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Role of silver nanoparticles and silver nanoclusters for the detection and removal of Hg(II)

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Silver metal, being a 3d transition metal in group 11 in the periodic table, is widely used in material science for its distinguished plasmonic properties. Nanoparticles (NPs) and nanoclusters (NCs) are widely used in sensing applications having a surface plasmon band and emissive properties, respectively. Mercury is one of the detrimental toxins and threats to various ecosystems. The distinction between nanoparticles and nanoclusters, the utility and toxicity of heavy metal mercury, fluorometric and colorimetric approaches to the recognition of mercury ions with NPs and NCs, the mechanism of detection, spot detection, and natural water sample analyses were illustrated in detail in this review article. Moreover, the sensing platform and analyte (Hg²⁺) fate were described for substantiating the mechanism. It was observed that NCs are mostly utilized for fluorometric approaches, while NPs are mostly employed for colorimetric approaches. Fluorometric detection is mainly quenching-based. However, sensing with enhancement was found in a few reports. Adulteration of other metals with silver particles often modifies the sensing platform.

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1. Introduction

Water contamination has become a global problem, and combating it requires a constant evaluation of water resource policies. Water pollution causes fatalities and illnesses worldwide, and approximately 14 000 people die daily as a consequence of it.¹ Water pollution is an issue in both established and developing nations. Weather, climate, soil type, flora, geology, flow conditions, groundwater, and human actions all have an impact on water purity. The most serious threat to water

purity comes from point sources of contamination, such as factories and towns. Water quality is influenced by mining, urbanization, and farmland. Other kinds of sources of contamination include sediments, nutrients, and poisonous contaminants.² Pollutants come in a variety of forms and have unique characteristics, such as storage pollutants including non-recyclable substances, polymers made from chemicals, and heavy metals.³

The following are some heavy metals with concentrations higher than 5 g cm⁻³ and are more prevalent daily. Some of the element names are Ti, V, Cr, Mn, Fe, Co, Ni, Cu, Zn, As, Mo, Ag, Cd, Sn, Pt, Au, Hg, and Pb.⁴ But mercury (Hg) is the most toxic metal, and diagnosing low-concentration of Hg is very

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challenging. Hg causes harmful effects on not only humans but also plants and animals. It is found that rivers are mostly contaminated with Hg. Among several sources of Hg, gold mining is the primary source of mercury pollution in worldwide coastal waters. Fossil fuels are also an important source. The convention of Minamata discussed mercury emissions into the atmosphere and the creation of methods to utilize less mercury.^{5,6}

Silver nanoparticles (AgNPs) and clusters (AgNCs) are well studied because of surface plasmons and intra/interbond transition.^{7,8} AgNPs are thus widely used for sensing applications based on colorimetry and fluorimetry. Sabela *et al.*⁹ reviewed colorimetric detection based on AgNPs. Lan *et al.*¹⁰ sensed ionic copper based on fluorescent AgNCs.

In our review article, we summarized the sensing of mercury calorimetrically and fluorometrically using AgNPs and AgNCs. We also illustrated the need for detection, the fate of silver particles and analytes, the mechanism of sensing, real sample analysis, spot detection for prototype applications, and the effect of adulteration.

2. Use of Hg²⁺

Hg is an element, that is liquid at the atmosphere's temperature. In different forms, Hg is used widely for biological, industrial, and domestic purposes.¹¹ In this review article, we focused on ionic mercury salts.

2.1 Cosmetics

Skin-lightening goods are available in a variety of forms such as lotions and soaps and are used without a doctor's guidance.¹² In cosmetic goods, Hg and Hg salts as HgCl₂ and HgO are utilized widely as skin-whitening substances.¹³ It was observed that the face treatments had a significantly greater Hg content than the body preparations,¹⁴ because Hg salts prevent melanin synthesis, producing a lighter complexion.¹⁵ Tyrosinase is necessary for producing melanin, a key component responsible for human skin color. Chen *et al.*¹⁶ discovered that HgCl₂ may directly block the tyrosinase. The sample results, obtained in the research literature, were 25.7 mg kg⁻¹ mercury

concentration in low-cost whitening cream products. Vermilion is a red color powder, made from mercury sulfide.¹⁷

2.2 In reaction

Metals have been applied frequently as catalysts for organic processes by chemists. Hg had been one of the earliest catalysts.¹⁸ Hg²⁺ acts as a catalyst in many organic reactions such as the hydroamination of alkynes¹⁹ and the amination of allyl alcohols.^{20,21} The synthesis of complex cyclic structures is very difficult. A large number of cyclizations are produced *via* transition-metal salts. Complex heterocyclic and carbocyclic compounds, which are hard to manufacture with other transition metal salts, have been successfully synthesized *via* cyclizations involving Hg(II) salts.²² Methane is converted into methanol not just in the presence of HgCl₂.²³ But mercuric sulfate is also used as a catalyst.²⁴

HgCl₂ can be used to stabilize chemical and analytical samples. Foreman *et al.*²⁵ demonstrated that continuous liquid-liquid extraction or closed-loop stripping produced organic components, recognized *via* analytical interferences of HgCl₂ preservatives in natural water.

2.3 Disinfecting agent

Surface sterilization is a key step in the creation of healthy and sustainable tissue culture explants.²⁶ Different amounts of HgCl₂ are employed for various lengths of time during plant tissue culture. Optimizing quantities is necessary to minimize contamination. *Clinacanthus nutans* is a medicinal herb used in various disease treatments.²⁷ *Clinacanthus nutans* is sterilized using a variety of agents. However, 0.2% HgCl₂ is the most efficient sterilization solution for sanitizing nodal explants of *Clinacanthus nutans*, used for *in vitro* growth.²⁸ *Aspidosperma polyneuron* is a high-valued wood tree, that gives nearby people a stable source of income. For contaminant control, 10 min of HgCl₂ (0.125%) disinfection was sufficient.^{28,29} To decrease microbial contamination, HgCl₂ is frequently used for surface sterilization in sugarcane tissue culture.³⁰

HgCl₂ is often typically employed in laboratories to destroy germs on explants due to its high sterilizing efficacy. Paul Das *et al.*³¹ performed a comprehensive study on the role of HgCl₂ as a disinfecting agent in the mixed culture.³¹ HgCl₂ is still used successfully as a disinfection solution on several forest and firewood species, that have major contamination concerns. Phenyl-mercury-nitrate is a disinfectant and antiseptic agent. It is non-toxic to animal tissue and may be applied to the skin and relieve wound healing.²⁹

2.4 Sensing platform

Hg is a wearable, wireless sensor platform for individuals with neuromotor illnesses including epilepsy, Parkinson's disease, and stroke to analyze their movement.³²

Hg²⁺ is also used in various sensing applications in the laboratory. For example, Askari *et al.*³³ synthesized graphene carbon dots with 1.8 nm size with high emission intensity, which was quenched selectively and sensitively with the help of



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Hg²⁺. Quenched fluorescence was restored selectively with thiocyanate (λ_{ex} 360 and λ_{em} 461 nm) to sense thiocyanate.

2.5 Energy source

India's coal has extremely high mercury levels. Indian coal has higher amounts of Hg than that from other nations. Hg, a common persistent worldwide toxin, is emitted from coal burning the most frequently in India. 50% of the 5500 tonnes of mercury, emitted into the environment annually come from coal-fired power stations. Mercury exists in three different forms in coal-burning power plant systems: pure Hg, Hg²⁺, and particulate Hg. Volatile Hg is present in large amounts in the geothermal steam, used to generate electricity. In cooling tower exhausts, a large portion of this Hg is released into the environment as pure Hg vapor. On a per megawatt (electric) basis, Hg emissions from geothermal power plants are equivalent to those from coal-fired power plants. According to estimations based on measurements from a few chosen power plants, Indian coal ash has an average mercury content of 0.53 mg kg⁻¹. Coal fly ash tends to have a greater concentration of Hg upon burning.^{34,35}

2.6 Gold mining

Small-scale gold extraction operations employ Hg in >50 poor nations. The discharges, that go along with it, have an impact on the planet and human health. Instead of a true alloying process, the wetting of Au by Hg is a phenomenon of somewhat deep sorption, involving some interpenetration of the two elements. Because the surface tension of Hg is lower than that of Au and greater than that of water, it sticks to the surface of Au particles. Hg also acts as a thick medium to sink Au, causing the lighter gangue material to float on the top. Only 0.06% of Au is soluble in Hg at 20 °C. Numerous amalgamation processes are

used in artisanal mining operations because amalgamation is simple to use and understand (Fig. 1).^{36,37}

When mercury is introduced to the grinding circuit, the greatest losses of Hg per unit of gold produced take place. Amalgamation employing miller mills in China results in losses of 14 to 20 parts of mercury for every part of generated gold (Fig. 2).³⁹

2.7 Others

Mercury is widely used in various preparations such as anti-bacterial agents, skin antiseptics diuretics, laxatives, and other ointments.¹⁸

Mercury is used in various forms and compositions for dental amalgam.⁴⁰ Mercury also has practical use in the study of porous solids. Determining the pore size distribution by mercury porosimeter is a well-established practice in the chemical industry.⁴¹

Inorganic mercury ions can be found in nature in a variety of colors, spanning from colorless oxides to browns and blacks with sulfide compounds. They are also widely employed in manufacturing. They have been used, as skin-brightening lotions, tooth eruption, and as antioxidants in some medications, most notably eyedrops. In the shape of the 'blue pill', HgCl (calomel) was extensively used as a purgative.⁴²

3. Nanocluster vs. nanoparticles

The distinction between metal nanoclusters (NCs) and nanoparticles (NPs). Because of their higher surface area (1–100 nm), nanoparticles have special characteristics. Surface plasmon resonance (SPR), which happens when light interacts with the electrons on the nanoparticle's surface, is one such characteristic. In contrast, metal nanoclusters are just tens to hundreds of atoms in size and are significantly smaller (around 2 nm). Due to their small size, nanoclusters have characteristics more akin to molecules, such as luminescence and catalysis, but lack the surface area required for surface plasmon resonance (SPR). In essence, nanoclusters act as a link between the actions of bigger nanoparticles and individual atoms.

4. AgNPs vs. AgNCs

The range of uses that silver nanoparticles (AgNPs) and nanoclusters (AgNCs) can have. AgNPs have several biological uses, including antibacterial and anti-inflammatory effects. They exhibit potential in catalysis and sensing as well. AgNPs have the additional ability to improve Raman signals for low-concentration molecule detection. On the other hand, AgNCs behave like molecules since they are smaller than 1 nm. Unlike bulk silver or AgNPs, they radiate strongly and absorb light across a wide range. Because of their compact size and high brightness, this special characteristic enables the development of accurate, adjustable fluorescent labels for bioimaging and sensing applications.

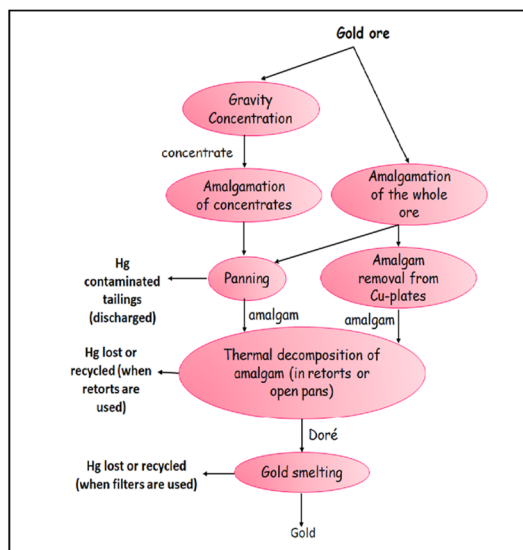


Fig. 1 Techniques for extracting gold using Hg in artisanal mining processes.

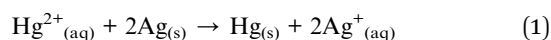




Fig. 2 Miners in Sulawesi, Indonesia, add one kilogram of mercury to each ball mill when kids play nearby the operational area. Reproduced with permission from ref. 38, copyright 2024, *J. Clean. Prod.*

5. Silver–mercury interaction

It is commonly known that, at the bulk scale, zero-valent mercury is produced by the stoichiometric 1:2 interaction between aqueous mercury(II) and silver metal(0) [eqn (1)].



The potential applications of the hyperstoichiometric interaction between mercury and silver at the nanoscale are significant since they span a variety of sectors. Hg^{2+} from water is reduced onto the AgNPs as a result of this interaction, which happens when the diameter of the AgNPs is lowered below 32 nm. For 11 nm AgNPs, the mercury-to-silver ratio reaches 1.125:1. This phenomenon has great potential for novel perspectives on nanoscale chemistry and substantial progress in wastewater treatment, improved chemical catalysis, and nanoscale system toxicity. It has been established that the hyperstoichiometry effect is related to the size of the AgNPs and occurs for both mercury nitrate and mercury acetate. Perhaps because nanosized silver particles have a higher surface energy than their bulk scale counterparts, the smaller AgNPs exhibit improved release and catalytic recycling of the silver ions released into the solution.⁴³

Several characterization techniques, including UV-vis spectroscopy, dynamic light scattering (DLS), zeta potential measurement, elemental mapping, and X-ray photoelectron spectroscopy (XPS), are used in the mechanism of direct SERS detection of Hg^{2+} without tagging. These characterizations show that the formation of a non-uniform mercury/Ag shell wrapping around AgNPs is the result of the rapid interaction between Hg^{2+} and AgNPs.⁴⁴

AgNPs' surface plasmon resonance (SPR) characteristic is impacted by this interaction, which lowers the substrates'

electromagnetic enhancement. AgNPs' zeta potential also decreases as a result of the contact, which has an impact on Raman reporter molecules' ability to adhere to AgNP surfaces. The XPS test findings demonstrate that the citrate has decreased Hg^0 and Hg^{2+} bound to it. Strong support for the mechanism of direct SERS detection of Hg^{2+} without tagging is provided by these experimental results.⁴⁴

Sun *et al.*⁴⁵ primarily offer a methodical theoretical investigation of Hg binding on the small neutral and charged Agn cluster ($n = 1-6$) using the DFT approach, as both supported and free Ag clusters are frequently employed as regenerable sorbents in real-world Hg removal research.

The density functional theory of mercury adsorption on a sequence of charged and neutral Agn ($n = 1-6$) clusters. The findings show that the charged state and cluster size have a major impact on the adsorption of mercury. The BEs rise and subsequently fall on the neutral clusters. Except for Ag, the binding energies for negatively charged Agn clusters exhibit an odd-even pattern, whereas the BEs for cationic clusters drop as cluster size increases. According to NBO analysis, in the case of anionic and neutral compounds, additionally, electrons go from the p orbital of mercury to the s orbital of silver. The prediction of Hg binding orientation for neutral and charged anions satisfactorily validates the rules of LUMO (HOMO for anions) established by Sun *et al.* (Fig. 3).⁴⁵

The surface plasmon mode shifts blue as a result of Hg adsorption on AgNPs. For smaller particles, the blue shift's magnitude is more noticeable.⁴⁶

6. Removal of mercury and adsorption via silver particles

A novel method for preconcentrating water by filtration and employing paper treated with NPs to detect Hg^{2+} in an



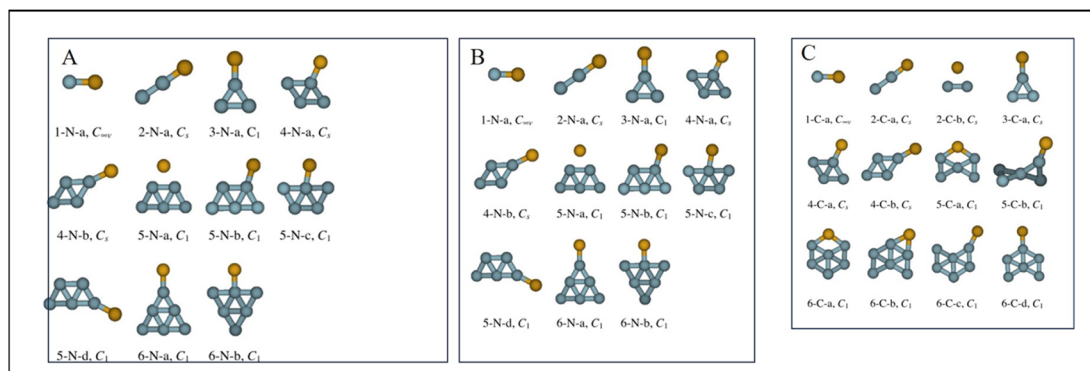


Fig. 3 (A) Optimized structures of neutral AgnHg complexes, $n \leq 6$. The symmetry point group is indicated. (B) Optimized structures of anionic AgnHg complexes, $n \leq 6$. The symmetry point group is indicated. (C) Optimized structures of cationic AgnHg complexes, $n \leq 6$. The symmetry point group is indicated.⁴⁵ Reproduced with permission from ref. 45, copyright 2024, *Chem. Phys. Lett.*

ultrasensitive manner. By reducing Hg^{2+} to Hg^0 without using chemical reagents, this approach amalgamates Hg^0 utilizing cellulose substrates that include *in situ*-synthesized AgNPs. The enhanced filters are then placed inside a direct mercury analyzer so that atomic absorption can determine the mercury content. The various NP synthesis techniques, filtering methods, filter paper properties, and other pertinent factors affect Hg solid-phase extraction.⁴⁷

By using filter paper that has been treated with AgNPs, the technique enables the direct analysis of mercury without the need for further digestion or desorption processes. This makes it possible to determine mercury at ultra-trace levels and streamlines the preconcentration procedure. Furthermore, for a 100 mL sample volume, the approach achieves a preconcentration factor of about 3500, with a limit of quantification (LOQ) of 0.8 ng L^{-1} and a limit of detection (LOD) of 0.2 ng L^{-1} . Relative standard deviation (RSD) represents the method's accuracy, which is 5% for repeatability and 9% for reproducibility. Moreover, the technique shows excellent selectivity for Hg^{2+} sorption, with recoveries for synthetic waters and verified reference materials ranging from 92% to 105%.⁴⁷ An inventive method for the synthesis of AgNPs using lignin as a reducing and stabilizing agent under solar light was presented by Zhang *et al.* The AgNPs were tangled in lignin after being placed onto the surface of lignin nanoparticles (LNPs) with an average size of around 14 nm and a consistent shape.⁴⁸

When it came to Hg^{2+} detection, the AgNP-LNP solution exhibited excellent sensitivity and selectivity. The ultra-sensitive colorimetric reaction to Hg^{2+} , which had a detection limit of 1.8 nM in tap water samples—much lower than the drinking water threshold of 10 nM for Hg^{2+} —demonstrated the sensitivity of the method. Additionally sensitive and selective for on-site Hg^{2+} detection in actual water samples was the AgNP-LNP solution. The absorbance results demonstrated the selectivity of the AgNP-LNP suspension, which had a stronger affinity for Hg^{2+} than other metal ions. As a result, it was a very useful sensor for detecting Hg^{2+} in real-world samples.⁴⁸

The adsorption capacity of synthetic zeolite (Na-X and Na-P1) generated from coal fly ash was insignificant for Hg, but it could

be significantly increased for Hg^0 after being impregnated with silver.^{49,50} Wang *et al.* reported ionic mercury adsorption and removal using organic framework passivated AgNPs from wastewater, acidic in nature (Table 1).⁵¹

7. Fluorometric detection concept

Electrical stimulation, which occurs during light absorption by the fluorophore, causes light to be emitted. When a molecule remains in the excitation state after light absorption, it produces fluorescence emission, which has a longer wavelength and lower energy.⁵² Fluoroionophore is one of the light sensor's luminescence and ionophore components.⁵³ Recognition and signaling systems are the primary components under evaluation. The former is in charge of discrimination and binding effectiveness, while the latter transforms information into an optical signal.⁵³ When the fluorescent sensor interacts with cations, variations in the photophysical properties of the fluorophore cause a variety of processes, including charge transfer, energy transfer, excimer formation, and so on.⁵⁴ The combination of silver particles and fluorescent units has increased sensor portability, stability, sensing range, and general performance (Fig. 4 and 5).⁵⁵

7.1 Fluorometric detection of Hg^{2+} with the silver nanocluster

7.1.1 Enhancement-based detection. In enhancement-based detection, fluorescent AgNCs can be formed *via* various biomolecules such as cytosine-rich DNA, amino acids, *etc.* DNA has several advantageous features for metal detection. DNA is incredibly reliable, cheap, susceptible to combinatorial selection, and simple to change. Aptamers and DNAzymes are the two primary functional DNA categories, created for metal sensing. A few metal-binding aptamers are known, although separating them is often quite challenging.⁵⁶

Deng *et al.*⁵⁷ formed a thymine–cytosine-rich DNA duplex for the formation of AgNCs. The interaction of DNA with Mercury gives stability to the DNA duplex and the configuration of a cytosine-rich loop (The λ_{ex} and λ_{em} were at 558 and 625 nm,



Table 1 Ionic mercury adsorption by AgNPs

NPs	Size	Hg removal capacity	Type of adsorption	Time	Rate constant
Covalent organic framework-AgNPs ⁵¹	200 nm	113 mg g ⁻¹	Langmuir	30 min	Pseudo-second-order

respectively). The thymine–cytosine-rich fluorescent probes work satisfactorily for mercury detection. Hg²⁺ can be detected by the lower limit concentration of 10 nM. The strong fluorescence shows from pH 5 to 7.⁵⁸

A quick and easy method for detecting metal ions is proposed by oligonucleotide-based radiant technologies, allowing for onsite detection and measurement of the metal-ion concentration.⁵⁹ Oligonucleotide probes, having -C6G5C6, ligated to AgNPs act as a capping agent. Guanines were introduced into cytosine fragments to enhance the fluorescence of AgNPs. DNA-capped AgNPs showed enhancement with Hg²⁺, while Cd²⁺ showed quenching. The advantage of this method has a lower LOD.⁶⁰

Amino acid and its components have a large active surface. So, amino acids are a promising platform for heavy metal sensing.⁶¹ Tryptophan is a type of amino acid and it is fluorescent. Bian *et al.*⁶² prepared tryptophan-capped AgNPs. Amino acid functionality and nitration of tryptophan give stability to the AgNPs. AgNPs were highly fluorescent (λ_{ex} 330 nm and λ_{em} 465 nm) and enhancement happened with Hg²⁺ selectively (LOD 0.658 pM and linear detection range 10 mM to 1 pM).

7.1.1.1 Mechanism based on enhancement. T–Hg²⁺–T pairing: Many novel methods for DNA-based Hg²⁺ sensors are now being developed as a result of the complementary interaction between thymine bases and mercury ions without the aforementioned restrictions. Thymine–thymine (T–T) mismatched DNA duplexes would particularly bind Hg²⁺ in an aqueous solution to produce stable T–Hg²⁺–T pairing DNA duplexes. T–Hg²⁺–T has

a binding constant, almost 4.14×10^6 times higher than an adenine–thymine base pair found in nature. Furthermore, the T–T base pair can only be stabilized by Hg²⁺ in this kind of T–Hg²⁺–T coupling, ensuring remarkable selectivity.^{63–66}

Release of self-quenching: A process in which the intensity of fluorescence emission is decreased is known as fluorescence quenching. Fluorescence quenching may be caused by several events, complex formation, electron transport, excited state reaction, and energy transfer.^{67,68}

A distinctive type of fluorescence quenching is self-quenching. In right-angle geometry, strong fluorescence self-quenching of highly concentrated fluorophore solutions is seen.^{69–71} The re-absorption effect is the main cause of fluorescence self-quenching. Moreover, the resonance energy transfer of the Förster type (home-transfer) is also an aspect. Self-quenching occurs when toluene has a high ambient tetraphenyl-porphyrin concentration (300 K).⁷¹ Water volume variations in lipid vesicles may be measured using self-quenching characteristics. At relatively high (mM) concentrations, self-quenching happens.⁷² Tryptophan's nitrosylation was important for the detection of Hg²⁺ ions. Trp-NO₂-AgNPs had the ability to self-quench. Such quenching was released after adding Hg²⁺ to the solution of AgNPs. A high static electrostatic repulsion was formed between AgNPs due to the binding of Hg²⁺, increasing the distance of AgNPs.⁶²

7.1.2 Quenching-based detection. The fluorescence probe is fluorescent AgNPs, templated with dihydroliipoic acid,^{73–75} guanine-rich DNA,⁷⁶ cytosine-rich DNA,^{77,78} polyinosinic, or

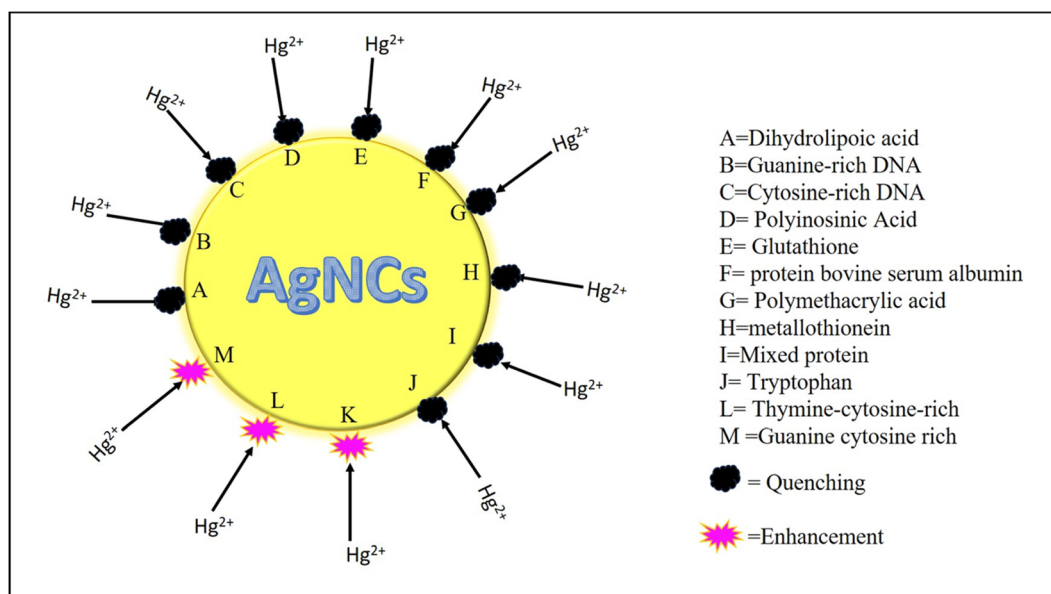


Fig. 4 Fluorescence-based sensing of mercury with AgNPs, with different capping agents.



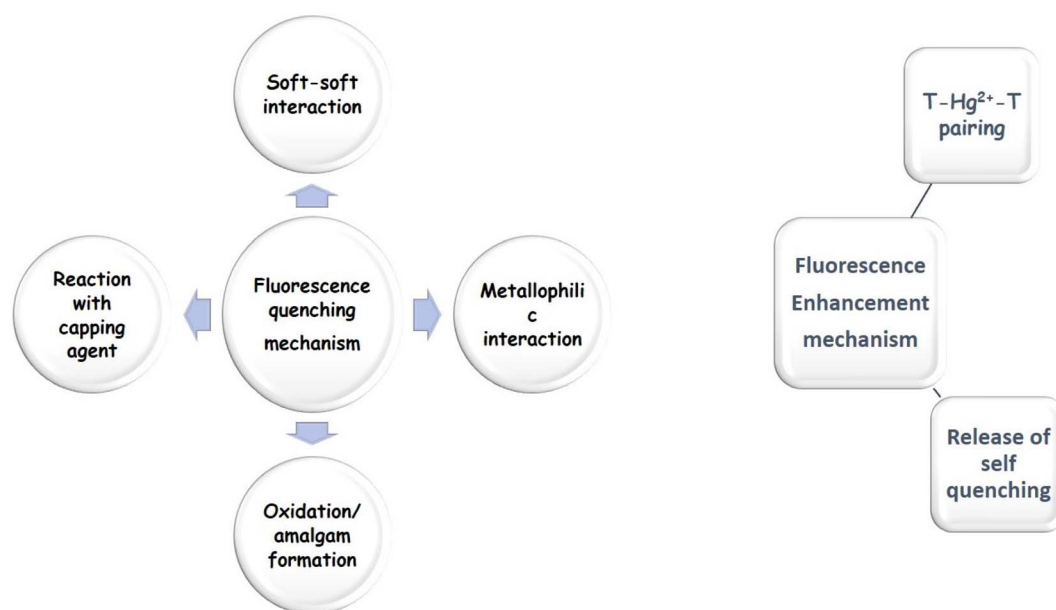


Fig. 5 Mechanism of fluorometric detection of Hg^{2+} with AgNCs.

polycytidylic acids, glutathione,^{79,80} protein bovine serum albumin,⁸¹ polymethacrylic acids,⁸² metallothionein (cysteine-rich protein),⁸³ mixed protein (bovine-serum-albumin and lysozyme)⁸⁴ in the context of quenching-based Hg^{2+} detection (Table 2). Lipoic acid is a natural antioxidant that interacts with harmful metals and adsorbs free radicals.⁷³ Lipoic acid is used as a stabilizing agent because lipoic acid is a little molecule with two thiol groups. In the presence of ultrasound conditions, silver may be reduced. The s-s linkage may rupture and convert into dihydrolipoic acid in lipoic acid.^{74,75} Hg^{2+} quenched the emission of dihydrolipoic acid-stabilized AgNCs selectively and sensitively. The Stern-Volmer graphic, shown by Bayen *et al.*⁷⁴ made it obvious that Hg^{2+} had a substantially steeper slope than the other metal ions. This demonstrated that the AgNCs were focused on detecting Hg^{2+} (Fig. 6 and 7).⁷⁴

DNA oligomers are especially intriguing templates for creating AgNCs because the DNA sequence may be changed to produce a variety of emission colors. Hg^{2+} caused a substantial quenching of the red peak (of DNA13 passivated AgNCs) and

a concurrent rise in the green peak, resulting in a visible fluorescence transition from orange to green.⁸⁶

7.1.2.1 Mechanism based on quenching. Soft-soft interaction: The extremely high lipophilicity of mercury may be the cause of its selective activity in the context of soft-soft contact (soft acid and soft base interaction: SASB). Due to SASB, mercury removed silver from the system^{74,75} Naaz *et al.* identified similar principals for their “knock-out” sensor of Hg^{2+} with 2–6 atom AgNCs.

Reaction with a capping agent: Carboxylic acids of the lipoic acid-capped AgNCs had the greatest affinity towards Hg^{2+} destabilizing AgNCs and fluorescence was quenched as reported by Bayen *et al.* ($\log b_4 = 17.6$, where b_4 is the complex formation constant).

Metallothionein (MT), a low-molecular-weight protein, has a unique dumbbell-shaped structure with a stable C-terminal α -domain and a reactive N-terminal β -domain. Because of the thiolate ligands in MT, Zn^{2+} in the α -domain was kept in its tetrahedrally coordinated state. However, Zn^{2+} in the β -domain

Table 2 AgNCs, with different capping agents, to sense Hg^{2+} via quenching-based detection

Capping agent	λ_{ex} and λ_{em} (nm)	LOD	Linear detection range	References
Dihydrolipoic acid	λ_{ex} 330 nm/ λ_{em} 448 nm	0.1 nM	—	74
Guanine-rich DNA	λ_{ex} 480 nm/ λ_{em} 560 nm	2.1 nM	6.0–160 nM	76
Cytosine-rich DNA	λ_{ex} 724 nm/ λ_{em} 768 nm	1.90×10^{-9}	1.90×10^{-9} to 1.90×10^{-8} M	77
Polyinosinic	λ_{ex} 545 nm/ λ_{em} 585 nm	3.2 nM	0.05–10 μM	85
Polycytidylic acids	λ_{ex} 500 nm/ λ_{em} 575 nm	9.0 nM	0.5–10 μM	85
Glutathione	λ_{ex} 527 nm/ λ_{em} 700 nm	0.1 nM to 10 mM	126–245 nM	79
Protein bovine serum albumin	λ_{ex} 400 nm/ λ_{em} 637 nm	10 nM	10 nM–5 μM	81
Polymethacrylic acids	λ_{ex} 510 nm/ λ_{em} 610 nm	10 nM	10 nM–20 μM	82
Metallothionein	λ_{ex} 385 nm/ λ_{em} 475 nm	—	0.2–5.0 μM	83
Mixed protein	λ_{ex} 470 nm/ λ_{em} 620 nm	0.7 nM	0–1 μM	84



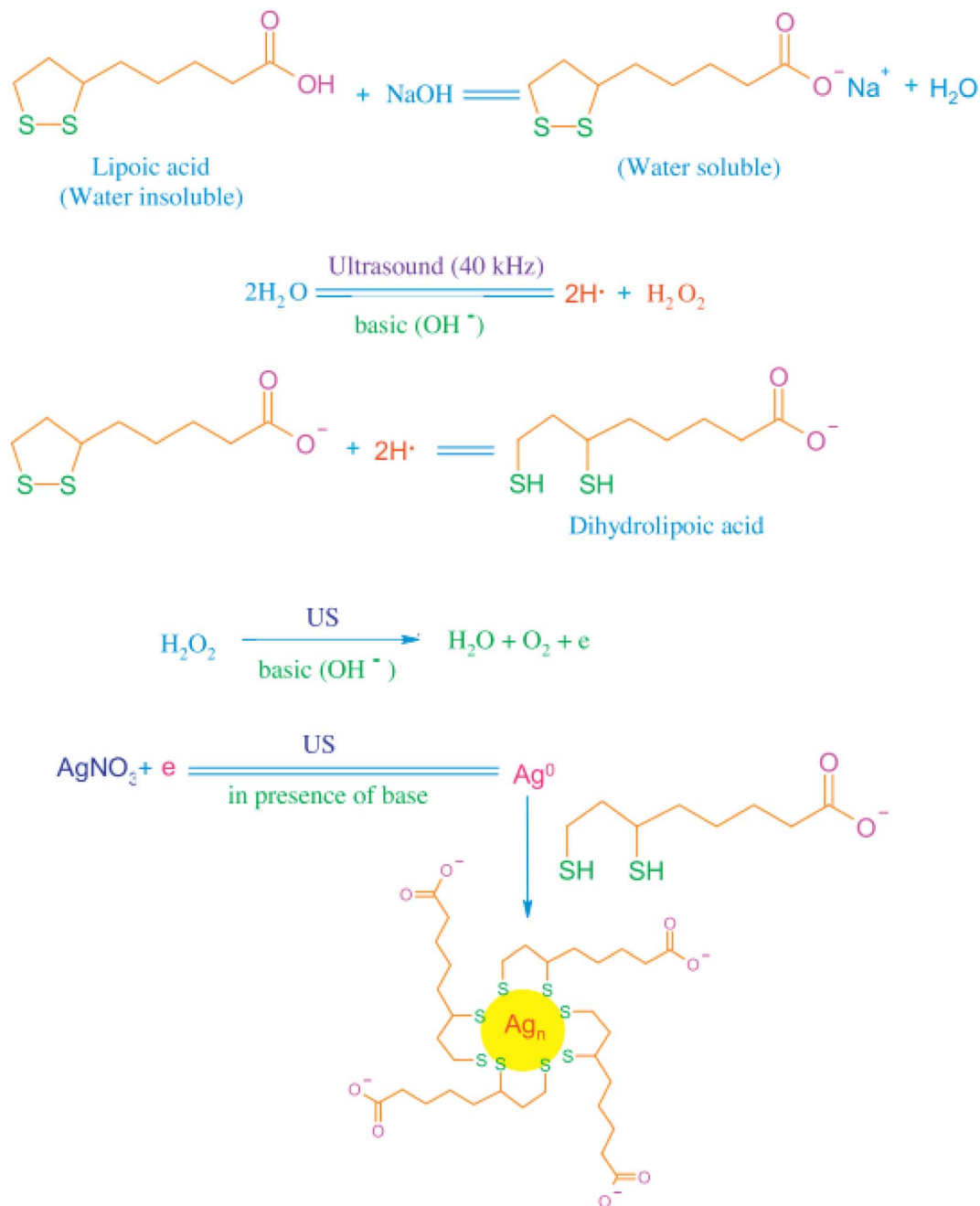


Fig. 6 Synthesis protocol for the preparation of water-soluble fluorescent AgNCs. Reproduced with permission from ref. 74, copyright 2024, *J. Environ. Chem. Eng.*

was preferentially replaced by silver, which was then reduced by sodium borohydride to create AgNCs.

The energy transfer between the AgNCs and the Hg^{2+} cysteine thiolates in the β -domain may allow coordinated Zn^{2+} in the α -domain to be exchanged for Hg^{2+} . The decrease in fluorescence emission was caused as a result. Fluorescence quenching was the result of the energy transfer between the AgNCs and the Hg^{2+} -cysteine thiolates in the α -domain.⁸³

Oxidation/amalgam formation: AgNCs were partially oxidized to Ag^+ due to electron withdrawal *via* Hg^{2+} ions with

amalgam formation. The emissive property of Hg^{2+} was substantially quenched.⁷⁴

Metallophilic interaction: Weak electrostatic interactions with low-valent closed shells $[(n-1)d^{10}ns^0]$ or pseudo-closed shells $[(n1)d^8ns^0]$ metal ions are referred to as metallophilic interactions. It is a well-known phenomenon that metallophilic connections can form in the structural chemistry of gold(i) derivatives, and it most likely results from dispersion forces that are amplified by relativistic effects. These interactions, whose strengths have been likened to hydrogen bonds, have a range of 2.8–3.5 Å. These interactions are predicted to happen in the



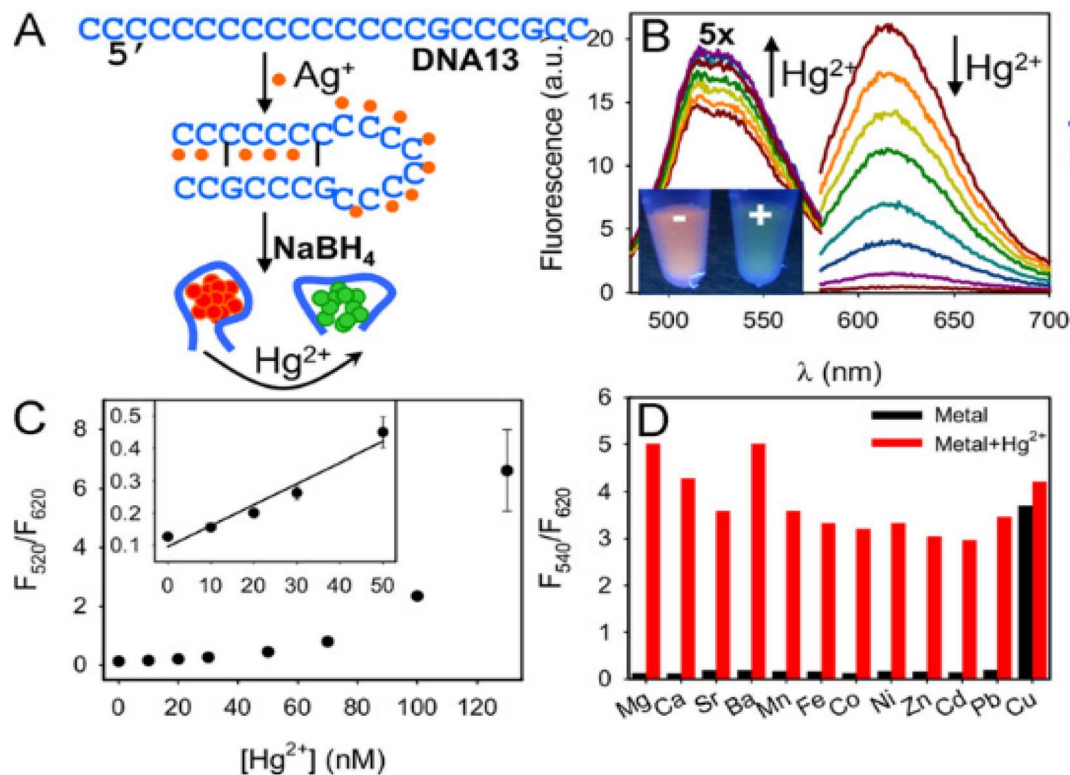


Fig. 7 (A) Synthesis of DNA13 passivated Ag clusters, (B) fluorescence spectra after the addition of Hg²⁺, (C) linear detection range, and (D) effect of competing metal ions. Reproduced with permission from ref. 86, copyright 2024, *Biosens Bioelectron.*

chemistry of Hg²⁺, which likewise has a 5d¹⁰ closed-shell electronic structure. These interactions were later found in a wide range of heterometallic/homometallic couples with supported or unsupported backbones.^{84,87–89}

Many research groups^{79,81,84} pointed out metallophilic interaction for Hg²⁺-induced quenching. Guo *et al.* illustrated 5d¹⁰(Hg²⁺)-4d¹⁰(Ag⁺) metallophilic interaction for protein-passivated AgNCs with Hg²⁺ for fluorescence quenching. In cytosine-rich DNA, fluorescence quenching resulted from high d¹⁰-d¹⁰ metallophilic interactions, 5d¹⁰(Hg²⁺)-4d¹⁰(Ag⁺).

7.2 Fluorometric detection of Hg²⁺ with silver nanoparticles

7.2.1 Enhancement-based detection. The most common organic cofactor found in nature is riboflavin (vitamin B2). It may exist in three different redox states: oxidized-electron, reduced, and completely reduced. Riboflavin is a fluorescent molecule. When it binds with AgNPs, fluorescence is quenched. However, in the presence of Hg²⁺, the fluorescence is enhanced.⁹⁰

Ganguly *et al.*⁹¹ prepared Schiff base from salicylaldehyde and 1,3 propylene-diamine. Such Schiff base worked as a capping agent for AgNPs, exhibiting silver-enhanced fluorescence (SEF). SEF was quenched selectively by dopamine and restored selectively by Hg²⁺. The introduction of Na₂-EDTA caused quenching due to the EDTA-Hg²⁺ adduct.

7.2.1.1 Mechanism-based on enhancement-based detection. In a turn-on situation thymine(T)-Hg-thymine(T) bond is

observed. So, Hg²⁺ selectively increased the fluorescence intensity.⁹⁰

7.2.2 Quenching-based detection. Kraithong *et al.*⁹² designed a Hg²⁺ selective fluorescence silver-based sensor using a tripodal ligand coupled to three 7-nitrobenzol-2-oxa-1,3-diazolyl (NBD) molecules (TNBD). With a LOD of 12.5 ppb, this sensor showed strong Hg²⁺ selectivity against other interfering ions.

Deng *et al.*⁹³ designed DNA-capped fluorescent AgNPs, which were highly selective for Hg²⁺, because of Ag/Hg amalgam formation (LOD 2.6 nM) (Fig. 8).

7.2.2.1 Mechanism-based on quenching. Reaction with a capping agent: The Fluorescence of NBD-capped AgNPs is drastically quenched by Hg²⁺ due to complexation with NBD. Having the paramagnetic properties of Hg²⁺, the electronic environment of NBD was changed leading to a turn of fluorescence.⁹²

It is also essential to know how Ag and Hg atoms interact chemically. Interaction is usually redox in nature. With Hg²⁺, nanoclusters lose their fluorescence by being oxidized and aggregated.

X-ray photoelectron spectroscopy was used to investigate the distinct and powerful interaction between AgNCs and Hg²⁺, which is caused by dispersion forces between closed-shell metal atoms. The silver 3d spectra were de-convoluted into components at 369.582 eV (Ag 3d_{5/2}) and 375.720 eV (Ag 3d_{3/2}), which are indicative of Ag ions when Hg²⁺ was introduced to the DNA-AgNCs. Hg endured the process as a doubly charged particle.



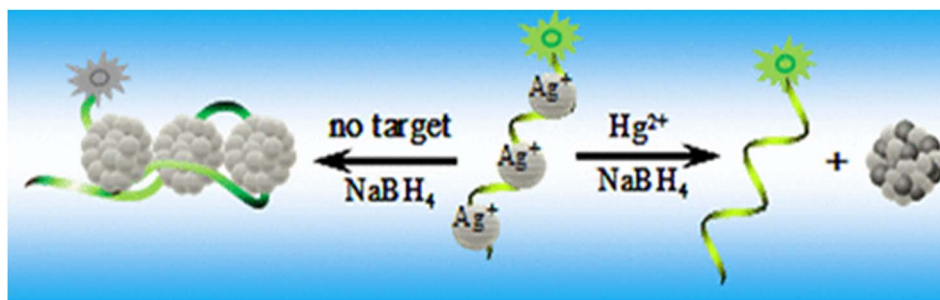


Fig. 8 Quenching of fluorescence with Hg^{2+} via DNA-AgNPs. Reproduced with permission from ref. 93, copyright 2024, *Anal. Chem.*

Fluorescence quenching was brought on by potent $5d^{10}(\text{Hg}^{2+})$ – $4d^{10}(\text{Ag}^+)$ metallophilic contacts.^{77,94} The TEM image clearly showed the formation of AgNPs from the small AgNCs, after Hg^{2+} treatment. Liu *et al.*⁸² theorized that the aggregation of AgNCs was driven by the coordination effect, because of the stronger interaction between Hg^{2+} and the $-\text{COOH}$ group, on the surfaces of AgNCs.

Riboflavin-capped AgNPs were not aggregated in the TEM picture, but they became aggregated after the addition of Hg^{2+} .⁹⁰

8. Concepts of colorimetric sensing

Localized Surface Plasmon Resonance (LSPR) is unique to noble metal nanoparticles (MNPs). The conduction electrons of MNPs collectively and tenaciously vibrate, when a certain frequency of electromagnetic radiation interacts with them. This resonance is known as an LSPR or SPR. Metal NPs and the environment around them were essential to LSPR. NPs can change their size, shape, and composition, changing the LSPR frequency, and their sensitivity to the dielectric properties of their surroundings with LSPR shifts. MNPs are unique due to these distinguishing LSPR characteristics, with gold and AgNPs being among the best in their class. Numerous sensors have been developed with this feature in mind. When MNPs interact with the target analyte, two different types of sensors are produced: an aggregation sensor (caused by the NPs' inter-particle plasmon coupling) and a refractive index sensor (caused by the medium's change in local refractive index—plasmon shift). The LSPR displacement from source frequency results from this. Additionally, there are additional types of sensors, including Surface Enhanced Raman Spectroscopy (SERS) and Metal Enhanced Fluorescence (MEF).^{95,96} Aggregation sensor is the subject of the current debate. Metal ion identification is easy, rapid, good, and cheap when MNPs are aggregated, causing a wide redshift of the LSPR band (Fig. 9).⁹⁷

8.1 Colorimetric detection of Hg^{2+} via AgNCs

Roy *et al.*⁷⁹ designed glutathione-capped AgNCs, which are reddish pink in color, and after the addition of Hg^{2+} , a colorless solution appeared. It was observed also through the naked eye (LOD 245 nM).

Liu *et al.*⁸² prepared pinked-colored AgNCs, templated with the sodium salt of polymethacrylic acid, employing

a hydrothermal technique. In the presence of Hg^{2+} , the pink color changed to colorless. A shift of the absorbance peak from 510 nm to 470 nm due to the color change was observed.

8.1.1 Mechanism for Hg^{2+} sensing via AgNCs. Liu *et al.*⁸² proposed an aggregation-induced mechanism for the observed color and absorbance changes after Hg^{2+} treatment.

8.2 Colorimetric detection of Hg^{2+} ions using AgNPs

8.2.1 Involving plant extract. For colorimetric detection of Hg^{2+} , many plant extracts with passivated AgNPs have been used, such as jicama root,⁹⁸ French lavender,⁹⁹ *Convolvulus cneorum's* aqueous leaf,¹⁰⁰ *Bistorta amplexicaulis*,¹⁰¹ onion¹⁰² Gum kondagogu,¹⁰⁸ Green tea,¹⁰⁴ Kokum fruit,¹⁰⁵ *Citrus japonica* (CJ) leaf,¹⁰⁶ and *Matricaria recutita* (Babunah),¹⁰⁷ *etc.* They are mostly used as reducing as well as stabilizing agents (Table 3).

Farhadi *et al.*⁹⁴ synthesized soap-root plants as a stabilizing agent for AgNPs. Because silver nanoparticles' surface plasmon resonance vibrations (SPR band) were excited, synthesized AgNPs appeared yellowish-brown in an aqueous solution. The color of the freshly prepared AgNPs solution changed from yellowish-brown to pale yellow upon the addition of Hg^{2+} solution. The color of the AgNPs solution then gradually decreased as the concentration of Hg^{2+} increased, and it eventually turned colorless upon the addition of a known concentration of $10^{-3} \text{ mol L}^{-1}$ mercury(II) (Fig. 10).

For example, using an environmentally friendly method, *Convolvulus cneorum's* aqueous leaf extract was used to create AgNPs. A possible colorimetric detection limit for hazardous Hg^{2+} , and Cr(VI) up to ppm was demonstrated using so-produced AgNPs. When Hg^{2+} , Cr(VI), and ammonia were added, the SPR peak of AgNPs underwent a noticeable blue shift.¹⁰⁰

8.2.2 Involving animal waste. Tirado-Guizar *et al.*¹⁰⁹ produced egg-white (EW) encapsulated AgNPs in a single step using crude EWs, a readily available reagent. These NPs were used to detect Hg^{2+} ions in solution using colorimetry. The color was yellowish-brown AgNPs. In the presence of Hg^{2+} , it became colorless, with a blue shift (LOD 300 nM) due to amalgam formation.

8.2.3 Involving lab chemicals. Several lab chemicals such as lignosulphonate (LS),¹¹⁰ *N*-Steroylethanolamine (NSEA),¹¹¹ cytosine triphosphate,¹¹² natural polymer (gelatine),¹¹³ polyvinyl alcohol (PVA) with alanine,¹¹⁴ mercury-specific



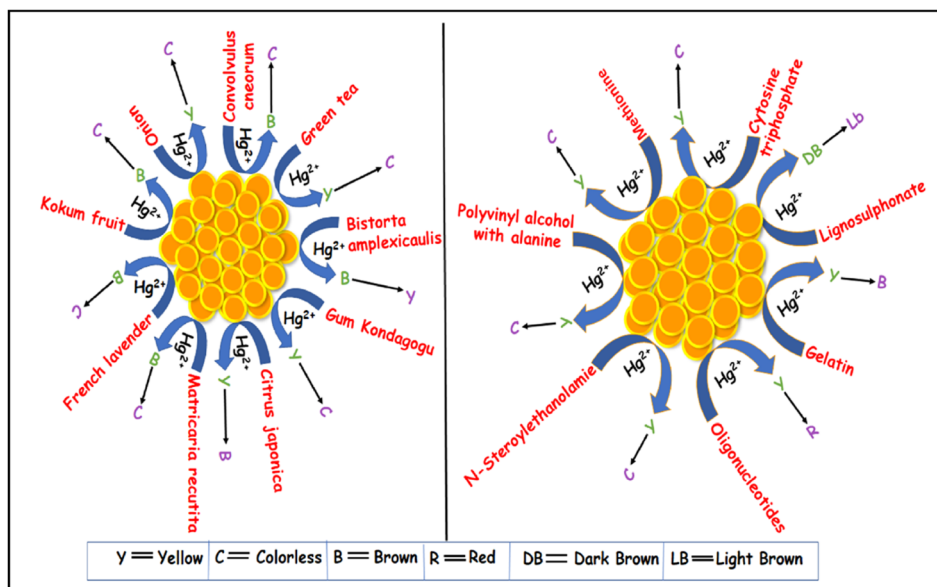


Fig. 9 Colorimetric-based sensing of mercury with AgNPs, with different capping agents.

oligonucleotides, cytosine triphosphate¹¹⁵ methionine,¹¹⁶ L-tyrosine¹¹⁷ were employed to synthesize and stabilize AgNPs (Table 4).

For example, Zhan *et al.*¹¹² developed a colorimetric platform for measuring Cr^{3+} and Hg^{2+} , using AgNPs capped with cytosine triphosphate (CTP). AgNO_3 was reduced in the presence of CTP in a single step to yield the CTP-capped AgNPs. It was shown that these AgNPs can interact with Cr^{3+} . A new, red-shifted band arose at 510 nm as a result, turning the material from yellow to red, and the SPR band at 390 nm lost some of its intensity. In further cases, the yellow color of AgNPs vanished gradually with increased $[\text{Hg}^{2+}]$, associated with little hypsochromic shift (detection range 0.625 μM to 5 μM and detection limit 0.125 μM).

Huang *et al.*¹¹⁷ synthesized L-tyrosine capped AgNPs for Hg^{2+} sensing, based on the Tyndall effect (TE) assay. The TE-inspired assay has a linear detection range for Hg^{2+} of 5 nM to 4 M.

8.2.4 Mechanism for Hg^{2+} sensing via silver nanoparticles. AgNPs and their composites are frequently used in biological and chemical sensing. SPR of AgNPs displays absorption bands between 370 and 470 nm wavelength. Surface functionalization has a substantial impact on the nanoparticles' stability.^{118–120} The functionalization preserves the nanostructure's surface chemistry, crystallinity, form, and size. AgNP aggregation and disaggregation processes need the presence of certain analytes. These aggregation and disaggregation processes alter the SPR band intensity and λ_{max} . Together with a red or blue shift in the SPR peak, these processes often result in a color change in the

Table 3 Colorimetric detection of Hg^{2+} ions using AgNPs, synthesized from plant extract

Plant extract	Color change (before and after Hg^{2+} treatment)	λ change (before and after Hg^{2+} treatment)	Linear range	LOD	References
Jicama root	Yellowish brown to colorless	From 427 nm to & hypsochromic	100 nM –200 μM		98
French lavender	Brown to colorless	From 440 nm to 290 nm hypsochromic	2.7 μM –200 μM	2.7 μM	99
<i>Convolvulus cneorum's</i> aqueous leaf	Brown to colorless	Blue shift 452–417	50 μM –200 mM	5 ppb	100
<i>Bistorta amplexicaulis</i>	Dark brown to light yellow	—	1×10^{-6} to 1×10^{-7} M	8.0×10^{-7} M	101
Onion	Yellow to colorless	From 400 nm to 290 nm hypsochromic shift	—	—	102
Gum kondagogu	Yellow to colorless	—	50–900 nM	4.9×10^{-8} mol L ⁻¹	103
Green tea	Brownish to transparent	424 nm	20 to 160 μM	—	104
Kokum fruit	Brown to colorless	426 nm	120–200 ppm	6.2 ppb	105
<i>Citrus japonica</i>	Yellow to brownish	—	0.3–7.3 μM	0.09 μM	106
<i>Matricaria recutita</i>	Yellowish brown to colorless	—	—	—	107



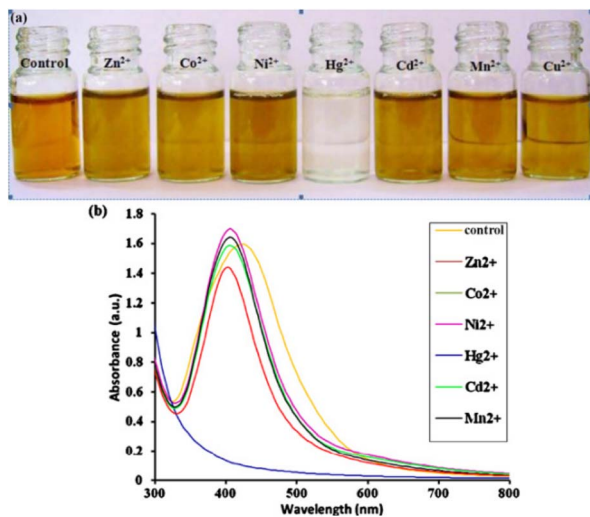


Fig. 10 (a) Digital image and (b) UV-vis spectra of synthetic AgNPs containing various transition-metal ions. Reproduced with permission from ref. 94, copyright 2024, *Sens. Actuators B: Chem.*

system.^{109,115} Moreover, other mechanisms are also reported such as amalgam formation^{104,107} redox reaction^{98,100,110,111} and the Tyndall effect (TE).¹¹⁷

8.2.4.1 Redox reaction. One putative mechanism for chlorophyll-AgNPs' interaction with Hg^{2+} ions is the oxidation of metallic Ag. Hg ions were associated with $-\text{CH}_3$ and $-\text{CH}_2$ at the surface of chlorophyll-AgNPs and exchanged electrons with the AgNPs. The color faded as a result. The SPR band widened and shifted to blue, and the absorption band at 290 nm picked up additional silver ions.⁹⁹

The possibility of a redox reaction, taking place at the surface of the AgNPs, was utilized to explain the putative mechanism for the selective identification of Hg^{2+} due to the difference between the standard potentials of 0.85 V (Hg^{2+}/Hg) and 0.8 V (Ag^+/Ag). When Hg^{2+} is added to the colloidal AgNPs solution, the organic capping agents on the surface of the AgNPs strengthen the electrostatic-ionic attractions between the NPs and Hg.¹⁰⁰ Anbazhagan *et al.*¹¹¹ also reported the selective detection of Hg^{2+} ions *via* the decoloration of *N*-steroylethanolamine-AgNPs due to redox reaction.

Due to the lower cohesive energy of Hg (0.69 eV) than that of Ag, reducing chemicals can create Hg atoms by reducing Hg^{2+} direct contact with AgNPs can result in the formation of Ag@Hg nanoalloys (2.95 eV). Newly created Hg atoms may migrate onto the silver surface, resulting in a slight blue shift and a diminution of the absorption band. Zhan *et al.*¹¹² synthesized cytosine triphosphate-capped AgNPs and the capping agent reduced Hg^{2+} to Hg atom. During this process, Hg had direct contact with AgNPs to form Ag@Hg nanoalloys.

8.2.4.2 Ag-Hg amalgam. A reaction between AgNPs and Hg ions produced amalgam. Amalgam formation can happen when Hg^{2+} and AgNPs interact chemically through under-potential deposition due to the negligibly tiny electrochemical potential difference between Hg^{2+} (0.85) and AgNPs (0.8 V). This suggests that the amalgam formation process is quite probable.^{104,105,107}

8.2.5 Aggregation. Thakur *et al.*¹¹⁰ designed sodium lignosulphonate-capped AgNPs for sensing higher concentrations of Hg^{2+} ions, achieved by color change. Another sign of the aggregation is the solution's gradual release of LS, as mercuric ions were added one at a time.

The polyvinylpyrrolidone-capped AgNPs added to methionine for colorimetric conjugate, play a significant role in the aggregation of the AgNPs because of the enhanced attraction of Hg^{2+} for the sulfur present in methionine. The selectivity of Hg^{2+} could not be observed without methionine. The Hg^{2+} ion caused the AgNPs to agglomerate, turning them colorless from pale yellow.

8.2.5.1 Tyndall effect (TE). The term Tyndall colloid was given by British scientist John Tyndall because it frequently suggests a "visible light path" that results from the scattering of a light beam by colloidal nanoparticles. The size or quantity of colloidal particles favorably impacts the TE intensity. AgNPs are preferred over AuNPs of the same size because of their less expensive material cost and other characteristics.^{121,122}

Visual tests are initially made using the silver colloid's TE. The TE-inspired test can detect Hg^{2+} ions with incredible sensitivity. It is about 5400 times more sensitive than surface plasmon resonance signaling. All you need is a smartphone and a laser pointer pen for portable quantification.¹¹⁷

For example, using hydrothermal tyrosine reduction, AgNPs were created, and they emitted an intense reddish signal due to

Table 4 Colorimetric detection of Hg^{2+} ions using AgNPs, synthesized from lab chemicals

Lab chemicals	Color change (before and after Hg^{2+} treatment)	λ change (before and after Hg^{2+} treatment)	Linear range	LOD	References
Lignosulphonate	Dark brown to light brown	403.5 nm/409 nm	1–20 ppm	0.7 ppm	110
<i>N</i> -Steroylethanolamine	Yellow to colorless	418 nm/gradual hypsochromic shift	—	—	111
Cytosine triphosphate	Yellow to colorless	390 nm/gradual hypsochromic shift	0.625 to 5 Mm	0.125 μM	112
Natural polymer	Generation of blue color	620 nm	0.50 to 800 nM	0.125 nM	113
Polyvinyl alcohol (PVA) with alanine	Brownish-yellow to colorless	—	200–1000 nM	143 nM	114
Mercury-specific oligonucleotides	Yellow to pale red	395 nm/570 nm	25–500 nM	17 nM	115
Methionine	Yellow to colorless	390 nm/Hypochromic shift	20–100 nm	—	116



TE (Fig. 11). As a result of the AgNPs being destroyed by the unique oxidation–reduction interactions between the analyte ions [$E^0(\text{Hg}^{2+}/\text{Hg}) = 0.85 \text{ V}$] and the nanoprobe [$E^0(\text{Ag}^+/\text{Ag}) = 0.80 \text{ V}$], the TE would be significantly decreased or even eliminated with the addition of Hg^{2+} . The variation in TE intensity was used to qualitatively or semi-quantitatively analyze the Hg^{2+} content and a Hg^{2+} detection limit of 0.85 nM was achieved.¹¹⁷

9. The fate of mercury and AgNPs

AgNPs are usually found to be aggregated and destabilized after the addition of Hg^{2+} in the silver hydrosol, as observed from various characterization tools. DLS, TEM, SEM, zeta potential, and XRD data often support the amalgam formation after the treatment of Hg^{2+} towards AgNPs.

For example, Alzahrani¹⁰² reported the green synthesis of AgNPs with a thin layer of capping materials, consisting of onion extract on their surface. TEM images of the particles were in the nano range with a spherical shape, great dispersion, and no aggregation, effectively stimulating the LSPR band's absorbance. After adding Hg^{2+} , the AgNPs were also evaluated by TEM examination. The findings showed that AgNPs were dramatically changed by the addition of Hg^{2+} . Catalytic interactions between Hg^{2+} and AgNPs caused the NPs to assemble and aggregate. Hg^{2+} , interacting with AgNPs, decreased the absorbance of the LSPR band in UV-vis spectra. AgNP aggregation did not exist together with other metal ions.

Gum kondagogu-capped¹⁰⁸ AgNPs underwent SEM-EDAX investigation before and after being exposed to Hg^{2+} . By lyophilizing the AgNPs colloidal solution before and after the reaction with Hg^{2+} and recording XRD bands, it was determined that Ag–Hg alloy was produced *via* analyzing standard silver-nanocomposite substance XRD patterns before and after Hg treatment. The face-centered cubic bulk metallic equivalents of the crystalline phases, produced by the creation of AgNPs in the Gum kondagogu medium, could be indexed to the (1 1 1), (2 0 0), (2 2 0), and (3 1 1) planes. All of the diffraction peaks are distinct, with the peak at (1 1 1) having the greatest strength.

The XRD study supports the creation of amalgam following the addition of Hg^{2+} . The lyophilized powder from AgNPs and Hg^{2+} reaction had an XRD pattern, nearly matched to Ag_2Hg_3 alloy, proving that Hg^{2+} had been reduced by Ag^0 .

The TEM picture showed scattered sphere-shaped nanocrystals with an average diameter of around 13 nm for the cytosine triphosphate (CTP)-capped AgNPs.¹¹² The average hydrodynamic diameter of scattered AgNPs was found to be $\sim 30 \text{ nm}$, which was a bit bigger than what was predicted using the TEM image. According to the TEM image, the irregular amalgam was formed after being incubated with Hg^{2+} as a result of Hg atom diffusion.¹¹² Hg^{2+} -stimulated AgNP aggregation was provided by TEM (reported by Wang *et al.*¹¹⁵), which revealed individual NPs in the absence of Hg^{2+} and aggregated NPs in the presence of Hg^{2+} (Fig. 12).

AgNPs can detect mercury ions as low as ppm level, when coupled with a ligand, such as sodium 3-mercapto-1-propane sulfonate(3-MPS). Schiesaro *et al.*¹²³ suggested the sensitivity of the nanosensor to Hg^{2+} in the 1 to 5 ppm range, using optical analysis, and substantiated the observation with several characterization tools (Fig. 13). The color was changed from yellow to colorless. The absorption spectra of AgNPs had a distinct maximum at 399 nm. With an increase of 1 ppm of Hg^{2+} , the absorption band was blue-shifted to 385 nm. As the amount of Hg^{2+} in the Ag hydrosol increased, the blue shift increased and reached its peak at 348 nm at 5 ppm of Hg^{2+} . The formation of homogeneous NPs of Ag–Hg alloys appeared to be triggered by the addition of Hg^{2+} .

The size of AgNPs increased in the presence of Hg^{2+} . DLS data supported 3-MPS ligand-capped-AgNPs to be non-aggregated than the same sample with 1 ppm Hg^{2+} . 3-MPS ligated AgNP/Hg aggregate structure was clarified by TEM morphological analysis, revealing the creation of Ag/Hg alloy NPs.

The EDS map investigation in conjunction with the STEM analysis brought up key points. Some AgNPs had dark sections with low Hg content and high porosity. Others included brilliant regions with high Hg concentrations and little porosity.

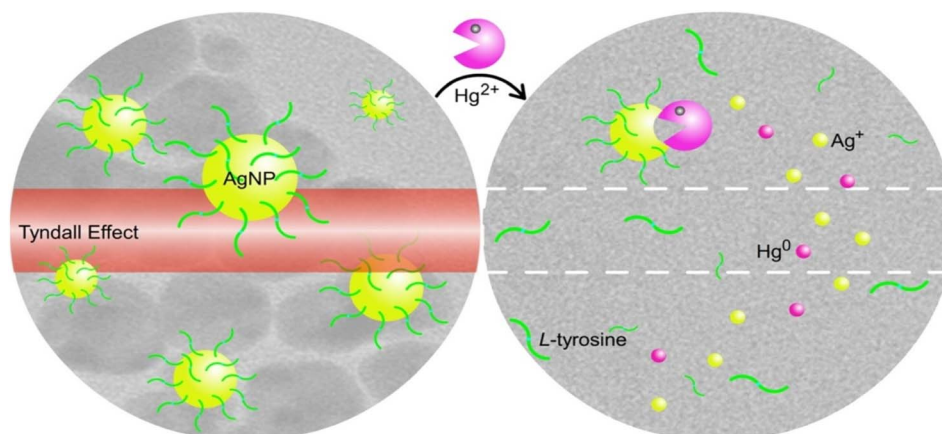


Fig. 11 Sensing of mercury with tyrosine-stabilized AgNPs involving the Tyndall effect. Reproduced with permission from ref. 117, copyright 2024, *Sens. Actuators B: Chem.*



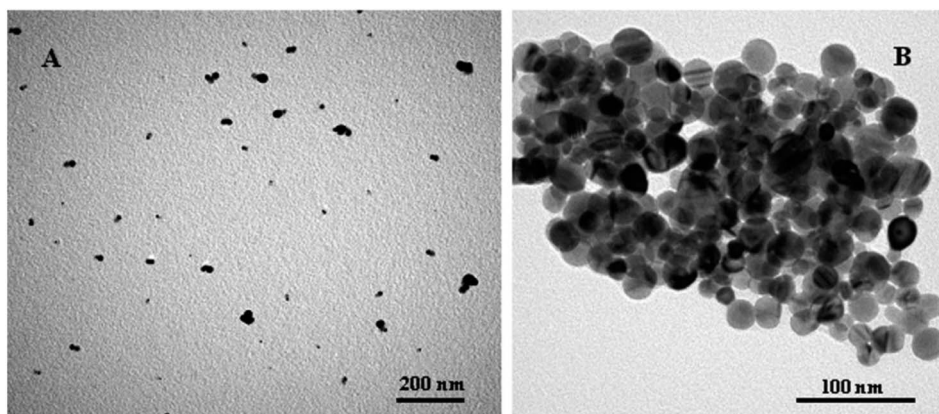


Fig. 12 TEM images of the AgNPs solution (0.075 nM) mixed with 300 nM MSO in the (A) absence or (B) presence of 500 nM Hg^{2+} after the addition of 60 mM NaCl. Reproduced with permission from ref. 115, copyright 2024, ACS Appl. Mater. Interfaces.

Furthermore, it was clear that the size of the AgNPs affected the dispersion of Hg atoms. In contrast to the smaller population, which had few or no Hg atoms interacting with the NPs, the larger population had a uniform distribution of Hg atoms across its whole volume, generating an amalgam of Ag and Hg rather than a core-shell structure.

The XPS spectra¹²³ of AgNPs were unsymmetric because of the capping of 3-MPS. The AgNPs were composed of at least two distinct types of Ag atoms. The extremely modest signal at greater binding energy was typically attributed to positively charged silver atoms, interacting with the ligand at the NP surface. Metallic Ag atoms in the NPs core had the lowest spin-orbit coupling binding energy (Ag $3d_{5/2}$ = 368.09 eV binding energy). The XPS spectra of Hg 4f were asymmetric with the addition of Hg, resulting in two distinct couples of spin-orbit components. The primary component (Hg $4f_{7/2}$ BE = 99.9 eV) was consistent with Hg^0 atoms for amalgams. The signal of lower intensity (about 10% of the total Hg 4f signal) with higher binding energy (around 101 eV) could be attributed to Hg^{2+} ions in coordination compounds or oxides.

10. Spot detection

There are some reports^{114,116} available in the literature for spot Hg^{2+} . Janani *et al.*¹¹⁴ synthesized polyvinyl alcohol-stabilized AgNPs. In the presence of alanine, Hg^{2+} can be detected by the colorimetric sensor. For making the test paper-based sensor, the Whatmann paper was submerged in AgNPs-alanine. To make the hydrogel for Hg^{2+} detection, agarose powder was dissolved in an AgNPs-alanine conjugate. To test the recognition depending on visual color change, a few drops of Hg^{2+} and other metal ions solutions were dripped onto the dried filter paper and agarose gel-based sensor. Hg^{2+} concentrations ranging from 1 nM to 1 mM were used to determine how the color gradient appeared.

11. Real sample analysis

NPs and NCs of Ag metal are widely used for selective and sensitive mercury detection. Though there are ample

reports^{77,92,98,108,112,114,117} available in the literature, it is also important to check their applicability in natural water samples (tap water, rainwater, sewage water, river, ocean, etc.). Hg is one of the serious contaminants in natural water, causing hazards in ecosystems human health, and the environment. The real sample analysis provides unequivocal evidence of the applicability of methods for prototype applications. To test the metal ion sensing potential of NSEA-AgNPs in ambient water bodies, Veerappan *et al.*¹¹¹ collected water samples from the normal tap (a nearby pond), disposal of a sewage plant (campus of SASTRA University in, Tamilnadu), and blood plasma. The addition of NSEA-AgNPs indicates the absence of Hg contamination in these samples.⁹⁸ They also spiked Hg^{2+} in natural samples and successfully determined [Hg^{2+}] with their proposed technique.

12. Effect of adulteration

The production of bimetallic Au/Ag NPs has also received increased attention recently, with the goal of combining their synergistic effects to attain more alluring benefits over monometallic ones for electrical, optical signals, and catalytic efficiency.^{124,125}

In the long-overlooked “silver effect” on gold catalysis, bimetallic AuAg NPs (Au–Ag BNPs) have substantially higher catalytic activity than Au nanoparticles alone. Additionally, some luminescent Au/AgNCs have been created or used by adding silver to alter the luminance of AuNCs.^{126,127} A Few reports^{128,129} are available on silver gold-doped nanoclusters and nanoparticles for Hg^{2+} sensing.

Ganguly *et al.*¹³⁰ designed a glutathione-capped core-shell giant cluster, with gold(i) in the core and silver(0) in the shell. The core-shell fluorescent giant cluster is highly stable with time, even in solid form (Fig. 14).¹³¹ This giant cluster was a sensitive and selective platform for Hg^{2+} based on fluorescence quenching. The metallophilic $5d^{10}(\text{Hg}^{2+})-4d^{10}(\text{Ag}^{\delta+})$ interaction as well as Hg^{2+} stimulated aggregation was ascribed for causing the fluorescence quenching and redshift (LOD 6 nM).



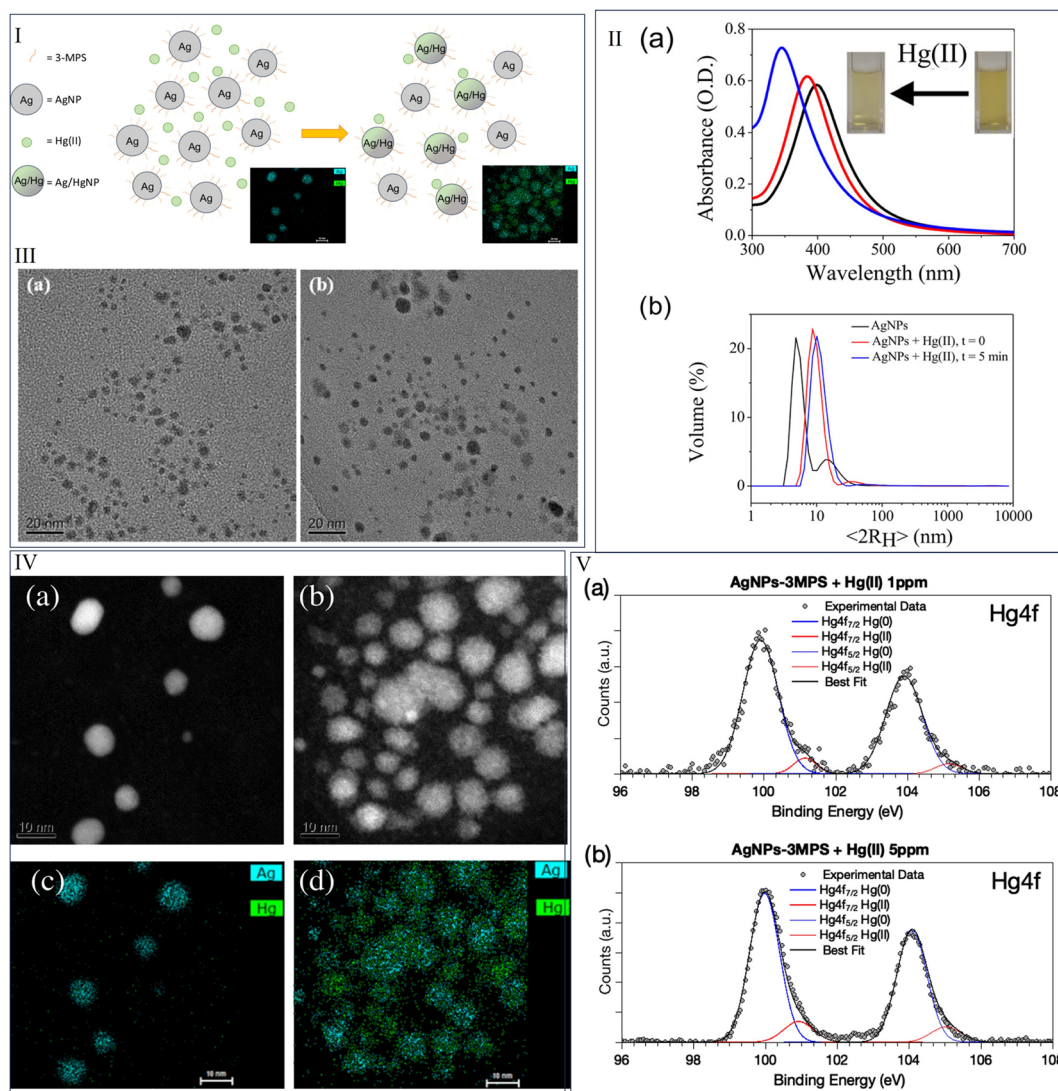


Fig. 13 (I) Formation of Ag–Hg amalgam after the treatment of Hg^{2+} to 3-MPS capped AgNPs; (II) (a) UV-visible spectra and (b) DLS spectra of 3-MPS capped AgNPs with the addition of Hg^{2+} ; (III) TEM image (a) before and (b) after the addition of Hg^{2+} ; (IV) (a) STEM image before addition of Hg^{2+} , (b) STEM image after addition of Hg^{2+} , (c) EDS mapping before addition of Hg^{2+} , and (d) EDS mapping after addition of Hg^{2+} ; (V) XPS spectra for the element mercury after the addition of Hg^{2+} at two different concentrations. Reproduced with permission from ref. 123, copyright 2024, *J. Phys. Chem. C*.

Zhang *et al.*¹³² synthesized bovine serum albumin-capped fluorescent bimetallic alloying gold–silver NCs for the selective sensing of Hg^{2+} and Cu^{2+} .

Mathaweesansurn *et al.*¹³³ designed a colorimetric sensor, Au–Ag bimetallic NPs for sensitive and selective detection of Hg^{2+} . Hg^{2+} was found to have a LOD of $0.526 + 0.001 \text{ mg L}^{-1}$ and a detection range of 0.5 to 80 mg L^{-1} . The practical application of bimetallic NPs for the detection of Hg^{2+} in real samples has also been confirmed.

Direct electrodeposition of the Ag–AuNPs onto the indium tin oxide glass substrate was reported by Tao *et al.*¹³⁴ In addition to serving as a reducing agent, the NPs may also act as a recognition platform to find Hg^{2+} . The Ag–AuNP surface had

a hypochromic shift in the LSPR band due to the Hg deposition (LOD 0.02 ppb and linear detection range 0.05 to 500 ppb).

Au–Fe doped AgI passivated with polyglycine was employed for colorimetric sensing of Hg^{2+} . The polyglycine assisted in stabilizing the Au–Fe doped AgI *via* hydrogen bonding. Hg^{2+} introduced a redox reaction, causing the formation of elemental Hg to coat the metal composite for sensing *via* amalgamation. Thus, optical density was decreased with the oxidation of metals involved in the sensing platform, and *in situ* produced reduced Hg was deposited on the oxidized metalized surface.¹²⁹ The LOD was observed to be 1.0 nM , while λ_{max} of 500 nm in the absorption spectra was decreased with $[\text{Hg}^{2+}]$.



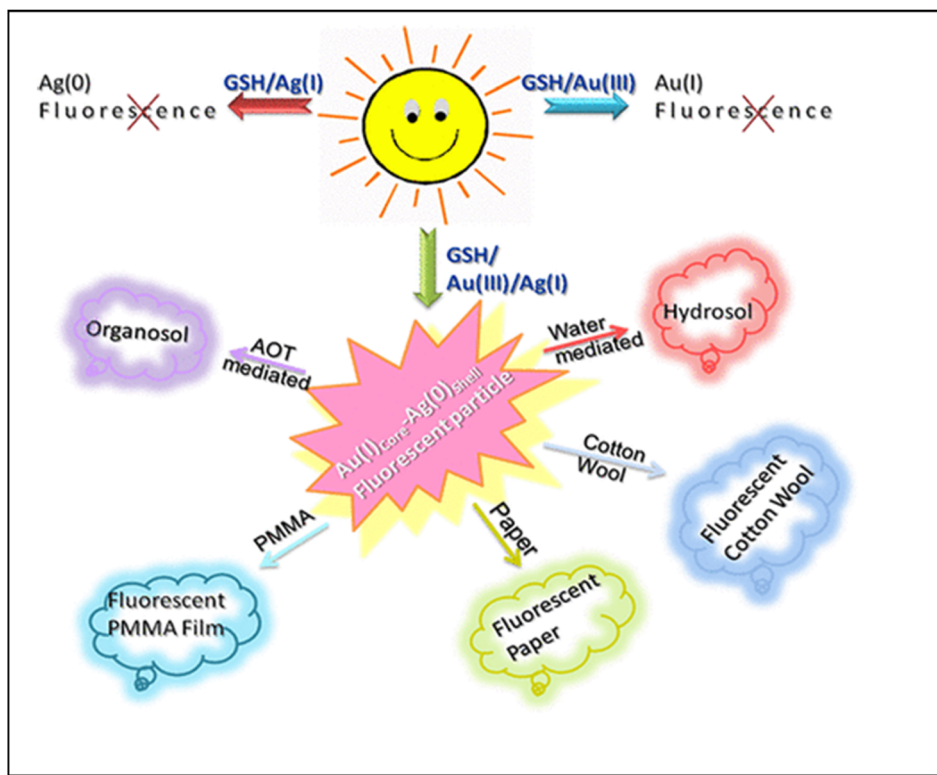


Fig. 14 Bimetallic gold silver giant clusters with strong emissive properties for versatile applications. Reproduced with permission from ref. 131, copyright 2024, Langmuir.

13. Conclusions and future perspectives

While NPs are primarily used for colorimetric techniques, NCs are predominantly used for fluorometric procedures. Numerous quenching-based fluorometric detection techniques exist. However, a few stories included sensing with enhancement. This review is a comparative account of AgNPs and AgNCs regarding physicochemical behaviors, sensing tactics, the fate of associated components, plausible mechanisms, spot detection, and real sample analyses. In contexts of prototype uses, such as in hospitals, labs, homes, and fields, novel AgNPs and AgNCs syntheses with non-toxic, stable, affordable, and biocompatible reagents are warranted for their simplicity, convenience of use, and low cost. This review will hopefully serve as an asset for the young researcher to venture into the field of hazardous material sensing and remediation technologies with sufficient literature background.

Conflicts of interest

There are no conflicts to declare.

References

1 F. N. Chaudhry and M. F. Malik, *J. Ecosyst. Ecography*, 2017, 7, 1–3.

2 S. Ahuja, *ACS Symp. Ser.*, 2020, 1352, 1–11.
 3 J. Mikes, M. Siglova, A. Cejkova, J. Masak and V. Jirku, *Water Sci. Technol.*, 2005, 52, 151–156.
 4 J. Briffa, E. Sinagra and R. Blundell, *Heliyon*, 2020, 6, e04691.
 5 M. Liu, Q. Zhang, T. Maavara, S. Liu, X. Wang and P. A. Raymond, *Nat. Geosci.*, 2021, 14, 672–677.
 6 A. C. (Thanos) Bourtsalas and N. J. Themelis, *Waste Manage.*, 2019, 85, 90–94.
 7 D. D. Evanoff and G. Chumanov, *ChemPhysChem*, 2005, 6, 1221–1231.
 8 N. Cao, J. Xu, H. Zhou, Y. Zhao, J. Xu, J. Li and S. Zhang, *Microchem. J.*, 2020, 159, 105406.
 9 M. Sabela, S. Balme, M. Bechelany, J. M. Janot and K. Bisetty, *Adv. Eng. Mater.*, 2017, 19, 1–24.
 10 G. Y. Lan, C. C. Huang and H. T. Chang, *Chem. Commun.*, 2010, 46, 1257–1259.
 11 D. M. Riley, C. A. Newby and T. O. Leal-Almeraz, *Risk Anal.*, 2006, 26, 1205–1221.
 12 B. Ladizinski, N. Mistry and R. V. Kundu, *Dermatol. Clin.*, 2011, 29, 111–123.
 13 T. Y. K. Chan, *Clin. Toxicol.*, 2011, 49, 886–891.
 14 A. Podgórska, A. Puścion-Jakubik, A. Grodzka, S. K. Naliwajko, R. Markiewicz-żukowska and K. Socha, *Molecules*, 2021, 26(13), 4088.
 15 H. H. Abbas, M. Sakakibara, K. Sera, Nurgahayu and E. Andayani, *Cosmetics*, 2020, 7, 58.



- 16 J. Chen, Y. Ye, M. Ran, Q. Li, Z. Ruan and N. Jin, *Front. Pharmacol.*, 2020, **11**, 1–10.
- 17 M. Cotte, J. Susini, N. Metrich, A. Moscato, C. Gratziu, A. Bertagnini and M. Pagano, *Anal. Chem.*, 2006, **78**, 7484–7492.
- 18 A. Leyva-Pérez and A. Corma, *Angew. Chem., Int. Ed.*, 2012, **51**, 614–635.
- 19 J. Barluenga and R. Rodes, *J. Chem. Soc., Perkin Trans. 1*, 1980, **1**, 2732–2737.
- 20 J. Barluenga, J. Perez-Prieto and D. Valencia, *Tetrahedron*, 1990, **46**, 2453–2460.
- 21 T. E. Müller, K. C. Hultzs, M. Yus, F. Foubelo and M. Tada, *Chem. Rev.*, 2008, **108**, 3795–3892.
- 22 S. Mandal, R. D. Chaudhari and G. Biswas, *Beilstein J. Org. Chem.*, 2021, **17**, 2348–2376.
- 23 R. A. Periana, D. J. Taube, E. R. Evitt, D. G. Löffler, P. R. Wentreck, G. Voss and T. Masuda, *Science*, 1993, **259**, 340–343.
- 24 X. Gang, H. Birch, Y. Zhu, H. A. Hjuler and N. J. Bjerrum, *J. Catal.*, 2000, **196**, 287–292.
- 25 W. T. Foreman, S. D. Zaug, L. M. Falres, M. G. Werner, T. J. Lelker and P. F. Rogerson, *Environ. Sci. Technol.*, 1992, **26**, 1307–1312.
- 26 S. N. Hashim, S. Z. Ghazali, N. J. Sidik, T. Chia-Chay and A. Saleh, *E3S Web of Conferences*, 2021, vol. 306, pp. 1–8.
- 27 A. Alam, S. Ferdosh, K. Ghafoor, A. Hakim, A. S. Juraimi, A. Khatib and Z. I. Sarker, *Asian Pac. J. Trop. Med.*, 2016, **9**, 402–409.
- 28 D. L. García Lozano, M. L. Ocampo Guerrero and N. Mesa López, *Rev. Colomb. Biotechnol.*, 2015, **17**, 76–84.
- 29 L. A. Weed and E. E. Ecker, *J. Infect. Dis.*, 1931, **51**, 309–314.
- 30 M. Tilahun, D. Mulugeta and S. Manju, *Afr. J. Biotechnol.*, 2013, **12**, 6282–6286.
- 31 M. Paul Das, L. Jeyanthi Rebecca, S. Sharmila and S. Chatterjee, *J. Chem. Pharm. Res.*, 2012, **4**, 4975–4978.
- 32 G. W. C. Konrad Lorincz, B.-rong Chen, S. P. Atanu Roy Chowdhury, P. Bonato and M. Welsh, *SenSys' 09: Proceedings of the 7th ACM Conference on Embedded Networked Sensor Systems.*, 2009, pp. 183–196.
- 33 F. Askari, A. Rahdar, M. Dashti and J. F. Trant, *J. Fluoresc.*, 2020, **30**, 1181–1187.
- 34 H. Agarwalla, R. N. Senapati and T. B. Das, *J. Environ. Sci.*, 2021, **100**, 28–33.
- 35 V. K. Rai, N. S. Raman and S. K. Choudhary, *Ind. J. Pure App. Biosci.*, 2013, **1**, 31–37.
- 36 M. M. Veiga, G. Angeloci-Santos and J. A. Meech, *Extr. Ind. Soc.*, 2014, **1**, 351–361.
- 37 M. M. Veiga and J. A. Meech, *Ambio*, 1995, **24**, 371–375.
- 38 M. M. Veiga, P. A. Maxson and L. D. Hylander, *J. Clean. Prod.*, 2006, **14**, 436–447.
- 39 A. J. Gunson and M. M. Veiga, *Environ. Pract.*, 2004, **6**, 109–120.
- 40 G. N. George, S. P. Singh, J. Hoover and I. J. Pickering, *Chem. Res. Toxicol.*, 2009, **22**, 1761–1764.
- 41 L. Moscou and S. Lub, *Powder Technol.*, 1981, **29**, 45–52.
- 42 F. B. Teixeira, A. C. A. De Oliveira, L. K. R. Leão, E. H. C. De Oliveira, M. E. Crespo-lopez and C. S. F. Maia, *Front. Mol. Neurosci.*, 2018, **11**, 1–11.
- 43 K. V. Katok, R. L. D. Whitby, T. Fukuda, T. Maekawa, I. Bezverkhy, S. V. Mikhalovsky and A. B. Cundy, *Angew. Chem., Int. Ed.*, 2012, **51**, 2632–2635.
- 44 W. Ren, C. Zhu and E. Wang, *Nanoscale*, 2012, **4**, 5902–5909.
- 45 L. Sun, A. Zhang, S. Su, H. Wang, J. Liu and J. Xiang, *Chem. Phys. Lett.*, 2011, **517**, 227–233.
- 46 T. Morris, H. Copeland, E. McLinden, S. Wilson and G. Szulczewski, *Langmuir*, 2002, **18**, 7261–7264.
- 47 I. de la Calle, I. Lavilla, H. Bartolomé-Alonso and C. Bendicho, *Spectrochim. Acta, Part B*, 2019, **161**, 105697.
- 48 Q. Zhang, C. Chen, G. Wan, M. Lei, M. Chi, S. Wang and D. Min, *Microchim. Acta*, 2019, **186**, 727.
- 49 M. Wdowin, M. M. Wiatros-Motyka, R. Panek, L. A. Stevens, W. Franus and C. E. Snape, *Fuel*, 2014, **128**, 451–457.
- 50 J. G. Yu, B. Y. Yue, X. W. Wu, Q. Liu, F. P. Jiao, X. Y. Jiang and X. Q. Chen, *Environ. Sci. Pollut. Res.*, 2016, **23**, 5056–5076.
- 51 L. Wang, H. Xu, Y. Qiu, X. Liu, W. Huang, N. Yan and Z. Qu, *J. Hazard. Mater.*, 2020, **389**, 121824.
- 52 B. Valeur and I. Leray, *Coord. Chem. Rev.*, 2000, **205**, 3–40.
- 53 L. Prodi, F. Bolletta, M. Montalti and N. Zaccheroni, *Coord. Chem. Rev.*, 2000, **205**, 59–83.
- 54 N. Bhardwaj, S. K. Bhardwaj, M. K. Nayak, J. Mehta, K. H. Kim and A. Deep, *TrAC, Trends Anal. Chem.*, 2017, **97**, 120–135.
- 55 C. D. G. J. R. Lakowicz, *Topics in Fluorescence Spectroscopy*, 1991, vol. 4.
- 56 W. Zhou, R. Saran and J. Liu, *Chem. Rev.*, 2017, **117**, 8272–8325.
- 57 L. Deng, Z. Zhou, J. Li, T. Li and S. Dong, *Chem. Commun.*, 2011, **47**, 11065–11067.
- 58 D. L. Ma, D. S. H. Chan, B. Y. W. Man and C. H. Leung, *Chem.-Asian J.*, 2011, **6**, 986–1003.
- 59 J. Wang, C. Du, P. Yu, Q. Zhang, H. Li and C. Sun, *Sens. Actuators, B*, 2021, **348**, 130707.
- 60 L. W. Tang, Y. Alias, R. Zakaria and P. Meng, *Crit. Rev. Anal. Chem.*, 2021, **53**, 869–886.
- 61 J. R. Wisniewski and F. Z. Gaugaz, *Anal. Chem.*, 2015, **87**, 4110–4116.
- 62 P. Bian, L. Xing, Z. Liu and Z. Ma, *Sens. Actuators, B*, 2014, **203**, 252–257.
- 63 G. Wang, G. Xu, Y. Zhu and X. Zhang, *Chem. Commun.*, 2014, **50**, 747–750.
- 64 Y. Miyake, H. Togashi, M. Tashiro, H. Yamaguchi, S. Oda, M. Kudo, Y. Tanaka, Y. Kondo, R. Sawa, T. Fujimoto, T. Machinami and A. Ono, *J. Am. Chem. Soc.*, 2006, **128**, 2172–2173.
- 65 R. C. Pawar, D. H. Choi, J. S. Lee and C. S. Lee, *Mater. Chem. Phys.*, 2015, **151**, 167–180.
- 66 Y. Tanaka, S. Oda, H. Yamaguchi, Y. Kondo, C. Kojima and A. Ono, *J. Am. Chem. Soc.*, 2007, **129**, 244–245.
- 67 J. B. Xiao, X. Q. Chen, X. Y. Jiang, M. Hilczler and M. Tachiya, *J. Fluoresc.*, 2008, **18**, 671–678.



- 68 X. Zhuang, T. Ha, H. D. Kim, T. Centner, S. Labeit and S. Chu, *Proc. Natl. Acad. Sci. U. S. A.*, 2000, **97**, 14241–14244.
- 69 R. Katoh, S. Sinha, S. Murata and M. Tachiya, *J. Photochem. Photobiol., A*, 2001, **145**, 23–34.
- 70 R. Liu, S. Duan, L. Bao, Z. Wu, J. Zhou and R. Yu, *Anal. Chim. Acta*, 2020, **1114**, 50–57.
- 71 M. Ghosh, S. Nath, A. Hajra and S. Sinha, *J. Lumin.*, 2013, **141**, 87–92.
- 72 S. Hamann, J. F. Kiilgaard, T. Litman, F. J. Alvarez-Leefmans, B. R. Winther and T. Zeuthen, *J. Fluoresc.*, 2002, **12**, 139–145.
- 73 Y. Liu, W. Zhu, D. Ni, Z. Zhou, J. H. Gu, W. Zhang, H. Sun and F. Liu, *J. Nanobiotechnol.*, 2020, **18**, 1–14.
- 74 S. P. Bayen, M. K. Mondal, S. Naaz, S. K. Mondal and P. Chowdhury, *J. Environ. Chem. Eng.*, 2016, **4**, 1110–1116.
- 75 S. Naaz and P. Chowdhury, *Sens. Actuators, B*, 2017, **241**, 840–848.
- 76 J. Peng, J. Ling, X. Q. Zhang, H. P. Bai, L. Zheng, Q. E. Cao and Z. T. Ding, *Spectrochim. Acta, Part A*, 2015, **137**, 1250–1257.
- 77 D. Lu, Z. Chen, Y. Li, J. Yang, S. Shuang and C. Dong, *Anal. Lett.*, 2015, **48**, 281–290.
- 78 A. Mao and C. Wei, *Microchim. Acta*, 2019, **186**, 541.
- 79 S. Roy, A. Baral and A. Banerjee, *ACS Applied Materials and Interface*, 2014, **6**, 4050–4056.
- 80 A. Baksi, M. S. Bootharaju, X. Chen, H. Häkkinen and T. Pradeep, *J. Phys. Chem. C*, 2014, **118**, 21722–21729.
- 81 C. Guo and J. Irudayaraj, *Anal. Chem.*, 2011, **83**, 2883–2889.
- 82 J. Liu, X. Ren, X. Meng, Z. Fang and F. Tang, *Nanoscale*, 2013, **5**, 10022–10028.
- 83 S. Hu, B. Ye, X. Yi, Z. Cao, D. Wu, C. Shen and J. Wang, *Talanta*, 2016, **155**, 272–277.
- 84 G. Yang, H. Zhang, Y. Wang, X. Liu, Z. Luo and J. Yao, *Sens. Actuators, B*, 2017, **251**, 773–780.
- 85 R. Z. Wang, D. L. Zhou, H. Huang, M. Zhang, J. J. Feng and A. J. Wang, *Microchim. Acta*, 2013, **180**, 1287–1293.
- 86 J. L. MacLean, K. Morishita and J. Liu, *Biosens. Bioelectron.*, 2013, **48**, 82–86.
- 87 J. B. King, M. R. Haneline, M. Tsunoda and F. P. Gabbai, *J. Am. Chem. Soc.*, 2002, **124**, 9350–9351.
- 88 T. Lasanta, J. M. López-De-Luzuriaga, M. Monge, M. E. Olmos and D. Pascual, *Chem.—Eur. J.*, 2013, **19**, 4754–4766.
- 89 S. Raju, H. B. Singh and R. J. Butcher, *Dalton Trans.*, 2020, **49**, 9099–9117.
- 90 B. Roy, P. Bairi and A. K. Nandi, *Analyst*, 2011, **136**, 3605–3607.
- 91 M. Ganguly, C. Mondal, J. Jana, A. Pal and T. Pal, *Langmuir*, 2014, **30**, 4120–4128.
- 92 S. Kraithong, N. Chailek, J. Sirirak, K. Suwatpipat, N. Wanichacheva and P. Swanglap, *J. Photochem. Photobiol., A*, 2021, **407**, 113064.
- 93 L. Deng, X. Ouyang, J. Jin, C. Ma, Y. Jiang, J. Zheng, J. Li, Y. Li, W. Tan and R. Yang, *Anal. Chem.*, 2013, **85**, 8594–8600.
- 94 K. Farhadi, M. Forough, R. Molaei, S. Hajizadeh and A. Rafipour, *Sens. Actuators, B*, 2012, **161**, 880–885.
- 95 F. Zarlaida and M. Adlim, *Microchim. Acta*, 2017, **184**, 45–58.
- 96 N. C. Pomal, K. D. Bhatt, K. M. Modi, A. L. Desai, N. P. Patel, A. Kongor and V. Kolivoška, *J. Fluoresc.*, 2021, **31**, 635–649.
- 97 C. Caro, P. M. Castillo, R. Klippstein, D. Pozo and A. P. Zaderenko, *Silver Nanopart.*, 2010, 201–225.
- 98 F. M. Lutfi, M. Juwita, P. R. Ibrahim, E. D. Rakhmawaty and R. Iman, *Res. J. Chem. Environ.*, 2018, **22**, 1–3.
- 99 D. Demirezen Yilmaz, D. Aksu Demirezen and H. Mihçioğur, *Surf. Interfaces*, 2021, **22**, 100840.
- 100 M. Ismail, M. I. Khan, K. Akhtar, J. Seo, M. A. Khan, A. M. Asiri and S. B. Khan, *J. Mater. Sci.: Mater. Electron.*, 2019, **30**, 7367–7383.
- 101 H. X. F. Ahemed and H. Kabir, *Front. Chem.*, 2020, **8**, 1–15.
- 102 E. Alzahrani, *J. Anal. Methods Chem.*, 2020, **2020**, 1–14.
- 103 L. Rastogi, R. B. Sashidhar, D. Karunasagar and J. Arunachalam, *Talanta*, 2014, **118**, 111–117.
- 104 P. Prema, V. Veeramani, K. Rameshkumar, M. K. Gatasheh, A. A. Hatamleh, R. Balasubramani and P. Balaji, *Environ. Res.*, 2022, **204**, 111915.
- 105 G. M. Sangaonkar, M. P. Desai, T. D. Dongale and K. D. Pawar, *Sci. Rep.*, 2020, **10**, 1–12.
- 106 S. Bhagat, H. Shaikh, A. Nafady, Sirajuddin, S. T. H. Sherazi, M. I. Bhangar, M. R. Shah, M. I. Abro, R. Memon and R. Bhagat, *J. Cluster Sci.*, 2022, **33**, 1865–1875.
- 107 I. Uddin, K. Ahmad, A. A. Khan and M. A. Kazmi, *Sens. Biosensing Res.*, 2017, **16**, 62–67.
- 108 L. Rastogi, R. B. Sashidhar, D. Karunasagar and J. Arunachalam, *Talanta*, 2014, **118**, 111–117.
- 109 A. Tirado-Guizar, G. Rodriguez-Gattorno, F. Paraguay-Delgado, G. Oskam and G. E. Pina-Luis, *MRS Commun.*, 2017, **7**, 695–700.
- 110 S. G. R. A. Thakur, *Iran. J. Mater. Sci. Eng.*, 2020, **17**, 80–89.
- 111 V. Anbazhagan, K. B. A. Ahmed and S. Janani, *Sens. Actuators, B*, 2014, **200**, 92–100.
- 112 L. Zhan, T. Yang, S. J. Zhen and C. Z. Huang, *Microchim. Acta*, 2017, **184**, 3171–3178.
- 113 Z. Sun, N. Zhang, Y. Si, S. Li, J. Wen, X. Zhu and H. Wang, *Chem. Commun.*, 2014, **50**, 9196–9199.
- 114 B. Janani, A. Syed, A. M. Thomas, A. H. Bahkali, A. M. Elgorban, L. L. Raju and S. S. Khan, *Optik*, 2020, **204**, 164160.
- 115 Y. Wang, F. Yang and X. Yang, *ACS Applied Materials and Interface*, 2010, **2**, 339–342.
- 116 S. Balasurya, A. Syed, A. M. Thomas, N. Marraiki, A. M. Elgorban, L. L. Raju, A. Das and S. S. Khan, *Spectrochim. Acta, Part A*, 2020, **228**, 117712.
- 117 J. Huang, X. Mo, H. Fu, Y. Sun, Q. Gao, X. Chen, J. Zou, Y. Yuan, J. Nie and Y. Zhang, *Sens. Actuators, B*, 2021, **344**, 130218.
- 118 P. Proposito, L. Burratti and I. Venditti, *Chemosensors*, 2020, **8**, 1–29.
- 119 S. K. Laliwala, V. N. Mehta, J. V. Rohit and S. K. Kailasa, *Sens. Actuators, B*, 2014, **197**, 254–263.
- 120 V. Vinod Kumar, S. Anbarasan, L. R. Christena, N. Saisubramanian and S. Philip Anthony, *Spectrochim. Acta, Part A*, 2014, **129**, 35–42.
- 121 W. Xiao, Z. Deng, J. Huang, Z. Huang, M. Zhuang, Y. Yuan, J. Nie and Y. Zhang, *Anal. Chem.*, 2019, **91**, 15114–15122.



- 122 Y. Z. K. Yuan, Y. Sun, F. Liang, F. Pan, M. Hu, F. Hua, Y. Yuan and J. Nie, *RSC Adv.*, 2022, **12**, 23379.
- 123 I. Schiesaro, L. Burratti, C. Meneghini, I. Fratoddi, P. Proposito, J. Lim, C. Scheu, I. Venditti, G. Iucci and C. Battocchio, *J. Phys. Chem. C*, 2020, **124**, 25975–25983.
- 124 M. J. Hostetler, C. J. Zhong, B. K. H. Yen, J. Anderegg, S. M. Gross, N. D. Evans, M. Porter and R. W. Murray, *J. Am. Chem. Soc.*, 1998, **120**, 9396–9397.
- 125 R. Ferrando, J. Jellinek and R. L. Johnston, *Chem. Rev.*, 2008, **108**, 845–910.
- 126 D. Wang, R. Cai, S. Sharma, J. Jirak, S. K. Thummanapelli, N. G. Akhmedov, H. Zhang, X. Liu, J. L. Petersen and X. Shi, *J. Am. Chem. Soc.*, 2012, **134**, 9012–9019.
- 127 J. Sun and Y. J. Haoxi Wua, *Nanoscale*, 2014, **6**, 5449–5457.
- 128 R. Gui, Y. Wang and J. Sun, *Microchim. Acta*, 2014, **181**, 1231–1238.
- 129 Y. Liu, Z. Xu, S. Zhu, A. Fakhri and V. Kumar Gupta, *J. Photochem. Photobiol., A*, 2022, **422**, 113522.
- 130 M. Ganguly, C. Mondal, J. Pal, A. Pal, Y. Negishi and T. Pal, *Dalton Trans.*, 2014, **43**, 11557–11565.
- 131 M. Ganguly, J. Pal, S. Das, C. Mondal, A. Pal, Y. Negishi and T. Pal, *Langmuir*, 2013, **29**, 10945–10958.
- 132 N. Zhang, Y. Si, Z. Sun, L. Chen, R. Li, Y. Qiao and H. Wang, *Anal. Chem.*, 2014, **86**, 11714–11721.
- 133 A. Mathaweesansurn, N. Vittayakorn and E. Detsri, *Molecules*, 2020, **25**, 4443.
- 134 H. Tao, Y. Lin, J. Yan and J. Di, *Electrochem. Commun.*, 2014, **40**, 75–79.

