	2.14×10^1	2%	2.35×10^0	9%
10	2.14×10^2	1%	6.86×10^1	2%	6.72×10^0	5%	6.55×10^0	5%	6.28×10^3	1%	1.12×10^2	2%	2.14×10^1	2%	2.41×10^0	13%
5	2.15×10^2	1%	6.90×10^1	1%	6.64×10^0	7%	6.64×10^0	7%	6.28×10^3	1%	1.13×10^2	3%	2.12×10^1	3%	2.36×10^0	7%
3	2.15×10^2	1%	$6.83 \times 10^1</$													

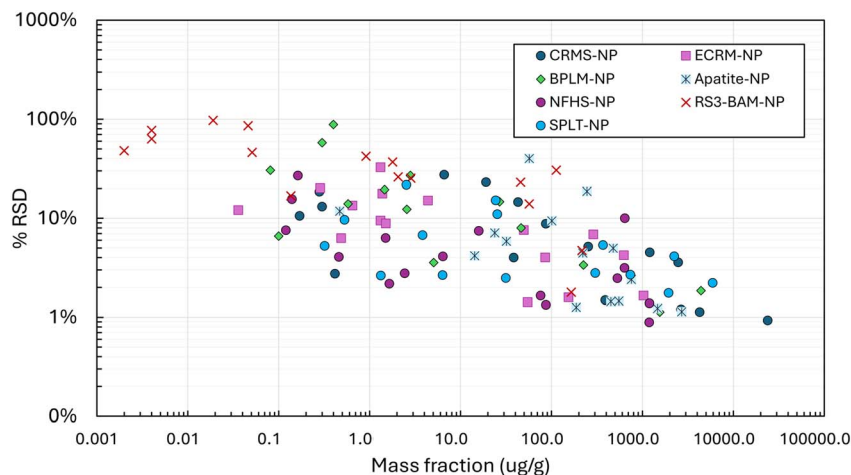


Fig. 4 RSD% based on 10 individual line scans performed for the entire set of nano-pellets (CRMS-NP, ECRM-752-1-NP, BPLM-NP, Apatite-NP, NFHS-2-NP, BAM-RS3-NP and SPLT-NP), plotted as a function of the mass fraction ($\mu\text{g g}^{-1}$) of the elements present.

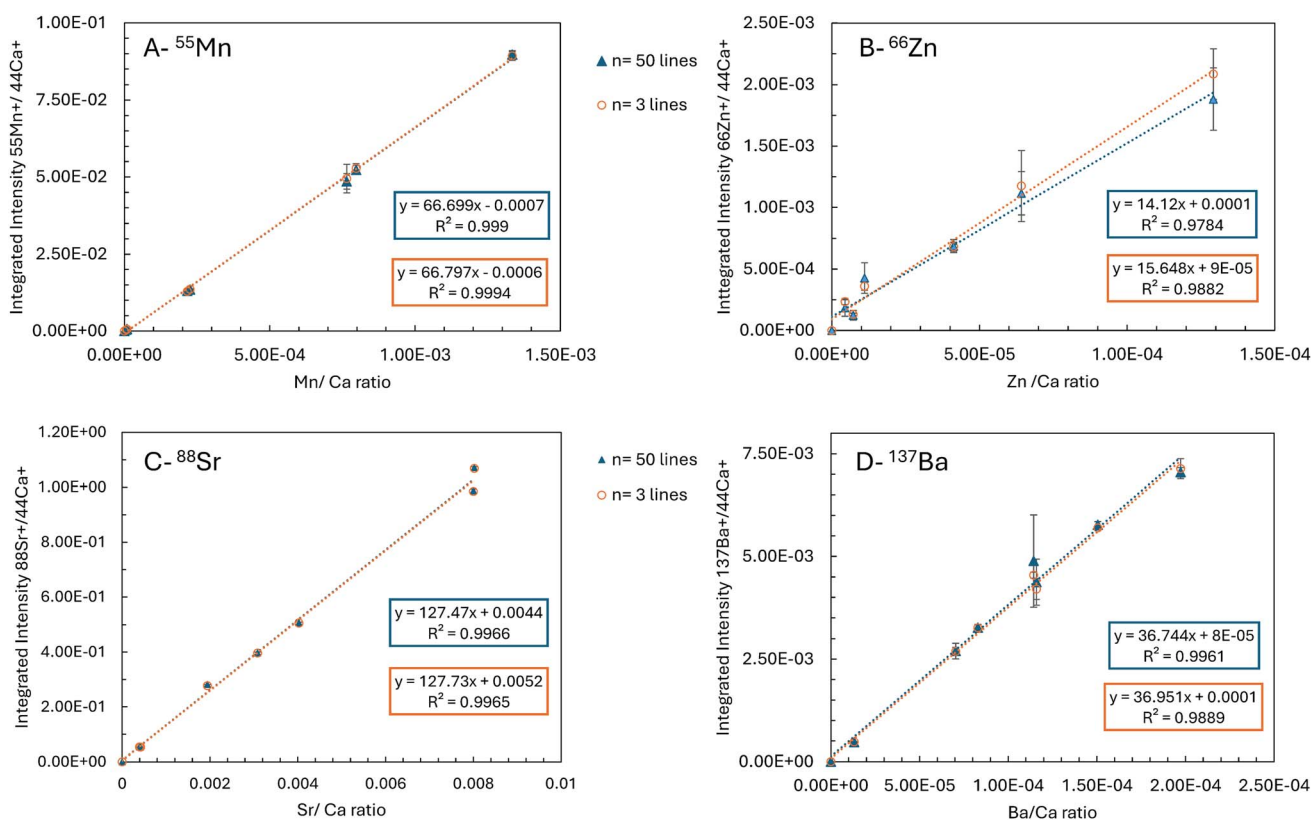


Fig. 5 Comparison of the linearity and slopes of calibration curves constructed based on the average signal intensities for 50 (triangles and blue boxes) and 3-line scans (circles and orange boxes) for (A) $^{55}\text{Mn}^+$, (B) $^{66}\text{Zn}^+$, (C) $^{88}\text{Sr}^+$, and (D) $^{137}\text{Ba}^+$, respectively, and normalized to $^{44}\text{Ca}^+$. Each point of the calibration curve represents the integrated intensity for a given nano-pellet, with the X/Ca ratios calculated based on the reference values. Error bars indicate the standard deviation (SD) of the average normalized signal intensities. In some cases, error bars are not visible due to the low SDs.

the linearity of all calibration curves, resulting in an average R^2 value of 0.9958 ± 0.0048 across all target elements. Also, small variations in laser beam focus during ablation could negatively affect the linearity which can be overcome when using internal standardization.

The recoveries obtained for the target elements in the remaining nano-pellets (Apatite-NP, SPLT-NP, BAM-RS3-NP, ECRM-752-1-NP and BPLM-NP) are shown in the SI (Table S2). The results highlight a clear relationship between the micro-homogeneity of nano-pellets and the accuracy of elemental



Table 3 Results for two of the nano-pellets investigated: CRMS-NP (upper pane) and NFHS-2-NP (lower pane). Summary of the calculated concentrations for the target elements by using different quantification approaches: (1) non-matrix-matched calibration against NIST SRM 610 with Ca as IS, (2) matrix-matched calibration using the full set of nano-pellets without IS, (3) matrix-matched calibration using the full set of nano-pellets with Ca as IS, and (4) matrix-matched calibration against a single nano-pellet standard (final column). NFHS-2-NP was used to quantify CRMS-NP, and vice versa. The propagated uncertainty is expressed as sd based on $n = 3$ lines used in the calibration, combined with the uncertainty from the certified concentration values (first column)

CRMS-NP	Reference value	Against NIST SRM 610/Ca (IS)				Against nano-pellets CAL				Against nano-pellets CAL/Ca (IS)				Against NFHS-2-NP/Ca (IS)						
		Conc. ($\mu\text{g g}^{-1}$)	sd	Conc. ($\mu\text{g g}^{-1}$)	RSD %	Recovery %	Conc. ($\mu\text{g g}^{-1}$)	sd	RSD %	Recovery %	Conc. ($\mu\text{g g}^{-1}$)	sd	RSD %	Recovery %	Conc. ($\mu\text{g g}^{-1}$)	sd	RSD %	Recovery %	LOD ($\mu\text{g g}^{-1}$)	LOQ ($\mu\text{g g}^{-1}$)
Na	4270	160	1398	16	1%	33%	55 780	63	1%	131%	4640	52	1%	109%	4920	230	5%	115%	55	180
Mg	23 940	690	21 570	200	1%	90%	25 050	230	1%	105%	23 980	220	1%	100%	26 540	920	3%	111%	1.7	5.8
Al	1207	38	441	20	5%	37%	2550	120	5%	211%	1140	52	5%	94%	1080	140	13%	89%		



Table 3 (Contd.)

NFHS-2-NP	Against NIST SRM 610/Ca (IS)				Against nano-pellets CAL				Against nano-pellets CAL/Ca (IS)				Against CRMS-NP			
	Certified value		Conc. ($\mu\text{g g}^{-1}$)		RSD %		Recovery %		Conc. ($\mu\text{g g}^{-1}$)		RSD %		Recovery %		Conc. ($\mu\text{g g}^{-1}$)	
	sd		sd		sd		sd		sd		sd		sd		sd	
Element	Conc. ($\mu\text{g g}^{-1}$)	sd	Conc. ($\mu\text{g g}^{-1}$)	sd	RSD %	Recovery %	Conc. ($\mu\text{g g}^{-1}$)	sd	RSD %	Recovery %	Conc. ($\mu\text{g g}^{-1}$)	sd	RSD %	Recovery %	Conc. ($\mu\text{g g}^{-1}$)	sd
Pr	0.460	0.040	0.340	0.014	4%	73%	0.480	0.014	3%	104%	0.452	0.013	3%	98%	0.410	0.050
Pb	1.650	0.050	1.370	0.030	2%	83%	1.030	0.027	3%	63%	1.600	0.042	3%	97%	1.624	0.030
Th	0.140	0.010	0.100	0.015	16%	69%	0.170	0.013	8%	123%	0.163	0.012	8%	116%	0.144	0.020
U	0.120	0.030	0.0660													

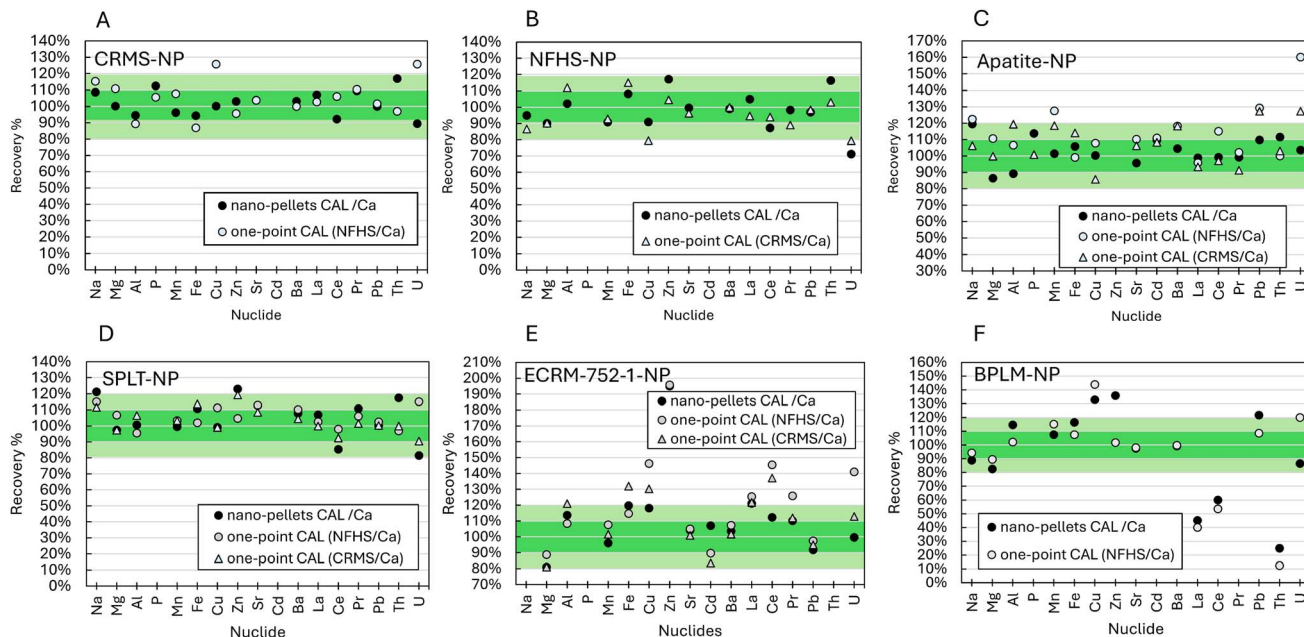


Fig. 6 Comparison of elemental recoveries – (determined value/reference value) $\times$ 100% – for target elements in different nano-pellets: (A) CRMS-NP, (B) NFHS-2-NP, (C) Apatite-NP, (D) SPLT-NP, (E) ECRM-752-1-NP, and (F) BPLM-NP. Recoveries are shown for calibrations performed using either the $N - 1$ nano-pellet method or one-point calibration (using NFHS-2-NP or CRMS-NP). Shading indicates deviation ranges from the reference value (dark and light green correspond to $\pm 10\%$ and $\pm 20\%$, respectively).

quantification (LOQs) in the $\mu\text{g g}^{-1}$ -range (Table S2), hindering the monitoring of trace elements (sub- $\mu\text{g g}^{-1}$ levels), especially in the low-mass range (due to the inherent lower sensitivity exhibited by TOF-based instrumentation). Nevertheless, Fig. 7 shows that for major and minor elements (above $\mu\text{g g}^{-1}$ levels) the bias from the reference value remained $<10\%$ (darker green) for most of the elements (*i.e.*, Na, Mg, Al, Mn, Fe, Sr and Ba), or $<20\%$ (light green) for some (*i.e.*, Cu and Pb) even when decreasing the spot size to $5 \times 5 \mu\text{m}^2$. These results confirm

that while smaller spot sizes reduce the overall sensitivity, especially critical when working with ICP-TOF-MS instrumentation, the micro-homogeneity of the nano-pellets is sufficient to provide reliable and accurate data for minor (particularly in the high-mass range) and major elements, though a full elemental overview is restricted under such conditions (*e.g.*, for Th or U, which were present at sub-ppm levels, the recoveries were less than 80% when using a 10×10 or $5 \times 5 \mu\text{m}^2$ laser beam).

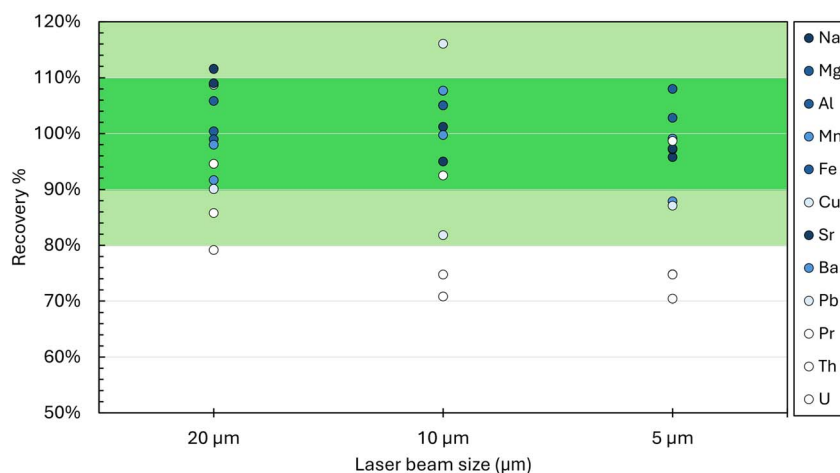


Fig. 7 Comparison of recoveries obtained using different laser beam sizes ($20 \mu\text{m} \times 20 \mu\text{m}$, $10 \mu\text{m} \times 10 \mu\text{m}$, and $5 \mu\text{m} \times 5 \mu\text{m}$) for major, minor, and trace elements in NFHS-2-NP. The intensity of the data point colour reflects the concentration levels: from highest (Na, Sr ≈ 1000 ppm) to sub-ppm levels; (Na, Sr) $>$ (Mg, Al, Fe) $>$ (Mn, Ba) $>$ (Cu, Pb) $>$ (Pt, Th, U).

Case study: quantitative multi-elemental mapping of CaCO_3 samples

In this section, quantitative multi-elemental mapping of two types of CaCO_3 -based biological samples—(1) fish otoliths and (2) bivalve shells—are presented. Data processing for these samples relied on the combined use of nano-pellets as matrix-matched standards for external calibration and Ca as an internal standard for correction of differences in the ablation yield. Both sample types are of particular interest in sclero-chronological studies, as archives for short-term climate reconstruction, for extracting (paleo)biological information, in the context of fisheries studies and for reconstructing environmental histories.^{46–48} To investigate the fine microstructural features of these materials, high spatial resolution is essential. While a detailed interpretation of the biological meaning of the data obtained is beyond the scope of this paper, relevant observations based on the case studies presented will be discussed briefly.

Otolith samples. Two otoliths from Atlantic cod (*Gadus morhua*), collected from two different waterbodies in Western Norway—Hardangerfjorden and Store Lungegårdsvannet (details provided in the Experimental section)—were quantitatively mapped using the LA-ICP-TOF-MS methodology developed. This method enabled quasi-simultaneous monitoring of nearly the entire elemental mass spectrum for every pixel, at a pixel acquisition rate of 50 pixels per second and with a lateral resolution of 20 μm (parameters summarized in Table 1). In this way, it was possible to distinguish growth rings, which exhibit considerable variation in thickness depending on the region of the otolith, being thinner near the core and thicker toward the edges. Also, when tracking along the vertical axis, the rings of the specimen appear more merged and were more difficult to distinguish individually compared to the horizontal view, as is displayed by the quantitative strontium (Sr) distribution shown in Fig. 8C.

The data processing workflow for generating quantitative elemental maps involved: (1) selection of the qualitative map of

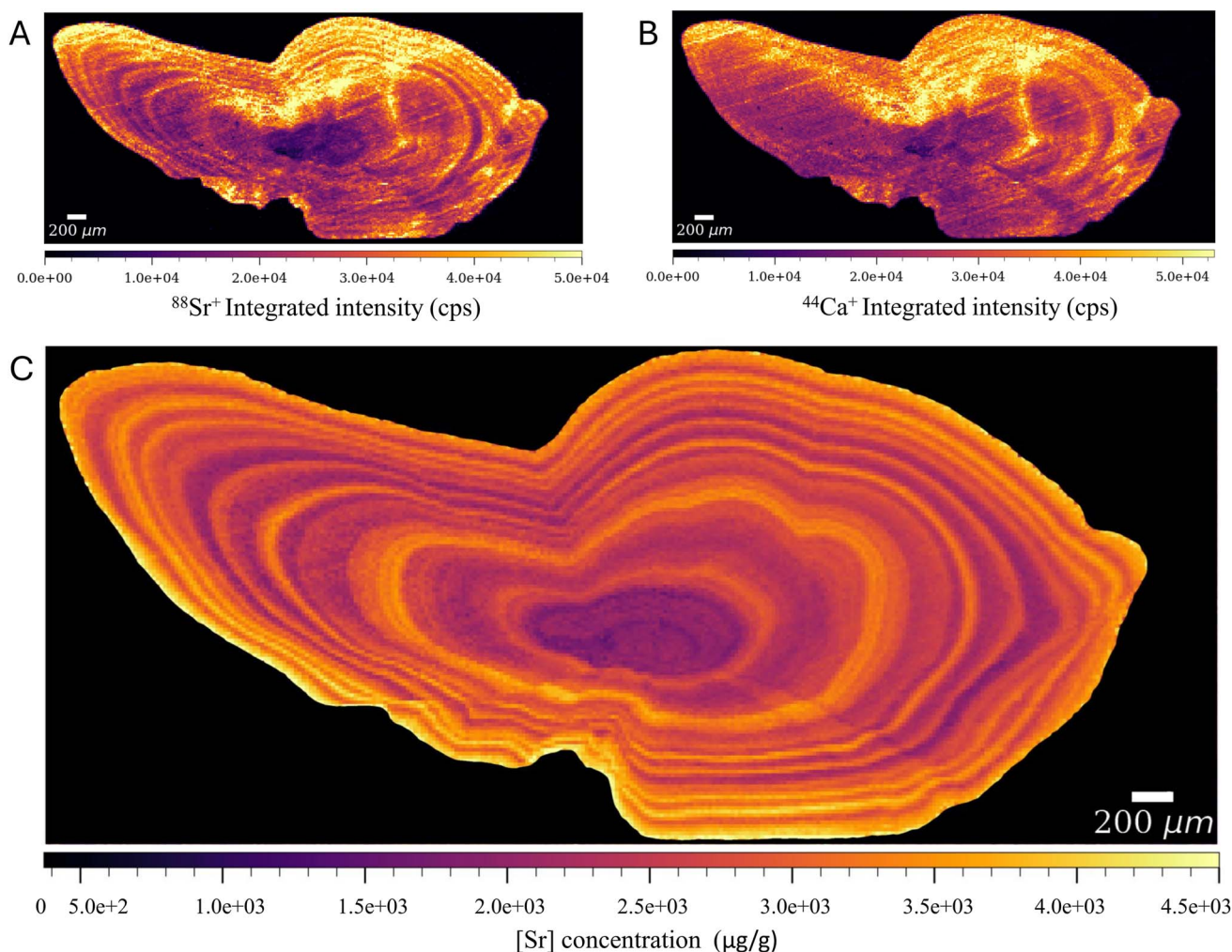


Fig. 8 LA-ICP-TOF-MS elemental maps obtained for the otolith from cod caught in the Store Lungegårdsvannet, Norway. (A) qualitative $^{88}\text{Sr}^+$ map, (B) qualitative $^{44}\text{Ca}^+$ map used for normalization and (C) quantitative map displaying the Sr distribution after Ca normalization and using the nano-pellets as external standards.

the nuclide of interest (*e.g.*, $^{88}\text{Sr}^+$) and of the internal standard ($^{44}\text{Ca}^+$) (Fig. 8A and B, respectively); (2) normalization of the target element's signal intensity at each pixel to that for $^{44}\text{Ca}^+$ to account for differences in ablation yield; (3) conversion of the normalized qualitative map into a quantitative one using matrix-matched external calibration against the nano-pellets ($n = 3$ lines per nano-pellet) and assuming a concentration of Ca of 40%.

Sr is typically used as a proxy for fish age. The distribution of Sr in the otolith displays concentric bands interpreted as chronological markers, as can be seen in the quantitative Sr map in Fig. 8C. However, it is also studied as a proxy for reconstructing environmental histories such as habitat shifts and waterbody changes.^{48,49} In fact, Sr incorporation in otoliths reflects the ambient water chemistry—mainly salinity and temperature variations.^{50,51} While these patterns may capture seasonal or episodic events, high lateral resolution mapping can reveal more Sr bands than actual annual growth increments. In this case, 15–17 Sr bands were identified in an otolith originating from a 6 year old fish, highlighting the enhanced time resolution offered by spatially resolved LA-ICP-TOF-MS mapping.² When examining the Mn distribution (Fig. S4A), it is still possible to distinguish an enrichment in the core regions corresponding to juvenile life (Mn concentration close to the LOQ of the method). Elevated Mn/Ca ratios were described to be related with hypoxic conditions (*i.e.*, reduced oxygen levels) typically in estuaries or sediment-rich areas.^{51–54} The combination of high Mn concentration with low Sr levels in the rings near to the core suggests a juvenile phase where the animal was exposed to low oxygen (high concentration of Mn) and low salinity (low concentration rings of Sr) water environments like hypoxic bottom waters or estuaries. Peaks in Mn can also

indicate metabolic activity or stress during key life events (*e.g.*, hatching, settlement).⁴⁸ The findings were compared to those for a second otolith from a cod sampled in the same waterbody (Lungegårdsvannet) that shows the same patterns: highly defined layers are observed based on the Sr distribution along the entire life of the fish, while Mn layers (higher Mn exposure) overlapping with low Sr concentration layers are only seen for early-life periods (Fig. S4B and C, respectively).

Notably, the diagonal lines—likely polishing artifacts—observed in the elemental maps of $^{88}\text{Sr}^+$ (Fig. 8A) and $^{44}\text{Ca}^+$ (Fig. 8B) are no longer visible after normalization (Fig. 8C). This demonstrates the effectiveness of IS normalization in correcting not only for matrix-related ablation yield variations (*e.g.*, due to crystallization or porosity) or instrumental drifts, but also for surface artifacts introduced during sample preparation.

For the other otolith from the cod caught in the Hardanger fjord in Norway, the Mn distribution was not revealed, but in the juvenile lines, an enrichment in Ba (Fig. 9B) coinciding with low-concentration rings of Sr (Fig. 9A) was observed. Ba is typically used as marker of specific habitat transitions.⁵⁰ While a high concentration of Sr is related to high salinity, a high concentration of Ba corresponds to low salinity environments (*i.e.*, fresh waters).⁴⁸ The co-localization of a low Sr and a high Ba concentration in the early growth rings suggests a juvenile phase where the cod inhabited shallower or more brackish environments (low salinity environments). A later migration to deeper, more saline waters (no Ba exposure) for foraging could explain the corresponding changes in element ratios recorded in the otolith. Taken together, these data underscore the value of quasi-simultaneous quantitative multi-elemental mapping in enhancing the interpretation of otolith chemistry covering

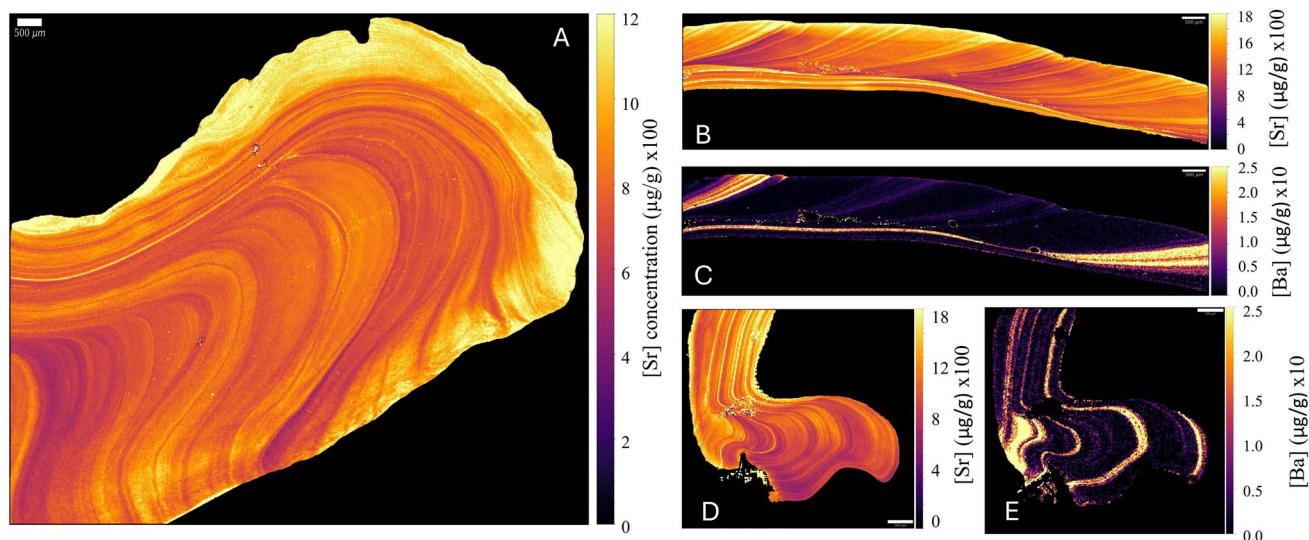


Fig. 10 Quantitative maps for Sr and Ba in two different bivalve shell types: (A) Sr distribution in oyster shell and (B) the Sr and (C) the Ba distribution in the upper part of the dorsoventral section along the axis of maximum growth and (D) the Sr and (E) the Ba distribution in the hinge area of the clam shell. The scale bar in all maps indicates 500 μm .

indicate chronological age, integrating multiple elemental ring distributions (*i.e.* Mn or Ba) enables a more nuanced reconstruction of the fish's environmental history. Noted that, fully interpreting the readings of each otolith and tracking the corresponding fish's life history requires extensive analysis, which is beyond the scope of this work.

Traditional 1D-analysis (line scanning) was compared to 2D-elemental mapping in all cases to emphasize the advantages of 2D-mapping in sclerochronological studies. Although a well-chosen 1D transect (*e.g.*, core-to-edge along the longest growth axis) can capture chronological information, it may not always represent the full spatial variability of elemental incorporation. Recent work by de Pontual *et al.*⁵⁵ demonstrated that otoliths can exhibit subtle asymmetries and heterogeneous distributions of certain elements, underscoring the limitations of relying solely on single transects. Our results are consistent with this view. As shown in Fig. 9B and C, the layer patterns for Sr and Ba differ depending on the direction of the line scan—whether from the core to the left (Fig. 9B), or from the core to the top (Fig. 9C) of the otolith. Notably, scans from the centre (juvenile layers) to the top show a loss of resolution, failing to capture the fine scale elemental distribution. In addition, as can be observed from the 2D-map (Fig. 9A and C), the layer arrangement on the left & right sides is not perfectly symmetrical. Consequently, 1D analysis can yield different patterns depending on scan orientation, potentially leading to misinterpretation of elemental (Fig. S5). By contrast, 2D-mapping provides a comprehensive view of the otolith structure, allowing potential spatial heterogeneities to be identified and interpreted more reliably.

To validate the quantitative results obtained *via* LA-ICP-TOF-MS mapping, a complementary bulk analysis was performed. From each fish, two otoliths were collected. One was embedded in epoxy resin for direct solid sampling by laser ablation, while

the second was acid-digested and used for validation through conventional solution ICP-MS, as described in the Experimental Section of the SI. The results were in the same range as the values obtained by solid sampling for Sr, Ba and Mn (Table S4).

Bivalve shells. Bivalve shells, which are common archives for short-term climate reconstructions, were mapped using the same method and workflow as used for otolith analysis. The quantitative distribution of target elements (Sr and Ba) was documented across different regions of oyster (*Crassostrea gigas*; Fig. 10A) and ocean quahog shell samples (*Arctica islandica*; Fig. 10A–E). The Sr/Ca and Ba/Ca ratios in *C. gigas* (~ 400 – $1200 \mu\text{g g}^{-1}$ for Sr/Ca and ~ 0 – $25 \mu\text{g g}^{-1}$ for Ba/Ca)^{56,57}

phytoplankton bloom events in spring and/or autumn, in accordance with observations in the literature.⁴⁴ Clear seasonal banding in Sr/Ca observed in *A. islandica* (Fig. 10C and E) corroborates the timing of these Ba/Ca peaks in this specimen. An important observation in these mollusc maps is that the concentration of Sr relative to Ca is not uniform in contemporary portions of the shell (see Fig. 10A for *C. gigas* and Fig. 10B for *A. islandica*). This corroborates previous studies by Freitas *et al.*⁶¹ who demonstrated such heterogeneity and like for the otoliths, this highlights the value of 2D-mapping for the interpretation of trace element profiles in mollusc shell material in terms of climate and environmental change.

Conclusions

The introduction of nanoparticulate pressed powder pellets has significantly enhanced the accuracy and precision attainable in quantitative LA-ICP-MS analysis of biogenic carbonates. Their higher micro-homogeneity owing to their smaller grain size permits higher reproducibility and accuracy in quantitative 2D-mapping at high spatial resolution. These findings underline the advantage of using matrix-matched nano-pellets. Unlike silicate glass reference materials such as NIST SRM 610, nano-pellets offer better matrix compatibility, resulting in lower quantification uncertainties. The use of Ca as an internal standard proved crucial not only for correcting for ablation yield differences between dissimilar matrices (*e.g.*, NIST glass *vs.* carbonate) but also for improving quantification among the different carbonate samples, which may exhibit slight but impactful matrix variations (*e.g.*, different crystallization or porosity) and for eliminating the effect of surface artifacts on the samples.

The micro-homogeneity assessments showed differences among set of the nano-pellets studied. While NFHS-2-NP, CRMS-NP, and Apatite-NP nano-pellets exhibited the highest micro-homogeneity for nearly all elements, higher RSDs and consequently poorer elemental recoveries were found for BAM-RS3-NP and BPLM-NP. It was noted that sub- $\mu\text{g g}^{-1}$ elemental concentrations in the nano-pellets yielded higher RSDs. Additionally, when decreasing laser beam sizes to reach very high lateral resolution (down to $5 \times 5 \mu\text{m}^2$), LODs and LOQs deteriorated, hampering detection and quantification of trace elements at concentrations below sub- $\mu\text{g g}^{-1}$, as well as increasing uncertainties for elements close to the LOD. This reduction in sensitivity associated with the use of a smaller laser beam spot and the inherently lower sensitivity of ICP-TOF-MS instrumentation compared to quadrupole-based systems jeopardize the detection of some minor and trace elements (often in the low $\mu\text{g g}^{-1}$ or even sub- $\mu\text{g g}^{-1}$ range). Nevertheless, the loss of sensitivity at ultrahigh lateral resolution could in principle be compensated for by increasing both the number of laser shots per pixel (*i.e.*, using a dosage >1) and the repetition rate accordingly. While this approach could boost signal intensity, it also exponentially increases the number of required laser pulses and introduces potential challenges with detector saturation. Therefore, dosage >1 approaches may be better suited for specific regions of interest (ROIs) where very high spatial detail is required.

Overall, this quantitative multi-elemental LA-ICP-TOF-MS 2D elemental mapping approach was successfully applied to two types of biogenic carbonate samples: otoliths and bivalve shells. This methodology enabled the quantitative visualization of growth bands and internal structures, allowing distinction of seasonal and sub-seasonal growth patterns. By combining Sr and Mn maps or Sr and Ba maps, the 2D reconstructions provide additional spatial context beyond that obtained from 1D transects, supporting a more robust interpretation of carbonate archives for tracking historical events within an animal's life-span. While Sr reflects seasonal-scale environmental variations in both archives, Mn and Ba serve as sensitive indicators of specific habitat conditions such as water oxygenation and phytoplankton blooms. Together, these tracers underline the utility of biogenic samples such as otoliths and bivalve shells as chronological environmental archives. Furthermore, comparison of 2D-maps with traditional 1D (line scanning) analyses revealed that, while 1D transects along the main growth axis can capture temporal Sr and Ba patterns, they may not consistently reflect finer-scale heterogeneities or asym

Supplementary information is available. See DOI: <https://doi.org/10.1039/d5ja00280j>.

Acknowledgements

A. L. P. and T. V. A. thank the Research Foundation Flanders for support under the form of their junior postdoctoral research fellowships (FWO 1276325N and 1218423N, respectively). N. J. W. acknowledges support from the Netherlands Research Council (NWO) for a personal VENI research fellowship (VI.Veni.222.354) and from the Flemish Research Council (FWO Science Prize Climate Research 2024). We would like to thank the following persons at the Institute of Marine Research, Norway for their contributions: Kai Kristoffer Lie for providing the otoliths of the Atlantic cod sampled in Hardanger, Åse Husebø for embedding and cutting the otoliths, and Erlend Langhelle for age determination of the Atlantic cod based on the otolith rings.

References

- 1 J. Vellekoop, D. Vanhove, I. Jelu, P. Claeys, L. C. Ivany, N. J. de Winter, R. P. Speijer and E. Steurbaut, *Palaeogeogr., Palaeoclimatol., Palaeoecol.*, 2025, **659**, 112627.
- 2 H. Tabouret, G. Bareille, F. Claverie, C. Pécuyer, P. Prouzet and O. F. X. Donard, *Mar. Environ. Res.*, 2010, **70**, 35–45.
- 3 D. Chew, K. Drost, J. H. Marsh and J. A. Petrus, *Chem. Geol.*, 2021, **559**, 119917.
- 4 Y. S. Liu, Z. C. Hu, M. Li and S. Gao, *Chin. Sci. Bull.*, 2013, **58**, 3863–3878.
- 5 A. Limbeck, P. Galler, M. Bonta, G. Bauer, W. Nischkauer and F. Vanhaecke, *Anal. Bioanal. Chem.*, 2015, **407**, 6593–6617.
- 6 S. Marali, B. R. Schöne, R. Mertz-Kraus, S. M. Griffin, A. D. Wanamaker, P. G. Butler, H. A. Holland and K. P. Jochum, *Palaeogeogr., Palaeoclimatol., Palaeoecol.*, 2017, **484**, 109–128.
- 7 H. A. O. Wang, D. Grolimund, C. Giesen, C. N. Borca, J. R. H. Shaw-Stewart, B. Bodenmiller and D. Günther, *Anal. Chem.*, 2013, **85**, 10107–10116.
- 8 S. J. M. Van Malderen, T. Van Acker and F. Vanhaecke, *Anal. Chem.*, 2020, **92**, 5756–5764.
- 9 T. Van Acker, S. J. M. Van Malderen, T. Van Helden, C. Stremtan, M. Šala, J. T. Van Elteren and F. Vanhaecke, *J. Anal. At. Spectrom.*, 2021, **36**, 1201–1209.
- 10 A. Gundlach-Graham and D. Günther, *Anal. Bioanal. Chem.*, 2016, **408**, 2687–2695.
- 11 I. Basabe-Mendizabal, R. Maeda, S. Goderis, F. Vanhaecke and T. Van Acker, *Anal. Chem.*, 2025, **97**, 6481–6488.
- 12 B. Hattendorf, C. Latkoczy and D. Günther, *Anal. Chem.*, 2003, **75**, 341–347.
- 13 K. P. Jochum, D. Scholz, B.

- Synchrotron X-Ray Elemental Distributions in Marine Bivalve Shells as Measured by Synchrotron X-Ray Fluorescence, *Biol. Bull.*, 1995, **188**, 57–67.
- 38 Y. Tamenori and T. Yoshimura, *Geochim. Cosmochim. Acta*, 2018, **237**, 357–369.
- 39 L. Hendriks, A. Gundlach-Graham, B. Hattendorf and D. Günther, *J. Anal. At. Spectrom.*, 2017, **32**, 548–561.
- 40 W. Boer, S. Nordstad, M. Weber, R. Mertz-Kraus, B. Hönlisch, J. Bijma, M. Raitzsch, D. Wilhelms-Dick, G. L. Foster, H. Goring-Harford, D. Nürnberg, F. Hauß, H. Kuhnert, F. Lugli, H. Spero, M. Rosner, P. van Gaever, L. J. de Noolijer and G. J. Reichart, *Geostand. Geoanal. Res.*, 2022, **46**, 411–432.
- 41 my-Standards, *myStandards-nano-pellets*, <https://www.my-standards.com/standards/carbonate-standards>, accessed 20 May 2025.
- 42 K. P. Jochum, U. Nohl, K. Herwig, E. Lammel, B. Stoll and A. W. Hofmann, *GeoReM: A New Geochemical Database for Reference Materials and Isotopic Standards*, <https://georem.mpch-mainz.gwdg.de/CiteGeorem.html>, accessed 24 June 2025.
- 43 D. Hippler, R. Witbaard, H. M. van Aken, D. Buhl and A. Immenhauser, *Palaeogeogr., Palaeoclimatol., Palaeoecol.*, 2013, **373**, 75–87.
- 44 N. J. de Winter, L. K. Dämmer, M. Falkenroth, G. J. Reichart, S. Moretti, A. Martínez-García, N. Höche, B. R. Schöne, K. Rodiouchkina, S. Goderis, F. Vanhaecke, S. M. van Leeuwen and M. Ziegler, *Geochim. Cosmochim. Acta*, 2021, **308**, 326–352.
- 45 M. Burger, L. Hendriks, J. Kaeslin, A. Gundlach-Graham, B. Hattendorf and D. Günther, *J. Anal. At. Spectrom.*, 2019, **34**, 135–146.
- 46 D. K. Moss, L. C. Ivany and D. S. Jones, *Paleobiology*, 2021, **47**, 551–573.
- 47 D. S. Jones, *Am. Sci.*, 1983, 384391.
- 48 P. Reis-Santos, B. M. Gillanders, A. M. Sturrock, C. Izzo, D. S. Oxman, J. A. Lueders-Dumont, K. Hüßy, S. E. Tanner, T. Rogers, Z. A. Doubleday, A. H. Andrews, C. Trueman, D. Brophy, J. D. Thiem, L. J. Baumgartner, M. Willmes, M. T. Chung, P. Charapata, R. C. Johnson, S. Trumble, Y. Heimbrand, K. E. Limburg and B. D. Walther, *Rev. Fish Biol. Fish.*, 2023, **33**, 411–449.
- 49 H. Tabouret, G. Bareille, F. Claverie, C. Pécheyran, P. Prouzet and O. F. X. Donard, *Mar. Environ. Res.*, 2010, **70**, 35–45.
- 50 C. Gauthier, J. A. D. Fisher, D. Robert and P. Sirois, *ICES J. Mar. Sci.*, 2024, **81**, 1221–1233.
- 51 P. D. Collingsworth, J. J. van Tassell, J. W. Olesik and E. A. Marschall, *Can. J. Fish. Aquat. Sci.</*