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Rational design of a multifunctional molecular dye for dual-modal NIR-II/photoacoustic imaging and photothermal therapy†

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Small-molecule based multifunctional probes play significant roles in biomedical science and possess high clinical translational ability. However, the preparation of these promising probes without complicated synthetic procedures remains a challenging task. Herein, we rationally designed a high-performance DD–A–DD scaffold molecular dye (SYL) with an intrinsic multifunctional ability and then incorporated it into DSPE-mPEG5000 to facilely construct biocompatible NIR-II fluorescent and photoacoustic (PA) dual-modal theranostic nanoprobes (SYL NPs) (~120 nm). *In vivo* studies confirmed that SYL NPs exhibited bright NIR-II fluorescence and PA signals in the tumor region with a promising signal to background ratio (S/B). Meanwhile, SYL NPs demonstrated significantly inhibited tumor growth under laser irradiation with no noticeable side effects. These promising results highlighted SYL NPs as a potential theranostic platform for cancer diagnosis (NIR-II region) and therapy.

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Introduction

Multifunctional fluorescent probes including multimodal imaging and theranostic probes have evolved into fast-growing research tools with goals of multidimensional visualization of early events in cancer diagnostics and assessment of therapeutic efficacy *in vivo*.^{1–8} Considering the translational potential of fluorescent probes in preclinical and clinical practice, small-molecule multifunctional probes remain as exceptional candidates owing to their high biocompatibility, rapid metabolism, and well-defined architectures.^{9–13} However, the widespread applications of such promising probes in biomedicine are heavily hampered as introducing different functional molecules into the molecular platform usually involves abundant and complex chemistry including multiple synthetic steps, tedious chromatographic separations, and issues of chemo-selectivity.^{14–17} Hence, continuous efforts should be made to design simple fluorophores with intrinsic multifunctional ability,

which have high significance and direct impact on the field of fluorescence imaging and biomedicine.^{18,19}

Over the past several years, fluorescence based imaging in the second near-infrared channel (NIR-II, 1000–1700 nm) has gained attention for its prominent merits and surpasses the conventional visible/NIR-I channels (700–900 nm) in terms of lower tissue scattering and minimal auto-fluorescence, thus leading to significant advances in imaging qualities including image depth and spatiotemporal resolution.^{20–38} More recently, based on a benzobisthiadiazole (BBTD) acceptor, a series of small molecular dyes with a donor–acceptor–donor (D–A–D) architecture have been developed. The reduced energy gap between the hybridized highest occupied molecular orbital (HOMO) and lowest unoccupied molecular orbital (LUMO) levels in these molecular systems shifted the fluorescence emission spectrum from the NIR-I region to the NIR-II channel.^{39–44} Benefiting from their excellent photo-stability and fluorescence features, diverse biological practices such as vascular/lymphatic and tumor fluorescence imaging with a high signal to background ratio (S/B) as well as precise fluorescence image-guided surgery indeed bring good news for biomedical applications in the NIR-II channel.^{45–60} Despite these excellent fluorescence imaging results, the potential utilization of these molecular dyes for multimodal imaging or even cancer theranostics is rarely explored.

In this study, an intrinsic multifunctional and high-performance dye (SYL) based on a DD–A–DD scaffold was rationally designed and facilely prepared (Fig. 1). The designed probe is expected to present the following advantages. Compared with most existing D–A–D based molecular dyes, SYL could achieve better fluorescence performance. This is due to the enhanced

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Fig. 1 Schematic illustration of the preparation of SYL NPs, as well as their multifunctional applications in NIR-II/PA dual-modal imaging and photothermal therapy.

electron-density and molar extinction coefficient due to the addition of a secondary electron donor (diphenylamine) to the molecular scaffold.⁶¹ Moreover, the dialkyl substituted fluorene moiety (first donor) inserted between the second donor and thiophene could extend the π -conjugated bridge and facilitate intramolecular charge transfer (ICT) (Fig. 1).⁶² 3,4-Ethylene dioxithiophene employed here as the spacer to distort the conjugated backbones could also minimize the intermolecular interactions as well as energy transfer from the molecular excited states to the nearby H_2O molecules.⁶³ The fluorophore SYL with substituted fluorene as the first donor and diphenylamine as the second donor showed the optimized optical performance with strong absorption around 800 nm. This result suggested that under 808 nm laser irradiation, the optical energy absorbed by SYL could be partly converted into heat to increase the temperature of the surrounding environment and could be utilized to trigger cancer cell death by highly effective and noninvasive photothermal therapy (PTT).^{64,65} In addition, the photothermal effect of SYL generated acoustic waves that could also be detected and converted into photoacoustic (PA) imaging signals. As a hybrid modality, the dual-modal NIR-II/PA imaging meets the requirements of both the superior S/B (signal to background) ratio and high penetration depth for delineating tumor margins, as well as determining the best photothermal treatment time after administration to realize an optimal photothermal effect in cancer therapy.⁶⁵ Therefore, the small molecular dye SYL could effectively integrate dual-modal imaging and photothermal therapeutic functions for cancer theranostics (Fig. 1).

To further endow SYL with excellent solubility and biocompatibility and high preferential passive accumulation at the tumor site, SYL was incorporated into a PEGylated surfactant DSPE-mPEG5000 to form nanoparticles (SYL NPs). Based on SYL NPs, *in vivo* dual-modal NIR-II fluorescence/PA tumor imaging and precise PTT have been performed with excellent results, which

could speed up the utilization of small molecular dyes based on D-A-D scaffolds for biomedical applications in near future.

Results and discussion

A new type of NIR-II small molecular dye, SYL, was designed by using an optimized DD-A-DD scaffold including 3,4-ethylenedioxythiophene and dialkylfluorene and diphenylamine donor units; both contributed to enhancing the fluorescence signals as compared to the previously reported D-A-D dye H1 (Fig. 2).⁴⁶ SYL was synthesized through a convergent route in 25% overall yield in 9 steps (Fig. 1) and well-characterized by $^1H/^{13}C$ -NMR and MALDI-TOF-MS (Fig. S23–S25†). Density functional theory (DFT) calculations with the B3LYP (Gaussian 09) functional using the 6-31G basis set were employed for calculating the HOMO and LUMO of both H1 and SYL, respectively. As shown in Fig. 2, the energy bandgap of SYL (1.36 eV) was higher than that of H1 (1.17 eV).

SYL demonstrated a hypsochromic shift in absorption and emission wavelengths as well as a higher NIR-II fluorescence

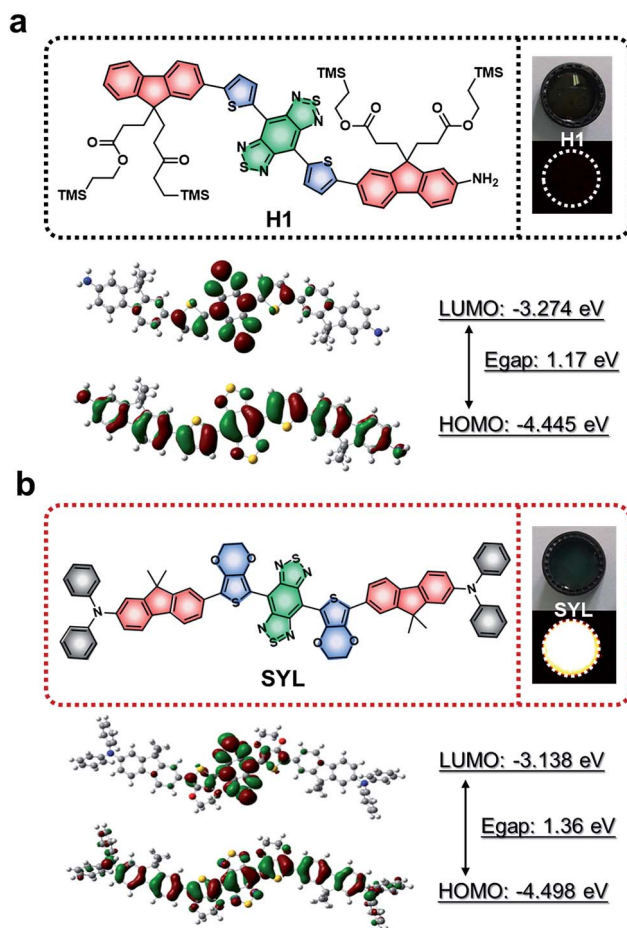


Fig. 2 Chemical structures and electronic properties of SYL and the previously reported NIR-II dye H1; (a) Frontier orbitals of H1, its associated energies (DFT using B3LYP Gaussian 09), and NIR-II fluorescence signals. (b) Chemical structures, NIR-II fluorescence signals and Frontier orbitals with associated energies of the novel NIR-II dye SYL.



signal output compared to **H1**. To enhance the capability of **SYL** for *in vivo* applications, **SYL** was incorporated into DSPE-mPEG5000 (1 : 9, w/w) for facile preparation of **SYL** NPs with $81.6 \pm 2.4\%$ encapsulation efficiency (Fig. S1†). The absorption and emission wavelengths of **SYL** NPs were detected at ~ 765 nm and ~ 976 nm, respectively (Fig. 3a). Based on transmission electron microscopy (TEM) and dynamic light scattering (DLS) results, the as-synthesized **SYL** NPs were discrete and uniform with an average size of ~ 120 nm, suggesting high monodispersity (Fig. 3b). The stability of the **SYL** NPs was tested in phosphate-buffered saline (PBS) containing 10% (v/v) fetal bovine serum (FBS) at 37°C . Even after 48 h, there was no apparent change in the particle-diameter distribution, suggesting the excellent *in vitro* stability of **SYL** NPs (Fig. S2†). Furthermore, the FDA-approved NIR-I theranostic agent

indocyanine green (ICG) was also incorporated into DSPE-mPEG5000 to serve as the control group, ICG NPs. **SYL** NPs exhibited high photo-stability with a negligible decay as compared with ICG NPs under continuous irradiation at 808 nm for 1 h (Fig. 3c). Also, **SYL** in DMSO and **SYL** NPs in different media did not show any significant change in fluorescence signal intensity upon irradiation (1 W cm^{-2}) up to 1 h (Fig. S3–S4†). The PA spectrum showed a broad band extending over a wide NIR-I region (700–900 nm, Fig. S5†) and hence **SYL** NPs could furnish a promising PA signal under 760 nm irradiation (Fig. S5†). Moreover, the PA amplitudes of **SYL** NPs showed excellent linearity with their molar concentration in a range from 0 to $100\text{ }\mu\text{g mL}^{-1}$ based on **SYL** (Fig. 3d).

The photothermal properties of **SYL** NPs were next tested using different power laser irradiations (1 to 2 W cm^{-2} , 808 nm). A significant temperature enhancement was observed, suggesting the potential of **SYL** NPs for photothermal therapy (Fig. S6†). In addition, the temperature was found to increase more rapidly with the increased concentrations of **SYL** NPs (0 to $50\text{ }\mu\text{g mL}^{-1}$) under 808 nm laser irradiation (power 1 W cm^{-2}) (Fig. 3e). The temperature of **SYL** NPs increased from 25.0°C to 68.2°C within 180 s of irradiation while there was no significant temperature rise in the control (PBS) group under the same conditions (Fig. 3e). The change in temperature upon laser irradiation was also monitored by using an infrared thermal camera (Fig. S7†). **SYL** NPs exhibited a reversible photothermal heating and natural cooling phenomenon up to five cycles without any significant change in photothermal conversion efficiency, suggesting their good photothermal stability (Fig. 3f). Compared with **SYL** NPs, ICG NPs were not stable enough under irradiation (Fig. 3f and S8†). The photothermal conversion efficiency of **SYL** and **SYL** NPs was $\sim 22.3\%$ and $\sim 21.8\%$, respectively (Fig. S9–S10 and the equation in the ESI†), higher than that of ICG (15.1%).⁶⁶

Given the excellent photothermal features, next we evaluated the therapeutic efficacy of **SYL** NPs in breast cancer 4T1 cells with or without laser irradiation. The therapeutic effect of **SYL** NPs was examined quantitatively by using CCK8 assay. As shown in Fig. 3g, more than 90% of cells were dead upon treatment with $20\text{ }\mu\text{g mL}^{-1}$ **SYL** NPs under 808 nm laser irradiation (power 1 W cm^{-2}). There was only a slightly higher effect on cell death as the concentration of **SYL** was increased from 20 to $40\text{ }\mu\text{g mL}^{-1}$. In contrast, no obvious cytotoxicity was observed in cells with **SYL** NPs up to $40\text{ }\mu\text{g mL}^{-1}$ concentration without light irradiation. Further, **SYL** NP mediated photothermal cytotoxicity was also demonstrated with calcein-AM and PI staining assays (Fig. 3h). The effect of inducing cancer cell death in 4T1 cells with the combination of **SYL** NPs and laser-irradiation *in vitro* was much stronger than that of **SYL** NPs or laser irradiation alone, showing their potential as an efficient phototherapeutic agent *in vivo*.

Next, we evaluated the fluorescence imaging (NIR-II) performance of **SYL** NPs *in vivo*, with ICG NPs as the control. Lymphatic system imaging was performed on living mice (Fig. S11a and b†). Compared to ICG NPs, the lymphatic vessel between the injection site and sentinel lymph node was clearly distinguished and exhibited a high S/B ratio after injection of **SYL** NPs, thus providing fast and precise mapping of the



Fig. 3 *In vitro* NIR-II, PA, photothermal properties and cell viabilities of **SYL** NPs. (a) UV-Vis-NIR absorption and fluorescence emission of **SYL** NPs; (b) DLS data and TEM image (inset) of **SYL** NPs. Scale bar: 200 nm; (c) photostability of **SYL** NPs and ICG NPs under continuous irradiation at 808 nm (1 W cm^{-2}) for 1 h; (d) PA intensity of **SYL** NPs as a function of concentration from 0 to $100\text{ }\mu\text{g mL}^{-1}$; (e) photothermal conversion behavior of **SYL** NPs in PBS solution at various concentrations under laser irradiation (1 W cm^{-2}); (f) the photothermal heating and natural cooling cycles of **SYL** NPs and ICG NPs in PBS solution ($25\text{ }\mu\text{g mL}^{-1}$, 1 W cm^{-2}); (g) cell viabilities of 4T1 cancer cells after incubation with **SYL** NPs with or without laser irradiation; (h) confocal fluorescence images of live (green) and dead (red) 4T1 cells with calcein-AM and PI staining after various treatments. Scale bar: $100\text{ }\mu\text{m}$.



lymphatic system by NIR-II fluorescence imaging. In addition, successful imaging of blood vessels was observed from the surrounding background normal tissue after injection of **SYL** NPs as compared with ICG NPs (Fig. S11c and d†).

Based on the above results, the efficacy of **SYL** NPs for passive targeting and dual-modal imaging of 4T1 tumors *in vivo* was further investigated. After the 4T1 tumor-bearing nude mice ($n = 3$) were intravenously injected with 100 μg of **SYL** NPs in PBS, NIR-II fluorescence images (Series III 900/1700 equipment, NIR-Optics Tech. Co. Ltd., Suzhou) and PA signals of living mice were constantly monitored. From both NIR-II fluorescence and PA images at 2, 8, 12 and 24 h post-injection, the tumor was clearly visualized from the surrounding background tissues and showed passive uptake of **SYL** NPs at all time points (Fig. 4a). The semi-quantitative analysis of both NIR-II and PA images further revealed that the tumor uptake of **SYL** NPs gradually increased with time and reached a maximum at 8 h (Fig. 4b). Thereafter, the signal from the localized **SYL** NPs at the tumor region site reduced gradually during 8–24 h. As seen in Fig. 4c, the S/B ratio of both NIR-II and PA imaging increased steadily in the first 2 h and reached a maximum value at 8 h post-injection. To further confirm this, *ex vivo* imaging of tumors and other major organs/tissues was performed 24 h post-injection. The NIR-II fluorescence signals in the excised tumor, liver and spleen were much brighter than those of other organs (Fig. S12†). These findings were consistent with the *in vivo* imaging results, demonstrating that **SYL** NPs possessed favorable *in vivo* distribution, higher tumor accumulation, and hepatobiliary clearance.

Guided by the excellent NIR-II fluorescence/PA imaging both *in vitro* and *in vivo*, the PTT efficacy of **SYL** NPs in mice was then investigated with PBS, PBS with 808 nm NIR laser irradiation and **SYL** NPs as negative controls. PBS (100 μL) and **SYL** NPs (100 $\mu\text{g}/100 \mu\text{L}$) were injected into 4T1 tumor-bearing nude mice respectively *via* the tail vein. As the maximum accumulation of

SYL NPs was around 8 h according to the *in vivo* imaging data, PTT treatment was performed at 8 h post-injection. As seen in Fig. 5a–c, after 120 s of irradiation (808 nm, power 1 W cm^{-2}), the temperature at the tumor site treated with **SYL** NPs increased to 50.1°C , which was much higher than that of other controls, *i.e.*, the PBS with 808 nm NIR laser irradiation group (38.6°C) (Fig. 5c). Body weight and tumor growth were recorded every other day until 19 days. No significant body weight loss was observed in all the tested groups after treatment (Fig. 5d). Moreover, the result of hematoxylin and eosin (H&E) stains of major organs excised at the 20th day indicated that **SYL** NP based PTT was biocompatible without obvious adverse effects (Fig. S13†). Only the tumor growth in the mice treated with both **SYL** NPs and 808 nm laser irradiation was obviously inhibited, whereas the growth of the tumors in the other three tested groups exhibited a likewise rapid growth trend (Fig. 5e). Additionally, the survival rate of the 4T1 tumor-bearing mice treated with **SYL** NPs and laser irradiation was higher than those of the other tested groups after 40 days (Fig. 5f), indicating the effective photo-thermal properties of **SYL** NPs. Further, H&E stains of dissected tumor tissues from the tested groups indicated significant tumor cell damage with nuclear membrane fragmentation and nucleus pyknosis in the **SYL** NP treated group under laser irradiation (Fig. S14†). For the other tested groups, there was no obvious

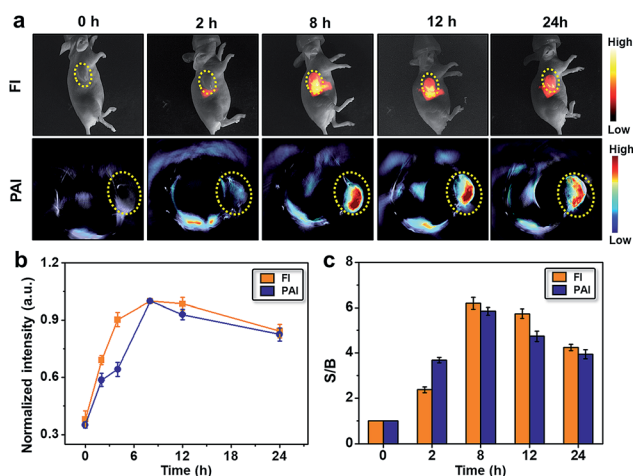


Fig. 4 *In vivo* multimodal images with **SYL** NPs. (a) *In vivo* NIR-II fluorescence (100 ms, $LP1000$, 82 mW cm^{-2}) and PA images of 4T1 tumor-bearing mice at 0, 2, 8, 12, and 24 h after tail vein injection of **SYL** NPs. Yellow circles indicate tumor sites; (b) semi-quantified fluorescence and PA intensities; (c) time-dependence of the signal-to-background ratio (FI & PA) after intravenous injection.

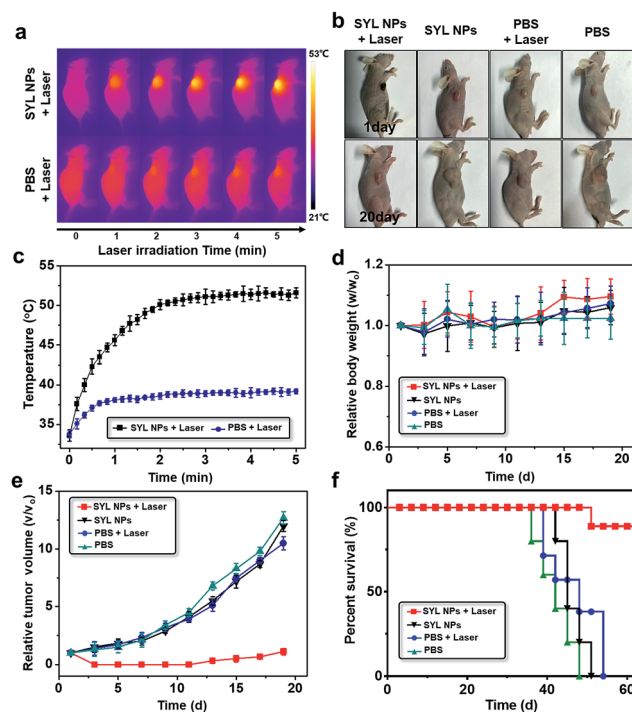


Fig. 5 Therapeutic effect of **SYL** NPs on tumor-bearing mice *via* tail vein injection. (a) IR thermal images of 4T-1 bearing mice injected with **SYL** NPs and PBS under irradiation (808 nm , 1 W cm^{-2}); (b) images of tumor-bearing mice 20 days post various treatments; (c) temperature changes in the subcutaneous tumor sites of mice treated with **SYL** NPs and PBS under continuous laser irradiation; (d) analysis of average body weights; (e) tumor growth inhibition curves for PBS alone, PBS & 808 nm laser irradiation, **SYL** NPs alone, and **SYL** NPs & 808 nm laser irradiation; (f) survival curves of the 4T1 tumor-bearing mice groups after different treatments.



tumor cell damage. The results suggested that **SYL** NPs under laser irradiation displayed high *in vivo* photothermal therapeutic efficiency in the 4T1 tumor-bearing model.

Conclusions

In summary, a novel DD–A–DD scaffold molecular dye **SYL** was designed and facilely synthesized with intrinsic multifunctional ability including dual-modal NIR-II fluorescence/PA imaging as well as photothermal therapeutic effects. The **SYL** NP based probe has the capability to passively target tumors in the 4T1 subcutaneous mouse model with relatively high fluorescence and photoacoustic signal outputs as well as an excellent S/B ratio. Additionally, the efficient tumor inhibition rate with minimal adverse side effects prolonged the life span of tumor-bearing mice in the **SYL** NP treated group under laser irradiation. These results highlighted that **SYL** NPs as a multifunctional platform could be an interesting candidate for cancer diagnosis (NIR-II) and therapy in future bioapplications. However, stimulated by hyperthermia at the tumor site, incompletely killed tumor cells might have chances of recurrence because of thermal resistance acquired by these residual tumor cells.⁶⁷ To improve PTT efficiency, some suggestions are recommended here. First, dyes with a longer emission wavelength and higher photothermal conversion efficiency should be designed, which can ensure enough penetration depth to cover the whole tumor volume and sufficient temperature upon irradiation to kill the cancer cells completely. Second, integrating PTT with other therapeutic modalities such as photodynamic therapy (PDT), chemotherapy or gene therapy could be beneficial to improve the overall treatment efficiency.

Ethical statement

The facilities used for animal studies have been accredited by the Association for Assessment and Accreditation of Laboratory Animal Care International (AAALAC International), and all subsequent procedures were approved by and performed in accordance with the Guidelines for the Care and Use of Laboratory Animals of the Chinese Animal Welfare Committee.

Conflicts of interest

There are no conflicts to declare.

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Notes and references

- 1 Y. Cai, Z. Wei, C. Song, C. Tang, W. Han and X. Dong, *Chem. Soc. Rev.*, 2019, **48**, 22–37.
- 2 J. Li and K. Pu, *Chem. Soc. Rev.*, 2019, **48**, 38–71.
- 3 M. C. Heffern, L. M. Matosziuk and T. J. Meade, *Chem. Rev.*, 2014, **114**, 4496–4539.
- 4 S. He, J. Song, J. Qu and Z. Cheng, *Chem. Soc. Rev.*, 2018, **47**, 4258–4278.
- 5 X. Huang, J. Song, B. C. Yung, X. Huang, Y. Xiong and X. Chen, *Chem. Soc. Rev.*, 2018, **47**, 2873–2920.
- 6 M. H. Lee, A. Sharma, M. J. Chang, J. Lee, S. Son, J. L. Sessler, C. Kang and J. S. Kim, *Chem. Soc. Rev.*, 2018, **47**, 28–52.
- 7 L. He, B. Dong, Y. Liu and W. Lin, *Chem. Soc. Rev.*, 2016, **45**, 6449–6461.
- 8 Y. Xing, J. Zhao, P. S. Conti and K. Chen, *Theranostics*, 2014, **4**, 290–306.
- 9 A. Louie, *Chem. Rev.*, 2010, **110**, 3146–3195.
- 10 Y. Sun, X. Ma, K. Cheng, B. Wu, J. Duan, H. Chen, L. Bu, R. Zhang, X. Hu, Z. Deng, L. Xing, X. Hong and Z. Cheng, *Angew. Chem., Int. Ed.*, 2015, **54**, 5981–5984.
- 11 H. S. Jung, P. Verwilt, A. Sharma, J. Shin, J. L. Sessler and J. S. Kim, *Chem. Soc. Rev.*, 2018, **47**, 2280–2297.
- 12 J. Ma, M. Yu, Z. Zhang, W. Cai, Z. Zhang, H. Zhu, Q. Cheng, Z. Tian and D. Pang, *Nanoscale*, 2018, **10**, 10699–10704.
- 13 R. Huang, M. Wang, Y. Zhu, P. S. Conti and K. Chen, *Curr. Top. Med. Chem.*, 2015, **15**, 795–819.
- 14 Y. Sun, X. Zeng, Y. Xiao, C. Liu, H. Zhu, H. Zhou, Z. Chen, F. Xu, J. Wang, M. Zhu, J. Wu, H. Zhang, Z. Deng, Z. Cheng and X. Hong, *Chem. Sci.*, 2018, **9**, 2092–2097.
- 15 L. Sun, J. Ding, W. Xing, Y. Gai, J. Sheng and D. Zeng, *Bioconjugate Chem.*, 2016, **27**, 1200–1204.
- 16 Y. Gai, G. Xiang, X. Ma, W. Hui, Q. Ouyang, L. Sun, J. Ding, J. Sheng and D. Zeng, *Bioconjugate Chem.*, 2016, **27**, 515–520.
- 17 H. Zhang, P. Desai, Y. Koike, J. Houghton, S. Carlin, N. Tandon, K. Touijer and W. A. Weber, *J. Nucl. Med.*, 2017, **58**, 29–35.
- 18 F. Ding, Y. Zhan, X. Lu and Y. Sun, *Chem. Sci.*, 2018, **9**, 4370–4380.
- 19 F. Ding, S. Chen, W. Zhang, Y. Tu and Y. Sun, *Bioorg. Med. Chem.*, 2017, **25**, 5179–5184.
- 20 G. Hong, A. L. Antaris and H. Dai, *Nat. Biomed. Eng.*, 2017, **1**, 0010.
- 21 Y. Fan, P. Wang, Y. Lu, R. Wang, L. Zhou, X. Zheng, X. Li, J. A. Piper and F. Zhang, *Nat. Nanotechnol.*, 2018, **13**, 941–946.
- 22 Q. Miao and K. Pu, *Adv. Mater.*, 2018, **30**, 1801778.
- 23 Kenry, Y. Duan and B. Liu, *Adv. Mater.*, 2018, **30**, 1802394.
- 24 C. Li and Q. Wang, *ACS Nano*, 2018, **12**, 9654–9659.
- 25 Y. Fan and F. Zhang, *Adv. Opt. Mater.*, 2019, **7**, 1801417.
- 26 Z. Xue, S. Zeng and J. Hao, *Biomaterials*, 2018, **171**, 153–163.



- 27 R. Wang, L. Zhou, W. Wang, X. Li and F. Zhang, *Nat. Commun.*, 2017, **8**, 14702.
- 28 S. Zhu, B. C. Yung, S. Chandra, G. Niu, A. L. Antaris and X. Chen, *Theranostics*, 2018, **8**, 4141–4151.
- 29 Y. Tang, Y. Li, X. Lu, X. Hu, H. Zhao, W. Hu, F. Lu, Q. Fan and W. Huang, *Adv. Funct. Mater.*, 2019, **29**, 1807376.
- 30 Y. Tang, Y. Li, X. Hu, H. Zhao, Y. Ji, L. Chen, W. Hu, W. Zhang, X. Li, X. Lu, W. Huang and Q. Fan, *Adv. Mater.*, 2018, **30**, 1801140.
- 31 X. Lei, R. Li, D. Tu, X. Shang, Y. Liu, W. You, C. Sun, F. Zhang and X. Chen, *Chem. Sci.*, 2018, **9**, 4682–4688.
- 32 C. Ruan, C. Liu, H. Hu, X. L. Guo, B. P. Jiang, H. Liang and X. C. Shen, *Chem. Sci.*, 2019, **10**, 4699–4706.
- 33 P. Sun, Q. Wu, X. Sun, H. Mao, W. Deng, W. Zhang, Q. Fan and W. Huang, *Chem. Commun.*, 2018, **54**, 13395–13398.
- 34 Y. Tang, Y. Li, Z. Wang, F. Pei, X. Hu, Y. Ji, X. Li, H. Zhao, W. Hu, X. Lu, Q. Fan and W. Huang, *Chem. Commun.*, 2019, **55**, 27–30.
- 35 F. Ding, Y. Fan, Y. Sun and F. Zhang, *Adv. Healthcare Mater.*, 2019, **8**, 1900260.
- 36 B. Li, L. Lu, M. Zhao, Z. Lei and F. Zhang, *Angew. Chem., Int. Ed.*, 2018, **57**, 7483–7487.
- 37 L. Liu, S. Wang, B. Zhao, P. Pei, Y. Fan, X. Li and F. Zhang, *Angew. Chem., Int. Ed.*, 2018, **57**, 7518–7522.
- 38 G. Xu, Q. Yan, X. Lv, Y. Zhu, K. Xin, B. Shi, R. Wang, J. Chen, W. Gao, P. Shi, C. Fan, C. Zhao and H. Tian, *Angew. Chem., Int. Ed.*, 2018, **57**, 3626–3630.
- 39 A. L. Antaris, H. Chen, K. Cheng, Y. Sun, G. Hong, C. Qu, S. Diao, Z. Deng, X. Hu, B. Zhang, X. Zhang, O. K. Yaghi, Z. R. Alamparambil, X. Hong, Z. Cheng and H. Dai, *Nat. Mater.*, 2016, **15**, 235–242.
- 40 A. L. Antaris, H. Chen, S. Diao, Z. Ma, Z. Zhang, S. Zhu, J. Wang, A. X. Lozano, Q. Fan, L. Chew, M. Zhu, K. Cheng, X. Hong, H. Dai and Z. Cheng, *Nat. Commun.*, 2017, **8**, 15269.
- 41 Y. Sun, C. Qu, H. Chen, M. He, C. Tang, K. Shou, S. Hong, M. Yang, Y. Jiang, B. Ding, Y. Xiao, L. Xing, X. Hong and Z. Cheng, *Chem. Sci.*, 2016, **7**, 6203–6207.
- 42 F. Ding, C. Li, Y. Xu, J. Li, H. Li, G. Yang and Y. Sun, *Adv. Healthcare Mater.*, 2018, **7**, 1800973.
- 43 Q. Yang, Z. Hu, S. Zhu, R. Ma, H. Ma, R. Ma, H. Wan, T. Zhu, Z. Jiang, W. Liu, L. Jiao, H. Sun, Y. Liang and H. Dai, *J. Am. Chem. Soc.*, 2018, **140**, 1715–1724.
- 44 S. Zhu, Q. Yang, A. L. Antaris, J. Yue, Z. Ma, H. Wang, W. Huang, H. Wan, J. Wang, S. Diao, B. Zhang, X. Li, Y. Zhong, K. Yu, G. Hong, J. Luo, Y. Liang and H. Dai, *Proc. Natl. Acad. Sci. U. S. A.*, 2017, **114**, 962–967.
- 45 K. Shou, C. Qu, Y. Sun, H. Chen, S. Chen, L. Zhang, H. Xu, X. Hong, A. Yu and Z. Cheng, *Adv. Funct. Mater.*, 2017, **27**, 1700995.
- 46 Y. Sun, M. Ding, X. Zeng, Y. Xiao, H. Wu, H. Zhou, B. Ding, C. Qu, W. Hou, A. Er-bu, Y. Zhang, Z. Cheng and X. Hong, *Chem. Sci.*, 2017, **8**, 3489–3493.
- 47 L. Tu, Y. Xu, Q. Ouyang, X. Li and Y. Sun, *Chin. Chem. Lett.*, 2019, DOI: 10.1016/j.ccl.2019.05.022.
- 48 Y. Sun, F. Ding, Z. Zhou, C. Li, M. Pu, Y. Xu, Y. Zhan, X. Lu, H. Li, G. Yang, Y. Sun and P. J. Stang, *Proc. Natl. Acad. Sci. U. S. A.*, 2019, **116**, 1968–1973.
- 49 X. Zhang, H. Wang, A. L. Antaris, L. Li, S. Diao, R. Ma, A. Nguyen, G. Hong, Z. Ma, J. Wang, S. Zhu, J. M. Castellano, T. Wyss-Coray, Y. Liang, J. Luo and H. Dai, *Adv. Mater.*, 2016, **28**, 6872–6879.
- 50 Q. Yang, Z. Ma, H. Wang, B. Zhou, S. Zhu, Y. Zhong, J. Wang, H. Wan, A. L. Antaris, R. Ma, X. Zhang, J. Yang, X. Zhang, H. Sun, W. Liu, Y. Liang and H. Dai, *Adv. Mater.*, 2017, **29**, 1605497.
- 51 H. Wan, J. Yue, S. Zhu, T. Uno, X. Zhang, Q. Yang, K. Yu, G. Hong, J. Wang, L. Li, Z. Ma, H. Gao, Y. Zhong, J. Su, A. L. Antaris, Y. Xia, J. Luo, Y. Liang and H. Dai, *Nat. Commun.*, 2018, **9**, 1171–1179.
- 52 S. Zhu, S. Herraiz, J. Yue, M. Zhang, H. Wan, Q. Yang, Z. Ma, Y. Wang, J. He, A. L. Antaris, Y. Zhong, S. Diao, Y. Feng, Y. Zhou, K. Yu, G. Hong, Y. Liang, A. J. Hsueh and H. Dai, *Adv. Mater.*, 2018, **30**, 1705799.
- 53 X. Zeng, Y. Xiao, J. Lin, S. Li, J. Nong, G. Xu, H. Wang, F. Xu, J. Wu, Z. Deng and X. Hong, *Adv. Healthcare Mater.*, 2018, **7**, 1800589.
- 54 J. Lin, X. Zeng, Y. Xiao, L. Tang, J. Nong, Y. Liu, H. Zhou, B. Ding, F. Xu, H. Tong, Z. Deng and X. Hong, *Chem. Sci.*, 2019, **10**, 1219–1226.
- 55 R. Tian, H. Ma, Q. Yang, H. Wan, S. Zhu, S. Chandra, H. Sun, D. O. Kiesewetter, G. Niu, Y. Liang and X. Chen, *Chem. Sci.*, 2019, **10**, 326–332.
- 56 K. Cheng, H. Chen, C. H. Jenkins, G. L. Zhang, W. Zhao, Z. Zhang, F. Han, J. Fung, M. Yang, Y. Jiang, L. Xing and Z. Cheng, *ACS Nano*, 2017, **11**, 12276–12291.
- 57 Y. Yuan, C. Zhang, M. Gao, R. Zhang, B. Tang and B. Liu, *Angew. Chem., Int. Ed.*, 2015, **54**, 1780–1786.
- 58 J. Qi, C. Sun, A. Zebibula, H. Zhang, R. K. Kwok, X. Zhao, W. Xi, J. W. Y. Lam, J. Qian and B. Tang, *Adv. Mater.*, 2018, **30**, 1706856.
- 59 Z. Sheng, B. Guo, D. Hu, S. Xu, W. Wu, W. H. Liew, K. Yao, J. Jiang, C. Liu, H. Zheng and B. Liu, *Adv. Mater.*, 2018, **30**, 1800766.
- 60 J. Qi, C. Sun, D. Li, H. Zhang, W. Yu, A. Zebibula, J. W. Y. Lam, W. Xi, L. Zhu, F. Cai, P. Wei, C. Zhu, R. T. K. Kwok, L. L. Streich, R. Prevedel, J. Qian and B. Z. Tang, *ACS Nano*, 2018, **12**, 7936–7945.
- 61 K. R. J. Thomas, Y. C. Hsu, J. T. Lin, K. M. Lee, K. C. Ho, C. H. Lai, Y. M. Cheng and P. T. Chou, *Chem. Mater.*, 2008, **20**, 1830–1840.
- 62 P. Shen, Y. Tang, S. Jiang, H. Chen, X. Zheng, X. Wang, B. Zhao and S. Tan, *Org. Electron.*, 2011, **12**, 125–135.
- 63 G. Qian and Z. Wang, *Chem.-Asian J.*, 2010, **5**, 1006–1029.
- 64 Q. Chen, J. Wen, H. Li, Y. Xu, F. Liu and S. Sun, *Biomaterials*, 2016, **106**, 144–166.
- 65 Y. Liu, P. Bhattarai, Z. Dai and X. Chen, *Chem. Soc. Rev.*, 2019, **48**, 2053–2108.
- 66 M. Zheng, P. Zhao, Z. Luo, P. Gong, C. Zheng, P. Zhang, C. Yue, D. Gao, Y. Ma and L. Cai, *ACS Appl. Mater. Interfaces*, 2014, **6**, 6709–6716.
- 67 Q. Wang, Y. Dai, J. Xu, J. Cai, X. Niu, L. Zhang, R. Chen, Q. Shen, W. Huang and Q. Fan, *Adv. Funct. Mater.*, 2019, DOI: 10.1002/adfm.201901480.

