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## Synthesis and characterization of fluorescent gold clusters encapsulated in PNIPAM and PAAM hydrogels for selective detection of mercury in water†

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The present work describes the synthesis of new nanomaterials based on gold nanoclusters (AuNCs) supported on poly(*N*-isopropylacrylamide) (PNIPAM) and polyacrylamide (PAAM) hydrogels, as well as their characterization in terms of structure and physicochemical properties. AuNCs were synthesized by incorporating the metallic precursor (Au<sup>3+</sup>) into the polymeric matrices and subsequently reducing it with UV-vis light ( $\lambda = 365$  nm). The materials were characterized by UV-vis, FT-IR, fluorescence spectroscopy, TEM and SEM. Likewise, the thermal properties and the swelling capacity of the different polymer matrices were determined. In addition, we present a comparative study with two nanomaterials regarding their behavior for the selective detection of Hg<sup>2+</sup> in water. Furthermore, it is demonstrated that the polymer matrices influence the nanomaterials' selectivity to mercury. As the AuNCs fluorescence incorporated in both matrices is quenched by Hg<sup>2+</sup>, the materials can be used to sense sensitively and selectively Hg<sup>2+</sup> in water with a detection limit of 4.1 nM and 1.3 nM for Au-PNIPAM and Au-PAAM NCs, respectively. Summarizing, our study presents a novel, easy and straightforward route to synthesized nanomaterials with unique optical and chemical properties, which could be used for the construction of a portable Hg<sup>2+</sup> detection device.

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### Environmental significance

Precise and efficient detection of mercury in water is a fundamental challenge to protect the environment and human health. In this work, we present a novel approach for the detection of mercury in water using gold nanoclusters supported on polymer matrices. Our approach has demonstrated high sensitivity and selectivity for the detection of mercury in water samples. The approach could have a significant impact on protecting the environment and human health by enabling early detection of mercury, which can help prevent exposure to this dangerous pollutant and reduce negative effects on aquatic life and the ecosystem. In addition, the technology we have developed could be applied in other areas where early and efficient detection of pollutants in the environment is required, which could have a positive impact on environmental protection and human health around the world.

## Introduction

Contamination of drinking water and food with heavy metals is a social problem in many regions of the world. Especially in the Pampas plains of our country, with a rural population of more than 1 000 000 inhabitants, soil and groundwater can be contaminated with Hg, Cr, and As.<sup>1</sup> The presence of heavy metals in water can be harmful to human health, for this reason the World Health Organization (WHO) has set the limit of Hg in drinking water at 0.001 mg L<sup>-1</sup>.<sup>2,3</sup> The determination and quantification of these contaminants demand expensive and sophisticated instruments and complicated sample preparation processes.<sup>4</sup> A fast, simple, and efficient alternative for the sensitive optical detection of heavy metals is the use of highly fluorescent metal nanoclusters (MNCs) supported on different matrices.<sup>5</sup> MNCs consist of only a few atoms of the metal (M<sub>n</sub>, n = 2–20) and are interesting because of their ultra-small

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size, excellent fluorescent properties, low toxicity, and high stability.<sup>6</sup> An interesting strategy is to synthesize MNCs supported on scaffolded matrices such as polymers, DNA, and hydrogels.<sup>7–9</sup> It is known that the matrix-MNC combination affects the optical properties of the systems, giving them great selectivity for the specific detection of some heavy metals in water by fluorescence quenching.<sup>10,11</sup> Several matrices can be employed, the hydrogels present several advantages such as high hydrophilicity, easy synthesis, and high swelling in water, among others.<sup>12,13</sup> In this sense, three-dimensional crosslinked networks of hydrophilic polymer chains where the free polymer segments are solvated by water constitute a hydrogel. Among the most studied hydrogels are those formed by different polyacrylamides such as poly(*N*-isopropylacrylamide) (PNIPAM) and polyacrylamide (PAAM).<sup>14–17</sup> Polyacrylamide gels can be easily fabricated into macroscopic shapes. Polymerization of an acrylamide monomer together with an appropriate cross-linker in a concentrated aqueous solution produces a solid gel that fills the mold or copies the shape of the solution container.<sup>18</sup> Since the radical polymerization produces a distribution of chain lengths with a random distribution of cross-links, the structures are a 3D network with open and connected nanopores.<sup>19</sup> One of the most interesting features is their capability of swelling greatly in water exposing the polymer chains to the aqueous media.<sup>13</sup> The ability of hydrogels to absorb water arises from hydrophilic functional groups attached to the polymer backbone while their resistance to dissolution arises from cross-links between network chains.<sup>20</sup> Water inside the hydrogel allows free diffusion of some solute molecules, while the polymer serves as a matrix to hold water together. Hydrogels may exhibit drastic volume changes in response to specific external stimuli, such as temperature, solvent quality, pH, electric field, *etc.*<sup>21</sup> Such is the case of PNIPAM that in addition to great swelling in water, PNIPAM presents thermosensitive properties since it undergoes a hydrophilic to hydrophobic phase transition at temperatures (32–34 °C).<sup>22</sup> In addition, the mechanical properties of hydrogels allow them to be used as matrix not only to contain active materials but also to sense different chemical species, such as heavy metals (As,<sup>23</sup> Fe<sup>24</sup>), dopamine,<sup>25</sup> among others.

Several research articles demonstrate that toxic heavy-metal ions may be detected by hydrogels through the spectroscopic method. For example, mercury can be detected using thymine-rich DNA-polyacrylamide hydrogels.<sup>26</sup> The Hg<sup>2+</sup> in the gel was detected by green fluorescence, after the binding mercury between two thymine bases and the addition of SYBR Green I dye. A reliable visual detection of 10 nM Hg<sup>2+</sup> ions is possible with additional value-added capabilities, such as mercury removal through interaction with acrylamide. On the other hand, other authors showed that heavy metal ions can be detected by the fluorescence of MNCs embedded in hydrogels, such as AuNC, AgNC, CuNC, *etc.*<sup>27–30</sup> Particularly, the fluorescence of gold nanocluster (AuNCs) is quenched through electron transfer from heavy-metal ions.<sup>27</sup> AuNC-based sensors provide a 5 nM linear detection range for Hg<sup>2+</sup> ions.<sup>31</sup> One strategy for MNCs preparation is to use pre-formed hydrogels. A pre-formed hydrogel acts as a microreactor, in which metal nanostructures are formed from

its precursor, after a series of required treatments. The metal ions can be absorbed by the hydrogels functional group by electrostatic or dipole ion interaction. During the absorption, the ion exchange occurs between metal ions present in the solution and the functional group present within the hydrogel. After absorption, metal ions can be reduced with reducing agents such as NaBH<sub>4</sub>,<sup>32</sup> H<sub>2</sub>,<sup>33</sup> citrate,<sup>34</sup> our UV-vis light,<sup>35,36</sup> which contribute to different particle sizes from nanoparticles to MNCs.<sup>37</sup> Therefore, it is expected that this synthetic method of matrix-MNCs allows the design and construction of metal ion sensors and photocatalysts for the remediation of contaminated water suitable for use, without the problem of being released into the water that is being purified. The use of MNCs-embedded hydrogels for the fluorescence detection of mercury ions in water offers numerous advantages and practical applications compared to other methods. The hydrogels can be designed with high affinity and selectivity towards the mercury ion, allowing highly sensitive and specific detection. They generally have a fast response time, which results in shorter detection times.

Moreover, the MNCs incorporated into the matrix could enable the construction of a portable and straightforward sampling material to mercury detection. In this way, the MNCs hydrogel can be transported to a specific location where sampling is required, allowed for water uptake, and then determine the Hg content in the laboratory. In addition, hydrogel-based sensors are relatively easy to use and do not require complex laboratory equipment, which increases their ease of use and portability.

This work describes the synthesis and physicochemical characterization of AuNCs supported on polymeric matrices based on PNIPAM and PAAM hydrogels. Also, the active AuNCs inside the matrix was employed and tested as a probe of concept for the development of a portable fluorescent sensor to detect and quantify heavy metals (Hg<sup>2+</sup>) in contaminated water. In summary, this work shows a simple, straightforward manner to produce a useful device that allows the detection of Hg<sup>2+</sup> in concentration above the allowed WHO limit in drinking water.

## Experimental

### Material and methods

**Materials.** *N*-isopropylacrylamide (NIPAM) and acrylamide (AAM) used as monomer was obtained from Scientific Polymer Products (purity > 99%). *N,N*-methylenebisacrylamide (BIS), *N,N,N',N'*-tetramethylethylenediamine (TEMED), and ammonium peroxydisulfate (APS) were purchased from Aldrich. Water used in this work was bidistilled. All other reagents are of analytical quality. HAuCl<sub>4</sub>, 99.99% was employed to obtain nanomaterials. NaCl, KCl, CaCO<sub>3</sub>, HgCl<sub>2</sub>, ZnSO<sub>4</sub>, AlCl<sub>3</sub> and FeCl<sub>3</sub> used for quenching tests was from Sigma-Aldrich quality (purity > 99%).

### Hydrogel synthesis

The *N*-isopropylacrylamide (PNIPAM, Scheme 1E) and poly(acrylamide) (PAAM, Scheme 1F) hydrogels were synthesized using the same method. Firstly, 1.5 mmol of the monomer (either NIPAM or PAAM), and 0.06 mmol of BIS





**Scheme 1** Chemical structure of reagents and polymeric hydrogels synthesized: (A) NIPAM, (B) AAM, (C) TEMED, (D) BIS, (E) PNIPAM, and (F) PAAM.

(as a crosslinking agent) were dissolved in 3 mL of distilled water, resulting in a 0.5 M monomer solution. The hydrogel free radical polymerization was carried out in a glass tube at room temperature (22 °C) for 3 h, using APS (0.01 mmol) and TEMED (0.03 mL) as redox initiators. After the polymerization, the hydrogel pieces were immersed in distilled water at room temperature for 48 h. The distilled water was renewed every hour to remove soluble by-products<sup>4,38</sup> Then, the hydrogels were dried for 48 h at 25 °C and moderate vacuum (0.1 atm) for later use.

**Synthesis of nanomaterials (Au-PNIPAM and Au-PAAM nano clusters).** The nanomaterials were synthesized *in situ*. First, the hydrogels were immersed in 0.5 mM Au<sup>3+</sup> solution and allowed to swell in the solution for 96 h, to obtain Au<sup>3+</sup>-PNIPAM and Au<sup>3+</sup>-PAAM materials. Next, the Au<sup>3+</sup>-hydrogels were dried in a vacuum at 30 °C for 48 h. After that, the Au<sup>3+</sup>-hydrogels were irradiated with a UV lamp (365 nm) for 3 h, to photo-reduce Au<sup>3+</sup>cation and, obtain the pure nanocomposites Au-PNIPAM and Au-PAAM NCs. The purified Au-PNIPAM and Au-PAAM NCs were stable for at least 3 months when stored at 4 °C in the dark.

**Materials characterization.** UV-vis spectra were recorded using a spectrophotometer Shimadzu 2401 with a sample holder thermostated at 25 °C.

The chemical composition of the hydrogels and the nanocomposites were analyzed by FTIR-ATR. The spectra were obtained by an ATR-FTIR Bruker Tensor 27 spectrometer in absorbance mode using air as a background. In the first place, the samples were dried for 48 h at 30 °C under a dynamic vacuum system.

Also, A Carl Zeiss EVO MA10 scanning electron microscope with an integrated EDX system was used to examine the morphology of the polymers and the nanocomposites. The materials were lyophilized to preserve the gel structure during drying, after that the samples were gold-coated for conductance.

A Fluoromax Horiba Plus spectrofluorometer was employed to analyze the fluorescent properties of the nanomaterials.

The thermo-responsive performance of hydrogels was characterized *via* differential scanning calorimetry (DSC). A Netzsch DSC-204-F1-Phoenix differential scanning calorimeter equipped with a cooling device was used to perform the test. The hydrogels were characterized at a 5.0 °C min<sup>-1</sup> heating and cooling rate for 2 cycles from -10 °C to 40 °C to study the effects of gold nanomaterials in the hydrogels.

Transmission electron microscopy (TEM) micrographs were recorded by using a Joel JEM-1400 microscope at 20–120 kV. For TEM analysis, a drop of AuNCs was suspended onto a copper-coated grid. The grids were dried in a desiccator for 24 h and then they were examined. Histogram AuNCs by number (*n*) were made with “ImageJ” program, which captures the scale of TEM micrographs.

### Hydrogels swelling kinetics

Swelling kinetic measurements were carried out for PNIPAM and PAAM hydrogels employing two different media: (i) distilled water, and (ii) solutions of gold nanoparticles precursor HAuCl<sub>4</sub> (Au<sup>3+</sup> 0.5 mM and Au<sup>3+</sup> 1 mM). Dried hydrogel disks, previously washed and weighed, were placed in the media at room temperature (20 °C). The samples were removed from the solution, at certain time intervals, superficially dried with tissue paper, weighed in an analytic balance, and placed back in the solution. The swelling percentage is calculated as a function of time according to eqn (1):<sup>13</sup>

$$\text{Swelling}(\%) = \frac{(W_s - W_d)}{W_d} \times 100 \quad (1)$$

where  $W_s$  stands for the sample weight of the swollen state at a given time and  $W_d$  is the dry sample weight. The equilibrium swelling percentage (% Sw<sub>eq</sub>) was calculated to measure the sample weight after 2 h until constant weight.



## Detection of heavy metals contaminants

A stock solution of  $\text{HgCl}_2$  (10 mM) was prepared, which was then diluted in bidistilled water to form 1  $\mu\text{M}$   $\text{Hg}^{2+}$  solutions. In  $\text{Hg}^{2+}$  detection experiments, an aliquot of dilute  $\text{Hg}^{2+}$  solution was taken and added to a vial containing Au-PNIPAM NCs to obtain a final volume of 1 mL. The same procedure was used for Au-PAAM NCs. The samples were incubated for 30 min at room temperature. The fluorescence spectra of the nanomaterials were recorded before and after the addition of the heavy metals.

## Results and discussion

### Polymer matrix characterization

Fig. 1A shows the absorption spectra of Au nanomaterials supported on PNIPAM (black line) and PAAM (red line) hydrogel, while the corresponding spectra of PNIPAM (black line) and PAAM (red line) pure hydrogels are shown in Fig. 1B. In Fig. 1A a strong absorption band can be observed between 500 and 600 nm. Similarities in both sets of spectra are observed in these figures. This band is associated with the surface plasmon resonance of Au nanoparticles (Au-NPs). The maximum absorption wavelengths of the SPR bands ( $\lambda_{\text{max}}$ ) are located at 540 nm and 544 nm for Au nanomaterials supported on PNIPAM and PAAM polymer matrices, respectively. Chen *et al.*<sup>9</sup> demonstrate that the bands located at 540–544 nm correspond to Au-NPs, and they are originate from Au nanomaterial complexes by charge transfer from the ligand to the metal. In addition, hydrogels without incorporated metals do not present an absorption band in this region of the spectrum (Fig. 1B). Furthermore, it is noted that for Au-NPs supported in PNIPAM, the bandwidth is large and the  $\lambda_{\text{max}}$  is shifted to the blue compared to the Au-NPs supported in PAAM. The SPR hypsochromic shift provides information about the size and stabilization of nanomaterials. According to Mie's theory,<sup>39</sup> there is a relationship between SPR maximum and size, the Au-NPs are smaller when there is a hypsochromic shift in the SPR maximum.<sup>40</sup> The different  $\lambda_{\text{max}}$  obtained could be suggesting that the size, growth, and, collective dipole oscillations of Au-free electrons depend on the stabilizing matrix chemical nature used in the synthesis.<sup>41</sup> The hydrogel's different

chemical properties depend on the monomer's chemical structure, which affects the nanomaterial optical properties incorporated into the hydrogel. In addition, the absence of an important shift of the maximum absorbance peak suggests a good Au-NPs dispersion within the hydrogels.

The hydrogels microstructure was investigated by SEM, examining the samples cross-sections. In Fig. 2, micrographs of the pristine hydrogels and gold-based nanomaterials are presented. The pure hydrogel and the metal-embedded material are compared before and after irradiation with UV light.

The SEM images (B, C, E, and F) correspond to the materials synthesized by impregnating the hydrogels with 0.5 mM metal precursor.

In the case of PNIPAM (Fig. 2A), the surface presents a regular and compact structure, while the  $\text{Au}^{3+}$ -PNIPAM and Au-PNIPAM NCs (Fig. 2B and C) show additional layered structures probably due to the presence of Au nanostructures. However, the Au nanostructures, due to their small size, cannot be seen. Comparing the PNIPAM-based nanomaterials (B and C), it can be noted that UV light irradiation modifies their structure. In this way, UV light not only contributes to the formation of metallic nanostructures but also causes the polymeric matrix framework to break. The irradiated  $\text{Au}^{3+}$ -PNIPAM hydrogel (Au-PNIPAM NCs) presents "holes" in the order of microns and is more porous and less compact as compared to pure PNIPAM and  $\text{Au}^{3+}$ -PNIPAM.

On the other hand, it can be observed that the PAAM hydrogel has a smooth surface (Fig. 2D). In contrast, PAAM-based materials containing gold exhibit a rough surface (Fig. 2E and F). It should be noted that no differences were observed between the materials with gold (Fig. 2E and F) before and after irradiation. The SEM images of the two hydrogels show a homogeneous polymer surface with similar surface properties. The SEM images of both hydrogels show a homogeneous polymer surface of similar surface characteristics. In this case, irradiation would not alter the microstructure of the hydrogel.

Also the chemical structure of the polymers was characterized by infrared spectroscopy. FT-IR measurements were performed for the pure hydrogels and their respective gold-based nanomaterials. The most relevant stretch mode bands for



Fig. 1 UV-vis spectra of: (A) Au nanomaterials obtained in PNIPAM (Au-PNIPAM NCs, black line) and PAAM (Au-PAAM NCs, red line) hydrogels. (B) PNIPAM (black line) and PAAM (red line) hydrogels.





Fig. 2 SEM images of the materials: (A) PNIPAM, (B) Au<sup>3+</sup>-PNIPAM, (C) Au-PNIPAM NCs, (D) PAAM, (E) Au<sup>3+</sup>-PAAM, and (F) Au-PAAM NCs.

polymeric hydrogels were identified by FT-IR. The analysis was focused on the stretching bands of the CH<sub>2</sub> and CH<sub>3</sub> and the isopropyl groups because important information about the Au-polymer matrix interaction strength can be obtained in this spectral region. Fig. 3 shows the spectra of pure PNIPAM and PMAA hydrogel and the gold-modified nanomaterials. Well, resolved bands are observed in the 2900 cm<sup>-1</sup> region for the polymeric materials based on PNIPAM, Fig. 3A. The 2980–2870 cm<sup>-1</sup> region is assigned to CH<sub>2</sub> and CH<sub>3</sub> stretches. In addition, the bands observed at 1540 and 1457 cm<sup>-1</sup> correspond to isopropyl groups, which confirms the presence of these groups in the materials. These bands are present not only in the pristine polymer but also remain in the Au-PNIPAM NCs indicating that the hydrogel backbone is not altered during the nanomaterials formation.

Pure PNIPAM (Fig. 3A, black line) shows two bands (doublet) at 1385 and 1365 cm<sup>-1</sup> corresponding to the umbrella mode of

isopropyl groups.<sup>42</sup> The appearance of this doublet is due to the presence of two independent CH<sub>3</sub> groups in different environments. The interaction between hydrogen atoms in two different methyl groups depends on whether these groups are moving closer to or further from each other.<sup>43</sup> In contrast the spectrum of Au-PNIPAM NCs (Fig. 3A, red line) shows a single band at 1377 cm<sup>-1</sup>, suggesting that the umbrella mode of the isopropyl group is perturbed when the material is modified with Au. Thus, this indicates the success of PNIPAM coating on the surface of Au nanostructures.<sup>44</sup> Furthermore, the results suggest that the freedom of movement of the isopropyl groups is restricted in Au-PNIPAM NCs. This interaction could be responsible for the stability of the nanostructures, avoiding the collapse of the system.

On the other hand, the PAAM-based materials only show changes in the 2900 cm<sup>-1</sup> region corresponding to the CH<sub>2</sub> and CH<sub>3</sub> stretching modes (Fig. 3B). The pure PAAM hydrogel

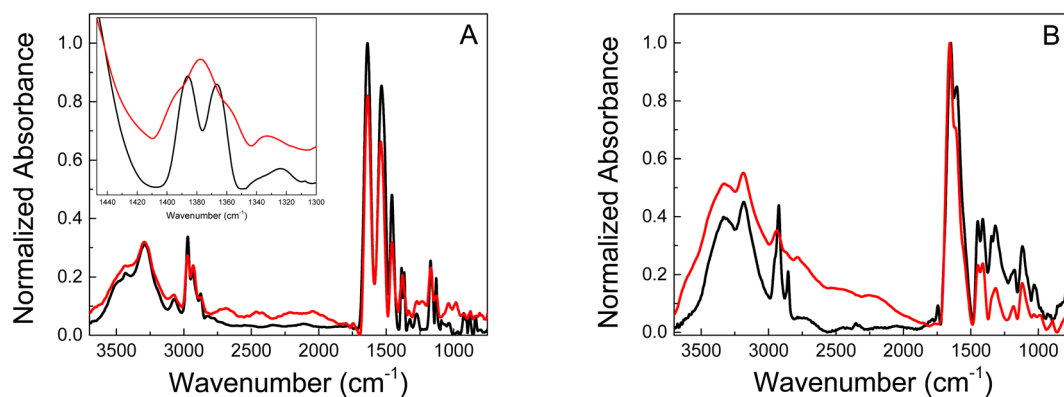


Fig. 3 FT-IR spectra of (A) pure PNIPAM (black) and Au-PNIPAM NCs (red) lyophilized materials. (B) Pure PAAM (black) and Au-PAAM NCs (red) lyophilized materials.



presents a narrow band at  $2920\text{ cm}^{-1}$  and a less intense one at  $2855\text{ cm}^{-1}$  (Fig. 3B, black line). In contrast, the Au-PAAM NCs (Fig. 3B, red line) shows a single band located at  $2926\text{ cm}^{-1}$  that is less defined and with lower relative intensity as compared to the pure PAAM matrix. Likewise, the band at  $2855\text{ cm}^{-1}$  disappears when the material contains gold nanostructures. The observed behavior suggests that the Au nanostructures interact with the alkyl groups of the polymeric matrix. Due to the magnitude of the observed changes, the interaction of the Au nanostructures with PAAM hydrogel would be stronger than with PNIPAM hydrogel.

Swelling is one of the most important parameters in hydrogel technology. The hydrogel swelling capacity was studied in two different media: (i) distilled water, (ii) gold precursor solution  $\text{HAuCl}_4$  ( $\text{Au}^{3+}$  0.5 mM). The swelling capacity (swelling%) was determined as a function of time according to eqn (1). Fig. 4 shows the PNIPAM (A) and PAAM (B) hydrogels swelling kinetic in the distilled water and metallic precursor solutions ( $\text{Au}^{3+}$  0.5 mM). Also, a swelling percentage in the equilibrium state ( $\% \text{Sw}_{\text{eq}}$ ) is shown in Table 1.

As shown in Fig. 4 and Table 1, the swelling capacity for the different polymeric matrices increase gradually with the time at room temperature. All samples exhibited exponential growth curves, characteristic of hydrophilic hydrogels. No significant differences are observed for the PNIPAM-based hydrogels in the equilibrium swelling value ( $\% \text{Sw}_{\text{eq}}$ ) in both mediums, which is approximately 2500% (Fig. 4A). On the other hand, it can be noted that the initial swelling velocity is different for each material.

The initial swelling rate for pure PNIPAM in water is lower than for the same hydrogel in the metal solution. The hydrogels swelling process is known to be controlled by three consecutive steps: the diffusion of the solvent molecule in the polymer system, the relaxation of the polymer chains, and the expansion of the polymer network in the solution.<sup>45</sup> Strong inter/intra-molecular interactions would greatly restrict the polymer chain relaxation, resulting in a slower swelling rate. The results suggest that the gold solution uptake diminish the inter/intra-molecular interactions in the PNIPAM, resulting in a high swelling rate in comparison with pure water.

In addition, the materials behavior is different for the PAAM hydrogels (Fig. 4B). The PAAM hydrogel swelling in  $\text{Au}^{3+}$  solutions has the higher equilibrium swelling value (2323%  $\text{Sw}_{\text{eq}}$ ), which could be due to the three-dimensional network having the lowest degree of cross-linking and/or the highest hydrophilic character, than pure PAAM (1677%  $\text{Sw}_{\text{eq}}$ ). Furthermore, it is observed that the metal incorporation decreases the initial swelling rate, indicating a strong interaction between the gold nanostructures and the polymer matrix. Also, the results also suggest that pure PAAM is less hydrophilic than  $\text{Au}^{3+}$ -PAAM.

The hydrogels swelling with different concentrations of metal precursor solutions was evaluated (Table 1).

It was found that the swelling capacity of PAAM-increases with the concentration of the metal solution. The presence of  $\text{Au}^{3+}$  in the materials enhance the  $\% \text{SW}_{\text{eq}}$  because it weakens the inter/intra-molecular interactions of the polymer chains, opens channels and pores, and thus allows an easy diffusion



Fig. 4 Swelling kinetics of materials based on (A) PNIPAM and (B) PAAM. The profiles correspond to the materials immersed in metal precursors solutions 0.5 mM (red line) and in bidistilled water (black line).

Table 1 Summary of the swelling of nanomaterials in different concentrations of metal solutions

Sample	Metal concentration Au (mM)	Initial weight $W_0$ (mg)	Final weight $W_d$ (mg)	Swelling (%)
PNIPAM	0	13.1	299.5	2386
PAAM	0	6.3	99.4	1677
$\text{Au}^{3+}$ -PNIPAM	0.5			2487
$\text{Au}^{3+}$ -PAAM	0.5	4.5	117	2323
$\text{Au}^{3+}$ -PAAM	1	9.6	267.1	2682.29
$\text{Au}^{3+}$ -PAAM	5	7.3	271.1	3613.69



into the material. This behavior could be due to that the gold 3+ charge of the metal cation makes the material more hydrophilic. Moreover, the same behavior was observed in the materials based on PNIPAM.

### Metal incorporation in polymeric matrices

A UV-vis spectroscopy study was carried out to determine the incorporation profiles of the metals in the different polymer matrices. Fig. 5 shows the UV-vis absorption spectra over time of the 0.5 mM solutions (initial concentration) of metal precursor where the PNIPAM and PAAM hydrogels were immersed.

In the UV-vis spectra of the gold solution show a band located between 230-300 nm corresponding to the absorption of  $\text{Au}^{3+}$ . It can be observed that the absorbance decreases with the time (Fig. 5A and B) indicating less quantity of  $\text{Au}^{3+}$  ions in the solution. This result confirms that the precursor metal concentration decreases in the solution in which the material is immersed due to the incorporation of  $\text{Au}^{3+}$  in both hydrogels. The data of % $S_{w_{eq}}$  and incorporation kinetics can also be used to calculate the  $\text{Au}^{3+}$  moles and the amount of water incorporated into the hydrogels. These calculated values can then be used to estimate the molality of  $\text{Au}^{3+}$  in each hydrogel. The amount of  $\text{Au}^{3+}$  incorporated in the hydrogels was 0.30 mol and

0.38 mol for  $\text{Au}^{3+}$ -PNIPAM and  $\text{Au}^{3+}$ -PAAM, respectively. According to the results, the PAAM hydrogel contains approximately 20% more metal precursor compared to PNIPAM.

In addition, the UV-vis spectra of the  $\text{Au}^{3+}$ -hydrogels show no SPR band (not shown), suggesting that Au NPs do not form in the PNIPAM and PAAM hydrogels without UV light irradiation. This conclusion is consistent with the UV-vis spectra showed in Fig. 5, which shows that the materials does not release nano-materials to the external solution for at least 30 hours.

The thermal properties of the pristine hydrogel PNIPAM and Au-PNIPAM NCs were evaluated by differential scanning calorimetry (DSC). The DSC thermogram of pure PNIPAM (Fig. 6A) shows an endothermic peak at 33.6 °C. PNIPAM is known to be a thermosensitive material with a lower critical solution temperature (LCST) of around 32 °C.<sup>22</sup> This peak is attributed to the endothermic process, which corresponds to the energy required to break the hydrogen bonds between the polymer and the water molecules. The thermogram of Au-PNIPAM NCs also shows an LCST proving that this new material is also thermosensitive (Fig. 6B). In the latter case, the peak presented at about 36.7–40.6 °C in the DSC thermogram (Fig. 6B) is similar to the LCST of pure PNIPAM (Fig. 6A). The differences in the LCST value is attributed to the fact that one material is a pristine NIPAM and the other, Au-PNIPAM NC is a nanocomposite of

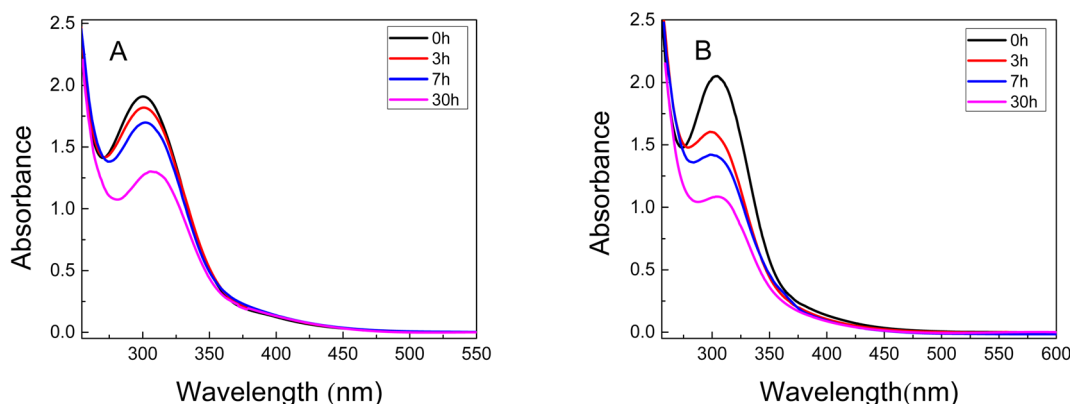


Fig. 5 UV-vis spectra of  $\text{Au}^{3+}$  solution at different time during the swelling of (A) PNIPAM (B) PAAM. 0 h (black line), 3 h (red line), 5 h (blue line), and 30 h (pink line).

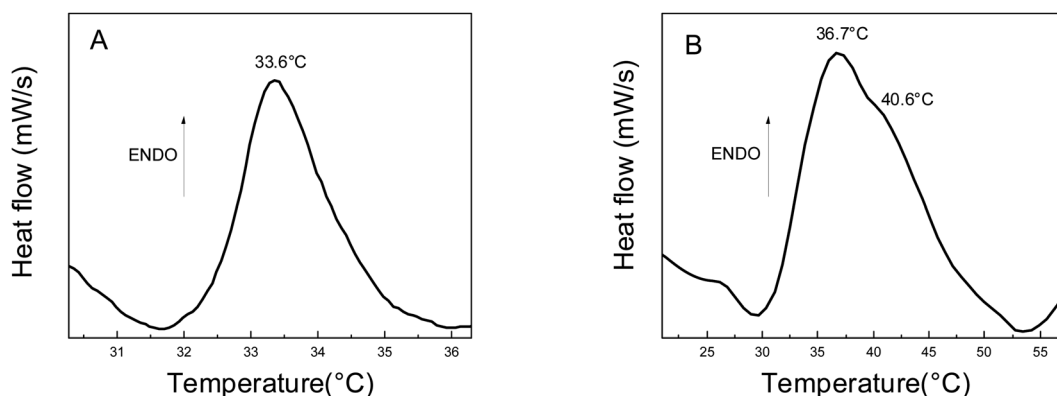


Fig. 6 DSC thermograms of (A) pure PNIPAM (B) Au-PNIPAM NCs.



PNIPAM and gold nanostructures inside of the 3D matrix. The effects of gold nanostructures on the network were not enough to destroy the balance of hydrophilic/hydrophobic interactions allowing the nanocomposite experiment the phase transition. Also, the endothermic peaks have a similar enthalpy, which is typical of hydrogen bonding interactions. However, it should be noted that Au-PNIPAM NCs has a higher phase transition temperature than pure hydrogel. This phenomenon could be attributed to the stronger electrostatic and steric interactions between gold nanomaterial and PNIPAM in the modified polymer network structures.<sup>46</sup> In addition, Schild and Tirrell found that phase transition temperatures are sensitive to the presence of different substances in the PNIPAM structure, such as comonomers, solutes, surfactants, *etc.*<sup>47</sup> Therefore, this could indicate a strong hydrogen bond interaction between water and gold nanostructures. The interaction would be stronger compared to the interaction between the water molecules and the polymeric matrix. Furthermore, the results show that the gold nanostructures remain incorporated into the polymer matrix after heating. It is important to mention that PAAM-based materials are not sensitive to temperature changes.

#### AuNCs incorporated in matrices

TEM images of the nanomaterials synthesized inside of the PMIPAM and PAAM are shown in Fig. 7. When PNIPAM hydrogels is employed as support Au-NPs are produced with a bimodal composed a main population with particles size distribution peaking at 6.5 nm and a less populated distribution peaking at 10.5 nm as shown in Fig. S1A.† In addition, numerous smaller AuNCs are observed in Fig. 7A. In the case of the material produced in PAAM hydrogels (Fig. 7B) a monodisperse and homogeneous Au-NP with an average diameter of approximately 7.5 nm is observed (Fig. S1B†). The AuNCs are too small to be observed by TEM and they are probably absorbed on the Au-NPs. However, their presence is confirmed by fluorescence spectroscopy since it is well known that Au-NPs do not fluoresce.

The fluorescent properties of the synthesized nanomaterials were also evaluated. Fluorescence was determined using a special holder and hydrated samples, recording the emission at a 45° angle. Excitation was performed at 535 nm, as the formed nanomaterials and all samples showed absorption between 500–550 nm. The fluorescence spectra of pure PNIPAM and PAAM hydrogels and Au-based nanomaterials are shown in Fig. 8. While the UV-irradiated Au-PNIPAM and Au-PAAM NCs (Fig. 8A and B, blue line) show strong fluorescence emission around 620 nm, the fluorescence of pristine hydrogels (Fig. 8A and B, black line) and non-irradiated Au<sup>3+</sup>-PNIPAM and Au<sup>3+</sup>-PAAM hydrogels (Fig. 8A and B, red line) is insignificant under the same conditions. Likewise, it can be noted that the emission intensity of Au-PAAM NCs is higher compared to Au-PNIPAM NCs. On the other hand, we found that the fluorescence of these nanomaterials depends on the pH of the solution in which the hydrogels are immersed (Fig. S2†). As shown in Fig. S2,† the Au-PNIPAM NCs show a maximum emission at pH 7, while the maximum emission of Au-PAAM takes place at pH 4. However, the fluorescence of the synthesized NCs in both matrices is detectable in the whole range of studied pH (pH = 4.0–10.0).

Several research studies had demonstrated that the AuNCs fluorescence depends on the pH.<sup>48–50</sup> In that sense Csapó *et al.*<sup>48</sup> shows that the nicotinamide-stabilized AuNCs present detectable fluorescence in the pH range 1–12, showing a fluorescence decrease at pH < 3.0 and pH > 10.0. Moreover, the decreasing of the fluorescence a low pH is attributed to the formation of aggregates of NAM-Au NCs and at high pH the clusters lose their stability again thanks to the high electrostatic shielding. Evidently the chemical structure and swelling degree of the hydrogel changes with pH, which affects the emission of AuNC. However, since the goal of this work is to develop a new fluorescent material to detect Hg<sup>2+</sup> in water as obtained from the tap or any natural water course, without further treatment, the sensitivity and selectivity experiments were all performed at pH 7.

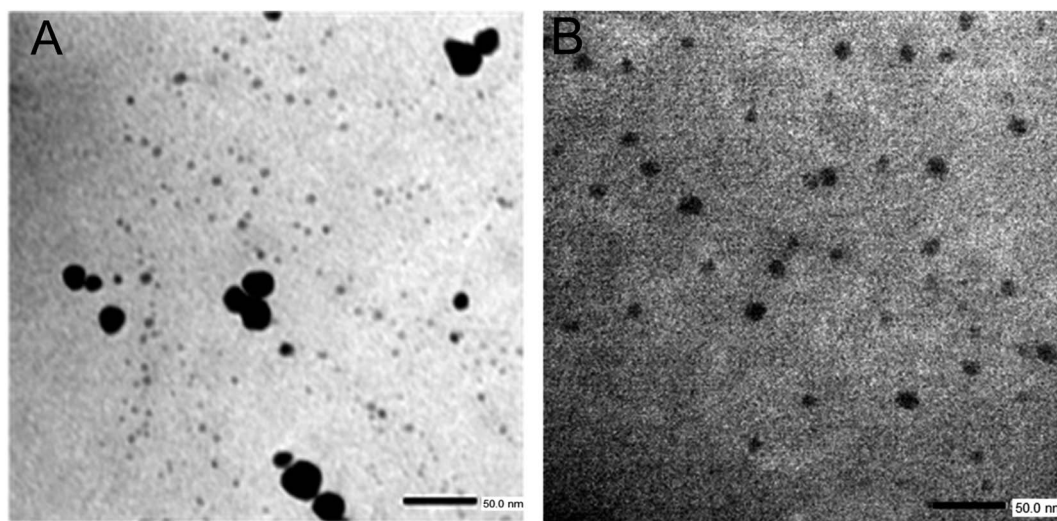


Fig. 7 TEM images of nanomaterials. (A) Au-PNIPAM NCs and (B) Au-PAAM NCs.







Fig. 8 Fluorescence spectra of (A) pure PNIPAM (black line), non-irradiated Au<sup>3+</sup>-PNIPAM (red line), irradiated Au-PNIPAM NCs (blue line). (B) Pure PAAM (black line), non-irradiated Au<sup>3+</sup>-PAAM (red line), and irradiated Au-PAAM NCs (blue line). Excitation wavelength: 535 nm.

As it is known Au-NPs do not exhibit fluorescence because the optical properties of Au-NPs are defined as collective electrons oscillations of hundreds of atoms supporting a surface plasmon (Fig. 1A), thus it is expected that hydrogels impregnated with Au-NPs will not fluoresce either. Therefore, the fluorescence of the materials is due to the formation of AuNCs in the UV-irradiated hydrogels. AuNCs consist of several to a few tens of Au atoms with a small size. Hence, cannot be observed in TEM and SEM images. It is well known that, when the size of the metal system is sufficiently reduced, quantum confinement effects appear in the material and make new optical and chemical properties arise.<sup>31,51</sup> The main effect of quantum confinement in metal nanoclusters is the appearance of HOMO–LUMO energy gaps due to a discretization of the energy levels within the nanocluster that behaves as a molecule-like system.<sup>52</sup> This energy discretization is responsible for molecular properties such as fluorescence. The origin of metal nanoclusters fluorescence has been attributed to the ligand-to-metal charge transfer or ligand-to-metal–metal charge transfer.<sup>53</sup> Therefore, the obtained results prove that the observed fluorescence originates from the gold core of the AuNCs supported on the PNIPAM and PAAM hydrogel. Also, the results demonstrate that the AuNCs are formed by photoreduction of the Au<sup>3+</sup> cations loaded inside of the hydrogel upon UV light

irradiation, as it has been previously shown for similar systems.<sup>54,55</sup>

In order to study deeply the spectroscopic properties of the new materials, the excitation–emission matrix (EEM) of both, Au-PNIPAM NCs and Au-PAAM NCs, were determined and are shown in Fig. 9. The EEM fluorescence spectroscopy enables the identification and characterization of complex mixtures of nanoclusters.<sup>51,56</sup> Using EEM spectroscopy, it is possible to visualize the complete fluorescence profile of each of the obtained cluster species. The EEMs recorded for both materials, Au-PNIPAM NCs and Au-PAAM NCs, show an island peaking at  $\lambda_{exc} = 520$  nm and  $\lambda_{em} = 612$  nm superimposed on a continuous background observed over a wide range of excitation–emission wavelengths. The continuous background is most likely due to the light scattering produced by the Au-NPs, while the islands at  $\lambda_{exc}/\lambda_{em} = 520/612$  nm are due to the actual fluorescence of the AuNCs incorporated into the materials. The Au-NPs supported on PNIPAM and PAAM polymer matrices show a SPR at 540 nm and 544 nm, respectively, as observed in the UV absorption spectra (Fig. 1A). However, the excitation wavelength at the maximum fluorescence of the AuNCs is blue shifted at  $\lambda_{exc} = 520$  nm (Fig. 9) and is not observed in the corresponding UV spectra. These results indicate that both, Au-NPs and AuNCs are produced upon UV irradiation of the Au<sup>3+</sup>-PNIPAM and Au<sup>3+</sup>-

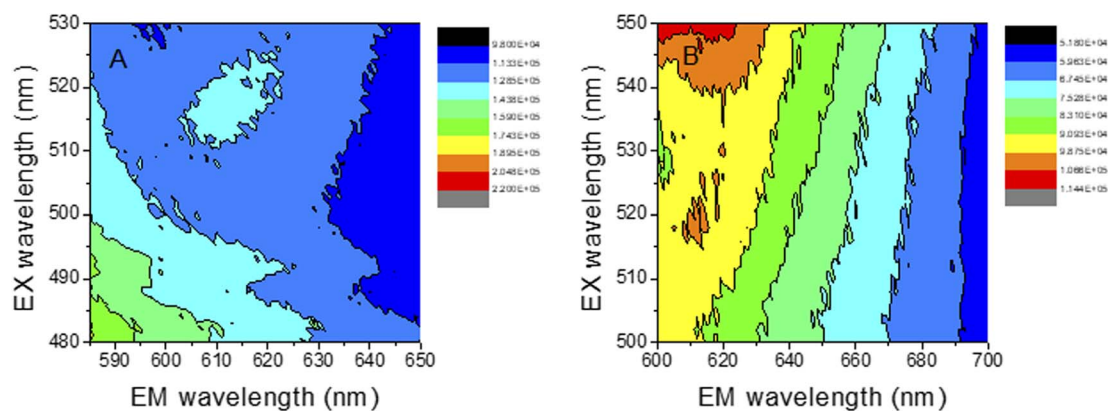


Fig. 9 AuNCs emission–excitation matrices, recorded at different emission and excitation wavelengths. (A) Au-PNIPAM NCs (B) Au-PAAM NCs.



PAAM polymer matrices. In addition, the fact that only the SPR of the Au-NPs is seen in the UV spectra (Fig. 1A) strongly suggests that AuNCs are a minor product of the synthesis compared to Au-NPs. Furthermore, the fluorescence quantum yield of the AuNCs is high enough to allow the detection of these minority species by fluorescence spectroscopy.

### Heavy metals detection

Liu *et al.* have shown that the fluorescent material (Ag-PMAA) based on silver NCs and poly(methacrylic acid) can be used to sense  $\text{Cr}^{3+}$  since this cation selectively quenches the clusters fluorescence in solution.<sup>57</sup> The materials reported in these work contain Au instead of Ag and different polymers that allow studying the quenching of the fluorescence by different heavy metal cations in the hydrogel. Also, the present work the AuNC are incorporated inside of the matrix polymer allowing to build a portable device to detect the contaminant. The performance of these materials as heavy metal sensors is estimated by the fluorescence quenching measurement.

As a probe of concept to test the performance of the new materials, heavy metal ions ( $\text{Hg}^{2+}$ ) were added to each Au-matrix NCs to test the AuNCs response. The irradiated samples were incubated for 30 min with  $\text{Hg}^{2+}$  solutions of different concentrations and the excitation was performed at 535 nm. The quenching of the fluorescence by  $\text{Hg}^{2+}$  was evaluated as the fluorescence intensity decay ( $I_0/I$ ) of AuNCs after ( $I$ ) and before ( $I_0$ ) adding metal ions at  $\lambda_{\text{em}} = 618$  nm. Fig. 10 shows the Au-PNIPAM NCs and Au-PAAM NCs fluorescence spectra in the presence of different concentrations of  $\text{Hg}^{2+}$  ions. As shown in Fig. 10A and B, the emission intensity of AuNCs is gradually reduced with increasing the  $\text{Hg}^{2+}$  concentration in the solution (pH: 7). A good linear Stern–Volmer correlation can be obtained ( $R = 0.999$  and  $R = 0.954$  for Au-PNIPAM and Au-PAAM NCs, respectively) based on the  $\text{Hg}^{2+}$  ions quenching effects over the range 10–100 nM, as indicated in the inset in Fig. 10. The limits of detection (LD) were calculated as  $\text{LD} = 3\sigma_b/m$ , where  $\sigma_b$  and  $m$  are the standard deviation of the blank and the slope of the Stern–Vomer graph, respectively. The LDs were determined to

be 4.1 nM and 1.3 nM for Au-PNIPAM and Au-PAAM NCs, respectively.

We notice that the presence of  $\text{Hg}^{2+}$  changes the absorbance of nanomaterials at 540 nm (Fig. S3<sup>†</sup>). This result would indicate that the fluorescence quenching is associated with an static quenching mechanism due to the formation of a complex between  $\text{Hg}^{2+}$  and the AuNCs. It is important to note that our results are in agreement with previous results by Chen *et al.*<sup>9</sup> These authors reported that the quenching of the AuNCs synthesized on PNIPAM microgels is produced by the deposition of  $\text{Hg}^{2+}$  ions on the surfaces of the AuNCs supported on the matrix. The possible formation of an Hg–Au amalgam would lead to an aggregation of the AuNCs and, consequently, to a progressive extinction of the fluorescent species with increasing  $\text{Hg}^{2+}$  concentration.

The selectivity of the assay was evaluated by challenging it with other environmentally relevant metal cations ( $\text{Na}^+$ ,  $\text{K}^+$ ,

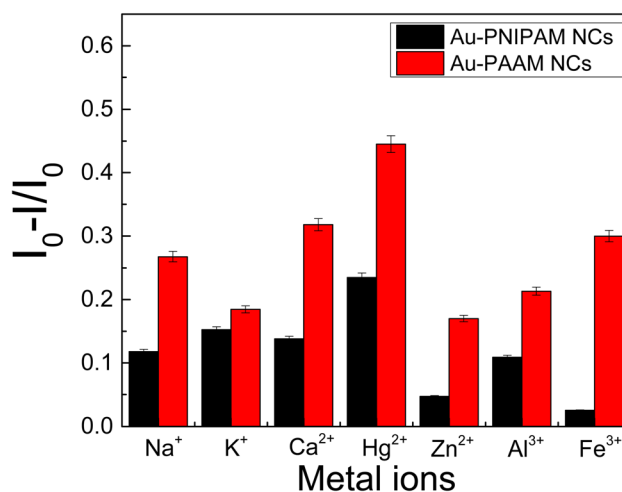


Fig. 11 Selectivity of Au-PNIPAM NCs (black bar) and Au-PAAM NCs (red bar) to  $\text{Hg}^{2+}$ . Plots of relative fluorescence intensity spectra [ $(I_0 - I)/I_0$ ] of the Au-NIPAM NCs and Au-PAAM NCs in the presence of  $\text{Hg}^{2+}$  (20 nM) and various metal ions (20 nM) in water at pH: 7, incubation time 30 min.



Fig. 10 Sensitivity of the (A) Au-PNIPAM NCs and (B) Au-PAAM NCs for  $\text{Hg}^{2+}$  at pH: 7 ( $\lambda_{\text{exc}} = 535$  nm). Initial fluorescence of the AuNCs (black line).  $\text{Hg}^{2+}$  concentrations: 10 nM (red line), 15 nM (blue line), 20 nM (pink line), 50 nM (board line), 100 nM (dark blue line) and 200 nM (green line) inset: Stern–Volmer plot ( $I_0/I$  vs.  $[\text{Hg}^{2+}]$ ) over the concentration range 10–100 nM. For the Stern–Volmer plot, the fluorescence intensity was determined at  $\lambda_{\text{em}} = 618$  nm and the reported values correspond to the average of three replicated measurements ( $\text{RSD} \leq 4\%$ ).



Table 2 Comparison of analytical parameters for detection of Hg<sup>2+</sup> at several methods

Technique	Nanomaterial	Stabilizer/support	Linear range	Detection limit (nM)	Ref.
Fluorescence	AuNCs	PNIPAM	10–100 nM	4.1 nM	This work
Fluorescence	AuNCs	PAAM	10–100 nM	1.3 nM	This work
Fluorescence	AuNCs	Cytidine	0.01–6 μM	30 nM	58
Fluorescence	AuNCs	Ovalbumin	0–10 μM	10 nM	59
Fluorescence	AuNCs	PNIPAM microgels	2–20 nM	1.7 nM	9
Fluorescence	NCQDs <sup>a</sup>	MGDA-MPD <sup>a</sup>	0–100 μM	0.9 μM	60
Colorimetric	AgNPs	Gelatin and Tween-20	0.01–0.5 mM	2.25 μM–0.65 μM	61

<sup>a</sup> NCQDs: nitrogen-doped carbon quantum dots. MGDA: methyl glycine diacetic acid trisodium salt, MPD: *m*-phenylenediamine.

Ca<sup>2+</sup>, Zn<sup>2+</sup>, Al<sup>3+</sup> and Fe<sup>3+</sup>). In this sense, the effect of each of these metal cations on the fluorescence quenching was evaluated by adding of 20 nM of the metal cations to samples containing AuNCs and 20 nM of Hg<sup>2+</sup> ions. The corresponding fluorescence spectra are shown in Fig. S4 and S5† and the plot of  $[I_0 - I]/I_0$  for each metal cation is shown in Fig. 11.

For both nanomaterials (Au-PNIPAM NC and Au-PAAM NC), the larger quenching effect is observed for Hg<sup>2+</sup> ions, although the selectivity is not as good as in other materials previously reported.<sup>9</sup> However, it must be noticed that the goal of this work was to develop a new platform to detect Hg<sup>2+</sup> directly from the tap or any course of natural water without any previous treatment and is the simplest way to be used as a first probe for drinkable water conditions in remote regions. In this sense, none buffer is used to adjust the pH to the optimum quenching fluorescence response and this fact could lead to the precipitation of the hydroxides of some of the metal cations probed which could mask the selectivity of the system.

Nevertheless, the detection limit of Hg<sup>2+</sup> by Au-PNIPAM NC and Au-PAAM NC are comparable and even lower than for other AuNCs, especially those using biomolecules for the synthesis of AuNC (see Table 2). In addition, the detection limits of the materials presented in this work are below the value recommended by the WHO for drinkable water.<sup>3</sup> Consequently, the AuNCs synthesized in this work reached acceptable validation parameters, exhibiting a good linear range and detection limit for been used as a first probe to ensure Hg<sup>2+</sup> free water, even though the selectivity of the method is not as good.

Finally, the results reveal that Au-PNIPAM NCs are more selective and less sensitive than Au-PAAM, while Au-PAAM are more sensitive and less selective for detecting Hg<sup>2+</sup> ions in water at pH: 7. In this regard, we consider that Au-PNIPAM NCs at pH: 7 are more adequate to detect Hg<sup>2+</sup> since they are more selective and the detection limit is still below the value recommended by the WHO.

## Conclusion

In this work, AuNCs were successfully synthesized and supported on two different polymeric matrices PNIPAM and PAAM. The materials were characterized by UV-Vis, FT-IR, fluorescence spectroscopy, TEM and SEM. Likewise, the thermal properties and the swelling capacity of the different polymer matrices were determined. The results reveal that the incorporation of Au<sup>3+</sup> in the PNIPAM and PAAM hydrogels modifies the structure and

physicochemical properties of the material. The Au<sup>3+</sup> presence in the hydrogels favors the water incorporation inside the polymeric network because it opens the channels and pores of the hydrogel. Moreover, the supported nanomaterial fluorescent properties (PNIPAM NCs and PAAM NCs) were determined. It is demonstrate that AuNCs are only formed when the Au<sup>3+</sup> incorporated material is irradiated with UV light. Also, the Au-PNIPAM NCs and Au-PAAM NCs exhibit fluorescence emission at *ca.* 620 nm when they are excited at 535 nm. In addition, our study demonstrates that the AuNCs supported on each polymeric matrix present a unique characteristic and optical signature. Furthermore, the AuNCs supported on the polymer matrices were used for the detection of Hg<sup>2+</sup> in water at pH: 7. The AuNCs showed a remarkably rapid response and high sensitivity for Hg<sup>2+</sup> and a moderate but higher selectivity as compared to other metal cations. The Stern–Volmer relationship graphs were linear over the range of 10–100 nM, and the corresponding LD was 4.1 nM and 1.3 nM for Au-PNIPAM and Au-PAAM NCs, respectively.

In summary, this research work shows an easy, one pot synthesis and characterization of new applied materials that are good candidates as potential material for detecting Hg<sup>2+</sup> in water at pH: 7 and it is worth exploring the possibility to build a portable device in the future.

## Conflicts of interest

There are no conflicts to declare.

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## References

- 1 E. E. Mariño, G. T. Ávila, P. Bhattacharya and C. J. Schulz, *J. South Am. Earth Sci.*, 2020, **100**, 102547.
- 2 C. Bilos, L. M. Tatone, C. N. Skorupka, M. J. Astoviza, C. L. Romero and J. C. Colombo, *Int. J. Environ. Health*, 2019, **9**(3), 271–287.



- 3 K. P. Lisha, A. Anshup and T. Pradeep, *Gold Bull.*, 2009, **42**(2), 144–152.
- 4 B. R. M. Alsehli, *Egypt. J. Chem.*, 2021, **64**(2), 869–881.
- 5 Z. Yan, Y. Cai, J. Zhang and Y. Zhao, *Measurement*, 2022, **187**, 110355.
- 6 J. P. Wilcoxon and B. L. Abrams, *Chem. Soc. Rev.*, 2006, **35**(11), 1162–1194.
- 7 J. Qiao, X. Mu, L. Qi, J. Deng and L. Mao, *Chem. Commun.*, 2013, **49**(73), 8030–8032.
- 8 H. Jiang, Z. Qin, Y. Zheng, L. Liu and X. Wang, *Small*, 2019, **15**(18), 1901170.
- 9 L.-Y. Chen, C.-M. Ou, W.-Y. Chen, C.-C. Huang and H.-T. Chang, *ACS Appl. Mater. Interfaces*, 2013, **5**(10), 4383–4388.
- 10 X.-P. Zhang, K.-Y. Huang, S.-B. He, H.-P. Peng, X.-H. Xia, W. Chen and H.-H. Deng, *J. Hazard. Mater.*, 2021, **405**, 124259.
- 11 M. Cui, Y. Zhao and Q. Song, *TrAC, Trends Anal. Chem.*, 2014, **57**, 73–82.
- 12 E. Caló and V. V. Khutoryanskiy, *Eur. Polym. J.*, 2015, **65**, 252–267.
- 13 M. V. Martinez, M. Molina and C. A. Barbero, *J. Phys. Chem. B*, 2018, **122**(38), 9038–9048.
- 14 C. A. Barbero, M. V. Martínez, D. F. Acevedo, M. A. Molina and C. R. Rivarola, *Macromol*, 2022, **2**(3), 440–475.
- 15 M. V. Martinez, M. A. Molina, S. B. Abel and C. A. Barbero, *MRS Adv.*, 2018, **3**(63), 3735–3740.
- 16 V. Packirisamy and P. Pandurangan, *BioTechniques*, 2022, **74**, 51–62.
- 17 P. Thoniyot, M. J. Tan, A. A. Karim, D. J. Young and X. J. Loh, *Advanced Science*, 2015, **2**(1–2), 1400010.
- 18 T.-H. Yang, *Recent Patents on Materials Science*, 2008, pp. 29–40.
- 19 M. A. Molina, C. R. Rivarola, M. C. Miras, D. Lescano and C. A. Barbero, *Nanotechnology*, 2011, **22**(24), 245504.
- 20 O. Okay, G. Gerlach and K.-F. Arndt, *Hydrogel Sensors and Actuators*, 2009, vol. 6, pp. 165–195.
- 21 K. S. Anseth, C. N. Bowman and L. Brannon-Peppas, *Biomaterials*, 1996, **17**(17), 1647–1657.
- 22 H. Feng, N. Tang, M. An, R. Guo, D. Ma, X. Yu, J. Zang and N. Yang, *J. Phys. Chem. C*, 2019, **123**(51), 31003–31010.
- 23 M. V. Martinez, R. C. Rodriguez, M. M. Bruno, D. F. Acevedo and C. A. Barbero, *J. Electrochem. Soc.*, 2018, **165**(16), H1021–H1027.
- 24 M. V. Martínez, C. R. Rivarola, M. C. Miras and C. A. Barbero, *Sens. Actuators, B*, 2017, **241**, 19–32.
- 25 J. Pereyra, M. V. Martinez, C. Barbero, M. Bruno and D. Acevedo, *J. Compos. Sci.*, 2019, **3**(1), 1–14.
- 26 N. Dave, M. Y. Chan, P.-J. J. Huang, B. D. Smith and J. Liu, *J. Am. Chem. Soc.*, 2010, **132**(36), 12668–12673.
- 27 X. Lei, H. Li, Y. Luo, X. Sun, X. Guo, Y. Hu and R. Wen, *J. Taiwan Inst. Chem. Eng.*, 2021, **123**, 79–86.
- 28 J. L. MacLean, K. Morishita and J. Liu, *Biosens. Bioelectron.*, 2013, **48**, 82–86.
- 29 J.-S. Shen, Y.-L. Chen, Q.-P. Wang, T. Yu, X.-Y. Huang, Y. Yang and H.-W. Zhang, *J. Mater. Chem. C*, 2013, **1**(11), 2092–2096.
- 30 Z. Zhao and Y. Li, *Colloids Surf., B*, 2020, **195**, 111244.
- 31 L.-Y. Chen, C.-W. Wang, Z. Yuan and H.-T. Chang, *Anal. Chem.*, 2015, **87**(1), 216–229.
- 32 O. Ozay, N. Aktas, E. Inger and N. Sahiner, *Int. J. Hydrogen Energy*, 2011, **36**(3), 1998–2006.
- 33 A. Contin, A. Biffis, S. Sterchele, K. Dörmbach, S. Schipmann and A. Pich, *J. Colloid Interface Sci.*, 2014, **414**, 41–45.
- 34 H. A. H. Alzahrani, M. A. Buckingham, W. P. Wardley, R. D. Tilley, N. Ariotti and L. Aldous, *Chem. Commun.*, 2020, **56**(8), 1263–1266.
- 35 H. Dong, J. F. Snyder, D. T. Tran and J. L. Leadore, *Carbohydr. Polym.*, 2013, **95**(2), 760–767.
- 36 X.-Q. Wu, X.-W. Wu, Q. Huang, J.-S. Shen and H.-W. Zhang, *Appl. Surf. Sci.*, 2015, **331**, 210–218.
- 37 F. Wahid, C. Zhong, H.-S. Wang, X.-H. Hu and L.-Q. Chu, *Polymers*, 2017, **9**(12), 636.
- 38 M. A. Molina, C. R. Rivarola and C. A. Barbero, *Eur. Polym. J.*, 2011, **47**(10), 1977–1984.
- 39 S. Mourdikoudis, R. M. Pallares and N. T. K. Thanh, *Nanoscale*, 2018, **10**(27), 12871–12934.
- 40 G. A. Monti, G. A. Fernández, N. M. Correa, R. D. Falcone, F. Moyano and G. F. Silbestri, *R. Soc. Open Sci.*, 2017, **4**(7), 170481.
- 41 G. A. Monti, N. M. Correa, R. D. Falcone, G. F. Silbestri and F. Moyano, *ChemistrySelect*, 2020, **5**(36), 11264–11271.
- 42 D. W. Mayo, in *Course Notes on the Interpretation of Infrared and Raman Spectra*, 2004, pp. 33–72.
- 43 I. J. D. Ebenezer, S. Ramalingam, C. R. Raja and P. C. J. Prabakar, *J. Nanotechnol. Adv. Mater.*, 2014, **2**, 11–25.
- 44 K. Kusolkamabot, P. Sae-ung, N. Niamnont, K. Wongravee, M. Sukwattanasinitt and V. P. Hoven, *Langmuir*, 2013, **29**(39), 12317–12327.
- 45 X.-F. Sun, Q. Zeng, H. Wang and Y. Hao, *Cellulose*, 2019, **26**(3), 1909–1922.
- 46 J. Yin and B. Deng, *J. Membr. Sci.*, 2015, **479**, 256–275.
- 47 H. G. Schild, *Prog. Polym. Sci.*, 1992, **17**(2), 163–249.
- 48 G. Gombár, D. Ungor, G. F. Samu, O. Dömötör and E. Csapó, *J. Mol. Liq.*, 2022, **359**, 119372.
- 49 L. Xi, X. Zhang, Y. Chen, J. Peng, M. Liu, D. Huo, G. Li and H. He, *Chem. Phys. Lett.*, 2022, **807**, 140085.
- 50 H. Zhuang, X. Jiang, S. Wu, S. Wang, Y. Pang, Y. Huang and H. Yan, *Sci. Rep.*, 2022, **12**(1), 6624.
- 51 M. I. Taccone, R. A. Fernández, F. L. Molina, I. Gústín, C. G. Sánchez, S. A. Dassie and G. A. Pino, *Phys. Chem. Chem. Phys.*, 2020, **22**(29), 16813–16821.
- 52 K. Kwak and D. Lee, *Acc. Chem. Res.*, 2019, **52**(1), 12–22.
- 53 N. Cao, J. Xu, H. Zhou, Y. Zhao, J. Xu, J. Li and S. Zhang, *Microchem. J.*, 2020, **159**, 105406.
- 54 L. E. Mulko, M. Rossa, J. P. Aranguren-Abrate and G. A. Pino, *Appl. Surf. Sci.*, 2019, **485**, 141–146.
- 55 F.-N. Wu, J. Zhu, G.-J. Weng, J.-J. Li and J.-W. Zhao, *J. Mater. Chem. C*, 2022, **10**(40), 14812–14833.
- 56 H. Ramsay, D. Simon, E. Steele, A. Hebert, R. D. Oleschuk and K. G. Stamplecoskie, *RSC Adv.*, 2018, **8**(73), 42080–42086.
- 57 S. Liu, F. Lu and J. J. Zhu, *Chem. Commun.*, 2011, **47**(9), 2661–2663.



## Paper

- 58 Y. Zhang, H. Jiang and X. Wang, *Anal. Chim. Acta*, 2015, **870**, 1–7.
- 59 H. Shi, M. Y. Ou, J. P. Cao and G. F. Chen, *RSC Adv.*, 2015, 5(105), 86740–86745.
- 60 G. Wang, S. Zhang, J. Cui, W. Gao, X. Rong, Y. Lu and C. Gao, *Spectrochim. Acta, Part A*, 2023, **286**, 122010.
- 61 L. Sulistiawaty, S. Sugiarti and N. Darmawan, *Indones. J. Chem.*, 2015, **15**(1), 1–8.

