

# Analytical Methods

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4 **1 Amplified electrochemiluminescence of lucigenin**  
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6 **2 triggered by electrochemically reduced graphene**  
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9 **3 oxide and its sensitively detection for bisphenol A**  
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16 **Abstract**

17 An amplified and stable electrochemiluminescent signal of lucigenin on  
18 electrochemically reduced graphene oxide contained sensing platform was observed.  
19 Herein, chitosan was used as the excellent dispersant, which in favor of the better  
20 dispersion of graphene oxide in solution and the better electrochemical reduction of  
21 graphene oxide. And the electrochemically reduced graphene oxide owned the  
22 outstanding conductivity which was propitious to the transfer of electrons, leading to  
23 the enhancement of electrochemiluminescent signals of lucigenin. Additionally,  
24 bisphenol A, as a classical foodborne pollutant, was firstly detected due to its strong  
25 inhibition phenomenon to the electrochemiluminescent response of lucigenin. Then  
26 we studied the quenched electrochemiluminescent system and the possible  
27 mechanism of the platform in detail. Under optimum conditions, the proposed  
28 electrochemiluminescent sensor exhibited a linear response range from  $1.0 \times 10^{-9}$  mol  
29  $L^{-1}$  to  $1.0 \times 10^{-4}$  mol  $L^{-1}$  with a low detection limit of  $3.0 \times 10^{-10}$  mol  $L^{-1}$  for bisphenol  
30 A, which might find promising applications in developing a new type of biosensor.

31  
32 **Keywords:** electrochemiluminescent, lucigenin, electrochemically reduced graphene  
33 oxide, chitosan, bisphenol A

## 1 Introduction

Electrochemiluminescence, also called as electrogenerated chemiluminescence, means that electric energy is converted into radioactive energy at the electrode surface through an applied potential.<sup>1,2</sup> Luminescent signals can be obtained from the excited states of an electrochemiluminescent (ECL) luminophore generated at electrode surfaces during the electrochemical reaction.<sup>1,2</sup> In recent years, there has been a steady growth of focus in electrochemiluminescence which is considered as an important and valuable detection method in analytical chemistry due to its high sensitivity, high selectivity and good reproducibility.<sup>3,4</sup> Furthermore, ECL sensors not only retain the advantages of chemiluminescence (CL) sensor, such as the excellent sensitivity and a wide dynamic concentration response range, but also own some additional advantages over the CL sensors.<sup>3,4</sup> Consequently, many researches had been done universally on the three ECL compounds, which include lucigenin (N,N-dimethyl- 9,9-biacridinium dinitrate), luminol and ruthenium complexes, particularly tris(2,2'-bipyridine) ruthenium(II) ( $\text{Ru}(\text{bpy})_3^{2+}$ ),<sup>1,5</sup> in which we found that, compared with the ECL reagents of luminol and  $\text{Ru}(\text{bpy})_3^{2+}$ , there were only a few literatures about ECL of lucigenin in aqueous media and analytical applications owe to the bad reproducibility. Takeo Ohsaka and Takeyoshi Okajima discovered that the sensitivity and reproducibility of ECL of lucigenin can be improved when doing some researches on the modified electrode with self-assembled monolayers and of solutions with surfactants.<sup>6</sup> Therefore, it is of great importance for extending analytical applications of lucigenin to find a new method for solving the problem of insoluble electrochemical

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4 58 luminescence absorption on the surface of electrode.  
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6 59 Tremendous attention has been paid to graphene together with its various derivatives,<sup>7</sup>  
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9 60 such as reduced graphene oxide (RGO),<sup>8,9</sup> graphene nanoribbon<sup>10</sup> and nitrogen-doped  
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11 61 graphene<sup>11</sup> in recent years because of their excellent electrical conductivity. In  
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14 62 especial, RGO, a material of two-dimensional material, is robust and popular owe to  
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16 63 its good dispersibility, convenient chemical modification, good mechanical and  
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19 64 thermal properties, high surface area, easy functionalization and good  
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21 65 biocompatibility.<sup>12</sup> These interesting properties make RGO a promising material for  
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24 66 extraordinary applications in the fields such as batteries<sup>13</sup>, catalysis<sup>14</sup>,  
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26 67 nanoelectronics<sup>15</sup> and supercapacitors<sup>16</sup>. Various methods including micromechanical  
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29 68 cleavage<sup>17</sup>, solvothermal synthesis<sup>18</sup>, epitaxial growth<sup>19,20</sup> and liquid-phase  
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31 69 exfoliation<sup>21-23</sup> have been proposed to prepare RGO. However, the widespread  
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34 70 applications of RGO are limited by high temperature, special equipment and  
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37 71 conditions, and tedious procedures during its preparation. Recently, there have been  
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39 72 reported that graphene sheets can be prepared through GO as a precursor in a bulk  
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41 73 route by non-composite conversion of GO into “chemically reduced” graphene oxide  
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44 74 making use of thermal treatment<sup>24</sup> or chemical reducing agents<sup>25</sup>, which may be cheap  
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47 75 and of high synthesized efficiency but poisonous and intricacy. More recently,  
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49 76 Srinivasan Sampath have successfully combined raman spectroscopy with  
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51 77 electrochemistry to accomplish electrochemical reduction of GO to RGO, and  
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54 78 realized an in situ raman spectroelectrochemical study.<sup>26</sup> Shaojun Dong have  
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57 79 actualized controlled synthesis of large-area and patterned electrochemically reduced  
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4 80 graphene oxide (ERGO) films.<sup>27</sup> Direct electrochemical reduction of single-layer GO  
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6 81 and subsequent functionalization with glucose oxidase have been achieved by Hua  
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8 82 Zhang and his coworkers,<sup>28</sup> which could obtain more highly conducting graphene that  
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10 83 possessed better biocompatibility on electrode materials. It is therefore of our interest  
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12 84 to develop an efficient, facile, environment friendly and low-cost electrochemical  
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14 85 method to produce RGO films. Herein, we dispersed GO in a certain concentration  
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16 86 chitosan (CS) solution and cast the GO-CS solution on the bare gold electrode, then  
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18 87 reduced the modified electrode at certain potential. Not only this modified electrode  
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20 88 amplified the ECL emission of lucigenin, but also evidently improved the stability of  
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22 89 the ECL response of lucigenin. Herein, CS was used to disperse GO and provide an  
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24 90 excellent environment to construct sensors due to its attractive properties, including  
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26 91 excellent high permeability toward water, film-forming ability, biocompatibility, good  
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28 92 adhesion, nontoxicity, biocompatibility, high mechanical strength and a susceptibility  
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30 93 to chemical modifications.<sup>29-32</sup> Besides, an ECL inhibition method for the  
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32 94 determination of trace bisphenol A (BPA) has been established based on the strong  
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34 95 inhibition phenomenon of BPA to the ECL values of lucigenin. Under the optimum  
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36 96 conditions, the sensor exhibited high sensitivity, good stability and specificity for the  
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38 97 quantitative detection of BPA, which may have promising applications for the  
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40 98 fabrication of ECL array sensors.  
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## 50 99 **2 Experimental**

### 51 100 **2.1 Reagents**

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56 101 Chitosan (CS), potassium ferricyanide ( $K_3Fe(CN)_6$ ), potassium ferrocyanide  
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4 102 (K<sub>4</sub>Fe(CN)<sub>6</sub>), potassium chloride (KCl), hydrochloric acid (HCl), acetic acid  
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6 103 (CH<sub>3</sub>COOH) and sodium acetate (CH<sub>3</sub>COONa) were purchased from Chemicals Co.,  
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9 104 Ltd. (Shanghai, China). 0.1 mol L<sup>-1</sup> acetate buffer solutions at various pH values  
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11 105 were prepared from the stock solutions of 0.1 mol L<sup>-1</sup> CH<sub>3</sub>COOH and CH<sub>3</sub>COONa  
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13 106 solution. Lucigenin was obtained from Fluka Chemie AG. 1.0×10<sup>-3</sup> mol L<sup>-1</sup> stock  
14  
15 107 solution of lucigenin was prepared by dissolving lucigenin in redistilled water. All  
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17 108 reagents were used without further purification. All aqueous solutions and the water  
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19 109 for rinsing were prepared from Ultrapure water from a Milli-Q plus system  
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21 110 (Millipore Co., >18 MX cm<sup>-1</sup>).  
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## 26 111 **2.2 Apparatus**

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28 112 A homemade ECL system including a BPCL Ultra-Weak Chemiluminescence  
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30 113 Analyzer (Institute of Biophysics, Chinese Academy of Sciences) for detecting light  
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32 114 emission and an Electrochemistry Working Station (Chenhua 660A, Shanghai, China)  
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34 115 was used to detect ECL intensity versus potential. All experiments were carried out  
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36 116 with a conventional three-electrode system in which a gold electrode (GE) (2 mm  
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38 117 diameter), KCl-saturated silver–silver chloride (Ag/AgCl) and a platinum wire acted  
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40 118 as the working, reference and counter electrodes, respectively. All potentials reported  
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42 119 here refer to the Ag/AgCl (sat. KCl) reference electrode. X-ray photoelectron  
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44 120 spectroscopy (XPS) measurements were carried out with ESCALAB 250 Å 1314.  
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46 121 Transmission Electron Microscope (TEM) measurements were executed with  
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48 122 JEM-2100F, Japan. The solution pH measurements were carried out on a pHS-3C  
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50 123 pH meter equipped with a composite pH glass electrode (Shanghai Dapu Instr. Inc.,  
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4 124 China), after careful pH-calibration procedures.  
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### 6 125 **2.3 Preparation of GO-CS colloidal suspensions** 7

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9 126 The preparation of GO was based on Hummers' method described in the literature.<sup>33</sup>  
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11 127 In a typical procedure, 0.2 g of graphite powder was added into 3.5 mL H<sub>2</sub>SO<sub>4</sub> and  
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13 128 stirred for 2 h, in the same time adding 0.6 g KMnO<sub>4</sub> slowly to maintain the  
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15 129 temperature of the solution below 293 K under constant stirring. The resulting  
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17 130 mixture was then stirred at 308 K for 30 min then 15 mL H<sub>2</sub>O and 2 mL 30% H<sub>2</sub>O<sub>2</sub>  
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19 131 solutions were added to terminate the oxidation. The mixture was then washed with 5%  
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21 132 HCl aqueous solution repeatedly until the pH became neutral. 16 mL water was added  
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23 133 into the resulting precipitate and sonicated well to obtain GO. The brown dispersion  
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25 134 was centrifuged at 4000 rpm for 2 h to remove any unexfoliated GO. Then 8 mg mL<sup>-1</sup>  
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27 135 GO suspension was prepared by dissolving 4 mg GO in 500 μL deionized water and  
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29 136 vibrating for 5 min at room temperature until obtaining a homogeneous GO solution.  
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31 137 A 1.0% CS solution was prepared by dissolving 1.0 g CS solid in 100 mL of 1.0 %  
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33 138 acetic acid solution and stirring for several hours at room temperature until reaching a  
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35 139 homogeneous CS solution, and the CS solution was stored in refrigerator when it was  
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37 140 not in use. A certain proportion GO-CS was prepared by mixing certain volume 8 mg  
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39 141 mL<sup>-1</sup> GO suspensions with certain volume 1.0% chitosan solution, and then vibrating  
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41 142 and sonicating until obtaining a homogeneous solution.  
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### 51 143 **2.4 Preparation and reduction of GO-CS modified gold electrode** 52

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54 144 The mechanism for fabrication of ECL sensor was shown in Schem. 1A. Before  
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56 145 modification, the bare gold electrode (GE) was firstly polished with 0.05 μm alumina  
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4 146 slurry, and next sonicated in deionized water to remove the trace remainder, then  
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6 147 immersed in freshly prepared Piranha solution (30% H<sub>2</sub>O<sub>2</sub> and concentrated H<sub>2</sub>SO<sub>4</sub>,  
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9 148 3:1 (v/v)) for 10 min, and finally electrochemically pre-treated by cyclic potential  
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11 149 scanning between 1.4 V and -0.2 V in 0.1 mol L<sup>-1</sup> H<sub>2</sub>SO<sub>4</sub> until cyclic voltammogram  
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13 150 of clean gold electrode was obtained. During modification, 3 μL prepared GO-CS  
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15 151 colloidal suspension was casted on the pre-treated gold electrode, dried under infrared  
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18 152 lamp and cooled to room temperature. Then, the GO-CS modified gold electrode was  
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21 153 reduced in CH<sub>3</sub>COOH-CH<sub>3</sub>COONa (pH 5.0) buffer with 0.1 mol L<sup>-1</sup> KCl solution at  
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24 154 -1.3 V for 400 s.

### 155 **3 Results and discussion**

#### 156 **3.1 The morphology of the reduced modified electrode**

157 The morphological and structural characterization of GO was performed by TEM. As  
158 shown in Fig.1A, we can clearly found that the GO possessed transparent and similar  
159 to silk veil large layered structure. The XPS of the GO-CS and ERGO-CS modified  
160 gold electrodes were exhibited in Fig. 1B and Fig. 1C, respectively. The peak at 284.5,  
161 286.4 and 288.2 eV, which stand for C-C, C-O and C=O bands in graphene-based  
162 materials, respectively. By comparing Fig. 1B with 1C, after electrochemical  
163 reduction, we can know that the peak intensity of functional groups that contain the  
164 element of oxygen decreased tremendously while that of carbon-carbon bond  
165 increased. The result confirmed that the effectiveness of the electrochemical reduction  
166 and the GO can be reduced through the electrochemical method in the CS modified  
167 layer.

### 168 3.2 Electrochemical behavior of various modified electrodes

169 As can be seen from Fig. 2A, there were cyclic voltammograms (CVs) of  
170 ERGO-CS/GE (a), GO-CS/GE (c) and GE (b). A classic pair of redox peaks was  
171 obtained at the bare gold electrode. Whereas, the introduced of GO/CS composite film  
172 significantly dwindled the oxidation/reduction of  $\text{Fe}(\text{CN})_6^{3-/4-}$  on the electrode, which  
173 due to not only the GO as the insulator impeded the transfer of electron in the  
174 electrode surface, but also the electrostatic repulsion between GO/CS composite and  
175 electrochemical probes. It is especially noteworthy that we could find the value of  
176 peak current of reduced modified electrode obtained at -1.3 V for 400 s was bigger  
177 than that of bare gold electrode. This is mainly due to the electrochemical reduction of  
178 ERGO possessed high conductivity, which would lead to the electron transfer in  
179 electrode surface and the reduction of negative charge in ERGO/CS membrane  
180 lessened the electrostatic repulsion to  $\text{Fe}(\text{CN})_6^{3-}$ . The result was well consistent with  
181 the phenomenon in early reports, such as, Navarro observed a three-order increase in  
182 conductivity when GO was chemically reduced using hydrazine to RGO, and Hua  
183 Zhang<sup>28</sup> found the redox peak of RGO-CS modified gold electrode in  $[\text{Fe}(\text{CN})_6]^{3-/4-}$   
184 was almost the same as that of bare GCE, which also indicated that the conductivity  
185 of the electrochemically RGO was as good as that of GCE.

186 Fig. 2B showed the CVs of the modified electrode in  $\text{K}_3[\text{Fe}(\text{CN})_6]$  solution with  
187 various potential scan rates. The relationship redox peak current of  $[\text{Fe}(\text{CN})_6]^{3-/4-}$  and  
188 the square root of scan rate assess the properties of electrode, described by following  
189 Randles–Sevcik equation<sup>34</sup>:

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$$i_p = (2.69 \times 10^5) n^{3/2} A D^{1/2} C_0 v^{1/2}$$

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191 In Fig. 2B inset, we can conclude that the anodic and cathodic peak currents varied  
192 linearly with the square root of the scan rate, suggesting that the reaction was a  
193 diffuse-controlled process.

194 The Randles circuit was selected to obtain the impedance data. As we all know, the  
195 resistance to charge transfer ( $R_{ct}$ ) and the diffusion impedance ( $W$ ) were both in  
196 parallel to the interfacial capacitance ( $C_{dl}$ ). The diameter of the semicircle changed  
197 corresponding to the interfacial electron-transfer resistance ( $R_{ct}$ ). From Fig. 2C, we  
198 could easily find that the diameter of the semicircle of bare gold electrode was the  
199 smallest and that of ERGO-CS modified electrode was much smaller than the GO-CS  
200 modified electrode. We can learn from the phenomenon that the conductivity of  
201 reduced electrode was better than the unreduced one and that ERGO-CS composite  
202 film could function as an efficient electron conducting tunnel between the electrode  
203 and electrolyte.

### 204 3.3 ECL characteristics of various modified electrodes

205 We could see from the Fig. 2D that the ECL values at the bare gold electrode and  
206 GO-CS/GE were very low. However, the ECL values of ERGO-CS/GE increased to  
207 4500, almost 9 folds compared with that of GO-CS/GE, demonstrating that  
208 ERGO-CS/GE had better conductivity. In addition, the electrochemiluminescence of  
209 lucigenin on ERGO-CS/GE occurred at a more positive initial voltage, indicating the  
210 ECL process can be carried out faster and easier. From the electrochemical behavior  
211 characterization of different modified films, we have achieved that the

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4 212 electrochemical reduction of GO generated ERGO has good conductivity, it could  
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6 213 promote the transfer rate of electron in ECL process which lead to faster and stronger  
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9 214 ECL intensity. Simultaneously, negatively charged chitosan was propitious to  
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11 215 positively charged lucigenin absorbed to electrode surface by electrostatic attraction,  
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13 216 which lead to the ECL intensity enhancement of lucigenin in ERGO-CS composite  
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15 217 film modified gold electrode.

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18 218 The ECL mechanism of lucigenin at  $-0.6$  V have been studied and could be  
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21 219 intuitively seen in Scheme 1B. Primarily, electrons were transferred from electrode to  
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23 220 the lucigenin and  $O_2$  through ERGO-CS, which led to the generation of  $Luc^+$  and  
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25 221  $O_2^{\cdot-}$ . The cathodic ECL herein was originated from the formation of excited-state  
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27 222  $NMA^*$  via the transformation of the dioxetane which generated by the reaction of  
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29 223  $Luc^{2+}$  and  $O_2^{\cdot-}$ .<sup>35-39</sup> The corresponding ECL mechanism could be formulated as  
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33 224 follows:



### 45 46 229 **3.4 Influence factors on ECL emission on ERGO-CS/GE**

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49 230 The parameters of the modified electrode affected not only the ECL intensity but also  
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51 231 the reproducibility of proposed ECL method. Thus we chose some parameters to  
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54 232 investigate in order to get the best analytical performance.

#### 55 56 233 **3.4.1 Effect of GO concentration**

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4 234 From Fig. 3A we could find the ECL intensity first increased and then decreased with  
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6 235 the increase in the concentrations of GO, and the maximum response was obtained at  
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8 236 the concentration of  $1.5 \text{ mg mL}^{-1}$ , which was attribute to that EGRO obtained by the  
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10 237 process of electrochemical reduce the GO was helpful for the electron transfer.  
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12 238 Besides, the increased surface area of electrode supported substance transportation  
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14 239 and made contribution to the enhanced ECL intensity. However, when the  
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16 240 concentration was higher than  $1.5 \text{ mg mL}^{-1}$ , the thicker modified membrane hindered  
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18 241 the mass transfer of reactants resulting in the decrease of ECL intensity,<sup>40</sup> therefore  
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20 242  $1.5 \text{ mg mL}^{-1}$  GO was selected as the optimal concentration for this electrochemical  
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22 243 sensor.  
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#### 28 244 **3.4.2 Reduction potential effect**

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31 245 As shown in Fig. 3B, only weak ECL signals could be found when the reduction  
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33 246 potential was lower than  $-1.0 \text{ V}$  maybe because of the trace amount of the ERGO.  
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35 247 Moreover, we found that in certain range, the higher the reduction potential, the  
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37 248 higher the ECL intensity, and then the ECL intensity will keep at a steady value after  
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39 249  $-1.3 \text{ V}$ , suggesting that the GO in the composite film could be almost reduced at  $-1.3$   
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41 250  $\text{V}$ .  
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#### 46 251 **3.4.3 Reduction time dependence**

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49 252 The reduction time influenced the ECL signals obviously. When the time was less  
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51 253 than  $400 \text{ s}$ , the ECL intensity increased with the reduction time and then reached the  
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53 254 largest at  $400 \text{ s}$ . More than  $400 \text{ s}$ , the ECL value decreased quickly, as we can see in  
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56 255 Fig. 3C. This may be due to that the electrode surface performance could cluster  
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4 256 sharply, which was harmful for the electron delivering, and the construction of CS  
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6 257 would be violated when reduction time was longer than 400 s at the optimal potential.  
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#### 9 258 **3.4.4 The influence of dispensed volume**

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11 259 As can be seen from Fig. 3D, the highest ECL intensity could be obtained when the  
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13 260 dispensed volume was 3  $\mu\text{L}$ . The possible reason was that too much modified volume  
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15 261 was unfavorable for the ECL behavior and resulted in the increase of non-conductive  
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17 262 GO resulting in weaker electron conduction and the thick membrane would hinder the  
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19 263 mass transfer of reactants.<sup>36</sup>  
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#### 22 264 **3.4.5 CS concentration dependence**

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25 265 From the picture (Fig. 3E) we can know that the concentration of CS used for  
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27 266 modified gold electrode had great influence on the ECL intensity. Firstly, the possible  
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29 267 reason could be that the higher the concentration of the CS, the better the dispersion  
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31 268 of GO in solution, and the easier for solution to form a membrane, which was in favor  
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33 269 of mass transfer. Secondly, negative charged CS was propitious to the accumulation of  
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35 270 lucigenin on the surface of electrode, which could also lead to the increase of ECL  
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37 271 intensity. We tried different concentrations of CS and found that when the value was  
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39 272 0.3%, the ECL was strongest, so 0.3% CS was selected for subsequent studies.  
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#### 46 273 **3.5 The linearity of lucigenin on the ERGO-CS/GE and GE**

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49 274 Fig. 4A revealed that the linear range of lucigenin on the ERGO-CS/GE is wider than  
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51 275 that on bare gold electrode, ranging from  $1.0 \times 10^{-7} \text{ mol L}^{-1}$  to  $1.0 \times 10^{-4} \text{ mol L}^{-1}$ , and the  
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53 276 regression equation on the ERGO-CS/GE was  $\lg(\text{ECL}) = 0.5174 \lg C + 6.0585$  ( $R^2 =$   
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55 277  $0.994$ ), where ECL is the ECL intensity; C is the concentration of lucigenin, while on  
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4 278 the bare gold electrode, only ranging from  $5.0 \times 10^{-6}$  mol L<sup>-1</sup> to  $1.0 \times 10^{-4}$  mol L<sup>-1</sup>, and  
5  
6 279 the regression equation was  $\lg(\text{ECL}) = 1.2918 \lg C + 8.6151$  ( $R^2 = 0.997$ ), where the  
7  
8  
9 280 ECL and C stay just the same as before.

### 281 **3.6 Stability and reproducibility of the modified electrode**

282 Fig. 4B showed the ECL intensity of ERGO-CS/GE in the optimal parameters. We  
283 knew from the figure that the ECL intensity remained almost the same under the  
284 optimum conditions scanning for 10 cycles and the relative standard deviation (RSD)  
285 of the ECL intensity was only 5%, which indicated that the prepared sensor possessed  
286 good stability and showed potential for practical applications. Reproducibility of the  
287 modified electrode was investigated in 20  $\mu\text{mol L}^{-1}$  lucigenin for three different  
288 ERGO-CS modified electrodes. Three measurements resulted in a relative standard  
289 derivation of 3.4%, showing good reproducibility of the fabrication protocol. Besides,  
290 when the electrode was not in use, it was stored in the shade at room temperature and  
291 measured every week. No obvious decrease in the photocurrent response to lucigenin  
292 was observed after 2 weeks, and 93% of the initial photocurrent response was  
293 maintained after one month. This was attribute to the improvement of hydrophilicity of  
294 the membrane on the modified electrode, which played a role in avoiding the  
295 absorption of resultant of lucigenin reaction which is of hydrophobicity. We can take  
296 advantage of the property that the ECL sensor was very steady and the chemical  
297 property of ERGO was stable in the following studies.

### 298 **3.7 Practical Detection**

299 BPA, an estrogenic endocrine-disrupting chemical, is used in the lacquer lining of

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4 300 metal food cans and water pipes, food packing and dental sealants, which has been  
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6 301 attracting more and more attention due to a large body of assured reports pointed out  
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8 302 that BPA can induce a variety of adverse effects including abnormal weight gain,  
9  
10 303 prostate cancer and insulin resistance in human. Recently, analysis of BPA has mainly  
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12 304 been accomplished by high performance liquid chromatography-mass spectrometry,<sup>41</sup>  
13  
14 305 gas chromatography-mass spectrometry,<sup>42</sup> and liquid chromatography,<sup>43</sup> with various  
15  
16 306 detectors. Nonetheless, they were low sensitivity, time consuming, and needing  
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18 307 troublesome sample preparation, highly qualified technicians and sophisticated  
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20 308 equipments.<sup>44</sup> Consequently, an eco-friendliness, fast and sensitive method based on  
21  
22 309 the strong inhibition phenomenon of BPA to the ECL values of lucigenin to  
23  
24 310 monitoring the trace level BPA in biological and environmental samples.

### 311 **3.7.1 Determination mechanism**

312 Scheme 1C demonstrated the possible mechanism about how the ECL signal of  
313 lucigenin was quenched by BPA. The possible quenching mechanism between  
314 lucigenin and BPA was similar to that between lucigenin and carboic acid<sup>37</sup> as well as  
315 aniline<sup>45</sup>. Chemical compound that has electron-donating group, especially  $\cdot\text{OH}$ ,  $\cdot\text{NH}_2$ ,  
316 would restrain the ECL of lucigenin. While compounds that have intense reduction  
317 ability, such as phenol compounds can act as good antioxygen which could strongly  
318 quench the ECL of lucigenin as well. From Fig.1 we could clearly know that the BPA  
319 possess two  $\cdot\text{OH}$ , BPA may be oxidized when  $\text{O}_2^{\cdot-}$  encountered the BPA and the  
320 chance of  $\text{O}_2^{\cdot-}$  coming across the  $\text{Luc}^{\cdot+}$  would be reduced simultaneity, as a result, the  
321 ECL values decreased after BPA being injected into lucigenin solution.

### 3.7.2 Condition optimization in the BPA detection

We could see that the ECL intensity was strongest when the pH value was 5.0 as Fig. 5A showed. This may be ascribed to that protonation involved in the electrochemical reduction process when at lower pH values,<sup>37</sup> however, when the pH value was lower than 5.0, the ECL intensity decreased, which could be attributed to that the CS and GO could dissolve in such a buffer as we all know that the CS could dissolve in the acid solution. Therefore, the ECL detection was performed in pH 5.0 buffer solutions. Besides, the effect of the KCl concentration in buffer solution (5.0) on the ECL intensity of the sensor was investigated, which revealed clearly that the concentration of KCl in buffer solution tremendously influenced the ECL intensity, the lower the lg (concentration of KCl), the stronger the ECL intensity in certain range. As clearly depicted in Fig. 5B, when the concentration of KCl was higher than  $0.01\text{ mol L}^{-1}$ , the background signal increased greatly.<sup>37</sup> The maximum ECL was obtained in  $1\times 10^{-4}\text{ mol L}^{-1}$  according to the figure. And when the concentration of KCl was lower than  $1\times 10^{-4}\text{ mol L}^{-1}$ , the tendency changed inconspicuously, so  $1\times 10^{-4}\text{ mol L}^{-1}$  was the optimum concentration of KCl. Finally, the concentration of lucigenin also produced a great influence on the ECL intensity. According to the study before, we know that, first, when the concentration was higher than  $1\times 10^{-4}\text{ mol L}^{-1}$ , the ECL background signal of lucigenin was quite high and it weakened the ECL intensity, which was harmful for achieving higher sensitivity. Second, higher concentration of lucigenin would produce more reductive product, which was of low solubility and would adsorb on the surface of modified

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4 344 electrode. Third, the lucigenin was expensive. Considering all mentioned above, we  
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6 345 selected  $1 \times 10^{-4}$  mol L<sup>-1</sup> as the optimal concentration of lucigenin.  
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### 9 346 **3.7.3 Calibration Curve, Linear Range, and Detection Limit.**

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11 347 As shown in Fig. 5C, on conditions of the optimum, the ECL intensity was inversely  
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13 348 proportional to the concentration of BPA, ranging from  $1.0 \times 10^{-9}$  to  $1.0 \times 10^{-4}$  mol L<sup>-1</sup>,  
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16 349 and the regression equation was  $\lg(\text{ECL}) = -423.94 \lg C - 944.2$  ( $R^2 = 0.9975$ ), where  
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19 350 ECL is the ECL intensity, C is the concentration of BPA. The relative standard  
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21 351 deviation for  $1.0 \times 10^{-9}$  mol L<sup>-1</sup> BPA was 3.29% (n = 5). The detection limit (defined  
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23 352 as the concentration that could be detected at the signal-to-noise ratio of 3) was  
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26 353  $3 \times 10^{-10}$  mol L<sup>-1</sup>.  
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### 28 354 **3.7.4 Interference**

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31 355 The practical utility of a sensor often depends upon its selectivity, thus, the influence  
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33 356 of some potential coexisting ions and other compounds in biological sample were  
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36 357 investigated. The responses of 500-fold for Na<sup>+</sup>, SO<sub>4</sub><sup>2-</sup>, NO<sub>3</sub><sup>-</sup>, 5-fold for maltose, citric  
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38 358 acid, sucrose, and 3-fold for uric acid and ascorbic acid had no evident influence on  
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41 359 the determination of BPA, suggesting the novel ECL sensor had high selectivity  
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44 360 toward BPA from other coexisting species.  
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## 46 361 **4. Conclusion**

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49 362 A simple, low cost and high efficiency electrochemical method was used to transform  
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51 363 GO into ERGO successfully. And a composite film made of GO and CS was used to  
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54 364 fabricate modified gold electrode, and then conventional cyclic voltammetry was  
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56 365 posed on the modified electrode to produce ERGO-CS composite film modified  
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4 366 electrode. The new ECL sensor we proposed displayed supersensitive and selective  
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6 367 detection of BPA through a lucigenin ECL signal amplification mechanism. Due to  
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9 368 ERGO possessed better conductivity than GO and it played a great role in this  
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11 369 platform, thereby, it could greatly avoid the adsorption of electrochemical reaction  
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14 370 products on the electrode and greatly improve the sensitivity of detecting BPA.  
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16 371 Furthermore, this material could be also used in other areas, such as electrochemistry,  
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19 372 immunoassay and so on. Accordingly, this text present an important strategy to design  
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21 373 ECL sensors and expand their applications of ultratrace amount environmental  
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24 374 sensing.

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27  
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33  
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4 466 **Figure captions**

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6 467 **Scheme 1** Schematic representation of (A) the preparation of the ERGO-CS modified  
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8 468 electrode, (B) the mechanism of ECL emission of lucigenin and (C) the possible  
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10 469 quenched ECL mechanism by BPA.

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13 470 **Figure 1** (A) TEM image of GO in CS solution, XPS images of the GO-CS (B) and  
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15 471 REGO-CS (C) modified electrodes.

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18 472 **Figure 2** (A) Cyclic voltammograms of REGO-CS modified electrode (a), bare gold  
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20 473 electrode (b), and the GO-CS modified gold electrode (c) in 5 mmol L<sup>-1</sup> [Fe(CN)<sub>6</sub>]<sup>3-</sup>  
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22 474 solution containing 0.3 mol L<sup>-1</sup> KCl, scan rate: 0.1 V/s; (B) Cyclic voltammograms of  
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24 475 reduced GO-CS modified electrode in 5mM K<sub>3</sub>[Fe(CN)<sub>6</sub>] solution containing 0.3 M  
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26 476 KCl with different scan rates and the linear relationship between the peak currents and  
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28 477 the square root of the scan rates (the inset in B), scan rate (a-k) is from 500 to 10  
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30 478 mV/s; (C) Nyquist plots of (a) GO-CS modified, (b) ERGO-CS modified, and (c) bare  
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32 479 gold electrodes in 5 mmol L<sup>-1</sup> [Fe(CN)<sub>6</sub>]<sup>3-</sup> solution containing 0.01 mol L<sup>-1</sup> KCl,  
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34 480 frequency range: 0.1 to 10 KHz; (D) The ECL of different electrodes in 0.1 mmol L<sup>-1</sup>  
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36 481 KCl containing 20 μmol L<sup>-1</sup> lucigenin with the scan rate of 0.1 V/s, (a) ERGO-CS  
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38 482 modified gold electrode, (b) bare gold electrode, (c) GO-CS modified gold electrode.

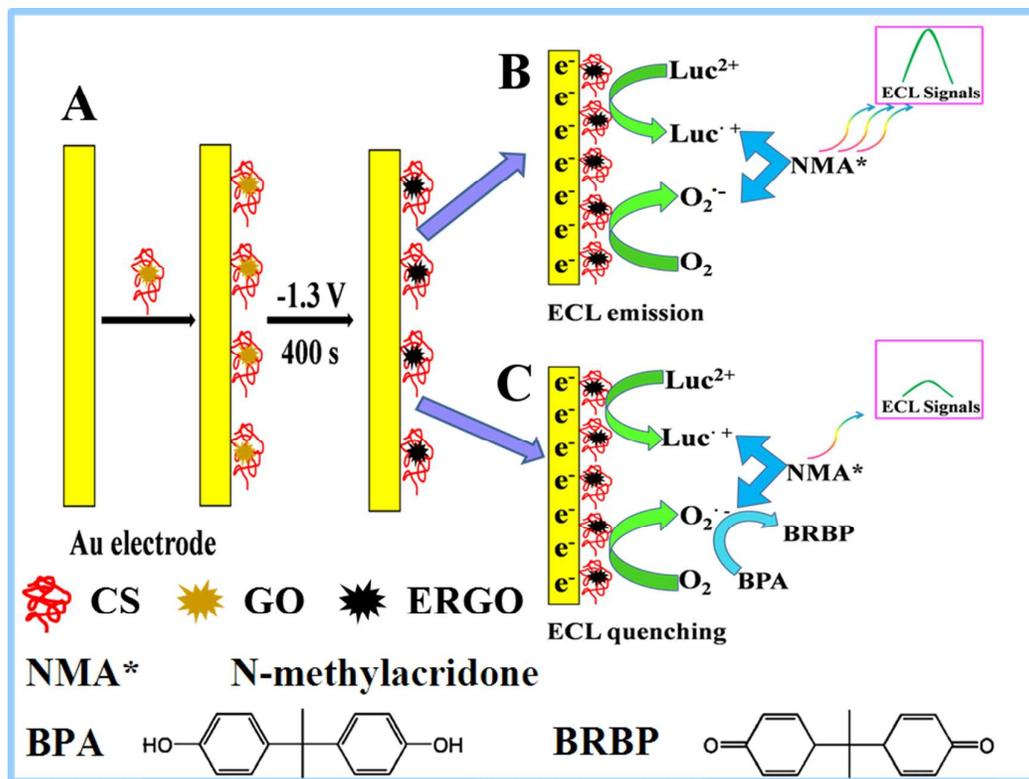
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41 483 **Figure 3** Effects of (A) the concentration of GO, (B) reduction potential, (C)  
42  
43 484 reduction time, (D) dispend volume, and (E) the concentration of CS on the ECL  
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45 485 intensity of ERGO-CS modified electrode in 0.01 mol L<sup>-1</sup> KCl containing 20 μmol L<sup>-1</sup>  
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47 486 lucigenin.

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50 487 **Figure 4** (A) Calibration plots of lucigenin at (a) ERGO-CS modified gold electrode  
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4 488 and (b) bare gold electrode, condition: KCl 0.1 mmol L<sup>-1</sup> KCl, Potential  
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6 489 window:-1.5-0 V, scan rate: 0.1 V/s; (B) ECL signals for duplicate measurements of  
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9 490 lucigenin at the concentration of 20 μmol L<sup>-1</sup> in 0.1 mmol L<sup>-1</sup> KCl on ERGO-CS  
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11 491 modified electrode with the scan rate of 0.1V/s.

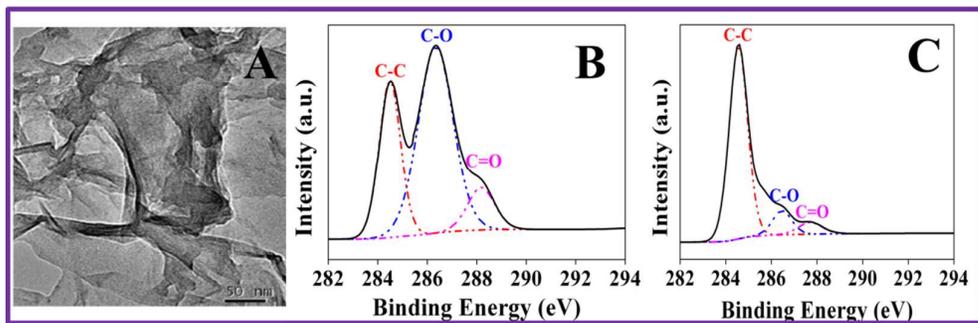
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14 492 **Figure 5** Effects of pH (A) and KCl concentration (B) on the lucigenin ECL  
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16 493 intensity. Condition: GO 1.5mg/ml with 0.3% CS, Reduction potential -1.3 V,  
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18 494 Reduction time 400s, Dispensed volume 3 μL ; (C) ECL responses of the proposed  
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21 495 sensor to different concentration (inter to outer is from 1 × 10<sup>-4</sup> to 1 × 10<sup>-9</sup> mol L<sup>-1</sup>) of  
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24 496 BPA ,and linear plot for BPA determination ( the inset in (C)).

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Scheme 1

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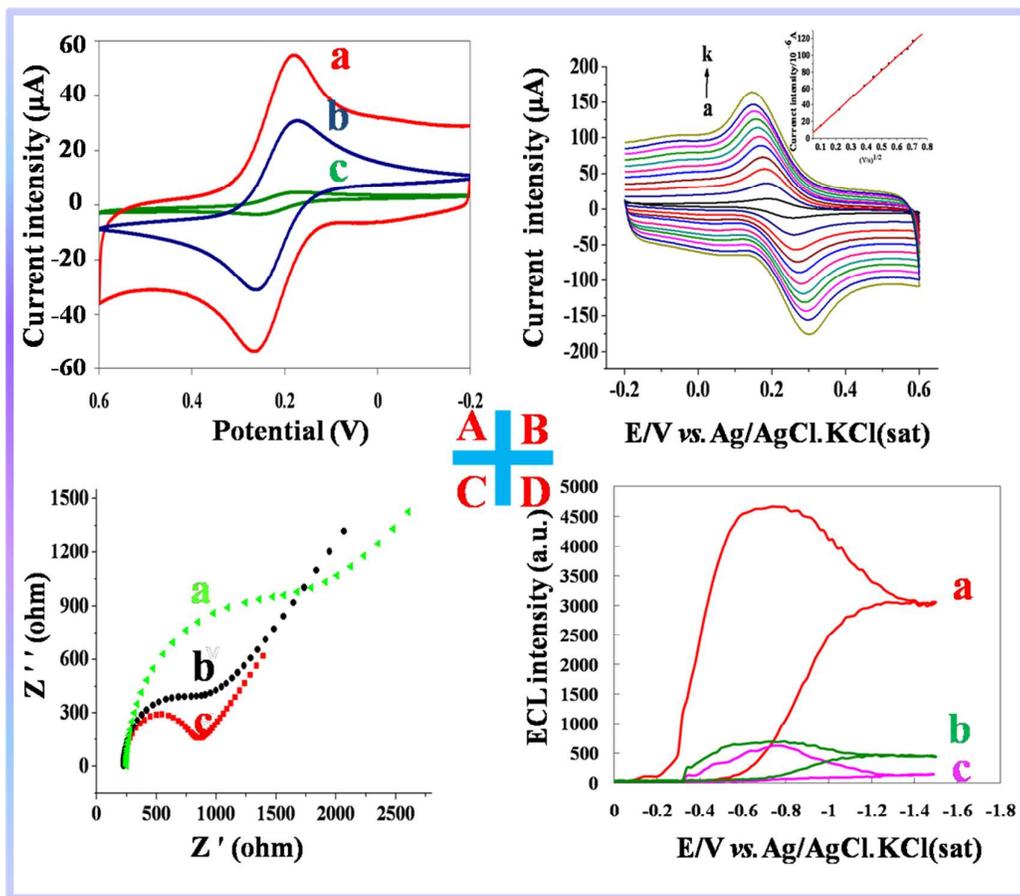


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Figure 1

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Figure 2

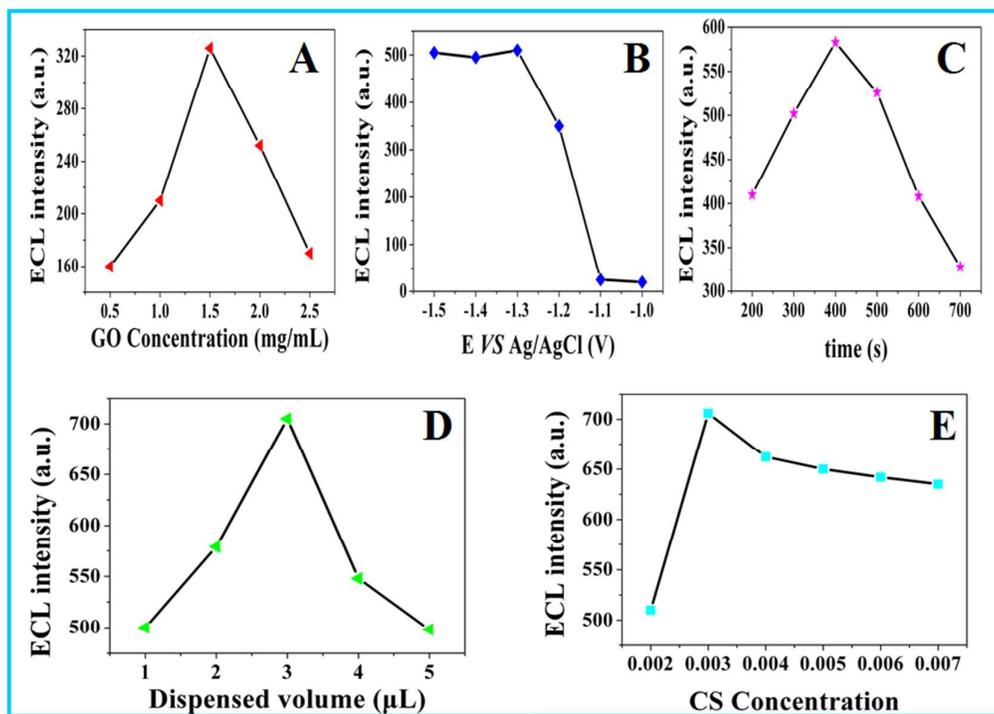


Figure 3

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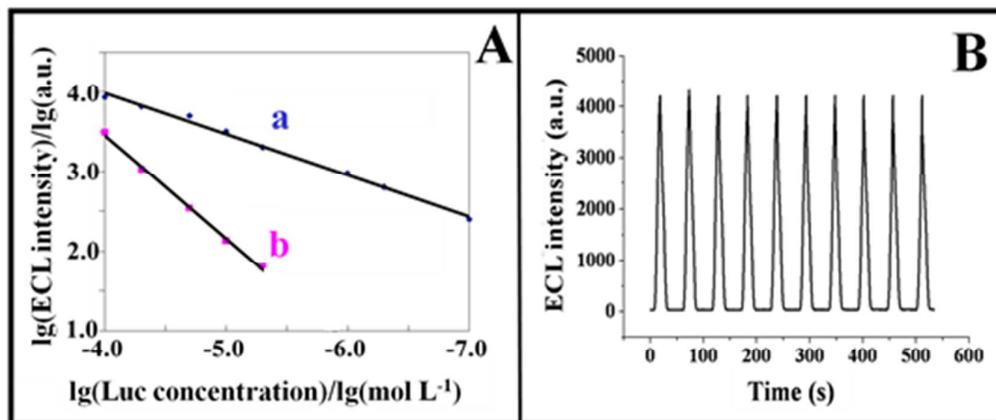


Figure 4

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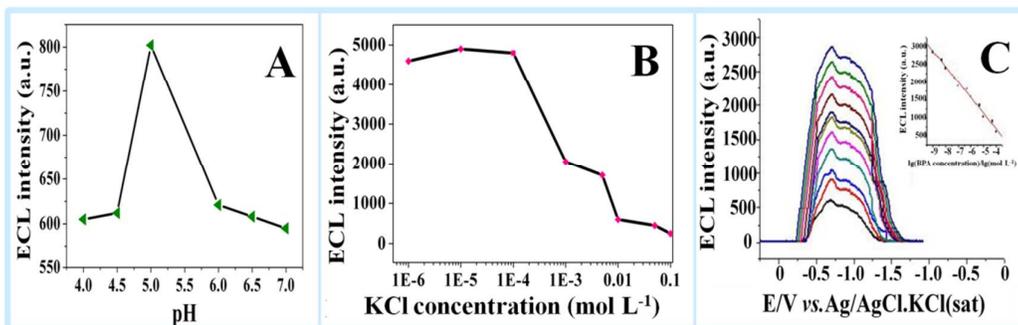
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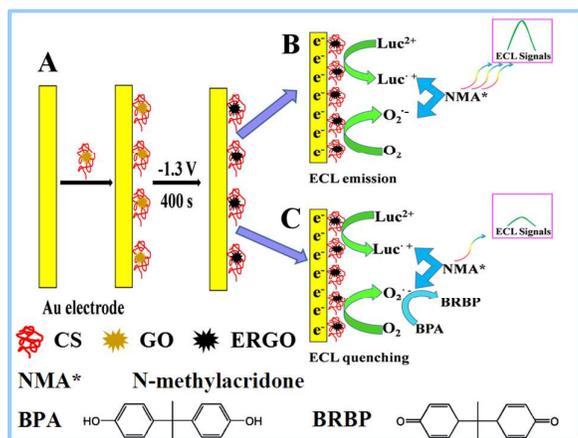
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3 536 An amplified and stable electrochemiluminescent signal of lucigenin on  
4 537 electrochemically reduced graphene oxide contained sensing platform was obtained  
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