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Tetrazine-*trans*-cyclooctene ligated lanthanide conjugates for biomedical imaging

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Lanthanide chelates and copper-free 'click' chemistry have important uses for targeted molecular imaging and therapeutic strategies. Herein, we report the complexation of lanthanides to a tetrazine-functionalised DO3A macrocyclic chelator and the tetrazine-TCO ligation between the lanthanide(DO3A-tetrazine) complexes and a TCO-PEG₄-functionalised rhodamine as a model agent. The luminescent and magnetic properties of the resultant dual-modal conjugates are described. The tetrazine moiety was found to sensitise terbium luminescence, resulting in a 'turn-off' effect upon its transformation to the dihydropyridazine linker, with the rhodamine moiety then dominating the fluorescence emissions. The T_1 relaxivities of Gd(DO3A-tetrazine) and Gd(DO3A-PEG₄-rhodamine) were found to be similar to [Gd(DOTA)]⁻ (Dotarem®). As a proof-of-concept *in vivo* test, the click conjugates were delivered to mice brains using the combination of focused ultrasound and microbubbles, with neuron uptake of the probes observed.

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Introduction

Simple and reliable methods for covalently linking molecular entities are essential in the development of targeted and multi-modal molecular imaging and therapeutic strategies. Covalent linkages are important for the introduction of targeting vectors to maximise signal at the sites of interest and minimise off-target responses and toxicity.^{1,2} Covalent linkages also enable the introduction of additional imaging modalities,^{3,4} therapeutic loads^{5,6} or chemical modifiers to optimise the physical⁷ and biological properties of imaging agents towards their specific applications.^{8,9}

The use of catalyst-free 'click' chemistry has received much interest for the modular combination of molecular fragments, with fast kinetics under mild conditions and the absence of toxic by-products.^{10–14} The exclusion of non-reactants simplifies purification procedures and is necessary where additional reagents would interact adversely with molecular components

or biological systems. Such scenarios include the undesired chelation of catalytic copper in place of a desired radiometal¹⁵ or changes in pH leading to the disruption of biomolecular structure and activity.^{16,17}

The inverse-electron demand Diels–Alder (IEDDA) reaction between *trans*-cyclooctene (TCO) and tetrazine has been demonstrated to be a highly effective and versatile metal-free conjugation method for biomedical applications.¹⁸ TCO-tetrazine conjugation has demonstrated high specificity and bioorthogonality, as well as fast kinetics (10^3 – 10^6 M⁻¹ s⁻¹) in biocompatible solvents at dilute concentrations (10 nM).^{19–21} Upon formation of the stable 1,4-dihydropyridazine linker and its subsequent oxidation to pyridazine, N₂ is released as the sole by-product.²² TCO-tetrazine conjugation is highly versatile for biological applications, with recent examples including radiolabelled microbubbles,²³ radioimmunoconjugates in a pre-targeting approach,^{24–26} and the *in vivo* labelling of biomarkers in cells^{27,28} and in organisms.²⁹

The lanthanide series holds several roles in biomedical applications. Several lanthanide isotopes are currently in clinical use or have demonstrated promise as radionuclides for: positron emission tomography (lanthanum-132, lanthanum-133, terbium-152);^{30–32} alpha therapy (terbium-149),³³ beta therapy (terbium-161, lutetium-177)^{34,35} and Auger therapy (terbium-161).³⁴ Gadolinium chelates are widely used as longitudinal (T_1) contrast agents in magnetic resonance imaging (MRI), while dysprosium(III) holds potential as a high-field transverse (T_2) contrast agent through Curie spin relaxation

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and as paramagnetically shifted (PARASHIFT) agents for MRI.^{36,37} Europium and terbium have been used as luminescent optical imaging probes,³⁸ with targeting moieties in some cases serving a dual purpose as a sensitiser for long-lived luminescence.^{39,40} Luminescence emission from lanthanide centres can be made switchable, as demonstrated by Zheng *et al.* using a terbium-quinolinone-tetrazine probe for luminogenic metabolic labelling.⁴¹ In this example, the IEDDA reaction of the tetrazine moiety with a TCO-glycan resulted in the loss of quenching by luminescence resonance energy transfer by the tetrazine moiety and resulted in 'turn-on' sensitised terbium emission.⁴¹

Free lanthanide ions have exhibited toxic effects, interfering with signal pathways mediated by the similarly sized Ca^{2+} ion,⁴² and gadolinium deposition in the brain has been reported.^{43,44} Effective macrocyclic chelators that can bind the metal centre without impinging on their functionality⁴⁵ are therefore essential for their use in biological systems.

The dodecane tetraacetic acid (DOTA) macrocycle has been extensively used as a lanthanide chelator, forming stable octadentate complexes through its four amine and four acetate moieties. The $[\text{Gd}(\text{DOTA})]^-$ complex is currently in widespread clinical use as the MRI contrast agent, gadoteric acid (Dotarem®).⁴⁶ A bifunctional analogue of DOTA, 1,4,7,10-tetraazacyclododecane-1,4,7-triacetic acid (DO3A), including coordination from a carbonyl in the amide linker to maintain octadentate coordination, has also been employed as a chelator for Ln^{3+} ions and the chemically-similar Y^{3+} ion.^{47–50} Notable examples of its clinical use include lutetium-177 vipivotide tetraxetan for the treatment of prostate-specific membrane antigen (PSMA)-positive tumours,⁴⁷ lutetium-177 oxodotreotide for the treatment of neuroendocrine tumours⁴⁸ and yttrium-90, terbium-161 or lutetium-177 edotreotide for the treatment of somatostatin receptor-positive tumours.^{49,50}

In 2013, Evans *et al.* reported the development of tetrazine-functionalised DOTA-based chelates for gallium-68, and subsequently demonstrated their use for the radiolabelling of the epidermal growth factor receptor-selective monoclonal antibody, Cetuximab, using both pre-targeting and direct conjugation strategies.^{51,52} Recently, we have reported the synthesis and properties of Gd, Eu and Tb(DO3A-tetrazine) and the IEDDA conjugation of the Gd(DO3A-tetrazine) with a cyclic RGD peptide shuttle.⁵³

In this work, we report the synthesis of Dy and Lu(DO3A-tetrazine), the conjugation of these, and previous, Ln complexes (Ln = Eu, Gd, Tb)⁵³ to a TCO-functionalised rhodamine fluorophore as a model agent (Scheme 1), and the luminescent and magnetic properties of the complexes and conjugates. An 'always-on' rhodamine-piperazine fluorescent tag was used as the complementary TCO-functionalised fragment. This resulted in analogous structures to the dual-modal MRI/optical Gd(DO3A-rhodamine) and Gd(DO3A-pip-rhodamine) probes we have previously reported for pH-responsivity within tumour cells and for neuron labelling respectively.^{54,55} In place of an ethylene bridge or direct amide coupling, a poly(ethylene glycol) (PEG) linker was incorporated due to its use in conjugates for biological applications as a water-soluble, inert, and biocompatible linker.⁵⁶ This approach illustrates a simple biorthogonal introduction of a fluorescent tag which could be extended to targeting groups or alternative imaging modalities.

To demonstrate their potential as dual-modal probes for *in vivo* imaging applications, we administered the conjugates to mice and carried out post-mortem assessments of its uptake in the brain and its clearance through the kidneys and liver. Analogous to our previous study,⁵⁵ we have delivered enhanced concentrations of our probes to the brain using the combination of focused ultrasound and microbubbles as most



Scheme 1 Synthesis of biorthogonal click reaction between Ln-tetrazine complex, Ln.L^1 and Rho-Pip-PEG₄-TCO, 10.



compounds larger than 400–500 Da are otherwise unable to enter the brain parenchyma.^{55,57,58} This non-invasive technique involves the ultrasound-driven oscillations of systemically-administered microbubbles to exert mechanical stress on the cerebral vasculature and transiently enhance blood–brain barrier permeability (Fig. 1).⁵⁹ This proof-of-concept study aims to demonstrate the potential of tetrazine-TCO ligation for the modular and biocompatible syntheses of lanthanide-based conjugates towards biomedical applications.

Results and discussion

Synthesis of lanthanide complexes

The octadentate DO3A-tetrazine macrocycle, **L**¹, was synthesised as previously reported (Schemes S1 and S2).^{51,53,60} The final addition of the chloroacetyl tetrazine arm to the *t*Bu₃DO3A-HBr macrocycle was carried out under milder conditions using NaHCO₃ (7 equiv.) and KI (0.1 equiv.) at 50 °C for 3 days, which resulted in reduced decomposition relative to previous observations.⁵³ The macrocycle was deprotected by TFA, followed by lanthanide complexation. The rhodamine-TCO moiety, **10**, was synthesised from an ‘always on’ rhodamine-piperazine, **9**, and a TCO-PEG₄-NHS ester.

The rhodamine-piperazine was synthesised as previously reported from rhodamine B and 1-Boc-piperazine by amide coupling using HBTU and triethylamine, followed by a subsequent TFA-catalysed Boc deprotection (Scheme S3).^{61,62} The rhodamine-TCO moiety, **10**, was formed by NHS-ester activated amide coupling between the commercially-available TCO-PEG₄-NHS and rhodamine-piperazine in anhydrous chloroform (Scheme S4).

Bioorthogonal ‘click’ chemistry

The final complexes were synthesised by IEDDA cycloaddition between Rho-Pip-PEG₄-TCO, **10**, and Ln(DO3A-tetrazine), **Ln.L**¹, in methanol at 25 °C (Scheme 1). Despite literature reports of the tetrazine-TCO cycloaddition reaching completion in as little as 15 minutes,⁵² we found the reaction between the two

moieties to be slow. The reaction was monitored by LCMS and found to require at least 24 hours before full consumption of the Rho-Pip-PEG₄-TCO starting material. These observations were most likely a result of low concentrations of reaction mixtures (5–10 mM) or potential isomerisation of the *trans*-cyclooctene to the less reactive *cis*-isomer prior to use.

Photophysical spectroscopy

The absorbance spectra of the tetrazine precursors in PBS (pH = 7.4) all exhibit absorbance at 265–268 nm and a shoulder at 330 nm (Fig. 2A), which each correspond to separate π - π^* transitions.^{63,64} The absorbance at 526 nm arises from the n - π^* transition and the observed pink colour of the complexes. Following cycloaddition, the absorbance profiles for the click products, **M.L**², are dominated by the rhodamine absorbance at 566 nm (Fig. 2B) and these compounds have molar extinction coefficients of 44 500–90 800 M⁻¹ cm⁻¹ in PBS (Table S3). This is in the expected region, given that rhodamine B has a molar extinction coefficient of 106 000 M⁻¹ cm⁻¹.⁶⁵ Analogously, the fluorescence spectra of the click products exhibited fluorescence solely arising from the rhodamine moiety, $\lambda_{em} = 590$ nm (Fig. 2C). All click products, **M.L**², display similar fluorescence quantum yields (ϕ_F) in the range of 0.17–0.2, determined relative to Rhodamine B, which has a $\phi_F = 0.5$ in ethanol (Table S3).^{66,67}

The long-lived photophysical emissions of the terbium and europium complexes were investigated by recording time-gated luminescence spectra (Fig. 2D and Fig. S2D). When excited at $\lambda_{ex} = 265$ nm through the tetrazine moiety, the Tb(DO3A-tetrazine) complex, **Tb.L**¹, displayed characteristic green terbium emission corresponding to the ⁵D₄ → ⁷F_J transitions, indicating that the tetrazine moiety can act as a weak lanthanide sensitizer.⁵³ This is expected as the absorption of the tetrazine (19 011 cm⁻¹) lies below the energy levels of the excited states for Tb³⁺ (⁵D₄ 20 500 cm⁻¹)⁶⁸ and Eu³⁺ (⁵D₁ and ⁵D₂ 19 000–21 000 cm⁻¹).⁶⁹ Weak sensitisation can be rationalised by the useful guideline for effective energy transfer, which requires that the lowest energy triplet state of the sensitizer is



Fig. 1 (A) *In vivo* experimental set-up for the focused ultrasound-assisted delivery of **Ln.L**² to the mouse brain. (B) Radially-oscillating microbubbles in response to ultrasound exert mechanical stress on the cerebral vasculature to disrupt the blood–brain barrier and enable the crossing of probes into the brain parenchyma. Created in Inkscape and <https://BioRender.com>.





Fig. 2 Absorbance spectra of the tetrazine precursors, **4**, L^1 and $Tb.L^1$ at 20 μM , $\lambda_{abs} = 265$ nm (A) and the rhodamine precursors, **9** and **10** and click products, L^2 and $Tb.L^2$ at 5 μM , $\lambda_{abs} = 566$ nm (B) in PBS buffer (pH = 7.4). (C) Fluorescence spectrum of the rhodamine precursor and click products **5**, L^2 and $Tb.L^2$ when $\lambda_{ex} = 566$ nm at 5 μM in PBS buffer (pH = 7.4). (D) Phosphorescence spectrum of the Tb complexes prior- ($Tb.L^1$, $\lambda_{ex} = 265$ nm) and post-IEDDA cycloaddition ($Tb.L^2$) excited at 265 nm and 568 nm at 20 μM in PBS (pH = 7.4) at 600 V and 1000 V.

around 1000–2000 cm^{-1} higher than that of the lanthanide emissive state. This facilitates efficient transfer from the ligand triplet state to the excited lanthanide state whilst limiting competitive back energy transfer.⁷⁰ Moreover, there are several reports of tetrazines acting as fluorescence quenchers for both inorganic and organic dyes.^{41,71,72}

Following IEDDA cycloaddition, the tetrazine is converted into a 1,4-dihydropyridazine, which is no longer able to act as a sensitizer for terbium. This is demonstrated by the almost complete loss of terbium emission for $Tb.L^2$ (Fig. 2D) following IEDDA when excited at 265 nm. This was also observed for the europium analogues, $Eu.L^1$ and $Eu.L^2$, where the red europium emission, corresponding to the $^5D_0 \rightarrow ^7F_3$ transition, is lost following cycloaddition (Fig. S2D). Previous work by our group has demonstrated the ability of the rhodamine moiety to act as an efficient sensitizer when excited at 310 nm.⁵⁴ We have also previously demonstrated lanthanide emission sensitization by Quin C1, an organic formyl peptide receptor 2/lipoxin A4 receptor (FPR2/ALX)-targeting group, and luminescence resonance energy transfer (LRET) from the terbium

centre to rhodamine.³⁹ The distance-dependency of this mechanism of energy transfer aligns with our observations in this case that the rhodamine is unable to sensitize the terbium centre when excited at $\lambda_{ex} = 566$ nm. The distance between the two moieties in $Tb.L^2$ is greater than in the work previously reported, and LRET is therefore poor.³⁹ Nevertheless, the presence of a ‘clickable’ tetrazine moiety in proximity to a lanthanide chelator has been shown to modulate the emission properties of the metal,⁴¹ and can likely be further exploited in other modular systems.

Relaxometric properties

The relaxivity (r_1) of the Gd complexes were measured using a 0.25 T Fast Field Cycling NMR relaxometer at frequencies from 0.01 to 10 MHz; while the relaxivity at 400 MHz was recorded on a 9.4 T MRI preclinical scanner in 0.5 mL solutions in PBS (pH = 7.4) in 3D-printed moulds. The effective Gd^{3+} ion concentration of samples was determined from bulk magnetic susceptibility shifts in 1H NMR spectra.^{73,74}





Fig. 3 (A) NMRD profiles for Gd complexes before and after IEDDA cycloaddition compared to Gd-DOTA, at 25 °C (solid symbols) and 37 °C (hollow symbols); (B) longitudinal relaxivity of Gd complexes at 9.4 T (400 MHz) for phantoms of 1 mL solution in PBS buffer at different concentrations at 25 °C. Concentrations corrected for Gd concentration using bulk magnetic susceptibility shifts.^{73,74}

From the ¹H NMR spectra obtained for the Eu and Tb complexes of L¹ (Fig. S20 and S21) and L² (Fig. S30 and S31), we observed that the Ln(DO3A-tetrazine), Ln.L¹, and Ln(DO3A-PEG₄-rhodamine), Ln.L², were present as the SAP conformation. This indicates that the less favourable conformation for Gd-based contrast agents for MRI compared with TSAP analogues. TSAP conformers have previously demonstrated approximately 50 times faster water exchange at the Gd centre, allowing relaxation to be transferred more rapidly to the bulk water.⁷⁴ Although previous work recorded lifetimes for Eu.L¹ and Tb.L¹ and calculated the hydration number, $q = 1$,⁵³ the hydration numbers for Ln.L² could not be determined due to the loss of lanthanide emission upon conjugation. However, given that the lanthanide coordination environment was unchanged in the formation of Ln.L² and remained octadentate, it is assumed that hydration number of Ln.L² conjugates would remain as 1, with one coordination site for H₂O. Measurements recorded on the fast field cycling relaxometer at 1 MHz at 25 °C show relaxation rates in the order of Gd.L² > Gd-DOTA > Gd.L¹ (Fig. 3A). The minor differences are likely due to the greater molecular weight of Gd.L² and, therefore, its reduced rotational correlation time. However, these differences become insignificant at a higher magnetic field strength, with minimal differences in the relaxivities between the three Gd complexes when measured with a 9.4 T MRI scanner (400 MHz) at 25 °C (Fig. 3B). This suggests that the relaxometric properties and expected MRI properties of the tetrazine-functionalised and conjugated Gd complexes are comparable to the Gd-DOTA clinical standard (Table 1).

Ultrasound-assisted *in vivo* delivery to mouse brains

The click conjugates, Gd.L², Tb.L² and Lu.L² were intravenously injected into wild-type mice ($n = 5$) and delivered to

Table 1 Longitudinal relaxivity data for Gd complexes at 25 °C and 37 °C

Complex	Temperature (°C)	r_1 ($\text{mM}^{-1} \text{s}^{-1}$)		
		0.01 MHz	10 MHz	400 MHz
Gd.L ¹	25	9.32	6.14	2.51
	37	8.44	5.28	—
Gd.L ²	25	10.11	7.44	2.60
	37	9.68	6.80	—
Gd-DOTA	25	9.53	5.13	2.80
	37	7.84	4.03	—

the left hemisphere of mouse brains using the combination of focused ultrasound and microbubbles. For the deliveries of Tb.L² and Lu.L², a peak-negative ultrasound pressure of 0.80 MPa was used, accounting for an 11% attenuation by the mouse skull,⁷⁵ and a dose of 15.6 mg kg⁻¹ was administered. 30 μm -thick formalin-fixed frozen brain sections were subsequently analysed by fluorescence microscopy. In the case of Gd.L², a lower peak-negative ultrasound pressure of 0.53 MPa was emitted and the dose of Gd.L² was doubled to 31.1 mg kg⁻¹. *Ex vivo* MRI scans using T1 Flash 3D and 2D, T1 mapping and T2 TurboRARE 3D sequences were first performed on mice heads prior to fluorescence imaging of brain sections.

The successful ultrasound-assisted disruption of the blood-brain barrier and delivery of Tb.L² and Lu.L² to mouse brains was evidenced by the detection of fluorescence signals from the rhodamine tag in the left hippocampus and thalamus (Fig. 4). Fluorescence signals with intensities greater than the background level were not observed in the contralateral right hemisphere control region, which was not exposed to the ultrasound beam and where blood-brain barrier permeability was





Fig. 4 (A) The left hippocampus of mouse brains was targeted with the focused ultrasound beam, highlighted as a light blue circle. (B) Fluorescence image showing both the left targeted and right control hippocampi following intravenous injection with Lu.L² (orange rectangle in part A). Created in Inkscape and <https://BioRender.com>.

not enhanced. Neuronal uptake in the ultrasound-treated region was observed and was particularly prominent in the dentate gyrus, layers V and VI of the entorhinal cortex and cerebellar granule cells (Fig. 5 and S35), with the identity of fluorescent cells as neurons confirmed by NeuN staining (Fig. S38–S40). These observations mirror those in our pre-

vious study with an amide-coupled Gd(DO3A-pip-rhodamine) analogue, with the cationic and lipophilic nature of the rhodamine moiety enabling permeation of the cell membrane and its uptake by neurons.⁵⁵

However, in the case of Gd.L², gadolinium-enhanced MR contrast was not detected, with no expected hyperintensity discernible in the region of the brain exposed to the ultrasound beam (Fig. S34). Although neuron uptake was observed in the entorhinal cortex and cerebellum in the left hemisphere, the intensity and area of fluorescence at these sites were much-reduced compared to the *in vivo* deliveries of Tb.L² and Lu.L². Rhodamine fluorescence signals were detected in kidney and liver sections from mice injected with Gd.L² and treated with focused ultrasound and microbubbles, but were absent in kidney (Fig. S36) and liver (Fig. S37) sections from a control mouse that was not injected with Gd.L² but was treated with focused ultrasound and microbubbles only. This suggested rapid clearance through renal and hepatic pathways within approximately 10 minutes following intravenous injection.

We speculate that the reduced fluorescence signals from Gd.L² observed in the brain compared with Tb.L² and Lu.L² are due to an increased washout of the unfixed probe from the tissues. Brain samples were immersed in solution for a longer duration (an additional 10 days on average) as part of the MR imaging procedure and underwent an increased number of solution changes. The absence of MR enhancement also suggests that the reduced sensitivity of MRI compared with fluorescence imaging requires increased gadolinium doses for MR enhancement, which may not be attainable using the probe and *ex vivo* MR imaging technique used in this study.



Fig. 5 (Left) Fluorescence images, prior to immunofluorescence staining, of horizontal, 30 μm-thick formalin-fixed frozen brain sections from mice injected with Lu.L² and treated with focused ultrasound and microbubbles and (Right) NeuN-positive staining of horizontal 30 μm-thick frozen sections from the same brain.

Conclusions

This proof-of-concept work describes the use of tetrazine-TCO ligation for synthesising dual-modal lanthanide probes in a



biocompatible and modular strategy. Although MRI/optical imaging was explored here, this conjugation method can be translated to lanthanide-based radiolabelling or the introduction of targeting moieties. The inclusion of the PEG chain enabled the click products, **Ln.L²**, to be soluble in PBS (pH = 7.4) for *in vivo* administration. To improve the applicability of this IEDDA conjugation, the stability of any TCO-bearing reagents would need to be investigated to ensure the cyclooctene remains as the *trans*-isomer to minimise reaction times, which would be especially pertinent to radiochemistry and pre-targeting applications.

The focused ultrasound-assisted delivery of the click products, **Ln.L²**, to the mouse brain and their neuronal uptake demonstrated their potential for *in vivo* applications. Enhanced doses were delivered to the targeted left hemisphere over the contralateral control hemisphere. The relaxivity of the **Gd.L²** complex was too low to be observed in mouse brains using *ex vivo* MRI, although rhodamine fluorescence from the **Tb.L²** and **Lu.L²** complexes was observed.

Overall, tetrazine-TCO ligation is a useful method of conjugation to produce lanthanide-based molecular imaging and therapeutic agents. Further applications will explore the use of luminescence quenching following ligation as a switchable probe and the use of *in vivo* pre-targeting strategies.

Experimental

Synthesis of 'Click' conjugates, **L²** and **Ln.L²**

DO3A-PEG₄-piperazine-rhodamine (L²). **L¹** (7.2 mg, 12.1 μmol, 1.1 eq.) and **10** (9.8 mg, 11.0 μmol, 1 eq.) were dissolved in H₂O (1.25 mL). The solution was stirred at 60 °C for 7 hours. The solvent was removed under vacuum. The crude product was purified by HPLC using prep HPLC gradient method 2 (Table S2). After lyophilisation, a dark purple solid (2.9 mg, 2.0 μmol, 18%) was obtained; ESI-LRMS (ES+): [C₇₈H₁₀₄N₁₂O₁₆]²⁺, (+2) *m/z* 734.9, ESI-HRMS (ES+): anal. For [C₇₈H₁₀₄N₁₂O₁₆]²⁺ [M + 2H]²⁺ calcd: 734.9037, found: 735.3635; UV-Vis (PBS, λ_{max}/nm): 569; *t_R* (prep) = 12.71–12.80 min.

Eu(DO3A-PEG₄-piperazine-rhodamine) (Eu.L²). **Eu.L¹** (10.0 mg, 0.011 mmol, 1.0 eq.) and **10** (12.2 mg, 0.017 mol, 1.5 eq.) were dissolved in MeOH (0.5 mL). The solution was stirred at 25 °C for 24 hours. The solvent was removed under vacuum. The crude product was dissolved in 3 : 1 H₂O : MeCN and purified by HPLC using prep HPLC gradient method 1 (Table S1). After lyophilisation, a dark purple solid **Eu.L²** (5.8 mg, 3.6 μmol, 33%) was obtained; ¹H NMR (400 MHz, CD₃OD) δ_H (ppm) 38.9, 36.4, 35.6, 34.4, 16.3, 14.1, -1.4, -1.8, -2.4, -3.6, -5.2, -6.4, -6.5, -9.4, -11.2, -12.7, -13.8, -14.3, -17.2, -18.1, -18.2, -18.7, -18.9 (only peaks outside of 0 to 10 ppm reported); ESI-LRMS (ES+): [C₇₈H₁₀₄N₁₂O₁₆Eu]⁺, (+) *m/z* 1617.7, ESI-HRMS (ES+): anal. For [C₇₈H₁₀₄N₁₂O₁₆Eu]⁺ [M + H]⁺ calcd: 1617.6906, found: 1617.6907; UV-Vis (PBS, λ_{max}/nm): 569; *t_R* (prep) = 8.26 min.

Gd(DO3A-PEG₄-piperazine-rhodamine) (Gd.L²). **Gd.L¹** (12.1 mg, 16 μmol, 1.0 eq.) was dissolved in H₂O : MeOH (1 : 1,

2 mL). **10** (15.0 mg, 16 μmol, 1.0 eq.) was added to the solution and stirred at 25 °C for 24 hours. The solvent was removed under vacuum and combined with the reaction in H₂O. The solvent was removed under vacuum. The crude product was dissolved in 3 : 1 H₂O : MeCN and purified by HPLC using prep HPLC gradient method 1 (Table S1). After lyophilisation, **Gd.L²** was obtained as a dark purple solid (7 mg, 4 μmol, 27%); ESI-LRMS (ES+): [C₇₈H₁₀₄N₁₂O₁₆Gd]²⁺, (+2) *m/z* 811.9, ESI-HRMS (ES+): anal. For [C₇₈H₁₀₄N₁₂O₁₆Gd]²⁺ [M + 2H]²⁺ calcd: 811.8501, found: 811.8503; UV-Vis (PBS, λ_{max}/nm): 569; *t_R* (prep) = 8.11–8.21 min.

Tb(DO3A-PEG₄-piperazine-rhodamine) (Tb.L²). **Tb.L¹** (12.2 mg, 16.0 μmol, 1.0 eq.) and **10** (10.0 mg, 11.0 μmol, 1.2 eq.) were dissolved in MeOH (0.5 mL). The solution was stirred at 25 °C for 24 hours. The solvent was removed under vacuum. The crude product was dissolved in 3 : 1 H₂O : MeCN and purified by HPLC using prep HPLC gradient method 1 (Table S1). After lyophilisation, a dark purple solid **Tb.L²** (14.0 mg, 9.0 μmol, 56%) was obtained; ¹H NMR (400 MHz, CD₃OD) δ_H (ppm) 277.0, 270.1, 255.8, 236.8, 230.3, 135.3, 128.2, 51.5, 45.8, 34.6, 30.3, 22.2, 15.6, 13.7, 13.6, 13.4, 13.0, 12.6, 12.3, 11.7, 11.1, 10.9, -25.2, -32.3, -40.4, -52.1, -61.4, -70.3, -73.3, -80.8, -104.5, -118.1, -125.5, -130.7, -148.4, -221.7, -226.3, -246.5, -259.7, -384.6, -389.5, -396.2, -428.3 (only peaks outside of -10 to 10 ppm reported); ESI-LRMS (ES+): [C₇₈H₁₀₄N₁₂O₁₆Tb]²⁺, (+2) *m/z* 812.4, ESI-HRMS (ES+): anal. For [C₇₈H₁₀₄N₁₂O₁₆Tb]²⁺ [M + 2H]²⁺ calcd: 812.3513, found: 812.3514; UV-Vis (PBS, λ_{max}/nm): 569; *t_R* (prep) = 8.20–8.24 min.

Dy(DO3A-PEG₄-piperazine-rhodamine) (Dy.L²). **Dy.L¹** (13.4 mg, 18.0 μmol, 1.0 eq.) was dissolved in MeOH (0.5 mL). **10** (16.1 mg, 18.0 μmol, 1.0 eq.) was added to the solution and stirred at 25 °C for 24 hours. The solvent was removed under vacuum. The crude product was dissolved in 3 : 1 H₂O : MeCN and purified by HPLC using prep HPLC gradient method 1 (Table S1). After lyophilisation, a purple solid **Dy.L²** (28.4 mg, 17.4 μmol, 96%) was obtained; ¹H NMR (500 MHz, CD₃OD) δ_H (ppm) 367.9, 347.1, 325.8, 238.0, 226.7, 25.5, -13.9, -14.3, -29.2, -38.1, -43.9, -51.8, -53.4, -60.7, -80.8, -97.3, -100.0, -117.0, -133.8, -159.3, -167.3, -193.4, -221.0 (only peaks outside of -10 to 10 ppm reported); ESI-LRMS (ES+): [C₇₈H₁₀₄N₁₂O₁₆Dy]⁺, (+) *m/z* 1628.7, ESI-HRMS (ES+): anal. For [C₇₈H₁₀₄N₁₂O₁₆Dy]⁺ [M + H]⁺ calcd: 1628.6985, found: 1628.6960; UV-Vis (PBS, λ_{max}/nm): 569; *t_R* (prep) = 8.17–8.30 min.

Lu(DO3A-PEG₄-piperazine-rhodamine) (Lu.L²). **Lu.L¹** (7.2 mg, 9.0 μmol, 1.0 eq.) was dissolved in MeOH (5 mL). **10** (8.0 mg, 9.0 μmol, 1.0 eq.) was added to the solution and stirred at 60 °C for 32 hours. The solvent was removed under vacuum. The crude product was purified by HPLC using prep HPLC gradient method 2 (Table S2). After lyophilisation, a purple solid **Lu.L²** (4.0 mg, 2.4 μmol, 29%) was obtained; ¹H NMR (500 MHz, D₂O) δ_H (ppm) 8.63 (s, 1H), 8.37 (d, *J* = 8.0 Hz, 2H), 7.80 (d, *J* = 7.7 Hz, 2H), 7.63 (t, *J* = 8.4 Hz, 3H), 7.45 (s, 4H), 7.41 (s, 1H), 7.33 (d, *J* = 7.6 Hz, 1H), 7.18 (s, 2H), 6.94 (d, *J* = 9.6 Hz, 2H), 4.63 (t, *J* = 16.3 Hz, 3H), 4.42 (d, *J* = 15.1 Hz, 1H),



4.35 (d, $J = 14.5$ Hz, 2H), 3.93–3.79 (m, 3H), 3.73–3.11 (m, 57H), 2.99–2.36 (m, 42H), 2.30 (d, $J = 13.3$ Hz, 1H), 2.24–2.18 (m, 1H), 2.17–1.92 (m, 4H), 1.92–1.40 (m, 7H), 1.19 (q, $J = 6.8$ Hz, 13H); ESI-LRMS (ES⁺): [C₇₈H₁₀₄N₁₂O₁₆Lu]²⁺, (+2) m/z 820.4, ESI-HRMS (ES⁺): anal. For [C₇₈H₁₀₄N₁₂O₁₆Lu]⁺ [M + H]⁺ calcd: 1639.7101, found: 1639.7040; UV-Vis (PBS, λ_{max} /nm): 569; t_R (prep) = 12.70–12.80 min.

Author contributions

H. C., W. L. K. C. and B. P. W. synthesised the compounds, G. T. M. purified the complexes. H. C., W. L. K. C. and G. T. M. performed the optical spectroscopy and relaxivity measurements. W. L. K. C., G. G. and S. V. M. performed the *in vivo* experiments and W. L. K. C. and C. Y. performed the histology. Y. Y. performed the MRI experiments and Y. Y. and G. T. M. processed the MRI data. N. J. L. supervised the work. All the authors contributed to the preparation and writing of the manuscript.

Conflicts of interest

There are no conflicts to declare.

Data availability

All the relevant research data is contained within the manuscript and supplementary information (SI). No databases have been used and no references to such databases are contained in the manuscript or SI. Supplementary information is available. See DOI: <https://doi.org/10.1039/d5qi01745a>.

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References

- 1 A. M. Morawski, G. A. Lanza and S. A. Wickline, Targeted contrast agents for magnetic resonance imaging and ultrasound, *Curr. Opin. Biotechnol.*, 2005, **16**, 89–92.
- 2 M. Srinivasarao, C. V. Galliford and P. S. Low, Principles in the design of ligand-targeted cancer therapeutics and imaging agents, *Nat. Rev. Drug Discovery*, 2015, **14**, 203–219.
- 3 S. Lee, J. Xie and X. Chen, Peptide-based probes for targeted molecular imaging, *Biochemistry*, 2010, **49**, 1364–1376.
- 4 A. Louie, Multimodality imaging probes: design and challenges, *Chem. Rev.*, 2010, **110**, 3146–3195.
- 5 A. Rudra, J. Li, R. Shakur, S. Bhagchandani and R. Langer, Trends in therapeutic conjugates: bench to clinic, *Bioconjugate Chem.*, 2020, **31**, 462–473.
- 6 Z. Fu, S. Li, S. Han, C. Shi and Y. Zhang, Antibody drug conjugate: the “biological missile” for targeted cancer therapy, *v*, 2022, **7**, 93.
- 7 I. Ekladios, Y. L. Colson and M. W. Grinstaff, Polymer-drug conjugate therapeutics: advances, insights and prospects, *Nat. Rev. Drug Discovery*, 2019, **18**, 273–294.
- 8 A. Kolate, D. Baradia, S. Patil, I. Vhora, G. Kore and A. Misra, PEG – a versatile conjugating ligand for drugs and drug delivery systems, *J. Controlled Release*, 2014, **192**, 67–81.
- 9 A. Wijesinghe, S. Kumari and V. Booth, Conjugates for use in peptide therapeutics: a systematic review and meta-analysis, *PLoS One*, 2022, **17**, e0255753.
- 10 N. J. Agard, J. A. Prescher and C. R. Bertozzi, A strain-promoted [3 + 2] azide–alkyne cycloaddition for covalent modification of biomolecules in living systems, *J. Am. Chem. Soc.*, 2004, **126**, 15046–15047.
- 11 J. M. Baskin, J. A. Prescher, S. T. Laughlin, N. J. Agard, P. V. Chang, I. A. Miller, A. Lo, J. A. Codelli and C. R. Bertozzi, Copper-free click chemistry for dynamic *in vivo* imaging, *Proc. Natl. Acad. Sci. U. S. A.*, 2007, **104**, 16793–16797.
- 12 J. C. Jewett, E. M. Sletten and C. R. Bertozzi, Rapid Cu-free click chemistry with readily synthesized biarylazacyclooctynes, *J. Am. Chem. Soc.*, 2010, **132**, 3688–3690.
- 13 C. S. McKay, J. Moran and J. P. Pezacki, Nitrones as dipoles for rapid strain-promoted 1,3-dipolar cycloadditions with cyclooctynes, *Chem. Commun.*, 2010, **46**, 931–933.
- 14 K. Lang, L. Davis, S. Wallace, M. Mahesh, D. J. Cox, M. L. Blackman, J. M. Fox and J. W. Chin, Genetic encoding of bicyclononynes and *trans*-cyclooctenes for site-specific protein labeling *in vitro* and in live mammalian cells via rapid fluorogenic Diels–Alder reactions, *J. Am. Chem. Soc.*, 2012, **134**, 10317–10320.
- 15 T. J. Wadas, E. H. Wong, G. R. Weisman and C. J. Anderson, Coordinating radiometals of copper, gallium, indium, yttrium, and zirconium for PET and SPECT imaging of disease, *Chem. Rev.*, 2010, **110**, 2858–2902.
- 16 M. Tollinger, K. A. Crowhurst, L. E. Kay and J. D. Forman-Kay, Site-specific contributions to the pH dependence of



- protein stability, *Proc. Natl. Acad. Sci. U. S. A.*, 2003, **100**, 4545–4550.
- 17 K. Talley and E. Alexov, On the pH-optimum of activity and stability of proteins, *Proteins*, 2010, **78**, 2699–2706.
 - 18 M. L. Blackman, M. Royzen and J. M. Fox, Tetrazine ligation: Fast bioconjugation based on inverse-electron-demand Diels-Alder reactivity, *J. Am. Chem. Soc.*, 2008, **130**, 13518–13519.
 - 19 J. C. T. Carlson, H. Mikula and R. Weissleder, Unraveling tetrazine-triggered bioorthogonal elimination enables chemical tools for ultrafast release and universal cleavage, *J. Am. Chem. Soc.*, 2018, **140**, 3603–3612.
 - 20 D. S. Liu, A. Tangpeerachaikul, R. Selvaraj, M. T. Taylor, J. M. Fox and A. Y. Ting, Diels-Alder cycloaddition for fluorophore targeting to specific proteins inside living cells, *J. Am. Chem. Soc.*, 2012, **134**, 792–795.
 - 21 J. Schoch, M. Staudt, A. Samanta, M. Wiessler and A. Jäschke, Site-specific one-pot dual labeling of DNA by orthogonal cycloaddition chemistry, *Bioconjugate Chem.*, 2012, **23**, 1382–1386.
 - 22 D. Svatunek, M. Wilkovič, L. Hartmann, K. N. Houk and H. Mikula, Uncovering the key role of distortion in bioorthogonal tetrazine tools that defy the reactivity/stability trade-off, *J. Am. Chem. Soc.*, 2022, **144**, 8171–8177.
 - 23 J. H. Teh, M. Braga, L. Allott, C. Barnes, J. Hernandez-Gil, M.-X. Tang, E. O. Aboagye and N. J. Long, A kit-based aluminium- ^{18}F fluoride approach to radiolabelled microbubbles, *Chem. Commun.*, 2021, **57**, 11677–11680.
 - 24 J.-P. Meyer, J. L. Houghton, P. Kozłowski, D. Abdel-Atti, T. Reiner, N. V. K. Pillarsetty, W. W. Scholz, B. M. Zeglis and J. S. Lewis, ^{18}F -Based pretargeted PET imaging based on bioorthogonal Diels-Alder click chemistry, *Bioconjugate Chem.*, 2016, **27**, 298–301.
 - 25 S. Poty, R. Membreno, J. M. Glaser, A. Ragupathi, W. W. Scholz, B. M. Zeglis and J. S. Lewis, The inverse electron-demand Diels-Alder reaction as a new methodology for the synthesis of ^{225}Ac -labelled radioimmunoconjugates, *Chem. Commun.*, 2018, **54**, 2599–2602.
 - 26 V. Shalgunov, S. Lopes van den Broek, I. V. Andersen, R. García Vázquez, N. R. Raval, M. Palner, Y. Mori, G. Schäfer, B. Herrmann, H. Mikula, N. Beschorner, M. Nedergaard, S. Syvänen, M. Barz, G. M. Knudsen, U. M. Battisti and M. M. Herth, Pretargeted imaging beyond the blood-brain barrier, *RSC Med. Chem.*, 2023, **14**, 444–453.
 - 27 H. E. Murrey, J. C. Judkins, C. W. am Ende, T. E. Ballard, Y. Fang, K. Riccardi, L. Di, E. R. Guilmette, J. W. Schwartz, J. M. Fox and D. S. Johnson, Systematic evaluation of bioorthogonal reactions in live cells with clickable HaloTag ligands: implications for intracellular imaging, *J. Am. Chem. Soc.*, 2015, **137**, 11461–11475.
 - 28 P. Werther, K. Yserentant, F. Braun, N. Kaltwasser, C. Popp, M. Baalman, D.-P. Herten and R. Wombacher, Live-cell localization microscopy with a fluorogenic and self-blinking tetrazine probe, *Angew. Chem., Int. Ed.*, 2020, **59**, 804–810.
 - 29 S. T. Laughlin, J. M. Baskin, S. L. Amacher and C. R. Bertozzi, In vivo imaging of membrane-associated glycans in developing zebrafish, *Science*, 2008, **320**, 664–667.
 - 30 E. Aluicio-Sarduy, T. E. Barnhart, J. Weichert, R. Hernandez and J. W. Engle, Cyclotron-produced ^{132}La as a PET imaging surrogate for therapeutic ^{225}Ac , *J. Nucl. Med.*, 2021, **62**, 1012–1015.
 - 31 B. J. B. Nelson, S. Ferguson, M. Wuest, J. Wilson, M. J. M. Duke, S. Richter, H. Soenke-Jans, J. D. Andersson, F. Juengling and F. Wuest, First in vivo and phantom imaging of cyclotron-produced ^{133}La as a theranostic radionuclide for ^{225}Ac and ^{135}La , *J. Nucl. Med.*, 2022, **63**, 584–590.
 - 32 C. Müller, A. Singh, C. A. Umbricht, H. R. Kulkarni, K. Johnston, M. Benešová, S. Senftleben, D. Müller, C. Vermeulen, R. Schibli, U. Köster, N. P. van der Meulen and R. P. Baum, Preclinical investigations and first-in-human application of ^{152}Tb -PSMA-617 for PET/CT imaging of prostate cancer, *EJNMMI Res.*, 2019, **9**, 68.
 - 33 C. Müller, C. Vermeulen, U. Köster, K. Johnston, A. Türler, R. Schibli and N. P. van der Meulen, Alpha-PET with terbium-149: evidence and perspectives for radiotheragnostics, *EJNMMI Radiopharm. Chem.*, 2016, **1**, 5.
 - 34 C. Müller, C. A. Umbricht, N. Gracheva, V. J. Tschan, G. Pellegrini, P. Bernhardt, J. R. Zeevaart, U. Köster, R. Schibli and N. P. van der Meulen, Terbium-161 for PSMA-targeted radionuclide therapy of prostate cancer, *Eur. J. Nucl. Med. Mol. Imaging*, 2019, **46**, 1919–1930.
 - 35 O. Sartor, J. de Bono, K. N. Chi, K. Fizazi, K. Herrmann, K. Rahbar, S. T. Tagawa, L. T. Nordquist, N. Vaishampayan, G. El-Haddad, C. H. Park, T. M. Beer, A. Armour, W. J. Pérez-Contreras, M. DeSilvio, E. Kpamegan, G. Gericke, R. A. Messmann, M. J. Morris and B. J. Krause, Lutetium-177-PSMA-617 for metastatic castration-resistant prostate cancer, *N. Engl. J. Med.*, 2021, **385**, 1091–1103.
 - 36 J. W. Bulte, C. Wu, M. W. Brechbiel, R. A. Brooks, J. Vymazal, M. Holla and J. A. Frank, Dysprosium-DOTA-PAMAM dendrimers as macromolecular T2 contrast agents - Preparation and relaxometry, *Invest. Radiol.*, 1998, **33**, 841–845.
 - 37 P. K. Senanayake, N. J. Rogers, K.-L. N. A. Finney, P. Harvey, A. M. Funk, J. I. Wilson, D. O'Hogain, R. Maxwell, D. Parker and A. M. Blamire, A new paramagnetically shifted imaging probe for MRI, *Magn. Reson. Med.*, 2017, **77**, 1307–1317.
 - 38 S. J. Butler, L. Lamarque, R. Pal and D. Parker, EuroTracker dyes: highly emissive europium complexes as alternative organelle stains for live cell imaging, *Chem. Sci.*, 2014, **5**, 1750–1756.
 - 39 T. Boltersdorf, F. N. E. Gavins and N. J. Long, Long-lived lanthanide emission via a pH-sensitive and switchable LRET complex, *Chem. Sci.*, 2021, **12**, 8740–8745.
 - 40 G. T. McMullon, A. Ezdoglian, A. C. Booth, P. Jimenez-Royo, P. S. Murphy, G. Jansen, C. J. van der Laken and S. Faulkner, Synthesis and characterization of folic acid-conjugated terbium complexes as luminescent probes for



- targeting folate receptor-expressing cells, *J. Med. Chem.*, 2024, **67**, 14062–14076.
- 41 J. Zheng, Q. Zhan, L. Jiang, D. Xing, T. Zhang and K.-L. Wong, A bioorthogonal time-resolved luminogenic probe for metabolic labelling and imaging of glycans, *Inorg. Chem. Front.*, 2020, **7**, 4062–4069.
- 42 M. Baykara, M. Ozcan, M. Bilgen and H. Kelestimur, Effects of gadolinium and gadolinium chelates on intracellular calcium signaling in sensory neurons, *Neurosci. Lett.*, 2019, **707**, 134295.
- 43 R. C. Semelka, J. Ramalho, A. Vakharia, M. AlObaidy, L. M. Burke, M. Jay and M. Ramalho, Gadolinium deposition disease: Initial description of a disease that has been around for a while, *Magn. Reson. Imaging*, 2016, **34**, 1383–1390.
- 44 T. Kanda, K. Ishii, H. Kawaguchi, K. Kitajima and D. Takenaka, High signal intensity in the dentate nucleus and globus pallidus on unenhanced T1-weighted MR images: relationship with increasing cumulative dose of a gadolinium-based contrast material, *Radiology*, 2014, **270**, 834–841.
- 45 T. J. Clough, L. Jiang, K.-L. Wong and N. J. Long, Ligand design strategies to increase stability of gadolinium-based magnetic resonance imaging contrast agents, *Nat. Commun.*, 2019, **10**, 1420.
- 46 J. Wahsner, E. M. Gale, A. Rodríguez-Rodríguez and P. Caravan, Chemistry of MRI contrast agents: current challenges and new frontiers, *Chem. Rev.*, 2019, **119**, 957–1057.
- 47 J. Fallah, S. Agrawal, H. Gittleman, M. H. Fiero, S. Subramaniam, C. John, W. Chen, T. K. Ricks, G. Niu, A. Fotenos, M. Wang, K. Chiang, W. F. Pierce, D. L. Suzman, S. Tang, R. Pazdur, L. Amiri-Kordestani, A. Ibrahim and P. G. Kluetz, FDA approval summary: Lutetium Lu 177 Vipivotide Tetraxetan for patients with metastatic castration-resistant prostate cancer, *Clin. Cancer Res.*, 2023, **29**, 1651–1657.
- 48 J. Strosberg, G. El-Haddad, E. Wolin, A. Hendifar, J. Yao, B. Chasen, E. Mittra, P. L. Kunz, M. H. Kulke, H. Jacene, D. Bushnell, T. M. O'Dorisio, R. P. Baum, H. R. Kulkarni, M. Caplin, R. Lebtahi, T. Hobday, E. Delpassand, E. Van Cutsem, A. Benson, R. Srirajaskanthan, M. Pavel, J. Mora, J. Berlin, E. Grande, N. Reed, E. Seregni, K. Oberg, M. Lopera Sierra, P. Santoro, T. Thevenet, J. L. Erion, P. Ruzniewski, D. Kwekkeboom and E. Krenning, Phase 3 trial of ¹⁷⁷Lu-Dotatate for midgut neuroendocrine tumors, *N. Engl. J. Med.*, 2017, **376**, 125–135.
- 49 A. Frilling, F. Weber, F. Saner, A. Bockisch, M. Hofmann, J. Mueller-Brand and C. E. Broelsch, Treatment with ⁹⁰Y- and ¹⁷⁷Lu-DOTATOC in patients with metastatic neuroendocrine tumors, *Surgery*, 2006, **140**, 968–976.
- 50 R. P. Baum, A. Singh, M. Benešová, C. Vermeulen, S. Gnesin, U. Köster, K. Johnston, D. Müller, S. Senftleben, H. R. Kulkarni, A. Türler, R. Schibli, J. O. Prior, N. P. van der Meulen and C. Müller, Clinical evaluation of the radiolanthanide terbium-152: first-in-human PET/CT with ¹⁵²Tb-DOTATOC, *Dalton Trans.*, 2017, **46**, 14638–14646.
- 51 H. L. Evans, L. Carroll, E. O. Aboagye and A. C. Spivey, Bioorthogonal chemistry for ⁶⁸Ga radiolabelling of DOTA-containing compounds, *J. Labelled Compd. Radiopharm.*, 2014, **57**, 291–297.
- 52 H. L. Evans, Q.-D. Nguyen, L. S. Carroll, M. Kaliszczak, F. J. Twyman, A. C. Spivey and E. O. Aboagye, A bioorthogonal ⁶⁸Ga-labelling strategy for rapid *in vivo* imaging, *Chem. Commun.*, 2014, **50**, 9557–9560.
- 53 B. Woolley, Y. Wu, L. Xiong, H.-F. Chau, J. Zhang, G.-L. Law, K.-L. Wong and N. J. Long, Lanthanide-tetrazine probes for bio-imaging and click chemistry, *Chem. Sci.*, 2025, **16**, 3588–3597.
- 54 C. Rivas, G. J. Stasiuk, J. Gallo, F. Minuzzi, G. A. Rutter and N. J. Long, Lanthanide(III) complexes of rhodamine-DO3A conjugates as agents for dual-modal imaging, *Inorg. Chem.*, 2013, **52**, 14284–14293.
- 55 S. V. Morse, T. Boltersdorf, B. I. Harriss, T. G. Chan, N. Baxan, H. S. Jung, A. N. Pouliopoulos, J. J. Choi and N. J. Long, Neuron labeling with rhodamine-conjugated Gd-based MRI contrast agents delivered to the brain via focused ultrasound, *Theranostics*, 2020, **10**, 2659–2674.
- 56 G. T. Hermanson, Chapter 18 - PEGylation and Synthetic Polymer Modification, in *Bioconjugate Techniques*, ed. G. T. Hermanson, Academic Press, 3rd edn, 2013, pp. 787–838.
- 57 K. Hynynen, N. McDannold, N. Vykhodtseva and F. A. Jolesz, Noninvasive MR imaging-guided focal opening of the blood-brain barrier in rabbits, *Radiology*, 2001, **220**, 640–646.
- 58 S. V. Morse, T. Boltersdorf, T. G. Chan, F. N. E. Gavins, J. J. Choi and N. J. Long, *In vivo* delivery of a fluorescent FPR2/ALX-targeted probe using focused ultrasound and microbubbles to image activated microglia, *RSC Chem. Biol.*, 2020, **1**, 385–389.
- 59 J. H. Bezer, P. Prentice, W. Lim Kee Chang, S. V. Morse, K. Christensen-Jeffries, C. J. Rowlands, A. S. Kozlov and J. J. Choi, Microbubble dynamics in brain microvessels, *PLoS One*, 2025, **20**, e0310425.
- 60 J. Hernández-Gil, M. Braga, B. I. Harriss, L. S. Carroll, C. H. Leow, M.-X. Tang, E. O. Aboagye and N. J. Long, Development of ⁶⁸Ga-labelled ultrasound microbubbles for whole-body PET imaging, *Chem. Sci.*, 2019, **10**, 5603–5615.
- 61 L. Bi, WIPO, Michigan Technological University, WO2014/063033 A2, 2014.
- 62 T. Boltersdorf, J. Ansari, E. Y. Senchenkova, J. Groeper, D. Pajonczyk, S. A. Vital, G. Kaur, J. S. Alexander, T. Vogl, U. Rescher, N. J. Long and F. N. E. Gavins, Targeting of Formyl Peptide Receptor 2 for *in vivo* imaging of acute vascular inflammation, *Theranostics*, 2020, **10**, 6599–6614.
- 63 M. Plugge, V. Alain-Rizzo, P. Audebert and A. M. Brouwer, Excited state dynamics of 3,6-diaryl-1,2,4,5-tetrazines. Experimental and theoretical studies, *J. Photochem. Photobiol., A*, 2012, **234**, 12–20.
- 64 F. Miomandre and P. Audebert, 1,2,4,5-Tetrazines: an intriguing heterocycles family with outstanding characteristics in the field of luminescence and electrochemistry, *J. Photochem. Photobiol., C*, 2020, **44**, 100372.



- 65 S. K. Sahoo and R. A. Jockusch, The fluorescence properties of cationic rhodamine B in the gas phase, *J. Photochem. Photobiol., A*, 2011, **220**, 173–178.
- 66 T. Karstens and K. Kobs, Rhodamine-B and rhodamine-101 as reference substances for fluorescence quantum yield measurements, *J. Phys. Chem.*, 1980, **84**, 1871–1872.
- 67 A. M. Brouwer, Standards for photoluminescence quantum yield measurements in solution (IUPAC Technical Report), *Pure Appl. Chem.*, 2011, **83**, 2213–2228.
- 68 S. Sato and M. Wada, Relations between Intramolecular energy transfer efficiencies and triplet state energies in rare earth β -diketone chelates, *Bull. Chem. Soc. Jpn.*, 2006, **43**, 1955–1962.
- 69 K. Binnemans, Interpretation of europium(III) spectra, *Coord. Chem. Rev.*, 2015, **295**, 1–45.
- 70 J.-C. G. Bünzli and S. V. Eliseeva, Basics of Lanthanide Photophysics, in *Lanthanide Luminescence: Photophysical, Analytical and Biological Aspects*, ed. P. Hänninen and H. Härmä, Springer, Berlin, Heidelberg, 2011, pp. 1–45.
- 71 A. Wiczorek, T. Buckup and R. Wombacher, Rigid tetrazine fluorophore conjugates with fluorogenic properties in the inverse electron demand Diels-Alder reaction, *Org. Biomol. Chem.*, 2014, **12**, 4177–4185.
- 72 H. Wu and N. K. Devaraj, Advances in tetrazine bioorthogonal chemistry driven by the synthesis of novel tetrazines and dienophiles, *Acc. Chem. Res.*, 2018, **51**, 1249–1259.
- 73 D. F. Evans, The determination of the paramagnetic susceptibility of substances in solution by nuclear magnetic resonance, *J. Chem. Soc.*, 1959, 2003–2005.
- 74 D. M. Corsi, C. Platas-Iglesias, H. van Bekkum and J. A. Peters, Determination of paramagnetic lanthanide(III) concentrations from bulk magnetic susceptibility shifts in NMR spectra, *Magn. Reson. Chem.*, 2001, **39**, 723–726.
- 75 S. V. Morse, A. N. Pouliopoulos, T. G. Chan, M. J. Copping, J. Lin, N. J. Long and J. J. Choi, Rapid short-pulse ultrasound delivers drugs uniformly across the murine blood-brain barrier with negligible disruption, *Radiology*, 2019, **291**, 458–465.

