


 Cite this: *Nanoscale*, 2024, **16**, 14953

One- and two-photon brightness of proteins interacting with gold. A closer look at gold–insulin conjugates†

 Dipankar Bain,^a Hao Yuan,^a Anna Priakowska,^b Agata Hajda,^b Charlene Bouanchaud,^a Fabien Chiot,^a Clothilde Comby-Zerbino,^a Virginie Gueguen-Chaignon,^c Vlasta Bonačić-Koutecký,^d Joanna Olesiak-Banska,^b Željka Sanader Maršić^b *^{d,e} and Rodolphe Antoine^b *^a

Red luminophores displaying large Stokes shift and high-quantum yields are obtained when gold salts are reacted with proteins under strongly alkaline conditions. Although bovine serum albumin (BSA) has mainly been used as a protein template, other attempts to prepare red luminophores have been proposed using other proteins. Here, we report on the structural characterization and nonlinear optical properties of insulin–gold conjugates. Such conjugates display strong luminescence at ~670 nm with quantum yields that reach 5.4%. They also display long luminescence lifetimes allowing efficient reactive oxygen species generation, with a quantum yield of ¹O₂ generation reaching 13%. In addition, they exhibit remarkable nonlinear optical properties and in particular a strong two-photon excited fluorescence (TPEF) cross section in the range of 800–1100 nm. By combining experimental studies and time-dependent density functional theory simulations (TD–DFT), we show the formation of insulin–Au(III) conjugates. The interaction of Au(III) ions with the aromatic rings of tyrosine induces charge transfer-like excitation in the visible range. Experimental investigations, together with molecular dynamics simulations of insulin and calculations of electronic properties in a model system, are performed to explore the origin of optical features and the structure–optical property relationship, leading the way to new concepts for nonlinear optics using protein–Au(III) conjugates.

 Received 18th April 2024,
 Accepted 11th July 2024
 DOI: 10.1039/d4nr01697a

rsc.li/nanoscale

Introduction

When gold salts are reacted with proteins at a basic pH, the reaction products exhibit large Stokes shift, red luminescence, and high quantum yields.^{1–8} In particular, many applications using mainly bovine serum albumin (BSA) as protein tem-

plates and gold have been reported as diagnostic and therapeutic platforms for biomedical applications.^{9,10}

Exemplified by BSA, two possible scenarios for this luminescence have been suggested: one scenario considers that BSA acts as a reacting and protecting “cage” in which gold ions are reduced by tyrosine residues under alkaline conditions and nucleate into a gold nanocluster, composed of dozens of gold atoms.¹¹ The other scenario asserts that the red luminophore formation involves Au(III) coordination in a specific domain of serum albumin.^{12,13} These observations and protein structural analyses with various proteins (bovine serum albumin, ovalbumin, trypsin, and insulin) show that red luminophore formation involves Au(III) coordination with a common motif within these proteins.¹⁴

Insulin is a small protein, or more precisely a polypeptide hormone comprising only 51 amino acids. Its function primarily lies in the regulation of insulin-responsive tissues and it is related to many diseases. Insulin has no free cysteine and only 4 tyrosine residues, so it can display unique binding motifs with gold and thus be a good candidate for producing red luminophores. Attempts to characterize prepared insulin–gold

^aInstitut lumière matière, UMR5306, Université Claude Bernard Lyon1-CNRS, Univ. Lyon 69622, Villeurbanne cedex, France. E-mail: rodolphe.antoine@univ-lyon1.fr

^bInstitute of Advanced Materials, Wrocław University of Science and Technology, Wybrzeże Wyspiańskiego 27, 50-370 Wrocław, Poland

^cUniversité Claude Bernard Lyon 1, CNRS UAR3444, Inserm US8, ENS de Lyon, SFR Biosciences, Lyon 69007, France

^dCenter of Excellence for Science and Technology-Integration of Mediterranean Region (STIM) at Interdisciplinary Center for Advanced Sciences and Technology (ICAST), University of Split, Poljička cesta 35, 21000 Split, Croatia

^eFaculty of Science, University of Split, Ruđera Boškovića 33, 21000 Split, Croatia. E-mail: zsm@pmfst.hr

† Electronic supplementary information (ESI) available: Experimental. Synthesis and purification. XPS spectra. ESI- and MALDI-MS spectra. DLS and CD spectra and analysis. PAGE gels. Optical spectra and ROS measurements. TD-DFT results. See DOI: <https://doi.org/10.1039/d4nr01697a>



samples by mass spectrometry have proved unsuccessful, emphasizing the difficulty of exploring the structure–optical property relationship taking place during the interaction of proteins with gold.^{5,15}

In this paper, we synthesize water-soluble insulin–gold conjugates. The goal of the current preparation was to develop highly luminescent gold–insulin conjugates. Although insulin functionalized nanoclusters were reported earlier, their preparation was time consuming whereas the current synthesis process is fast and completed within 4 hours. Moreover, apart from the new preparations, the mass spectrometric characterization¹⁶ and quantitative analysis of two-photon fluorescence properties¹⁷ are new findings which have also been correlated with theoretical calculations. In particular, our quantitative analysis of two-photon properties will be very attractive for two-photon bioimaging applications.

Results and discussion

Synthesis and characterization of insulin–gold conjugate

Here, we synthesized highly luminescent insulin–gold conjugates using a one pot “green” synthesis method with a modification of the protocol reported previously.⁵ The reaction was carried out at physiological temperature (37 °C) at elevated pH (~11.5), as described in detail in the Methods (see the ESI Fig. S1 and S2†). The pH of the insulin solution was in the range of pH ~3.5–4.0. Insulin has low solubility in water, thus when we added insulin to the Au³⁺ solution, the latter transformed into yellowish white aggregates. Quantitative XPS measurements were compatible with one gold and one insulin stoichiometry in gold–insulin conjugates (see Table S1 and Fig. S3†). We also tried to use XPS to investigate the valence state of the gold atoms in gold–insulin conjugates. A doublet peak structure should be observed with fitted binding energies of ~87.4 and ~91.4 eV for Au(III) conjugates (respectively ⁴f_{7/2} and ⁴f_{5/2} doublet peaks). However, the ⁴f_{5/2} peak of Au(0) was also located at 87.6 eV.¹⁸ The facts that the content of gold (0.3%) is very low and there is overlap of 4f peaks between different oxidation states of gold led to a lack of clear evidence of Au(III) by XPS, while mass spectrometry measurements led to an unambiguous assignment (see below). As already mentioned in the Introduction, early attempts to measure the mass of the as prepared insulin–Au NCs were either unsuccessful⁵ or only one gold atom attached to insulin was detected in mass spectra.¹⁵ We continued the mass characterization of insulin–gold using high-resolution electrospray mass spectrometry (HR-ESI-MS). The full ESI-MS spectrum of the insulin–gold conjugates in ESI is shown in Fig. S4,† where charge states from 2+ to 4+ can be observed. For insulin, charge states *n*+ are given by extra protonations [Insu + *n*H]^{*n*+}. The isotopic pattern of insulin (4+ charge state, [Insu + 4H]⁴⁺) is reproduced using the following formula: C₂₅₇H₃₈₃N₆₅O₇₇S₆ (see insets in Fig. 1). When Au is complexed with insulin (4+ charge state), thanks to the isotopic distribution analysis, we observe that 3 hydrogen ions are removed from insulin. Then



Fig. 1 Zoom of the variable temperature-nano electrospray ionization (VT-nano-ESI) MS spectrum of insulin–gold conjugate, corresponding to the 4+ charge state (Insu for Insulin). Insets. Isotopic pattern (black: experimental, and red: simulation) of insulin (C₂₅₇H₃₈₃N₆₅O₇₇S₆) on the left and of insulin with Au(III) (C₂₅₇H₃₈₀N₆₅O₇₇S₆Au) on the right. The other minor peaks are mostly alkali adducts.

the isotopic pattern of insulin is reproduced using the following formula: C₂₅₇H₃₈₀N₆₅O₇₇S₆Au (see insets in Fig. 1). The number of H atoms is reduced from 383 to 380 assigned to the 3 hydrogen ions removed from the insulin. At the pH value of 11.5 used for the synthesis, phenolic groups of tyrosine are deprotonated. One can thus assume that deprotonations on insulin occur on tyrosine residues.

The interaction between insulin and gold was also investigated using circular dichroism (CD) spectroscopy. Fig. S5† shows the CD spectra of pure insulin and insulin–Au conjugates. The pure insulin in neutral pH showed two negative absorption bands at ~211 and ~224 nm (in fair agreement with the reported CD spectrum of human insulin).¹⁹ When Au was added to insulin, only the first negative band at ~209 nm remained, and a broad negative band emerged at ~350 nm. The popular method for estimating protein secondary structure fractions from CD spectra (implemented in the software package CDSSTR)²⁰ was tested on insulin and insulin–Au conjugates. The results presented in Fig. S6† indicate that the native structure of insulin is not much affected by the presence of gold (Table S2†). Only the local alpha-helix environment is affected (where 3 of the 4 tyrosine residues are in helix motifs, see Fig. 3a). It is noteworthy that crystallization tests were carried out, but no crystals were formed in the presence of gold.

Although the presence of one gold atom bound to insulin was demonstrated by HR-ESI-MS, we cannot rule out the formation of oligomers (as expected since insulin is known to easily form multimers). Indeed, in a previous study, we used a simple and reliable fluorescence-based technique to measure the hydrodynamic size of monolayer protected gold nanoclusters.^{8,21} This technique uses the relation between the time resolved fluorescence anisotropy decay and the hydrodynamic volume. The comparison of sizes obtained between bare proteins and gold containing proteins allowed us to address



the volume changes induced by the formation of oligomers for insulin during gold incorporation.⁸ Interestingly, Dixon and Egusa further evidenced the formation of oligomers (approximately between dimers and tetramers) for insulin during gold incorporation, by performing native gel electrophoresis of protein–Au(III) conjugates.¹⁴ We performed native gel electrophoresis on our as-synthesized insulin–gold conjugates, also evidencing the tendency towards oligomerization during gold incorporation (see Fig. S7†). Oligomer formation was further observed by mass spectrometry (both MALDI²² and ESI-MS, see Fig. S8†). Interestingly, the VT-nano-ESI source was used to fix the sample temperature at 17 °C, thereby slowing down the aggregation.²³ In parallel, we performed DLS analysis on insulin and insulin–gold conjugates, see Fig. S9.† As can be seen, dynamic light scattering shows that the mean sizes of insulin and insulin–Au conjugates are 3.8 nm and 7 nm, respectively. Thus, the presence of gold in the sample affects insulin oligomerization. To sum up, the as-synthesized insulin–gold conjugates are mainly composed of insulin–Au(III) complex resulting in oligomers (ranging from monomers to trimers).

Optical properties of the insulin–gold conjugate

Linear optical properties. We recorded the absorption and photoluminescence spectra of insulin–gold conjugates, as shown in Fig. S10.† The as-synthesized insulin–Au conjugates show an absorption hump around 510 nm. The excitation spectrum exhibits a similar band corresponding to an absorption band at 510 nm which indicates that the PL band is excited by the HOMO–LUMO transition. We observed an intense emission peak at 667 nm. The emission spectra of Au–insulin only changed in intensity when the conjugates were irradiated at different wavelengths. The Stokes shift can reach at maximum 300 nm (see Fig. S11†). We recorded the luminescence quantum yield (QYs) using the integrating sphere method and found it to be 5.40%, with observed lifetimes fitted at 237 ns and 1723 ns (Fig. S12†).

Insulin–gold conjugates exhibit a long photoluminescence lifetime, suggesting that they can be capable of singlet oxygen generation,^{24–28} as demonstrated for other protein template gold systems.^{26,27} As a typical mechanism of singlet oxygen generation, after photoexcitation to an excited singlet state, the photosensitizer undergoes intersystem crossing to its triplet state, followed by energy transfer to ground-state (triplet) dioxygen (O₂). Generally, excited triplet states are relatively long-lived compared with excited singlet states because of the spin-forbidden nature of the T₁ → S₀ transition. Here, the generation of singlet oxygen was triggered by the excitation of the conjugates with a cw laser emitting at 473 nm with different exposure times. With 1,3-diphenylisobenzofuran (DPBF, an absorption probe for detection of ¹O₂), ¹O₂ generation can be monitored by a change in the absorption of DPBF at 412 nm. The absorption spectra and the change in the absorption of DPBF monitored over time are presented in Fig. S13a).† The rate of ¹O₂ generation was obtained by the initial DPBF concentration change over time ($\Delta[\text{DPBF}]/\Delta t$, at a shorter time where the slope has a linear behavior) divided by the concentration of the conjugates. To

demonstrate the efficiency of insulin–Au for ¹O₂ production, we compared ¹O₂ production by insulin–Au to that of the conventional dye photosensitizer, new methylene blue (NMB). Under 473 nm irradiation, a high ¹O₂ production efficiency was observed for insulin–Au (¹O₂ generation rate of 1.5 ± 0.3 compared with a value for NMB of 2.3). The quantum yield of singlet oxygen generation allows normalization by the absorbance at 473 nm of the cw laser. Using a comparative method (as described in Methods and Fig. S13†), we were able to evaluate the ¹O₂ generation quantum yield of insulin–Au conjugates at $13 \pm 5\%$. However, we cannot exclude that the generation of oxidizing species (H₂O₂, radicals) upon irradiation may also occur. To better address this point, we used a DCF dye²⁵ for probing the oxidizing agents produced following photoexcitation. We confirm that insulin–Au conjugates are prone to producing such oxidizing agents in addition to ¹O₂ species. Fig. S13(c)† displays the fluorescence spectra of DCF dye without and with 10 min of laser irradiation. The strong increase in fluorescence intensity is a signature of the generation of type I species.

Non-linear optical properties. We measured the two-photon excited luminescence (TPEL) spectra over a wide range of 800–1100 nm excitation wavelengths. Based on TPEL, we calculated the two-photon absorption cross-sections (TPA) – σ_2 . The features observed in TPA cross-sections as a function of wavelength are fairly comparable with the features observed in one-photon excitation spectra when plotted at doubled wavelength, for which resonance effects in a two-photon process may occur.^{29–31} Indeed, significant resonance enhancement occurs when the excitation energy of a one-photon absorption state is close to half that of a TPA state (“double resonance” condition, see Scheme S1†). Two photon brightness can be calculated by multiplying the TPA cross-sections by the one-photon luminescence QY (0.054) and the values range from 0.08 to 0.32 GM for excitation wavelengths in the range 800–1100 nm (Fig. 2).

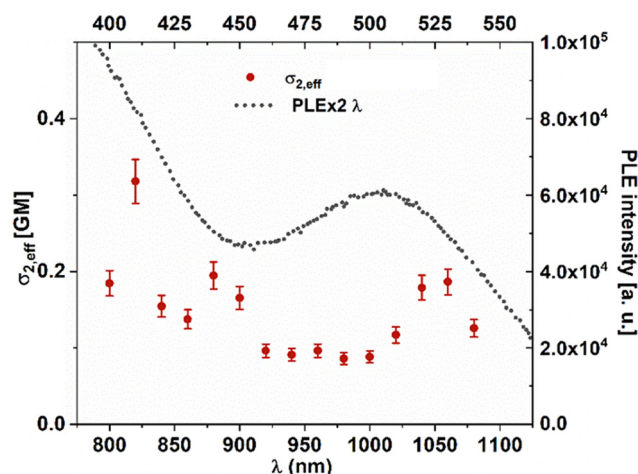


Fig. 2 Two-photon brightness ($\sigma_{2,\text{eff}}$) spectra of insulin–gold conjugate (red dots, left and bottom axes) correlated with the photoluminescence excitation spectrum of insulin–gold (black dotted line, right and upper axes). Brightness, defined as the product of absorption cross-section and fluorescence quantum yield.



enhancement of the luminescence properties, in both the linear and nonlinear optical regimes. They present long luminescence lifetimes enable efficient reactive oxygen species generation that may create oxidative stress, pointing to potential antimicrobial applications.^{25,36} Regarding other bio-applications, it was demonstrated by Liu *et al.*⁵ that the blood glucose-lowering activities of humulin R and insulin–Au nanoclusters showed no significant differences, which implies that the as prepared insulin–Au nanoclusters retained bioactivity in reducing blood glucose. We postulate that similar blood glucose-lowering activities would be reported with the present insulin–Au(III) conjugates, since the native structure of insulin is not greatly affected by the presence of a single Au(III) in the conjugate. Detailed structural characterization allows us to suggest that deprotonations on insulin occur on tyrosine residues. The interaction of Au(III) ions with the aromatic rings of tyrosine induces charge transfer-like excitation³⁷ in the visible range. Deprotonated phenolic groups in tyrosine residues serve as anchors for stabilizing Au(III) ions. Model calculations can establish new concepts for nonlinear optics using protein–Au(III) conjugates. In particular, engineering mutations in proteins with tyrosines will allow more selective control of the number and the position of Au(III) ions in proteins, thereby opening up new routes for the rational control of their photophysical properties. Finally, we point out the unique character of insulin having no free cysteine and only 4 tyrosine residues, thus it serves as an ideal template for coordinating Au(III). Other proteins have high numbers of tyrosine (and other aromatic acid groups) and free cysteines where the reduction of gold (Au(III) → Au(I) → Au(0)) enters into competition and where nanoclusters can be produced and anchored.

Author contributions

R. A. conceived the initial idea and coordinated the work. D. B., H. Y. synthesized and prepared the nanoclusters. C. B., C. C. Z. and F. C. recorded and analyzed mass spectra. H. Y. conducted ROS experiments with nanoclusters. A. H. A. P. and D. B. conducted circular dichroism and nonlinear optical experiments, supervised by J. O. B., Z. S. M., and V. B. K. performed and analyzed the theoretical results. V. G. C. carried out crystallization tests and DLS measurements. R. A., D. B., and H. Y. wrote the paper. All authors provided critical feedback and helped to shape the final manuscript.

Data availability

The data supporting the findings of this study are available within the article and its ESI.† Additional data that support the findings of this study are available from the corresponding author upon reasonable request. This includes computational codes and raw data files generated during the study.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

D. B. is grateful for post-doc fellowship donated by Agence Nationale de la Recherche (project MANBAMM, ANR-21-CE29-0020). H. Y. is grateful for PhD fellowships donated by the China Scholarship Council (CSC). C. B. is grateful for a PhD fellowship donated by the Agence Nationale de la Recherche (project nanoGOLD, ANR-22-CE29-0022). R. A. and H. Y. acknowledge Shanghai Science and Technology Innovation Program (22520712500) for support. R. A. and D. B. acknowledge Agence Nationale de la Recherche (project MANBAMM, ANR-21-CE29-0020) for support. R. A. and C. C. Z. acknowledge Agence Nationale de la Recherche (project MOONSTONE, ANR-22-CE42-0031) for support. R. A., C. B. and F. C. acknowledge Agence Nationale de la Recherche (project nanoGOLD, ANR-22-CE29-0022) for support. R. A., D. B., H. Y., A. H., A. P., and J. O. B. acknowledge PHC POLONIUM (Project: 49252PM, “Gold nanoclusters in chiral nanoassemblies – nonlinear optical properties”) for support. VBK acknowledges the STIM-REI project, contract KK.01.1.1.01.0003. The authors would like to thank Carol Grossiord from Science et Surface (<https://www.science-et-surface.fr>) for XPS spectra. We acknowledge the contribution of SFR Biosciences (Université Claude Bernard Lyon 1, CNRS UAR3444, Inserm US8, ENS de Lyon) Protein Science Facility.

References

- 1 N. Goswami, K. Zheng and J. Xie, *Nanoscale*, 2014, **6**, 13328–13347.
- 2 R. Antoine, D. Maysinger, L. Sancey and V. Bonačić-Koutecký, *Commun. Chem.*, 2022, **5**, 47.
- 3 N. Alkudaisi, B. A. Russell, B. Jachimska, D. J. S. Birch and Y. Chen, *J. Mater. Chem. B*, 2019, **7**, 1167–1175.
- 4 F. Wen, Y. Dong, L. Feng, S. Wang, S. Zhang and X. Zhang, *Anal. Chem.*, 2011, **83**, 1193–1196.
- 5 C.-L. Liu, H.-T. Wu, Y.-H. Hsiao, C.-W. Lai, C.-W. Shih, Y.-K. Peng, K.-C. Tang, H.-W. Chang, Y.-C. Chien, J.-K. Hsiao, J.-T. Cheng and P.-T. Chou, *Angew. Chem., Int. Ed.*, 2011, **50**, 7056–7060.
- 6 X. Xia, Y. Long and J. Wang, *Anal. Chim. Acta*, 2013, **772**, 81–86.
- 7 H. Shi, M. Y. Ou, J. P. Cao and G. F. Chen, *RSC Adv.*, 2015, **5**, 86740–86745.
- 8 A. Soleilhac, F. Bertorelle and R. Antoine, *Spectrochim. Acta, Part A*, 2018, **193**, 283–288.
- 9 I. Zare, D. M. Chevrier, A. Cifuentes-Rius, N. Moradi, Y. Xianyu, S. Ghosh, L. Trapiella-Alfonso, Y. Tian, A. Shourangiz-Haghighi, S. Mukherjee, K. Fan and M. R. Hamblin, *Mater. Today*, 2023, **66**, 159–193.
- 10 V. Bonačić-Koutecký, X. Le Guével and R. Antoine, *ChemBioChem*, 2023, **24**, e202200524.



- 11 J. Xie, Y. Zheng and J. Y. Ying, *J. Am. Chem. Soc.*, 2009, **131**, 888–889.
- 12 J. M. Dixon, J. Tomida and S. Egusa, *J. Phys. Chem. Lett.*, 2020, **11**, 3345–3349.
- 13 J. M. Dixon and S. Egusa, *J. Am. Chem. Soc.*, 2018, **140**, 2265–2271.
- 14 J. M. Dixon and S. Egusa, *J. Phys. Chem. Lett.*, 2021, **12**, 2865–2870.
- 15 M. Shamsipur, E. Babaei, M.-B. Gholivand, F. Molaabasi, F. Mousavi, A. Barati, B. Hajipour Vardom, B. Shojaedin-Givi and H. Naderi-Manesh, *ACS Appl. Nano Mater.*, 2023, **6**, 5939–5951.
- 16 C. Comby-Zerbino, X. Dagany, F. Chirot, P. Dugourd and R. Antoine, *Mater. Adv.*, 2021, **2**, 4896–4913.
- 17 F. Bertorelle, C. Moulin, A. Soleilhac, C. Comby-Zerbino, P. Dugourd, I. Russier-Antoine, P.-F. Brevet and R. Antoine, *ChemPhysChem*, 2018, **19**, 165–168.
- 18 S. Müllegger, W. Schöfberger, M. Rashidi, T. Lengauer, F. Klappenberger, K. Diller, K. Kara, J. V. Barth, E. Rauls, W. G. Schmidt and R. Koch, *ACS Nano*, 2011, **5**, 6480–6486.
- 19 S. G. Melberg and W. C. Johnson Jr., *Proteins: Struct., Funct., Bioinf.*, 1990, **8**, 280–286.
- 20 L. Whitmore and B. A. Wallace, *Nucleic Acids Res.*, 2004, **32**, W668–W673.
- 21 A. Soleilhac, F. Bertorelle, C. Comby-Zerbino, F. Chirot, N. Calin, P. Dugourd and R. Antoine, *J. Phys. Chem. C*, 2017, **121**, 27733–27740.
- 22 H. Yuan, D. Lima, C. Comby-Zerbino, C. Bouanchaud, F. Chirot, D. Bain, S. Zhang and R. Antoine, *Int. J. Mass Spectrom.*, 2024, **503**, 117276.
- 23 W. Mohammad, K. D. Wegner, C. Comby-Zerbino, V. Trouillet, M. P. Ogayar, J.-L. Coll, R. Marin, D. J. Garcia, U. Resch-Genger, R. Antoine and X. Le Guével, *J. Mater. Chem. C*, 2023, **11**, 14714–14724.
- 24 H. Fakhouri, M. P. Bakulić, I. Zhang, H. Yuan, D. Bain, F. Rondepierre, P.-F. Brevet, Ž. S. Maršić, R. Antoine, V. Bonačić-Koutecký and D. Maysinger, *Commun. Chem.*, 2023, **6**, 97.
- 25 H. Yuan, M. Theresa, D. Bain, H. Fakhouri, K. Sreekanth, A. Ravi, S. Thomas, N. Kalarikkal, R. Antoine and E. K. Radhakrishnan, *Inorg. Chem. Commun.*, 2024, **159**, 111799.
- 26 V. Poderys, G. Jarockyte, S. Bagdonas, V. Karabanovas and R. Rotomskis, *J. Photochem. Photobiol., B*, 2020, **204**, 111802.
- 27 M. Yamamoto, I. Osaka, K. Yamashita, H. Hasegawa, R. Arakawa and H. Kawasaki, *J. Lumin.*, 2016, **180**, 315–320.
- 28 H. Kawasaki, S. Kumar, G. Li, C. Zeng, D. R. Kauffman, J. Yoshimoto, Y. Iwasaki and R. Jin, *Chem. Mater.*, 2014, **26**, 2777–2788.
- 29 Z. Hu and L. Jensen, *Chem. Sci.*, 2017, **8**, 4595–4601.
- 30 I. Russier-Antoine, F. Bertorelle, N. Calin, Ž. Sanader, M. Krstić, C. Comby-Zerbino, P. Dugourd, P.-F. Brevet, V. Bonačić-Koutecký and R. Antoine, *Nanoscale*, 2017, **9**, 1221–1228.
- 31 Ž. Sanader, M. Krstić, I. Russier-Antoine, F. Bertorelle, P. Dugourd, P.-F. Brevet, R. Antoine and V. Bonačić-Koutecký, *Phys. Chem. Chem. Phys.*, 2016, **18**, 12404–12408.
- 32 R. Antoine, F. Bertorelle, M. Broyer, I. Compagnon, P. Dugourd, A. Kulesza, R. Mitrić and V. Bonačić-Koutecký, *Angew. Chem., Int. Ed.*, 2009, **48**, 7829–7832.
- 33 B. Bellina, I. Compagnon, F. Bertorelle, M. Broyer, R. Antoine, P. Dugourd, L. Gell, A. Kulesza, R. Mitrić and V. Bonačić-Koutecký, *J. Phys. Chem. C*, 2011, **115**, 24549–24554.
- 34 I. Russier-Antoine, F. Bertorelle, A. Kulesza, A. Soleilhac, A. Bensalah-Ledoux, S. Guy, P. Dugourd, P.-F. Brevet and R. Antoine, *Prog. Nat. Sci.: Mater. Int.*, 2016, **26**, 455–460.
- 35 I. Compagnon, A.-R. Allouche, F. Bertorelle, R. Antoine and P. Dugourd, *Phys. Chem. Chem. Phys.*, 2010, **12**, 3399–3403.
- 36 S. Chakraborty, P. Sagarika, S. Rai, C. Sahi and S. Mukherjee, *ACS Appl. Mater. Interfaces*, 2021, **13**, 36938–36947.
- 37 R. Antoine, T. Tabarin, M. Broyer, P. Dugourd, R. Mitrić and V. Bonačić-Koutecký, *ChemPhysChem*, 2006, **7**, 524–528.

