

REVIEW

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Recent progress in nanomaterials for bacteria-related tumor therapy

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Many studies suggest that tumor microbiome closely relates to the oncogenesis and anti-tumor responses in multiple cancer types (e.g., colorectal cancer (CRC), breast cancer, lung cancer and pancreatic cancer), thereby raising an emerging research area of bacteria-related tumor therapy. Nanomaterials have long been used for both cancer and bacterial infection treatment, holding great potential for bacteria-related tumor therapy. In this review, we summarized recent progress in nanomaterials for bacteria-related tumor therapy. We focus on the types and mechanisms of pathogenic bacteria in the development and promotion of cancers and emphasize how nanomaterials work. We also briefly discuss the design principles and challenges of nanomaterials for bacteria-related tumor therapy. We hope this review can provide some insights into this emerging and rapidly growing research area.

1. Introduction

Increasing evidence suggests that there are various bacteria existing in tumor tissues, and the intratumoral microbiome plays important roles in tumor genesis, progression, metastasis, immune response and chemosensitivity.^{1–3} For example, intratumoral bacteria can directly cause DNA damage, leading to increased gene mutations, causing cancer and promoting the growth of cancer cells.^{4,5} The intratumoral bacteria can also convert chemotherapy drugs into inactive forms⁶ or upregulate tumor cell autophagy,⁷ causing chemotherapy resistance. Moreover, intratumoral bacteria can affect the tumor immune microenvironment, inhibiting anti-tumor immune responses and making the tumors more aggressive.⁸

To date, the use of antibiotics remains the most effective and widely used strategy against bacterial infections, which has shown benefits for bacteria-related tumor therapy.⁹ However, systemic or oral administration of antibiotics lacks anti-bacterial specificity, causing undesired dysbiosis of gut microbiota.^{10,11} Moreover, the poor accumulation of free antibiotics in the infected tumor tissues requires a large dose and repeated administration of antibiotics to efficiently kill bac-

teria, which can enhance gut dysbiosis and develop drug resistance.¹² The rapid developments in nanotechnology and materials science provide advanced strategies against both bacterial and cancer.^{13,14} In comparison with small molecules, nanomaterials have a higher surface-to-volume ratio, providing a good tailorable surface for functionality modifications.¹⁵ Nanomaterials themselves can directly exhibit antibacterial or antitumor activities, such as silver nanoparticles,¹⁶ carbon-based nanomaterials,¹⁷ nanozymes¹⁸ and polycationic nanomaterials.¹⁹ Meanwhile, nanomaterials including mesoporous silica nanomaterials,²⁰ liposomes²¹ and polymeric nanoparticles²² can also be used as drug carriers to effectively deliver antibacterial and anti-tumor drugs to infection or tumor sites, improving the therapeutic efficacy and reducing side effects of the free drugs.^{23,24} Therefore, nanomaterials certainly hold great potential for bacteria-related tumor therapy.

In this review, the recent progress in nanomaterials for bacteria-related tumor therapy is summarized. The main types and mechanisms of pathogenic bacteria in the development and promotion of cancers are first introduced. Then, we summarize the current nanomaterials for the treatment of bacteria-related tumors including CRC, breast cancer, lung cancer and pancreatic cancer, and discuss the mechanisms. Finally, we briefly discussed the challenges in the area of development of nanomaterials for bacteria-related tumor therapy. We hope this review will provide some insights into this emerging and rapidly growing research area.

2. Pathogenic bacteria in tumors

The microbial composition in tumors shows cases distinct patterns (Fig. 1). For example, rectal tumors predominantly

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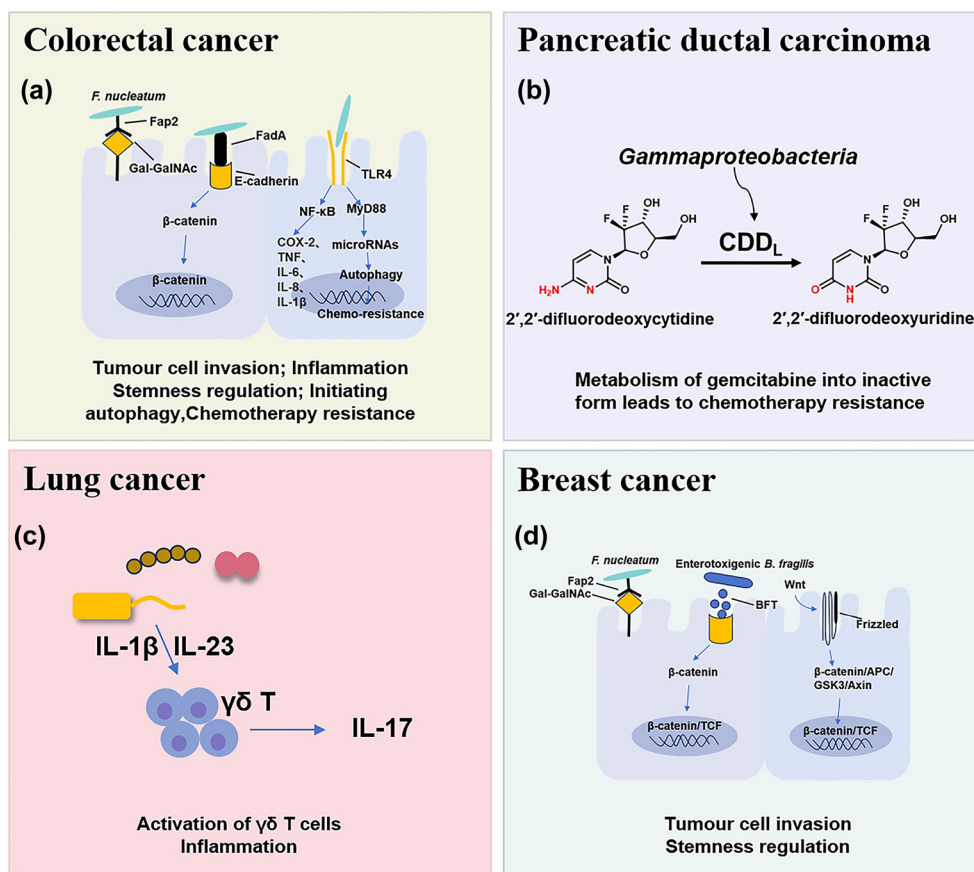


Fig. 1 The main types of pathogenic bacteria in cancers and their mechanisms in tumorigenesis. (A) Fap2 secreted by *F. nucleatum* can invade tumor cells. *F. nucleatum* activates β -catenin signaling by binding to E-cadherin using its specific FadA adhesion protein, which subsequently enhances the expression of inflammatory genes. *F. nucleatum* may activate TLR4 innate immune signaling and subsequently reduce certain microRNAs to initiate the autophagy pathway, which induces colorectal cancer chemoresistance. (B) Tumor-related bacteria metabolize gemcitabine into an inactive form, which leads to chemotherapy resistance. (C) Lung bacteria can induce inflammation by producing $\gamma\delta$ T cells and IL-17. (D) Breast-associated bacteria can secrete adhesion proteins to invade breast cells and increase the expression level of the Notch reaction genes.

harbor Firmicutes and Bacteroidetes.²⁵ Pancreatic cancer exhibits a dominance of *Proteus* akin to the normal duodenal microbiome, possibly indicating bacterial migration from the duodenum through the pancreatic duct.^{3,26} Proteobacteria and Firmicutes are present across all cancer types, while their ratio (P/F) varies among tumors, potentially holding significance in understanding individual cancer microbiomes.²⁷ The non-gastrointestinal tumors such as breast, lung, and ovarian cancers display the presence of actinomycetes such as corynebacteriaceae and micrococcaceae.¹ These microbial distinctions in different cancers provide insights into potential relationships between microbial presence and tumor development. Here, we briefly discuss the main types of pathogenic bacteria and their mechanisms in the oncogenesis of cancers including CRC, pancreatic cancer, lung cancer and breast cancer.

2.1 CRC

CRC is one of the most common causes of cancer death in the world, and chemotherapy and immunotherapy are most used clinically for the treatment of CRC.²⁸ The intestine contains

approximately 3×10^{13} bacterial cells that are symbiotic with the host, which play important roles in the occurrence and treatment of CRC and have attracted increasing interest in recent years.²⁹

***Fusobacterium nucleatum* (*F. nucleatum*).** *F. nucleatum* is a Gram-negative anaerobic bacterium, which is ubiquitous in the human oral cavity. Many studies suggest that *F. nucleatum* is enriched in human colon cancer relative to non-cancerous colon tissue, closely relating to the oncogenesis of CRC.³⁰ For example, *F. nucleatum* secretes adhesive lectin Fap2, which can mediate the colonization and invasion of colorectal cancer cells by binding to the host factor D-galactose- β (1-3)-N-acetyl-D-galactosamine (Gal-GalNAc) overexpressed in the early and metastatic stages of tumors.³¹ *F. nucleatum* can also adhere to neoplastic colonic epithelial cells by producing amyloid FadA,^{32,33} which can modulate E-cadherin and activate E-cadherin/ β -catenin signaling, leading to increased expression of transcription factors, oncogenes, Wnt genes, and inflammatory genes, as well as growth stimulation of CRC cells.³⁴

In mouse models of CRC, *F. nucleatum* can drive a pro-inflammatory tumor microenvironment and suppress anti-cancer immune responses. *F. nucleatum* can activate the NF- κ B pathway, and upregulate pro-inflammatory factors (COX-2, TNF, IL-6, IL-8, and IL-1 β),³⁵ thus promoting chronic inflammation and infiltrating intra-tumoural myeloid cells to enhance tumorigenesis.³⁶ In addition, some studies have preliminarily found that *F. nucleatum* may activate TLR4 innate immune signaling, subsequently downregulation certain microRNAs to initiate the autophagy pathway, and reduce the sensitive response of *F. nucleatum*-treated CRC cells to 5-FU and oxaliplatin chemotherapy through autophagy.^{7,37} Moreover, it was found that short-chain fatty acids (SCFAs), a derivative metabolite of *F. nucleatum*, can further promote its role in carcinogenesis by changing the function of the immune system.³⁸ SCFAs can cause numerous changes in different types of immune cells such as Treg cells, innate lymphoid cells type 3 (ILC3s), neutrophils and dendritic cells, and usually affect the immune system in a receptor-mediated way.³⁹

***Bacteroides fragilis* (*B. fragilis*).** *B. fragilis* accounts for a small fraction of total gut bacteria, estimated at about 0.1% to 0.5%, but it is considered an important symbiont and can serve as an effective pathogen.³ Enterotoxigenic *B. fragilis* (ETBF) secretes *B. fragilis* toxin (BFT) stimulates carcinogenesis by activating host colonic epithelial cell (CEC) NF- κ B and E-cadherin pathways while additionally recruiting procarcinogenic myeloid inflammation and inducing mucosal IL-17 production.⁴⁰

***Escherichia coli* (*E. coli*).** *E. coli* is a common resident bacterium in the human intestine, most of which are non-pathogenic and only a small part can cause diseases under certain conditions.² For example, *pks*⁺ *E. coli*, which produces enterotoxin-polyketide synthase, can alkylate DNA and produce DNA adducts, inducing DNA damage in colonic epithelial cells.⁴⁵ Co-colonization of ETBF and *pks*⁺ *E. coli* was found in the biopsy tissues of patients with familial adenomatous polyposis (FAP), which indicated that the two virulence factors may have synergistic effects.⁴¹

2.2 Pancreatic cancer

Pancreatic ductal carcinoma (PDAC) is a highly malignant tumor with a hidden location, low early diagnosis rate, poor efficacy of various treatments, and extremely poor prognosis.⁴² The survival rate within five years after diagnosis is less than 10%.⁴³ Many studies have found that the intratumoral microbial composition of long-term survival (LTS) patients and short-term survival (STS) patients with PDAC is varied, and regulation of the tumor microbiome can affect tumor growth and immune infiltration.⁴⁴

Gammaproteobacteria (GP). GP can express the bacterial enzyme cytidine deaminase (CDD_L) to metabolize gemcitabine, a chemotherapy drug commonly used to treat pancreatic cancer, into an inactive form, 2',2'-difluorodeoxyuridine, ultimately causing chemotherapy resistance.⁶ Additionally, depleting the microbiome with antibiotics promotes T-cell prolifer-

ation and immune activation, including tumor responsiveness to checkpoint inhibitor therapy, and prevents preinvasive and invasive pancreatic ductal adenocarcinoma.⁴⁵

2.3 Lung cancer

Lung cancer is the most common primary malignant tumor and the 1st cause of cancer deaths worldwide.⁴⁶ Because of the presence of mucosa, lung tissue attracts a large number of microorganisms to colonize, such as *Staphylococcus aureus*, *Streptococcus pneumoniae*, and *Pseudomonas aeruginosa*.⁴⁷ Due to the limitation of technology, few viable microbial cells can be isolated from healthy lungs, so the intervention mechanism of lung microflora is rarely studied.

Tsay *et al.*⁴⁸ demonstrated that *Streptococcus* and *Veillonella* enriched in the lower respiratory tract of lung cancer patients affect the proliferation, survival and tissue invasion of lung cancer cells by up-regulating RK/MAPK and PI3K/AKT pathways. Jin *et al.*⁴⁹ used the KP mouse model of lung adenocarcinoma (LUAD) driven by *Kras* activation point mutation and *p53* deletion, which clearly proved that the increase of lung bacteria and ecological imbalance would accelerate LUAD by producing γ DT cells and IL-17. The evidence is mounting that bacterial microbe-associated molecular patterns (MAMPs) and lipoteichoic acid (LTAs) will lead to the activation of the transcription factors, such as NF- κ B and AP-1, thus inducing inflammation and promoting lung cancer.⁴⁹

2.4 Breast cancer

Breast cancer is the most common type of cancer and the leading cause of cancer death in women.⁵⁰ Fu *et al.*⁵¹ reported that the tumors of spontaneous mouse mammary tumor (BT) model mice contained a large number of intracellular bacteria, similar to human breast cancer. Under the physiological conditions, these intracellular bacteria can move with cancer cells in the circulation and play a key role in tumor metastasis and colonization.⁵¹

***F. nucleatum*.** *F. nucleatum* can utilize the expressed lectin Fap2 to bind to Gal-GalNAc to colonize breast cancer tissue and reduce CD4⁺ and CD8⁺ T cell infiltration in a Fap2-dependent manner.⁵²

***B. fragilis*.** ETBF can be detected not only in the intestinal tract, but also in the breast, and can strongly induce the growth and metastatic progression of tumor cells implanted in the mammary ducts.⁵³ Parida *et al.*⁵³ discovered that short-term exposure to BFT elicits a "BFT memory" with long-term implications, functionally mediated by the β -catenin and Notch1 pathways. It can increase the expression level of several canonical and noncanonical Notch-responsive genes in breast cancer cells.

3. Nanomaterials for bacteria-related tumor therapy

As discussed above, tumor-associated bacteria can metabolize drugs into inactive forms, induce drug resistance, and regulate

the response of the immune system, thus having a significant impact on the therapeutic effect.⁵⁴ With the continuous development and perfection of nano-materials, nano-drug delivery systems including liposomes, polymeric micelles, and inorganic nanoparticles provide new possibilities for both bacterial infection and cancer therapies by improving the pharmacokinetic behavior of drugs, increasing the stability of drugs, and realizing targeted drug delivery and controlled release.⁵⁵ Liposomes are spherical nanoparticles composed of phospholipid bilayer membranes, and their size is usually between 50 and 500 nanometers. Liposome nanoparticles can encapsulate hydrophilic and hydrophobic drugs and release drugs in tumor cells, thus achieving targeted drug delivery and controlled release.⁵⁶ A polymeric micelle is a kind of nanoparticle formed by polymer self-assembly, which can encapsulate and release chemotherapy drugs and has good biocompatibility.⁵⁷ Mesoporous silica nanoparticles are considered one of the most potential nano-carriers of antibacterial drugs because of their porosity, large specific surface area and easy modification. Gold nanoparticles have also been studied for cancer

treatment because of their excellent antibacterial mechanism.⁵⁸ Nanozymes with oxidase activity can selectively catalyze the tumor microenvironment and generate a large amount of ROS, so it is widely used in the treatment of tumors.⁵⁹

3.1 Chemotherapy

Chemotherapy is one of the most commonly used and effective means to treat cancer. The basic principle is that chemical drugs enter the tumors to inhibit and kill cancer cells, thus achieving the purpose of treating cancer.⁵⁵ Previous studies have shown that tumor-related bacteria can lead to chemotherapy resistance by activating autophagy or inducing drug inactivation.⁵⁴ Therefore, the use of nano-drugs to regulate bacteria closely related to cancer progression and chemotherapy resistance can be used as an important method to improve the efficacy of chemotherapy.

Conventional liposome generally lacks self-adaptability which is important for the tumor accumulation of nanomedicines. Shi *et al.*⁶⁰ synthesized a new type of pH-responsive liposome 2-(4-((1,5-bis(octadecyloxy)-1,5-dioxopentan-2-yl) carba-

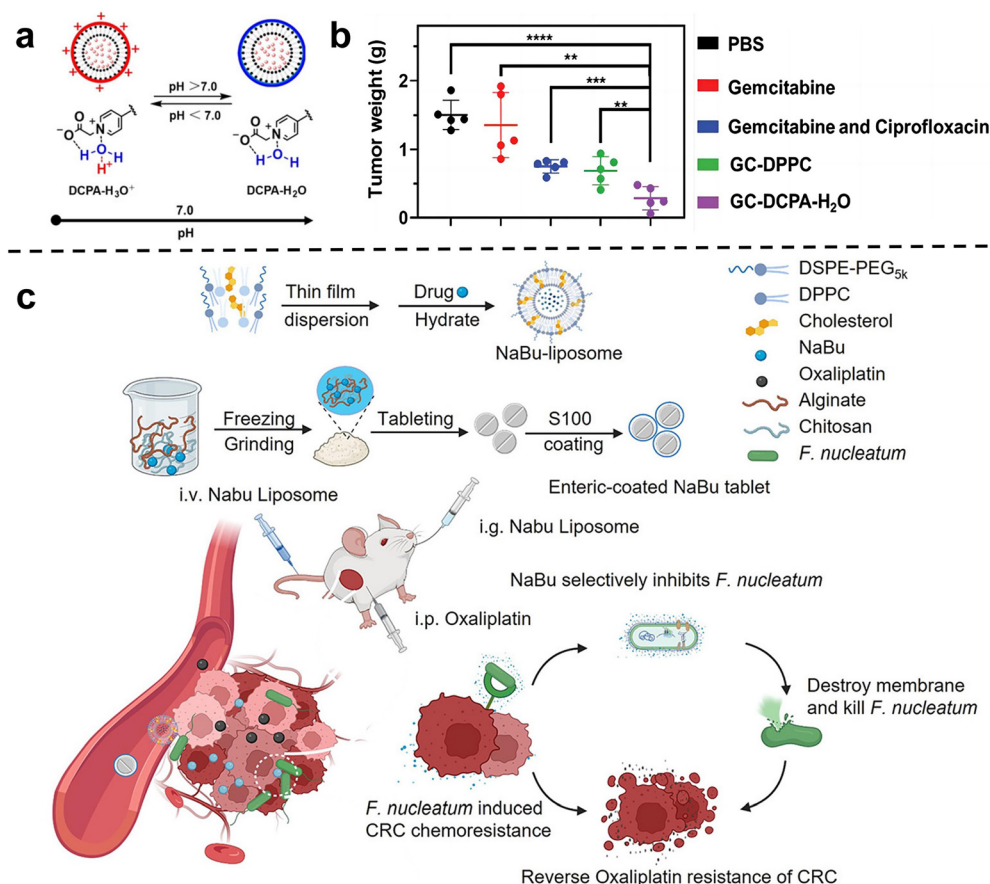


Fig. 2 (a) DCPA-H₂O has the function of pH response and is negatively charged in a normal physiological environment, and will be protonated rapidly in a tumor or infected site (pH < 6.8). Reproduced with permission from ref. 60. Copyright 2021, Wiley. (b) Tumor weight at sacrifice. Treatment either consisted of tail-vein injections of PBS, free gemcitabine or gemcitabine combined with ciprofloxacin in solution, GC-DPPC or self-targeting GC-DCPA-H₂O liposomes. Reproduced with permission from ref. 61. Copyright 2023, Wiley. (c) Schematic illustration of the NaBu-liposome to regulate *F. nucleatum*-induced oxaliplatin chemoresistance in colorectal cancer. Reproduced with permission from ref. 62. Copyright 2023, Elsevier.

moyl) pyridin-1-ium-1-yl) acetate, abbreviated DCPA in 2021, which can complex with water molecules through hydrogen bonds ($\text{DCPA-H}_2\text{O}$). Due to its high HOMO binding energy, it can be rapidly protonated in acidic environments. Therefore, DCPA liposomes circulating in the blood can quickly target the surface of negatively charged bacteria or tumor cells and release the loaded drugs. The strong self-targeting properties make $\text{DCPA-H}_2\text{O}$ liposomes a very promising nanocarrier for the treatment of bacteria-related tumors. Recently, they used DCPA liposomes to co-deliver the antibiotic ciprofloxacin and the chemotherapy drug gemcitabine to bacteria-related tumors (Fig. 2a and b).⁶¹ *In vivo* and *in vitro* results show that GC-DCPA- H_2O can accumulate in tumor tissue very fast and eliminate tumor bacteria with high efficiency, thereby preventing the inactivation of gemcitabine and significantly inhibiting tumor growth.

Chen *et al.*⁶² tried to use sodium butyrate (NaBu), the main product of intestinal microbial fermentation, to treat bacterially infected colorectal cancer (Fig. 2c). However, since sodium butyrate is a small hydrophilic molecule, it is easily

excreted in the body. In order to improve bioavailability, sodium butyrate was embedded in liposomes and injected intravenously into subcutaneous colorectal tumors, orthotopic colorectal tumors, and spontaneously formed colorectal tumors in *F. nucleatum*-infected mice. They showed that the liposome-embedded sodium butyrate can effectively inhibit the growth of *F. nucleatum* in colorectal tumors and reduce *F. nucleatum*-induced chemotherapy resistance, achieving effective tumor chemotherapy, and prolonging the overall survival of mice.

Zhang *et al.*⁶³ isolated a phage strain from human saliva that can specifically lyse *F. nucleatum*. This phage was used to design a bio-non-biological hybrid nanomaterial, that is, the phage was modified in dextran nanoparticles (DNPs) encapsulating the anti-colorectal cancer drug irinotecan (IRT)(Fig. 3a). The results showed that in mice with orthotopic colorectal tumors or spontaneously formed colorectal tumors, oral or intravenous administration could inhibit the proliferation of *F. nucleatum* and significantly improve the efficiency of CRC chemotherapy treatment. Chen *et al.*⁶⁴ designed nitroreduc-

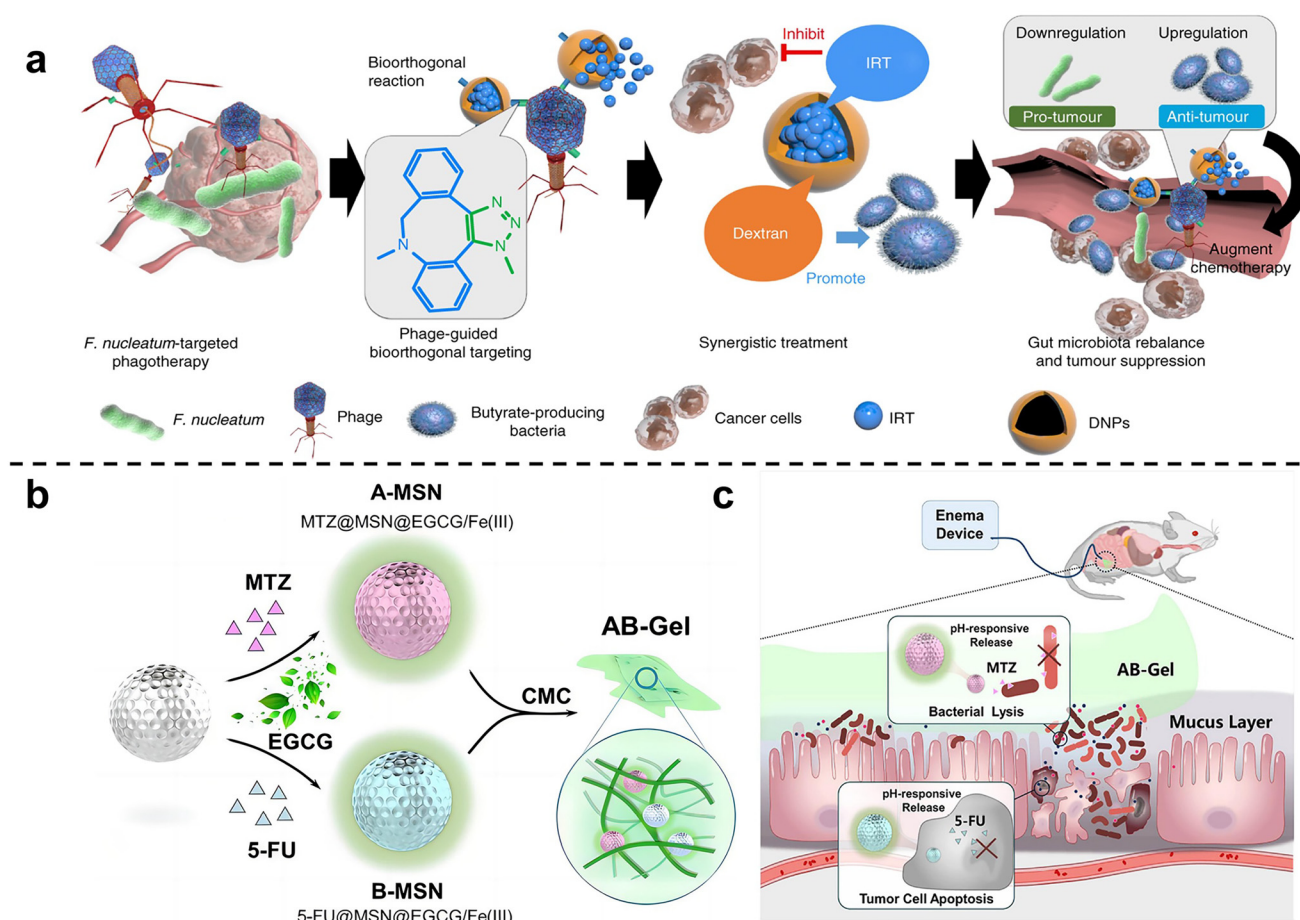


Fig. 3 (a) Illustration of the phage-guided biotic-abiotic hybrid nanosystem and its mechanism. Reproduced with permission from ref. 63. Copyright 2019, Springer Nature. (b) EGCG/Fe(III) network encapsulated MSN loaded with MTZ and 5-FU, mixing into CMC, constructed as the AB-Gel. (c) AB-Gel was directly perfused into the colon to the tumor site, which not only realized the rapid and accurate release of drugs but also reduced the side effects, thus increasing the chemotherapy effect. Reproduced with permission from ref. 66. Copyright 2021, Elsevier.

tase-mediated supramolecular self-assembly. After enzyme catalysis, the precursor undergoes a self-elimination reaction to form a gel, which self-assembles into supramolecular nanofibers. The supramolecular nanofibers then target and capture *F. nucleatum* to inhibit intratumoral bacteria and enhance subsequent chemotherapy.

Chen *et al.*⁶⁵ used dendritic mesoporous silicas (DMSNs) to load silver nanoparticles (Ag NPs), and combined with chemotherapy to treat *F. nucleatum*-infected colorectal cancer. DMSN was firstly modified with Ag⁺ through sulfhydryl bonds, and then Ag NPs were grown *in situ* in the mesopores of SH-DMSNs through the sodium borohydride rapid reduction method. Finally, the chemotherapy drug epirubicin (EPI) was loaded to form antibacterial and antitumor Ag@DMSNs-EPI NPs (ADEN for short). The designed antibacterial-antitumor nanomedicines can effectively treat colorectal cancer by clearing *F. nucleatum* within tumors and reshaping the tumor micro-environment. Chen *et al.*⁶⁶ used mesoporous silica nanoparticles to prepare a drug delivery gel (Fig. 3b and c). The antibiotic metronidazole and the chemotherapy drug 5-fluorouracil (5-FU) were loaded in MSN and then blended with carboxymethylcellulose (CMC) to obtain the anti-CRC gel

AB-Gel. *In vitro* experiments showed that antibiotic-loaded A-MSN has strong antibacterial activity, and 5-FU-loaded B-MSN has strong anti-cancer ability. *In vivo* studies further confirmed that combined microbiota modulation can significantly improve the efficacy of colorectal cancer treatment.

In order to inhibit chemotherapy resistance caused by commensal bacteria in lung tumors, Han *et al.*⁶⁷ constructed an inhalable gallium-based metal nanoparticle coated with capsular polysaccharidecp (CP), defined as GaTa-cpNPs (Fig. 4a). Tannin (Ta) is a natural polyphenol with many phenolic hydroxyl groups. GaTa NPs were successfully synthesized by self-assembly through a coordination reaction between tannin and gallium metal ions. The *Streptococcus pneumoniae* capsular polysaccharide coating can enable GaTa-cp NPs to effectively adhere to lung tissue and masquerade as normal tissue components to reduce immune clearance in the body. In addition, they also included the chemotherapy drug epiposide to verify the drug delivery and tumor treatment capabilities of GaTa-cp NPs. In the acidic environment of the tumor, Ga³⁺ and the drug Eto are released. In multiple *Escherichia coli* Nissle 1917 (*EcN*)-infected mouse lung cancer models, local lung microbiota can be effectively depleted to inhibit bacterial-

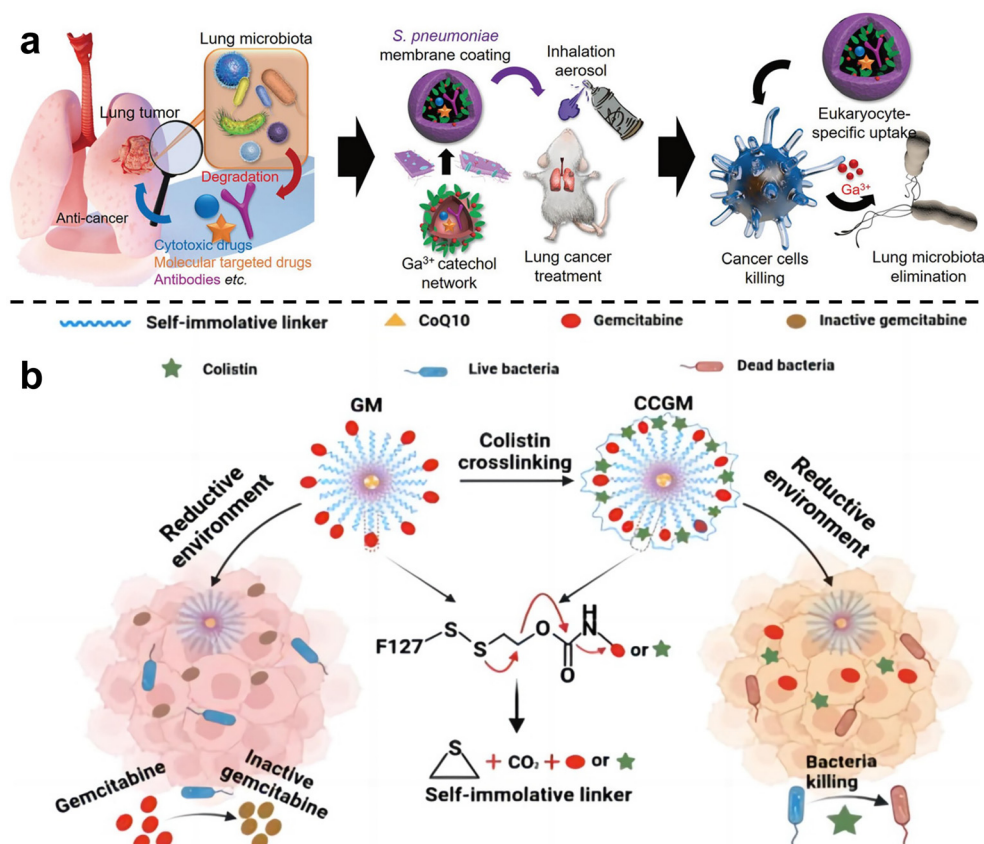


Fig. 4 (a) Scheme for the synthesis of the inhalable capsular polysaccharide (CP)-camouflaged gallium-based metal-organic network (MON). It can effectively eliminate various microorganisms in the lung tumor model, thus reducing the drug resistance induced by bacteria. Reproduced with permission from ref. 67. Copyright 2023, Wiley. (b) Schematic illustration of CCGM eliminating bacterial-induced cancer drug resistance. Reproduced with permission from ref. 68. Copyright 2022, American Chemical Society.

induced chemotherapy resistance, thereby enhancing lung cancer treatment. Recently, Qiu *et al.*⁶⁸ used micelles to jointly deliver antibiotics and anticancer drugs, called colistin cross-linked gemcitabine micelles (CCGM) for *EcN*-infected breast cancer therapy (Fig. 4b). Once CCGM is delivered into the tumor microenvironment, it triggers a glutathione response, and the self-immolating linker is disconnected, resulting in the release of colistin and gemcitabine, thereby inhibiting the growth of intratumoral bacteria and eliminating bacterial-induced tumor resistance to enhance tumor chemotherapy.

Recently, our group has developed a series of polymeric nanosystems to overcome *F. nucleatum* caused issues including chemotherapy resistance and immunosuppression in CRC therapy. Based on our decadal works on supramolecular assembly, we rationally designed a size-switchable supramole-

cular nanomedicine (PG-Pt-LA/CB[7]) to effectively reverse *F. nucleatum*-induced chemotherapy resistance of CRC to oxaliplatin (Fig. 5a–f).⁶⁹ This was the first report on using a natural fatty acid lauric acid (LA) as an antibacterial agent to selectively kill *F. nucleatum*, which overcomes the drawbacks of broad-spectrum antibiotics. In detail, LA and platinum(IV) oxaliplatin prodrug (OxPt-COOH) were coupled with hyperbranched poly(glycidol) (PG) and self-assembled to form PG-Pt-LA nanoparticles, and then cucurbit[7]uril (CB[7]) was added to induce supramolecular assembly. The PG-Pt-LA/CB[7] (~210 nm) is stable in blood with a long half-life, while disassembles in response to the CRC microenvironment and turns into small PG-Pt-LA NPs (~10 nm) to penetrate the tumor deeply. *In vivo* results confirmed that LA has a significant effect in eliminating *F. nucleatum* present in solid CRC tumors

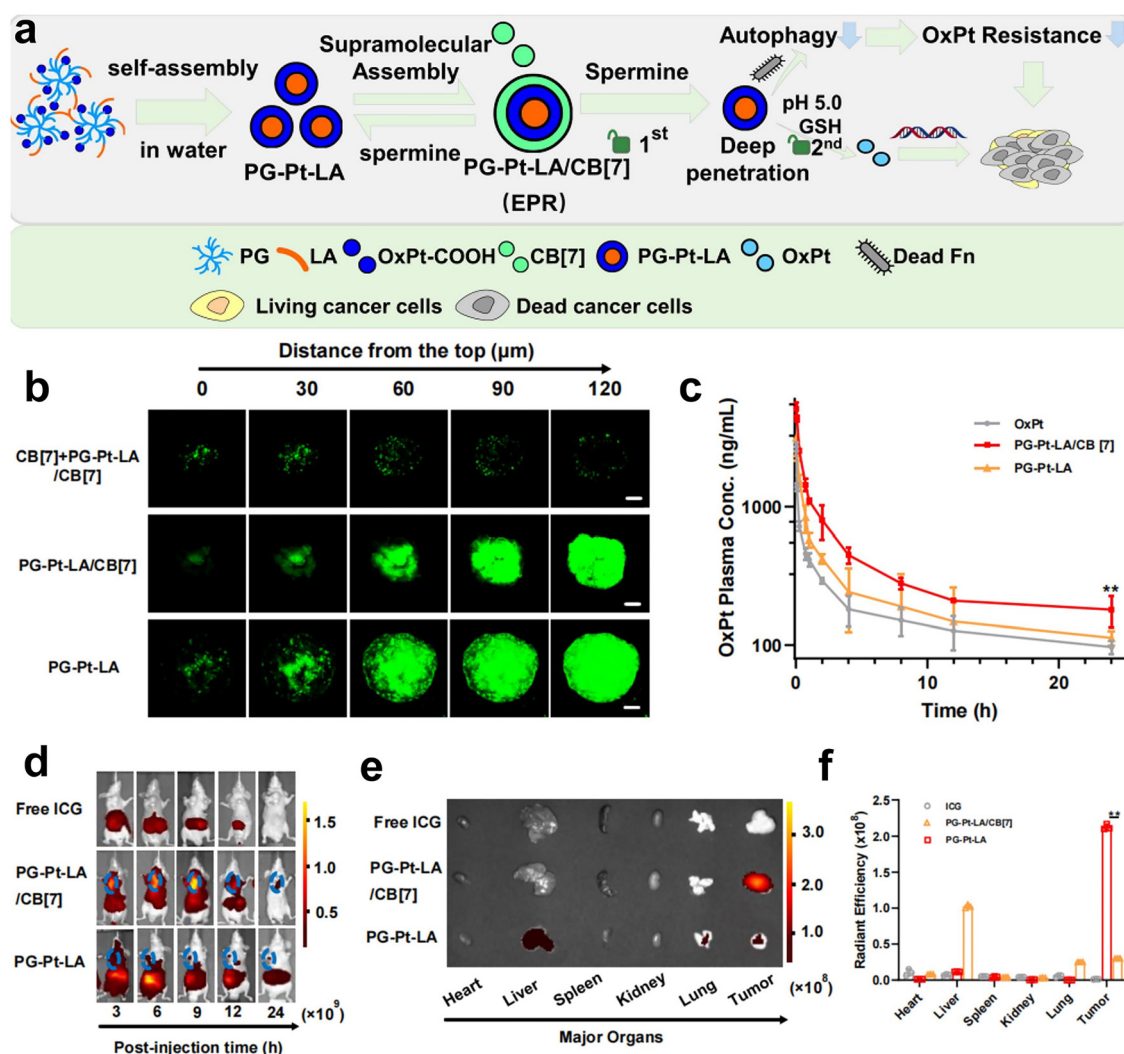


Fig. 5 (a) The assembly diagram of supramolecular nanomedicine. (b) CLSM observation on the HT29 3D multicellular spheroids subjected to prior incubation of CB[7] with the aim of depletion of spermine, followed by different treatments. (c) Pharmacokinetics of OxPt, PG-Pt-LA/CB[7] and PG-Pt-LA upon intravenous dosage. (d) Change of biodistributions of free ICG, ICG-labeled PG-Pt-LA/CB[7] and ICG-labeled PG-Pt-LA with time. (e) Fluorescence imaging and (f) quantitative analysis of the dissected organs and tumors after different treatments. Reproduced with permission from ref. 69. Copyright 2022, Elsevier.

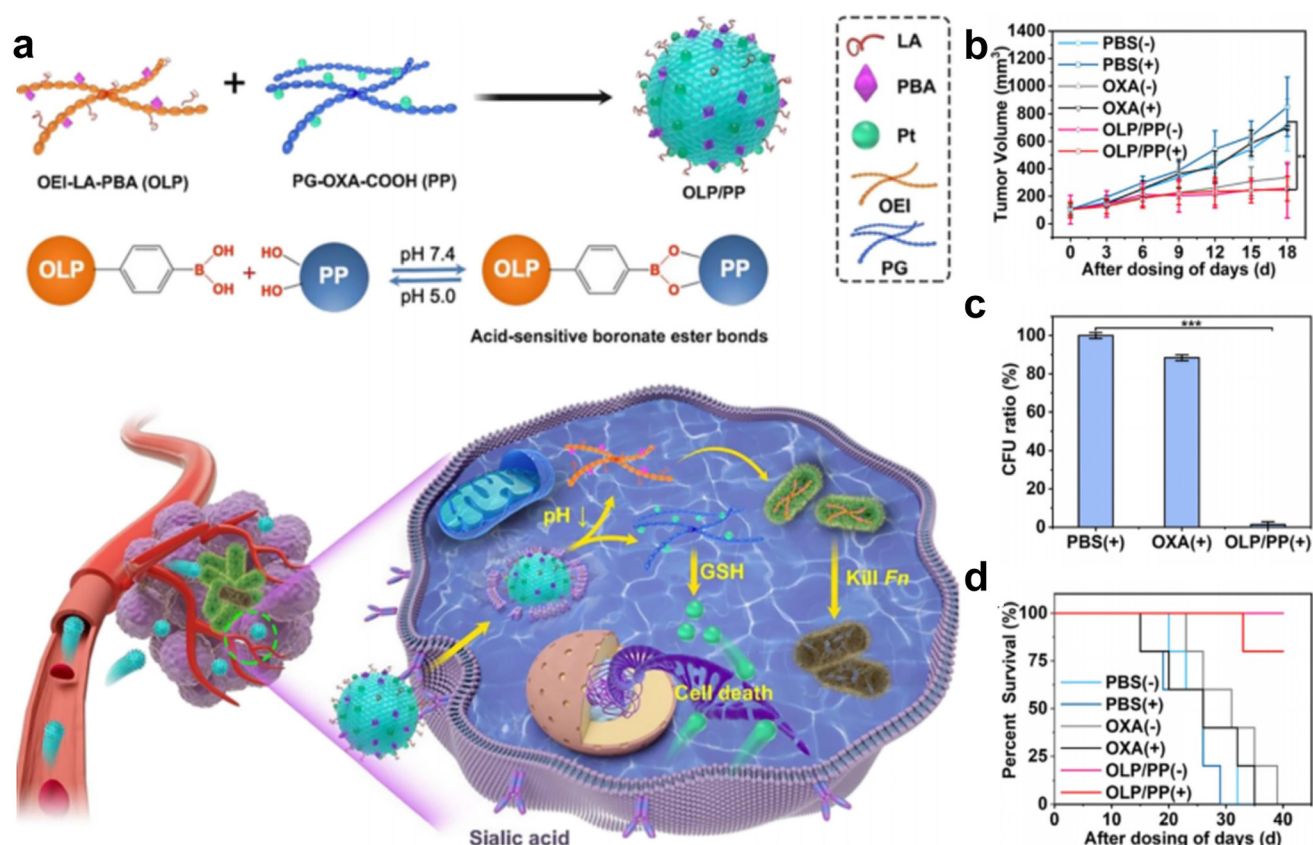


Fig. 6 (a) Schematic illustration of OLP/PP for the treatment of CRC tumors colonized with *F. nucleatum*. (b) Tumor volume, (c) quantification of *F. nucleatum* in tumors and (d) survival fractions of the *F. nucleatum*-infected HT29 tumor-bearing mice under diverse treatments. Reproduced with permission from ref. 70. Copyright 2023, American Chemical Society.

and can reduce chemotherapy resistance, thereby increasing the therapeutic effect of chemotherapy drugs on tumors.

Intracellular *F. nucleatum* was hard to eliminate using free antibiotics due to its inability to traverse cell membranes. In a recent study, Li *et al.*⁷⁰ fabricated an acidity-responsive nanomaterial that can deliver LA and OxpT-COOH into tumor cells to kill the intratumoral bacteria and treat cancer (Fig. 6a–d). OLP and PP can self-assemble in water to form stable nanoassembly, in which the PBA can specifically bind to tumor cells and facilitate receptor-mediated endocytosis. After cellular uptake, the OLP/PP nanoassembly can disassemble in response to the tumor's acidic microenvironment (pH 5.0) to trigger the release of drugs. The results showed that the constructed OLP/PP nanoassembly can effectively eliminate both extracellular and intracellular *F. nucleatum* and inhibit tumor growth.

3.2 Immunotherapy

Cancer immunotherapy aims to activate the immune system of patients and kill cancer cells by their own immune function.⁵⁵ This new treatment method is expected to achieve the goal of long-term survival with a tumor or a complete cure of tumor and is becoming one of the important development directions

in the field of cancer treatment. Recent studies have shown that the enrichment of *F. nucleatum* in colorectal cancer can drive the formation of tumor microenvironment in immunosuppression.⁷¹ As such, eliminating tumor-associated bacteria may be an effective strategy to improve the effect of tumor immunotherapy.

Chen *et al.*⁷² designed a liposome-based nanomedicine that mimics *F. nucleatum* to reverse immunotherapy resistance for the infected CRC treatment (Fig. 7a and b). The antibiotic colistin is loaded into liposomes, and then fused with the cytoplasmic membrane FM of *F. nucleatum* to obtain colistin-loaded FM fusion liposomes (colistin-LipoFM). Since the surface of the nuclear-cytoplasmic membrane (FM) contains Fap-2, it can specifically target Gal-GalNAc that was over-expressed on the surface of colorectal tumor cells, allowing colistin to effectively accumulate at the site of the tumor infection without affecting intestinal microorganisms and selectively kill tumor-colonizing *F. nucleatum*. Recently, Huang *et al.*⁷³ also designed an antibiotic silver-tinidazole complex encapsulated liposomes (LipoAgTNZ) (Fig. 7c and d). This LipoAgTNZ can accurately target anaerobic bacteria in infected tumors, release silver particles and tinidazole in the acidic tumor microenvironment, and kill *F. nucleatum* in the tumor.

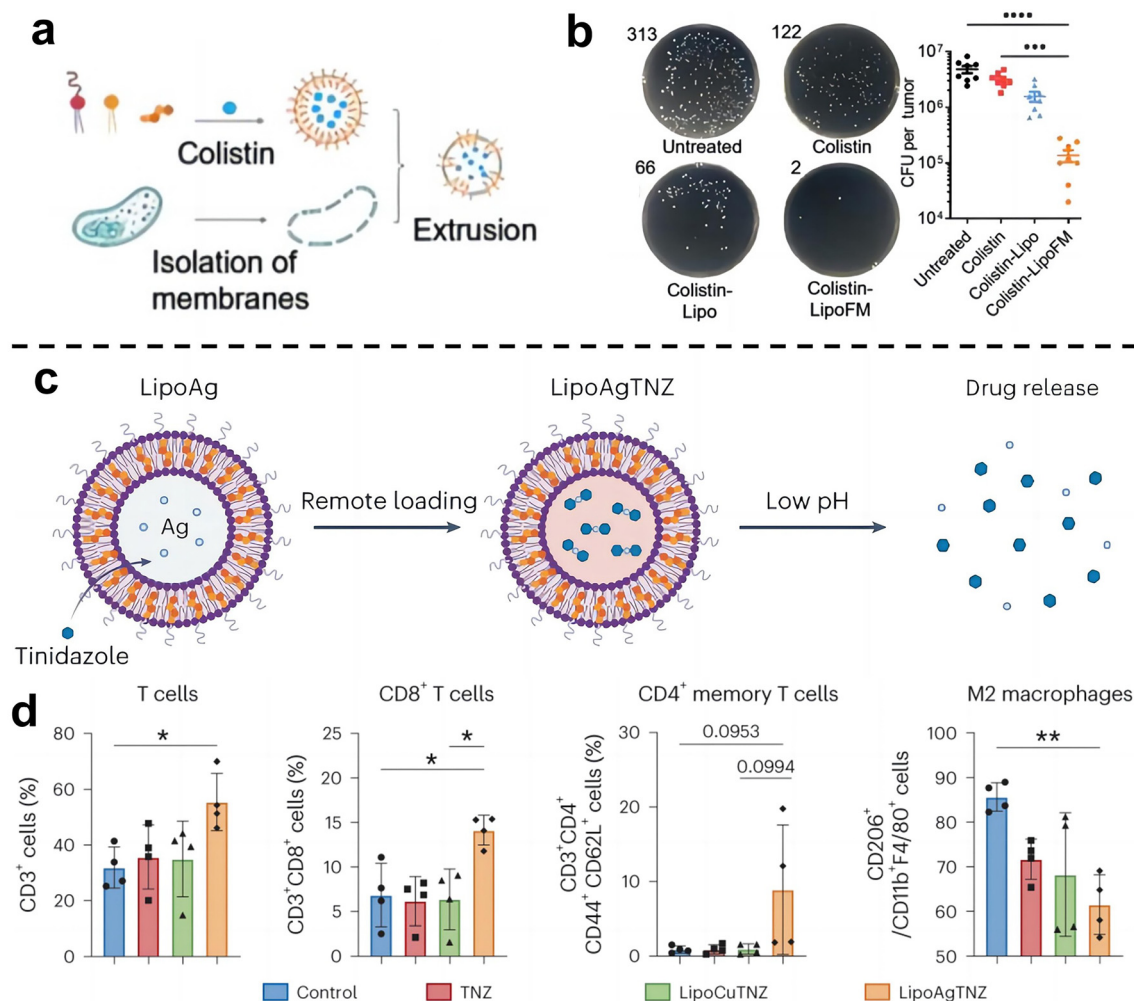


Fig. 7 (a) Synthetic legend of Colistin-LipoFM. (b) Colony plate images (left) and the colony-forming units (CFUs) (right) showing the *F. nucleatum* abundance in homogenized tumors of mice treated with colistin-LipoFM, colistin-Lipo and free colistin. Reproduced with permission from ref. 72. Copyright 2023, Wiley. (c) Illustration of remote loading by a silver nitrate gradient and drug release in response to low pH. (d) Analyses of the immune cell population in the tumor microenvironment after treatment. Reproduced with permission from ref. 73. Copyright 2023, Springer Nature.

Interestingly, they found that eliminating bacteria in colorectal cancer tumors can expose microbial epitopes, thus activating immune responses to enhance tumor treatment efficacy.

Zhang *et al.*⁷⁴ used hyaluronic acid (HA)-modified photoluminescent organosilica nanodots (OSiNDs) to deliver gemcitabine (Gem) and the antibiotic ciprofloxacin (Cip) into bacterially infected mouse colon tumors (Fig. 8a). After treatment with Gem/Cip@SiPNG@HA, the killed bacteria promoted the maturation of antigen-presenting dendritic cells (DCs), while Gem reduced the levels of MDSCs, thereby triggering T cells enhanced cancer immunotherapy. Song *et al.*⁷⁵ designed a bacteria-targeting nanomaterial to target bacteria living in tumor sites. Bacterial lipophosphocholic acid antibodies extracted from bacterial walls were modified on mesoporous silica nanoparticles, which could precisely target LTA on the bacterial surface within tumors and deliver anti-tumor drugs. Ma *et al.*⁷⁶ also designed an inhalable, dual-loaded mesoporous silica nanoparticle (MSN@DOX-AMP) to treat lung cancer

(Fig. 8b). Efficient loading of anti-tumor antibiotic doxorubicin (DOX) and antimicrobial peptide HHC36 using physical adsorption and thiol-ene click chemistry. In mice with lung cancer with *Staphylococcus aureus* as a commensal bacterium, it was demonstrated that MSN@DOX-AMP could strongly destroy the tumor/bacteria symbiosis system of lung cancer, achieving better therapeutic effects that compared with treating tumors or bacteria alone.

3.3 Synergetic therapy

After carefully designed nano-system with two or more treatments can be skillfully combined to act on tumors together, thus producing remarkable synergistic effects. This combination therapy strategy can not only significantly improve the therapeutic effect, make the tumor inhibition rate reach more than 90% within 20 days, but also greatly reduce the toxic and side effects so that patients can bear fewer side effects during the treatment.⁷⁷

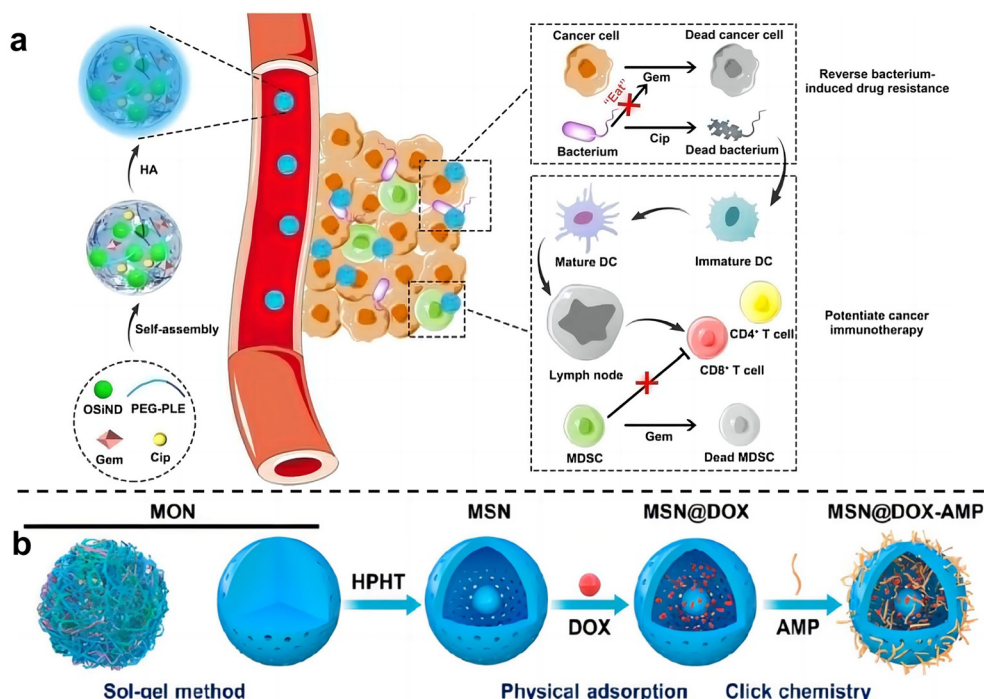


Fig. 8 (a) Schematic diagram illustrates the preparation and mechanism of Gem/Cip@SiPNG, which can reverse bacterial-induced chemotherapy resistance and potentiate cancer immunotherapy. Reproduced with permission from ref. 74. Copyright 2021, Wiley. (b) Preparation of the dual-drug-delivery mesoporous silica nanoparticles (MSNs). Reproduced with permission from ref. 76. Copyright 2023, American Chemical Society.

Qu *et al.*⁷⁸ developed a gold nanoparticle-based antibacterial nanoplateform by using bovine serum albumin (BSA) as a carrier to improve biocompatibility and stability (Au@BSA-CuPpIX) (Fig. 9a). Gold nanoparticles (Au NPs) are captured through biomimetic mineralization and then modi-

fied with an antibacterial metallopephyrin (CuPpIX) sonosensitizer to synthesize nanoparticles with ultra-small diameters. Reactive oxygen species (ROS) are generated under the action of ultrasound, and *in vivo* experiments showed that Au@BSA-CuPpIX can effectively kill bacteria in tumors and

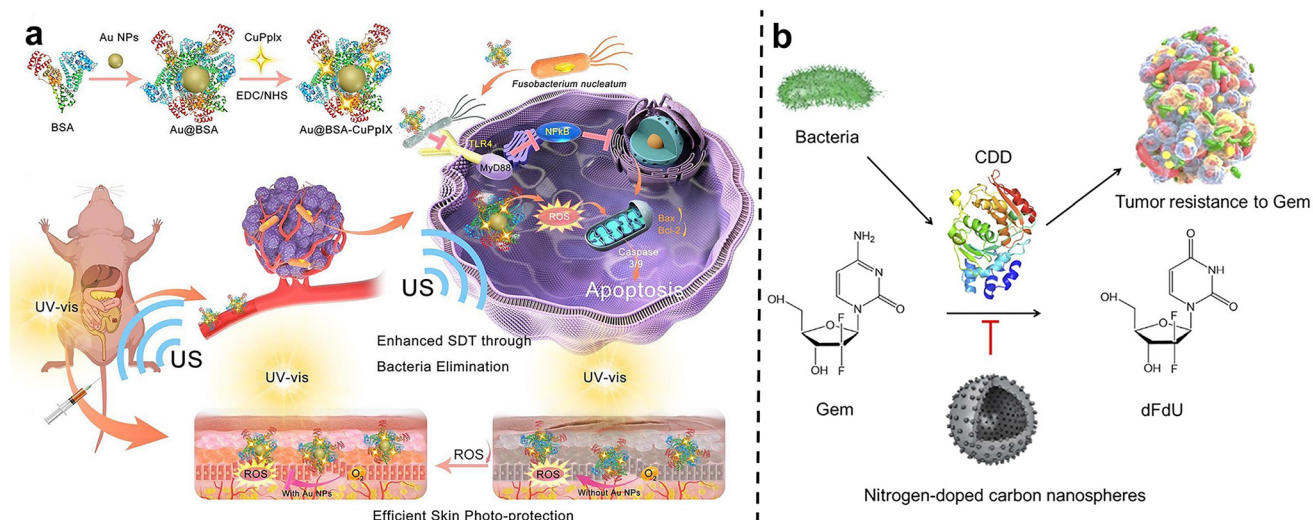


Fig. 9 (a) The scheme describes the synthesis and mechanism of Au@BSA-CuPpIX. Au@BSA-CuPpIX, which can effectively eliminate *F. nucleatum* to reduce the level of apoptosis inhibitor protein in cancer cells, and enhance the tumor treatment effect of SDT. Au NPs can reduce the phototoxicity of CuPpIX in the skin, thus reducing skin damage. Reproduced with permission from ref. 78. Copyright 2023, American Chemical Society. (b) Schematic of N-CS-induced inhibitory effects against CDD expressed by bacteria to metabolize Gem into dFdU. Reproduced with permission from ref. 80. Copyright 2022, Elsevier.

improve the therapeutic effect of colorectal cancer. In addition, the wrapped gold nanoparticles reduce the phototoxicity of metalloporphyrins and avoid severe inflammation and metal damage.

Wang *et al.*⁷⁹ pioneered the construction of a highly biomimetic artificial protein-supported copper single-atom nanozyme (BSA-Cu SAN). Due to the multivalent nature of copper, BSA-Cu SAN can produce catalytic H_2O_2 to generate ROS, consume glutathione, kill intratumoral *F. nucleatum*, and effectively restore the autophagy level of tumor cells elevated by *F. nucleatum*. At the same time, bovine serum albumin is loaded on the nanozyme, which gives the nanozyme high hydrophilicity and biocompatibility, solving the problem of

high metal ion toxicity, and achieving satisfactory tumor treatment effects. Xi *et al.*⁸⁰ used N-doped carbon spheres (N-CSs) as a colorectal cancer treatment system and found that they have the dual functions of nanozymes and nanoinhibitors, capable of reversing bacterial-induced tumor drug resistance and improving immunity in pathogen-symbiotic tumors (Fig. 9b). First, N-CSs, as a peroxidase-like nanozyme, can catalyze H_2O_2 in the tumor microenvironment to produce highly cytotoxic hydroxyl radicals ($\cdot OH$) to kill cancer cells. Secondly, N-CSs, as nanoinhibitors, inhibited bacteria at tumor sites, successfully reversing bacterial-induced gemcitabine resistance and restoring tumor susceptibility to gemcitabine. It provides an effective way to overcome tumor resistance induced by

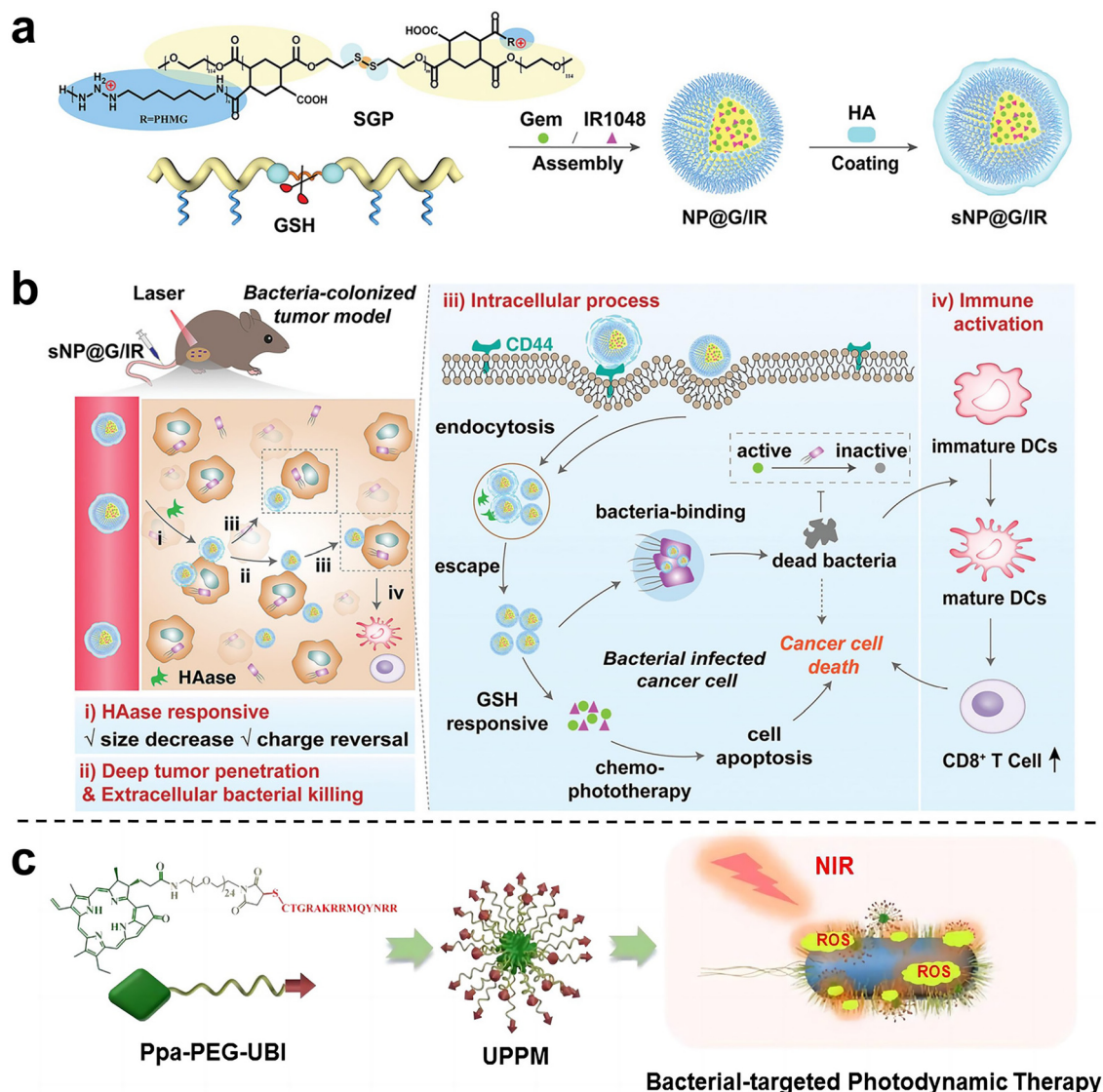


Fig. 10 (a) Preparation process of sNP@G/IR. (b) Schematic diagram illustrating that sNP@G/IR can kill bacteria and activate immunity. After the HAase response, the particle size of sNP@G/IR decreases and the charge is reversed, which can achieve deep penetration of the tumor and bacterial killing. After rapid endocytosis, NP@G/IR is beneficial for the clearance of intracellular bacteria and immune activation. Reproduced with permission from ref. 81. Copyright 2023, Wiley. (c) Schematic illustration of the preparation process of UPPM micelles. Reproduced with permission from ref. 82. Copyright 2023, American Chemical Society.

intratumoral bacteria and to synergize chemotherapy with catalytic therapy.

Wang *et al.*⁸¹ proposed and synthesized a dual-cascade response nanoparticle (sNP@G/IR) with a shell-core structure that can eliminate intra-tumor bacteria and achieve effective drug delivery for *EcN*-infected pancreatic cancer therapy (Fig. 10a and b). Firstly, an amphiphilic GSH-responsive polymer (SGP) with good antibacterial activity and biocompatibility was synthesized, which contained the chemotherapy drug Gem and the photothermal agent IR1048 and was self-assembled to form NP@G/IR. It was further coated with hya-

luronic acid (HA) with a tumor-targeting effect to obtain dual cascade reaction nanoparticles (sNP@G/IR). This inhibits the growth of bacterially colonized tumors by eliminating tumor-resident intracellular bacteria and killing cancer cells with precise delivery. Liu *et al.*⁸² synthesized an amphiphilic polymer by coupling hydrophilic polyethylene glycol (PEG) with the poorly water-soluble photosensitizer pyropheophorbide-a (Ppa) and the antimicrobial peptide ubiquicidin (UBI) (Fig. 10c), and then self-assembled into polymer micelles (UPPM) in the aqueous solution. UBI peptides endow UPPM with excellent bacterial targeting capabilities. Under light,

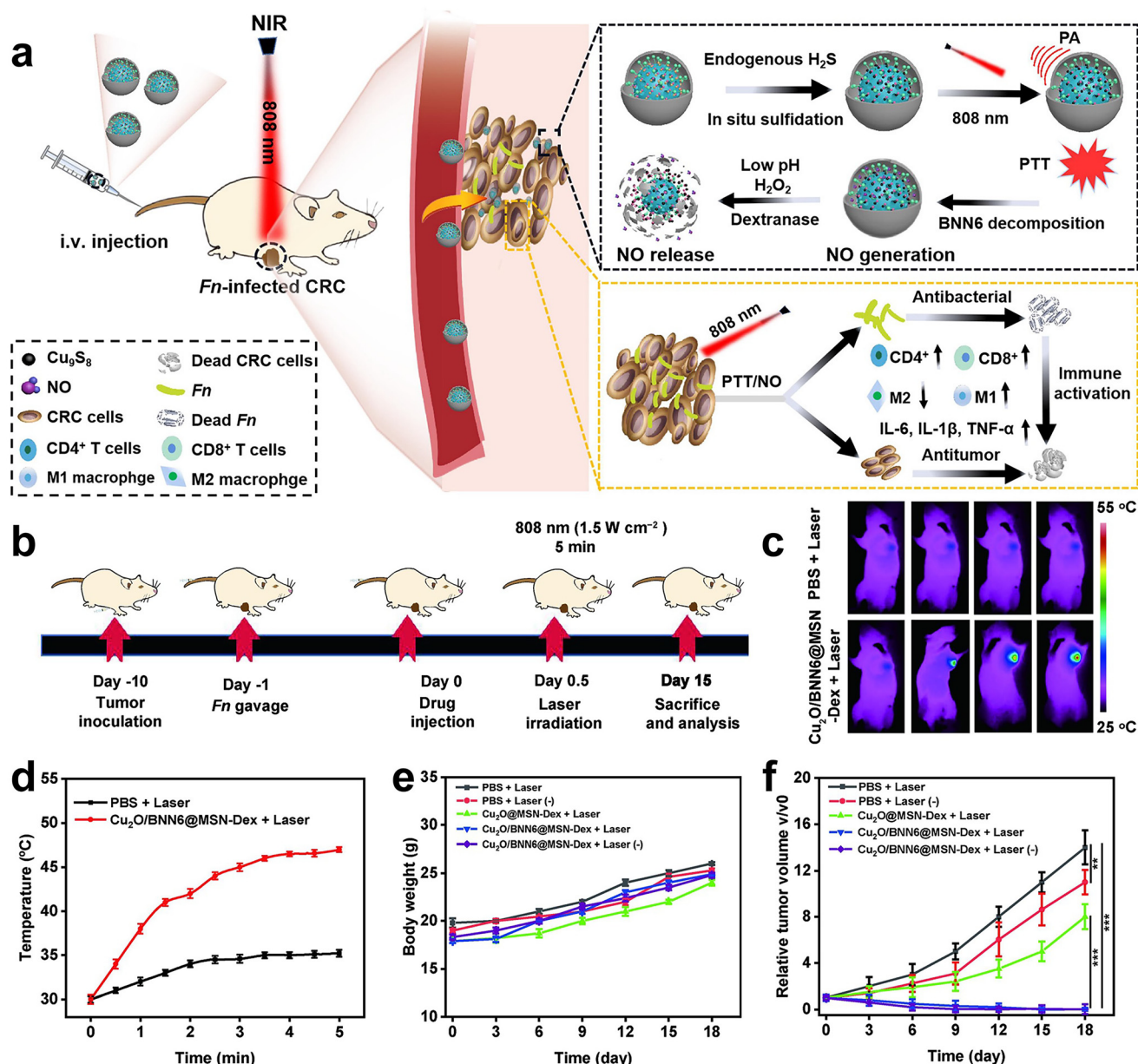


Fig. 11 (a) Schematic diagram of the photothermal and gas combined therapy with $\text{Cu}_2\text{O}/\text{BNN6}@\text{MSN-Dex}$. (b) Scheme showing the experimental schedule. (c) *In vivo* fluorescence imaging and (d) temperature versus time curves of *F. nucleatum*-infected HT29 tumor mice treated with PBS or $\text{Cu}_2\text{O}/\text{BNN6}@\text{MSN-Dex}$ under 808 nm laser irradiation. (e) Body weight and (f) tumor growth curves of *F. nucleatum*-infected mice bearing HT29 tumors after different treatments. Reproduced with permission from ref. 83. Copyright 2023, Elsevier.

UPPM can kill bacteria by inducing bacterial membrane damage, thereby effectively inhibiting Gem metabolism. In subcutaneous PDAC animal models injected with bacteria, UPPM can eradicate intratumoral bacteria *via* photodynamic therapy without affecting the intestinal microbiota.

Immunotherapy is promising for the treatment of CRC, however, the therapeutic efficiency is also affected by *F. nucleatum*. Recently, Xin *et al.*⁸³ developed an *in situ* activated anti-tumor and antibacterial nanoplatfrom (Cu₂O/BNN6@MSN-Dex) that allows combined photothermal and gas therapy to enhance *F. nucleatum*-infected colorectal cancer immunotherapy (Fig. 11a–f). Boronized mesoporous silica nanoparticles MSN-PBA were first synthesized, and then cuprous oxide (Cu₂O) and *N,N'*-di-sec-butyl-*N,N'*-dinitroso-1,4-phenylene diamine (BNN6) were encapsulated in MSN-PBA. Finally, polyhydroxydextran (Dex) was coated through dynamic cross-linking boronic acid bonds to obtain Cu₂O/BNN6@MSN-Dex. Cu₂O/BNN6@MSN-Dex can accumulate into tumor tissues and target tumor cells through the PBA ligand. After penetrating into the tumor, the loaded Cu₂O will be sulfated by endogenous H₂S overexpressed in tumors, resulting in copper sulfide with significant photoacoustic (PA) and photothermal (PT) properties. Under 808 nm laser irradiation, non-invasive PTT was performed with *in situ* local hyperthermia and induced BNN6 to produce NO. Dextran makes Cu₂O/BNN6@MSN-Dex have good biocompatibility, responding in the tumor microenvironment, and is decomposed by glucanase, which is beneficial to the release of NO. *In vivo* and *in vitro* experiments demonstrated that combined photothermal and NO gas treatment can specifically eliminate *F. nucleatum* within tumors and significantly inhibit tumor growth, providing an effective strategy to enhance CRC treatment.

4. Conclusions and perspective

In this review, we summarized the key research progress in the field of nanomaterials for bacteria-related tumor therapy. Current studies mostly focus on targeting the tumor-commensal bacteria model and using nanomaterials to eradicate cancer-associated microorganisms to enhance tumor treatment effects. It is obvious that this strategy has shown prosperous significance. One key challenge for the construction of such nanomaterials is how to effectively integrate antibacterial and antitumor functions into a single system. More importantly, it is optimum for antibacterial agents to selectively exert antibacterial effects on intratumoral bacteria to prevent microbiome imbalance caused by unselective sterilization.

Currently, there is relatively little research on the mechanism of the progression of bacteria colonization in tumor tissue. This should be a long process with many pathological changes. However, current animal models are too artificial. For example, when building a bacterial related tumor model, researchers usually use intratumoral injection or intravenous injection of a single bacterial species. Although this method is

simple and feasible, it cannot truly simulate the diversity of the intratumoral microbiota and the complex bacteria-tumor interaction. Therefore, more attention needs to be paid to building a more authentic bacterial related tumor model in order to obtain more accurate and in-depth research results. Moreover, the effectiveness of such a therapeutic strategy should be further evaluated using the above models. Overall, this is an emerging research area which is worthy of attention. We believe more innovative methods and strategies will emerge in the near future, which will be helpful in improving tumor treatment effects and improving patients' quality of life.

Author contributions

Fuping Zhang: writing–original draft, figures designing and investigation. Shuo Yang: investigation. Shuyu Wang, Feihe Ma and Hui Gao: writing–reviewing and editing, supervision, and funding acquisition.

Conflicts of interest

There are no conflicts to declare.

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References

- 1 D. Nejman, I. Livyatan, G. Fuks, N. Gavert, Y. Zwing, L. T. Geller, A. Rotter-Maskowitz, R. Weiser, G. Mallel, E. Gigi, A. Meltser, G. M. Douglas, I. Kamer, V. Gopalakrishnan, T. Dadosh, S. Levin-Zaidman, S. Avnet, T. Atlan, Z. A. Cooper, R. Arora, A. P. Cogdill, Md A. W. Khan, G. Ologun, Y. Bussi, A. Weinberger, M. Lotan-Pompan, O. Golani, G. Perry, M. Rokah, K. Bahar-Shany, E. A. Rozeman, C. U. Blank, A. Ronai, R. Shaoul, A. Amit, T. Dorfman, R. Kremer, Z. R. Cohen, S. Harnof, T. Siegal, E. Yehuda-Shnaidman, E. N. Gal-Yam, H. Shapira, N. Baldini, M. G. I. Langille, A. Ben-Nun, B. Kaufman, A. Nissan, T. Golan, M. Dadiani, K. Levanon, J. Bar, S. Yust-Katz, I. Barshack, D. S. Peeper, D. J. Raz, E. Segal, J. A. Wargo, J. Sandbank, N. Shental and R. Straussman, *Science*, 2020, **368**, 973–980.
- 2 G. El Tekle and W. S. Garrett, *Nat. Rev. Cancer*, 2023, **23**, 600–618.

- 3 R. J. Knippel, J. L. Drewes and C. L. Sears, *Cancer Discovery*, 2021, **11**, 2378–2395.
- 4 P. J. Dziubańska-Kusibab, H. Berger, F. Battistini, B. A. M. Bouwman, A. Iftekhhar, R. Katainen, T. Cajuso, N. Crosetto, M. Orozco, L. A. Aaltonen and T. F. Meyer, *Nat. Med.*, 2020, **26**, 1063–1069.
- 5 C. Pleguezuelos-Manzano, J. Puschhof, R. Bonnet, P. Quirke, M. Meyerson, E. Cuppen, R. van Boxtel and H. Clevers, *Nature*, 2020, **580**, 269–273.
- 6 L. T. Geller, M. Barzily-Rokni, T. Danino, O. H. Jonas, N. Shental, D. Nejman, N. Gavert, Y. Zwang and Z. A. Cooper, *Science*, 2017, **357**, 1156–1160.
- 7 T. Yu, F. Guo, Y. Yu, T. Sun, D. Ma, J. Han, Y. Qian, I. Kryczek, D. Sun, N. Nagarsheth, Y. Chen, H. Chen, J. Hong, W. Zou and J.-Y. Fang, *Cell*, 2017, **170**, 548–563.
- 8 C. Gur, Y. Ibrahim, B. Isaacson, R. Yamin, J. Abed, M. Gamliel, J. Enk, Y. Bar-On, N. Stanitsky-Kaynan, S. Copenhagen-Glazer, N. Shussman, G. Almogy, A. Cuapio, E. Hofer, D. Mevorach, A. Tabib, R. Ortenberg, G. Markel, K. Miklič, S. Jonjic, C. A. Brennan, W. S. Garrett, G. Bachrach and O. Mandelboim, *Immunity*, 2015, **42**, 344–355.
- 9 F. Baquero and B. R. Levin, *Nat. Rev. Microbiol.*, 2020, **19**, 123–132.
- 10 F. Peyrusson, H. Varet, T. K. Nguyen, R. Legendre, O. Sismeiro, J.-Y. Coppée, C. Wolz, T. Tenson and F. Van Bambeke, *Nat. Commun.*, 2020, **11**, 2200.
- 11 L. Maier, C. V. Goemans, J. Wirbel, M. Kuhn, C. Eberl, M. Pruteanu, P. Müller, S. Garcia-Santamarina, E. Cacace, B. Zhang, C. Gekeler, T. Banerjee, E. E. Anderson, A. Milanese, U. Löber, S. K. Forslund, K. R. Patil, M. Zimmermann, B. Stecher, G. Zeller, P. Bork and A. Typas, *Nature*, 2021, **599**, 120–124.
- 12 J. G. Liu, O. Gefen, I. Ronin, M. Bar-Meir and N. Balaban, *Science*, 2020, **367**, 200–204.
- 13 Y. Liu, L. Shi, L. Su, H. C. van der Mei, P. C. Jutte, Y. Ren and H. J. Busscher, *Chem. Soc. Rev.*, 2019, **48**, 428–446.
- 14 X. Kong, Y. Qi, X. Wang, R. Jiang, J. Wang, Y. Fang, J. Gao and K. C. Hwang, *Prog. Mater. Sci.*, 2023, **134**, 101070.
- 15 A. Gupta, S. Mumtaz, C.-H. Li, I. Hussain and V. M. Rotello, *Chem. Soc. Rev.*, 2019, **48**, 415–427.
- 16 N. Dasgupta and C. Ramalingam, *Environ. Chem. Lett.*, 2016, **14**, 477–485.
- 17 H. Sun, N. Gao, K. Dong, J. Ren and X. Qu, *ACS Nano*, 2014, **8**, 6202–6210.
- 18 M. Liang and X. Yan, *Acc. Chem. Res.*, 2019, **52**, 2190–2200.
- 19 Y. Zhu, C. Xu, N. Zhang, X. Ding, B. Yu and F. J. Xu, *Adv. Funct. Mater.*, 2018, **28**, 1706709.
- 20 B. Li, Y. Liao, X. Su, S. Chen, X. Wang, B. Shen, H. Song and P. Yue, *J. Nanobiotechnol.*, 2023, **21**, 325.
- 21 S. Dong, Y. Bi, X. Sun, Y. Zhao, R. Sun, F. Hao, Y. Sun, Y. Wang, X. Li, W. Deng, X. Liu, J. Ha, L. Teng, P. Gong, J. Xie, B. Y. S. Kim, Z. Yang, W. Jiang and L. Teng, *Small*, 2022, **18**, 2107690.
- 22 M. Li, Y. Wang, L. Zhang, Q. Liu, F. Jiang, W. Hou, Y. Wang, H. Fang and Y. Zhang, *ACS Nano*, 2023, **17**, 16703–16714.
- 23 P. P. Kalelkar, M. Riddick and A. J. García, *Nat. Rev. Mater.*, 2021, **7**, 39–54.
- 24 M. Bar-Zeev, Y. D. Livney and Y. G. Assaraf, *Drug Resistance Updates*, 2017, **31**, 15–30.
- 25 C. C. Wong and J. Yu, *Nat. Rev. Clin. Oncol.*, 2023, **20**, 429–452.
- 26 V. Chandra and F. McAllister, *Gut*, 2021, **70**, 1419–1425.
- 27 J. An, H. Kwon and Y. J. Kim, *J. Clin. Med.*, 2023, **12**, 2216.
- 28 R. L. Siegel, N. S. Wagle, A. Cercek, R. A. Smith and A. Jemal, *Ca-Cancer J. Clin.*, 2023, **73**, 233–254.
- 29 M. S. R. Rajoka, H. M. Mehwish, Y. Xiong, X. Song, N. Hussain, Q. Zhu and Z. He, *Trends Food Sci. Technol.*, 2021, **107**, 240–251.
- 30 N. Wang and J.-Y. Fang, *Trends Microbiol.*, 2023, **31**, 159–172.
- 31 J. Abed, J. E. M. Emgård, G. Zamir, M. Faroja, G. Almogy, A. Grenov, A. Sol, R. Naor, E. Pikarsky, K. A. Atlan, A. Mellul, S. Chaushu, A. L. Manson, A. M. Earl, N. Ou, C. A. Brennan, W. S. Garrett and G. Bachrach, *Cell Host Microbe*, 2016, **20**, 215–225.
- 32 M. Xu, M. Yamada, M. Li, H. Liu, S. G. Chen and Y. W. Han, *J. Biol. Chem.*, 2007, **282**, 25000–25009.
- 33 M. R. Rubinstein, X. Wang, W. Liu, Y. Hao, G. Cai and Y. W. Han, *Cell Host Microbe*, 2013, **14**, 195–206.
- 34 Q. Meng, Q. Gao, S. Mehrazarin, K. Tangwanichgapong, Y. Wang, Y. Huang, Y. Pan, S. Robinson, Z. Liu, A. Zangiabadi, R. Lux, P. N. Papapanou, X. E. Guo, H. Wang, L. E. Berchowitz and Y. W. Han, *EMBO Rep.*, 2021, **22**, 52891.
- 35 A. Janney, F. Powrie and E. H. Mann, *Nature*, 2020, **585**, 509–517.
- 36 A. D. Kostic, E. Chun, L. Robertson, J. N. Glickman, C. A. Gallini, M. Michaud, T. E. Clancy, D. C. Chung, P. Lochhead, G. L. Hold, E. M. El-Omar, D. Brenner, C. S. Fuchs, M. Meyerson and W. S. Garrett, *Cell Host Microbe*, 2013, **14**, 207–215.
- 37 S. Zhang, Y. Yang, W. Weng, B. Guo, G. Cai, Y. Ma and S. Cai, *J. Exp. Clin. Cancer Res.*, 2019, **38**, 14.
- 38 M. Nomura, R. Nagatomo, K. Inoue, K. Doi, J. Shimizu, K. Baba, T. Saito, S. Matsumoto and M. Muto, *Ann. Oncol.*, 2019, **30**, 509.
- 39 G. Li, J. Lin, C. Zhang, H. Gao, H. Lu, X. Gao, R. Zhu, Z. Li, M. Li and Z. Liu, *Gut Microbes*, 2021, **13**, 1968257.
- 40 W. T. Cheng, H. K. Kantilal and F. Davamani, *Malays. J. Med. Sci.*, 2020, **27**, 9–21.
- 41 C. M. Dejea, P. Fathi, J. M. Craig, A. Boleij, R. Taddese, A. L. Geis, X. Wu, C. E. D. Shields, E. M. Hechenbleikner, D. L. Huso, R. A. Anders, F. M. Giardiello, E. C. Wick, H. Wang, S. Wu, D. M. Pardoll, F. Housseau and C. L. Sears, *Science*, 2018, **359**, 592–597.
- 42 Y. Wang, G. Yang, L. You, J. Yang, M. Feng, J. Qiu, F. Zhao, Y. Liu, Z. Cao, L. Zheng, T. Zhang and Y. Zhao, *Mol. Cancer*, 2019, **18**, 173.
- 43 D. Iain, *Nat. Rev. Gastroenterol. Hepatol.*, 2018, **15**, 328.
- 44 E. Riquelme, Y. Zhang, L. Zhang, M. Montiel, M. Zoltan, W. Dong, P. Quesada, I. Sahin, V. Chandra, A. S. Lucas,

- P. Scheet, H. Xu, S. M. Hanash, L. Feng, J. K. Burks, K. Do, C. B. Peterson, D. Nejman and F. McAllister, *Cell*, 2019, **178**, 795–806.
- 45 S. Pushalkar, M. Hundeyin, D. Daley, C. P. Zambirinis, E. Kurz, A. Mishra, N. Mohan, B. Aykut, M. Usyk, L. E. Torres, G. Werba, K. Zhang, Y. Guo, Q. Li, N. Akkad, S. Lall, B. Wadowski, J. Gutierrez, M. Janal, A. Saxena, X. Li, D. Cohen, R. B. Sartor, D. Saxena and G. Miller, *Cancer Discovery*, 2018, **8**, 403–416.
 - 46 A. Leiter, R. R. Veluswamy and J. P. Wisnivesky, *Nat. Rev. Clin. Oncol.*, 2023, **20**, 624–639.
 - 47 W. Barcik, R. C. T. Boutin, M. Sokolowska and B. B. Finlay, *Immunity*, 2020, **52**, 241–255.
 - 48 J. J. Tsay, B. G. Wu, I. Sulaiman, K. Gershner, R. Schluger, Y. Li, T. Yie, P. Meyn, E. Olsen, L. Perez, B. Franca, J. Carpenito, T. Iizumi, M. El-Ashmawy, M. Badri, J. T. Morton, N. Shen, L. He, G. Michaud, S. Rafeq, J. L. Bessich, R. L. Smith, H. Sauthoff, K. Felner, R. Pillai, A. Zavitsanou, S. B. Koralov, V. Mezzano, C. A. Loomis, A. L. Moreira, W. Moore, A. Tsirigos, A. Heguy, W. N. Rom, D. H. Sterman, H. I. Pass, J. C. Clemente, H. Li, R. Bonneau, K. Wong, T. Papagiannakopoulos and L. N. Segal, *Cancer Discovery*, 2021, **11**, 293–307.
 - 49 C. Jin, G. K. Lagoudas, C. Zhao, S. Bullman, A. Bhutkar, B. Hu, S. Ameh, D. Sandel, X. S. Liang, S. Mazzilli, M. T. Whary, M. Meyerson, R. Germain, P. C. Blainey, J. G. Fox and T. Jacks, *Cell*, 2019, **176**, 998–1013.
 - 50 H. Sung, J. Ferlay, R. L. Siegel, M. Laversanne, I. Soerjomataram, A. Jemal and F. Bray, *Ca-Cancer J. Clin.*, 2021, **71**, 209–249.
 - 51 A. Fu, B. Yao, T. Dong, Y. Chen, J. Yao, Y. Liu, H. Li, H. Bai, X. Liu, Y. Zhang, C. Wang, Y. Guo, N. Li and S. Cai, *Cell*, 2022, **185**, 1356–1372.
 - 52 L. Parhi, T. Alon-Maimon, A. Sol, D. Nejman, A. Shhadeh, T. Fainsod-Levi, O. Yajuk, B. Isaacson, J. Abed, N. Maalouf, A. Nissan, J. Sandbank, E. Yehuda-Shnaidman, F. Ponath, J. Vogel, O. Mandelboim, Z. Granot, R. Straussman and G. Bachrach, *Nat. Commun.*, 2020, **11**, 3259.
 - 53 S. Parida, S. Wu, S. Siddharth, G. Wang, N. Muniraj, A. Nagalingam, C. Hum, P. Mistriotis, H. Hao, C. C. Talbot, K. Konstantopoulos, K. L. Gabrielson, C. L. Sears and D. Sharma, *Cancer Discovery*, 2021, **11**, 1138–1157.
 - 54 S. Bullman, C. S. Peadarallu, E. Sicinska, T. E. Clancy, X. Zhang, D. Cai, D. Neuberg, K. Huang, F. Guevara, T. Nelson, O. Chipashvili, T. Hagan, M. Walker, A. Ramachandran, B. Diosdado, G. Serna, N. Mulet, S. Landolfi, S. R. Y. Cajal, R. Fasani, A. J. Aguirre, K. Ng, E. Élez, S. Ogino, J. Tabernero, C. S. Fuchs, W. C. Hahn, P. Nuciforo and M. Meyerson, *Science*, 2017, **358**, 1443–1448.
 - 55 W. Mu, Q. Chu, Y. Liu and N. Zhang, *Nano-Micro Lett.*, 2022, **12**, 142.
 - 56 K. Forier, K. Raemdonck, S. C. D. Smedt, J. Demeester, T. Coenye and K. Braeckmans, *J. Controlled Release*, 2014, **190**, 607–623.
 - 57 X. Sun, G. Wang, H. Zhang, S. Hu, X. Liu, J. Tang and Y. Shen, *ACS Nano*, 2018, **12**, 6179–6192.
 - 58 K. Zheng, M. I. Setyawati, D. T. Leong and J. Xie, *ACS Nano*, 2017, **11**, 6904–6910.
 - 59 L. Zhang, H. Wang and X. Qu, *Adv. Mater.*, 2023, 2211147.
 - 60 D. Wang, G. Yang, H. C. van der Mei, Y. Ren, H. J. Busscher and L. Shi, *Angew. Chem., Int. Ed.*, 2021, **60**, 17714–17719.
 - 61 D. Wang, Y. Cao, G. Yang, S. Zhang, H. C. van der Mei, Y. Ren, T. G. van Kooten, D. A. de Groot, J. J. de Haan, L. Shi and H. J. Busscher, *Adv. Funct. Mater.*, 2023, **33**, 2215153.
 - 62 L. Chen, R. Zhao, Z. Kang, Z. Cao, N. Liu, J. Shen, C. Wang, F. Pan, X. Zhou, Z. Liu, Y. Yang and Q. Chen, *J. Controlled Release*, 2023, **363**, 43–56.
 - 63 D. Zheng, X. Dong, P. Pan, K. Chen, J. Fan, S. Cheng and X. Zhang, *Nat. Biomed. Eng.*, 2019, **3**, 717–728.
 - 64 J. Chen, P. Zhang, Y. Zhao, J. Zhao, X. Wu, R. Zhang, R. Cha, Q. Yao and Y. Gao, *Sci. Adv.*, 2022, **8**, 2789.
 - 65 Q. Chen, Y. Zhu, K. Wang, X. Wang, Y. Zhang, M. Zhou, J. Du, X. Qu, Z. Shi, Y. Zhang, Y. Chen and H. Qin, *Nano Today*, 2023, **52**, 101994.
 - 66 Z. Chen, J. Li, P. Pan, P. Bao, X. Zeng and X. Zhang, *Nano Today*, 2021, **41**, 101329.
 - 67 Z. Han, Q. Chen, D. Zheng, K. Chen, Q. Huang, Z. Zhuang and X. Zhang, *Adv. Mater.*, 2023, **35**, 2302551.
 - 68 Q. Qiu, D. Lu, G. Liu, X. Yang, J. Li, H. Ren, J. Liu, B. Sun and Y. Zhang, *Bioconjugate Chem.*, 2022, **33**, 1944–1952.
 - 69 X. Yan, F. Ma, Q. Chen, X. Gou, X. Li, L. Zhang and H. Gao, *Chem. Eng. J.*, 2022, **450**, 137605.
 - 70 X. Li, Y. Ma, Y. Xin, F. Ma and H. Gao, *ACS Appl. Mater. Interfaces*, 2023, **15**, 14164–14172.
 - 71 X. Zheng, G. Zhu, D. Pan, H. Li, K. Xiao, Q. Gong, Z. Gu, K. Luo and W. Li, *Adv. Mater.*, 2024, **36**, 2308977.
 - 72 L. Chen, R. Zhao, J. Shen, N. Liu, Z. Zheng, Y. Miao, J. Zhu, L. Zhang, Y. Wang, H. Fang, J. Zhou, M. Li, Y. Yang, Z. Liu and Q. Chen, *Adv. Mater.*, 2023, **35**, 2306281.
 - 73 M. Wang, B. Rousseau, K. Qiu, G. Huang, Y. Zhang, H. Su, C. Le Bihan-Benjamin, I. Khati, O. Artz, M. B. Foote, Y. Cheng, K. Lee, M. Z. Miao, Y. Sun, P. Bousquet, M. Hilmi, E. Dumas, A. Hamy, F. Rey, L. Lin, P. M. Armistead, W. Song, A. Vargason, J. C. Arthur, Y. Liu, J. Guo, X. Zhou, J. Nguyen, Y. He, J. P.-Y. Ting, A. C. Anselmo and L. Huang, *Nat. Biotechnol.*, 2023, DOI: [10.1038/s41587-023-01957-8](https://doi.org/10.1038/s41587-023-01957-8).
 - 74 X. Zhang, X. Chen, Y. Guo, G. Gao, D. Wang, Y. Wu, J. Liu, G. Liang, Y. Zhao and F. Wu, *Angew. Chem., Int. Ed.*, 2021, **60**, 14013–14021.
 - 75 W. Song, D. Zheng, S. Zeng, X. Zeng and X. Zhang, *ACS Nano*, 2022, **16**, 17402–17413.
 - 76 Z. Ma, H. Wang, Z. Shi, F. Yan, Q. Li, J. Chen, Z. Cui, Y. Zhang, X. Jin, Y. Jia and L. Wang, *ACS Nano*, 2023, **17**, 5740–5756.
 - 77 W. Fan, B. Yung, P. Huang and X. Chen, *Chem. Rev.*, 2017, **117**, 13566–13638.
 - 78 X. Qu, F. Yin, M. Pei, Q. Chen, Y. Zhang, S. Lu, X. Zhang, Z. Liu, X. Li, H. Chen, Y. Zhang and H. Qin, *ACS Nano*, 2023, **17**, 11466–11480.

- 79 X. Wang, Q. Chen, Y. Zhu, K. Wang, Y. Chang, X. Wu, W. Bao, T. Cao, H. Chen, Y. Zhang and H. Qin, *Signal Transduction Targeted Ther.*, 2023, **8**, 277.
- 80 J. Xi, Y. Wang, X. Gao, Y. Huang, J. Chen, Y. Chen, L. Fan and L. Gao, *Nano Today*, 2022, **43**, 101395.
- 81 X. Kang, F. Bu, W. Feng, F. Liu, X. Yang, H. Li, Y. Yu, G. Li, H. Xiao and X. Wang, *Adv. Mater.*, 2022, **34**, 2206765.
- 82 R. Liu, H. Yang, S. Qu, P. Yang, X. Zhi, Y. Xu, Z. Dai and L. Qian, *Aggregate*, 2023, **5**, e423.
- 83 Y. Xin, Y. Yu, M. Su, X. Li, M. Elsbahy and H. Gao, *J. Controlled Release*, 2023, **359**, 69–84.