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Targeting of the mitochondrion by dinuclear thiolato-bridged arene ruthenium complexes in cancer cells and in the apicomplexan parasite *Neospora caninum*

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A library of 18 dinuclear-thiolato bridged arene ruthenium complexes, some of which with demonstrated activity against cancer cells, was screened for activity against a transgenic *Neospora caninum* strain that constitutively expresses beta-galactosidase. Initial assessments were done at concentrations of 2500, 250, 25 and 2.5 nM, and 5 compounds were further evaluated with regard to their half maximal proliferation-inhibiting concentration (IC₅₀). Among those, [(η⁶-*p*-MeC₆H₄Prⁱ)₂Ru₂(μ₂-SC₆H₄-*p*-CH₃)₃]Cl (**1**), [(η⁶-*p*-MeC₆H₄Prⁱ)₂Ru₂(μ₂-SC₆H₄-*p*-Bu^t)₃]Cl (**2**) and [(η⁶-*p*-MeC₆H₄Prⁱ)₂Ru₂(μ₂-SCH₂C₆H₄-*p*-Bu^t)₂(μ₂-SC₆H₄-*p*-OH)]BF₄ (**9**) inhibited *N. caninum* proliferation with low C₅₀ values of 15, 5 and 1 nM, respectively, while [(η⁶-*p*-MeC₆H₄Prⁱ)₂Ru₂(μ₂-SC₆H₄-*p*-OH)₃]Cl (**3**) and [(η⁶-*p*-MeC₆H₄Prⁱ)₂Ru₂(μ₂-SC₆H₄-*p*-mco)₃]Cl (**5**, mco = 4-methylcoumarinyl) were less active (IC₅₀ = 280 and 108 nM, respectively). These compounds did not affect human foreskin fibroblast (HFF) host cells at dosages of 5 μM and above, but impaired proliferation of the human ovarian carcinoma cell line A2780 (IC₅₀ values of 130 nM (**1**), 30 nM (**2**), 530 nM (**3**), 7730 nM (**5**), 130 nM (**9**)). A2780 cancer cells were treated with complexes **1**, **2**, and **5**, and biodistribution analysis using inductively coupled plasma mass spectrometry (ICP-MS) showed that most of the drugs accumulated in the mitochondrial fractions. Transmission electron microscopy showed that the parasite mitochondrion is the primary target also in *N. caninum* tachyzoites, but these compounds, when applied at 200 nM for 15 days *in vitro*, did not act parasitocidal. Complexes **1**, **2** and **9** applied orally at 2 and 10 mg kg⁻¹ day⁻¹ during 5 days in a neosporosis mouse model did not reduce parasite load and did not limit parasite dissemination to the central nervous system. In accordance with these results, ICP-MS carried out on different organs of mice orally administrated with complexes **1** and **9**, demonstrated that the drugs were readily absorbed, and after 3 and 48 h, were mainly detected in liver and kidney, but were largely absent from the brain. Thus, dinuclear thiolato-bridged arene ruthenium complexes exhibit interesting activities against *N. caninum in vitro*, but further modifications of these promising molecules are required to improve their bioavailability and pharmacokinetic properties in order to exert a pronounced and selective effect against *N. caninum in vivo*.

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Significance to metallomics

Ruthenium(II) complexes have attracted attention because of their favorable antiparasitic properties. Many different structural scaffolds are being investigated but their mode of action and targets remain largely unknown. Here, we report on the antiparasitic properties of dinuclear-thiolato bridged arene ruthenium complexes against *N. caninum*. We show that their activity and uptake can be correlated with their lipophilicity and we demonstrate their ability to selectively target the mitochondria of the parasite. *In vivo* experiments also reveal that the compounds accumulated in liver and kidneys, but were absent from the brain. However, the compounds did not act as parasitocidal.

Introduction

Cancer cells and apicomplexan parasites such as *Neospora caninum* and *Toxoplasma gondii* share a number of distinct biological features.¹ They both live and multiply within a host organism, and they do not immediately kill their hosts, but rather scavenge metabolites and nutrients to satisfy their own needs.^{2,3} Intracellular apicomplexans have evolved distinct mechanisms to interfere with the cell death machinery of their host cell, by affecting distinct signalling pathways.⁴ Similar to the rapidly replicating stages (tachyzoites) of these apicomplexans, cancer cells exhibit a seemingly unlimited proliferative potential. Tissue cyst-forming stages of these parasites (bradyzoites) and benign tumours survive within their hosts for extended periods of time, mostly without causing extensive pathology, and they achieve this goal by (i) disseminating into immune compromised tissues and (ii) exhibiting immunomodulatory properties that skew host immunity to their own favour.⁵

In contrast to the research on novel anti-cancer compounds, drug discovery in parasitology is a neglected field.⁶ While novel drugs have been developed against the classical neglected diseases such as malaria,⁷ trypanosomiasis^{8–10} and leishmaniasis,^{11,12} novel options for the treatment of protozoan diseases that affect farm animals are largely missing.¹³ Due to the intrinsic similarities between parasites and cancer cells, a reasonable starting point for the discovery of novel anti-parasitic drug candidates is to examine compounds that are being developed against cancer. While metal-based drugs are well-established as anti-cancer agents,^{14,15} the currently used drugs against protozoan parasites are organic compounds, and do not contain any metal atoms. However, based on different kinetic, geometric and electronic properties, metal complexes and organometallic complexes in particular can undergo reactions that are different from organic agents that could be used against parasites,^{16–18} as exemplified by the anti-malarial drug candidate Ferroquine.¹⁹ The application of platinum anti-tumour agents clearly demonstrates that metal agents can be developed into drugs with a high impact in therapy.²⁰ Potentially, the same could be done for targeting protozoan parasites.

Dinuclear thiolato-bridged arene ruthenium complexes $[(\eta^6\text{-}p\text{-MeC}_6\text{H}_4\text{Pr}^1)_2\text{Ru}_2(\mu_2\text{-SR})_3]^+$ and $[(\eta^6\text{-}p\text{-MeC}_6\text{H}_4\text{Pr}^1)_2\text{Ru}_2(\mu_2\text{-SR}^1)(\mu_2\text{-SR}^2)]^+$ were shown to exhibit high toxicity against several types of cancer cells *in vitro* (IC₅₀ values as low as 30 nM).^{21–25} *In vivo*, one of these complexes, $[(\eta^6\text{-}p\text{-MeC}_6\text{H}_4\text{Pr}^1)_2\text{Ru}_2(\mu_2\text{-SC}_6\text{H}_4\text{-}p\text{-Bu}^1)_3]\text{Cl}$ (**2**, DiRu-1), prolonged significantly the survival rate of mice experimentally inoculated with tumour cells.^{26,27} Some other mononuclear ruthenium complexes were shown to exhibit promising *in vitro* activities against *T. gondii* and *N. caninum*,^{28,29} and

we have recently reported on the activity of a panel of dinuclear thiolato-bridged arene ruthenium complexes against *T. gondii* tachyzoites.³⁰ Some of these compounds efficiently inhibited host cell invasion, and had an IC₅₀ in the low nanomolar range, low host cell toxicity, and exposure of parasites to these compounds led to distinct alterations in the parasite mitochondrion.³⁰

Neospora caninum is an apicomplexan, which belongs to the family Sarcocystidae. The parasite is distributed worldwide and infects a broad range of animals including cattle, sheep, goats, deer, horses and dogs.³¹ *Neospora caninum* is the causative agent of neosporosis, a serious disease associated with abortion, stillbirth and maternal infertility in cattle and neurological disorders in dogs. Dogs and other canids represent the definitive hosts of *N. caninum*, where sexual development takes place. Due to its close resemblance to *T. gondii*, which is of high medical as well as veterinary medical importance, *N. caninum* was recurrently misdiagnosed as such until Dubey and colleagues succeeded in isolating the parasite in the late 1980ies.³² In contrast to *T. gondii*, *N. caninum* has never been shown to be transmitted from animals to humans.³¹ The prevalence of *N. caninum* varies considerably in different regions, however, the annual economic impact of neosporosis, based on data of 10 countries, was calculated to be around 1.28 billion US dollars.³³

Vaccination and chemotherapy have been identified as economically promising options for control of neosporosis, provided suitable targets and effective reagents can be made available.^{34,35} Although considerable efforts have been undertaken to develop vaccines,^{31,36} success with recombinant vaccines has been limited,^{37–39} and only live vaccines using attenuated *N. caninum* strains have shown promising efficacy so far.⁴⁰ As an alternative approach, treatment with anti-parasitic drugs could be envisaged, carried out either during pregnancy or right after birth, with the goal to eliminate the parasite before it can cause extensive damage to the next generation.

We here comparatively assessed the effects of a series of 18 thiolato-bridged dinuclear arene ruthenium complexes, consisting of 7 cationic trithiolato complexes $[(\eta^6\text{-}p\text{-MeC}_6\text{H}_4\text{Pr}^1)_2\text{Ru}_2(\mu_2\text{-SC}_6\text{H}_4\text{-}R)_3]\text{Cl}$ (**1–7**), 6 cationic mixed trithiolato complexes $[(\eta^6\text{-}p\text{-MeC}_6\text{H}_4\text{Pr}^1)_2\text{Ru}_2(\mu_2\text{-SCH}_2\text{C}_6\text{H}_4\text{-}R)(\mu_2\text{-SC}_6\text{H}_4\text{-}R)]\text{BF}_4$ (**8–13**), 4 neutral dithiolato complexes $[(\eta^6\text{-}p\text{-MeC}_6\text{H}_4\text{Pr}^1)_2\text{Ru}_2(\mu_2\text{-SCH}_2\text{C}_6\text{H}_4\text{-}p\text{-}R)_2\text{Cl}_2]$ (**14–17**), and one neutral monothiolato complex $[(\eta^6\text{-}p\text{-MeC}_6\text{H}_4\text{Pr}^1)_2\text{Ru}_2(\mu\text{-Cl})(\mu_2\text{-SCH}_2\text{C}_6\text{H}_4\text{-}p\text{-NO}_2)]$ (**18**) in the human ovarian cancer cell line A2780 and in tachyzoites of the intracellular apicomplexan *N. caninum* *in vitro*, and confirm that the mitochondrion is the primary target in both. In addition, we report on the limited efficacy of $[(\eta^6\text{-}p\text{-MeC}_6\text{H}_4\text{Pr}^1)_2\text{Ru}_2(\mu_2\text{-SC}_6\text{H}_4\text{-}p\text{-CH}_3)_3]\text{Cl}$ (**1**), $[(\eta^6\text{-}p\text{-MeC}_6\text{H}_4\text{Pr}^1)_2\text{Ru}_2(\mu_2\text{-SC}_6\text{H}_4\text{-}p\text{-Bu}^1)_3]\text{Cl}$ (**2**)



and $[(\eta^6\text{-}p\text{-MeC}_6\text{H}_4\text{Pr}^i)_2\text{Ru}_2(\mu_2\text{-SCH}_2\text{C}_6\text{H}_4\text{-}p\text{-Bu}^t)_2(\mu_2\text{-SC}_6\text{H}_4\text{-}p\text{-OH})]\text{BF}_4$ (**9**) in an experimental neosporosis mouse model, and studied the biodistribution of complex **1** in different organs of mice.

Materials and methods

Chemicals and synthesis of ruthenium complexes.

All reagents for chemical synthesis were commercially available and were used as received. Complexes **1–18** and their *in vitro* efficacy against *T. gondii* tachyzoites were previously reported.³⁰ All complexes were isolated as chloride or tetrafluoroborate salts, were stable orange to red solids, and were dried in vacuum. The analytical data matched those previously reported in the literature.^{21–23,41}

Culture of mammalian cells and parasite cultures

If not stated otherwise, all tissue culture media were purchased from Gibco-BRL (Zurich, Switzerland), and biochemical reagents were from Sigma (St. Louis, MO). Human foreskin fibroblasts (HFF) and Vero cells (green monkey kidney epithelial cells) were maintained in RPMI-medium containing 10% fetal calf serum (FCS) (Gibco-BRL, Zürich, Switzerland) and antibiotics as described earlier.³⁷ The human ovarian carcinoma A2780 cell line was cultured in RPMI 1640 medium (Gibco) supplemented with 10% FCS, 100 U mL⁻¹ penicillin, 100 µg mL⁻¹ streptomycin at 37 °C and 6% CO₂. *N. caninum* beta-gal (transgenic *N. caninum* Nc-1) expressing the beta-galactosidase gene from *E. coli*,⁴² and the *N. caninum* Spain7 isolate (Nc-Spain7) were maintained in Vero cells, and were isolated and separated from their host cells as described.⁴³

In vitro assessment of drug efficacy against *N. caninum* tachyzoites

To study the effects of compounds against *N. caninum*-beta-Gal tachyzoites *in vitro*, 0.5 mM stock solutions of the complexes were prepared in water, sterile filtered, and stored at 4 °C. The stability of the complexes in water solutions and in mixtures water/DMSO has been previously established.^{24,44,45} Drug assays were performed using HFF as host cells.²⁹ In short, 5×10^3 HFF per well were grown to confluence in a 96 well plate in phenol-red free culture medium at 37 °C with 5% CO₂. Cultures were infected with freshly isolated *N. caninum* beta-gal tachyzoites (1×10^3 per well) and drugs were added at the time point of infection. Initial assessments were done by exposing

parasite cultures to 2500 nM, 250 nM, 25 nM or 2.5 nM of each compound for a period of three days, or water was added as a control. For IC₅₀ determinations, 5 selected complexes (**1–3**, **5** and **9**; see Table 1) were added at concentrations ranging between 0 and 2000 nM. After three days at 37 °C/5% CO₂, plates were centrifuged at $500 \times g$, medium was removed, and cell cultures were exposed to PBS containing 0.05% Triton-X-100. After addition of 10 µL of 5 mM chlorophenol red-β-D-galactopyranoside (CPRG; Roche Diagnostics, Rotkreuz, Switzerland) dissolved in PBS, the absorption shift was measured at 570 nm wavelength at various time points on a VersaMax multiplate reader (Bucher Biotec, Basel, Switzerland). The activity, measured as the release of chlorophenol red over time, was proportional to the number of live parasites down to 50 per well as determined in pilot assays. IC₅₀ values were calculated after the logit-log-transformation of relative growth and subsequent regression analysis by the corresponding software tool contained in the Excel software package (Microsoft, Seattle, WA).

For long term treatment assays, *N. caninum* infected HFF grown in T25 culture flasks were exposed to 250 nM of complexes **1**, **2** or **9** for a period of 15 days, with medium changes and addition of fresh compounds every 3 days. At the end, the cultures were washed with medium and were further maintained in medium devoid of drugs. Regrowth of parasites was monitored on a daily basis by light microscopy.

Measurements of cytotoxicity

Cytotoxicity assays on non-infected confluent HFF were performed by Alamarblue assay as previously described.³⁰ The assessments of selected compounds on A2780 cells have been described earlier.^{21–23,41}

Transmission electron microscopy (TEM)

HFF (5×10^4 per inoculum) cultured in T25 tissue culture flasks for 24 h were infected with 10^5 *N. caninum* beta-gal tachyzoites, and 200 nM of complexes **1**, **2** or **9** were added at 24 h post-infection. After 6, 24 and 48 h, cells were harvested using a cell scraper, and they were placed into the primary fixation solution (2.5% glutaraldehyde in 100 mM sodium cacodylate buffer pH 7.3) for 2 h. Specimens were then washed 2 times in cacodylate buffer and were post-fixed in 2% OsO₄ in cacodylate buffer for 2 h, followed by washing in water, pre-staining in saturated uranyl acetate solution, and step wise dehydration in ethanol. They were then embedded in Epon 812-resin, and processed for

Table 1 log *P* values, Hammett constants σ_p , and efficacies of dinuclear thiolato-bridged arene ruthenium complexes against *N. caninum* tachyzoites, and for comparative purposes, of *T. gondii* tachyzoites, human foreskin fibroblasts (HFF) and the human ovarian carcinoma cell line A2780

Complex ^a	<i>N. caninum</i> beta-gal IC ₅₀ (nM)	<i>T. gondii</i> beta-gal IC ₅₀ ^b (nM)	HFF IC ₅₀ ^b (µM)	A2780 IC ₅₀ ^c (nM)	log <i>P</i> (RSH) ^c	$\sigma_p(R)^{c,d}$
1	15 ± 1	34 ± 4	800	130 ± 10	2.98 ± 0.28	−0.17
2	5 ± 1	62 ± 10	> 1000	30 ± 10	4.21 ± 0.29	−0.20
3	280 ± 50	540 ± 60	ND	530 ± 20	1.68 ± 0.29	−0.37
5	108 ± 19	120 ± 20	ND	7730 ± 1500	2.83 ± 0.42	ND
9	1 ± 0.3	1.2 ± 0.5	5	130 ± 3	3.51 ± 0.32	−0.20; −0.37

^a Chloride salts of complexes **1–3** and **5** or tetrafluoroborate salts of complex **9** were used for all experiments. ^b Data from ref. 30. ^c Data from ref. 21–23. ^d For complex **9**, two Hammett constants σ_p are given, one for each group R¹ and R².



TEM as described.^{30,46} Specimens were viewed on a Phillips CM12 transmission electron microscope operating at 60 kV.

Studies on drug uptake in human carcinoma A2780 cells

A2780 cells were maintained in RPMI 1640 medium (Gibco) supplemented with 10% fetal calf serum (FCS, Gibco) 100 U mL⁻¹ penicillin, 100 µg mL⁻¹ streptomycin at 37 °C and 6% CO₂. Seven days prior to treatments, 1 × 10⁶ cells were seeded in 175 cm² cell culture flask and grown until 80% of confluency, and were incubated with the target complexes (previously dissolved in 0.1% DMSO at a concentration of 1.0 µM for 4 h). Working concentration and incubation time were chosen in order to avoid extended cell mass lost due to the high cytotoxicity of the complexes and Ru amount that could afford a significant determination. After this, the cultures were washed in PBS, trypsinized, and after centrifugation (500 g for 5 min at 4 °C) and re-suspension in PBS, the pellet was collected. Mitochondria were isolated using a mitochondria isolation kit (Cat. Nr.: MITISO2, Sigma Aldrich) following the producer's instructions. Briefly, the collected pellets were re-dissolved in 1.5 mL of extraction buffer (delivered with the kit) and were incubated for 15 min on ice. The samples were homogenized with a pre-chilled dounce homogenizer (7 mL, tight pestle A, 30 strokes) and centrifuged at 600g for 10 min at 4 °C. The supernatant was transferred in a fresh tube and centrifuged at 11 000g for 10 min at 4 °C. The obtained pellets represented mitochondrial fractions. Nuclei of A2780 cells were obtained following an established procedure with minor modifications.⁴⁷ All the fractions were isolated from the same cellular sample as the mitochondrial fraction for direct comparative purposes. After homogenization, the pellet obtained was redissolved in 2 mL of a sucrose solution (0.25 M sucrose, 10 mM MgCl₂) and layered with 2 mL of a second hypertonic sucrose solution (0.35 M sucrose, 0.5 mM MgCl₂). The suspension was centrifuged at 1450g and 4 °C for 5 min. The pellet was re-suspended in 3 mL of the second sucrose solution and centrifuged at 1450g and 4 °C for 5 min to obtain the pure nuclear extract. All the steps of the isolation procedure were monitored by phase contrast microscopy. The supernatant phases discarded during the isolation of nuclei and mitochondria procedures were collected and formed the "residual" fraction. An aliquot of crude lysate supernatant, nuclear, mitochondrial and residual fraction was each used for protein quantification using the Bradford method.⁴⁸ The isolated samples were then lyophilized on an Alpha 2-4 LD plus (CHRIST) and stored at 4 °C. The distribution of ruthenium in different subcellular fractions was quantified by inductively coupled plasma mass spectrometry (ICP-MS) as described below.

Inductively coupled plasma mass spectrometry (ICP-MS)

ICP-MS measurements were performed on an Agilent QQQ 8800 Triple quad ICP-MS spectrometer (Agilent Technologies) with an ASX200 autosampler (Agilent Technologies), equipped with standard nickel cones and a "micro-mist" quartz nebulizer fed with 0.3 mL min⁻¹ analytic flow (as a 2% HNO₃ aqueous solution). Ruthenium was measured against a Ru single element standard (Merck 170347) and verified by a control (Agilent5188-6524 PA Tuning 2). Ruthenium content of the samples was

determined by means of a 7-step serial dilution in the range between 0 and 100 ppb in Ru ($R > 0.99$) with a background equivalent concentration of BEC: 3.3 ppt and a detection limit of DL: 5.4 ppt. The isotope Ru⁹⁹ (12.76% abundance) ¹⁰¹Ru (17.06% abundance) was evaluated in "no-gas" mode and He-gas mode. Spiking the samples with 1% methanol (to account for eventual carbon content from the biological samples) resulted in equivalent values within error ranges. A solution of Indium (500 ppb) and Tungsten (500 ppb) was used as internal standard. The results are expressed as ng Ru per mg protein (correction due to the different mass of the observed cellular compartments or organs), as mean ± standard deviation error of different independent experiments.

Treatment of *N. caninum*-infected mice with complexes 1, 2 and 9

Infection and treatment experiments were approved by the Animal Welfare Committee of the Canton of Bern under the license BE115/14. All animals were handled in strict accordance with practices made to minimize suffering. Balb/c mice each, which were purchased from Charles River Laboratories (Sulzfeld, Germany) and maintained under conventional day/night cycle housing conditions with water and food *ad libitum*. The mice were used for the experiment after two weeks of acclimatization. Mice were separated into experimental groups of 6 mice each, and were infected subcutaneously with 10⁵ cell culture-derived tachyzoites of the Nc-Spain7 strain.⁴³ Prior to the start of treatments, complexes were emulsified in corn oil by vortexing and ultrasound bath at 37 °C. At 48 h post infection, treatments were initiated. For each compound, one group received 2 mg of complexes 1, 2 or 9 per kg bodyweight per day for 5 consecutive days, the other group was given 10 mg of complexes 1, 2 or 9 per kg bodyweight per day, and a third group received corn oil only (placebo). All treatments were applied in 100 µL by oral gavage. Following infection, all mice were closely monitored for clinical signs by using a standardized score sheet. Mice were inspected daily with regard to their coat condition (ruffled coat = score 1; starry stiff coat = score 2), weight loss (10% loss = score 1; 15–20% weight loss = score 4), and behaviour (hunched appearance, walking in circles, head tilt, apathy and ataxia, all = score 1). Animals were euthanized when the score exceeded 3 points. The experiment was terminated latest at 21 days post-infection. Lungs and brain were collected to determine parasite loads by real-time PCR as previously described,⁴³ and cerebral and lung parasite burdens were compared between groups by the non-parametric Kruskal-Wallis test.

Uptake and biodistribution of complex 1 in mice

Complex 1 was emulsified in corn oil by vortexing and ultrasound bath at 37 °C. Six Balb/c mice received the drug by oral gavage of 100 µL at a dose of 10 mg kg⁻¹ of body weight and two control mice received corn oil only. One control and three treated mice were euthanized at 3 h and at 48 h post-administration. Whole blood, liver, kidney, lungs and brain were collected, lyophilized and the weights of the lyophilized samples were determined. The distribution of ruthenium in the different organs was quantified by ICP-MS as described above.



Results

In vitro activities of arene ruthenium complexes against *N. caninum* tachyzoites

A set of 18 ruthenium complexes (see Fig. 1) were screened for their potential to inhibit *N. caninum* tachyzoite proliferation in HFF monolayers. The symmetrical Ru(II) trithiolato complexes $[(\eta^6\text{-}p\text{-MeC}_6\text{H}_4\text{Pr}^i)_2\text{Ru}_2(\mu_2\text{-SC}_6\text{H}_4\text{-}p\text{-CH}_3)_3]\text{Cl}$ (1), $[(\eta^6\text{-}p\text{-MeC}_6\text{H}_4\text{Pr}^i)_2\text{Ru}_2(\mu_2\text{-SC}_6\text{H}_4\text{-}p\text{-Bu}^t)_3]\text{Cl}$ (2), $[(\eta^6\text{-}p\text{-MeC}_6\text{H}_4\text{Pr}^i)_2\text{Ru}_2(\mu_2\text{-SC}_6\text{H}_4\text{-}p\text{-OH})_3]\text{Cl}$ (3) and $[(\eta^6\text{-}p\text{-MeC}_6\text{H}_4\text{Pr}^i)_2\text{Ru}_2(\mu_2\text{-SC}_6\text{H}_4\text{-}p\text{-mco})_3]\text{Cl}$ (5, mco = 4-methylcoumarinyl), bearing one type of thiol ligand^{21,23} and the “mixed” Ru(II) trithiolato complex $[(\eta^6\text{-}p\text{-MeC}_6\text{H}_4\text{Pr}^i)_2\text{Ru}_2(\mu_2\text{-SCH}_2\text{C}_6\text{H}_4\text{-}p\text{-Bu}^t)_2(\mu_2\text{-SC}_6\text{H}_4\text{-}p\text{-OH})]\text{BF}_4$ (9), bearing two different thiol ligands²² inhibited the proliferation of *N. caninum* tachyzoites *in vitro* with IC₅₀ values between 1 and 280 nM, complex 9 being the most efficient (1 ± 0.3 nM) and complex 3 (280 ± 50 nM) the least active (Table 1). The trithiolato complexes 4, 6 and 7, the mixed complexes 8 and 10–13, the dithiolato complexes 14–17, and the monothiolato complex 18 had no measurable anti-parasitic activity or were toxic for HFF already at concentrations of 250 nM or 2500 nM. Overall, these compounds exhibited similar activities against *N. caninum* as for *T. gondii* tachyzoites, with the exception of complex 2, which was 10 times more active against *N. caninum* than against *T. gondii* (Table 1). The activity of the compounds against *N. caninum* parallels to a certain extent the results previously found against several cancer cell lines: the IC₅₀ values of 7 were two orders of magnitude larger than that of the other complexes, and the mono- and dithiolato complexes were found to be only moderately cytotoxic against cancer cell lines (IC₅₀ values between 0.2 and 2.5 μM).

In contrast, HFF used here as host cells for intracellular proliferation of *N. caninum* tachyzoites, appeared largely unaffected at several thousand-fold higher concentrations [selective toxicity index: > 50 000 for complex 1, > 160 000 for complex 2, and > 5000 for complex 9 (Table 1)]. These compounds did not act parasiticidal: long-term drug treatments of infected HFF with complexes 1, 2 and 9 at 250 nM during a period of up to 15 days did not eliminate all parasites in a culture, since regrowth of tachyzoites was observed 5–10 days after releasing drug pressure in all three instances.

Inductively coupled plasma mass spectrometry (ICP-MS) shows that ruthenium complexes target the mitochondrion in A2780 cancer cells

The presence of the non-physiological metal ruthenium in dinuclear thiolato-bridged arene ruthenium complexes allows for the use of ICP-MS to detect and quantify the uptake of the drug candidates into cells and different subcellular fractions.^{49,50} Complexes 1, 2 and 5 were chosen for these studies, 1 and 2 being active and selective, and complex 5 representing a less active compound. Subcellular fractionation was carried out using treated A2780 cancer cells. Strikingly, already after a relative short incubation time of 4 h, the large majority of the ruthenium taken up by these cells was found to accumulate in the mitochondrial fraction (*ca.* 90, 97 and 71% for complexes 1, 2 and 5, respectively) (see Fig. 2).

Also, the total values for ruthenium were much higher in fractions originating from complex 2 treated cells compared to the others, indicating that complex 2 is taken up much more efficiently, which perfectly parallels with the respective lipophilicity of the three complexes (Table 1 & Fig. 2).

Transmission electron microscopy (TEM) identifies the mitochondrion as the primary target of ruthenium complexes in *N. caninum* tachyzoites

Due to the lack of reliable subcellular fractionation procedures for *N. caninum*, the effects of ruthenium complexes in this parasite were studied by TEM (Fig. 3 and 4). In cultures maintained in the absence of ruthenium complexes, tachyzoites were located intracellularly and proliferated within a parasitophorous vacuole (PV), surrounded by a distinct PV membrane. Tachyzoites exhibited their typical structural features such as rhoptries, dense granules and micronemes, and the parasite mitochondria filled with a structured electron dense matrix and cristae could be readily identified (Fig. 3A and B). In drug-treated cultures, changes evolved rapidly, and were most pronounced within the mitochondrial matrix, which appeared less electron dense in some parasites already after 6 h of treatment with complexes 1 and 2 (Fig. 3C and D) and also complex 9 (Fig. 4A). The other organelles appeared non-affected. After 24 h, as exemplified here for complex 9, the changes became more evident, and the mitochondrion with its characteristic cristae and electron dense matrix was replaced by large vacuoles, which were either seemingly empty (Fig. 4B), or filled with tightly spaced membrane stacks resembling autophagosomes (Fig. 4C). However, the outer membrane of the mitochondria was still intact, and parasites maintained their overall shape and structural features. From 72 h onwards (Fig. 4D), the majority of parasitophorous vacuoles contained largely lysed or distorted parasites, with only few of the typical organelles still recognizable (Fig. 4D). These changes were similar upon treatments with all three complexes 1, 2 and 9. However, the host cell mitochondria were still structurally intact and associated with the PV membrane (Fig. 4D).

Effects of ruthenium complexes 1, 2 and 9 in the neosporosis mouse model

For assessment of *in vivo* anti-parasitic effects of these ruthenium complexes, initial studies were carried out on complexes 1 and 2. Balb/c mice were infected subcutaneously with 10⁵ NcSpain-7 tachyzoites, a dose that does not induce severe clinical signs in non-pregnant mice,⁴³ and after 48 h, daily treatments by gavage (either 2 mg kg⁻¹ or 10 mg kg⁻¹ of drugs emulsified in corn oil for 5 days) were initiated. Parasite loads in lungs and brain were determined by real-time PCR at 21 dpi (Fig. 5). In the placebo group (infected, but treated with corn oil only), two mice showed symptoms compatible to neosporosis, such as ruffled coat and body tilting, but they did not have to be euthanized as the clinical score was below the threshold. In the group treated with complexes 1 at 2 mg kg⁻¹ day⁻¹, one mouse exhibited a ruffled coat and survived, whereas in the group treated with 10 mg kg⁻¹ days⁻¹, 2 out of 6 mice had to be euthanized on days 14 and 17 dpi. In the group treated with complex 2, one mouse treated



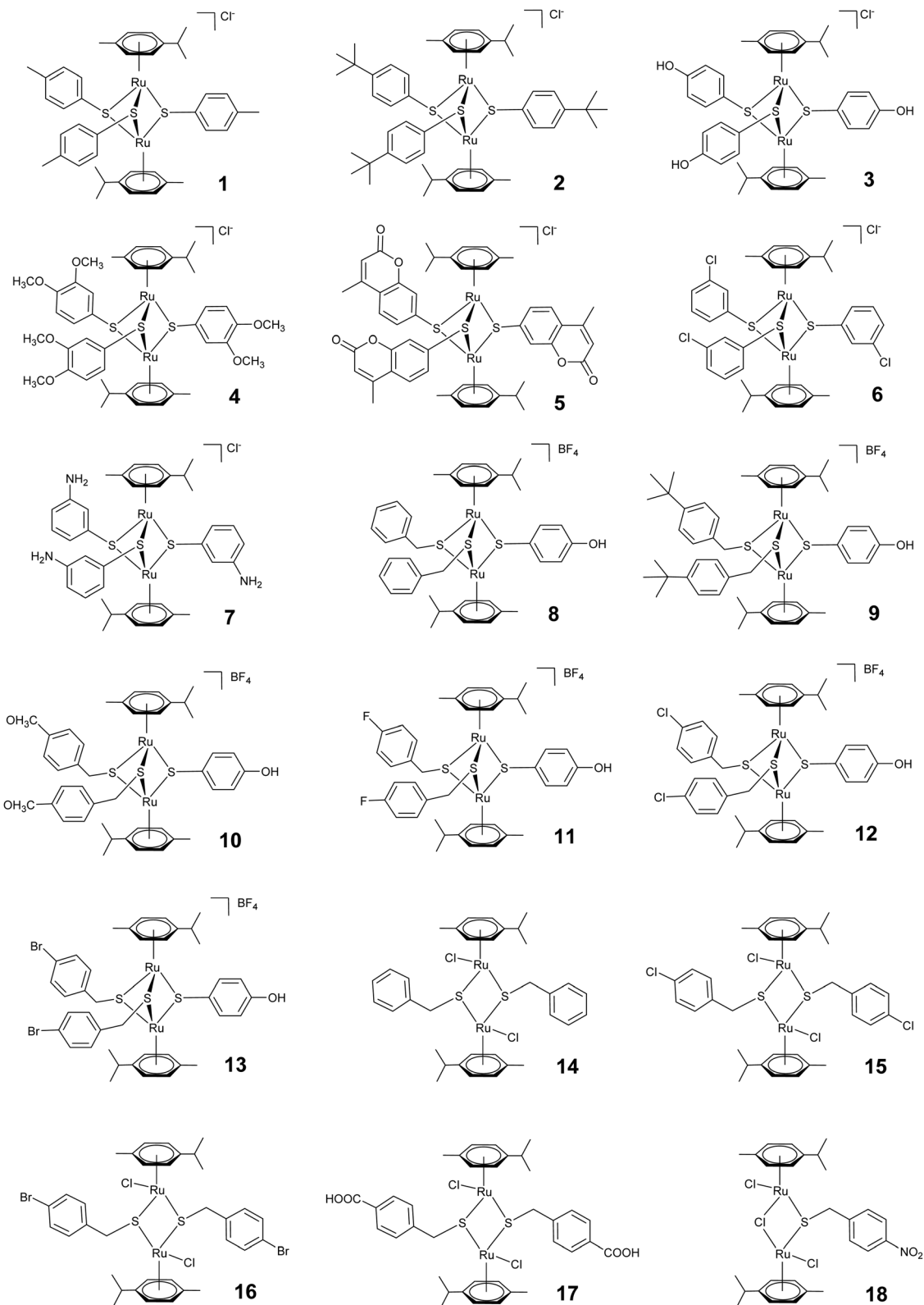


Fig. 1 Structures of the investigated dinuclear thiolato-bridged arene ruthenium complexes. Complexes **1–7** are symmetrical cationic dinuclear trithiolato complexes (one thiolato ligand), **8–13** are “mixed” cationic dinuclear trithiolato complexes (two different thiolato ligands), **14–17** are neutral dithiolato dinuclear complexes, and **18** is a neutral monothiolato dinuclear complex.



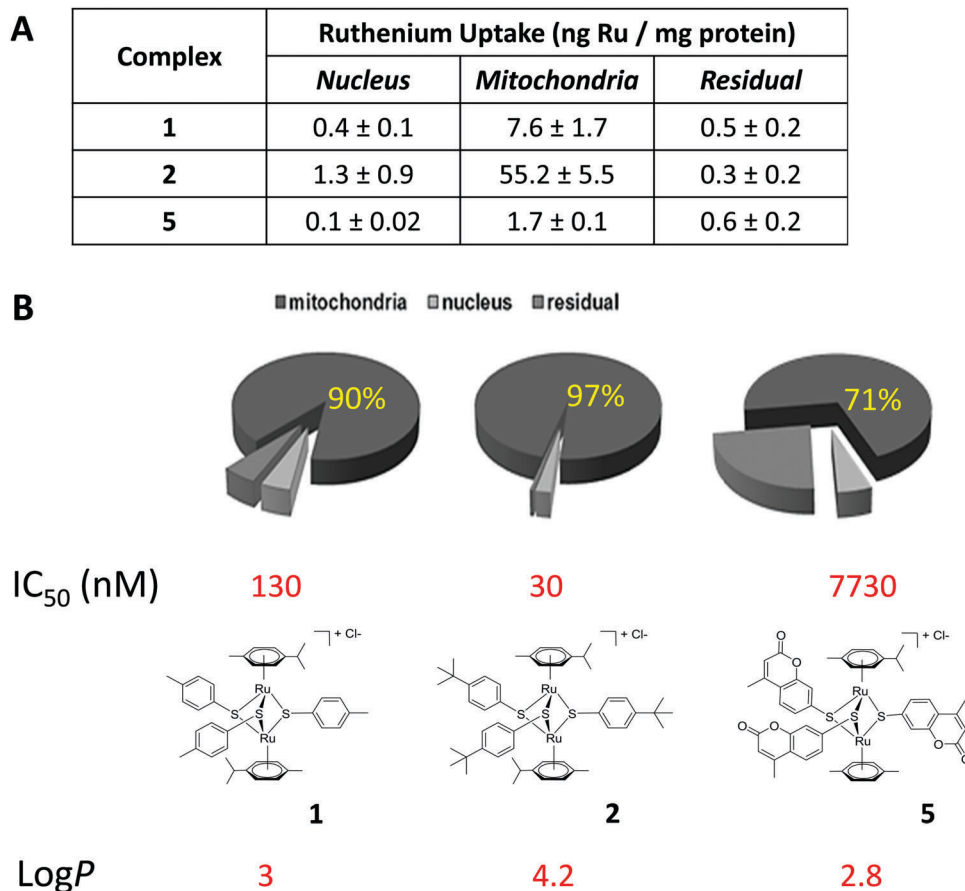


Fig. 2 Total uptake (A) and relative cellular biodistribution (B) of ruthenium in A2780 ovarian carcinoma cells treated for 4 h with 1 μ M complex **1**, **2** and **5**.

with 2 mg kg⁻¹ exhibited clinical signs and survived until the end of the experiment, while 3 out of 6 mice did in the group treated with 10 mg kg⁻¹ complex **2** for 5 days. All the remaining mice survived until 21 dpi. No statistically significant effects of drug treatments on parasite load could be detected in lungs and in the CNS in any of the drug treated groups. However, the higher rate of fatalities in the drug treated groups compared to the placebo group indicates that adverse side effects could play a role (Fig. 5).

Due to the low IC₅₀ of complex **9** (1 ± 0.3 nM), this compound was also evaluated *in vivo* in 2 independent experiments (Fig. 6). First, mice were treated with 2 mg kg⁻¹, and in a second experiment with 10 mg kg⁻¹ for 5 days. In placebo groups (infected, but treated with corn oil only), one mouse each showed symptoms compatible to neosporosis, but they did not have to be euthanized. None of the mice in the group treated with 2 mg kg⁻¹, and one of the mice treated with 10 mg kg⁻¹, died during this experiment. Determination of the parasite load in lungs and CNS by real time PCR revealed that the average parasite levels in the two organs of drug treated mice were lower compared to the placebo group. However, these values were not statistically significant (Fig. 6). Overall, the ruthenium complexes developed herein exhibited only limited *in vivo* efficacy in the neosporosis mouse model.

Biodistribution of ruthenium complex **1** indicates low drug levels in the CNS

Since *in vivo* studies in mice consistently resulted in high level brain infection comparable to the placebo treatments, we performed a biodistribution study. ICP-MS biodistribution analysis was performed in mice that had received a single dose of 10 mg kg⁻¹ of complex **1** by oral gavage. Ruthenium was identified in blood, liver, kidney and lungs at levels above those detected for a placebo-treated mouse, demonstrating the drug absorption after oral administration (Fig. 7). The majority of the ruthenium was detected in the liver at 3 h and in the kidney at 48 h post-administration, but no drug was detected in the brain. These results are in agreement with those previously reported for complex **2** (also known as *diruthenium-1* or *DiRu-1*) which had been intraperitoneally injected into mice.²⁶

Discussion

The plethora of compounds that have been developed against cancer has paved the way for numerous new and innovative approaches to combat parasitic diseases that affect not only humans, but also animals. There has been much recent interest in the development of therapeutic transition metal-based complexes in part fuelled by the clinical success of the platinum(II)



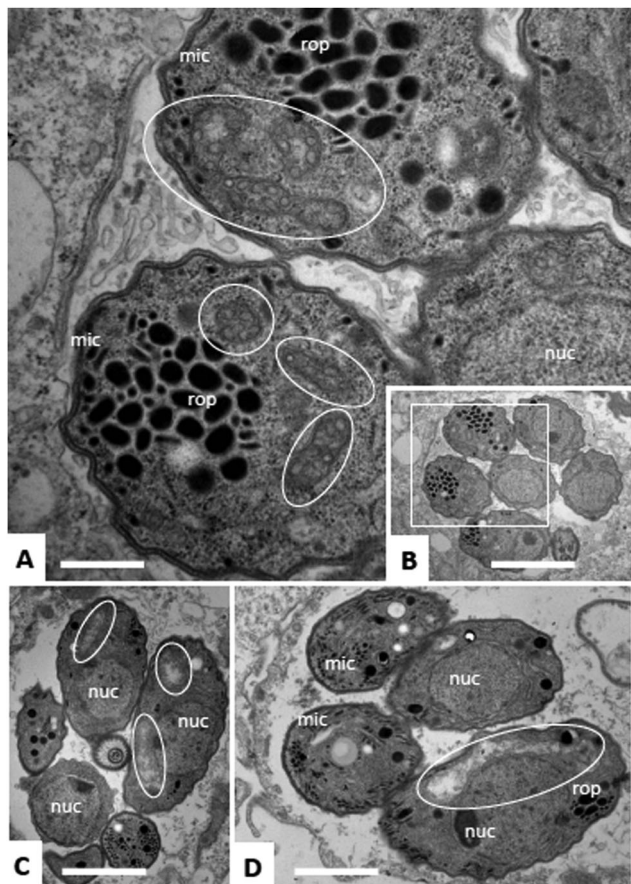


Fig. 3 TEM of non-treated *N. caninum* infected control HFF cultures (A and B) and cultures exposed to complex **1** (B) and complex **2** (C) for 6 h. The framed area in B is enlarged in A. *N. caninum* tachyzoites are located within a parasitophorous vacuole, and the secretory organelles such as micronemes (mic), rhoptries (rop) and dense granules (dg) are clearly visible. Mitochondria are encircled, and primary signs of mitochondrial distortion such as loss of the electron dense matrix are evident already at 6 hours of drug exposure (C and D); nuc = nucleus. Bar = 0.5 μm in A; 2.2 μm in B; 1.8 μm in C; 1.2 μm in D.

anticancer drug, cisplatin. Yet, known platinum drugs are limited by their high toxicity, severe side-effects, and incidences of drug resistance. Organometallic ruthenium-arene complexes have risen to prominence as a pharmacophore due to the success of other ruthenium drug candidates in clinical trials.⁵¹ Substantial activities of arene ruthenium complexes have been reported in infectious diseases caused by bacteria^{52,53} against protozoan parasites including *T. gondii*^{29,30} and *Plasmodium falciparum*,^{54,55} and against helminths such as *Schistosoma mansoni*^{56,57} and *Echinococcus multilocularis*.²⁸ In addition, a series of ruthenium-clotrimazole (ctz) complexes displayed high and specific *in vitro* activity against *Leishmania major* and *Trypanosoma cruzi*.⁵⁸ Other metal complexes containing Ag, Pt, Pd and Au, already in the market for cancer therapy, treatments of bacterial infections or as anti-inflammatory agents, have been proposed for the treatment of trypanosomatid-induced infections.¹⁰ A recent review by Gasser *et al.* provides an overview of metal complexes against tropical neglected diseases.⁵⁹

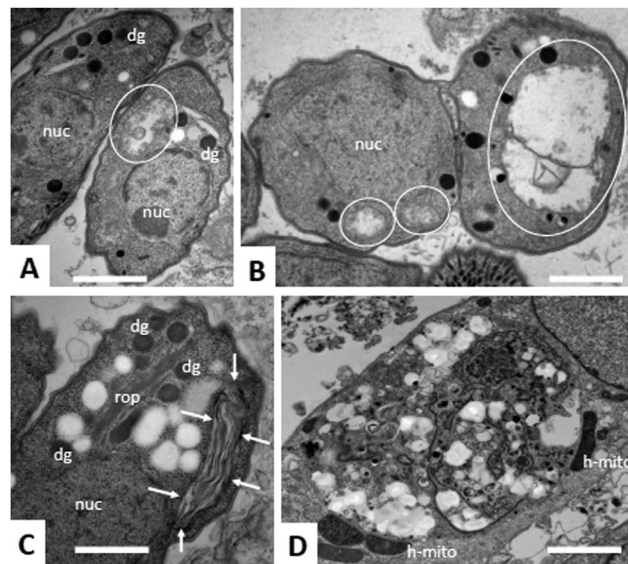


Fig. 4 TEM of *N. caninum* infected HFF cultures treated with complex **9** for 6 h (A), 24 h (B) and 72 h (C and D). Encircled areas mark the mitochondrion or residues thereof. The designation h-mito indicates the presence of structurally intact host cell mitochondria associated with the parasitophorous vacuole membrane; nuc = nucleus, dg = dense granules, rop = rhoptries. Arrows in C point towards lamellar membrane stacks. Bars = 1 μm .

We here report on the *in vitro* and *in vivo* activities of 18 dinuclear thiolato-bridged arene ruthenium complexes against the apicomplexan parasite *N. caninum*. *N. caninum* causes substantial losses in the bovine dairy and meat industry, and no vaccine and no drugs are currently on the market to limit these losses.³¹ Of the 18 complexes investigated herein, three were identified that exhibited IC_{50} values between 1 and 62 nM against *N. caninum* tachyzoites, and IC_{50} values between 30 and 530 nM against the ovarian cancer cell line A2780.^{21–23,41} The mono- and dithiolato complexes **13–18** had been also found to be only moderately cytotoxic *in vitro* against cancer cell lines (IC_{50} values between 0.2 and 2.5 μM),^{25,60,61} reflecting their propensity to readily hydrolyse and their reactivity towards biomolecules compared to their trithiolato counterparts.^{25,45}

As indicated in Table 1 for the ovarian carcinoma cell line A2780, the activity of the complexes against *N. caninum* largely mirrors the findings obtained earlier in several cancer cells,^{21–23,41} and in the closely related apicomplexan parasite *Toxoplasma gondii*.³⁰ Generally, the *in vitro* activity of anticancer and anti-parasitic drugs can be related in part to their lipophilic character. The resulting hydrophobicity may contribute to an increased uptake of the compound by the cells or the parasites, thereby enhancing the antiproliferative activity. We have previously evidenced a correlation between lipophilicity and cytotoxic activity for these types of dinuclear arene ruthenium complexes.^{21–23,25} Complexes with $\log P$ parameters up to the value 4 show a steady decrease of their IC_{50} values, which increase again for very lipophilic complexes with $\log P$ values above 4.5, the tendency being the same for all cell lines investigated.^{21–23,25} The optimum lipophilicity for these complexes was found to be for $\log P$ values in



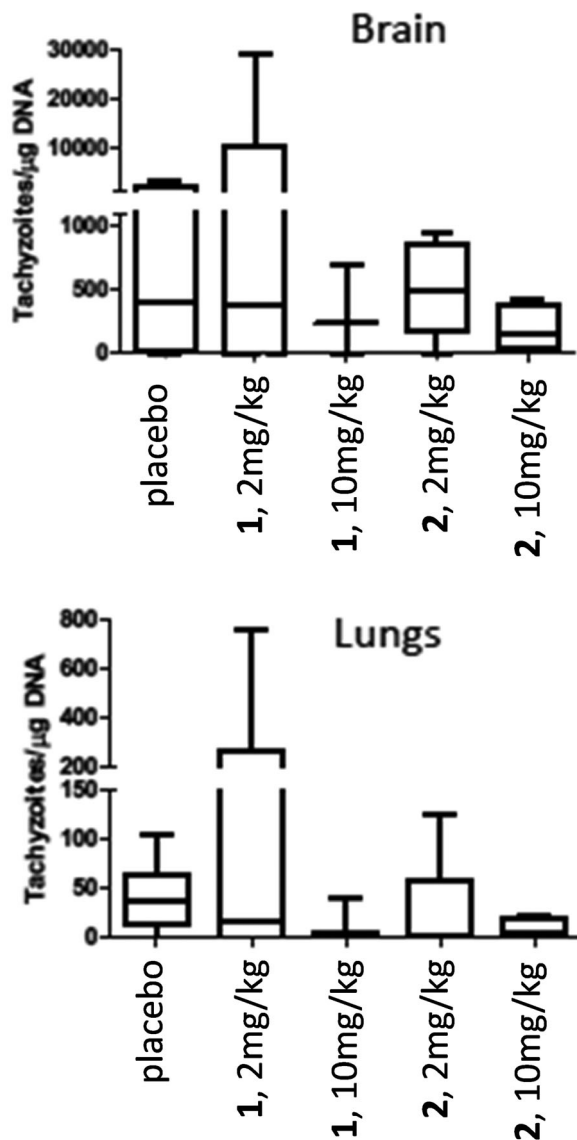


Fig. 5 Efficacy of complexes **1** and **2** in *N. caninum* infected Balb/c mice, treated with oral gavage of either 2 mg kg⁻¹ bodyweight for 5 days (A) or 10 mg kg⁻¹ bodyweight during 5 days. Placebo groups received corn oil only. Parasite load in brain and lungs is shown as box plots (no. of tachyzoites/μg DNA).

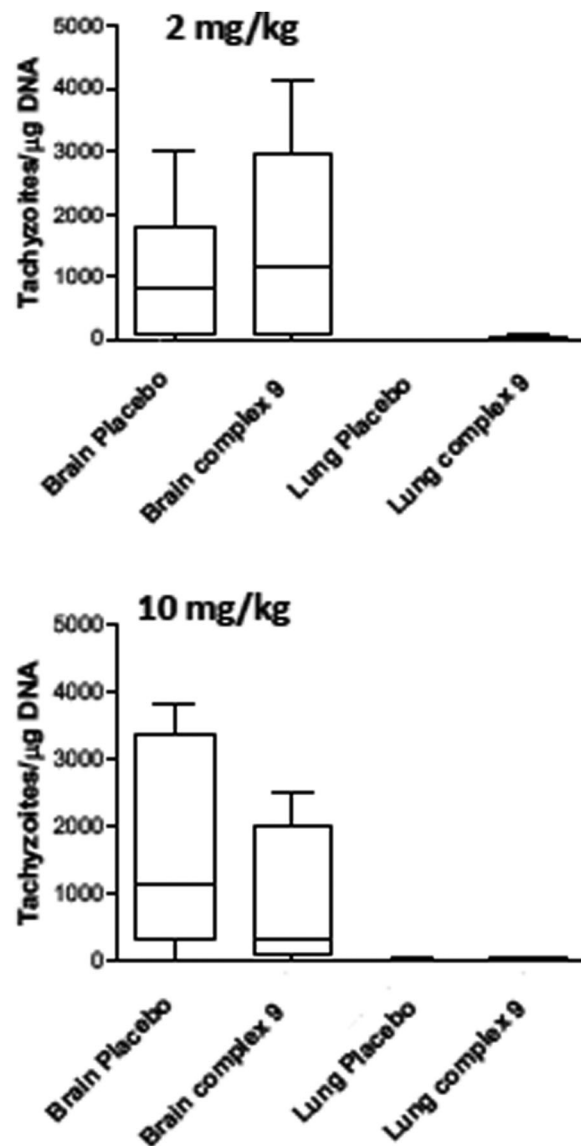


Fig. 6 Box plots of parasite loads (no of tachyzoites/μg DNA) in complex **9** treated *N. caninum* infected Balb/c mice. The drug was applied by oral gavage of either 2 mg kg⁻¹ bodyweight for 5 days (A) or 10 mg kg⁻¹ bodyweight during 5 days. Placebo groups received corn oil only.

the range between 3.5 and 4.0, whereas complexes with log *P* values lower than 3.5 systematically exhibited a steady increase of their IC₅₀ values, presumably due to an insufficient cellular uptake, since the less lipophilic compounds have more difficulties to cross cell membranes or are more rapidly excreted by exocytosis.

A similar tendency can be found in this study against *N. caninum* and previously against *T. gondii*.³⁰ Complexes with log *P* values in the range between 3.0 and 4.0 (**1**, **2** and **9**) showed the highest cytotoxicities, whereas complexes with log *P* values lower than 3.0 (**3–8** and **10–13**) belong to the less active ones of the series. If the Hammett constants σ_p are included, our results obtained on cancer cells show that the complexes having Hammett constants in the range $-0.2 < \sigma_p < 0$ and log *P* values above 3.0 have the lowest IC₅₀ values.^{21–23,25}

This tendency seems to be also valid against *N. caninum* and *T. gondii*, as shown in Table 1.

In both cancer cells and the apicomplexan parasite *N. caninum*, the mitochondrion represents the major target. This was shown by ICP-MS in drug treated A2870 cells and by performing TEM of drug-treated infected HFF. The disruption of mitochondria has been investigated as a potential novel chemotherapeutic mechanism for cancer treatment, because it circumvents upstream apoptotic pathways that may be mutated or lacking in cancer cells.⁶² Moreover, cancer cells have higher mitochondrial membrane potentials, rendering them more susceptible to mitochondrial perturbations than non-immortalized cells.⁶³ On the basis of these factors, numerous mitochondria-targeting agents have been developed in order to disrupt the





Fig. 7 *In vivo* biodistribution of complex **1** after oral administration to mice. Complex **1** was emulsified in corn oil and administered to Balb/c mice by oral gavage at a dose of 10 mg kg^{-1} of body weight. Mice were euthanized at 3 h and 48 h post-administration, whole blood, liver, kidney, lungs and brain were collected, lyophilized, and ruthenium was quantified by ICP-MS. Means \pm SEM after subtraction of results from a placebo-treated mice are shown.

mitochondrial membrane potential and further permeabilise the mitochondrial outer membrane. It has been reported that some ruthenium(II) complexes can induce mitochondria-mediated apoptosis in cancer cells.⁶⁴

In *N. caninum*, ICP-MS could not be carried out due to the lack of protocols for obtaining pure mitochondrial fractions, but TEM clearly indicated that treatments with ruthenium complexes **1**, **2** and **9** are characterized by severe ultrastructural alterations in the mitochondrial matrix and cristae early on, indicating that also in these cells the mitochondrion is the primary target. Compounds that affect the mitochondrial integrity in apicomplexan parasites have been identified earlier. For instance, low nanomolar concentrations of the naphthoquinone buparvaquone inhibited *N. caninum* proliferation *in vitro* at concentrations similar to ruthenium complexes, and buparvaquone also impaired vertical transmission in the pregnant neosporosis mouse model, but did not affect CNS infection in dams.¹³ Another interesting class are endochin-like quinolones (ELQs), which target the cytochrome *bc*₁ complex of the mitochondrial electron transport chain in protozoan parasites.⁶⁵ Cytochrome *bc*₁ facilitates the transfer of electrons from ubiquinol to cytochrome *c* and contains both oxidative (*Q*_o) sites and reductive (*Q*_i) catalytic sites. The anti-malarial drug atovaquone, also used for the treatment of toxoplasmosis, and buparvaquone, are classical *Q*_o site inhibitors.⁶⁶ More recently, novel endochin analogues with a diphenylether side chain at the third position, and various substitutions at position 5, 6 or 7 of the quinolone-ring were synthesized.^{67,68} One of these compounds, ELQ-400,⁶⁹ targets both *Q*_o and *Q*_i sites of cytochrome *bc*₁ of *S. cerevisiae* and potentially other organisms.⁷⁰ ELQ-400 was one of the compounds of outstanding activity against *N. caninum* (IC₅₀ 2 nM) that was identified during a screening of the open-source MMV (Medicines for Malaria Venture) Pathogen Box.⁷¹ One of the ELQs with excellent efficacy against *T. gondii* is ELQ-316,⁶⁵ exhibiting *in vitro* IC₅₀ values below 1 nM, and *in vivo* affecting not only tachyzoites, but also tissue cysts.⁶⁷

In vivo studies on complexes **1**, **2** and **9** were carried out using a standardized non-pregnant Balb/c mouse model for the assessment of acute and cerebral *N. caninum* infection. In this model, mice normally experience mild to medium clinical signs during the acute phase of infection (day 1–10 post infection), then immunity takes over and parasites are largely controlled, with the exception of the brain, which is an immune-privileged site.^{43,72} This is reflected by the fact that the parasite load in the lung tissues is clearly lower compared to the brain in all experiments. For all three complexes, the results indicated that they did not significantly impact on the parasite load, neither in the lungs nor in the brain. In addition, several mice from the treated groups had to be euthanized during prior to termination of the experiment on day 21 p.i.

A possible explanation for the observed failure of these complexes to exert activity in mice could be found, as exemplified for complex **1** in this study, in the respective biodistribution pattern after oral gavage, which was investigated by ICP-MS, showing that this compound is well absorbed and detectable in whole blood after 3 h, but eliminated from blood after 48 h. Instead, ruthenium was detected in the liver, lungs and kidneys after 48 h, but at no point in the brain tissue. Similar findings were obtained earlier when complex **2** was injected intraperitoneally into mice.²⁶ These findings are important as they suggest that these compounds may be ineffective against cerebral parasitic stages found in cyst-forming coccidia such as *Neospora* and *Toxoplasma*.

Conclusion

In conclusion, this study demonstrated that dinuclear thiolato-bridged arene ruthenium complexes target the mitochondrion in cancer cells as well as in the proliferative stages of apicomplexan parasites such as *N. caninum*. For future development of this class of compounds, it will be of interest to investigate which parts of the mitochondria (*i.e.* DNA, membrane, *etc.*), or potentially also other targets mediate the efficacy. The lipophilicity as well as the ligands undoubtedly play an important role for future design of dinuclear thiolato-bridged arene ruthenium complexes and conjugates. Additional studies, including a broader panel of parasites, as well as new dinuclear thiolato-bridged arene ruthenium conjugates bearing known anti-parasitic drugs such as atovaquone and buparvaquone, are needed to define precisely the antiparasitic effectiveness and potential of these ruthenium complexes as well as their molecular targets and modes of action. Overall, our work opens new avenues in the field of parasitology, as it clearly demonstrates, using metallomics studies, the importance of targeting mitochondria in parasites.

Conflicts of interest

There are no conflicts of interest to declare.

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