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Unveiling the impact of oxidation-driven endogenous protein interactions on the dynamics of amyloid- β aggregation and toxicity†

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Cytochrome *c* (Cyt *c*), a multifunctional protein with a crucial role in controlling cell fate, has been implicated in the amyloid pathology associated with Alzheimer's disease (AD); however, the interaction between Cyt *c* and amyloid- β (A β) with the consequent impact on the aggregation and toxicity of A β is not known. Here we report that Cyt *c* can directly bind to A β and alter the aggregation and toxicity profiles of A β in a manner that is dependent on the presence of a peroxide. When combined with hydrogen peroxide (H₂O₂), Cyt *c* redirects A β peptides into less toxic, off-pathway amorphous aggregates, whereas without H₂O₂, it promotes A β fibrillization. The mechanisms behind these effects may involve a combination of the complexation between Cyt *c* and A β , the oxidation of A β by Cyt *c* and H₂O₂, and the modification of Cyt *c* by H₂O₂. Our findings demonstrate a new function of Cyt *c* as a modulator against A β amyloidogenesis.

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Introduction

Alzheimer's disease (AD) is a neurodegenerative disorder that causes memory loss and cognitive impairment.^{1–3} A major pathological hallmark of AD is the deposition of senile plaques primarily composed of amyloid- β (A β) aggregates. A β self-assembles to generate oligomers, protofibrils, and amyloid fibrils.^{3–5} Among various A β species, structured oligomers are

reported to be toxic, with the ability to directly and indirectly disturb intracellular and extracellular systems.⁴ Specifically, A β oligomers can induce oxidative damage, including lipid peroxidation by membrane permeabilization, while oxidative stress can increase A β production by activating β -secretase.^{6–8} This process propagates a circular cascade of A β accumulation and oxidative damage. Recent studies suggest that the intertwined networks associated with A β and cellular components lead to the complicated pathological nature of AD.³ Metal ions, such as Cu(I/II) and Zn(II), bind to A β peptides, modify their aggregation pathways, and stabilize toxic structured oligomers.^{9,10} Furthermore, the overproduction of reactive oxygen species (ROS) triggered by dysregulated redox-active metal ions bound and unbound to A β results in oxidative stress with the subsequent impact towards neuronal death.^{11,12}

Cytochrome *c* (Cyt *c*) is a globular protein that contains 104 amino acid residues with a covalently attached heme group as a cofactor.¹³ Cyt *c* exerts different functions depending on its cellular localization and conditions and, thus, plays the pleiotropic role in biological systems.¹⁴ In general, Cyt *c* serves as an electron shuttle in the mitochondrial respiratory chain as well as a scavenger against ROS.¹⁵ In addition to mitochondria, Cyt *c* is also found in the cytosol and is associated with cell differentiation and proliferation.¹⁵ Moreover, extracellular Cyt *c* is reported to promote the survival of hypoxic neurons.¹⁶ Under oxidative stress, Cyt *c* is released from mitochondria to the cytosol.^{13,14} When the concentration of cytosolic Cyt *c* exceeds the threshold, apoptosis initiates through activating caspases and exhibiting its peroxidase-like activity.^{15,17} A β is proposed to trigger the efflux of Cyt *c* from mitochondria to the cytosol and,

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consequently, cause oxidative stress and apoptosis.^{18,19} Based on these findings, the reactivities of Cyt *c* with A β have been recently explored.^{20–22} For example, Cyt *c* significantly attenuated ROS generated by the A β –heme complexes *via* direct electron transfer.²⁰ Studies with A β mutants and Cyt *c* exhibited that metal-free A β and metal-bound A β have distinct electrostatic interactions with Cyt *c*.²¹ The mechanism of how Cyt *c* affects the pathology associated with A β is not known to date, however.

We questioned whether Cyt *c* directly interacts with A β and modifies its aggregation and toxicity profiles. Thus, we evaluated the impact of Cyt *c* on the aggregation of both metal-free A β and metal-bound A β under normal conditions and oxidative stress as well as the cytotoxicity of the resultant A β species.

Mechanistic details in the distinct reactivity of Cyt *c* towards A β amyloidogenesis in the absence and presence of ROS, including their contacts, conformational changes, and oxidative modifications, were probed. Overall, our work illuminates a novel modulative role of Cyt *c* in the A β -related pathology of AD.

Results and discussion

Effects of Cyt *c* on the aggregation of metal-free and metal-bound A β_{42} with and without ROS

The influence of Cyt *c* on the aggregation of metal-free A β_{42} in the absence and presence of H₂O₂ used as a ROS was first monitored by gel electrophoresis with western blotting

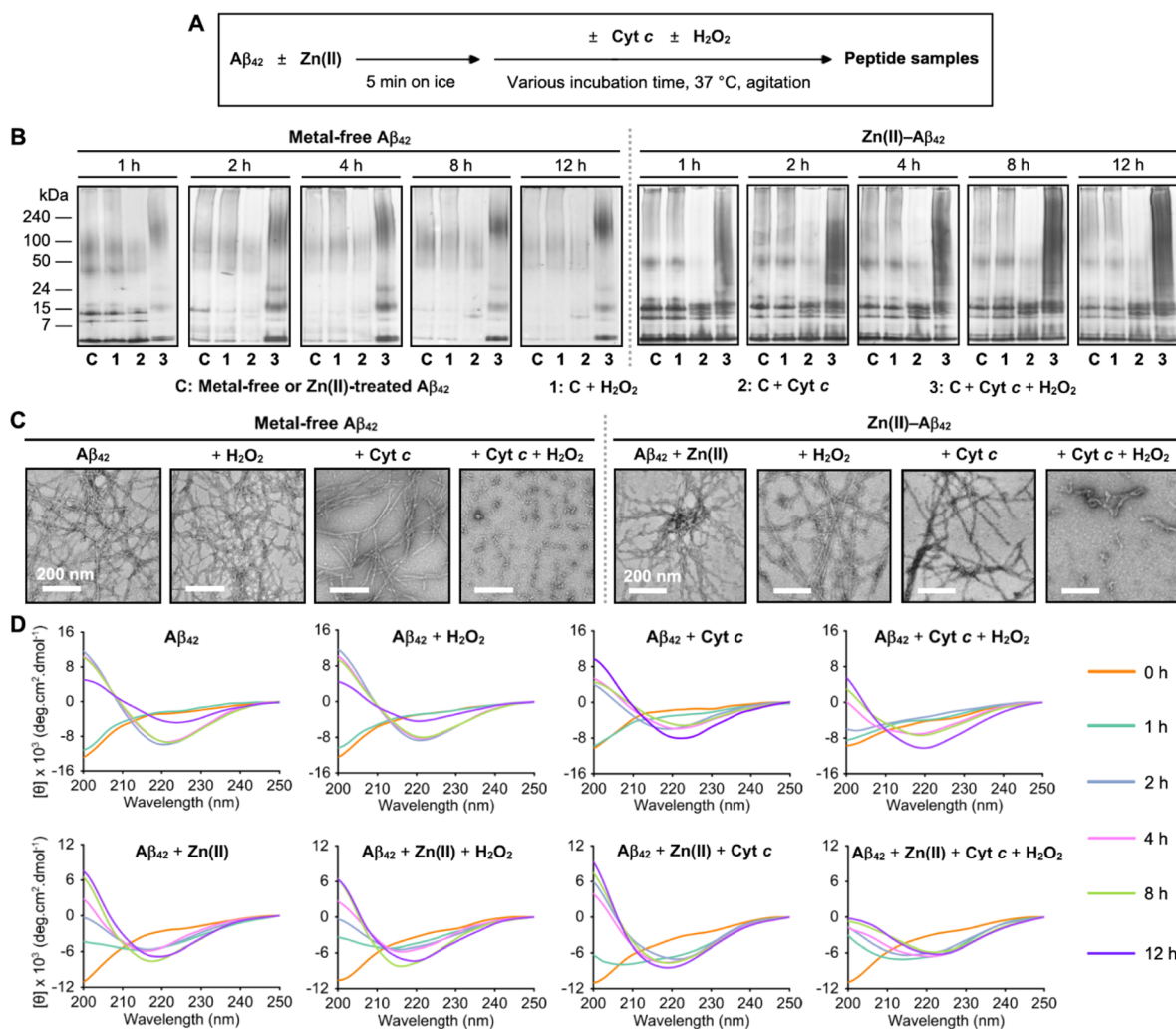


Fig. 1 Effects of Cyt *c* on the aggregation of metal-free and Zn(II)-bound A β_{42} in the absence and presence of H₂O₂. (A) Scheme of A β_{42} aggregation experiments with and without Zn(II). (B) Size distribution of the resultant A β_{42} species at various incubation time points analyzed by gel/western blot using an anti-A β antibody (6E10). (C) Morphology of the peptide aggregates produced after 12 h incubation visualized by TEM. Scale bars = 200 nm. Conditions: [A β_{42}] = 25 μ M; [Zn(II)] = 25 μ M; [Cyt *c*] = 25 μ M; [H₂O₂] = 200 μ M; 150 mM HEPES, pH 7.4; 37 °C; constant agitation (250 rpm). (D) Change in the secondary structures of metal-free and Zn(II)-treated A β_{42} upon aggregation in the presence of Cyt *c* and H₂O₂ observed by CD spectroscopy. A β_{42} was incubated with either Cyt *c*, H₂O₂, or both for 0, 1, 2, 4, 8, and 12 h. The spectra of A β_{42} with Cyt *c* were obtained by subtracting the features of Cyt *c* under the same conditions. In 20 mM sodium phosphate (NaPi), pH 7.4, 150 mM NaF that was chosen to avoid the optical interference from the buffer, the aggregation behavior of A β_{42} was similar to that in the HEPES buffer (Fig. S1†). Conditions: [A β_{42}] = 40 μ M; [Zn(II)] = 40 μ M; [Cyt *c*] = 4 μ M; [H₂O₂] = 200 μ M; 20 mM NaPi, pH 7.4, 150 mM NaF; 37 °C; constant agitation (250 rpm).



(gel/western blot) using an anti-A β antibody (6E10) to determine the molecular weight (MW) distribution of the resultant A β species, as described in Fig. 1A and B. The band intensities observed in low MWs of A β_{42} species in a range of *ca.* 4–15 kDa were weakened upon incubation, indicative of its aggregation. When A β_{42} was treated with Cyt *c*, the aggregation of A β_{42} was accelerated showing the disappearance of overall bands after 2 h incubation. Distinct from A β_{42} with and without Cyt *c*, an increased amount of A β_{42} species ranging from 100 to 240 kDa and the bands at low MWs (*ca.* 4–24 kDa) were observed upon treatment of A β_{42} with both Cyt *c* and H₂O₂. The size distribution of A β_{42} was not significantly changed by H₂O₂ only. Furthermore, the morphology of A β_{42} aggregates produced with either Cyt *c*, H₂O₂, or both was visualized by transmission electron microscopy (TEM). As depicted in Fig. 1C, the samples of A β_{42} with and without H₂O₂ showed long and thin fibrils after 12 h incubation. When A β_{42} was incubated with Cyt *c*, long and thick fibrils were detected. Notably, the addition of both Cyt *c* and H₂O₂ led the generation of amorphous A β_{42} aggregates as well as short and flexible protofibrils. Therefore, these results support that Cyt *c* can alter the aggregation of A β_{42} in the absence and presence of H₂O₂ to different extents. It should be noted that the impact of Cyt *c* on the aggregation kinetics of A β_{42} was not observed by a fluorescent assay because of its signal interference.

To verify the conformational change of A β_{42} upon treatment of either Cyt *c*, H₂O₂, or both, the samples were further analyzed by circular dichroism (CD) spectroscopy. We measured the far-UV CD spectra of A β_{42} incubated with Cyt *c* in an A β_{42} -to-Cyt *c* ratio of 10 : 1 that was selected to limit Cyt *c*'s signals. At this stoichiometry, Cyt *c* could affect the aggregation of A β_{42} (Fig. S2†). As shown in Fig. 1D and S3,† the CD spectra of A β_{42} species with or without H₂O₂ upon incubation exhibited a single negative peak at *ca.* 220 nm, indicative of the formation of amyloid fibrils, and an increase in the β -sheet content to *ca.* 46%. Neither Cyt *c* nor H₂O₂ significantly influenced the change in the secondary structures of A β_{42} . Different from A β_{42} with and without either Cyt *c* or H₂O₂, A β_{42} added with both of them formed the β -sheet structure in a slower manner. The overall results obtained by gel/western blot, TEM, and CD measurements manifest that Cyt *c* can alter the size distribution, morphology, and secondary structures of A β_{42} upon aggregation in a peroxide-dependent manner.

As metal ions affect the aggregation of A β *via* coordination to A β forming metal-bound A β (metal-A β) complexes,^{1,2,9,10} we additionally assessed the impact of Cyt *c* on the aggregation of Zn(II)-A β_{42} with and without H₂O₂, as illustrated in Fig. 1A and B. It should be noted that we did not probe how Cyt *c* affects the aggregation of Cu(II)-A β_{42} in a peroxide-dependent manner because the reaction of Cu(II) with H₂O₂ interferes with our analysis.⁵ The sample containing Zn(II)-A β_{42} and Cyt *c* indicated a decrease in the amount of A β_{42} aggregates (*ca.* 24 to 240 kDa) upon incubation. When Zn(II)-A β_{42} was treated with both Cyt *c* and H₂O₂, the smearing in a range of 4–240 kDa was visualized in the gel, which was different from those from the samples of Zn(II)-A β_{42} with and without addition of either Cyt *c* or H₂O₂. As shown in Fig. 1C, a mixture of short and flexible protofibrils and

amorphous aggregates was detected by incubation of Zn(II)-A β_{42} with both Cyt *c* and H₂O₂, compared to long fibrils produced by Zn(II)-A β_{42} with and without either Cyt *c* or H₂O₂. As expected, the sample of Zn(II)-A β_{42} with Cyt *c* and H₂O₂ exhibited the reduced β -sheet content to *ca.* 24%, relative to that of Zn(II)-A β_{42} only or Zn(II)-A β_{42} with either Cyt *c* or H₂O₂ (*ca.* 40%), as illustrated in Fig. 1D and S4.† These observations confirm the formation of less structured A β_{42} aggregates. Collectively, our aggregation investigations corroborate that Cyt *c* can vary the aggregation behaviors of both metal-free A β_{42} and Zn(II)-bound A β_{42} with and without H₂O₂ to different extents. Particularly, Cyt *c* accelerates the aggregation of A β_{42} in the absence of H₂O₂ and redirects A β peptides into off-pathway less structured assemblies when H₂O₂ is present.

Complexation between Cyt *c* and A β_{42}

To illuminate how Cyt *c* affects the aggregation of A β_{42} , the direct interactions between Cyt *c* and A β_{42} were first analyzed by electrospray ionization-mass spectrometry (ESI-MS), a soft ionization method for characterizing protein complexes.²³ As depicted in Fig. 2B, new peaks at 1688 and 1878 *m/z* corresponding to the complex between Cyt *c* and A β_{42} with +10 and +9 charge states, respectively, were detected upon treatment of A β_{42} with Cyt *c*. The Cyt *c*-A β_{42} adduct in a 1 : 1 ratio was mainly observed, with the complexes with the Cyt *c*-to-A β ratios of 1 : 2 and 2 : 1 (Fig. S5†). The binding of Cyt *c* to A β_{42} was further investigated by isothermal titration calorimetry (ITC), as shown in Fig. S6.† An endothermic ITC isotherm was obtained and the ITC peak intensity decreased as Cyt *c* was titrated into the solution of A β_{42} , indicating their intermolecular interactions. The ITC peaks exhibited the two components, *i.e.*, the early sharp peak and the late broad peak. The broad ITC peaks may be attributed to the aggregation of A β_{42} by Cyt *c*, which would be further boosted because of rigorous stirring in an ITC cell. The ITC measurements revealed a heat change resulting from both Cyt *c*-A β_{42} interaction and A β_{42} aggregation, which would hamper the accurate fitting analysis to obtain thermodynamic parameters for intermolecular interactions.

Next, the binding properties of Cyt *c* with A β_{42} were probed by two-dimensional (2D) nuclear magnetic resonance (NMR) spectroscopy employing either ¹⁵N-labeled A β_{42} or Cyt *c*. As presented in Fig. 2C, when Cyt *c* was added into uniformly ¹⁵N-labeled A β_{42} , moderate chemical shift perturbations (CSPs) were detectable near or in the self-recognition and C-terminal regions of A β_{42} (*e.g.*, Glu11, Lys16, Val18, Val39, and Ile41), respectively, that are critical for A β aggregation.² To further analyze the binding site of Cyt *c* towards A β_{42} , we employed ¹⁵N-labeled Cyt *c* with 104 amino acid residues that were expressed and purified as previously described.²⁴ Upon incubation of ¹⁵N-labeled Cyt *c* with A β_{42} , CSPs were indicated at several amino acid residues of Cyt *c*, including Gly1, Asp2, Lys5, Gly6, Lys8, Ile11, His18, Gly34, Phe36, Thr40, Tyr46, Leu68, Lys72, and Ile95, as displayed in Fig. 2D and S7.† This suggests that A β_{42} has multiple contacts onto the whole region of Cyt *c*. It should be noted that no significant differences in the CSPs or signal intensity of ¹⁵N-labeled A β_{42} were observed over 12 h incubation



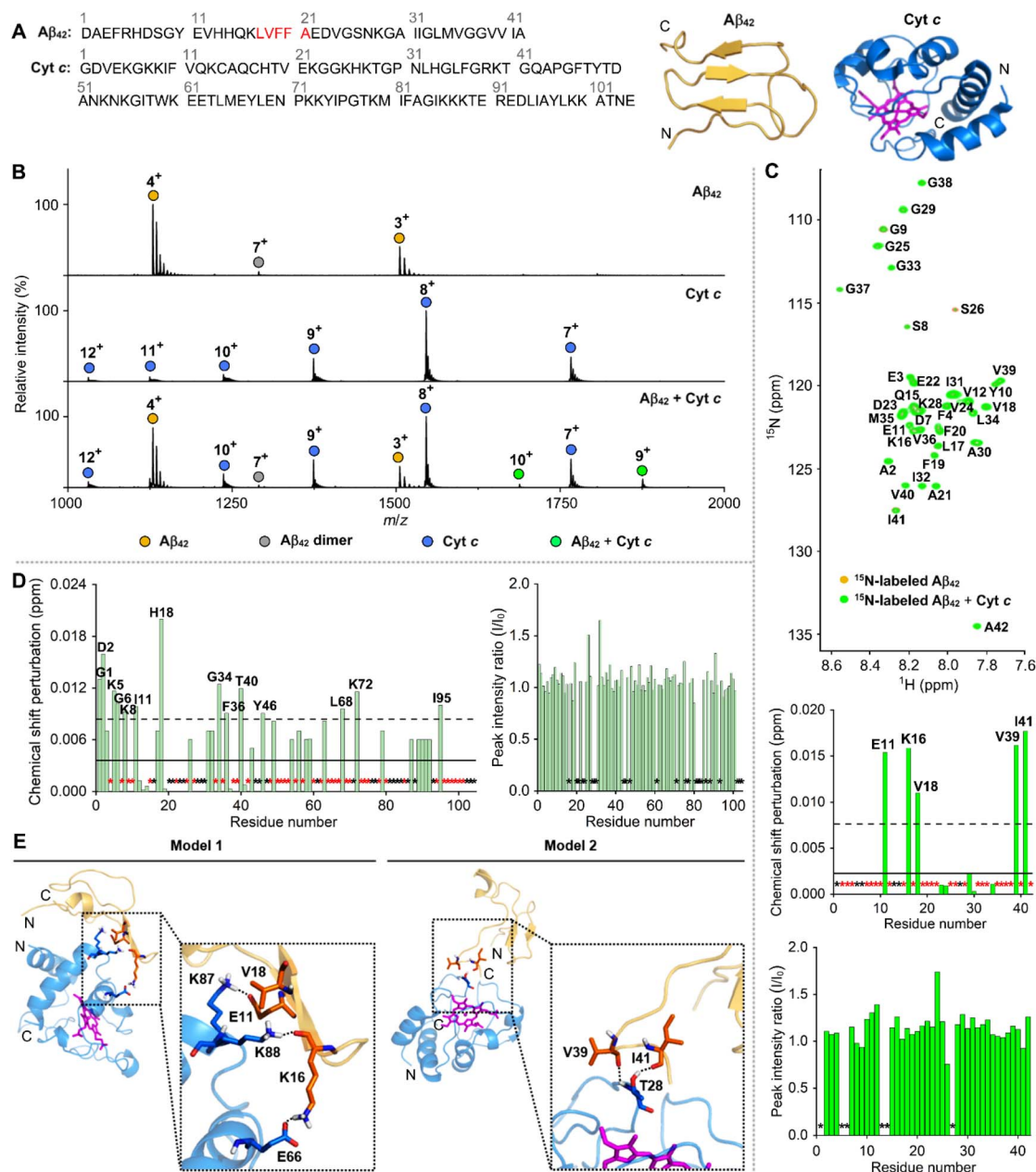


Fig. 2 Interaction between Cyt c and Aβ₄₂. (A) Amino acid sequences and structures of Cyt c (PDB 1HRC)²⁸ and Aβ₄₂.²⁹ The heme group in Cyt c and the self-recognition site of Aβ₄₂ are highlighted in pink and red, respectively. (B) Complexation between Cyt c and Aβ₄₂ detected by ESI-MS. Conditions: [Aβ₄₂] = 100 μM; [Cyt c] = 100 μM; 20 mM ammonium acetate, pH 7.4; 37 °C; incubation for 30 min. The samples were diluted by 10-fold prior to injection to the mass spectrometer. (C) Two-dimensional (2D) ¹H-¹⁵N band-selective optimized flip angle short transient-heteronuclear multiple quantum correlation (SOFAST-HMQC) NMR spectra (800 MHz) of ¹⁵N-labeled Aβ₄₂ monomer with and without Cyt c. The average of chemical shift perturbations (CSPs) and the average plus one standard deviation are presented with solid and dashed lines, respectively. Black asterisks represent the amino acid residues that cannot be resolved for analysis, and red asterisks indicate the amino acid residues without detectable CSPs. Conditions: [¹⁵N-labeled Aβ₄₂] = 40 μM; [Cyt c] = 200 μM; 150 mM HEPES, pH 7.4; 10% v/v D₂O; 10 °C. (D) Analysis of the CSPs and the change in the peak intensity observed within ¹⁵N-labeled Cyt c by addition of Aβ₄₂. The 2D ¹H-¹⁵N heteronuclear single quantum coherence (HSQC) NMR spectra of ¹⁵N-labeled Cyt c with and without Aβ₄₂ monomer are depicted in Fig. S7.† (E) Representative models of the Cyt c-Aβ₄₂ interfaces from the trajectories of MD simulations. Initial conformations of Cyt c (PDB 1HRC; blue)²⁸ and Aβ₄₂ (yellow)²⁹ were used for binding studies. The heme group in Cyt c is highlighted in pink. Model 1 and 2 of the Cyt c-Aβ₄₂ adducts are illustrated in Fig. S9.† Possible hydrogen bonds are depicted with dashed black lines.

(Fig. S8†), indicating that insoluble aggregates did not form under our experimental conditions.

The structural heterogeneity of intrinsically disordered Aβ limits the experimental determination of high-resolution local

and global conformations. Molecular dynamics (MD) simulations guided by experimental data can present a detailed picture for protein-protein interactions.²⁵⁻²⁷ To investigate the potential binding modes of Cyt c to Aβ₄₂, MD simulations were carried out

with an X-ray crystal structure of Cyt *c* (PDB 1HRC)²⁸ and an A β ₄₂ conformer acquired from the MD simulation data,²⁹ as shown in Fig. 2A. Two possible Cyt *c*-A β ₄₂ dimeric interfaces associated with two majorly interacting regions (e.g., Glu11, Lys16, and Val18; Val39 and Ile41) of A β ₄₂ were shown in 2D NMR studies (Fig. 2E and S9†). It should be noted that our MD simulations did not consider the CSPs in Cyt *c* obtained upon addition of A β ₄₂ because they were detected in most of its amino acid residues. The model on the left involves Glu11, Lys16, and Val18 close to the self-recognition site in A β ₄₂ as the major binding region with Cyt *c*. The carboxylate group of Glu11 forms a hydrogen bond with a length of 1.9 Å against the side chain of Lys87 in Cyt *c*. Furthermore, the side chain of Lys16 in A β ₄₂ interacts with Glu66 in Cyt *c* in an average length of 1.9 Å and its backbone carbonyl group exhibits an additional hydrogen bond with Lys88 in Cyt *c*. The hydrophobic Val18 residue of A β ₄₂ resides between two positioned alkyl chains of the aforementioned Lys87 and Lys88 in Cyt *c*. In the other probable model, Thr28 in Cyt *c* plays a key role in interacting with the C-terminal region of A β ₄₂ that is anchored to Cyt *c* by the following two hydrogen bonds: (i) the backbone carbonyl group of Val39 in A β ₄₂ and the backbone amide moiety of the Thr28 in Cyt *c*; (ii) the backbone carbonyl group of Ile41 in A β ₄₂ and the side chain hydroxyl group of Thr28 in Cyt *c*. It should be noted that we chose the initial structure of A β ₄₂ for MD simulations from conformational ensembles identified based on a combination of experimental and theoretical data.²⁹ The monomeric A β ₄₂ structure, which occupied the largest population in the conformational ensembles, was selected as a template. Although the initial structure of A β ₄₂ can influence the trajectories and results of MD simulations, our computational results may provide valuable insights into the molecular interaction between Cyt *c* and A β ₄₂. Overall, our ESI-MS, ITC, NMR, and computational studies support that Cyt *c* can interact with A β ₄₂ in a direct but relatively weak binding manner.

Oxidation of A β ₄₂ by Cyt *c* and H₂O₂

Given that ROS cause oxidative modifications of proteins and, consequently, alter their structures and functions,³⁰ we evaluated whether Cyt *c* could oxidize A β ₄₂ in the presence of H₂O₂. Like other heme-containing peroxidases, Cyt *c* with H₂O₂ can catalytically oxidize a wide range of substrates *via* high-valent Fe(IV) intermediates.³¹ The peroxidase-like activity of Cyt *c* was measured by the ABTS assay [ABTS = 2,2'-azinobis(3-ethylbenzthiazoline-6-sulfonate)].³² Cyt *c* with H₂O₂ noticeably induced the oxidation of ABTS in a concentration- and time-dependent manner while H₂O₂ was not able to significantly oxidize ABTS under our experimental conditions, which suggests its peroxidase-like activity (Fig. S10†).

Moving forward, we probed the oxidation of A β ₄₂ in the presence of both Cyt *c* and H₂O₂ by ESI-MS. The peaks at 1133, 1137, and 1139 *m/z*, assigned to be [A β ₄₂ + O + 4H]⁴⁺, [A β ₄₂ + 2O + 4H]⁴⁺, and [A β ₄₂ + Na + O + 3H]⁴⁺, respectively, were detected, as displayed in Fig. 3A. The peak intensities associated with oxidized A β ₄₂⁴⁺ species produced by Cyt *c* and H₂O₂ were enhanced, compared to those by H₂O₂ only. To identify the oxidation sites of A β ₄₂ by both Cyt *c* and H₂O₂, tandem MS

(ESI-MS²) with collision-induced dissociation (CID) was performed. A mass shift of 16 Da from *y*₈ and *b*₁₄, as presented in Fig. 3B and C, indicated that both His14 and Met35 could be oxidized. Oxidized A β ₄₂⁴⁺ by H₂O₂ at 1133 *m/z* exhibited Met35 as the oxidation site according to a mass shift of 16 Da from *b*₃₅ and *y*₈, as depicted in Fig. 3D. Moreover, in the case of Zn(II)-added A β ₄₂, the oxidation sites of A β ₄₂ by both Cyt *c* and H₂O₂ were identical with those illustrated above (Fig. S11†). These observations regarding A β oxidation are consistent with previous reports regarding oxidative modifications onto the Met and His residues in A β species isolated from amyloid plaques.^{33,34} The Met35 residue in A β is highly susceptible to the oxidation to sulfoxide or sulfone forms upon exposure to H₂O₂.^{35,36} Oxidized Met35 can alter the solubility, aggregation, and cytotoxicity of A β *via* decreasing the hydrophobicity of its C-terminal region and varying its secondary structures.^{35–37} It

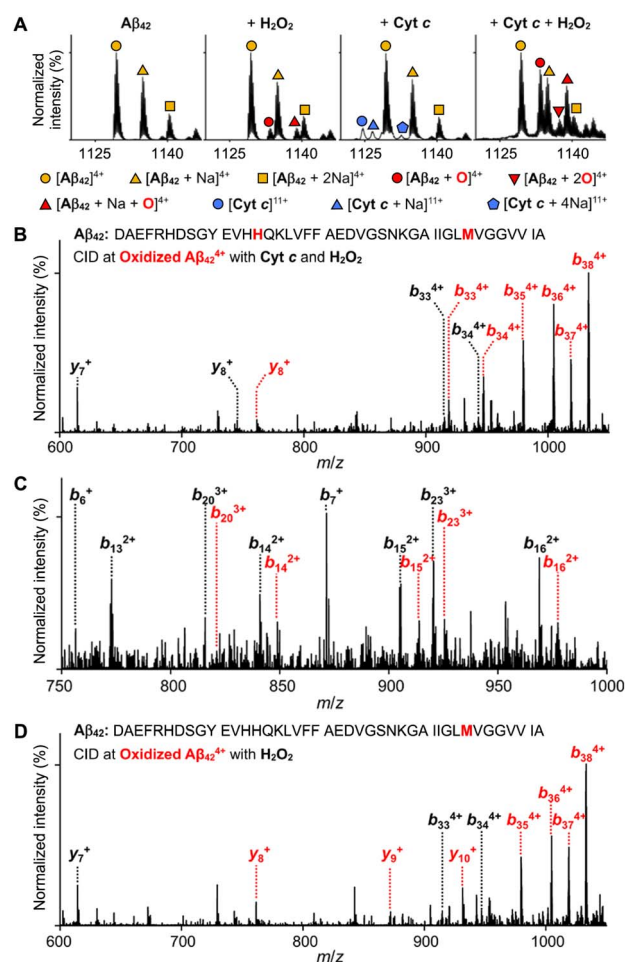
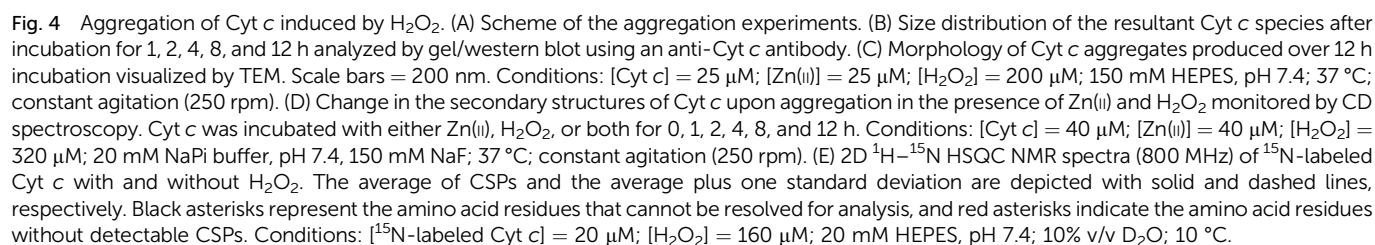


Fig. 3 Oxidation of A β ₄₂ in the presence of Cyt *c* and H₂O₂ analyzed by ESI-MS and ESI-MS². (A) ESI-MS spectra of +4-charged A β ₄₂ treated with either Cyt *c*, H₂O₂, or both. Oxidized A β ₄₂⁴⁺ obtained upon treatment with both Cyt *c* and H₂O₂. (B) 1133 *m/z* (oxidized A β ₄₂⁴⁺); (C) 1139 *m/z* (an adduct of oxidized A β ₄₂⁴⁺ with Na⁺) were analyzed by ESI-MS². (D) ESI-MS² spectrum of oxidized A β ₄₂⁴⁺ produced by H₂O₂. Conditions: [A β ₄₂] = 100 μ M; [Cyt *c*] = 100 μ M; [H₂O₂] = 1.6 mM; 20 mM ammonium acetate, pH 7.4; 37 °C; incubation for 30 min; without agitation. The 10-fold diluted samples were injected to the mass spectrometer.

H₂O₂ is reported to induce the oxidative modifications and conformational fluctuations of Cyt *c*, which ultimately enhances



its peroxidase-like activity and triggers its aggregation.^{39–41} We questioned whether H₂O₂ triggers the aggregation of Cyt *c* under our experimental conditions with the consequent impact on the aggregation of metal-free and Zn(II)-bound Aβ₄₂. Thus, the size distribution, morphology, and change in the secondary structures of the resultant Cyt *c* species upon incubation were analyzed by gel/western blot, TEM, dynamic light scattering (DLS), and CD spectroscopy, as summarized in Fig. 4 and S12–S16.† In the samples of Cyt *c* with H₂O₂ regardless of the presence of Zn(II), the smearing bands in the gels ranging from ca. 35 to 240 kDa were observed within 1 h, indicative of the formation of its aggregates, with the bands at ca. 12 and 24 kDa corresponding to monomeric and dimeric Cyt *c*, as illustrated in Fig. 4B. Additionally, amorphous Cyt *c* aggregates were visualized by TEM when Cyt *c* was added with H₂O₂ (Fig. 4C). Moreover, the hydrodynamic radius (*R*_H) of the resultant Cyt *c* species was determined by DLS. In the samples of Cyt *c* without H₂O₂ in both the absence and presence of Zn(II), the DLS peaks were centered at the *R*_H of ca. 1.7 nm, which is similar to the size of monomeric Cyt *c* reported in the previous study (Fig. S12 and S13†).⁴² DLS data also showed no significant change in the *R*_H value over 12 h incubation, indicating that Cyt *c* did not aggregate without H₂O₂. In the case of the samples of Cyt *c* and H₂O₂ with and without Zn(II), however, new peaks with the *R*_H values larger than 100 nm were observed after 1 h incubation (Fig. S14 and S15†). Upon 12 h of incubation, two peaks were detected in the DLS profile. One peak was centered at ca. 7 nm, while the other was centered at 300–400 nm. These results suggest a heterogeneous distribution of Cyt *c* aggregates. We further examined the samples upon incubation by CD spectroscopy. The far-UV CD spectra of samples containing Cyt *c* and H₂O₂ with and without Zn(II) exhibited significant changes, as depicted in Fig. 4D. Structural analyses of the CD spectra revealed a decrease in the α-helical content from ca. 36% to ca. 29%, while the amount of random coil increased from ca. 30% to ca. 40%, as shown in Fig. S16,† consistent with the previous report.³¹ Collectively, our results obtained by gel/western blot, TEM, DLS, and CD spectroscopy suggest the formation of amorphous Cyt *c* aggregates in the presence of H₂O₂ with and without Zn(II). Note that the treatment of both metal-free Aβ₄₂ and Zn(II)-Aβ₄₂ without H₂O₂ did not induce the aggregation of Cyt *c* (Fig. S17†).

The effect of H₂O₂ on the structural alteration of Cyt *c* was further investigated by 2D ¹H–¹⁵N NMR spectroscopy. As presented in Fig. 4E, upon addition of H₂O₂ into ¹⁵N-labeled Cyt *c*, moderate CSPs of Asp2, Lys5, Gly6, Ile9, Phe10, Asn31, Thr40, Gly56, Trp59, Gly60, Thr63, Leu68, Met80, Lys87, and Arg91 were observed. Furthermore, H₂O₂ significantly decreased the overall peak intensity of Cyt *c*, which could be resulted from the H₂O₂-mediated generation of amorphous NMR-invisible Cyt *c* aggregates. When ¹⁵N-labeled Cyt *c* was incubated with both H₂O₂ and Aβ₄₂, the Asp2, Ile9, Lys27, Lys39, Thr40, Gly56, and Arg91 residues indicated relatively significant CSPs, compared to those with H₂O₂ only (Fig. S18†). H₂O₂ and Aβ₄₂ greatly enhanced the peak intensity of several amino acid residues, indicating that Aβ₄₂ can interact with amorphous Cyt *c* aggregates. It should be noted that no substantial difference in the

CSP and signal intensity of ¹⁵N-labeled Aβ₄₂ was observed by treatment of H₂O₂ (Fig. S19A†), in good agreement with the conclusion drawn from our gel/western blot, TEM, and CD measurements. Upon treatment of Cyt *c* and H₂O₂, the Asp23 and Gly29 residues of ¹⁵N-labeled Aβ₄₂ were subject to greater CSPs, as depicted in Fig. S19B.† The peak intensity was also noticeably reduced, which may be caused by the co-aggregation of Aβ₄₂ and Cyt *c*.

To determine whether amorphous Cyt *c* aggregates and native Cyt *c* modify the aggregation of Aβ₄₂ in a distinct manner, we prepared amorphous Cyt *c* aggregates by incubating Cyt *c* with H₂O₂ for 12 h, as illustrated in Fig. S20A.† After incubation with preformed amorphous Cyt *c* aggregates, the change in the band intensity of monomeric Aβ₄₂ over time was different from that with or without native Cyt *c* (Fig. S20B†). In addition, the smearing in a range of 50–240 kDa indicated the distinct influence of amorphous Cyt *c* aggregates on Aβ₄₂ aggregation, relative to that of Cyt *c* with and without H₂O₂, which was also confirmed by morphological variations observed from TEM (Fig. S20C†). These observations manifest that the modification of Cyt *c* triggered by H₂O₂ was another driving force for modifying Aβ₄₂ aggregation.

Influence of Cyt *c* and H₂O₂ on the cytotoxicity induced by metal-free Aβ₄₂ and Zn(II)-Aβ₄₂

To evaluate the cytotoxicity of metal-free and Zn(II)-added Aβ₄₂ species produced by the addition of Cyt *c* and H₂O₂, we conducted the MTT assay [MTT: 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide] employing a human neuroblastoma SH-SY5Y cell line. For the cell viability assay, we prepared metal-free and Zn(II)-treated Aβ₄₂ aggregates by 12 h incubation with Cyt *c* in the absence and presence of H₂O₂, and cells were incubated with the resultant aggregates for 24 h, as depicted in Fig. 5. Cyt *c* with and without H₂O₂ generated metal-free Aβ₄₂ aggregates that showed less cytotoxicity by approximately 15% and 7%, respectively, than those untreated with either Cyt *c*, H₂O₂, or both. In the case of preformed Zn(II)-Aβ₄₂ aggregates, cell viability was improved by about 6% and 20%, respectively,

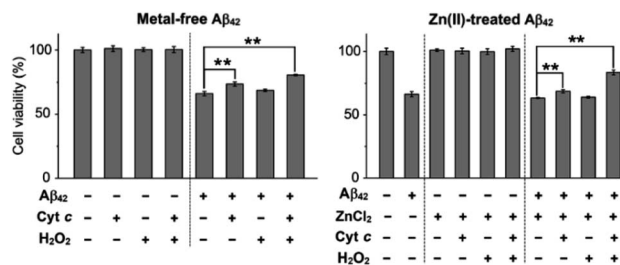


Fig. 5 Influence of Cyt *c* on the cytotoxicity induced by metal-free and Zn(II)-treated Aβ₄₂ species with and without H₂O₂. SH-SY5Y cells were treated with metal-free and Zn(II)-added Aβ₄₂ species generated by 12 h treatment with either Cyt *c*, H₂O₂, or both followed by 24 h incubation. Cell viability, determined by the MTT assay, was calculated in comparison to that with an equivalent amount of the buffered solution. Conditions: [Aβ₄₂] = 10 μM; [Cyt *c*] = 10 μM; [Zn(II)] = 10 μM; [H₂O₂] = 80 μM; 150 mM HEPES, pH 7.4. Data are represented as mean ± s.e.m. ***P* < 0.01 by Student's *t*-test.



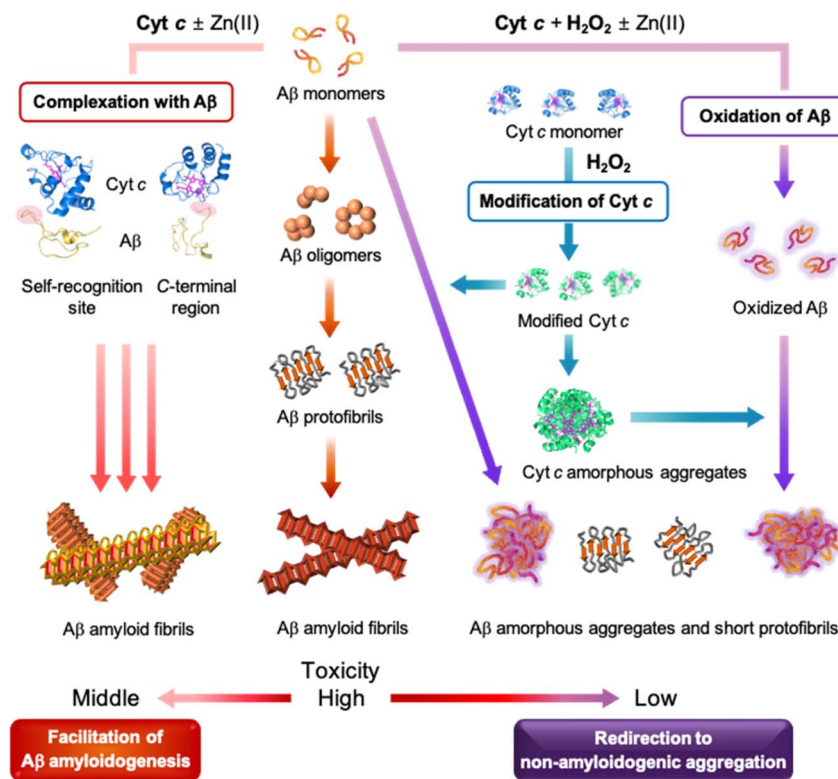


Fig. 6 Potential mechanisms for the impact of Cyt c on Aβ aggregation in the absence and presence of H₂O₂. Cyt c facilitates Aβ aggregation through their adduct formation in the absence of H₂O₂. Upon incubation of Cyt c with Aβ in the presence of H₂O₂, off-pathway amorphous and less toxic Aβ aggregates are generated by three possible mechanisms, including (i) the complexation between Cyt c and Aβ, (ii) the oxidation of Aβ by Cyt c and H₂O₂, and (iii) the modification of Cyt c induced by H₂O₂.

with addition of Cyt c in the absence and presence of H₂O₂. These results illustrate that the aggregates of metal-free Aβ₄₂ and Zn(II)-Aβ₄₂ produced with Cyt c have distinct cytotoxicity depending on the presence of H₂O₂, which is supported by the aggregation studies described in Fig. 1. Notably, amorphous metal-free and Zn(II)-treated Aβ₄₂ aggregates obtained with both Cyt c and H₂O₂ shown in Fig. 1C were less toxic than those with either Cyt c or H₂O₂ and without both. It should be noted that the amounts of Cyt c and H₂O₂ used for cell studies did not cause cytotoxicity under our experimental conditions (Fig. S21†). Our cell and aggregation results suggest the regulatory reactivity of Cyt c towards the aggregation and toxicity of Aβ particularly in the presence of ROS.

Conclusions

Cyt c is a crucial protein that influences various cellular processes linked to respiration, apoptosis, and redox signalling and, thus, is highly relevant to the development of neurodegenerative diseases such as AD.^{13,14,43} Recent findings suggest the reactivity of Cyt c with Aβ;^{20–22} however, details on whether and how Cyt c affects the aggregation and toxicity of Aβ are not known thus far. Through our investigations, we demonstrate, *for the first time*, that Cyt c can modulate the amyloidogenesis of both metal-free Aβ and metal-bound Aβ in a peroxide-dependent manner, to the best of our knowledge. Our findings contribute to a deeper

understanding of the relationship between Cyt c and the aggregation and toxicity of Aβ, which may have important implications for the prevention and treatment of AD.

Our comprehensive studies shed light on the influence of Cyt c on the size distribution, morphology, and secondary structures of both metal-free Aβ and Zn(II)-Aβ to different extents. Our results illustrate that the presence of ROS such as H₂O₂ changes the impact of Cyt c on Aβ amyloidogenesis. As summarized in Fig. 6, Cyt c accelerates Aβ peptides into amyloids, while in the presence of H₂O₂, it redirects Aβ peptides into less toxic off-pathway aggregates. These effects are observed regardless of the presence of Zn(II) and may be driven by a combination of possible mechanisms: (i) the complexation between Cyt c and Aβ, (ii) the oxidation of Aβ by Cyt c and H₂O₂, and (iii) the modification of Cyt c by H₂O₂. Our experimental studies with computer simulations support that Cyt c directly interacts with the self-recognition and C-terminal regions of Aβ that are critical for Aβ aggregation,^{2,4} in a relatively weak binding manner, which results in facilitating Aβ amyloid formation. Furthermore, Cyt c with H₂O₂ promotes oxidative modifications onto the His14 and Met35 residues in Aβ and, thus, alters its aggregation behavior. Lastly, amorphous Cyt c aggregates are formed in the presence of H₂O₂, consistent with previously reported studies,^{31,40} which can notably modify the aggregation of Aβ. The aggregation of metal-free and Zn(II)-bound Aβ in the absence and presence of Cyt c and H₂O₂ is highly complex, involving the generation of diverse



aggregates at intermediate incubation time points. Future investigations would be valuable to elucidate the physiological and pathological activities of intermediate aggregates in biological systems. Our work validates the novel protective of Cyt *c* in A β amyloidogenesis and demonstrates its ability to modulate the aggregation and toxicity profiles of both metal-free and metal-bound A β .

Data availability

All experimental details and data supporting the findings of this study are available within the paper and its ESI.† The data are also available from the corresponding authors upon reasonable request.

Author contributions

Z. D., Y.-H. L., and M. H. L. designed the research. Z. D. and E. N. performed CD, TEM, ESI-MS, UV-Vis, biochemical assays, and cell studies with data analyses. Y. L. and Y.-H. L. conducted 2D ^1H - ^{15}N NMR, ITC, and DLS experiments and analyzed the data. M. H. and M.-H. B. carried out MD simulations with analysis. T. M. performed the expression and purification of ^{15}N -labeled recombinant A β_{42} and K. I. provided ^{15}N -labeled Cyt *c*. Z. D., E. N., M. H., and M. H. L. wrote the manuscript with input from all authors.

Conflicts of interest

There are no conflicts to declare.

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References

- 1 I. W. Hamley, *Chem. Rev.*, 2012, **112**, 5147–5192.
- 2 K. P. Kepp, *Chem. Rev.*, 2012, **112**, 5193–5239.
- 3 M. G. Savellieff, G. Nam, J. Kang, H. J. Lee, M. Lee and M. H. Lim, *Chem. Rev.*, 2019, **119**, 1221–1322.
- 4 S. J. Lee, E. Nam, H. J. Lee, M. G. Savellieff and M. H. Lim, *Chem. Soc. Rev.*, 2017, **46**, 310–323.
- 5 J. Han, Z. Du and M. H. Lim, *Acc. Chem. Res.*, 2021, **54**, 3930–3940.
- 6 D. A. Butterfield, A. M. Swomley and R. Sultana, *Antioxid. Redox Signaling*, 2013, **19**, 823–835.
- 7 L. Zuo, B. T. Hemmelgarn, C. C. Chuang and T. M. Best, *Oxid. Med. Cell. Longevity*, 2015, **2015**, 604658.
- 8 C. Cheignon, M. Tomas, D. Bonnefont-Rousselot, P. Faller, C. Hureau and F. Collin, *Redox Biol.*, 2018, **14**, 450–464.
- 9 E. Atrian-Blasco, P. Gonzalez, A. Santoro, B. Alies, P. Faller and C. Hureau, *Coord. Chem. Rev.*, 2018, **375**, 38–55.
- 10 P. Faller, C. Hureau and O. Berthoumieu, *Inorg. Chem.*, 2013, **52**, 12193–12206.
- 11 K. J. Barnham, C. L. Masters and A. I. Bush, *Nat. Rev. Drug Discovery*, 2004, **3**, 205–214.
- 12 F. Collin, C. Cheignon and C. Hureau, *Biomarkers Med.*, 2018, **12**, 201–203.
- 13 R. Santucci, F. Sinibaldi, P. Cozza, F. Polticelli and L. Fiorucci, *Int. J. Biol. Macromol.*, 2019, **136**, 1237–1246.
- 14 D. Alvarez-Paggi, L. Hannibal, M. A. Castro, S. Oviedo-Rouco, V. Demicheli, V. Tortora, F. Tomasina, R. Radi and D. H. Murgida, *Chem. Rev.*, 2017, **117**, 13382–13460.
- 15 C. Garrido, L. Galluzzi, M. Brunet, P. E. Puig, C. Didelot and G. Kroemer, *Cell Death Differ.*, 2006, **13**, 1423–1433.
- 16 H. Liu, S. M. Sarnaik, M. D. Manole, Y. Chen, S. N. Shinde, W. Li, M. Rose, H. Alexander, J. Chen, R. S. Clark, S. H. Graham and R. W. Hickey, *Resuscitation*, 2012, **83**, 1491–1496.
- 17 K. Gonzalez-Arzola, A. Velazquez-Cruz, A. Guerra-Castellano, M. A. Casado-Combreras, G. Perez-Mejias, A. Diaz-Quintana, I. Diaz-Moreno and M. A. De la Rosa, *FEBS Lett.*, 2019, **593**, 3101–3119.
- 18 H. S. Kim, J. H. Lee, J. P. Lee, E. M. Kim, K. A. Chang, C. H. Park, S. J. Jeong, M. C. Wittendorp, J. H. Seo, S. H. Choi and Y. H. Suh, *Neuroreport*, 2002, **13**, 1989–1993.
- 19 S. Morais Cardoso, R. H. Swerdlow and C. R. Oliveira, *Brain Res.*, 2002, **931**, 117–125.
- 20 C. Ghosh, S. Mukherjee and S. G. Dey, *Chem. Commun.*, 2013, **49**, 5754–5756.
- 21 A. Sarkar, K. Sengupta, S. Chatterjee, M. Seal, P. Faller, S. G. Dey and A. Dey, *ACS Omega*, 2018, **3**, 13994–14003.
- 22 M. Seal, C. Ghosh, O. Basu and S. G. Dey, *J. Biol. Inorg. Chem.*, 2016, **21**, 683–690.
- 23 A. J. Heck and R. H. Van Den Heuvel, *Mass Spectrom. Rev.*, 2004, **23**, 368–389.
- 24 K. Sakamoto, M. Kamiya, T. Uchida, K. Kawano and K. Ishimori, *Biochem. Biophys. Res. Commun.*, 2010, **398**, 231–236.
- 25 P. H. Nguyen, A. Ramamoorthy, B. R. Sahoo, J. Zheng, P. Faller, J. E. Straub, L. Dominguez, J. E. Shea, N. V. Dokholyan, A. De Simone, B. Ma, R. Nussinov, S. Najafi, S. T. Ngo, A. Loquet, M. Chiricotto, P. Ganguly, J. McCarty, M. S. Li, C. Hall, Y. Wang, Y. Miller, S. Melchionna, B. Habenstein, S. Timr, J. Chen, B. Hnath, B. Strodel, R. Kaye, S. Lesne, G. Wei, F. Sterpone, A. J. Doig and P. Derreumaux, *Chem. Rev.*, 2021, **121**, 2545–2647.
- 26 G. J. Morgan, *Biophys. Chem.*, 2022, **281**, 106711.
- 27 P. H. Nguyen and P. Derreumaux, *Biophys. Chem.*, 2020, **264**, 106421.



- 28 G. W. Bushnell, G. V. Louie and G. D. Brayer, *J. Mol. Biol.*, 1990, **214**, 585–595.
- 29 W. Yang, B. S. Kim, Y. Lin, D. Ito, J. H. Kim, Y.-H. Lee and W. Yu, *bioRxiv*, preprint, 2021, 2021.2008.2023.457317, DOI: [10.1101/2021.08.23.457317](https://doi.org/10.1101/2021.08.23.457317).
- 30 N. T. Moldogazieva, I. M. Mokhosoev, N. B. Feldman and S. V. Lutsenko, *Free Radical Res.*, 2018, **52**, 507–543.
- 31 V. Yin, G. S. Shaw and L. Konermann, *J. Am. Chem. Soc.*, 2017, **139**, 15701–15709.
- 32 J. Han, H. J. Lee, K. Y. Kim, G. Nam, J. Chae and M. H. Lim, *Proc. Natl. Acad. Sci. U. S. A.*, 2020, **117**, 5160–5167.
- 33 J. Dong, C. S. Atwood, V. E. Anderson, S. L. Siedlak, M. A. Smith, G. Perry and P. R. Carey, *Biochemistry*, 2003, **42**, 2768–2773.
- 34 C. S. Atwood, X. Huang, A. Khatri, R. C. Scarpa, Y. S. Kim, R. D. Moir, R. E. Tanzi, A. E. Roher and A. I. Bush, *Cell. Mol. Biol.*, 2000, **46**, 777–783.
- 35 L. Hou, I. Kang, R. E. Marchant and M. G. Zagorski, *J. Biol. Chem.*, 2002, **277**, 40173–40176.
- 36 A. S. Johansson, J. Bergquist, C. Volbracht, A. Paivio, M. Leist, L. Lannfelt and A. Westlind-Danielsson, *Neuroreport*, 2007, **18**, 559–563.
- 37 L. Triguero, R. Singh and R. Prabhakar, *J. Phys. Chem. B*, 2008, **112**, 7123–7131.
- 38 C. Hureau and P. Faller, *Biochimie*, 2009, **91**, 1212–1217.
- 39 N. Tomaskova, P. Novak, T. Kozar, M. Petrencakova, D. Jancura, G. Yassaghi, P. Man and E. Sedlak, *Int. J. Biol. Macromol.*, 2021, **174**, 413–423.
- 40 N. H. Kim, M. S. Jeong, S. Y. Choi and J. H. Kang, *Mol. Cells*, 2006, **22**, 220–227.
- 41 M. Hashimoto, A. Takeda, L. J. Hsu, T. Takenouchi and E. Masliah, *J. Biol. Chem.*, 1999, **274**, 28849–28852.
- 42 M. Saxena, Y. Delgado, R. K. Sharma, S. Sharma, S. Guzman, A. D. Tinoco and K. Griebenow, *PLoS One*, 2018, **13**, e0195542.
- 43 A. Guerra-Castellano, I. Marquez, G. Perez-Mejias, A. Diaz-Quintana, M. A. De la Rosa and I. Diaz-Moreno, *Int. J. Mol. Sci.*, 2020, **21**, 8483–8502.

