



**Showcasing research from Associate Professor Takao Yasui and Professor Yoshinobu Baba's laboratory, Graduate School of Engineering, Nagoya University, Nagoya, Japan.**

**Tailoring ZnO nanowire crystallinity and morphology for label-free capturing of extracellular vesicles**

An effective strategy to fabricate various morphologies and well-defined crystal structures composed of mixed phases of ZnO nanowire was accomplished *via* an ammonia-assisted hydrothermal method. The label-free capturing of cancer cell-derived extracellular vesicles (EVs) obtained at high surface-to-volume ratio and phase mixture takes advantage of the tuneable electrostatic properties of the nanowire. Analysis with nanoscale spatial resolution reveals a zinc blend phase related to more EV adsorption localized at the upper and lower parts of the nanowires. This research provides new insights into the functionalities of ZnO nanowire and can be applied to high performance devices for EV isolation and cancer detection technology.

**As featured in:**



See Takao Yasui *et al.*, *Nanoscale*, 2022, **14**, 4484.



Cite this: *Nanoscale*, 2022, **14**, 4484

# Tailoring ZnO nanowire crystallinity and morphology for label-free capturing of extracellular vesicles†

Piyawan Paisrisarn,<sup>a</sup> Takao Yasui,<sup>a,b,c</sup> Zetao Zhu,<sup>a,c</sup> Annop Klamchuen,<sup>d</sup> Panita Kasamechongchun,<sup>d</sup> Tuksadon Wutikhun,<sup>d</sup> Visittapong Yordsri<sup>e</sup> and Yoshinobu Baba<sup>a,c,f</sup>

Zinc oxide (ZnO) nanowires have shown their potential in isolation of cancer-related biomolecules such as extracellular vesicles (EVs), RNAs, and DNAs for early diagnosis and therapeutic development of diseases. Since the function of inorganic nanowires changes depending on their morphology, previous studies have established strategies to control the morphology and have demonstrated attainment of improved properties for gas and organic compound detection, and for dye-sensitized solar cells and photoelectric conversion performance. Nevertheless, crystallinity and morphology of ZnO nanowires for capturing EVs, an important biomarker of cancer, have not yet been discussed. Here, we fabricated ZnO nanowires with different crystallinities and morphologies using an ammonia-assisted hydrothermal method, and we comprehensively analyzed the crystalline nature and oriented growth of the synthesized nanowires by X-ray diffraction and selected area electron diffraction using high resolution transmission electron microscopy. In evaluating the performance of label-free EV capture in a microfluidic device platform, we found both the crystallinity and morphology of ZnO nanowires affected EV capture efficiency. In particular, the zinc blende phase was identified as important for crystallinity, while increasing the nanowire density in the array was important for morphology to improve EV capture performance. These results highlighted that the key physicochemical properties of the ZnO nanowires were related to the EV capture performance.

Received 2nd November 2021

Accepted 24th January 2022

DOI: 10.1039/d1nr07237d

[rsc.li/nanoscale](http://rsc.li/nanoscale)

## Introduction

At the nanoscale, device properties and performance are no longer dependent solely on the material type, but also on material geometry. Nanowires, anisotropic structures with diameters from one to hundreds of nanometers and lengths of tens to hundreds of microns, provide a unique platform for

harnessing nanoscale phenomena.<sup>1</sup> Semiconductor nanowires are a broad class of materials which, through controlled growth and organization, have led to a number of novel nanoscale mechanical, electrochemical, thermoelectric, photonic and electronic devices.<sup>2–4</sup> In order to produce and utilize these properties for nanowires, reliable and controllable nanowire syntheses are required. A hydrothermal method offers flexible control over nanowire crystal structure and growth orientation, enabling fabrication of high-quality single-crystalline materials with control of diameter, length, composition, and phase.<sup>5</sup> When ZnO nanowires are hydrothermally synthesized, they are single crystals, and as a result of their narrow diameter, the nanowires have an extremely high surface area allowing highly sensitive interaction with various substances.<sup>6–9</sup> This high surface-to-volume ratio makes the nanowires very sensitive to changes in surface chemistry and permits rapid interaction with comparatively small sizes of biomolecules including extracellular vesicles (EVs) and nucleic acids giving rise to their prevalence in isolation and detection technology development.<sup>10–15</sup>

The physical phenomena in the radial and axial directions and the high surface-to-volume ratios of the nanowires have

<sup>a</sup>Department of Biomolecular Engineering, Graduate School of Engineering, Nagoya University, Furo-cho, Chikusa-ku, Nagoya 464-8603, Japan.

E-mail: [yasui@chembio.nagoya-u.ac.jp](mailto:yasui@chembio.nagoya-u.ac.jp)

<sup>b</sup>Japan Science and Technology Agency (JST), PRESTO, 4-1-8 Honcho, Kawaguchi, Saitama 332-0012, Japan

<sup>c</sup>Institute of Nano-Life-Systems, Institutes of Innovation for Future Society, Nagoya University, Furo-cho, Chikusa-ku Nagoya 464-8603, Japan

<sup>d</sup>National Nanotechnology Center (NANOTEC), National Science and Technology Development Agency (NSTDA), Pathum Thani 12120, Thailand

<sup>e</sup>National Metal and Materials Technology Center (MTEC), National Science and Technology Development Agency (NSTDA), Pathum Thani 12120, Thailand

<sup>f</sup>Institute of Quantum Life Science, National Institutes for Quantum and Radiological Science and Technology, Anagawa 4-9-1, Inage-ku, Chiba 263-8555, Japan

†Electronic supplementary information (ESI) available. See DOI: 10.1039/d1nr07237d



been controlled and that has led to an explosion of applications utilizing these structures. Getting preferential nucleation at the ZnO (0001) polar plane and suppressing the lateral growth, such as of the (1010) plane, are key essentials to produce a well-defined nanowire structure.<sup>16–18</sup> Many growth parameters have been explored to manipulate the morphology of hydrothermally synthesized ZnO nanowires including the addition of auxiliary agents such as organic ligands, metal ion impurities and ammonia ions.<sup>19–24</sup> The solution-grown ZnO nanowires generally adopt the wurtzite crystal structure and grow along the *c* axis with a (0002) top facet and (1010) side facets. At pH = 11 basic conditions, it was found that the top facet is negatively charged and the side facets are positively charged, enabling control of the nanowire dimensions based on the facet-selective electrostatic interaction;<sup>25</sup> and therefore, the radial growth of colloidal nanowires can be effectively suppressed. In other words, control of the nanowire mean length and diameter can be achieved by optimizing the concentration of ammonia ions, for example. Several studies have investigated physicochemical properties of the developed high aspect ratio ZnO nanowires in various applications including photocatalysis, a piezoelectric transducer, a UV detector, photovoltaic devices, and gas and chemical sensors.<sup>26–31</sup> To the best of our knowledge, the correlation of ZnO nanowires in terms of crystal structure and morphology with biomolecule capture has not been discussed until now.

Here, we propose a methodology using oxide nanowires with different crystallinities and morphologies synthesized by ammonia-assisted hydrothermal growth to evaluate capture performance of extracellular vesicles (EVs) and we investigate a correlation between surface crystallinity and numbers of the captured EVs. The EVs are nano- to micro-sized vesicles secreted from all cells, including cancer cells, and they are released into their environment to act as intercellular

messengers.<sup>32–34</sup> Since they enclose components from the parent cells, including nucleic acids and proteins, EVs are being examined as sources of biomarkers in various diseases, including neurodegenerative diseases, cardiovascular diseases and cancers.<sup>35–38</sup> EVs are also being explored for the delivery of therapeutic payloads to specific cells or tissues in cancer therapeutics,<sup>39,40</sup> owing to their distinct biological features. Cancer cell-derived EVs have the same membrane orientation as the cells: they are exposed at their surfaces to lipids and organotropic metastasis-induced transmembrane proteins.<sup>41</sup> The enrichment of these proteoglycans has led to the idea that EVs are charged molecules.<sup>42,43</sup> We have initiated studies exploiting ZnO nanowire integrated microfluidic devices for label-free capture of EVs through surface charge-based capture which can collect massive numbers of EVs and their molecular cargos.<sup>10,14</sup> However, comprehensive studies of nanowires with a diverse crystal structure and morphology system have not been made yet.

## Experimental

### Synthesis of ZnO nanowires

We fabricated ZnO nanowires on Si substrates *via* a seed-assisted hydrothermal process. First, double-microchannel patterns ( $20 \times 2 \text{ mm}^2$ ) were formed by photolithography onto each silicon (100) substrate and a 20 nm thick seed layer was deposited with an electron cyclotron resonance (ECR) sputtering system (Elionix, EIS-200ERT-YN). High melting-point Cr-based alloy with purity of 99.999% was used as a sputtering target. The substrates with seed layer were oxidized in an oven at 400 °C for 2 h to prepare them as a scaffold for the ZnO nanowires. After seed layer preparation, the growth solutions were prepared using 15 mM hexamethylenetetramine (HMTA) and 15 mM zinc nitrate hexahydrate ( $\text{Zn}(\text{NO}_3)_2 \cdot 6\text{H}_2\text{O}$ ) as the precursors, and 0.8 M ammonium solution was added for ultra-lengthening of ZnO nanowires. The nanowires were grown at 95 °C with varying duration times (3 h, 6 h and 9 h). We renewed the growth solutions every 3 h to maintain the concentrations.

### Fabrication of the microfluidic device

The dried nanowires grown on the substrate were treated with oxygen plasma. A sheet of 30  $\mu\text{m}$ -depth polydimethylsiloxane (PDMS) was attached to the substrate. The sheet was patterned with two microchannels ( $20 \times 2 \text{ mm}^2$ ) and 0.05 mm inlet and outlet holes at the end of channels for sample injection.

### Characterization of ZnO nanowires

The morphology and the composition of ZnO nanowires were characterized with a field emission scanning electron microscope (FESEM; Carl Zeiss, Supra 40VP), a high-resolution transmission electron microscope (HRTEM; JEOL, JEM-2100) and an energy dispersive X-ray spectroscopy (EDS) system (Oxford Instruments, Ultim Max 100). For the cross-sectional SEM-EDS analysis and the single-nanowire HRTEM analysis,



**Takao Yasui**

*Takao Yasui is an Associate Professor of Department of Biomolecular Engineering, Graduate School of Engineering at Nagoya University, a PRESTO Researcher at the Japan Science and Technology Agency (JST), and a co-founder and a Nanodevice Director at Craif Inc. In 2011, he received his PhD in the Department of Applied Chemistry, Graduate School of Engineering from Nagoya University. He has received over*

*35 awards for his contributions in the field of oxide nanowire microfluidics. His research interests are focused on designing, fabricating, and characterizing oxide nanowire microfluidics for applications in healthcare, diagnosis, sensing biomolecules and engineering biology.*





we utilized an accelerating voltage of 15 kV and 200 kV, respectively. The EDS mapping images were integrated for 100 cycles. The peaks of Zn K $\alpha$  (8.6313 keV), O K $\alpha$  (0.5249 keV), Si K $\alpha$  (1.7398 keV), and Cr K $\alpha$  (5.4117 keV) and were chosen to construct the elemental mapping images. An X-ray diffraction (XRD) analyzer (Bruker, D8 Advance) was used to examine the crystalline nature and oriented growth of synthesized ZnO nanowires. The conditions were set to be Cu K $\alpha$  XRD source,  $\lambda = 0.1542$  nm, accelerating voltage of 40 kV, acceleration current 40 mA, scan step  $0.02^\circ$ , scan rate 0.5 s per step, and scan range 20 to  $80^\circ$ . The XRD patterns were analyzed using Diffrac.EVA 5.0.

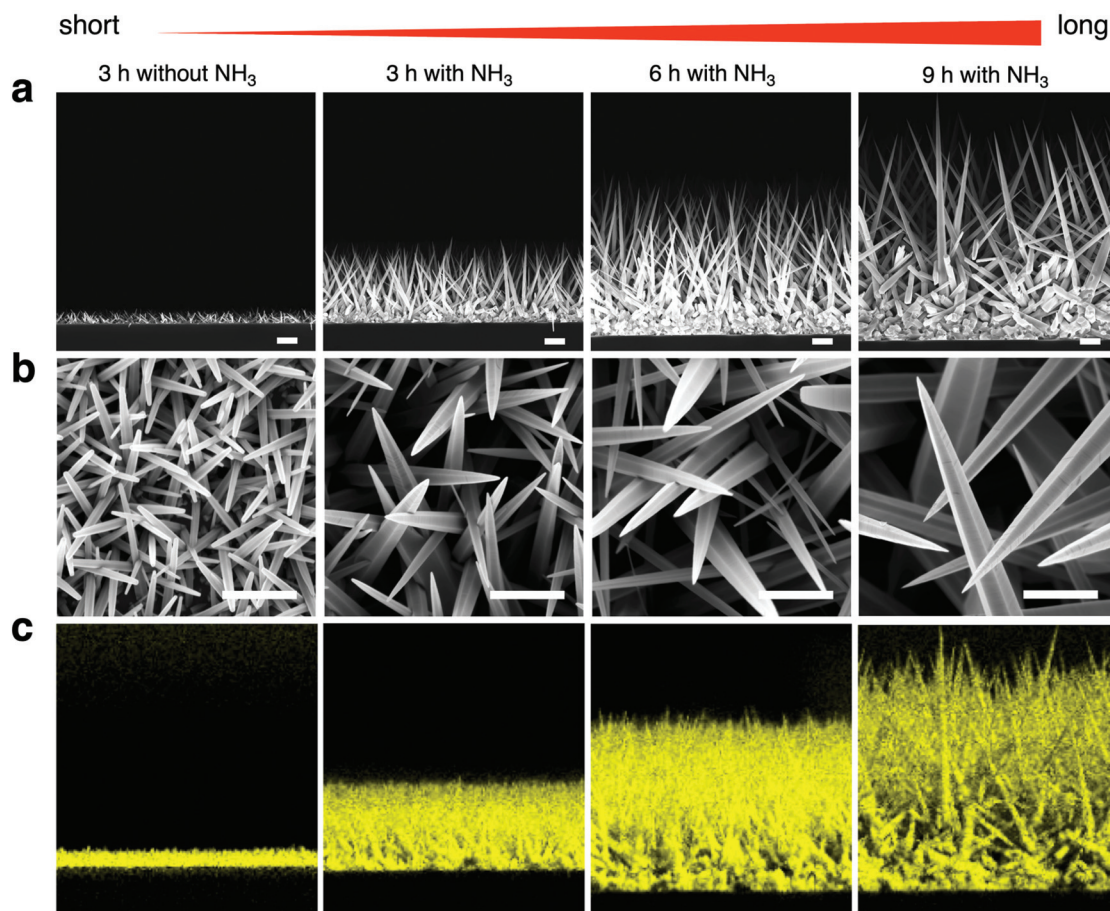
### Cell culture and extraction of EV by ultracentrifugation

We cultured human breast adenocarcinoma cells (MDA-MB-231) in DMEM supplemented with 10% exosome-depleted FBS and 1% penicillin streptomycin.  $2 \times 10^6$  cells were seeded into 15 mL of complete medium in a 75 cm<sup>2</sup> culturing flask and incubated at 37 °C and with 5% CO<sub>2</sub>. After 48 h, the cell medium was taken from the culturing flask and filtered through a 0.22  $\mu$ m filter to remove cellular debris. Next, the filtered medium was ultracentrifuged at 110 000g, 4 °C, 80 min. After discarding the supernatant, we added 5 mL

of 0.22  $\mu$ m filtered PBS to wash the EV pellets, and this solution was ultracentrifuged again with the same conditions. After discarding the supernatant, 1 mL of 0.22  $\mu$ m filtered PBS was added to collect EVs. The EV suspension was stored at 4 °C until use.

### Evaluation of EV capture efficiency

We integrated a nanowire device with a dual-channel syringe pump to continuously infuse EV sample with the flow rate of 10  $\mu$ L min<sup>-1</sup>. 250  $\mu$ L of EV-suspended PBS was supplied to the nanowire device to isolate EVs onto each nanowire. EV concentration and size were analysed using a nanoparticle tracking analysis system (Malvern Panalytical, NanoSight LM10). After appropriate dilutions, video data were collected 5 times for a 60 s time period for each video. Camera level and detection threshold were set to 13 and 5, respectively. Data analysis was performed automatically by NanoSight NTA 3.2 software. To observe EVs on nanowire surfaces, the device was left to dry naturally at room temperature. The PDMS sheet was removed from the substrate, the captured EVs on the nanowire surfaces were directly observed using FESEM at 5 kV of accelerating voltage without any fixing or coating processes.



**Fig. 1** (a) FESEM images of cross-sectional nanowires for different growth conditions. The scale bars are 2  $\mu$ m. (b) FESEM images of top-view nanowires. The scale bars are 1  $\mu$ m. (c) Elemental mapping of Zn K $\alpha$  (8.6313 keV) highlighted in yellow, as a main constituent of the nanowires.

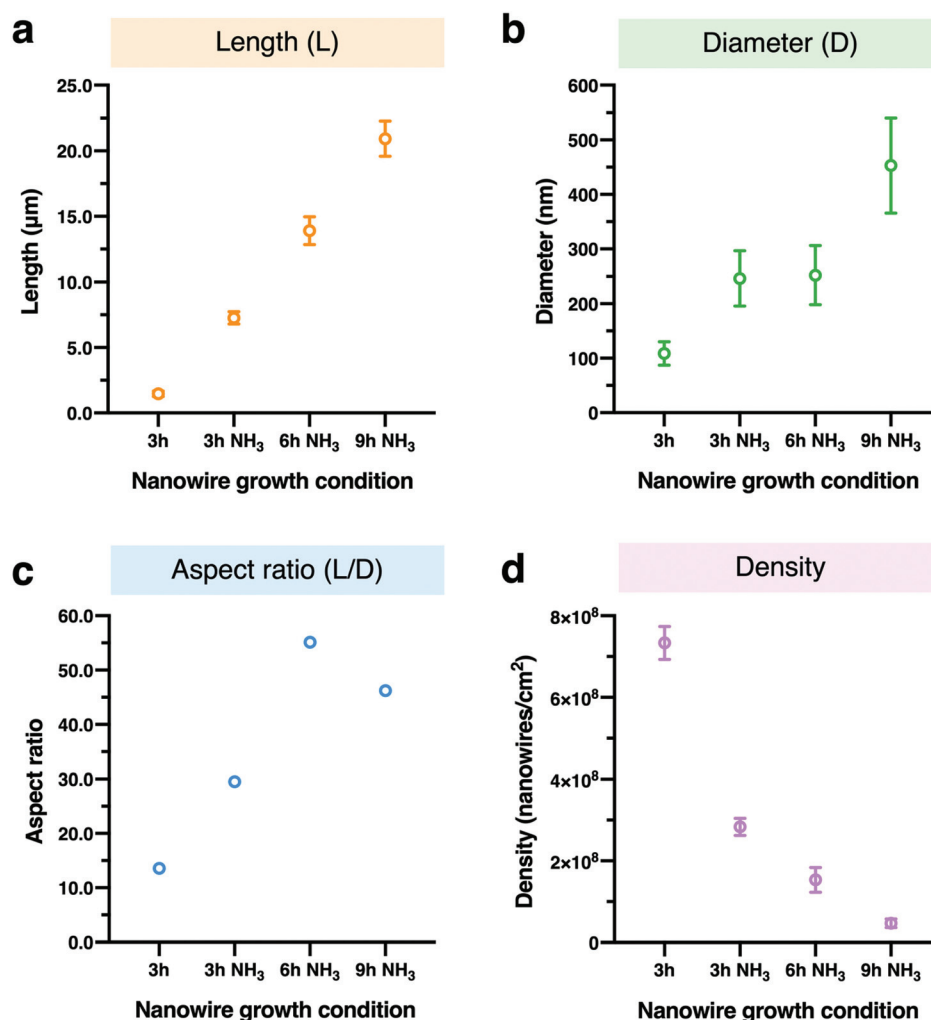


## Results and discussion

For controlling the morphology of ZnO nanowires, we introduced ammonia solution into the ZnO precursor growth solution during the growth step and hydrothermally fabricated nanowires in varying growth duration times for 3, 6, and 9 h. Ammonium hydroxide is an additive that has been widely used to enhance the nanowire growth rate along the *c*-axial ZnO plane *via* face-selective electrostatic crystal growth inhibition.<sup>25</sup> As ammonia concentration was reported to critically affect the morphology of nanowires,<sup>26,44,45</sup> we evaluated the optimum concentration of ammonia for our experimental conditions and found the concentration of ammonia at 0.8 M provided the longest nanowires (Fig. S1†). The pH values in growth solutions with ammonia addition (Fig. S2†) indicated the presence of OH<sup>−</sup> ions during the time before nanowire growth (initial) to after it (final). Morphology observations showed that the nanowires obtained were larger in length and

diameter in comparison to nanowires grown without ammonia addition, and nanowire length could be enlarged by increasing the growth duration time (Fig. 1a and b). The EDS mapping images of Zn K $\alpha$  as a main constituent of the synthesized nanowires confirmed that various lengths of ZnO nanowires could be obtained by using the ammonia-assisted hydrothermal fabrication method (Fig. 1c and Fig. S3†).

To clearly reflect the effect of ammonia in controlling the morphology of nanowires, we examined the length, diameter, aspect ratio and density of the synthesized nanowires as a function of growth duration times (Fig. 2). Both length and diameter increased with increasing growth time (Fig. 2a and b), especially, for the 3 h growth time, the addition of NH<sub>3</sub> lead to nanowires about 5-fold longer than those grown without NH<sub>3</sub> in the reaction mixture. Also, the nanowire lengths with ammonia addition increased with time in a linear manner; however, the nanowires grown for 9 h clearly tended to have a large diameter, indicating the occurrence of



**Fig. 2** The dependence of the morphology of ZnO nanowires on four nanowire characteristics. (a) Statistic length and (b) diameter of nanowires. The error bars are defined by standard deviation values ( $n = 100$ ). (c) Aspect ratio (length/diameter) of nanowires. (d) Density of nanowires calculated by counting the tips of nanowires in a known area. The error bars are defined by standard deviation values ( $n = 490, 190, 100, 30$  counted nanowires at each condition, respectively).



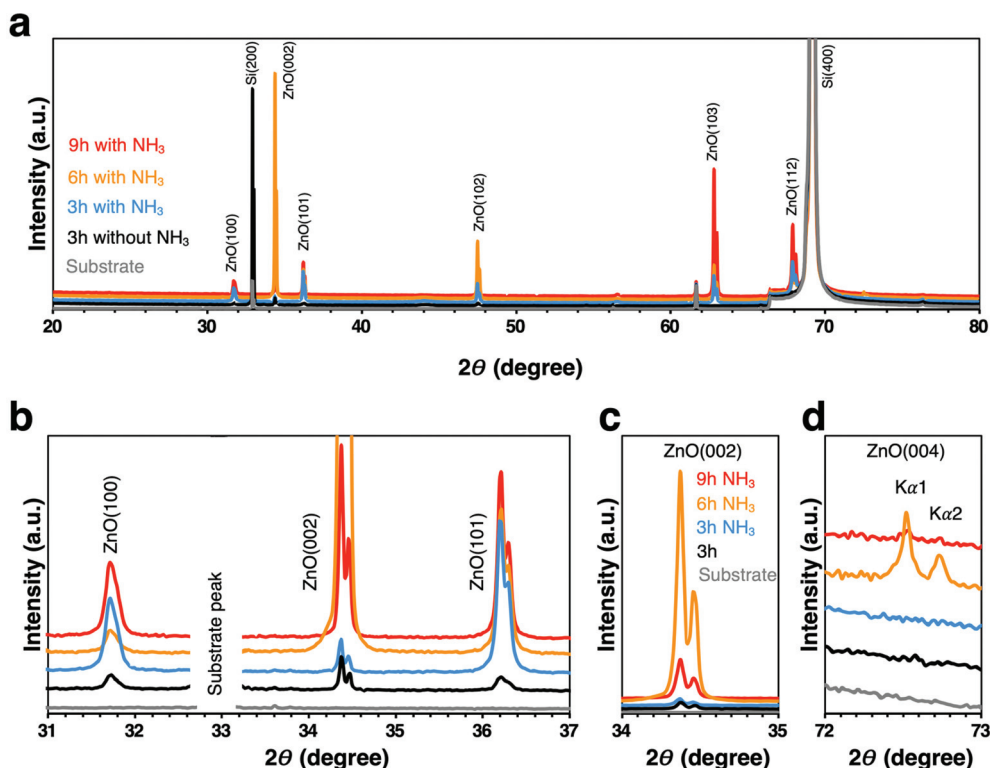
the (1010) radial plane growth. In addition, we noticed that the nanowires grown for 6 h with  $\text{NH}_3$  addition had the highest aspect ratio of over 55, calculated from the ratio of length to diameter, after which the ratio decreased (Fig. 2c). Furthermore, the density of ZnO nanowires decreased with increasing the growth time due to the lateral growth effect and the competition among nanowires (Fig. 4d and Fig. S4†). Although the precursor supply for the ZnO nanowire growth was replenished with fresh solution to maintain the concentration and growth rate, the diameter of the nanowires still continued to increase and they were expected to eventually attach together to form a ZnO film.<sup>46</sup> When the nanowires grow longer in random directions, only a few wires with a high growth rate will sustainably grow because nanowires with their orientation close to the substrate will be restricted by neighboring nanowires.<sup>18,47</sup> These results indicated the significant role of ammonia and the effect of growth time on the nanowire morphology: the length and diameter of the nanowires increased with increasing growth time but the aspect ratio and density decreased.

Since the morphological changes to the nanowires are expected to have a significant effect on the crystal structure of the nanowires, we next examined the crystallinity of the nanowires with different morphologies synthesized under each growth condition. Normalized XRD spectra showed the single crystallinity of these nanowires; however different preferential orientations were observed (Fig. 3a). The three main character-

istic peaks (100), (002), (101) of wurtzite ZnO phase in the  $2\theta = 31^\circ$  to  $37^\circ$  range demonstrated alterations of the dominant ZnO-related peak for each condition (Fig. 3b and Table 1). When ammonia was added for the 3 h growth time, the ZnO nanowires started to grow preferentially in the orientation (101) higher than (002), in contrast, the dominant peak shifted back to (002) direction at 6 h and 9 h with  $\text{NH}_3$  addition, and the highest intensity (002) peak was observed at 6 h with  $\text{NH}_3$  addition (Fig. 3c). Besides, the ZnO (004) peak which is the characteristic peak of the cubic zinc blende phase<sup>48,49</sup> was observed at 6 h and 9 h with  $\text{NH}_3$  addition (Fig. 3d). This was attributed to the fact that the synthesis of ZnO nanostructures under a thermodynamic condition generally results in the wurtzite structure since the wurtzite phase is thermodynamically more stable than the zinc blende phase.<sup>50</sup> However, compared with the wurtzite ZnO phase, zinc blende

**Table 1** Comparative intensity ratio of the characteristic XRD peaks

Sample	Peaks ratio		
	(002)/(100)	(002)/(101)	(100)/(101)
3 h without $\text{NH}_3$	1.7	1.8	1.1
3 h with $\text{NH}_3$	0.6	0.3	0.5
6 h with $\text{NH}_3$	23.6	6.9	0.3
9 h with $\text{NH}_3$	2.1	1.1	0.5
6 h with $\text{NH}_3$ 500 °C	2.1	0.7	0.3



**Fig. 3** (a) Normalized full spectra XRD patterns. (b) Main characteristic peaks of ZnO wurtzite phase. (c) Enlarged ZnO (002) plane. (d) Enlarged ZnO (004) plane.





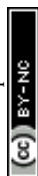
ZnO has lower ionicity and higher crystallographic symmetry, and zinc blende ZnO is expected to have higher carrier mobility and electrical gain.<sup>48</sup> The presence of different phases in the nanowires may play a role in the capture of EVs by these nanowires, because the chemical and physical properties of nanowires are expected to be significantly influenced by their crystal structure. The above results clearly showed that the effect of ammonia occurred not only on the morphology of the ZnO nanowires but also on their crystallinity, especially in the growth rate of ZnO nanowires in the vertical *c*-axis direction, where the NH<sub>3</sub> addition for 6 h had the highest growth rate.

The differences in local crystallinity caused by the nanowire morphological change in the upper, middle, and lower regions were investigated (Fig. 4), because the nanowires grown with NH<sub>3</sub> additions were found to have a needle-like shape, with the upper part of the nanowires being sharper and smaller in

diameter than the lower part. Electron diffraction patterns indicated that the structures of ZnO nanowires with different morphologies synthesized under each growth condition were a single crystal. In case of 3 h growth without NH<sub>3</sub> addition, the nanowires grow mainly only along the (0002) direction without any significant lateral sidewall growth for all positions. For the 3 h growth with NH<sub>3</sub> addition, the nanowires tended to have varying diameters from upper to lower parts but the selective area electron diffraction (SAED) measurements yielded consistent diffraction patterns. Distinct patterns among the nanowire regions were identified for the cases of 6 and 9 h growth with NH<sub>3</sub> addition. At the short 3 h growth time, the wurtzite growth mode was dominant; whereas, the larger diameter nanowires grown with longer reaction times (6 h and 9 h) had a zinc blende crystal structure. In other words, a transition to the zinc blende growth mode occurred at longer times. We



**Fig. 4** (a–d) Low-resolution TEM images, and HRTEM images in three regions of i. upper, ii. middle and iii. lower of a single ZnO nanowire, where (a) is a nanowire grown for 3 h without NH<sub>3</sub> addition; (b) is a nanowire grown for 3 h with NH<sub>3</sub> addition; (c) is a nanowire grown for 6 h with NH<sub>3</sub> addition; and (d) is a nanowire grown for 9 h with NH<sub>3</sub> addition. The insets are SAED patterns of each HRTEM micrograph. The scale bars are 50 nm and 2 nm in low-resolution TEM and HRTEM images, respectively.



investigated the inter planar spacing and diffraction planes from XRD and HRTEM for further understanding of the crystal structure of the nanowires (Table 2).

The above results highlighted that the growth time with  $\text{NH}_3$  addition not only affected the lateral diameter but also the growth direction and crystal plane. The anisotropic growth structure of the nanowires is caused by the highest growth rate occurring along the  $c$ -axis, which has a high surface energy but a small area leading to the wire-like structures.<sup>51</sup> A possible explanation for the wurtzite phase in the nanowires is that side walls of wurtzite have a lower surface energy than the zinc blende nanowires,<sup>52–54</sup> and the successful phase transition we achieved was attributed to alterations in nanowire surface energy at different growth conditions with  $\text{NH}_3$  addition (Fig. S6b†). Although the difference in atomic arrangement on a nanoscale is subtle, it is expected to cause a large difference

in the physical and chemical properties of the nanomaterials, such as their EV capture efficiency.<sup>55,56</sup> We anticipated that the preferential capture of the negatively charged EVs<sup>14,43,57</sup> onto the positively charged surfaces with different crystal planes of ZnO nanowires would lead to the higher EV capture efficiency.

Finally, we demonstrated the effect on EV capture capability for ZnO nanowires fabricated by the ammonia-assisted hydrothermal growth. To do this, we integrated nanowires with PDMS microchannels to fabricate nanowire microfluidic devices (Fig. 5a). The experimental setup of a double-channel nanowire microfluidic device connected with an automatically continuous flow syringe pump supplied EVs into the device (Fig. S5†). The nanowires grown with  $\text{NH}_3$  additions resulted in the efficacy of EV capture being increased. Further, the nanowires grown for 6 h, with the highest aspect ratio, achieved the highest performance of 68% after which the

**Table 2** The inter planar spacing and diffraction planes. ZB represents zinc blende

Sample		Peak $2\theta$	$d$ -Spacing calculated from XRD (nm)	$d$ -Spacing calculated from HRTEM (nm)	$Hkl$	Structure
3 h without $\text{NH}_3$	Upper	34.37	0.2609	0.2598	002	
		56.31	0.1634	0.1602	110	
		67.91	0.1380	0.1359	112	
	Middle	34.37	0.2609	0.2570	002	
		56.31	0.1634	0.1591	110	
		67.91	0.1380	0.1352	112	
	Lower	34.37	0.2609	0.2570	002	
		56.31	0.1634	0.1580	110	
		67.91	0.1380	0.1352	112	
3 h with $\text{NH}_3$	Upper	34.37	0.2609	0.2598	002	
		56.31	0.1634	0.1602	110	
		67.91	0.1380	0.1359	112	
	Middle	34.37	0.2609	0.2598	002	
		56.31	0.1634	0.1591	110	
		67.91	0.1380	0.1359	112	
	Lower	34.37	0.2609	0.2598	002	
		56.31	0.1634	0.1602	110	
		67.91	0.1380	0.1352	112	
6 h with $\text{NH}_3$	Upper	34.37	0.2609	0.2584	002	
		47.49	0.1914	0.1897	102	
		56.31	0.1634	0.1596	110	
		67.91	0.1380	0.1371	112	
	Middle	34.37	0.2609	0.2635	002	
		56.31	0.1634	0.1577	110	
		67.91	0.1380	0.1363	112	
	Lower	31.28	0.2859	0.2887	100	
		34.37	0.2609	0.2488	002	
		56.31	0.1634	0.1585	110	
		67.91	0.1380	0.1363	112	
9 h with $\text{NH}_3$	Upper	34.37	0.2609	0.2562	002	
		56.31	0.1634	0.1602	110	
		67.91	0.1380	0.1352	112	
		72.52	0.1303	0.1294	004	
	Middle	34.37	0.2609	0.2555	002	
		56.31	0.1634	0.1577	110	
		67.91	0.1380	0.1365	112	
	Lower	36.21	0.2480	0.2436	101	
		47.49	0.1914	0.1886	102	







**Fig. 5** (a) Schematic images of the nanowire microfluidic device and EVs captured by nanowires. (b) The comparative EV capture efficiency and (c) NTA-based numbers of captured vesicles obtained using ZnO nanowire devices. Initial concentration of EVs was  $1.0 \times 10^9$  (particles per mL). Error bars indicate the standard deviation ( $n = 6$ ). (d) The comparative capture capacity per unit area of a nanowire for each nanowire condition. (e) Colored FESEM images visualize the adsorption of EVs (pink) on a ZnO nanowire surface (blue) after the EV capture experiment from 6 h with NH<sub>3</sub> and 3 h without NH<sub>3</sub> conditions, respectively. Scale bars are 100 nm.



efficacy dropped at 9 h (Fig. 5b and c). The FESEM images visualizing EVs on the ZnO nanowire surface after the EV capture experiment showed that EVs were directly captured along the nanowire surface (Fig. 5e). We attributed the efficacy enhancement to the fact that nanowires which were long in the vertical could interact with each EV species; however, the capture rate did not increase monotonically within this regime. This was due to the decreased density of nanowires (Fig. 2d), which resulted in fewer possibilities for capture events of EVs onto nanowire surfaces.

In addition to the change in EV capture efficiency due to nanowire morphological changes, we also found a change in EV capture efficiency due to changes in nanowire crystallinity. The nanowires grown with  $\text{NH}_3$  addition for 9 h showed the lowest capture efficiency due to the low density of the nanowires, but the contribution of a single nanowire to EV capture was qualitatively found to be the highest, indicating that changes in the crystallinity of the nanowires were also important (Fig. 5d). To confirm the effect of the nanowire crystallinity on EV capture, we prepared ZnO nanowires of wurtzite ZnO phase with the same morphology by annealing the nanowires at 500 °C for 2 h (ref. 58–60) after nanowire growth since this annealing process is known to cause the phase transition from zinc blende to wurtzite (Fig. S6a† and Table 1). The nanowires grown with  $\text{NH}_3$  addition for 6 h followed by the annealing process exhibited lower EV capture efficiency than those grown without the annealing process (Fig. 5b and c). And also, the contribution of a single nanowire to EV capture showed that annealing process significantly decreased the number of captured EVs on a single nanowire, and then the zinc blende phase increased the number of captured EVs on a single nanowire (Fig. 5d and Table 2). Our results highlighted that designing and controlling the crystal structure of ZnO nanowires could be achieved by using  $\text{NH}_3$  addition to enhance the capability for label-free capture of EVs *via* the variations of their electrostatic interactions with the ZnO crystal plane. However,  $\text{NH}_3$  addition and the growth time also affected the morphology of nanowires leading to lower efficiency as the nanowires being larger in diameter and lower in density. Further studies are needed in order to achieve the optimum crystal structure and morphologies of nanowires for a specific application.

## Conclusions

This is the first study to identify the effects of crystallinity and morphology of ZnO nanowires grown hydrothermally with  $\text{NH}_3$  addition on their electrostatic properties for capturing of EVs. As found from results of the XRD and HRTEM analyses, the crystal structures of all samples had the main characteristics of the ZnO wurtzite phase, but they were also mixed with zinc blende phase when grown under some conditions, making them suitable for EV capture on the nanowires. Considering the crystallinity and the morphology of the nanowires, the nanowires grown for 6 h with  $\text{NH}_3$  addition offered the highest EV capture performance due to the presence of

zinc blende phase and high density of the nanowires. These findings are expected to facilitate the realization of tunable nanowires with high potential suitability for integration into advanced label-free EV capture devices which will open up opportunities for further fundamental studies (*i.e.*, adsorption mechanisms by biomolecules) and alternative applications in high-performance integrated nanowire devices.

## Author contributions

P. Paisrisarn contributed to conceptualization, formal analysis, investigation, methodology, visualization, writing – original draft and writing–review & editing. T. Yasui contributed to conceptualization, funding acquisition, methodology, project administration, supervision and writing–review & editing. Z. Zhu contributed to formal analysis, investigation and methodology. A. Klamchuen contributed to project administration, resources and supervision. P. Kasamechongchun, T. Wutikhun and V. Yordsri contributed to formal analysis, investigation and methodology. Y. Baba contributed to conceptualization, funding acquisition, project administration and supervision.

## Conflicts of interest

There are no conflicts to declare.

## Acknowledgements

This research was supported by the Japan Science and Technology Agency (JST) PRESTO (JPMJPR19H9), JST SICORP (JPMJSC19E3), the New Energy and Industrial Technology Development Organization (NEDO) JPNP20004, the JSPS Grant-in-Aid for Scientific Research (S) 18H05243, the JSPS Grant-in-Aid for Scientific Research (B) 21H01960, the JSPS Grant-in-Aid for Exploratory Research 20K21124, the JSPS Grant-in-Aid for Transformative Research Area (A) 21H05778, the JSPS Grant-in-Aid for Scientific Research on Innovative Areas “Chemistry for Multimolecular Crowding Biosystems”, a Medical Research and Development Program (AMED grand no. JP21he2302007 and JP21zf0127004) and research grants from each of the following: Advanced Technology Institute Research grants 2019, the Nanotechnology Platform Program (Molecule and Material Synthesis) of the Ministry of Education, Culture, Sports, Science and Technology (MEXT), and the Cooperative Research Program of the “Network Joint Research Center for Materials and Devices”.

## Notes and references

- 1 R. G. Hobbs, N. Petkov and J. D. Holmes, *Chem. Mater.*, 2012, **24**, 1975–1991.
- 2 B. Tian, T. J. Kempa and C. M. Lieber, *Chem. Soc. Rev.*, 2009, **38**, 16–24.



- 3 N. P. Dasgupta, J. W. Sun, C. Liu, S. Brittman, S. C. Andrews, J. Lim, H. W. Gao, R. X. Yan and P. D. Yang, *Adv. Mater.*, 2014, **26**, 2137–2184.
- 4 P. C. McIntyre and A. F. i Morral, *Mater. Today Nano*, 2020, **9**, 100058.
- 5 V. Gerbreder, M. Krasovska, E. Sledziskis, A. Gerbreder, I. Mihailova, E. Tamanis and A. Ogurcovs, *CrystEngComm*, 2020, **22**, 1346–1358.
- 6 P. Ambhorkar, Z. Wang, H. Ko, S. Lee, K.-I. Koo, K. Kim and D.-I. D. Cho, *Micromachines*, 2018, **9**, 679.
- 7 N. Caicedo, R. Leturcq, J.-P. Raskin, D. Flandre and D. Lenoble, *Sens. Actuators, B*, 2019, **297**, 126602.
- 8 B. Tian and C. M. Lieber, *Chem. Rev.*, 2019, **119**, 9136–9152.
- 9 A. M. Nahhas, *Am. J. Nanomater.*, 2021, **8**, 18–31.
- 10 T. Yasui, T. Yanagida, S. Ito, Y. Konakade, D. Takeshita, T. Naganawa, K. Nagashima, T. Shimada, N. Kaji, Y. Nakamura, I. A. Thiodorus, Y. He, S. Rahong, M. Kanai, H. Yukawa, T. Ochiya, T. Kawai and Y. Baba, *Sci. Adv.*, 2017, **3**, e1701133.
- 11 T. Suwatthanarak, I. A. Thiodorus, M. Tanaka, T. Shimada, D. Takeshita, T. Yasui, Y. Baba and M. Okochi, *Lab Chip*, 2021, **21**, 597.
- 12 M. Musa, T. Yasui, Z. Zhu, K. Nagashima, M. Ono, Q. Liu, H. Takahashi, T. Shimada, A. Arima, T. Yanagida and Y. Baba, *Anal. Sci.*, 2021, **37**, 1139–1145.
- 13 H. Takahashi, T. Yasui, H. Kashida, K. Makino, K. Shinjo, Q. Liu, T. Shimada, S. Rahong, N. Kaji, H. Asanuma and Y. Baba, *Nanotechnology*, 2021, **32**, 255301.
- 14 T. Yasui, P. Paisrisarn, T. Yanagida, Y. Konakade, Y. Nakamura, K. Nagashima, M. Musa, I. A. Thiodorus, H. Takahashi, T. Naganawa, T. Shimada, N. Kaji, T. Ochiya, T. Kawai and Y. Baba, *Biosens. Bioelectron.*, 2021, **194**, 113589.
- 15 H. Takahashi, T. Yasui, A. Klamchuen, N. Khemasiri, T. Wuthikhun, P. Paisrisarn, K. Shinjo, Y. Kitano, K. Aoki, A. Natsume, S. Rahong and Y. Baba, *Nanomaterials*, 2021, **11**, 1768.
- 16 Y. He, T. Yanagida, K. Nagashima, F. W. Zhuge, G. Meng, B. Xu, A. Klamchuen, S. Rahong, M. Kanai, X. M. Li, M. Suzuki, S. Kai and T. Kawai, *J. Phys. Chem. C*, 2013, **117**, 1197–1203.
- 17 V. Strano, R. G. Urso, M. Scuderi, K. O. Iwu, F. Simone, E. Ciliberto, C. Spinella and S. Mirabella, *J. Phys. Chem. C*, 2014, **118**, 28189–28195.
- 18 P. Kasamechong, M. Horprathum, K. Boonpavanitchakul, N. Supaka, P. Prompinit, W. Kangwansupamonkon, A. Somboonkaew, J. Wetcharungsri, S. Pratontep, S. Porntheeraphat and A. Klamchuen, *Phys. Status Solidi A*, 2015, **212**, 394–400.
- 19 Q. Huang, L. Fang, X. Chen and M. R. Saleem, *J. Alloys Compd.*, 2011, **509**, 9456–9459.
- 20 L.-Y. Chen, Y.-T. Yin, C.-H. Chen and J.-W. Chiou, *J. Phys. Chem. C*, 2011, **115**, 20913–20919.
- 21 T. Demes, C. Terner, F. Morisot, D. Riassetto, M. Legallais, H. Roussel and M. Langlet, *Appl. Surf. Sci.*, 2017, **410**, 423–431.
- 22 R. Hao, X. Deng, J. Zhang, H. Jang, Y. Zeng and H. Guo, *Proc. 2017 Symp. Piezoelectricity Acoust. Waves Device Appli. SPAWDA*, 2017, 345–349.
- 23 R. Parize, J. D. Garnier, E. Appert, O. Chaix-Pluchery and V. Consonni, *ACS Omega*, 2018, **3**, 12457–12464.
- 24 J. Liu, K. Nagashima, H. Yamashita, W. Mizukami, J. Uzuhashi, T. Hosomi, M. Kanai, X. Zhao, Y. Miura, G. Zhang, T. Takahashi, M. Suzuki, D. Sakai, B. Samransuksamer, Y. He, T. Ohkubo, T. Yasui, Y. Aoki, J. C. Ho, Y. Baba and T. Yanagida, *Commun. Mater.*, 2020, **1**, 58.
- 25 J. Joo, B. Y. Chow, M. Prakash, E. S. Boyden and J. M. Jacobson, *Nat. Mater.*, 2011, **10**, 596–601.
- 26 T. H. Kwon, K. Kim, S. H. Park, A. A. Annamalai and M. J. Lee, *Int. J. Nanotechnol.*, 2013, **10**, 681.
- 27 L. Liu, K. Hong, X. Ge, D. Liu and M. Xu, *J. Phys. Chem. C*, 2014, **118**, 15551–15555.
- 28 R. Kumar, O. Al-Dossary, G. Kumar and A. Umar, *Nano-Micro Lett.*, 2015, **7**, 97–120.
- 29 J. Wang, X. Li, C. Teng, Y. Xia, J. Xu, D. Xie, L. Xiang and S. Komarneni, *J. Mater. Chem. C*, 2016, **4**, 5755–5765.
- 30 S. V. Kurudirek, K. C. Pradel and C. J. Summers, *J. Alloys Compd.*, 2017, **702**, 700–709.
- 31 A. C. Campos, S. C. Paes, B. S. Correa, G. A. Cabrera-Pasca, M. S. Costa, C. S. Costa, L. Otubo and A. W. Carbonari, *ACS Appl. Nano Mater.*, 2020, **3**, 175–185.
- 32 M. Yanez-Mo, P. R. Siljander, Z. Andreu, A. B. Zavec, F. E. Borrás, E. I. Buzas, K. Buzas, E. Casal, F. Cappello, J. Carvalho, E. Colas, A. Cordeiro-da Silva, S. Fais, J. M. Falcon-Perez, I. M. Ghobrial, B. Giebel, M. Gimona, M. Graner, I. Gursel, M. Gursel, N. H. Heegaard, A. Hendrix, P. Kierulf, K. Kokubun, M. Kosanovic, V. Kralj-Iglic, E. M. Kramer-Albers, S. Laitinen, C. Lasser, T. Lener, E. Ligeti, A. Line, G. Lipps, A. Llorente, J. Lotvall, M. Mancek-Keber, A. Marcilla, M. Mittelbrunn, I. Nazarenko, E. N. M. N. Hoen, T. A. Nyman, L. O'Driscoll, M. Olivan, C. Oliveira, E. Pallinger, H. A. Del Portillo, J. Reventos, M. Rigau, E. Rohde, M. Sammar, F. Sanchez-Madrid, N. Santarem, K. Schallmoser, M. S. Ostfeld, W. Stoorvogel, R. Stukelj, S. G. Van der Grein, M. H. Vasconcelos, M. H. Wauben and O. De Wever, *J. Extracell. Vesicles*, 2015, **4**, 27066.
- 33 G. van Niel, G. D'Angelo and G. Raposo, *Nat. Rev. Mol. Cell Biol.*, 2018, **19**, 213–228.
- 34 L. M. Doyle and M. Z. Wang, *Cells*, 2019, **8**, 727.
- 35 R. Xu, A. Rai, M. Chen, W. Suwakulsiri, D. W. Greening and R. J. Simpson, *Nat. Rev. Clin. Oncol.*, 2018, **15**, 617–638.
- 36 L. Han, E. W. F. Lam and Y. Sun, *Mol. Cancer*, 2019, **18**, 59.
- 37 Y. Gao, Y. Qin, C. Wan, Y. Sun, J. Meng, J. Huang, Y. Hu, H. Jin and K. Yang, *Front. Oncol.*, 2021, **11**, 638357.
- 38 L. K. Chin, T. Son, J.-S. Hong, A.-Q. Liu, J. Skog, C. M. Castro, R. Weissleder, H. Lee and H. Im, *ACS Nano*, 2020, **14**, 14528–14548.
- 39 S. Sharma, M. K. Masud, Y. V. Kaneti, P. Rewatkar, A. Koradia, M. S. A. Hossain, Y. Yamauchi, A. Popat and C. Salomon, *Small*, 2021, **17**, e2102220.
- 40 Z. Zhuo, J. Wang, Y. Luo, R. Zeng, C. Zhang, W. Zhou, K. Guo, H. Wu, W. Sha and H. Chen, *Acta Biomater.*, 2021, **134**, 13–31.





- 41 A. Hoshino, B. Costa-Silva, T.-L. Shen, G. Rodrigues, A. Hashimoto, M. T. Mark, H. Molina, S. Kohsaka, A. Di Giannatale, S. Ceder, S. Singh, C. Williams, N. Soplop, K. Uryu, L. Pharmer, T. King, L. Bojmar, A. E. Davies, Y. Ararso, T. Zhang, H. Zhang, J. Hernandez, J. M. Weiss, V. D. Dumont-Cole, K. Kramer, L. H. Wexler, A. Narendran, G. K. Schwartz, J. H. Healey, P. Sandstrom, K. J. Labori, E. H. Kure, P. M. Grandgenett, M. A. Hollingsworth, M. de Sousa, S. Kaur, M. Jain, K. Mallya, S. K. Batra, W. R. Jarnagin, M. S. Brady, O. Fodstad, V. Muller, K. Pantel, A. J. Minn, M. J. Bissell, B. A. Garcia, Y. Kang, V. K. Rajasekhar, C. M. Ghajar, I. Matei, H. Peinado, J. Bromberg and D. Lyden, *Nature*, 2015, **527**, 329–335.
- 42 S. A. Melo, L. B. Luecke, C. Kahlert, A. F. Fernandez, S. T. Gammon, J. Kaye, V. S. LeBleu, E. A. Mittendorf, J. Weitz, N. Rahbari, C. Reissfelder, C. Pilarsky, M. F. Fraga, D. Piwnica-Worms and R. Kalluri, *Nature*, 2015, **523**, 177–182.
- 43 H. Zhang, D. Freitas, H. S. Kim, K. Fabijanic, Z. Li, H. Chen, M. T. Mark, H. Molina, A. B. Martin, L. Bojmar, J. Fang, S. Rampersaud, A. Hoshino, I. Matei, C. M. Kenific, M. Nakajima, A. P. Mutvei, P. Sansone, W. Buehring, H. Wang, J. P. Jimenez, L. Cohen-Gould, N. Paknejad, M. Brendel, K. Manova-Todorova, A. Magalhães, J. A. Ferreira, H. Osório, A. M. Silva, A. Massey, J. R. Cubillos-Ruiz, G. Galletti, P. Giannakakou, A. M. Cuervo, J. Blenis, R. Schwartz, M. S. Brady, H. Peinado, J. Bromberg, H. Matsui, C. A. Reis and D. Lyden, *Nat. Cell Biol.*, 2018, **20**, 332–343.
- 44 C. Xu, P. Shin, L. Cao and D. Gao, *J. Phys. Chem. C*, 2010, **114**, 125–129.
- 45 Q. L. Liu, T. Yasui, K. Nagashima, T. Yanagida, M. Hara, M. Horiuchi, Z. T. Zhu, H. Takahashi, T. Shimada, A. Arima and Y. Baba, *J. Phys. Chem. C*, 2020, **124**, 20563–20568.
- 46 M. Law, L. E. Greene, J. C. Johnson, R. Saykally and P. D. Yang, *Nat. Mater.*, 2005, **4**, 455–459.
- 47 Y. Y. Zhang, M. K. Ram, E. K. Stefanakos and D. Y. Goswami, *J. Nanomater.*, 2012, **2012**, 624520.
- 48 A. Ashrafi and C. Jagadish, *J. Appl. Phys.*, 2007, **102**, 071101.
- 49 N. Munoz-Aguirre, L. Martinez-Perez, S. Munoz-Aguirre, L. A. Flores-Herrera, E. V. Hernandez and O. Zelaya-Angel, *Materials*, 2019, **12**, 3314.
- 50 X. Huang, M. G. Willinger, H. Fan, Z. L. Xie, L. Wang, A. Klein-Hoffmann, F. Girgsdies, C. S. Lee and X. M. Meng, *Nanoscale*, 2014, **6**, 8787–8795.
- 51 S. Boubenia, A. S. Dahiya, G. Poulin-Vittrant, F. Morini, K. Nadaud and D. Alquier, *Sci. Rep.*, 2017, **7**, 15187.
- 52 H. J. Joyce, J. Wong-Leung, Q. Gao, H. H. Tan and C. Jagadish, *Nano Lett.*, 2010, **10**, 908–915.
- 53 K. Ikejiri, Y. Kitauchi, K. Tomioka, J. Motohisa and T. Fukui, *Nano Lett.*, 2011, **11**, 4314–4318.
- 54 A. Jenichen, C. Engler and B. Rauschenbach, *Surf. Sci.*, 2013, **613**, 74–79.
- 55 M. Ghosh, D. Karmakar, S. Basu, S. N. Jha, D. Bhattacharyya, S. C. Gadkari and S. K. Gupta, *J. Phys. Chem. Solids*, 2014, **75**, 543–549.
- 56 K. Momeni and H. Attariani, *Phys. Chem. Chem. Phys.*, 2014, **16**, 4522–4527.
- 57 G. Midekessa, K. Godakumara, J. Ord, J. Viil, F. Lättekivi, K. Dissanayake, S. Kopanchuk, A. Rinken, A. Andronowska, S. Bhattacharjee, T. Rinken and A. Fazeli, *ACS Omega*, 2020, **5**, 16701–16710.
- 58 Z. N. Kayani, F. Saleemi and I. Batool, *Appl. Phys. A*, 2015, **119**, 713–720.
- 59 M. R. Parra and F. Z. Haque, *J. Mater. Res. Technol.*, 2014, **3**, 363–369.
- 60 E. Y. Shaba, J. O. Jacob, J. O. Tijani and M. A. T. Suleiman, *Appl. Water Sci.*, 2021, **11**, 48.

