



Cite this: *Org. Biomol. Chem.*, 2017, **15**, 3276

Towards the biodegradation pathway of fosfomycin†

K. Pallitsch, *^a A. Schweifer, ‡ A. Roller^b and F. Hammerschmidt^a

Three functionalised propylphosphonic acids were synthesised to study C–P bond cleavage in *R. huakuii* PMY1. (*R*)-1-Hydroxy-2-oxopropylphosphonic acid [(*R*)-**5**] was prepared by chiral resolution of (±)-dimethyl 1-hydroxy-2-methylallylphosphonate [(±)-**12**], followed by ozonolysis and deprotection. The *N*-(*L*-alanyl)-substituted (1*R*,2*R*)-2-amino-1-hydroxypropylphosphonic acid **10**, a potential precursor for 2-oxopropylphosphonic acid (**5**) in cells, was obtained by coupling the aminophosphonic acid with benzotriazole-activated *Z*-*L*-alanine and hydrogenolytic deprotection. (1*R**,2*R**)-1,2-Dihydroxy-3,3,3-trifluoropropylphosphonic acid, a potential inhibitor of C–P bond cleavage after conversion into its 2-oxo derivative in the cell, was accessed from trifluoroacetaldehyde hydrate *via* hydroxypropanenitrile **21**, which was silylated and reduced to the aldehyde (±)-**23**. Diastereoselective addition of diethyl trimethylsilyl phosphite furnished diastereomeric α-siloxyphosphonates. The less polar one was converted to the desired racemic phosphonic acid (±)-(1*R**,2*R**)-**9** as its ammonium salt.

Received 6th March 2017,
Accepted 21st March 2017

DOI: 10.1039/c7ob00546f

rsc.li/obc

Introduction

Phosphonates are characterised by a direct covalent phosphorus to carbon bond, making them extremely stable against enzymatic cleavage, while at the same time being the reason for their potent bioactivity.^{1,2} Their favourable properties led to the use of both synthetic and biogenic phosphonates in medicine and agriculture. Among the economically most important phosphonates phosphinothricin (**1**),³ used as a herbicide and fosfomycin (**2**),⁴ utilised as a clinical antibiotic should be mentioned (Fig. 1).

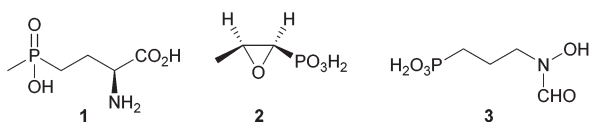


Fig. 1 Some biogenic phosphonates of agricultural and medicinal importance.

^aUniversity of Vienna, Institute of Organic Chemistry, Währingerstrasse 38, 1090 Vienna, Austria. E-mail: katharina.pallitsch@univie.ac.at; Tel: +43 (0) 1 4277 52123 or 52105

^bUniversity of Vienna, Institute of Inorganic Chemistry, Währingerstrasse 42, 1090 Vienna, Austria

† Electronic supplementary information (ESI) available: ¹H, ¹³C and ³¹P NMR spectra of the synthesised compounds, X-ray data. CCDC 1522706 for **25b**. For ESI and crystallographic data in CIF or other electronic format see DOI: 10.1039/c7ob00546f

‡ Deceased.

Fosmidomycin (**3**) is clinically evaluated for the treatment of malaria alone and in combination therapy, but has not yet reached the market.⁵ Despite these useful properties, phosphonates were thought to have little environmental importance for a long time. Lately, phosphonate catabolism in marine environments was shown to heavily influence the global phosphate and carbon cycle,⁶ contradicting this theory. Biodegradation of methylphosphonic acid was shown to be the major source of methane under aerobic conditions in ocean surface waters. In general, phosphonate degradation was shown to be quite common among microbes: studies on the distribution of the genes responsible for phosphonate breakdown in marine environments showed 40% of the studied bacterial genomes to contain one or more pathways for phosphonate degradation.⁷

This is not surprising regarding the low solubility of inorganic phosphate at ambient temperature and neutral pH⁸ and the fact that phosphorus is a vital nutrient for all life forms. Dissolved organic phosphates (DOP) often represent the only available source of phosphate in marine ecosystems. Therefore, nature evolved mechanisms to use them as a source of inorganic phosphate. DOPs mainly consist of phosphate esters (which are easy to hydrolyse), but also contain about 25% phosphonates, which can be used as a phosphate source too.⁹ Very specialised strategies are necessary to cleave the intrinsic C–P bond. This seemingly simple transformation is always the result of very unusual enzymatic pathways, of which three general types are known at present.^{2,10} While the most important pathway, called the C–P lyase pathway, follows a radical mechanism and has a very broad substrate scope, the



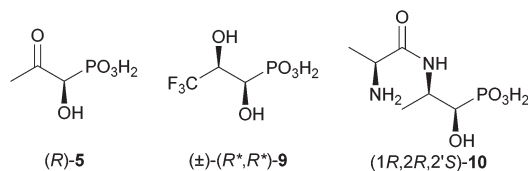


Fig. 2 Target structures to study the biodegradation of fosfomycin.

two other known pathways (hydrolytic cleavage and oxidative cleavage) have a very narrow substrate specificity. There is also striking evidence for the existence of other, until now unknown pathways.¹¹

In this article we describe the synthesis of three interesting target structures (Fig. 2), which will be used to study the catabolism of fosfomycin by *Rhizobium huakuii* PMY1, which is a microorganism that is able to use fosfomycin as the sole phosphorus and carbon source.¹²

We focus on the possibility of hydrolytic cleavage of the C–P bond by attack of an active site nucleophile of a hydrolase at the phosphorus atom after activation. This hypothesis is based on the findings of Quinn *et al.* who reported the isolation of *Rhizobium huakuii* PMY1.¹² To unravel the stepwise mineralisation of fosfomycin, they prepared putative intermediates of the catabolic pathway, among those also some isotopically labelled compounds.^{13,14} The synthesised compounds were tested as phosphorus/carbon source for the bacterium. Furthermore, cell lysates were incubated with the compounds.

These initial studies revealed the first aspects of fosfomycin metabolism by *R. huakuii* PMY1. It was shown that the first enzymatic step in its breakdown is a region- and stereospecific net opening of the epoxide ring by water at C-2 to give (1*R*,2*R*)-1,2-dihydroxypropylphosphonic acid (**4**) (Scheme 1).

The detailed mechanism for this process remains unknown. Of all other tested compounds none supported the growth of a culture of *R. huakuii* PMY1, except hydroxyacetone (**8**). However, *in vitro* C–P bond cleavage activity was detected with racemic 1-hydroxy-2-oxopropylphosphonic acid (**5**, see Fig. 2 for the (*R*)-enantiomer).¹⁴ The experiments with the labelled compounds support the hypothesis that **4** is oxidised to the corresponding 2-oxo compound **5**.¹³ As it has a carbonyl

group in the β-position to the phosphonic group, a hydrolytic mechanism for the cleavage of the C–P bond is indicated.⁷ This hypothesis forms the basis of our synthetic experiments.

Results and discussion

Proposed biodegradation pathway for fosfomycin

Based on the findings from the above described studies,^{13,14} we suggest a hypothetical biodegradation route for fosfomycin in *Rhizobium huakuii* PMY1 (Scheme 1) following a hydrolytic mechanism. We assume that dihydroxypropylphosphonic acid (*R,R*)-**4** is formed by opening of the epoxide ring by an NAD⁺ dependent dehydrogenase to give (*R*)-1-hydroxy-2-oxopropylphosphonic acid [(*R*)-**5**]. The subsequent C–P bond cleavage yielding hydroxyacetone and phosphate is presumed to involve iminium ion **6**. The latter is generated as an intermediate from the ω-amino group of an active site lysine of the putative hydrolase and the carbonyl group of (*R*)-**5**. This iminium ion is reminiscent of a similar one detected during the C–P bond cleavage of phosphonoacetaldehyde by phosphonatease.¹⁵

Here we present the synthesis of three compounds that can be used to study the final C–P bond cleavage step in the catabolism of fosfomycin by *R. huakuii* PMY1.

In previous studies a cell lysate released inorganic phosphate from racemic 1-hydroxy-2-oxopropylphosphonic acid [(±)-**5**], indicating conversion by an enzyme of *R. huakuii* PMY1.¹⁴ However, (±)-**5** did not support the growth of this bacterium. As the (*R*)-enantiomer is most probably formed *in vivo* from diol **4** by a dehydrogenase, it was assumed that either cellular uptake did not take place or that the (*S*)-enantiomer acts as a growth inhibitor.¹⁴ Therefore, both enantiomers of **5** will be needed to test them with a cell lysate and a growing culture of the bacterium separately. It is noteworthy that hydroxypropylphosphonate epoxidase (HppE) catalysing the final step of fosfomycin biosynthesis transforms both diastereomeric (1*R*,2*R/S*)-1-hydroxy-2-aminopropylphosphonic acids into (*R*)-**5** *in vitro*.¹⁶

An alternative to (*R*)-**5** would be the use of a pro-metabolite. We reasoned that (*R,R*)-2-amino-1-hydroxypropylphosphonic acid [(*R,R*)-**16**] could be a potential precursor for (*R*)-**5**, as it

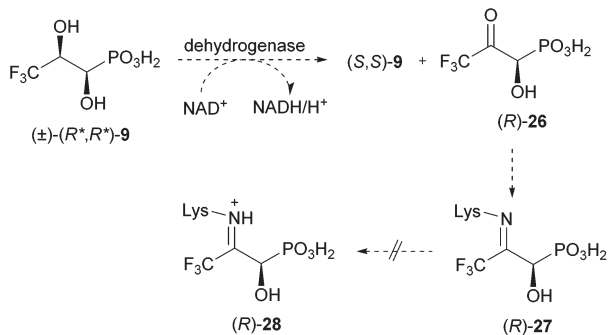


Scheme 1 Proposed biodegradation of fosfomycin by *R. huakuii* PMY1. Known conversions are denoted with full arrows, assumed transformations with dashed arrows.



could be converted by a transaminase. No phosphate was released from (*R,R*)-**16** (for the structure see Scheme 5) as the sole carbon and phosphorus source in growth experiments.¹⁷ However, there was a small detectable C–P cleavage activity with cell-extracts. It is likely that the putative precursor did not enter the bacterial cells. Thus, we envisaged a potential biochemical precursor, which will possibly be incorporated more easily by *R. huakuii* PMY1. Bacteria contain a number of transporters for amino acids, di-, tri- and oligopeptides. For example, the phosphonic acid analogue of L-alanine, a bactericidal compound, was incorporated into di-, tri- and tetrapeptides to extensively study its facilitated uptake and release by bacteria.¹⁸ Also, phosphinothricin (**1**), a widely used herbicide, easily enters bacterial cells as the tripeptide bialaphos with two L-alanines attached to the amino group.¹⁹ This tripeptide is split inside the cells by a peptidase to release phosphinothricin, the inhibitor of glutamine synthetase. Similarly, to facilitate the uptake of (*R,R*)-**16** by *R. huakuii* PMY1,²⁰ it was converted to its dipeptide (1*R*,2*R*,2'*S*)-**10** with alanine (Fig. 2). For simplicity, we started with the most simple pro-metabolite, a dipeptide. However, in case this is not incorporated by *R. huakuii* PMY1, we will aim for the corresponding tripeptide.

As described above, we presume that the cleavage of the C–P bond of 1-hydroxy-2-oxopropylphosphonic acid (**5**) involves the formation of an imine (see Scheme 1). The latter is probably protonated (**6**) to facilitate the attack of an active site nucleophile at the phosphorus atom (**7**) in analogy to phosphonate-catalysed reactions.²¹ Thus, C–P bond cleavage is induced by the formation of a Schiff's base. To prove this hypothesis, we decided to prepare a putative enzyme inhibitor. We assume that 3,3,3-trifluoro-1-hydroxy-2-oxopropylphosphonic acid (**26**, Scheme 2) would allow formation of a similar imine as in the case of the nonfluorinated analogue. However, the resulting Schiff's base (**27**) will be too electron deficient at the nitrogen (because of the electron withdrawing effect of the CF₃ group) to be protonated. The putative hydrolase, covalently modified at the active site lysine, will be blocked. Knowing that (1*R*,2*R*)-dihydroxypropylphosphonic acid [(1*R*,2*R*)-**4**] is taken up by *R. huakuii* PMY1 and metabolised, we opted for the synthesis of its racemic 3,3,3-trifluoro-methyl analogue (**9**) as a stable precursor for 3,3,3-trifluoro-1-



Scheme 2 Proposed inhibition mode of fosfomycin catabolism by (±)-(R*,R*)-**9** in *R. huakuii* PMY1.

hydroxy-2-oxopropylphosphonic acid (**26**). Oxidation of **9** inside the cell is proposed to occur upon the action of the same dehydrogenase, which is proposed to convert **4** to **5** (compare Scheme 1 and 2).

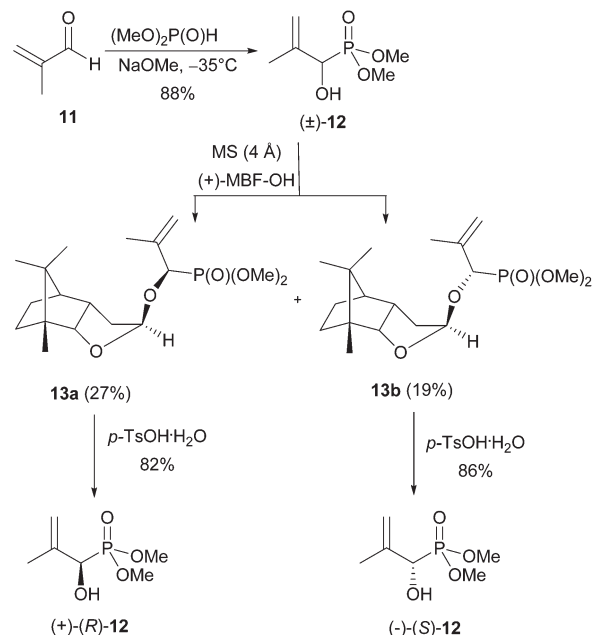
If the enzyme performing cleavage of the C–P bond is blocked, accumulation of (*R*)-3,3,3-trifluoro-1-hydroxy-2-oxopropylphosphonic acid (**26**) would be a possible detectable consequence. This would be additional information pointing towards the presented mechanism of C–P bond cleavage as the final steps in fosfomycin degradation by *R. huakuii* PMY1.

(*R*)- and (*S*)-1-Hydroxy-2-oxopropylphosphonic acid

Here we describe only the synthesis of the (*R*)-enantiomer of **5**, as the (*S*)-enantiomer can be prepared similarly just before use. The synthesis of (*R*)-1-hydroxy-2-oxopropylphosphonic acid [(*R*)-**5**] was accomplished in 5 steps from racemic dimethyl 1-hydroxy-2-methylallylphosphonate [(±)-**12**], which was obtained by a literature procedure from methacrolein and dimethyl phosphite (Scheme 3).¹⁴

The crude racemate (±)-**12** was resolved into its enantiomers using Noe's lactol,²² which reacts with alcohols in the presence of a catalytic amount of *p*TSA.²³ Here, a mixture of three diastereomeric acetals was formed. The major components were the two axially²² substituted acetals **13a** and **13b**, which are favoured due to the anomeric effect. They were separated by flash chromatography. Only one equatorially substituted acetal was observed in trace amounts. It was easily removed from the major ones.

The yield of the two main products varied slightly depending on whether Noe's lactol was used in its monomeric [(+)-MBF-OH] or dimeric [(+)-MBF-O-MBF] form for the derivatisation. In general the monomer gave slightly higher yields (27 and 19% isolated yield for **13a** and **13b**, respectively). The



Scheme 3 Chiral resolution of (±)-**12** using Noe's lactol.



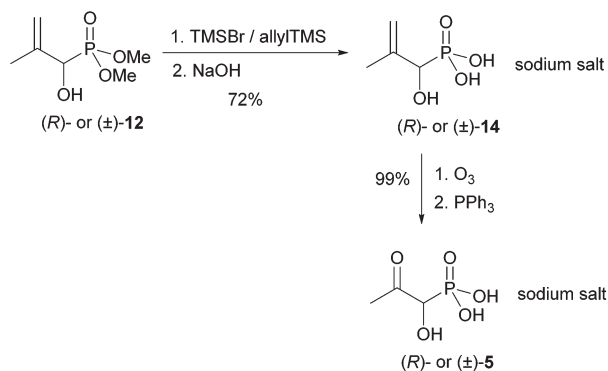
enantiomeric hydroxyphosphonates (+)- and (-)-**12** were then obtained by deprotection of the two acetals **13a** and **13b**, with *p*TSA·H₂O in a mixture of methanol and CH₂Cl₂ (82% and 86% yield).

Esterification of small samples of the enantiomers with (*S*)-Mosher chloride showed them to be enantiomerically pure (*ee* > 99%). As the phosphorus atom of the (*R*)-Mosher ester derived from enantiomer (+)-**12** resonated at higher field in the ³¹P NMR spectrum than that derived from (-)-**12**, (+)-**12** has (*R*)-configuration.²⁴ The levorotary enantiomer, derived from **13b** has therefore (*S*)-configuration. Hydroxyphosphonate (*R*)-**12** was deprotected at the phosphorus atom at 0 °C using TMSBr in combination with allyltrimethylsilane (Scheme 4) followed by an aqueous workup with neutralisation and lyophilisation.

This furnished the sodium salt of (*R*)-1-hydroxy-2-methylallylphosphonic acid [(*R*)-**14**] in 72% yield. This sodium salt of (*R*)-**14** was ozonolysed at -78 °C. Lyophilisation gave the desired sodium salt of (*R*)-**5** as a colourless powder in quantitative yield. Samples of the racemic product (±)-**5** were obtained similarly for comparative purposes.

As the sodium salts of (*R*)- and (*S*)-1-hydroxy-2-oxopropylphosphonic acid [(*R*)- and (*S*)-**5**] will be used for biodegradation studies, their stability towards racemisation in aqueous media at neutral and basic pH was tested. A sample of racemic (±)-**5** was dissolved in D₂O and the hydrogen/deuterium exchange at C-1 was monitored as a function of time at neutral and basic pH. Surprisingly, racemisation proceeded very slowly at pH 7 at ambient temperature (10–12% during the first 24 h, complete H/D-exchange within 5 days). However, racemisation was faster (50% H/D exchange after 6 h at room temperature) at pH > 8.5.

Further, the stability of the P–C bond was tested at acidic pH.²⁵ A sample of oxophosphonate **5** was dissolved in D₂O and acidified to pH 4.5 by addition of DCl/D₂O. The phosphonate/phosphate ratio was determined by ³¹P NMR spectroscopy, which showed formation of less than 5% phosphate from the phosphonate within the first 24 h. These findings suggest that the enantiomers of 1-hydroxy-2-oxopropylphosphonic acid (**5**) are stable for biological experiments at or around pH 7.



Scheme 4 Final transformations for the synthesis of (*R*)- and (±)-**5**.

A phosphonic acid dipeptide

The phosphonodipeptide (1*R*,2*R*,2'*S*)-**10** is easily accessible from commercially available fosfomycin (**2**) (as its disodium salt) in three steps (Scheme 5).

Ammonolysis of fosfomycin at 60 °C²⁶ delivered a mixture of isomeric aminophosphonates (*S,S*)-**15** and (*R,R*)-**16**. The needed regioisomer (*R,R*)-**16**, which could be easily separated from (*S,S*)-**15** by ion exchange chromatography, was coupled with benzotriazole-activated, *Z*-protected L-alanine [(*S*)-**18**].²⁷ The latter was prepared from *Z*-L-alanine and benzotriazole in 88% yield.

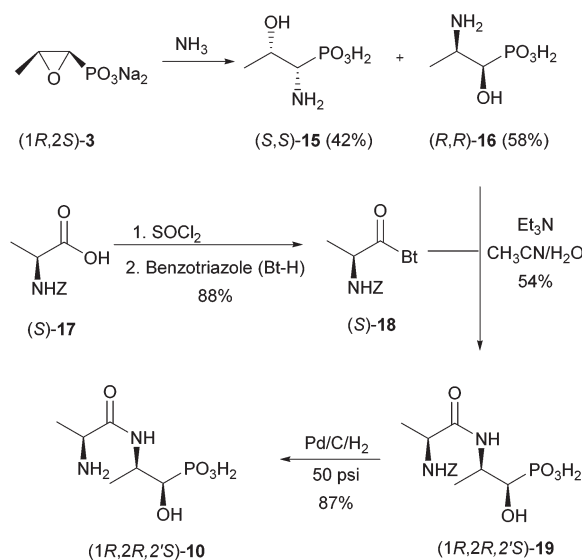
Aminophosphonic acid (*R,R*)-**16** and activated L-alanine (*S*)-**18** were coupled in a mixture of CH₃CN/H₂O (7 : 3) in the presence of one equivalent of Et₃N to give the *Z*-protected dipeptide **19** in 54% yield. An unidentified by-product and unreacted aminophosphonic acid were detected in the reaction mixture. The unknown by-product could neither be removed from the *Z*-protected dipeptide **19** nor could its formation be suppressed.

Finally, the *Z*-group was removed hydrogenolytically at 50 psi using Pd on charcoal (10% Pd) as a catalyst to give (1*R*,2*R*,2'*S*)-**10** as a white solid (87% yield). Crystallisation of the dipeptide from H₂O/MeOH removed the major portion of all impurities at the expense of yield.

A potential trifluorinated inhibitor of P–C bond cleavage in fosfomycin biodegradation

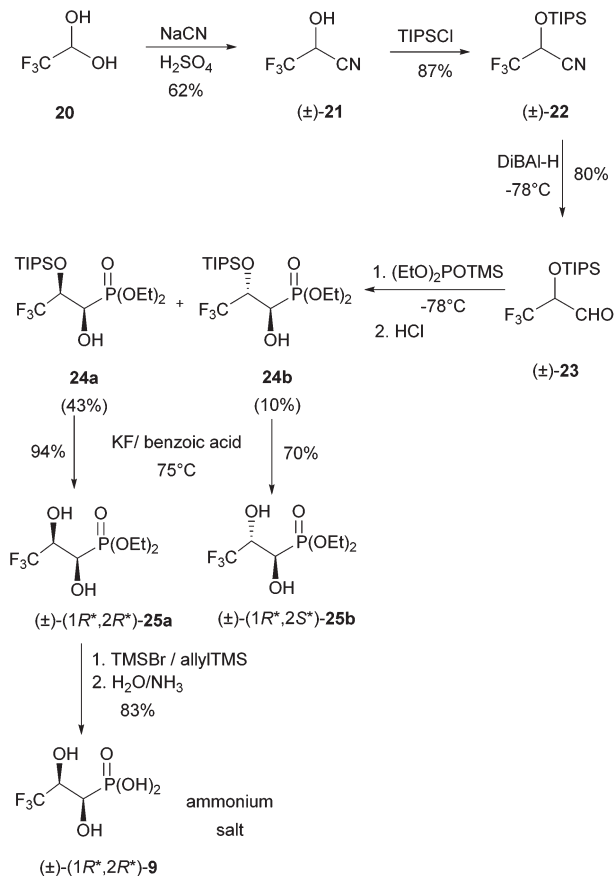
Racemic (*R**,*R**)-3,3,3-trifluoro-1,2-dihydroxypropylphosphonic acid [(±)-(*R**,*R**)-**9**] was prepared from commercially available trifluoroacetaldehyde hydrate (**20**) and NaCN in six steps (Scheme 6).

This dihydroxyphosphonate has already been synthesised by dihydroxylation of diethyl *E*-3,3,3-trifluoroprop-1-enylphosphonate with OsO₄²⁸ or KMnO₄.²⁹ We developed this new



Scheme 5 Synthesis of phosphonodipeptide (1*R*,2*R*,2'*S*)-**10**.





Scheme 6 Synthetic sequence towards (±)-(1R*,2R*)-9.

access as it will allow the synthesis of the compound labelled at C-1 with ^{13}C derived from Na^{13}CN . First sodium cyanide was added to trifluoroacetaldehyde at 0 °C under acidic conditions. The crude, racemic trifluorinated hydroxynitrile (±)-21 was distilled (62% yield, as judged by ^1H NMR spectroscopy) and used for the next step immediately. Due to the low stability of hydroxynitrile (±)-21, the hydroxyl group was protected using chlorotriisopropylsilane/imidazole. Protection proceeded in 87% yield and the resulting product (±)-22 was purified by flash chromatography and stored at room temperature. The TIPS-protected hydroxynitrile was subsequently reduced to the corresponding aldehyde 23 with DiBAL-H at -78°C followed by acidic workup. The isolated, crude aldehyde 23 was not purified by flash chromatography, as it decomposed on silica gel. Therefore, diethyl trimethylsilyl phosphite was added to a solution of the aldehyde at -78°C (Abramov reaction). The reaction mixture was allowed to warm to room temperature overnight and the TMS groups of the resulting α -(trimethylsiloxy)phosphonates were removed by addition of a few drops of concentrated HCl to the ethanolic reaction mixture. A mixture of two racemic, diastereomeric phosphonates, 24a (43%, less polar) and 24b (10%, more polar) resulted, which was easily separable by column chromatography. The configurations of 24a and 24b could be assigned after TIPS deprotection (see the following paragraph).

Attempted removal of the TIPS group from the less polar (and major) phosphonate 24a with concentrated HF (40%) in acetonitrile caused its decomposition. Therefore, desilylation was performed under milder conditions with *in situ* generated HF from a mixture of potassium fluoride and benzoic acid in acetonitrile at 75 °C for 24 h. The diol (1R*,2R*)-25a could be purified by flash chromatography to give a colourless, crystalline solid.

The more polar (and minor) phosphonate 24b was deprotected similarly. The obtained diol 25b could be crystallised from heptanes/dichloromethane to give colourless crystals, which were suitable for X-ray crystallography. It showed that the two hydroxyl groups are *anti*-orientated (see the ESI†). This findings allowed assignment of the (1R*,2R*)-configuration to 24a and the (1R*,2S*)-configuration to 24b as given in Scheme 6. Satisfyingly, the major isomer 25a is the one needed for studying C–P bond cleavage. Only this phosphonate was deprotected using bromotrimethylsilane and allyltrimethylsilane in 1,2-dichloroethane at room temperature over a period of three days. Aqueous workup with the addition of NH_3 , followed by lyophilisation gave the ammonium salt of (1R*,2R*)-3,3,3-trifluoro-1,2-dihydroxypropylphosphonic acid [(1R*,2R*)-9], in 83% yield.

Experimental section

General experimental

^1H , ^{13}C and ^{31}P NMR spectra were recorded in CDCl_3 , d_4 -methanol, d_8 -toluene or D_2O on a Bruker Avance DRX 400 (^1H : 400.13 MHz, ^{13}C : 100.61 MHz, ^{31}P : 161.98 MHz), AV 400 (^1H : 400.27 MHz, ^{13}C : 100.65 MHz, ^{31}P : 162.03 MHz) or AV600 Avance 3 (^1H : 600.25 MHz, ^{13}C : 150.95 MHz, ^{31}P : 242.97 MHz) spectrometer. Proton NMR chemical shifts were referenced to residual CHCl_3 (δ_{H} 7.24), CHD_2OD (δ_{H} 3.31), $\text{C}_6\text{D}_5\text{CD}_2\text{H}$ (δ_{H} 2.09) or HOD (δ_{H} 4.80). ^{13}C NMR spectra were referenced to CDCl_3 (δ_{C} 77.23), CD_3OD (δ_{C} 49.15) and $\text{C}_6\text{D}_5\text{CD}_3$ (δ_{C} 21.4); ^{31}P NMR spectra to external H_3PO_4 (85%) (δ_{P} 0.0). Chemical shifts (δ) are given in ppm and coupling constants (J) in Hz. IR spectra were run on a Bruker VERTEX 70 IR spectrometer in ATR mode or from solution on a silicon disc.³⁰ Optical rotations were measured at 20 °C on a PerkinElmer 341 polarimeter in a 1 dm cell. $[\alpha]_{\text{D}}$ values are given in $10^{-1} \text{ deg cm}^2 \text{ g}^{-1}$. Melting points were determined on a Leica Galen III Reichert Thermovar instrument and were uncorrected.

TLC was carried out on 0.25 mm thick Merck plates coated with silica gel 60 F254. Spots were visualised by UV light and/or dipping the plate into a solution of $(\text{NH}_4)_6\text{Mo}_7\text{O}_{24}\cdot 4\text{H}_2\text{O}$ (25.0 g) and $\text{Ce}(\text{SO}_4)_2\cdot 4\text{H}_2\text{O}$ (1.0 g) in aqueous 10% H_2SO_4 (500 mL), followed by heating with a heat gun. In case an amino-functionality was present in the molecule, the spots were visualised by dipping the plate into a solution of ninhydrin (0.2 w/v%) in ethanol followed by heating with a heat gun. Flash (column) chromatography was performed with Merck silica gel 60 (230–400 mesh).



Preparation of the sodium salt of (*R*)-1-hydroxy-2-oxopropylphosphonic acid [(*R*)-5]

(±)-Dimethyl 1-hydroxy-2-methylallylphosphonate [(±)-12]. A methanolic solution of NaOMe (0.1 mL, sat.) was added to a stirred solution of methacrolein (**11**, 0.701 g, 0.83 mL, 10.0 mmol) and dimethyl phosphite (1.10 g, 0.92 mL, 10.0 mmol) in Et₂O (20 mL) at -35 °C under argon. After 30 min concentrated H₂SO₄ (4 drops) was added. The reaction mixture was allowed to warm to room temperature and the solvent was removed under reduced pressure. The crude substance was dissolved in a mixture of toluene-CH₂Cl₂ (10 mL, 1:1), filtered and dried *in vacuo* to yield α-hydroxyallylphosphonate (±)-12 (1.579 g, 88%) as a colourless oil. The product was used for the following reaction without further purification. The recorded spectra are identical to those reported in the literature;¹⁴ δ_P (161.98 MHz, CDCl₃) 25.2 (s).

Acetals of (±)-dimethyl 1-hydroxy-2-methylallylphosphonate derived from Noe's lactol and its dimer [13a and 13b].²³ After stirring the crude α-hydroxyphosphonate [(±)-12, 2.673 g, 15 mmol] over molecular sieves (4 Å) in dry CH₂Cl₂ (60 mL) for 15 min, the mixture was cooled to 0 °C. (+)-Noe's lactol [(+)-MBF-OH, 2.944 g, 15 mmol] and *p*-toluenesulfonic acid monohydrate (0.094 g, 0.6 mmol) were added and stirring was continued for 1 h. Molecular sieves were removed by filtration, the reaction mixture was extracted with an aqueous solution of NaHCO₃ (3 × 20 mL, sat.), and then the organic layer was separated and dried (MgSO₄). The solution was concentrated under reduced pressure. The residue was flash chromatographed [ethyl acetate-hexanes 1:1 → ethyl acetate, *R*_f (**13a**, ethyl acetate-hexanes 1:1) 0.36 and *R*_f (**13b**, ethyl acetate-hexanes 1:1) 0.29] to yield the two diastereomeric acetals **13a** (1.452 g, 27%) and **13b** (1.005 g, 2.8 mmol, 19%) as colourless oils.

The acetalisation with Noe's lactol dimer [(+)-MBF-OH dimer] can be performed similarly, except that only 0.5 equivalents of the dimer were used relative to hydroxyphosphonate (±)-12.

Diastereomer 13a. [α]_D²⁰ +130.77 (*c* 0.91, acetone). Found: C, 60.32; H, 8.72; P 8.64. Calc. for C₁₈H₃₁O₅P: C, 45.49; H, 8.11; P 8.64. ν_{max}/cm⁻¹ 2954, 1451, 1262, 1181, 1040; δ_H (400.27 MHz, d₈-toluene) 5.17 (1 H, broadened d, *J*_{HP} 4.7, O-CH-O), 5.15-5.10 (1 H, m, CH₂=), 5.04-5.00 (1 H, m, CH₂=), 4.68 (1 H, d, *J*_{HH} 18.7, CH-O), 4.43 (1 H, dd, *J*_{HP} 9.5, *J*_{HH} 1.6, CH-P), 3.53 (3 H, d, *J*_{HP} 10.4, O-CH₃), 3.48 (3 H, d, *J*_{HP} 10.5, O-CH₃), 2.88-2.77 (1 H, m, CH), 1.96-1.91 (3 H, m, CH₃-C(=CH₂-), 1.90-1.82 (1 H, m, CH), 1.76-1.61 (2 H, m, 2 × CH), 1.50-1.35 (2 H, m, 2 × CH), 1.27 (1 H, ddd, *J*_{HH} 13.5, *J*_{HH} 9.2, *J*_{HH} 4.2, CH), 1.11 (1 H, tdd, *J*_{HH} 12.2, *J*_{HH} 3.8, *J*_{HH} 1.5, CH), 0.90 (3 H, s, CH₃), 0.89 (3 H, s, CH₃), 0.81 (3 H, s, CH₃); δ_C (100.61 MHz, d₈-toluene) 140.9 (s, C_q), 117.0 (d, *J*_{CP} 12.2, CH₂=), 106.8 (d, *J*_{CP} 12.2, O-CH-O), 90.9 (s, CH-O), 75.5 (d, *J*_{CP} 169.1, CH-P), 54.1 (d, *J*_{CP} 6.7, P-OCH₃), 53.8 (s, C_q), 53.4 (d, *J*_{CP} 6.3 Hz, P-OCH₃), 49.8 (s, C_q), 48.7 (s, CH), 41.5 (s, CH), 33.4 (s, CH₂), 27.8 (s, CH₂), 21.9 (s, CH₃), 21.8 (s, CH₂), 20.6 (s, CH₃), 19.7 (s, CH₃), 15.9 (s, CH₃); δ_P (162.03 MHz, d₈-toluene) 22.0 (s).

Diastereomer 13b. [α]_D²⁰ +98.80 (*c* 0.92, acetone). ν_{max}/cm⁻¹ 2954, 1458, 1259, 1182, 1099, 1033, 985; δ_H (400.27 MHz, d₈-toluene) 5.65 (1 H, d, *J*_{HP} 4.8, O-CH-O), 5.23-5.18 (1 H, m, CH₂=), 4.97-4.90 (1 H, m, CH₂=), 4.49 (1 H, d, *J*_{HH} 13.3, CH-O), 4.25 (1 H, dd, *J*_{HP} 9.5, *J*_{HH} 1.5, CH-P), 3.49 (3 H, d, *J*_{HP} 10.3, O-CH₃), 3.47 (3 H, d, *J*_{HP} 10.3 Hz, O-CH₃), 2.86-2.76 (1 H, m, CH), 1.97-1.92 (3 H, m, CH₃-C(=CH₂-), 1.90-1.67 (3 H, m, CH₃), 1.47-1.33 (2 H, m, 2 × CH), 1.32-1.22 (1 H, m, CH), 1.15-1.05 (1 H, m, CH), 0.85 (3 H, s, CH₃), 0.83 (3 H, s, CH₃), 0.78 (3 H, s, CH₃); δ_C (100.61 MHz, d₈-toluene) 142.6 (d, *J*_{CP} 1.7, =C_q), 114.7 (d, *J*_{CP} 11.0, CH₂), 110.4 (d, *J*_{CP} 5.5, O-CH-O), 91.3 (s, CH-O), 76.7 (d, *J*_{CP} 161.8, CH-P), 53.721 (2 C, d, *J*_{CP} 6.9, 2 × O-CH₃), 53.720 (s, C_q), 53.71 (d, *J*_{CP} 6.9, O-CH₃), 49.82 (s, C_q), 48.79 (s, CH), 41.5 (s, CH), 33.8 (s, CH₂), 27.8 (s, CH₂), 21.9 (s, CH₃), 21.8 (s, CH₂), 21.0 (s, CH₃), 19.8 (s, CH₃), 15.9 (CH₃); δ_P (162.32 MHz, d₈-toluene) 22.6 (s).

Deprotection of diastereomers 13a and 13b. *p*-Toluenesulfonic acid monohydrate (0.076 g, 0.4 mmol) was added to a stirred solution of diastereomer **13a** (1.452 g, 4.1 mmol) in a mixture of dry CH₂Cl₂ (30 mL) and dry methanol (9 mL) at room temperature. The resulting solution was stirred for 2.5 h before addition of a spatula tip of solid NaHCO₃. Stirring was continued for 10 min, and then the reaction mixture was filtered and concentrated under reduced pressure. The residue was purified by flash chromatography (ethyl acetate, *R*_f 0.23) to give (+)-12 (0.598 g, 82%) as a colourless oil. The product was crystallised from diisopropyl ether to give colourless crystals; mp 47-49 °C (from diisopropyl ether), [α]_D²⁰ +2.81 (*c* 0.930 in acetone), [α]₃₆₅²⁰ +9.72 (*c* 0.930 in acetone).

(-)-12 was prepared similarly starting from diastereomer **13b** (1.005 g, 2.8 mmol) as a colourless solid (0.424 g, 86%); mp 47-49 °C (from diisopropyl ether); [α]_D²⁰ -2.80 (*c* 0.965 in acetone), [α]₃₆₅²⁰ -9.95 (*c* 0.965 in acetone). The measured spectroscopic data of both enantiomers were identical to those of the racemic compound.

The absolute configuration of the obtained enantiomeric α-hydroxyphosphonates was determined by esterification using (*S*)-Mosher chloride, following a literature procedure to give the corresponding (*R*)-Mosher esters.²⁴ Relevant NMR signals are denoted below for both diastereomeric Mosher esters.

(*R*)-Mosher ester derived from (+)-12. δ_H (400.13 MHz, CDCl₃) 3.54 (3 H, q, *J*_{HF} 1.0, O-CH₃ of Mosher ester), 1.91 (3 H, br s, CH₃-C(=CH₂-); δ_P (161.98 MHz, CDCl₃) 19.27 (s).

(*R*)-Mosher ester derived from (-)-12. δ_H (400.13 MHz, CDCl₃) 3.57 (3 H, q, *J*_{HF} 1.0, O-CH₃ of Mosher ester), 1.81 (3 H, br s, CH₃-C(=CH₂-); δ_P (161.98 MHz, CDCl₃) 19.73 (s).

These NMR data allow assignment of the (*R*)-configuration to (+)-12 and (*S*) to (-)-12.

Sodium salt of (*R*)-1-hydroxy-2-methylallylphosphonic acid [(*R*)-14]. Allyltrimethylsilane (0.503 g, 0.64 mL, 4.4 mmol) and bromotrimethylsilane (1.183 g, 1.02 mL, 7.7 mmol) were added to a stirred solution of dimethyl phosphonate (*R*)-12 (0.235 g, 1.3 mmol) in 1,2-dichloroethane (2.3 mL) under argon at 0 °C. Stirring was continued for 1 h at 0 °C. All volatile components were removed under reduced pressure and the residue was dissolved in H₂O (10 mL). After adjusting the pH



to 7 with aqueous NaOH (1 M), the resulting solution was lyophilised to yield the sodium salt of (R)-**14** as a white powder (0.184 g, 72%, yield was calculated for the disodium salt); $[\alpha]_{\text{D}}^{20}$ -3.8 (c 0.990 in water); $\nu_{\text{max}}/\text{cm}^{-1}$ 3226, 2832, 2729, 2324, 1648, 1128, 1075, 1022, 987; δ_{H} (400.27 MHz, D₂O) 5.10 (1 H, broadened d, J_{HP} 3.0, CH₂=), 5.04 (1 H, br. s, CH₂=), 4.28 (1 H, d, J_{HP} 13.7, CH-P), 1.90 (3 H, s, CH₃); δ_{C} (100.65 MHz, D₂O) 143.8 (d, J_{CP} 3.5, C_q), 112.3 (d, J_{CP} 9.8, CH₂=), 73.9 (d, J_{CP} 149.7, CH-P), 19.2 (d, J_{CP} 1.5, CH₃); δ_{P} (162.02 MHz, D₂O) 16.4 (s).

Sodium salt of (R)-1-hydroxy-2-oxopropylphosphonic acid [(R)-5]. The sodium salt of α -hydroxyallylphosphonic acid (R)-**14** (0.139 g, 0.7 mmol) was dissolved in H₂O (0.7 mL) and the solution was diluted with methanol (7 mL). Ozone was bubbled through the stirred solution at -78 °C for 4 min until the reaction mixture turned cobalt blue. Subsequently air was bubbled through the solution to remove excess ozone. Ph₃P (0.186 g, 0.7 mmol) dissolved in CH₂Cl₂ (1 mL), was added and stirring was continued at -78 °C for 10 min. The cooling bath was removed and the mixture was allowed to come to room temperature. The solvent was removed under reduced pressure and the residue was dissolved in water (7 mL). The resulting solution was extracted with CH₂Cl₂ (3 \times 7 mL) and residual organic solvents were removed from the aqueous phase under reduced pressure. The pH of the remaining solution was adjusted to 7 with aqueous NaOH (0.5 M solution) and lyophilised to yield the sodium salt of 1-hydroxy-2-oxopropylphosphonic acid [(R)-5] (0.140 g, quant., yield was calculated for the disodium salt) as a white powder; $[\alpha]_{\text{D}}^{20}$ -94.8 (c 1.010 in water); $\nu_{\text{max}}/\text{cm}^{-1}$ 3171, 1690, 1357, 1070, 964; δ_{H} (400.27 MHz, D₂O) 8.51 (0.06 H, broad s, formate), 4.65 (1 H, d, J_{HP} 19.0, CH-P), 2.38 (3 H, s, CH₃), δ_{C} (100.65 MHz, D₂O) 212.3 (d, J_{CP} 7.0, C=O), 80.0 (d, J_{CP} 120.4, CH-P), 27.1 (s, CH₃); δ_{P} (162.03 MHz, D₂O) 9.4 (s, 93 mol%, desired product), 3.7 (s, 2 mol%, unknown byproduct), 1.17 (s, 5 mol%, phosphate). The product has to be stored at -25 °C.

Preparation of (+)-N-[(S)-alanyl]- (1R,2R)-2-amino-1-hydroxypropylphosphonic acid [(1R,2R,2'S)-10]

Benzotriazole-activated Z-(S)-alanine [S-(18)].³³ Benzotriazole (2.702 g, 22.7 mmol) was dissolved in dry CH₂Cl₂ (150 mL) and thionyl chloride (0.594 g, 0.36 mL, 5.0 mmol) was added dropwise. The resulting solution was refluxed for 30 min and then cooled to room temperature by means of an ice bath. Z-(S)-Alanine (**17**, 1.116 g, 5.0 mmol) was added at once and the reaction mixture was stirred at room temperature for 3.5 h. During the course of the reaction a white precipitate was formed. The mixture was extracted with aqueous solutions of Na₂CO₃ (3 \times 60 mL, 5% in water) and NH₄Cl (2 \times 60 mL, sat.). Afterwards it was dried (Na₂SO₄) and concentrated *in vacuo*. The crude product (1.420 g, 88%) was crystallised from hexanes/1,2-dichloroethane to yield colourless needles of **18** (0.976 g, 60%); mp 113–114 °C (lit.²⁵ 114–115 °C); $[\alpha]_{\text{D}}^{20}$ -76.5 (c 0.98 in CHCl₃); δ_{H} (CDCl₃, 400.27 MHz) 8.20 (1 H, broadened d, J_{HH} 8.3, benzotriazole = Bt), 8.08 (1 H, d, J_{HH} 8.3, Bt), 7.61 (1 H, ddd, J_{HH} 8.3, J_{HH} 7.2, J_{HH} 1.0, Bt), 7.47 (1 H, ddd, J_{HH} 8.3,

J_{HH} 7.2, J_{HH} 1.0, Bt), 7.44–7.05 (5 H, m, 5 \times CH from Z), 5.79 (1 H, quin, J_{HH} 7.4, CH-N), 5.48 (0.8 H, broadened d, J_{HH} 4.9, NH), 5.07 (2 H, AB-system: J_{AB} 12.2, CH₂), 1.63 (3 H, d, J_{HH} 7.1, CH₃).

N-(Z-L-Alanyl)-(1R,2R)-2-amino-1-hydroxypropylphosphonic acid [(1R,2R,2'S)-19]. Water (15 mL) and acetonitrile (35 mL) were added to (1R,2R)-2-amino-1-hydroxypropylphosphonic acid [(R,R)-**16**, 0.310 g, 2.0 mmol] prepared from commercially available fosfomycin.^{34,26} Afterwards Et₃N (0.242 g, 0.33 mL, 2.4 mmol) was added and the reaction mixture was warmed up slightly to completely dissolve the phosphonic acid. After the addition of Z-(S)-alanyl-benzotriazole [(S)-**18**] the solution was stirred at room temperature for 3.5 h. The degree of conversion was monitored by TLC (silica gel; eluent: *i*PrOH/H₂O/NH₃ (25 w/w%) 6 : 3 : 1; R_{f} (product) 0.64, R_{f} (unknown impurity) 0.70, R_{f} (**16**) 0.24, R_{f} (**18**) 0.90, R_{f} (benzotriazole) 0.87, R_{f} (**17**) 0.79).

The reaction mixture was concentrated *in vacuo* and dissolved in water (20 mL), extracted with CH₂Cl₂ (4 \times 30 mL) and the aqueous phase was concentrated to about 5 mL under reduced pressure. The residual solution was applied to a column filled with Dowex 50 W \times 8, H⁺ (90 mL) and eluted with water. The fractions containing the product (TLC: silica gel, eluent: *i*PrOH/H₂O/NH₃ (25 w/w%) 6 : 3 : 1, product moves with the front) were collected and concentrated *in vacuo* to give the crude Z-protected peptide **19** (0.389 g, 1.1 mmol, 54%; and two impurities of an unknown structure), which was used for the next step without further purification. δ_{H} (400.27 MHz, D₂O) 7.55–7.36 (5 H, m, 5 \times CH from Z), 5.18 (2 H, s, CH₂), 4.33–4.08 (2 H, m, 2 \times CH), 3.84 (1 H, br. s, CH), 1.40 (3 H, d, J_{HH} 7.0, CH₃ alanyl residue), 1.24 (3 H, d, J_{HH} 5.3, CH₃ phosphonic acid); δ_{P} (162.03 MHz, D₂O) 19.6 (s, 91 mol%, product), 11.7 (s, 7 mol%, unknown impurity), 10.1 (s, 2 mol%, unknown impurity).

(+)-N-(L-Alanyl)-(1R,2R)-2-amino-1-hydroxypropylphosphonic acid [(1R,2R,2'S)-10]. N-(Z-L-Alanyl)-(1R,2R)-2-amino-1-hydroxypropyl-phosphonic acid [(1R,2R)-**19**, 0.360 g, 1.0 mmol] was suspended in a mixture of water (10 mL) and ethanol (15 mL). Pd (180 mg, 10% on charcoal) was added as a catalyst and the Z-group was removed by hydrogenolysis (50 psi, 2 h). The catalyst was removed by filtration through Celite (moistened and rewashed with water/ethanol 1 : 1). The filtrate was concentrated *in vacuo* to give a white solid (0.196 g, 87%) which was purified by crystallisation. The product was suspended in methanol (2.0 mL) and drops of water were added until a clear solution was obtained, which was stored at 4 °C to give colourless crystals of the desired phosphonodipeptide (1R,2R,2'S)-**10** [TLC: *i*PrOH/H₂O/NH₃ (25 w/w%) 6 : 3 : 1, R_{f} 0.35]; mp 256–260 °C (decomp.); $[\alpha]_{\text{D}}^{20}$ $+21.57$ (c 0.830 in H₂O); $\nu_{\text{max}}/\text{cm}^{-1}$ 3264, 3121, 2813, 1666, 1576, 1503, 1447, 1131, 1056; δ_{H} (400.27 MHz, D₂O) 4.34–4.23 (1 H, m, CH-C-P), 4.11 (1 H, q, J_{HH} 7.1, CH of the alanyl residue), 3.76 (1 H, dd, J_{HP} 10.1, J_{HH} 4.7, CH-P), 1.56 (3 H, d, J_{HH} 7.1, CH₃ of the alanyl residue), 1.31 (d, J_{HH} 6.8, CH₃ of phosphonic acid); δ_{C} (100.65 MHz, D₂O) 169.9 (s, C=O), 70.8 (d, J_{CP} 155.5 Hz, CH-P), 49.3 (s, CH-C=O), 47.4 (d, J_{CP} 3.6 Hz, CH-C-P), 17.3 (d, J_{CP} 8.0, CH₃ of phosphonic acid), 16.4 (s, CH₃ of the alanyl residue);



δ_P (162.03 MHz, D₂O) 16.62 (s, 97 mol%, product), 15.53 (s, 3 mol%, unknown impurity).

Preparation of the ammonium salt of (\pm)-(1*R**,2*R**)-3,3,3-trifluoro-1,2-dihydroxypropylphosphonic acid [(\pm)-9]

(\pm)-3,3,3-Trifluoro-2-hydroxypropanenitrile [(\pm)-21]. Sodium cyanide (1.862 g, 38 mmol) was dissolved in water (10 mL) and the resulting solution was cooled to 0 °C. After dropwise addition of trifluoroacetaldehyde hydrate (**20**, 3.064 g, 20 mmol, 75 w/w% in water) and H₂SO₄ (13 mL, 3 M), the solution was stirred for 24 h at room temperature. The reaction mixture was extracted with ethyl acetate (4 × 20 mL). The combined organic layers were dried (Na₂SO₄), concentrated under reduced pressure and dried *in vacuo*. The residue was purified by distillation (90–98 °C/45 mm Hg) to yield hydroxynitrile (\pm)-**21** (2.961 g, 53 mol% **21** in an admixture with EtOAc as judged by ¹H NMR; calculated yield from NMR: 1.835 g, 62%) as a colourless liquid. The crude substance was used for the following reaction without further purification. δ_H (400.27 MHz, CDCl₃) 4.85 (2 H, br. s, 2H, OH and CH); δ_H (400.27 MHz, CDCl₃ + D₂O): δ = 4.77 (1 H, q, J_{HF} 5.8, CH).

(\pm)-3,3,3-Trifluoro-2-(triisopropylsilyloxy)propane nitrile [(\pm)-22].³¹ 3,3,3-Trifluoro-2-hydroxypropanenitrile [(\pm)-**21**, 1.489 g, 7.3 mmol] (calculated from ¹H NMR, see above) was dissolved in dry DMF (12 mL) under argon at ambient temperature. Imidazole (2.395 g, 35.5 mmol) and chlorotriisopropylsilane (3.429 g, 17.8 mmol) were added subsequently and the resulting mixture was stirred at room temperature for 15 h and at 50 °C for 2 h. Water (50 mL) was added and the reaction mixture was extracted with diisopropyl ether (3 × 20 mL). The combined organic layers were washed with water (30 mL), dried (Na₂SO₄) and concentrated under reduced pressure. The residue was purified by flash chromatography (hexanes–CH₂Cl₂ 4:1, R_f 0.67) to give silyl ether (\pm)-**22** (1.779 g, 6.32 mmol, 87%) as a colourless oil; found: C, 51.36; H, 7.92; N, 4.88. Calc. for C₁₂H₂₂F₃NOSi: C, 51.22; H, 7.88; N, 4.98; ν_{max}/cm^{-1} 3407, 2949, 1703, 1347, 1266, 1204, 1153, 1109; δ_H (400.27 MHz, CDCl₃) 4.88 (1 H, q, J_{HF} 5.2, CH), 1.25–1.14 (3 H, m, 3 × (CH₃)₂CH–Si), 1.10, 1.08 (18 H, overlapping doublets, J 5.7, J 5.6, 6 × CH₃ of TIPS); δ_C (100.65 MHz, CDCl₃) 121.3 (q, J_{CF} 283.0, CF₃), 113.8 (s, CN), 63.3 (q, J_{CF} 38.3, CH–CF₃), 17.70 (3 C, s, 3 × CH₃ of TIPS), 17.65 (3 C, s, 3 × CH₃ of TIPS), 12.1 (3 C, s, 3 × (CH₃)₂CH–Si).

(\pm)-3,3,3-Trifluoro-2-(triisopropylsilyloxy)propanal [(\pm)-23]. DiBAL-H (4.92 mL, 1 M solution in hexanes) was added to a stirred solution of (\pm)-3,3,3-trifluoro-2-(triisopropylsilyloxy)propanenitrile [(\pm)-**22**, 1.141 g, 4.1 mmol] in dry toluene (12 mL) at –78 °C under argon. The resulting reaction mixture was stirred at –78 °C for 1 h. Then Et₂O (30 mL), an aqueous solution of NH₄Cl (30 mL, sat.) and H₂SO₄ (20 mL, 1.5 M) were added to the cooled solution and the resulting biphasic mixture was stirred at room temperature for 20 min. The phases were separated and the aqueous layer was extracted with Et₂O (3 × 20 mL). The combined organic portions were washed with water (30 mL), dried (MgSO₄) and concentrated under reduced pressure. The residue was dissolved in dry toluene (20 mL) and

concentrated again to remove residual water. The oily residue was dried *in vacuo* to give the crude aldehyde (\pm)-**23** (0.934 g, 80%) as an oil. The crude product was used without further purification in the following step; δ_H (400.27 MHz, CDCl₃) 9.55 (1 H, quin, J_{HH} and J_{HF} 2.0, CHO), 4.36 (1 H, qd, J_{HF} 7.0, J_{HH} 2.0, CF₃CH), 1.15–1.05 (3 H, m, 3 H, 3 × (CH₃)₂CH–Si), 1.03 (9 H, d, J_{HH} 4.3, 3 × CH₃ of TIPS), 1.01 (9 H, d, J_{HH} 4.3, 3 × CH₃ of TIPS).

(\pm)-(1*R**,2*R**)- and (\pm)-(1*R**,2*S**)-Diethyl 3,3,3-trifluoro-1-hydroxy-2-(triisopropylsilyloxy)-propylphosphonate [(\pm)-(1*R**,2*R**)-24a and (\pm)-(1*R**,2*S**)-24b]. Diethyl trimethylsilyl phosphite (1.451 g, 6.9 mmol, 1.58 mL) was added dropwise to a stirred solution of (\pm)-3,3,3-trifluoro-2-(triisopropylsilyloxy)propanal [(\pm)-**23**, 1.612 g, 5.7 mmol] under argon at –78 °C in dry toluene (20 mL). The mixture was allowed to warm up to room temperature and stirring was continued for 15 h. Then, it was concentrated under reduced pressure and dried *in vacuo*. The residual oil was dissolved in ethanol (30 mL) and concentrated HCl (11 drops) was added. The solution was stirred for 2 h at room temperature and then concentrated under reduced pressure. Water (10 mL) was added and the mixture was extracted with ethyl acetate (3 × 20 mL). The combined organic phases were dried (MgSO₄) and concentrated under reduced pressure to yield a mixture of the two racemic, diastereomeric, hydroxyphosphonates **24a** and **24b**.

The diastereomers were separated by flash chromatography [hexanes–acetone 3:1, R_f (1*R**,2*R**)-**24a**) 0.21, R_f (1*S**,2*R**)-**24b**) 0.13] to yield (\pm)-(1*R**,2*R**)-**24a** (0.858 g, 43% calculated starting from **22**) and (\pm)-(1*S**,2*R**)-**24b** (0.154 g, 10% calculated starting from **22**).

(\pm)-(1*R**,2*R**)-**24a**. Mp 63–64 °C (from hexanes); found: C, 45.53; H, 8.05. Calc. for C₁₆H₃₄F₃O₅PSi: C, 45.49; H, 8.11; ν_{max}/cm^{-1} 3268, 2945, 2869, 1462, 1264, 1164, 1140, 1025, 971; δ_H (400.13 MHz, CDCl₃) 4.63–4.53 (1 H, m, CF₃CH), 4.25–4.11 (4 H, m, 2 × OCH₂CH₃), 4.07 (1 H, m, CH–P), 2.84 (1 H, d, J_{HH} 9.8, OH), 1.33 (6 H, t, J_{HH} 7.1, 2 × OCH₂CH₃), 1.28–1.17 (3 H, m, 3 × (CH₃)₂CHSi), 1.11 and 1.09 (18 H, 2 × d, J_{HH} 3.5, 6 × CH₃ of TIPS); δ_C (100.61 MHz, CDCl₃) 124.3 (qd, J_{CF} 284.9, J_{CP} 21.9, CF₃), 69.6 (qd, J_{CF} 31.1, J_{CP} 3.2, CF₃–C), 66.1 (dd, J_{CP} 169.3, J_{CF} 1.9, CH–P), 63.6 (d, J_{CP} 7.6, OCH₂CH₃), 62.8 (d, J_{CP} 7.6, OCH₂CH₃), 18.3 (3 C, s, 3 × CH₃ of TIPS), 18.1 (3 C, s, 3 × CH₃ of TIPS), 16.7 (d, J_{CP} 5.7, OCH₂CH₃), 16.6 (d, J_{CP} 6.0, OCH₂CH₃), 13.0 (3 C, s, 3 × (CH₃)₂CHSi); δ_P (162.03 MHz, CDCl₃) 20.9 (s).

(\pm)-(1*S**,2*R**)-**24b**. Mp 51–52 °C (from hexanes); found: C, 45.70; H, 7.97. Calc. for C₁₆H₃₄F₃O₅PSi: C, 45.49; H, 8.11; ν_{max}/cm^{-1} 3266, 2946, 2870, 1467, 1262, 1177, 1140, 1054, 1026; δ_H (400.13 MHz, CDCl₃) 4.56–4.46 (1 H, m, CF₃CH), 4.25–4.11 (5 H, m, 2 × OCH₂CH₃ and CH–P), 2.90 (1 H, br. s, OH), 1.33 (6 H, td, J_{HH} 7.0, J_{HP} 0.3, 2 × OCH₂CH₃), 1.20–1.03 (21 H, m, 3 × (CH₃)₂CHSi + 3 × (CH₃)₂CHSi); δ_C (100.61 MHz, CDCl₃) 124.1 (qd, J_{CF} 284.1, J_{CP} 3.4, CF₃), 72.3 (qd, J_{CF} 30.9, J_{CP} 6.2, CF₃–C), 70.5 (d, J_{CP} 161.1, CH–P), 63.4 (d, J_{CP} 6.4, OCH₂CH₃), 63.3 (d, J_{CP} 6.4, OCH₂CH₃), 18.1 (3 C, s, 3 × CH₃ of TIPS), 18.0 (3 C, s, 3 × CH₃ of TIPS), 16.6 (2 C, d, J_{CP} 5.6, 2 × OCH₂CH₃), 12.7 (3 C, s, 3 × (CH₃)₂CHSi); δ_P (162.03 MHz, CDCl₃) 19.9 (s).



(±)-(1*R**,2*R**)-Diethyl 3,3,3-trifluoro-1,2-dihydroxypropylphosphonate [(±)-(1*R**,2*R**)-25a].^{29,30,32} The silylated phosphonate (±)-(1*R**,2*R**)-24a (0.775 g, 1.83 mmol) was dissolved in dry acetonitrile (40 mL) together with potassium fluoride²³ (0.521 g, 9.0 mmol) and benzoic acid (1.098 g, 9.0 mmol). The resulting mixture was stirred at 70 °C for 16 h under argon. The mixture was allowed to cool to room temperature and then concentrated under reduced pressure. The crude product was purified by flash chromatography (ethyl acetate, *R*_f 0.45) to yield the desired racemic diol (±)-(1*R**,2*R**)-25a (0.458 g, 94%).

(±)-(1*R**,2*S**)-Diethyl 3,3,3-trifluoro-1,2-dihydroxypropylphosphonate [(±)-(1*R**,2*S**)-25b]. (±)-(1*R**,2*S**)-24b (163 mg, 0.39 mmol) was deprotected similarly to give racemic diol (±)-(1*R**,2*S**)-25b (73 mg, 70%) as a colourless solid. It was recrystallised (heptanes/CH₂Cl₂, room temperature to 4 °C) to give colourless crystals of which the structure could be determined by X-ray crystallography.

(±)-(1*R**,2*R**)-25a. Mp 38–40 °C (from hexanes–CH₂Cl₂) (lit.,²⁹ 38 °C); found: C, 31.43; H, 5.18. Calc. for C₇H₁₄F₃O₅F₃P: C, 31.59; H, 5.30; $\nu_{\max}/\text{cm}^{-1}$ 3255, 2985, 1276, 1259, 1166, 1137, 1029; δ_{H} (400.13 MHz, CDCl₃) 5.40 (1 H, br. s, OH), 4.75 (1 H, br. s, OH), 4.32–4.14 (6 H, m, 2 × OCH₂CH₃, CF₃CH and CH–P), 1.34 (6 H, t, *J*_{HH} 7.1, 2 × OCH₂CH₃); δ_{C} (100.61 MHz, CDCl₃) 124.4 (qd, *J*_{CF} 284.0, *J*_{CP} 22.8, CF₃), 69.3 (qd, *J*_{CF} 31.1, *J*_{CP} 3.1, CF₃–C), 66.8 (dd, *J*_{CP} 165.8, *J*_{CF} 1.0, CH–P), 64.3 (d, *J*_{CP} 7.0, OCH₂CH₃), 63.9 (d, *J*_{CP} 7.4, OCH₂CH₃), 16.6 (d, *J*_{CP} 5.3, OCH₂CH₃), 16.5 (d, *J*_{CP} 5.3, OCH₂CH₃); δ_{P} (161.98 MHz, CDCl₃) 21.8 (q, *J*_{PF} 4.3).

(±)-(1*R**,2*S**)-25b. δ_{H} (600.25 MHz, CDCl₃) 4.68 (1 H, br. d, *J*_{HH} 6.4, OH), 4.31–4.10 (6 H, m, 2 × OCH₂CH₃, CF₃CH and CH–P), 3.33 (1 H, br. t, *J*_{HH} 7.5, OH), 1.37 (3 H, t, *J*_{HP} 7.1, OCH₂CH₃), 1.34 (3 H, t, *J*_{HP} 7.1, OCH₂CH₃); δ_{C} (150.95 MHz, CDCl₃) 124.3 (qd, *J*_{CF} 282.8, *J*_{CP} 14.5, CF₃), 71.46 (qd, *J*_{CF} 30.9, *J*_{CP} 1.5, CF₃–C), 67.11 (d, *J*_{CP} 162.0, CH–P), 64.1 (d, *J*_{CP} 7.1, OCH₂CH₃), 63.80 (d, *J*_{CP} 7.1, OCH₂CH₃), 16.6 (d, *J*_{CP} 5.7, OCH₂CH₃), 16.5 (d, *J*_{CP} 5.8, OCH₂CH₃); δ_{P} (242.97 MHz, CDCl₃) 20.9 (br s).

(±)-(1*R**,2*R**)-3,3,3-Trifluoro-1,2-dihydroxypropylphosphonic acid, ammonium salt [(±)-(1*R**,2*R**)-9]. A mixture of dihydroxyphosphonate (±)-(1*R**,2*R**)-25a (0.330 g, 1.24 mmol), bromotrimethylsilane (1.470 g, 1.27 mL, 9.6 mmol) and allyltrimethylsilane (0.571 g, 0.72 mL, 5.0 mmol) in 1,2-dichloroethane (7.5 mL) was stirred for 3 d at room temperature. All volatiles were removed under reduced pressure (room temperature, then 35 °C/0.5 mm Hg). The residual liquid was dissolved in water–ethanol 1:1 (10 mL) and stirred for 3 h at room temperature. The solution was concentrated again to give a white powdery solid. Water (3 mL) and ammonia solution (5 drops, 25 w/w% in water) were added. The solution was lyophilised to give the ammonium salt of racemic (±)-(1*R**,2*R**)-9 (0.219 g, 83%, calculated yield for the diammonium salt); mp 172–175 °C; $\nu_{\max}/\text{cm}^{-1}$ 3019, 2816, 1437, 1264, 1114, 1038, 970; δ_{H} (400.13 MHz, D₂O) 4.42 (1 H, qdd, *J*_{HF} 7.6, *J*_{HP} 4.8, *J*_{HH} 0.9, CF₃–CH), 3.99 (1 H, dd, *J*_{HP} 13.0, *J*_{HH} 0.9, CH–P); δ_{C} (100.61 MHz, D₂O), 125.9 (qd, *J*_{CF} 282.4, *J*_{CP} 19.2, CF₃), 69.7 (qd, *J*_{CF} 30.0, *J*_{CP} 1.5, CF₃–C), 66.6 (d, *J*_{CP} 147.6, CH–P); δ_{P} (161.98 MHz, D₂O) 15.7 (s).

Conclusions

In order to unravel the biodegradation pathway of fosfomycin, which is a biogenic phosphonate of high medicinal importance a set of three phosphonic acids was synthesised. These will help to either underpin or rule out our hypothesis for the final step in fosfomycin metabolism: the cleavage of the C–P bond. The synthesised compounds will be used as probes in future experiments with cell lysates and growing cultures of *R. huakuii* PMY1, a bacterium which is able to use fosfomycin as the sole phosphorus and carbon source.

Acknowledgements

We greatly acknowledge the financial support by the Austrian Science Fund (FWF): P27987-N28. We thank S. Felsing for recording NMR spectra and J. Theiner for performing combustion analyses.

References

- R. L. Hilderbrand, *The Role of Phosphonates in Living Systems* 1983, CRC Press Inc., Boca Raton, 1983.
- (a) G. P. Horsman and D. L. Zechel, *Chem. Rev.*, 2016, DOI: 10.1021/acs.chemrev.6b00536; (b) J. P. Chin, J. W. McGrath and J. P. Quinn, *Curr. Opin. Chem. Biol.*, 2016, **31**, 50.
- A. K. White and W. W. Metcalf, *Annu. Rev. Microbiol.*, 2007, **61**, 379.
- F. M. Kahan, J. S. Kahan, P. J. Cassidy and H. Kropp, *Anal. N. Y. Acad. Sci.*, 1974, **235**, 364.
- (a) J. Wiesner, S. Borrmann and H. Jomaa, *Parasitol. Res.*, 2003, **90**(Suppl. 2), 71; (b) S. Sooriyaarachchi, R. Chofor, M. D. P. Risseeuw, T. Bergfors, J. Pouyez, C. S. Dowd, L. Maes, J. Wouters, T. A. Jones, S. Van Calenbergh and S. L. Mowbray, *ChemMedChem*, 2016, **11**, 1.
- (a) S. S. Kamat, H. J. Williams, L. J. Dangott, M. Chakrabarti and F. M. Raushel, *Nature*, 2013, **497**, 132; (b) W. W. Metcalf, B. M. Griffin, R. M. Cicchillo, J. Gao, S. C. Janga, H. A. Cooke, B. T. Circello, B. S. Evans, W. Martens-Habbena, D. A. Stahl and W. A. van der Donk, *Science*, 2012, **337**, 1104; (c) S. C. Peck and W. A. van der Donk, *Curr. Opin. Chem. Biol.*, 2013, **17**, 580; (d) D. M. Karl, L. Beversdorf, K. M. Björkman, M. J. Church, A. Martinez and E. F. DeLong, *Nat. Geosci.*, 2008, **1**, 473.
- J. F. Villarreal-Chiu, J. P. Quinn and J. W. McGrath, *Front. Microbiol.*, 2012, **3**, 19.
- M. A. Pasek, J. M. Sampson and Z. Atlas, *Proc. Natl. Acad. Sci. U. S. A.*, 2014, **111**, 15468.
- L. C. Kolowith, E. D. Ingall and R. Brenner, *Limnol. Oceanogr.*, 2012, **46**, 309.
- J. P. Chin, J. W. McGrath and J. P. Quinn, *Curr. Opin. Chem. Biol.*, 2016, **31**, 50.
- (a) J. L. Ford, N. O. Kaakoush and G. L. Mendz, *Antonie van Leeuwenhoek*, 2010, **97**, 51; (b) E. M. Fox and G. L. Mendz,



- Enzyme Microb. Technol.*, 2006, **40**, 145; (c) G. L. Mendz, F. Megraud and V. Korolik, *Arch. Microbiol.*, 2005, **183**, 113.
- 12 J. W. McGrath, F. Hammerschmidt and J. P. Quinn, *Appl. Environ. Microbiol.*, 1998, **64**, 356.
- 13 J. W. McGrath, F. Hammerschmidt, H. Kählig, F. Wuggenig, G. Lamprecht and J. P. Quinn, *Chem. – Eur. J.*, 2011, **17**, 13341.
- 14 J. W. McGrath, F. Hammerschmidt, W. Preusser, J. P. Quinn and A. Schweifer, *Org. Biomol. Chem.*, 2009, **7**, 1944.
- 15 S.-L. Lee, T. W. Hepburn, W. H. Swartz, H. L. Ammon, P. S. Mariano and D. Dunaway-Mariano, *J. Am. Chem. Soc.*, 1992, **114**, 7346.
- 16 W. Chang, M. Dey, P. Liu, S. O. Mansoorabadi, S. Moon, Z. K. Zhao, C. L. Drennan and H. Liu, *Nature*, 2013, **496**, 114.
- 17 J. Quinn and F. Hammerschmidt, unpublished results.
- 18 F. A. Atherton, M. J. Hall, C. H. Hassall, R. W. Lambert and P. S. Ringrose, *Antimicrob. Agents Chemother.*, 1979, **15**, 677.
- 19 F. Hammerschmidt and H. Kählig, *J. Org. Chem.*, 1991, **56**, 2364.
- 20 N. Simon, *Biochim. Biophys. Acta*, 2015, **1850**, 488.
- 21 S. S. Kamat and F. M. Raushel, *Curr. Opin. Chem. Biol.*, 2013, **17**, 589.
- 22 (a) C. R. Noe, *Chem. Ber.*, 1982, **115**, 1576; (b) C. R. Noe, *Chem. Ber.*, 1982, **115**, 1591; (c) C. R. Noe, M. Knollmüller, B. Oberhauser, G. Steinbauer and E. Wagner, *Chem. Ber.*, 1986, **119**, 729.
- 23 F. Hammerschmidt and H. Völlenke, *Liebigs Ann. Chem.*, 1989, 577.
- 24 F. Hammerschmidt and Y. Li, *Tetrahedron*, 1994, **50**, 10253.
- 25 B. L. Wanner, *Biodegradation*, 1994, **5**, 175.
- 26 L. M. van Staalduinen, F. R. McSorley, K. Schiessl, J. Séguin, P. B. Wyatt, F. Hammerschmidt, D. L. Zechel and Z. Jia, *Proc. Natl. Acad. Sci. U. S. A.*, 2014, **111**, 5171.
- 27 A. R. Katritzky, K. Suzuki and S. K. Singh, *Synthesis*, 2004, 2645.
- 28 W. Cen, X. Dai and Y. Shen, *J. Fluorine Chem.*, 1993, **65**, 49.
- 29 C. Yuan, Z. Jinfeng, Y. Yuan, C. Li and W. Zhang, *J. Fluorine Chem.*, 2006, **127**, 44.
- 30 W. Mikenda, *Vib. Spectrosc.*, 1992, **3**, 327.
- 31 K. K. Ogilvie, E. A. Thompson, M. A. Quilliam and J. B. Westmore, *Tetrahedron Lett.*, 1974, **33**, 2865.
- 32 P. Kumar, K. Ohkura, D. Beiki, L. I. Wiebe and K. Seki, *Chem. Pharm. Bull.*, 2003, **51**, 399.
- 33 A. R. Katritzky, A. A. Shetopalov and K. Suzuki, *ARKIVOC*, 2005, (vii), 36.
- 34 F. Hammerschmidt, G. Bovermann and K. Bayer, *Liebigs Ann. Chem.*, 1990, 1055.

