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Automated glycan assembly of peptidoglycan backbone fragments†

Pietro Dallabernardina,^a Valentina Benazzi,^{a,b} Jon D. Laman,^c
Peter H. Seeberger^{a,d} and Felix F. Loeffler^{ID} ^{*,a}

We report the automated glycan assembly (AGA) of different oligosaccharide fragments of the bacterial peptidoglycan (PGN) backbone. Iterative addition on a solid support of an acetyl glucosamine and a new muramic acid building block is followed by cleavage from the solid support and final deprotection providing 10 oligosaccharides up to six units.

Peptidoglycans (PGN) are an essential component of the cell wall of both Gram-positive and Gram-negative bacteria. PGN serve a structural function, counteracting osmotic pressure and maintaining cell shape, and protect against external threats.^{1,2} The main PGN structure is made of linear glycan strands cross-linked by short peptide chains (Fig. 1b).³ Several structural modifications at both the glycan part, such as acetylation, and the stem peptide are possible in order to give rise to hundreds of structural variants.⁴ Nevertheless, the structure of the glycan backbone is generally conserved and it is composed of two alternating amino sugars (Fig. 1a), *N*-acetylglucosamine (GlcNAc) and *N*-acetylmuramic acid (MurNAc), linked by a β -1-4 bond. The D-lactoyl group of each MurNAc residue is connected through an amide bond to a stem peptide that can be up to five amino acid residues long.⁵ Many modifications can occur at the peptide chain,² in particular at the third amino acid. The majority of Gram-positive bacteria have an L-Lys at position three, while the majority of Gram-negative bacteria have a non-proteinogenic amino acid A₂pm (2,6-diaminopimelic acid). The final dipeptide might present some modifications due to acquired resistance to antibiotics.⁶ Stem pep-

tides from different glycan strands may be cross-linked either directly or by an interpeptide bridge consisting of one to seven amino acid residues.²

In the past two decades, PGNs drew interest not only as a possible target for antibiotics,⁷ but also for their role as immunostimulatory molecules,^{8–11} as well as in mouse brain development and behavior.¹² It was suggested that PGN fragments have many diverse roles as signalling molecules, including communication between bacteria and bacteria with the host, as well as pathogenesis in animals and plants.⁵ PGNs have also been correlated with some autoimmune and chronic inflammatory diseases.^{13–16} In particular, some studies^{14,17–19} have demonstrated that peptidoglycan fragments are present within phagocytes in the brain of multiple sclerosis patients and that anti-PGN antibodies are present in cerebrospinal fluid during active disease, suggesting a possible link between

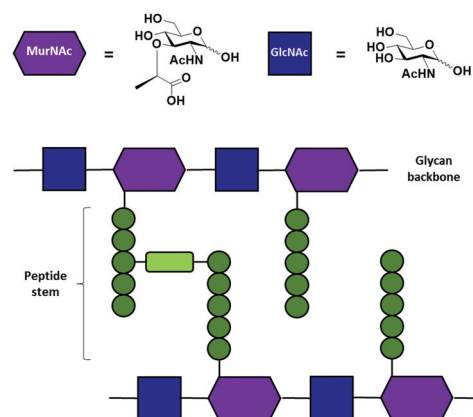


Fig. 1 (a) Symbolic representation and structures of the two amino sugars, *N*-acetylglucosamine (GlcNAc) and *N*-acetylmuramic acid (MurNAc). (b) Schematic representation of the peptidoglycan structure. The different glycan strands are branched on the MurNAc residues with a sequence of amino acids (dark green circles) called stem peptide. An interpeptide bridge (light green box) may connect the two stem peptides.

^aDepartment of Biomolecular Systems, Max Planck Institute of Colloids and Interfaces, Am Muehlenberg 1, 14476 Potsdam, Germany.
E-mail: felix.loeffler@mpikg.mpg.de

^bUniversity of Pavia, Department of Organic Chemistry, V.le Torquato Taramelli, 10, 27100 Pavia, Italy

^cDepartment of Pathology & Medical Biology, University Medical Center Groningen, Groningen, The Netherlands

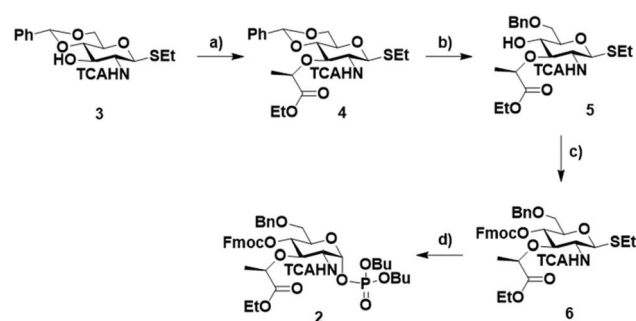
^dFreie Universität Berlin, Institute of Chemistry and Biochemistry, Arnimallee 22, 14195 Berlin, Germany

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Fluorenylmethoxycarbonyl (Fmoc) was installed at C4 and used as a temporary protecting group for β 1–4 chain elongation. A lactic ester was installed in position three of the BB 2 as precursor for the lactic acid moiety of the muramic acid. All other positions were protected with benzyl (Bn) permanent protecting groups. The phosphate leaving group was preferred to other leaving groups due to observed higher coupling efficiency in AGA for glucosamine BBs.²⁸ The two desired BBs 1 and 2 are accessible starting from the unprotected glucos-

Subsequently, the Fmoc carbonate was cleaved from the structure, preparing the monosaccharide for the installation of



Scheme 2 Synthesis of the MurNAC BB 2. Reagents and conditions: (a) (–)-ethyl (S)-2-trifluoromethylsulfonyloxy propionate, NaH, DMF/CH₂Cl₂, 0 °C → rt, 53%; (b) Me₃N·BH₃, BF₃·OEt₂, CH₃CN, 0 °C → rt, 65%; (c) FmOCl, pyridine, CH₂Cl₂, rt, 93%; (d) (BuO)₂P(O)OH, NIS, TfOH, 0 °C → rt, 83%.



Scheme 3 Automated glycan assembly of PGN oligosaccharides. Reagents and conditions: (a) 1 × 5 equiv. BB1, TMSOTf, CH₂Cl₂, −30 °C (5 min) → −15 °C (35 min); (b) 1 cycle of a 10% acetic anhydride and 2% methanesulfonic acid in CH₂Cl₂, 25 °C; (c) 3 cycles of 20% piperidine in DMF, 25 °C (5 min) (Module B); (d) CH₂Cl₂, *hν* (365 nm); (e) LiOH, THF/1,4-dioxane/H₂O, 2–24 h; (f) H₂, Pd/C, H₂O/HOAc, 16–48 h. **8**: 27%; **9**: 42%; **10**: 20%; **11**: 10%; **12**: 11%; **13**: 9%; **14**: 3%; **15**: 8%; **16**: 3%; **17**: 2%.

the next unit. This cycle of coupling, capping, and deprotection was repeated until the desired sequence was achieved. Once AGA was completed, the oligosaccharides were cleaved from the resin using UV light (365 nm) in a continuous flow photo-reactor.³⁵ After purification of the desired oligosaccharides from the deletion sequences, the basic hydrolysis of the lactoyl esters to carboxylic acids was performed using LiOH in a solution of THF/dioxane/H₂O.³¹ The remaining protecting groups were cleaved by hydrogenolysis with Pd/C and H₂. Under these conditions, the TCA group was reduced to acetamide. This last step was more problematic, since after several days of hydrogenolysis, we still observed the presence of multiple chlorine atoms by MALDI. Therefore, for the hexamers were hydrogenated under high pressure to achieve complete conversion in 48 h. The relatively low yield is caused by the inefficiency of the photocleavage from the resin together with the rather low reactivity of the building blocks, which led to the presence of deletion sequences. Finally, ten different PGN glycan backbones of alternating MurNAc and GlcNAc moieties, equipped with an amino-linker were obtained: the two monomers **8** and **9**, two disaccharides **10** and **11**, two trimers **12** and **13**, two tetramers **14** and **15**, and two hexamers **16** and **17**.

Conclusions

Ten oligosaccharide fragments of the PGN backbone were synthesized. The use of a new MurNAc BB enabled the fast synthesis of a collection of oligosaccharides up to hexamers. These oligosaccharides can be used to create glycan microarrays for the characterization of peptidoglycan directed antibodies. Advancing the idea from recently published approaches, where molecules on microarrays were modified directly on the glass slide after *in situ* synthesis³⁷ or printing,³⁸ we will follow a similar strategy. We envision to couple different stem peptide variants from solution, forming the amide bond directly with the arrayed oligosaccharides. These arrays will support the identification of epitopes recognized by anti-PGN antibodies, and can potentially help to close current knowledge gaps in autoimmune disorders.

Conflicts of interest

There are no conflicts to declare.



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