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Enzyme-loaded Fe³⁺-doped ZIF-90 particles as catalytic bioreactor hybrids for operating catalytic cascades†

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Fe³⁺-doped ZIF-90 (Fe³⁺-ZIF-90), a metal–organic framework (MOF), was synthesized and characterized. The MOF particles reveal peroxidase-like activity reflected by catalyzing the H₂O₂ oxidation of 3,3',5,5'-tetramethylbenzidine, TMB, to TMB⁺. Integration of the two enzymes, β-galactosidase, β-Gal, and glucose oxidase, GOx, in the Fe³⁺-ZIF-90 provides an organized framework allowing the operation of a three-catalyst cascade, where the β-Gal-catalyzed oxidation of lactose yields glucose and galactose, and the resulting glucose is aerobically oxidized by GOx to gluconic acid and H₂O₂, followed by the Fe³⁺-ZIF-90-catalyzed H₂O₂ oxidation of TMB to TMB⁺. The coupled bienzyme/nanozyme cascade in the MOFs is ca. 5-fold enhanced, as compared to a homogeneous mixture of the catalytic constituents. The enhanced catalytic activity of the enzyme cascades in the MOFs is attributed to the confined reaction framework, allowing product channeling across the multienzyme constituents and overcoming diffusion barriers. Moreover, the enzymes, acetylcholine esterase, AChE, and choline oxidase, ChOx, are encapsulated in the confined porous Fe³⁺-ZIF-90 particles. The catalytic cascade where the neurotransmitter acetylcholine is hydrolyzed by AChE followed by the stepwise ChOx-catalyzed oxidation of choline to betaine and H₂O₂, and the Fe³⁺-ZIF-90-catalyzed oxidation of TMB to colored TMB⁺ by H₂O₂ is demonstrated. The three-catalyst cascade is ca. 5-fold enhanced as compared to the mixture of separated catalysts. The integrated three-catalyst AChE/ChOx/Fe³⁺-ZIF-90 particles are applied as colorimetric sensors detecting the neurotransmitter acetylcholine and probing AChE inhibitors. The novelty of the systems is reflected by the assembly of multienzyme catalytic Fe³⁺-ZIF-90 hybrids in confined environments as bioreactor frameworks driving effective biocatalytic cascades.

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Introduction

Metal–organic frameworks (MOFs) find growing interest as functional materials for catalysis,^{1,2} separation,^{3,4} gas storage,^{5,6} water capture,^{7–9} clean energy,¹⁰ food safety,^{11,12} and environment remediation.^{13,14} Their high loading capacity, high surface area, and chemical functionalization enable their diverse applications for analytical and medical uses, such as sensors,¹⁵ imaging,^{16–18} and drug carriers for controlled release.^{19–22}

The catalytic functions of MOF particles can be engineered by three different strategies, including: (i) the use of the metal-ions integrated into the MOF as catalytic sites. For example, Cr³⁺ or Fe³⁺-MIL-100 MOF particles catalyzed the acetalization of benzaldehyde²³ and Co²⁺-ZIF-67 catalyzed the transformation of epoxide into cyclic carbonate in the presence of CO₂.²⁴ (ii) The anchoring of auxiliary metal-ions or complexes to the ligands composing the MOFs. For example, ligation of Cu²⁺ to the bipyridine ligand associated with UiO-type MOFs yielded a peroxidase-like catalyst oxidizing NADH to NAD⁺ or dopamine to aminochrome by H₂O₂,²⁵ and conjugation of a Cu²⁺-Schiff-base complex to an UiO-66-NH₂(Zr) framework catalyzed the Knoevenagel condensation and Michael addition processes.²⁶ (iii) The integration of homogeneous catalysts, catalytic nanoparticles/clusters or biocatalysts into the MOFs. For example, a Pd(II)-bipyridine complex encapsulated in UiO-67 MOF particles catalyzed the Heck coupling reaction between aryl halides and olefins or the Suzuki–Miyaura coupling of aryl chlorides and aryl boronic acids.²⁷ Also, Pd nanoparticles integrated into MIL-101-NH₂ MOFs catalyzed the hydrogenation of

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nitroaromatic substrates,²⁸ and gold nanoclusters loaded in UiO-66 catalyzed the oxidative esterification of furfural.²⁹

The encapsulation of biomolecules within the MOF matrices is particularly interesting since it provides a means to stabilize the recyclable biocatalysts and enables the triggered release of biomolecule loads using stimuli-responsive MOFs. Nevertheless, since many of the MOFs are synthesized at high temperatures and organic solvents, the encapsulation of biomolecules in the MOF matrices during their synthesis is hampered by their thermal deactivation during the encapsulation process. An interesting sub-family of MOF particles, particularly as frameworks for encapsulation of biomolecules, is the family of zeolitic imidazolate framework (ZIF) particles.^{30,31} The biocompatibility of ZIF particles and their preparation in aqueous solution at room temperature make these particles ideal matrices for encapsulating biomolecules. Indeed, basic Zn²⁺-imidazolate frameworks (ZIF-8) provided a versatile matrix for the encapsulation of biomolecules, such as enzymes,^{32–35} antibodies,³⁶ protein drugs (*e.g.*, insulin and vaccines),^{37,38} and DNA.^{39,40} Indeed, multienzymes were integrated into ZIF-8 matrices, and the advantages of the confined media on the cascaded catalysis were demonstrated.^{33,41} Nevertheless, the ZIF-8 matrices are not free of limitations. Their sensitivity to pH, degradability under acidic conditions, the lack of catalytic activities of the Zn²⁺ composing the frameworks, and the lack of surface functionalities allowing the further modification of the frameworks are serious limitations. Different strategies to overcome these difficulties were reported. These included the synthesis of ZIF particles composed of catalytic ions, *e.g.*, Co²⁺-ZIF-67,⁴² the stabilization of the ZIF structures by polymer coatings such as polyaniline,⁴³ and the construction of catalytic ZIF structures with functionalized imidazolate ligands, such as carboxaldehyde-modified imidazolate that binds diamine tethers and stabilizes Au nanoparticles for catalyzed hydrogenation of nitro-aromatic compounds.⁴⁴ Also, Zn²⁺-ZIF composites were doped with catalytic metal ions, such as Fe³⁺, Cu²⁺ or Mn²⁺ and the resulting hybrid ZIF structures revealed diverse catalytic activities, such as the aerobic oxidation of benzylic hydrocarbon⁴⁵ or cycloaddition reactions.⁴⁶ Moreover, by tethering auxiliary catalysts, such as hemin/G-quadruplex, to carboxaldehyde groups associated with ZIF-90 units, the ZIF hybrid could guide catalytic reactions, *e.g.*, the catalyzed H₂O₂ oxidation of *N*-hydroxy-L-arginine to citrulline.⁴⁷ Furthermore, structural changes induced by the ZIF particles on the encapsulated biocatalysts were reported to change the catalytic features of the catalytic loads.^{48–50} Different applications of the ZIF particles for sensing,^{51,52} controlled drug release^{53,54} and cascaded catalysis^{33,41} were demonstrated.

Here we wish to report the synthesis of Fe³⁺-doped ZIF-90 (Fe³⁺-ZIF-90) nanozymes exhibiting peroxidase-like catalytic functions and the assembly of multienzyme-loaded Fe³⁺-ZIF-90 MOFs. The biocatalyst/MOF composites are used as functional hybrids operating diverse catalytic cascades. Specifically, we integrate into the ZIF particles different single type enzymes, *e.g.*, glucose oxidase (GOx) or choline oxidase (ChOx), and coupled multienzymes, consisting of β -galactosidase (β -Gal)/GOx or acetylcholine esterase (AChE)/ChOx, which operate

biocatalytic cascades. We describe the conjugation of the biocatalytic cascades to the peroxidase-like catalytic functions of the synthetic Fe³⁺-doped catalytic sites of the Fe³⁺-doped MOF particles. Beyond the novel coupling of biocatalytic cascades to synthetic catalytic sites of the MOF structures to operate a multimodal catalytic bioreactor, we introduce the systems for sensing applications. Specifically, we address the use of the AChE/ChOx/Fe³⁺-ZIF-90 hybrid as a functional composite probing the activity of AChE, an important catalyst for metabolic degradation of the acetylcholine neurotransmitter. Furthermore, we introduce the use of the AChE/ChOx/Fe³⁺-ZIF-90 hybrid system to probe the effect of a 1,5-bis(4-allyldimethylammoniumphenyl)pentane-3-one dibromide (BW284C51) inhibitor, a nerve gas simulator, on the activity of AChE, thereby demonstrating the potential use of the system for sensitive detection of hazardous nerve gas. In the different coupled enzyme/nanozyme systems, we emphasize the importance of the confined reaction media of the frameworks on the coupled catalytic cascades by comparing the catalytic performance of the integrated assemblies to the catalytic cascades driven by the diffusive and separated enzyme/nanozyme constituents in bulk solution media.

Results and discussion

The Fe³⁺-ZIF-90 MOFs were prepared according to the reported method,⁵⁵ with slight modification, by mixing Zn(NO₃)₂ and FeCl₃ with 2-imidazolecarboxaldehyde and polyvinylpyrrolidone in a water/ethanol solution and stirring the mixture at room temperature, as shown in Fig. 1A. The particles, *ca.* 1 μ m in diameter, exhibit a rhombic dodecahedral structure, as shown in Fig. 1B. The powder X-ray diffraction (PXRD) spectrum of the particles, as shown in Fig. 1C, overlaps the spectrum of ZIF-90 (PDF 02-107-8543), indicating that the crystalline structure of ZIF-90 is retained in the doped particles. The deconvoluted Fe 2p X-ray photoelectron spectroscopy (XPS) spectra in Fig. 1D indicate that the doped iron exists in the Fe³⁺-state (for the XPS spectra of the other elements included in the particles, see Fig. S1†). The inductively coupled plasma mass spectrometry (ICP-MS) measurement indicates a *ca.* 5% loading of the doped ions (170.7 μ g mg⁻¹ of Zn²⁺; 7.3 μ g mg⁻¹ of Fe³⁺). The N₂ adsorption/desorption measurement (Fig. S2†) reveals a Brunauer–Emmett–Teller (BET) specific surface area of 317.65 m² g⁻¹.

The Fe³⁺-ZIF-90 particles demonstrate peroxidase-like activity reflected by the catalyzed H₂O₂ oxidation of 3,3',5,5'-tetramethylbenzidine, TMB, generating the blue-colored oxidized TMB, TMB⁺, as shown in Fig. 1E. In addition, the Fe³⁺-ZIF-90 demonstrates other peroxidase-like activities, such as NADH peroxidase function, as shown in Fig. S3.† The reactive oxygen species (ROS) generated by the Fe³⁺-ZIF-90 particles in the presence of H₂O₂ and their participation in the oxidation of TMB were characterized spectroscopically, revealing the formation of hydroxyl radicals ($\cdot$ OH), as shown in Fig. 1F, G and S4.† Fig. 1F and S4† depict the effective temporal depletion of the absorbance of 1,3-diphenylisobenzofuran (DPBF) by the ROS generated in the presence of both Fe³⁺-ZIF-90 and H₂O₂ as





Fig. 1 (A) Synthesis of the Fe³⁺-ZIF-90 framework and its schematic peroxidase-like function catalyzing the oxidation of TMB to TMB^{•+} by H₂O₂. (B) SEM image of the Fe³⁺-ZIF-90 MOF particles. (C) XRD spectrum of the Fe³⁺-ZIF-90 particles: top—experimental results; bottom—spectral band of ZIF-90 from the PDF database. (D) Deconvoluted Fe 2p XPS spectra of the Fe³⁺-ZIF-90 MOF particles. (E) Panel I: time-dependent concentration changes of TMB^{•+} using TMB, 2 mM, and the Fe³⁺-ZIF-90 catalyst, 100 μg mL⁻¹, in the presence of variable concentrations of H₂O₂: (i) 0 mM; (ii) 0.1 mM; (iii) 0.2 mM; (iv) 0.5 mM; (v) 1 mM; (vi) 2 mM; (vii) 5 mM; (viii) 10 mM. Panel II: rates of TMB^{•+} formation in the presence of Fe³⁺-ZIF-90, 100 μg mL⁻¹, TMB, 2 mM, and variable concentrations of H₂O₂. (F) Time-dependent absorbance changes of DPBF, 60 μM, as a result of ROS agent formation: (i) in the presence of Fe³⁺-ZIF-90, 100 μg mL⁻¹, and H₂O₂, 10 mM; (ii) in the presence of Fe³⁺-ZIF-90 particles, 100 μg mL⁻¹, and in the absence of H₂O₂. (G) EPR spectra corresponding to: panel I—(i) [•]OH radicals generated by Fe³⁺-ZIF-90 and H₂O₂; (ii) and (iii) control systems consisting of only Fe³⁺-ZIF-90 and only H₂O₂; (iv) [•]OH generated by Fe³⁺-ZIF-90/H₂O₂ and quenched by dimethyl sulfoxide; (v) [•]OH generated by the Fe³⁺-ZIF-90/H₂O₂ system quenched by TMB. In all experiments [•]OH is trapped by BMPO. Panel II—(i) the spectrum of TMB^{•+} formed in the presence of Fe³⁺-ZIF-90/H₂O₂ and TMB; (ii) and (iii) control systems demonstrating the lack of TMB^{•+} in systems composed of only Fe³⁺-ZIF-90/TMB or H₂O₂/TMB. Panel III—high resolution spectra of TMB^{•+}: (i) generated by the Fe³⁺-ZIF-90/H₂O₂/TMB system; (ii) computationally simulated TMB^{•+}; (iii) the chemical structure of TMB^{•+}.

compared to the low absorbance changes in the presence of the Fe³⁺-ZIF-90 and in the absence of H₂O₂. Fig. 1G displays electron paramagnetic resonance (EPR) spectra using 5-*tert*-butoxycarbonyl-5-methyl-1-pyrroline-*N*-oxide (BMPO) as the trapping agent, confirming the catalyzed formation of [•]OH as ROS

products and their participation in the oxidation of TMB. In panel I, the characteristic [•]OH spectrum, in the presence of both Fe³⁺-ZIF-90 particles and H₂O₂, curve (i), is observed while control experiments show that no radicals are formed in the presence of particles and in the absence of H₂O₂, curve (ii), and

in the presence of H_2O_2 and in the absence of MOFs, curve (iii). Panel I further reveals the formation of $\cdot\text{OH}$ by the quenching effect of dimethyl sulfoxide, curve (iv), and their participation in the oxidation of TMB, curve (v). Furthermore, the reaction of $\cdot\text{OH}$ with TMB is confirmed by the formation of $\text{TMB}^{+\cdot}$, panel II. The high resolution EPR spectrum of $\text{TMB}^{+\cdot}$ (generated in the absence of BMPO) is depicted in panel III, curve (i), and the computationally simulated spectrum is displayed in curve (ii). The results are consistent with the reported spectrum of $\text{TMB}^{+\cdot}$, and the computational spectrum allows the mapping of the electron distribution in the molecular framework,⁵⁶ as presented in the inset structure (iii) (for the mechanistic path corresponding to the Fe^{3+} -ZIF-90 catalyzed generation of $\cdot\text{OH}$, see ESI P. S11†).

The intrinsic peroxidase activities of the Fe^{3+} -doped ZIF-90 frameworks were then implemented to operate the MOF-mediated biocatalytic cascade by integrating enzymes in the Fe^{3+} -ZIF-90 frameworks. In the first step, GOx was integrated into the frameworks during the course of the particle synthesis, as shown in Fig. 2A. Two methods were employed to estimate the loading of the enzyme in the particles: (i) the residual content of GOx in the solution after encapsulation of the enzyme in the particles was evaluated using the GOx/horseradish peroxidase (HPR) standard probing cascade and

an appropriate calibration curve (see Fig. S5† and accompanying discussion for details). The loading of GOx in the Fe^{3+} -ZIF-90, using this method, was estimated to be $97.4 \mu\text{g mg}^{-1}$. (ii) GOx was modified with fluorescein isothiocyanate, FITC. As shown in Fig. S6,† panels I–III display the fluorescence confocal microscopy image of the FITC-modified GOx integrated into the particles. Using a calibration curve relating the fluorescence intensity of FITC-GOx at different concentrations of the labeled enzyme (Fig. S7A and B†) and probing the fluorescence intensity of residual content of the enzyme in solution after encapsulation in the framework (Fig. S7C†), the loading of the fluorescent GOx in the particles was evaluated to be $94.2 \mu\text{g mg}^{-1}$ particles, in good agreement with the loading determined by method (i). The GOx-loaded Fe^{3+} -ZIF-90 particles were characterized by SEM (Fig. 2B) and PXRD (Fig. 2C) demonstrating that the morphology and crystallinity of the ZIF-90 were retained after integration of the enzyme into the framework.

The GOx-loaded Fe^{3+} -ZIF-90 MOFs were then implemented as a catalytic reactor for the cascaded glucose-driven oxidation of TMB, as shown in Fig. 2A. In this system, the GOx-catalyzed aerobic oxidation of glucose yields gluconic acid and H_2O_2 , and the Fe^{3+} -ZIF-90 catalyzes the oxidation of TMB to $\text{TMB}^{+\cdot}$ by the generated H_2O_2 . As shown in Fig. 2D, panel I presents the time-dependent concentration changes of $\text{TMB}^{+\cdot}$ generated by the integrated frameworks in the presence of variable concentrations of glucose. The rate curve corresponding to the GOx-integrated system is displayed in panel II, and the respective $V_{\text{max}} = 0.45 \mu\text{M min}^{-1}$ was derived. In addition, Fig. S8† shows the time-dependent concentration changes of $\text{TMB}^{+\cdot}$, in the presence of glucose (5 mM), using different amounts of GOx loads in the integrated frameworks. As the loading amount increases, the glucose-driven catalytic oxidation of TMB to $\text{TMB}^{+\cdot}$ by the GOx-integrated system is enhanced.

The advantages of operating catalytic cascades in confined reaction media, as compared to diffusional operating enzyme cascades, attracted substantial research interest as a means of mimicking native enzyme cascades in biologically confined environments,⁵⁷ e.g., cell. The spatial proximity of the catalysts in the confined media overcomes diffusional barriers of the substrates/products tunneling between the catalysts participating in the catalytic cascade by providing directional and effective interconnective communication between the catalysts. Indeed, diverse biocatalytic cascades operating in confined media, such as tethered enzymes on programmed DNA nanostructures,^{58–61} encapsulation of enzymes in microdroplets^{62,63} or polymersomes,⁶⁴ demonstrated the advantages of operating the biocatalytic cascades in the organized assemblies. The ability of integrating multienzyme agents in the porous metal-organic frameworks and particularly the feasibility to couple the biocatalytic cascade to the peroxidase-like catalytic functions of the Fe^{3+} -ZIF-90 particles, provide a versatile means to engineer new biocatalyst/ Fe^{3+} -ZIF-90 composite reactors demonstrating the advantages of the confined porous particles as a hybrid reactor system driving the catalytic system. This is exemplified in Fig. 3A with the integration of two enzymes, β -Gal and GOx, in the Fe^{3+} -ZIF-90 porous particles. This bienzyme/ Fe^{3+} -ZIF-90 composite allows, in principle, to

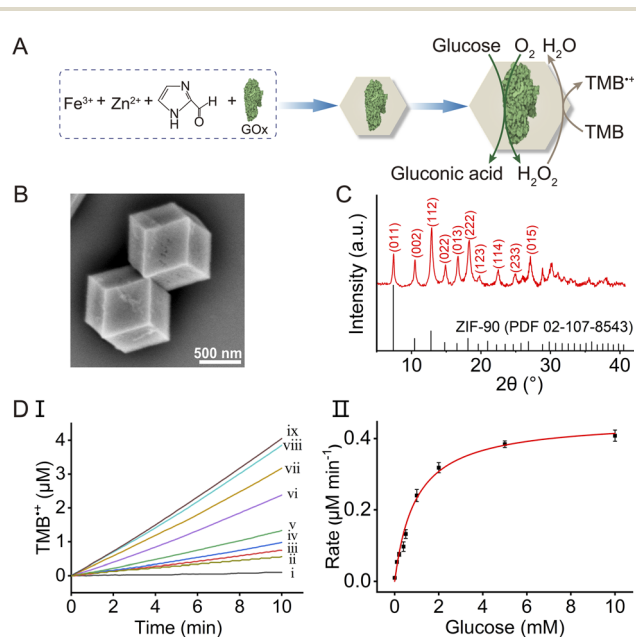


Fig. 2 (A) Schematic synthesis of GOx-loaded Fe^{3+} -ZIF-90 particles and the bioreactor activities towards the cascaded GOx-catalyzed aerobic oxidation of glucose, followed by the Fe^{3+} -ZIF-90-catalyzed oxidation of TMB to $\text{TMB}^{+\cdot}$ by the generated H_2O_2 . (B) SEM image of GOx-loaded Fe^{3+} -ZIF-90 particles. (C) XRD spectrum of the GOx-loaded Fe^{3+} -ZIF-90 particles: top—experimental results; bottom—spectral band from the PDF database. (D) Panel I: time-dependent concentration changes of $\text{TMB}^{+\cdot}$ upon operating the GOx-loaded Fe^{3+} -ZIF-90 framework, $100 \mu\text{g mL}^{-1}$, and TMB, 2 mM, in the presence of variable concentrations of glucose: (i) 0 mM; (ii) 0.1 mM; (iii) 0.2 mM; (iv) 0.4 mM; (v) 0.5 mM; (vi) 1 mM; (vii) 2 mM; (viii) 5 mM; (ix) 10 mM. Panel II: rates of TMB oxidation to $\text{TMB}^{+\cdot}$ by the GOx-loaded Fe^{3+} -ZIF-90/TMB system as a function of glucose concentration.

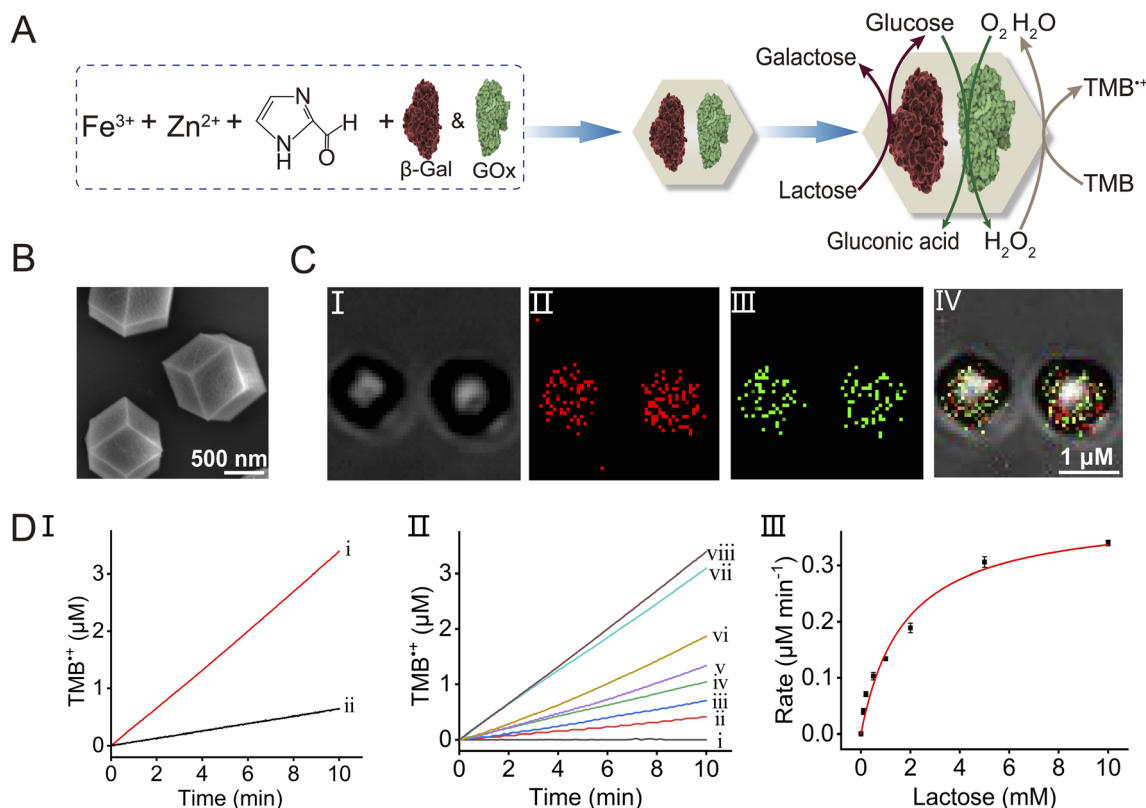


Fig. 3 (A) Schematic assembly of the two enzymes (β -Gal and GOx) into the porous Fe^{3+} -ZIF-90 MOF particles and their use as a three-catalyst cascade driving the β -Gal-catalyzed hydrolysis of lactose, the subsequent aerobic oxidation of glucose, and the cascaded Fe^{3+} -ZIF-90-catalyzed oxidation of TMB by H_2O_2 generated by the biocatalytic cascade. (B) SEM image of the β -Gal/GOx-loaded Fe^{3+} -ZIF-90 MOFs. (C) Bright field and fluorescence confocal microscopy images corresponding to the Atto 565-labeled β -Gal/FITC-labeled GOx loaded in the Fe^{3+} -ZIF-90: panel I—bright field image; panel II—imaging the Atto 565-labeled β -Gal ($\lambda_{\text{ex}} = 561 \text{ nm}$, $\lambda_{\text{em}} = 585 \text{ nm}$); panel III—imaging FITC-labeled GOx ($\lambda_{\text{ex}} = 488 \text{ nm}$, $\lambda_{\text{em}} = 525 \text{ nm}$); panel IV—overlapped image. (D) Panel I—time-dependent concentration changes of TMB^{++} upon operating the lactose-driving three-catalyst cascade using: (i) the β -Gal/GOx-loaded Fe^{3+} -ZIF-90 MOFs as an integrated assembly; (ii) the separated β -Gal, GOx and Fe^{3+} -ZIF-90 constituents in the homogeneous aqueous phase at concentrations identical to those in (i). Panel II—time-dependent concentration changes of TMB^{++} upon operating the β -Gal/GOx/ Fe^{3+} -ZIF-90 cascade in the presence of variable concentrations of lactose: (i) 0 mM; (ii) 0.1 mM; (iii) 0.2 mM; (iv) 0.5 mM; (v) 1 mM; (vi) 2 mM; (vii) 5 mM; (viii) 10 mM. Panel III—rates of TMB^{++} formation catalyzed by the β -Gal/GOx/ Fe^{3+} -ZIF-90 system in the presence of variable concentrations of lactose.

drive the three-catalyst cascade in the confined frameworks, where β -Gal acts as a primary biocatalyst for the catalyzed hydrolysis of lactose to galactose and glucose. The glucose product is then tunneled to GOx as a second enzyme, catalyzing the aerobic oxidation of glucose to gluconic acid and H_2O_2 . The resulting H_2O_2 is then transferred to the porous Fe^{3+} -ZIF-90 nanozyme that catalyzes the oxidation of TMB to TMB^{++} . Accordingly, the two enzymes, β -Gal and GOx, were integrated into the Fe^{3+} -ZIF-90 frameworks within the process of the formation of the particles (see the ESI Experimental section† for details). Fig. 3B depicts the SEM image of the resulting bienzyme-loaded particles, demonstrating that the rhombic dodecahedral crystallinity of the ZIF-90 is retained. The successful entrapment of the enzymes in the Fe^{3+} -ZIF-90 particles was confirmed by quantitative evaluation of the enzyme loading in the frameworks and by demonstrating the operation of the catalytic cascade by the particles (*vide infra*). Towards this goal, β -Gal was labeled with the Atto 565 fluorophore ($\lambda_{\text{ex}} = 561 \text{ nm}$, $\lambda_{\text{em}} = 585 \text{ nm}$) and GOx was labeled with FITC ($\lambda_{\text{ex}} =$

488 nm, $\lambda_{\text{em}} = 525 \text{ nm}$). Fig. 3C depicts the bright field, fluorescence confocal microscopy and merged images of the particles, demonstrating that the two enzymes were, indeed, integrated into the Fe^{3+} -ZIF-90 particles. By monitoring the fluorescence of the residual enzymes in the solution after preparing the particles (and knowing the initial concentrations of the enzymes) and employing appropriate calibration curves relating the fluorescence intensities of the fluorophore-labeled enzyme to their contents (Fig. S7, S9 and S10†), the loading of the enzymes in the particles was evaluated. The loading of β -Gal and GOx was determined to be $82.6 \mu\text{g mg}^{-1}$ and $96.1 \mu\text{g mg}^{-1}$, respectively. The β -Gal/GOx-loaded Fe^{3+} -ZIF-90 particles were then applied to activate the three-catalyst cascade. Fig. 3D, panel I shows the time-dependent concentration changes of TMB^{++} originating upon operation of the three-catalyst cascaded frameworks, β -Gal/GOx/ Fe^{3+} -ZIF-90, in the presence of lactose, 10 mM (curve (i)). For comparison, the formation of TMB^{++} by the diffusional mixture consisting of separated β -Gal, GOx and Fe^{3+} -ZIF-90 (at identical concentrations presented in



the hybrid composite) is presented in curve (ii). A 5-fold enhancement of the three-catalyst cascade in the frameworks, as compared to the diffusive cascaded process of the separated constituents, is observed. The effective operation of the cascade by the MOFs is attributed to the confined environment provided by the porous particles. The spatial proximity between the catalysts provides effective substrate/product transport tunneling pathways intercommunicating the catalysts, thereby overcoming the diffusive barrier present in solution, resulting in the effective operation of the three-catalyst cascade in the MOF particles. As shown in Fig. 3D, panel II depicts the time-dependent concentration changes of TMB^{++} generated by the cascaded $\beta\text{-Gal}/\text{GOx}/\text{Fe}^{3+}\text{-ZIF-90}$ in the presence of different concentrations of lactose. As the concentration of lactose increases, the rates of the generation of TMB^{++} are enhanced. As shown in Fig. 3D, panel III presents the rates of TMB^{++} formation as a function of lactose concentration. A saturation curve is

observed demonstrating a $V_{\text{max}} = 0.40 \mu\text{M min}^{-1}$ for the catalytic cascade operated by the composite $\beta\text{-Gal}/\text{GOx}$ -loaded $\text{Fe}^{3+}\text{-ZIF-90}$ MOFs.

In the next step, we searched for an application of operating a biocatalytic cascade in the $\text{Fe}^{3+}\text{-ZIF-90}$ frameworks that utilizes the advantages of operating a bienzyme/nanozyme cascaded reaction in the confined $\text{Fe}^{3+}\text{-ZIF-90}$ microenvironment. This is exemplified in Fig. 4 with the engineering of bienzyme-loaded $\text{Fe}^{3+}\text{-ZIF-90}$ frameworks composed of acetylcholine esterase, AChE, and choline oxidase, ChOx, with the vision that operation of the three-catalyst cascade in the frameworks could be implemented for sensing applications. Acetylcholine is a major neurotransmitter that signals cholinergic neurons. It is rapidly hydrolyzed by the serine protease, AChE, and any perturbation of binding acetylcholine to the neuronal synaptic membrane or inhibition of AChE can lead to neurological disorders, such as movement perturbation,



Fig. 4 (A) Schematic assembly of the two enzymes (AChE and ChOx) in the porous $\text{Fe}^{3+}\text{-ZIF-90}$ particles and their use as a three-catalyst cascade driving the AChE-catalyzed hydrolysis of acetylcholine, the subsequent aerobic oxidation of choline and the cascaded $\text{Fe}^{3+}\text{-ZIF-90}$ -catalyzed oxidation of TMB by H_2O_2 generated by the biocatalytic cascade. (B) Panel I: SEM image of the AChE/ChOx-loaded $\text{Fe}^{3+}\text{-ZIF-90}$ MOFs. (C) XRD spectrum of the AChE/ChOx-loaded $\text{Fe}^{3+}\text{-ZIF-90}$ particles: top—experimental results; bottom—spectral bands of ZIF-90 from the PDF database. (D) Bright field and fluorescence confocal microscopy images corresponding to the Atto 565-labeled AChE/FITC-labeled ChOx loaded in the $\text{Fe}^{3+}\text{-ZIF-90}$: panel I—bright field image; panel II—imaging the Atto 565-labeled AChE ($\lambda_{\text{ex}} = 561 \text{ nm}$, $\lambda_{\text{em}} = 585 \text{ nm}$); panel III—imaging FITC-labeled ChOx ($\lambda_{\text{ex}} = 488 \text{ nm}$, $\lambda_{\text{em}} = 525 \text{ nm}$); panel IV—overlapped image. (E) Time-dependent concentration changes of TMB^{++} upon operating the acetylcholine-driving three-catalyst cascade using: (i) the AChE/ChOx-loaded $\text{Fe}^{3+}\text{-ZIF-90}$ MOFs as an integrated assembly; (ii) the separated AChE, ChOx and $\text{Fe}^{3+}\text{-ZIF-90}$ constituents in the homogeneous aqueous phase at concentrations identical to those in (i). (F) Panel I—time-dependent concentration changes of TMB^{++} upon operating the AChE/ChOx/ $\text{Fe}^{3+}\text{-ZIF-90}$ cascade in the presence of variable concentrations of acetylcholine: (i) 0 mM; (ii) 0.1 mM; (iii) 0.2 mM; (iv) 0.5 mM; (v) 1 mM; (vi) 2 mM; (vii) 5 mM; (viii) 10 mM. Panel II—rates of TMB^{++} formation by the AChE/ChOx/ $\text{Fe}^{3+}\text{-ZIF-90}$ system in the presence of variable concentrations of acetylcholine.



paralysis and cognition.⁶⁵ Perturbations of the levels of acetylcholine were reported to be associated with Alzheimer's and Parkinson's diseases as well as paranoid schizophrenia.^{66–68} Moreover, blockage of the cholinergic neurons or inhibiting AChE by organophosphorus pesticides or chemical warfare toxins may lead to severe neuronal disorders. Indeed, monitoring the levels of acetylcholine in plasma or the activity of AChE in the presence of potential inhibitors is an important analytical issue. Different analytical tools, such as mass spectrometry,⁶⁹ high-performance liquid chromatography⁷⁰ and radiolabeling,⁷¹ were employed to quantitatively detect acetylcholine. Also, diverse optical, electrochemical and enzymatic biosensing platforms following AChE activity were developed.^{72,73} For example, optical sensing platforms include surface plasma resonance,⁷⁴ chemiluminescence,⁷⁵ colorimetric assays⁷⁶ and fluorescence sensing assays using semiconductor quantum dots.⁷⁷ Also, electrochemical sensing platforms including amperometric biosensor devices,⁷⁸ field-effect^{79,80} transistors and photoelectrochemical sensors⁸¹ following AChE activities and AChE inhibitors were reported. However, the present sensing platforms suffer from limited sensitivity, lack of desirable integration and circuit complexity. Thus, the integration of the sensing platform as a cascaded circuit in the MOF systems could provide a sensitive sensing platform due to the integrated nature of the system in the confined microenvironment. Fig. 4A depicts schematically the configuration of the three-catalyst cascaded frameworks. The two enzymes, AChE and ChOx, are integrated into the Fe³⁺-ZIF-90 MOFs. The AChE catalyzes the hydrolysis of acetylcholine, yielding acetic acid and choline as products. The aerobic oxidation of choline catalyzed by ChOx generates betaine and H₂O₂. The subsequent Fe³⁺-ZIF-90-catalyzed H₂O₂ oxidation of TMB to colored TMB⁺, provides an optical means to follow the catalytic cascade. The inhibition of AChE is, then, anticipated to affect the catalytic cascade. Evidently, appropriate engineering of the three-catalyst component, the structural and functional characterization of the framework and its response to inhibiting agents are essential for the assembly of the sensing platform. Accordingly, a stepwise process to assemble the sensor was adopted. In the first step, the single enzyme, ChOx, was integrated into the Fe³⁺-ZIF-90 MOFs. The ChOx-loaded particles and the accompanying aerobic oxidation of choline were characterized. Fig. S11† summarizes the structural and functional features of the ChOx-loaded Fe³⁺-ZIF-90 system. Evidently, the morphology and crystallinity of the Fe³⁺-ZIF-90 frameworks are preserved upon incorporation of ChOx. Also, the ChOx integrated into the composite demonstrates choline concentration-dependent activity. In the next step, the two enzymes, AChE and ChOx, were integrated into the Fe³⁺-ZIF-90 matrices, as shown in Fig. 4A. The morphology and crystallinity of the bienzyme-loaded particles were similar to those of the bare unloaded particles, as shown in Fig. 4B and C. The loading of the Fe³⁺-ZIF-90 particles by the two enzymes and the quantitative loading degree were evaluated by labeling the two enzymes with two different fluorophores and by complementary application of fluorescence confocal microscopy. ChOx was labeled with FITC ($\lambda_{\text{ex}} = 488 \text{ nm}$, $\lambda_{\text{em}} = 525 \text{ nm}$) and AChE was modified with Atto

565 ($\lambda_{\text{ex}} = 561 \text{ nm}$, $\lambda_{\text{em}} = 585 \text{ nm}$). Fig. 4D depicts the bright field and fluorescence confocal microscopy images using the different Atto 565 and FITC fluorescence channels and the merged image, panels I–IV. Evidently, the two enzymes were encapsulated in the Fe³⁺-ZIF-90 particles. For quantitative evaluation of the loading degree by the two fluorophore-modified enzymes in the frameworks, the residual fluorescence intensities of FITC–ChOx and Atto 565–AChE in the bulk solution after encapsulation were recorded. Knowing the original concentrations of the enzymes prior encapsulation and using appropriate calibration curves relating the concentrations of the fluorophore-modified enzymes and their associated fluorescence intensities, the contents of the Atto 565–AChE and FITC–ChOx in the MOF particles were evaluated to be 81.5 $\mu\text{g mg}^{-1}$ particles and 86.8 $\mu\text{g mg}^{-1}$ particles, respectively (see Fig. S12 and S13†). Moreover, the loading of the two enzymes in the Fe³⁺-ZIF-90 particles was confirmed by operating the three-catalyst cascade, as shown in Fig. 4E, curve (i), depicting the time-dependent concentration changes upon formation of TMB⁺ by the system. The control experiment, excluding ChOx and AChE from the particles, prohibited the formation of TMB⁺, confirming that the bienzyme-activated cascade is, indeed, operating. Furthermore, Fig. 4E, curve (ii) depicts the temporal formation of TMB⁺ in a diffusional mixture composed of separated AChE, ChOx and Fe³⁺-ZIF-90 particles (at identical concentrations to those of the integrated AChE/ChOx/Fe³⁺-ZIF-90 particles). The cascaded generation of TMB⁺ by the integrated assembly is *ca.* 5-fold enhanced as compared to the diffusional catalyst mixture, demonstrating the advantages of operating the catalytic cascade in the confined environment of the particles. Fig. 4F presents the time-dependent concentration changes of TMB⁺ generated by the integrated three-catalyst reactor in the presence of variable concentrations of acetylcholine. As the concentration of acetylcholine increases, the catalytic cascade is enhanced (Fig. 4F, panel I). Using the extinction coefficient of TMB⁺ ($\epsilon = 39\,000 \text{ M}^{-1} \text{ cm}^{-1}$), the rates of TMB⁺ formation at different acetylcholine concentrations were evaluated (Fig. 4F, panel II), demonstrating the capacity of the AChE/ChOx/Fe³⁺-ZIF-90 to act as an optical platform for sensing acetylcholine. The detection limit for analyzing acetylcholine was determined to be 78 μM (for comparison of the acetylcholine sensing capacity by the AChE/ChOx/Fe³⁺-ZIF-90 framework, as compared to other sensing platforms, see Table S1†).

The results demonstrate that the integrated AChE/ChOx/Fe³⁺-ZIF-90 system provides an effective sensing platform for probing acetylcholine quantitatively. Furthermore, we find that the sensing capacity of the integrated framework is affected by the concentration of AChE in the frameworks, as shown in Fig. S14.† This suggests that the sensing performance of the AChE/ChOx/Fe³⁺-ZIF-90 frameworks will be affected by AChE inhibiting agents (*e.g.*, chemical warfare agents). Accordingly, the effect of the 1,5-bis(4-allyldimethylammoniumphenyl)pentane-3-one dibromide (BW284C51) inhibitor,⁸² a well-established inhibitor of AChE and a model for the chemical warfare agent, on the sensing performance of the three-catalyst cascade was examined. Fig. 5A presents the time-dependent concentration changes of the generated



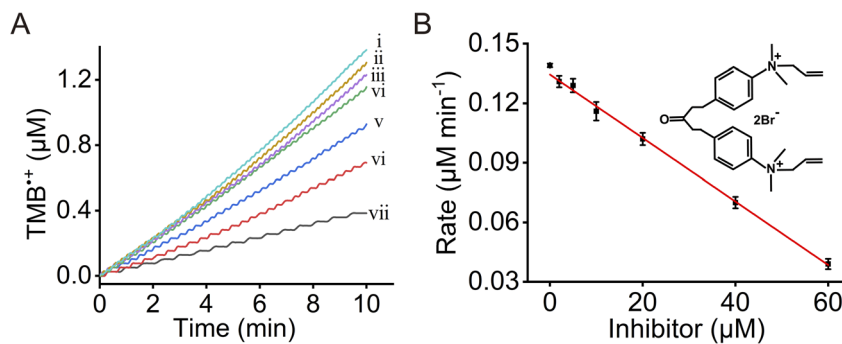


Fig. 5 (A) Time-dependent concentration changes of TMB^{•+} generated by the AChE/ChOx/Fe³⁺-ZIF-90 particles in the presence of acetylcholine, 10 mM, and variable concentrations of the 1,5-bis(4-allyldimethylammoniumphenyl)pentane-3-one dibromide (BW284C51) inhibitor: (i) 0 μM; (ii) 2 μM; (iii) 5 μM; (iv) 10 μM; (v) 20 μM; (vi) 40 μM; (vii) 60 μM. (B) Rates of TMB^{•+} formation by the AChE/ChOx/Fe³⁺-ZIF-90 MOFs in the presence of variable concentrations of the BW284C51 inhibitor.

TMB^{•+}. As the concentration of the BW284C51 inhibitor increases, the activity of the AChE/ChOx/Fe³⁺-ZIF-90 catalytic cascade decreases. Fig. 5B depicts the rates of TMB^{•+} formation catalyzed by the AChE/ChOx/Fe³⁺-ZIF-90 sensing platform in the presence of different concentrations of the BW284C51 inhibitor. The inhibitor could be sensed by the catalytic cascade with a detection limit of 1.69 μM.

Conclusions

The present study has introduced Fe³⁺-doped ZIF-90 particles as functional frameworks for the assembly of biocatalyst/nanozyme composites acting as nanoreactors for the operation of catalytic cascades. The important functional features of the nanoreactor were as follows: (i) the Fe³⁺-ZIF-90 particles revealed peroxidase-like activities. (ii) The porous structure of the particles and their synthesis in an aqueous environment at room temperature enabled the high-loading encapsulation of multienzyme systems in the particles without affecting their activities. (iii) Coupling of the enzyme cascades with the peroxidase-like activities of the frameworks enabled the operation of three-catalyst cascades. (iv) The advantages of operating the enzymes in the confined porous frameworks, overcoming diffusional barriers present in the mixture of separated constituents of the catalytic cascades, were demonstrated. These included enhanced catalytic cascades in the confined particles as compared to the analogous diffusional assemblies. (v) The operation of the AChE/ChOx/Fe³⁺-ZIF-90 three-catalyst cascade was applied to develop a sensing framework monitoring agents inhibiting AChE, thereby providing a detection platform for the identification of hazardous chemical warfare. The concept of encapsulation of enzymes into the metal ion-doped ZIF to yield catalytic nanozyme-based framework systems can be extended to many other enzymes and other metal-ion dopants of the particles. Moreover, the ZIF-90 includes carboxaldehyde functionalities that could provide anchoring sites for chemical functionalities such as aptamers or antibodies. Such hybrid systems are envisaged to provide superior systems for sensing platforms and therapeutic applications.

Data availability

The data supporting this article have been included as part of the ESI.†

Author contributions

I. Willner and X. Chen conceived and designed the experiments. J. Wang carried out the synthesis and catalytic activity test of the materials. Y. Qin, R. Carmieli and V. Gutkin contributed to the characterization studies. Z. Zhang and E. Pikarsky participated in the analysis of the results. I. Willner, J. Wang and X. Chen designed the figures and co-wrote the manuscript. All authors reviewed and approved the final manuscript.

Conflicts of interest

There are no conflicts to declare.

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References

- 1 M. Yoon, R. Srirambalaji and K. Kim, *Chem. Rev.*, 2011, **112**, 1196–1231.
- 2 J. Liu, L. Chen, H. Cui, J. Zhang, L. Zhang and C.-Y. Su, *Chem. Soc. Rev.*, 2014, **43**, 6011–6061.
- 3 L.-Z. Cai, Z.-Z. Yao, S.-J. Lin, M.-S. Wang and G.-C. Guo, *Angew. Chem., Int. Ed.*, 2021, **60**, 18223–18230.
- 4 M. Y. Gao, B. Q. Song, D. Sensharma and M. J. Zaworotko, *SmartMat*, 2020, **2**, 38–55.
- 5 D. Alezi, Y. Belmabkhout, M. Suyetin, P. M. Bhatt, Ł. J. Weseliński, V. Solovyeva, K. Adil, I. Spanopoulos, P. N. Trikalitis, A.-H. Emwas and M. Eddaoudi, *J. Am. Chem. Soc.*, 2015, **137**, 13308–13318.
- 6 H. Daglar, H. C. Gulbalkan, G. Avci, G. O. Aksu, O. F. Altundal, C. Altintas, I. Erucar and S. Keskin, *Angew. Chem., Int. Ed.*, 2021, **60**, 7828–7837.



- 7 H. Furukawa, F. Gándara, Y.-B. Zhang, J. Jiang, W. L. Queen, M. R. Hudson and O. M. Yaghi, *J. Am. Chem. Soc.*, 2014, **136**, 4369–4381.
- 8 J. Canivet, A. Fateeva, Y. Guo, B. Coasne and D. Farrusseng, *Chem. Soc. Rev.*, 2014, **43**, 5594–5617.
- 9 Y. Han, P. Das, Y. He, D. C. Sorescu, K. D. Jordan and N. L. Rosi, *J. Am. Chem. Soc.*, 2022, **144**, 19567–19575.
- 10 L. Jiao and H.-L. Jiang, *Chem*, 2019, **5**, 786–804.
- 11 P.-L. Wang, L.-H. Xie, E. A. Joseph, J.-R. Li, X.-O. Su and H.-C. Zhou, *Chem. Rev.*, 2019, **119**, 10638–10690.
- 12 P. S. Sharanyakanth and M. Radhakrishnan, *Trends Food Sci. Technol.*, 2020, **104**, 102–116.
- 13 X.-J. Kong and J.-R. Li, *Engineering*, 2021, **7**, 1115–1139.
- 14 T. He, X.-J. Kong, Z.-X. Bian, Y.-Z. Zhang, G.-R. Si, L.-H. Xie, X.-Q. Wu, H. Huang, Z. Chang, X.-H. Bu, M. J. Zaworotko, Z.-R. Nie and J.-R. Li, *Nat. Mater.*, 2022, **21**, 689–695.
- 15 L. E. Kreno, K. Leong, O. K. Farha, M. Allendorf, R. P. Van Duyne and J. T. Hupp, *Chem. Rev.*, 2011, **112**, 1105–1125.
- 16 D. Liu, R. C. Huxford and W. Lin, *Angew. Chem., Int. Ed.*, 2011, **50**, 3696–3700.
- 17 J. Della Rocca, D. Liu and W. Lin, *Acc. Chem. Res.*, 2011, **44**, 957–968.
- 18 K. M. L. Taylor, A. Jin and W. Lin, *Angew. Chem., Int. Ed.*, 2008, **47**, 7722–7725.
- 19 W. H. Chen, W. C. Liao, Y. S. Sohn, M. Fadeev, A. Cecconello, R. Nechushtai and I. Willner, *Adv. Funct. Mater.*, 2017, **28**, 1705137.
- 20 W.-H. Chen, X. Yu, A. Cecconello, Y. S. Sohn, R. Nechushtai and I. Willner, *Chem. Sci.*, 2017, **8**, 5769–5780.
- 21 W. H. Chen, X. Yu, W. C. Liao, Y. S. Sohn, A. Cecconello, A. Kozell, R. Nechushtai and I. Willner, *Adv. Funct. Mater.*, 2017, **27**, 1702102.
- 22 P. Horcajada, R. Gref, T. Baati, P. K. Allan, G. Maurin, P. Couvreur, G. Férey, R. E. Morris and C. Serre, *Chem. Rev.*, 2011, **112**, 1232–1268.
- 23 J. N. Hall and P. Bollini, *ACS Catal.*, 2020, **10**, 3750–3763.
- 24 R. R. Kuruppathparambil, T. Jose, R. Babu, G.-Y. Hwang, A. C. Kathalikkattil, D.-W. Kim and D.-W. Park, *Appl. Catal., B*, 2016, **182**, 562–569.
- 25 W. H. Chen, M. Vázquez-González, A. Kozell, A. Cecconello and I. Willner, *Small*, 2017, **14**, 1703149.
- 26 F. Ghobakhloo, D. Azarifar, M. Mohammadi, H. Keypour and H. Zeynali, *Inorg. Chem.*, 2022, **61**, 4825–4841.
- 27 L. Chen, S. Rangan, J. Li, H. Jiang and Y. Li, *Green Chem.*, 2014, **16**, 3978–3985.
- 28 L. Li, Z. Li, W. Yang, Y. Huang, G. Huang, Q. Guan, Y. Dong, J. Lu, S.-H. Yu and H.-L. Jiang, *Chem*, 2021, **7**, 686–698.
- 29 H. Wang, X. Liu, W. Yang, G. Mao, Z. Meng, Z. Wu and H.-L. Jiang, *J. Am. Chem. Soc.*, 2022, **144**, 22008–22017.
- 30 M. Eddaoudi, D. F. Sava, J. F. Eubank, K. Adil and V. Guillermin, *Chem. Soc. Rev.*, 2015, **44**, 228–249.
- 31 J. Yao and H. Wang, *Chem. Soc. Rev.*, 2014, **43**, 4470–4493.
- 32 S. Liang, X.-L. Wu, J. Xiong, M.-H. Zong and W.-Y. Lou, *Coord. Chem. Rev.*, 2020, **406**, 213149.
- 33 W.-H. Chen, M. Vázquez-González, A. Zoabi, R. Abu-Reziq and I. Willner, *Nat. Catal.*, 2018, **1**, 689–695.
- 34 E. Gkaniatsou, C. Sicard, R. Ricoux, L. Benahmed, F. Bourdreux, Q. Zhang, C. Serre, J. P. Mahy and N. Steunou, *Angew. Chem., Int. Ed.*, 2018, **57**, 16141–16146.
- 35 V. Gascón, C. Carucci, M. B. Jiménez, R. M. Blanco, M. Sánchez-Sánchez and E. Magner, *ChemCatChem*, 2017, **9**, 1182–1186.
- 36 Q. Zhang, J. Liang, A. Bongers, J. J. Richardson, K. Liang and Z. Gu, *Adv. Sci.*, 2023, **10**, 2206546.
- 37 W.-H. Chen, G.-F. Luo, M. Vázquez-González, R. Cazelles, Y. S. Sohn, R. Nechushtai, Y. Mandel and I. Willner, *ACS Nano*, 2018, **12**, 7538–7545.
- 38 F. Peng, Y. Xiang, H. Wang, Y. Hu, R. Zhou and Y. Hu, *Small*, 2022, **18**, 2204011.
- 39 A. Poddar, J. J. Conesa, K. Liang, S. Dhakal, P. Reineck, G. Bryant, E. Pereiro, R. Ricco, H. Amenitsch, C. Doonan, X. Mulet, C. M. Doherty, P. Falcato and R. Shukla, *Small*, 2019, **15**, 1902268.
- 40 K. Wei, M. He, J. Zhang, C. Zhao, C. Nie, T. Zhang, Y. Liu, T. Chen, J. Jiang and X. Chu, *Angew. Chem., Int. Ed.*, 2023, **62**, e202307025.
- 41 T. Man, C. Xu, X.-Y. Liu, D. Li, C.-K. Tsung, H. Pei, Y. Wan and L. Li, *Nat. Commun.*, 2022, **13**, 305.
- 42 C. Duan, Y. Yu and H. Hu, *Green Energy Environ.*, 2022, **7**, 3–15.
- 43 Y. Wu, Y. Qin, M. Muppudathi, R. Carmieli, M. Fadeev, W. Lei, M. Xia and I. Willner, *Adv. Funct. Mater.*, 2023, **34**, 2306929.
- 44 Y. Zhong, P. Liao, J. Kang, Q. Liu, S. Wang, S. Li, X. Liu and G. Li, *J. Am. Chem. Soc.*, 2023, **145**, 4659–4666.
- 45 N. Nagarjun and A. Dhakshinamoorthy, *New J. Chem.*, 2019, **43**, 18702–18712.
- 46 C. Duan, Y. Feng, Y. Xie, M. Ding and J. Yao, *Microporous Mesoporous Mater.*, 2023, **347**, 112351.
- 47 C. Chen, M. Vázquez-González, M. P. O'Hagan, Y. Ouyang, Z. Wang and I. Willner, *Small*, 2022, **18**, 2104420.
- 48 W. Liang, K. Flint, Y. Yao, J. Wu, L. Wang, C. Doonan and J. Huang, *J. Am. Chem. Soc.*, 2023, **145**, 20365–20374.
- 49 J. Guo, L. Yang, Z. Gao, C. Zhao, Y. Mei and Y.-Y. Song, *ACS Catal.*, 2020, **10**, 5949–5958.
- 50 Y.-M. Li, J. Yuan, H. Ren, C.-Y. Ji, Y. Tao, Y. Wu, L.-Y. Chou, Y.-B. Zhang and L. Cheng, *J. Am. Chem. Soc.*, 2021, **143**, 15378–15390.
- 51 J. Qin, M. Cho and Y. Lee, *ACS Appl. Mater. Interfaces*, 2019, **11**, 11743–11748.
- 52 M. Kim, M. J. Han, H. Lee, P. Flouda, D. Bukharina, K. J. Pierce, K. M. Adstedt, M. L. Buxton, Y. H. Yoon, W. T. Heller, S. Singamaneni and V. V. Tsukruk, *Angew. Chem., Int. Ed.*, 2023, **62**, e202305646.
- 53 B. Liu, F. Hu, J. Zhang, C. Wang and L. Li, *Angew. Chem., Int. Ed.*, 2019, **58**, 8804–8808.
- 54 J. Shen, M. Ma, M. Shafiq, H. Yu, Z. Lan and H. Chen, *Angew. Chem., Int. Ed.*, 2022, **61**, e202113703.
- 55 Y. Qin, Y. Ouyang, J. Wang, X. Chen, Y. S. Sohn and I. Willner, *Nano Lett.*, 2023, **23**, 8664–8673.
- 56 P. D. Josephy, T. Eling and R. P. Mason, *J. Biol. Chem.*, 1982, **257**, 3669–3675.



- 57 M. Vázquez-González, C. Wang and I. Willner, *Nat. Catal.*, 2020, **3**, 256–273.
- 58 O. I. Wilner, Y. Weizmann, R. Gill, O. Lioubashevski, R. Freeman and I. Willner, *Nat. Nanotechnol.*, 2009, **4**, 249–254.
- 59 O. I. Wilner, S. Shimron, Y. Weizmann, Z.-G. Wang and I. Willner, *Nano Lett.*, 2009, **9**, 2040–2043.
- 60 J. Fu, M. Liu, Y. Liu, N. W. Woodbury and H. Yan, *J. Am. Chem. Soc.*, 2012, **134**, 5516–5519.
- 61 T. A. Ngo, E. Nakata, M. Saimura and T. Morii, *J. Am. Chem. Soc.*, 2016, **138**, 3012–3021.
- 62 Y. Elani, R. V. Law and O. Ces, *Nat. Commun.*, 2014, **5**, 5305.
- 63 H. Tan, S. Guo, N.-D. Dinh, R. Luo, L. Jin and C.-H. Chen, *Nat. Commun.*, 2017, **8**, 663.
- 64 D. M. Vriezema, P. M. L. Garcia, N. S. Oltra, N. S. Hatzakis, S. M. Kuiper, R. J. M. Nolte, A. E. Rowan and J. C. M. van Hest, *Angew. Chem., Int. Ed.*, 2007, **46**, 7378–7382.
- 65 Ł. J. Walczak-Nowicka and M. Herbet, *Int. J. Mol. Sci.*, 2021, **22**, 9290.
- 66 H. Shinotoh, H. Namba, K. Fukushi, S.-I. Nagatsuka, N. Tanaka, A. Aotsuka, T. Ota, S. Tanada and T. Irie, *Ann. Neurol.*, 2000, **48**, 194–200.
- 67 X. Zhang, L. Lu, S. Liu, W. Ye, J. Wu and X. Zhang, *Int. J. Biochem. Cell Biol.*, 2013, **45**, 265–272.
- 68 N. C. Inestrosa, A. Alvarez, C. A. Pérez, R. D. Moreno, M. Vicente, C. Linker, O. I. Casanueva, C. Soto and J. Garrido, *Neuron*, 1996, **16**, 881–891.
- 69 W. Lee, N. H. Park, Y. C. Lee, K.-H. Kim and J. Hong, *TrAC, Trends Anal. Chem.*, 2018, **106**, 159–168.
- 70 C. Eva, M. Hadjiconstantinou, N. H. Neff and J. L. Meek, *Anal. Biochem.*, 1984, **143**, 320–324.
- 71 K. Kawashima, T. Hayakawa, Y. Kashima, T. Suzuki, K. Fujimoto and H. Oohata, *J. Neurochem.*, 2006, **57**, 882–887.
- 72 A. V. Alex and A. Mukherjee, *Microchem. J.*, 2021, **161**, 105779.
- 73 J. Ahlawat, M. Sharma and C. S. Pundir, *Microchem. J.*, 2023, **190**, 108620.
- 74 M. Zayats, A. B. Kharitonov, S. P. Pogorelova, O. Lioubashevski, E. Katz and I. Willner, *J. Am. Chem. Soc.*, 2003, **125**, 16006–16014.
- 75 N. Xia, Q. Wang and L. Liu, *Sensors*, 2014, **15**, 499–514.
- 76 D.-M. Liu, B. Xu and C. Dong, *TrAC, Trends Anal. Chem.*, 2021, **142**, 116320.
- 77 R. Gill, L. Bahshi, R. Freeman and I. Willner, *Angew. Chem., Int. Ed.*, 2008, **47**, 1676–1679.
- 78 S. Shakil, D. Yuan and M. Li, *J. Electrochem. Soc.*, 2024, **171**, 067512.
- 79 M.-S. Chae, Y. K. Yoo, J. Kim, T. G. Kim and K. S. Hwang, *Sens. Actuators, B*, 2018, **272**, 448–458.
- 80 L. H. Hess, A. Lyuleeva, B. M. Blaschke, M. Sachsenhauser, M. Seifert, J. A. Garrido and F. Deubel, *ACS Appl. Mater. Interfaces*, 2014, **6**, 9705–9710.
- 81 V. Pardo-Yissar, E. Katz, J. Wasserman and I. Willner, *J. Am. Chem. Soc.*, 2002, **125**, 622–623.
- 82 S. Olivera-Bravo, I. Ivorra and A. Morales, *Br. J. Pharmacol.*, 2009, **144**, 88–97.

