


Cite this: *RSC Adv.*, 2020, 10, 41353

Recent advances in the pharmacological diversification of quinazoline/quinazolinone hybrids

Prashant S. Auti,[†] Ginson George[†] and Atish T. Paul ^{‡*}

Due to the pharmacological activities of quinazoline and quinazolinone scaffolds, it has aroused great interest in medicinal chemists for the development of new drugs or drug candidates. The pharmacological activities of quinazoline and its related scaffolds include anti-cancer, anti-microbial, anti-convulsant, and antihyperlipidaemia. Recently, molecular hybridization technology is used for the development of hybrid analogues with improved potency by combining two or more pharmacophores of bioactive scaffolds. The molecular hybridization of various biologically active pharmacophores with quinazoline derivatives resulted in lead compounds with multi-faceted biological activity wherein specific as well as multiple targets were involved. The present review summarizes the advances in lead compounds of quinazoline hybrids and their related heterocycles in medicinal chemistry. Moreover, the review also helps to intensify the drug development process by providing an understanding of the potential role of these hybridized pharmacophoric features in exhibiting various pharmacological activities.

Received 31st July 2020
Accepted 27th October 2020

DOI: 10.1039/d0ra06642g

rsc.li/rsc-advances

1. Introduction

Nowadays, “one drug one target one disease approach” is not appropriate in complex diseases such as cancer and infectious diseases.^{1,2} This conventional approach of disease treatment cannot overcome the phenomenon of drug resistance. Consequently, polypharmacy may prove beneficial by the use of two or more drugs that interact simultaneously at different targets. However, the use of multiple drug combinations is often linked with drug–drug interactions and altered pharmacokinetics, leading to severe unintended outcomes that may result in the failure of the treatment. Altered pharmacokinetics and pharmacodynamics of drugs used in combination may be due to the induction or inhibition of hepatic enzymes, which are responsible for drug metabolism.¹ Hence, in order to modulate more than one disease target simultaneously, molecular hybrids were designed by combining two or more known active pharmacophores.³ A molecular hybrid is a combination of two or more independently acting pharmacophores that are covalently linked. It can be achieved by the method of linking or framework integration to form one molecule, having increased the pharmacological activity.⁴ The strategy of designing such hybrid

molecules is widely known as molecular hybridization. The molecular hybridization strategy is emerging as a novel approach, involving the conglomeration of two or more pharmacophores in one scaffold to develop hybrid multi-functional molecules.⁵ The presence of two or more biologically active pharmacophores in one unit will synergize the biological activity as well as increase its ability to interact with more than one biological target.⁶ Due to such properties of hybrid analogues, it is possible to develop combination therapies using single multi-functional molecules.

Quinazoline is a nitrogen containing fused heterocycle, having four isomeric forms, *i.e.*, quinazoline, quinoxaline, cinnoline, and phthalazine, depending upon the position of the nitrogen atom in the heterocyclic ring system.⁷ Carbonyl linkage on the quinazoline ring will give rise to quinazolinone (4(3*H*)-quinazolinone & 2(1*H*)-quinazolinone).

Amongst all the isomeric forms, quinazoline and quinazolinone are medicinally important nitrogen heterocycles, displaying diverse biological activities including anti-microbial,⁸ anti-convulsant,^{9,10} anti-cancer,¹¹ anti-malarial,¹² anti-hypertensive,¹³ anti-inflammatory,¹⁴ anti-diabetic,¹⁵ anti-tumor,¹⁶ anti-cholinesterase,¹⁷ dihydrofolate reductase inhibition,¹⁸ cellular phosphorylation inhibition,¹⁹ and kinase inhibitory activities.²⁰ There are many synthetic and natural product-based drugs, containing quinazoline and quinazolinone moiety, which are used clinically for treating various disease conditions (Fig. 1 and 2).²¹

Hybrid analogues containing quinazoline/quinazolinone and other biologically active pharmacophores may result in drugs with increased potency and reduced drug resistance. There are many reports available on the design of various

Laboratory of Natural Product Chemistry, Department of Pharmacy, Birla Institute of Technology and Science, Pilani (BITS Pilani), Pilani Campus, Pilani, 333 031, Rajasthan, India. E-mail: atish.paul@pilani.bits-pilani.ac.in

[†] Equal contribution.

[‡] Present address: Laboratory of Natural Product Chemistry, Department of Pharmacy, Birla Institute of Technology and Science, Pilani (BITS Pilani), Pilani Campus, Pilani – 333031, Rajasthan, India.



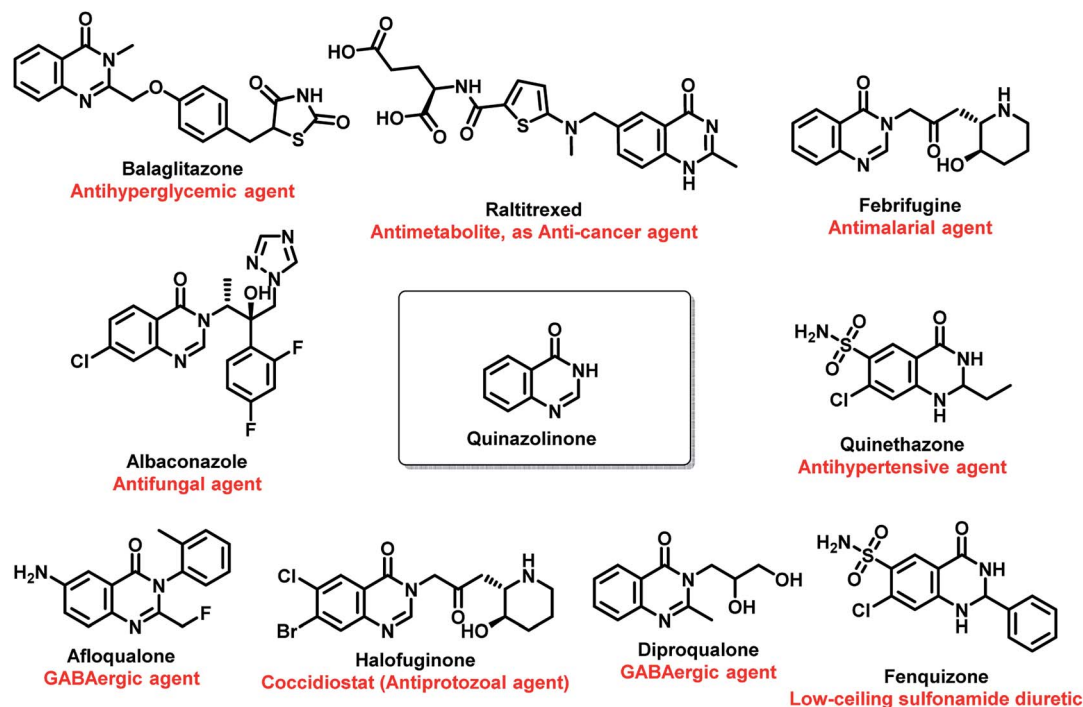


Fig. 1 Pharmacological importance of quinazolinone-based drugs.

quinazoline/quinazolinone-based hybrid analogues against a variety of disease conditions.

In the current review, different strategies and approaches that were employed in the design of quinazoline/4(3*H*)-quinazolinone-

based hybrids is discussed along with their pharmacological activities (in the areas of cancer, infectious diseases, diabetes, and CNS disorders) and the effect of various substituents on the pharmacological activity.

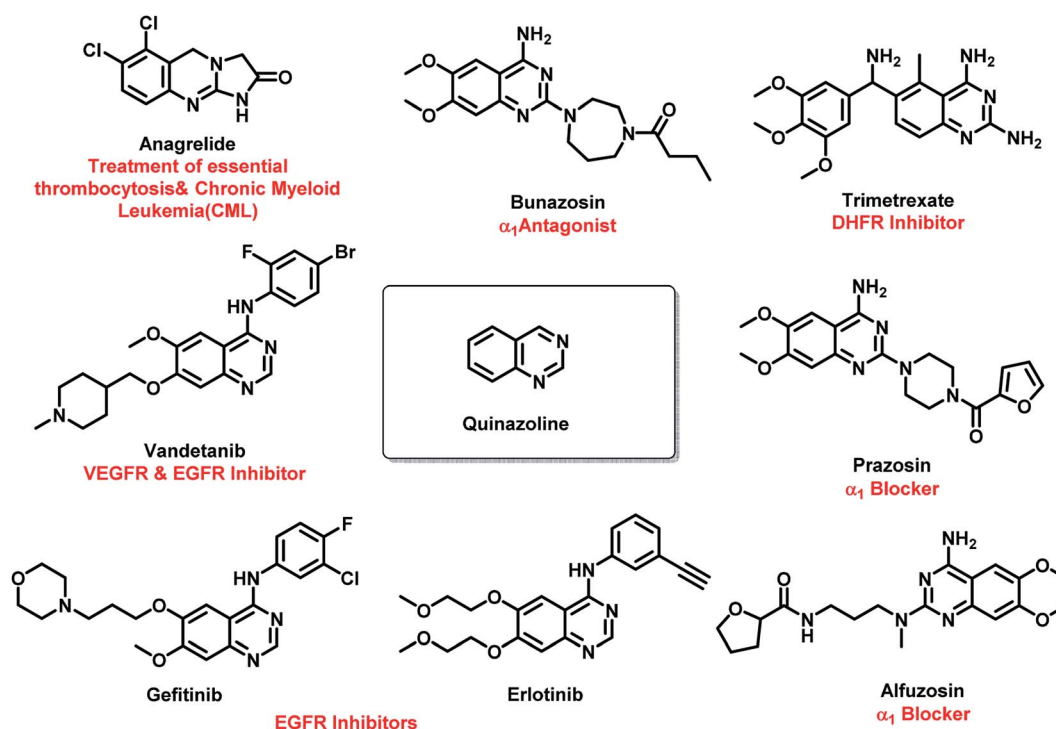
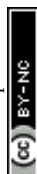
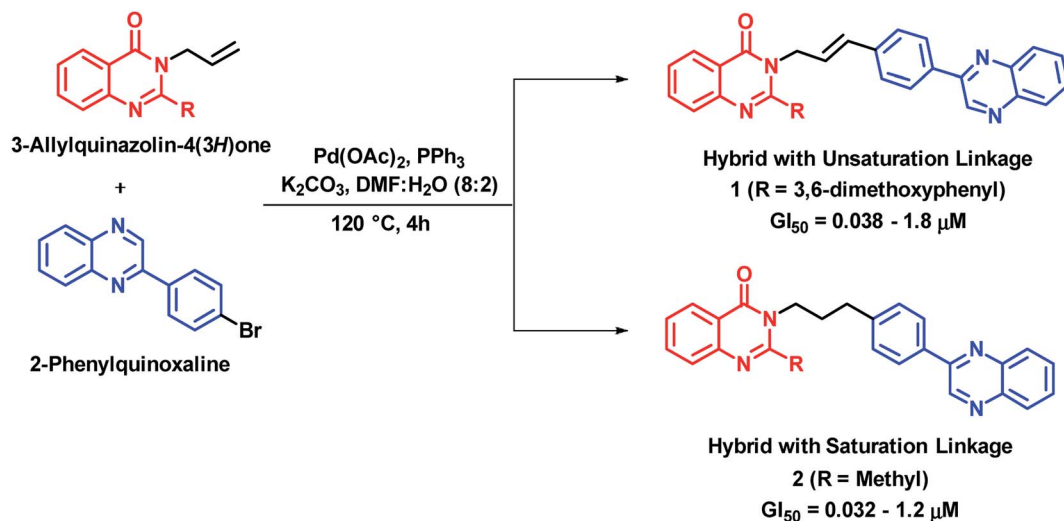


Fig. 2 Pharmacological importance of quinazoline-based drugs.





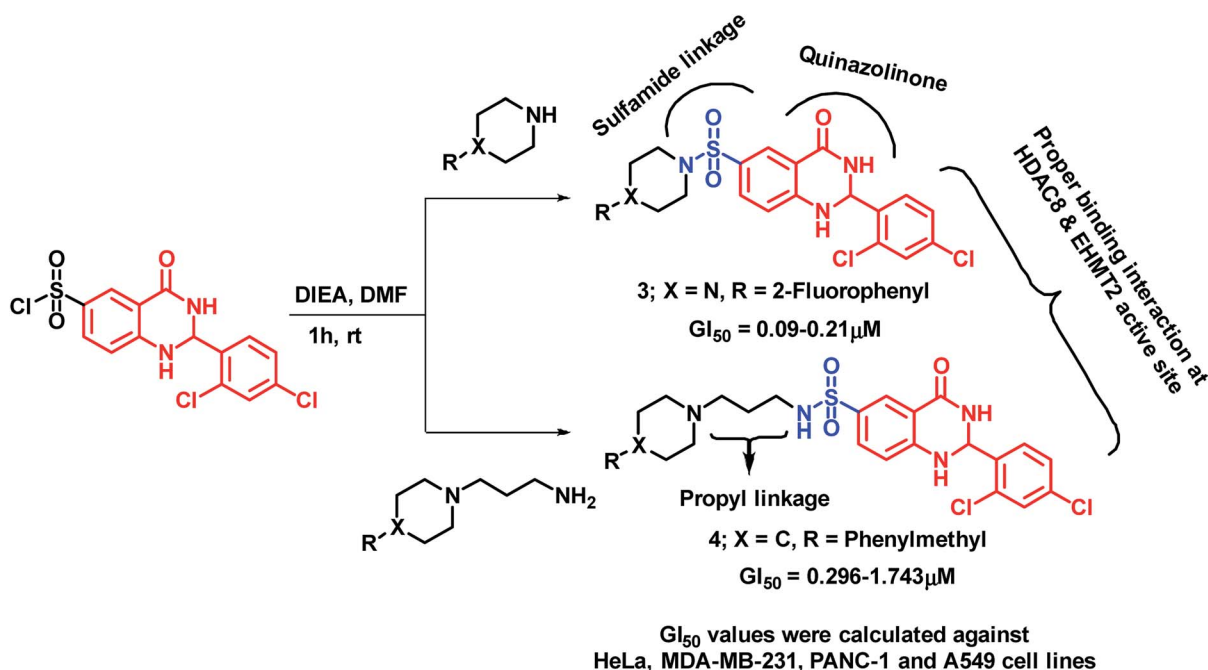
Scheme 1 Synthetic scheme for quinazolinone-phenylquinoxaline hybrids.

2. Anti-cancer hybrids

Cancer is a cell cycle disease, associated with uncontrolled cell division of abnormal cells. It continues to be one of the major health problems all over the world. There are many anti-cancer drugs in the market but still the demand is evergreen due to drug resistance. Hybrid drugs with multiple mechanism of action may be of great value for cancer treatment. Some of the hybrid analogues based on quinazoline and quinazolinone moiety are discussed below.

Palem *et al.* synthesized novel hybrid analogues by the combination of quinazolinone and allylphenyl quinoxaline and

evaluated their anti-cancer potential against 4 cancer cell lines (HeLa, MIA-PACA, MDA-MB-231, and IMR32).²² Palladium (Pd)-catalyzed Heck cross coupling was utilized for the coupling of halogenated quinoxaline with substituted 3-allylquinazolinones. Two series of hybrid analogues (with alkenyl and alkyl linkages) were synthesized. These compounds of both the series have shown significant to moderate anti-cancer activity ($\text{GI}_{50} = 0.08\text{--}0.2 \mu\text{M}$) against all the four cancer cell lines, as compared with doxorubicin ($\text{GI}_{50} = 0.023\text{--}0.097 \mu\text{M}$) and paclitaxel ($\text{GI}_{50} = 0.025\text{--}0.091 \mu\text{M}$), which were used as standards. Compound 1 with alkenyl linker ($\text{GI}_{50} = 0.038\text{--}1.8 \mu\text{M}$) and compound 2 with alkyl linker ($\text{GI}_{50} = 0.032\text{--}1.2 \mu\text{M}$) were most potent against



Scheme 2 Synthetic scheme for sulfamide-linked quinazolinone hybrids.

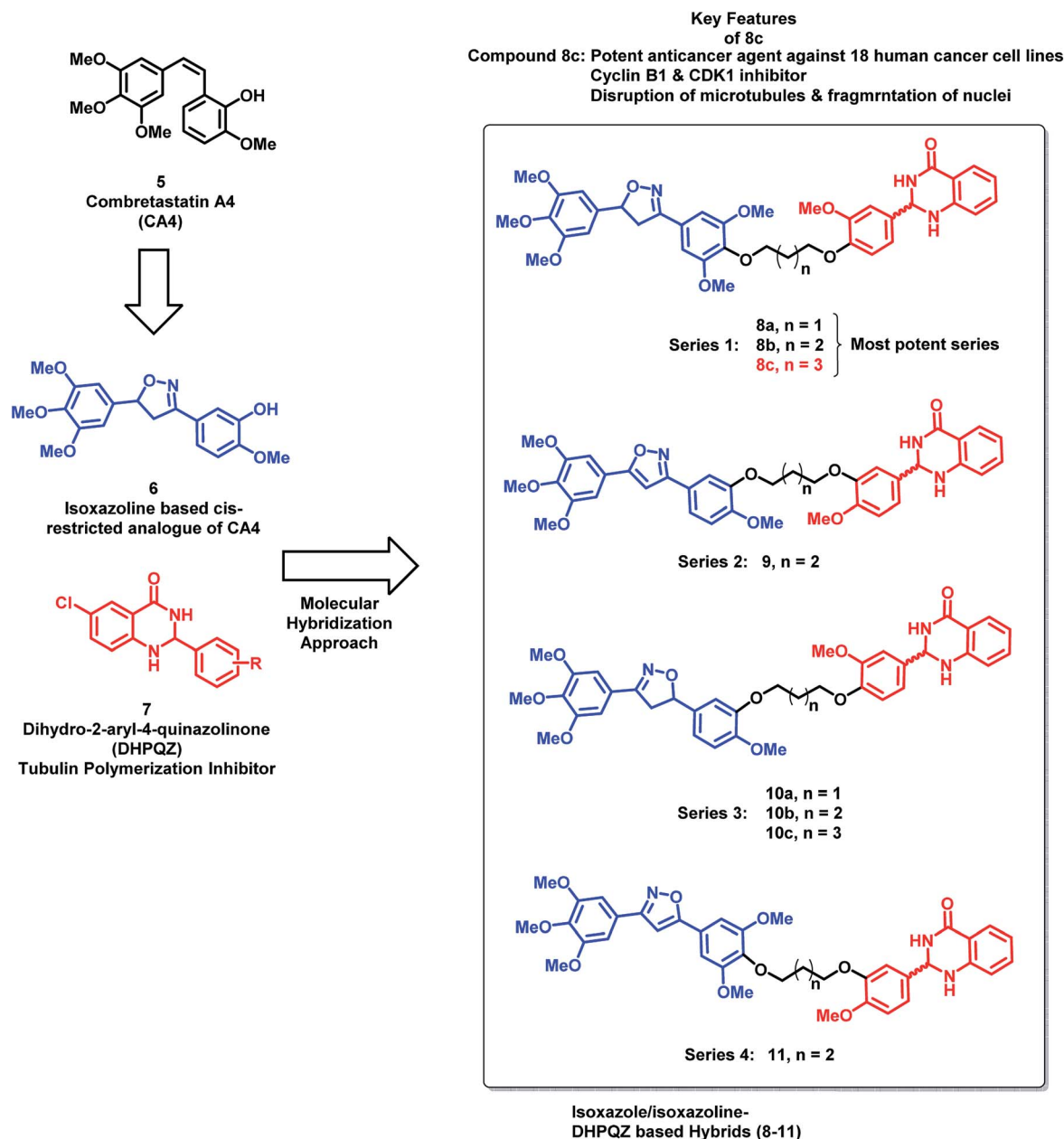


Fig. 3 Designing strategy for oxazoline- and dihydroquinazolinone-based hybrids.

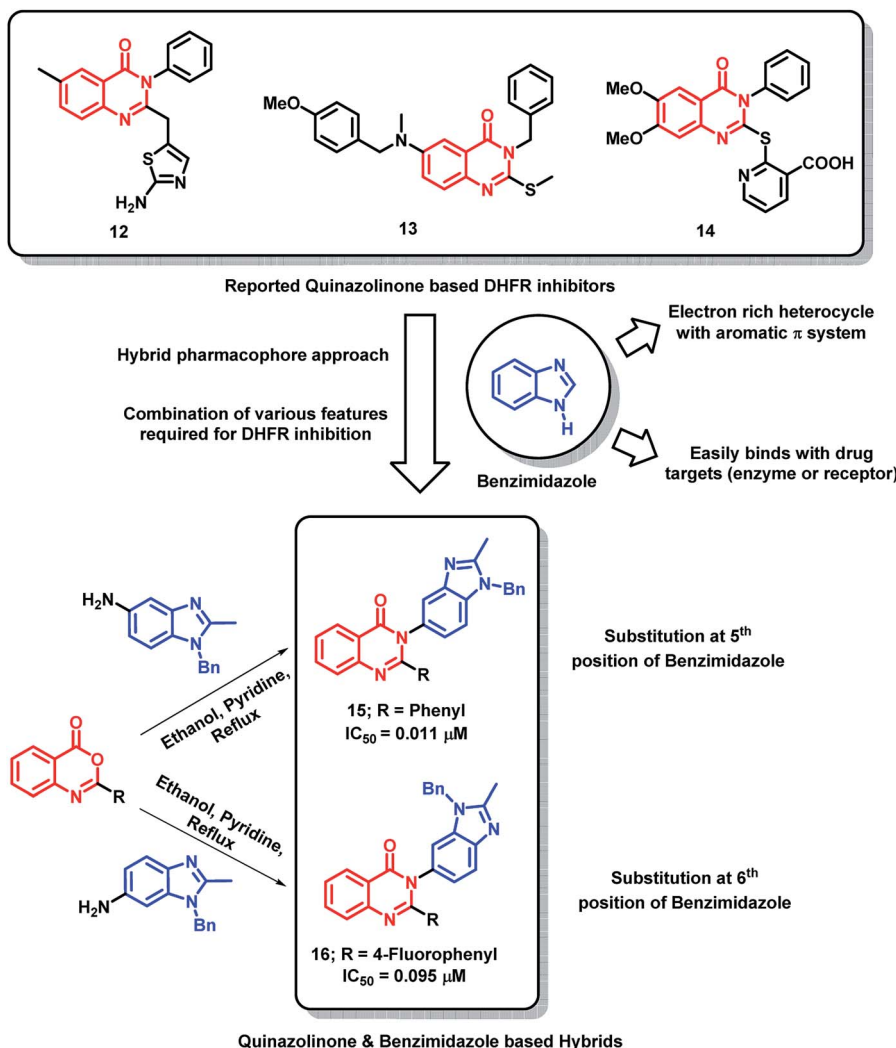
above cell lines (Scheme 1). The designed analogues were found to intercalate with DNA, when analyzed by docking interactions with the DNA molecule (AGACGTCT)₂ (PDB code: 1N37).

Venkatesh *et al.* have designed and synthesized sulphamide-linked dihydroquinazolinone hybrids and evaluated their anti-cancer potential on various cancer cell lines (HeLa, MDA-MB-231, PANC-1, A549 (ref. 23)). Series of compounds, with or without aliphatic linkage, have shown significant growth inhibition against all the tested cell lines with GI₅₀ in the range of 0.045–6.94 μM. Amongst them, compound 3 was found to be potent (GI₅₀ in the range of 0.09–0.21 μM). It was found that the alkyl linkage is also important from the activity point of view, as compound 4 with propyl linkage has shown comparable potency

(GI₅₀ in the range of 0.296–1.743 μM) as that of compound 3 (Scheme 2).

Kamal *et al.*²⁴ have designed hybrid analogues based on 2,3-dihydro-2-aryl-4-quinazolinones (DHPQZ) and diaryl-substituted isoxazoline/isoxazole, wherein both the scaffolds were reported for tubulin polymerization inhibitory activity.^{25–28} Four series of hybrid analogues were designed and synthesized by linking DHPQZ and isoxazoline/isoxazole with the alkyl chain linkers of varied chain length (Fig. 3) compound 8c was found to have potential anti-cancer activity, when tested against 18 human cancer cell lines. Compound 8c was found to cause significant disruption of microtubules in MCF-7 cells along with nuclear fragmentation (analyzed by DAPI staining). Further, the flow



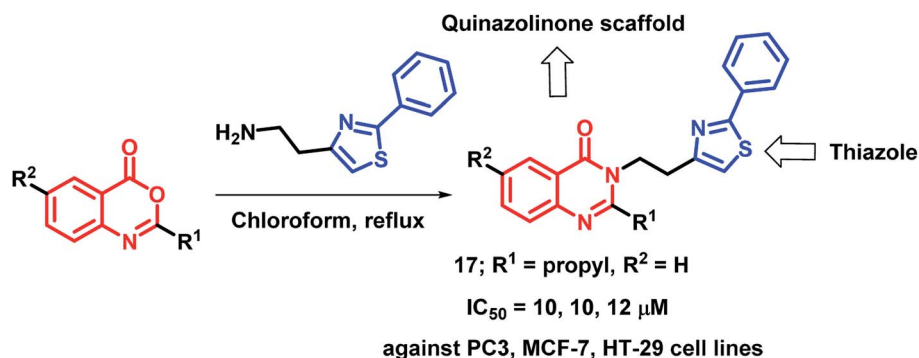


Scheme 3 Synthetic and design strategy for quinazolinone- and benzimidazole-based hybrids as DHFR inhibitors.

cytometric analysis of **8c** suggested the G2/M phase cell cycle arrest.

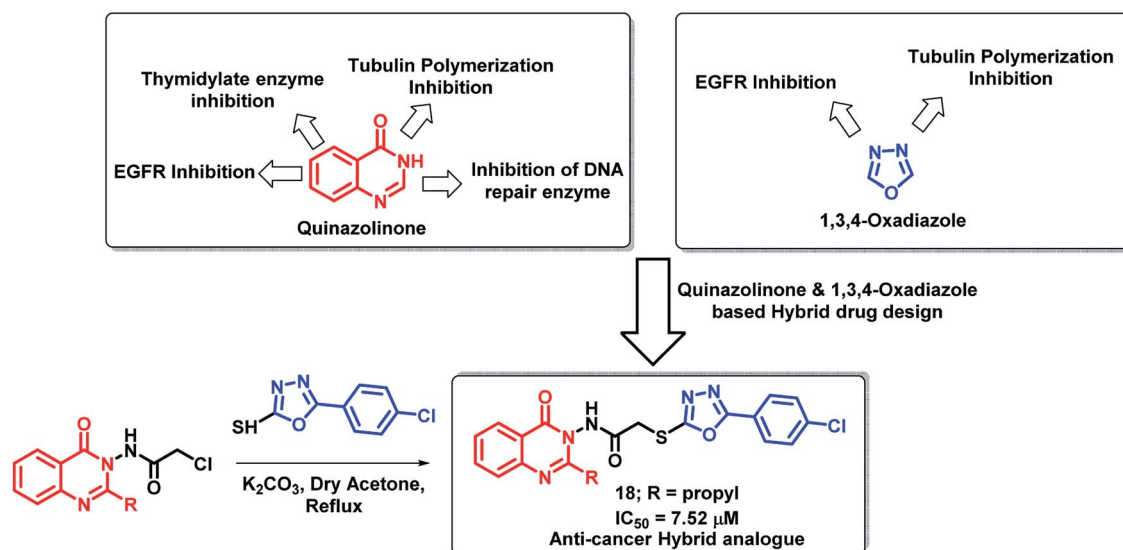
In the search for potent DHFR inhibitors, Paul *et al.* have modified existing quinazolinone-based DHFR inhibitors (**12–14**)^{29–31} with the aim of increasing the hydrophobic π -interaction

with the DHFR receptor.³² For such an attempt, the phenyl ring at the 3rd position of quinazolinone was replaced by benzimidazole, which is beneficial for exhibiting additional π -interactions due to aromatic π -system with electron rich properties (Scheme 3). Among all the synthesized analogues, hybrid analogue **15** was



Scheme 4 Synthetic scheme for quinazolinone- and thiazole-based hybrids.

Anti-cancer Mechanism of Quinazolinone & Triazole scaffolds

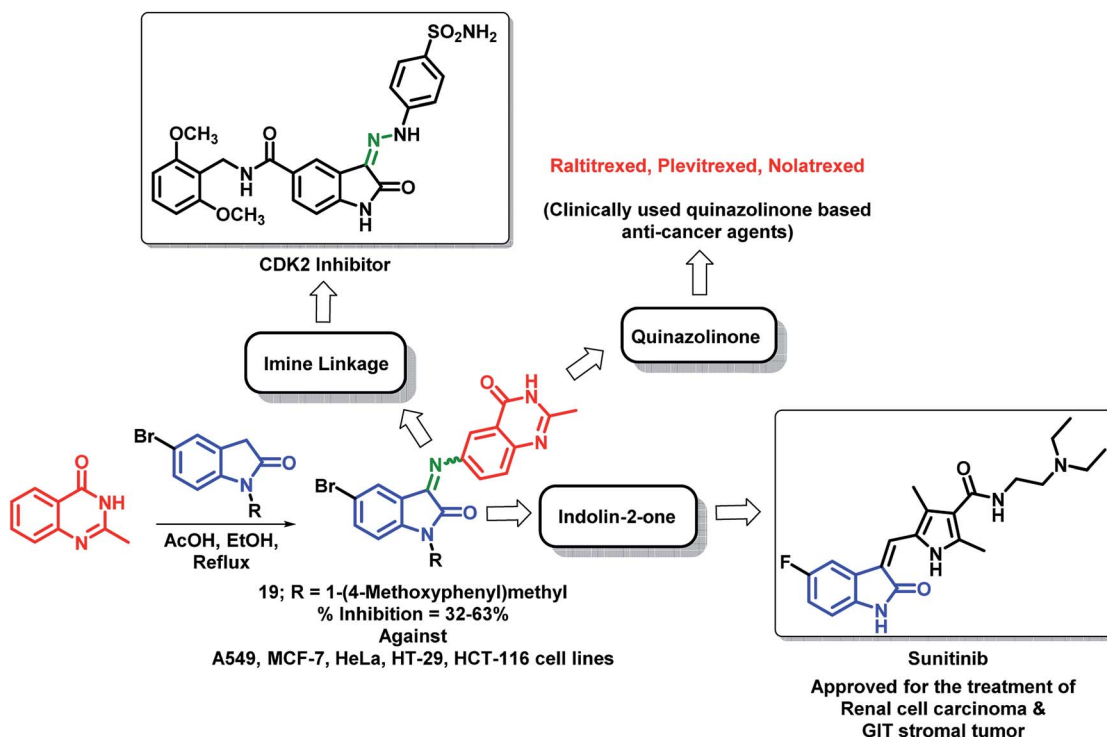


Scheme 5 Synthetic and design strategy for quinazolinone-oxadiazole hybrids.

found to be superior in DHFR inhibition with an IC₅₀ of 0.011 μM, as compared with methotrexate (IC₅₀ of 0.02 μM). From the SAR study, it was found that the phenyl ring is essential at 2nd position of quinazolinone for effective DHFR inhibition. Also, the electron-donating groups at the *para* position of the phenyl ring lead to decreased inhibitory potential. Analogues with 5-substituted benzimidazole at 3rd position of quinazolinone have shown

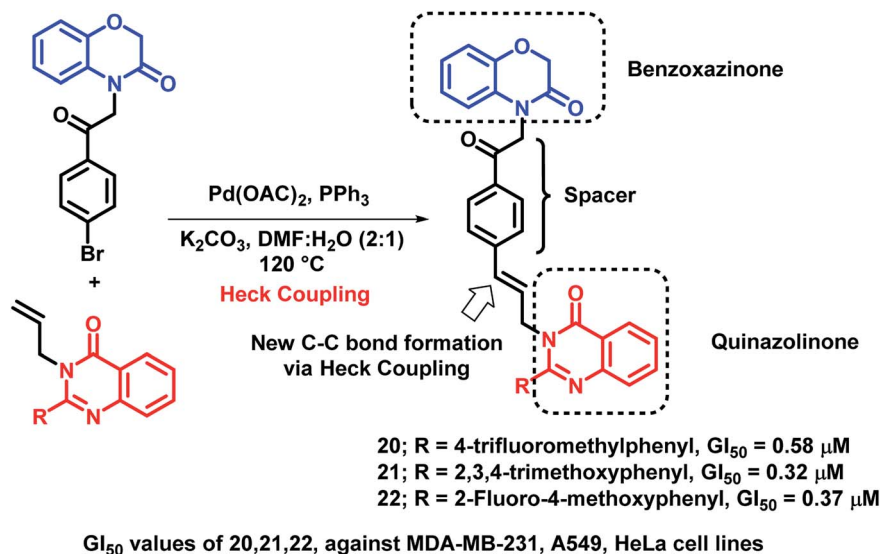
greater inhibition compared to that of 6-substituted ones (Scheme 3), which states the importance of orientation of benzimidazole on DHFR inhibition.

By considering the various structural aspects of clinically-approved thiazole-containing anti-cancer drugs (tiazofurin and bleomycin), Hosseinzadeh *et al.* designed hybrid analogues containing thiazole and quinazolinone moieties.³³ After anti-



Scheme 6 Synthetic strategy for quinazolinone and indolin-2-one hybrids, showing the importance of each structural unit.



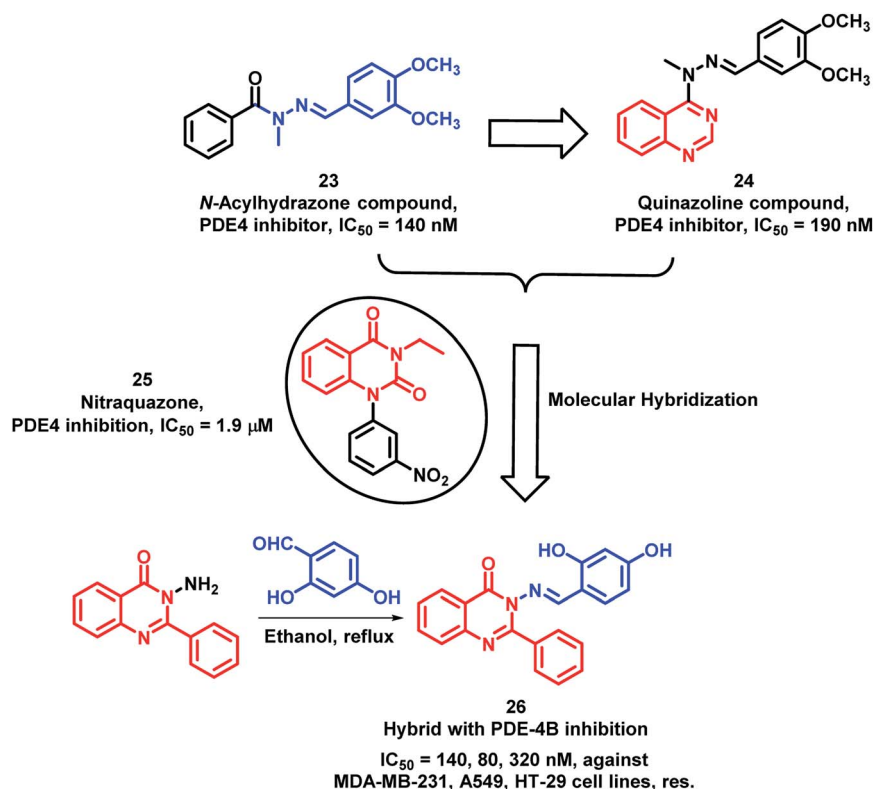


Scheme 7 Synthetic scheme for benzoxazinone and quinazolinone hybrids.

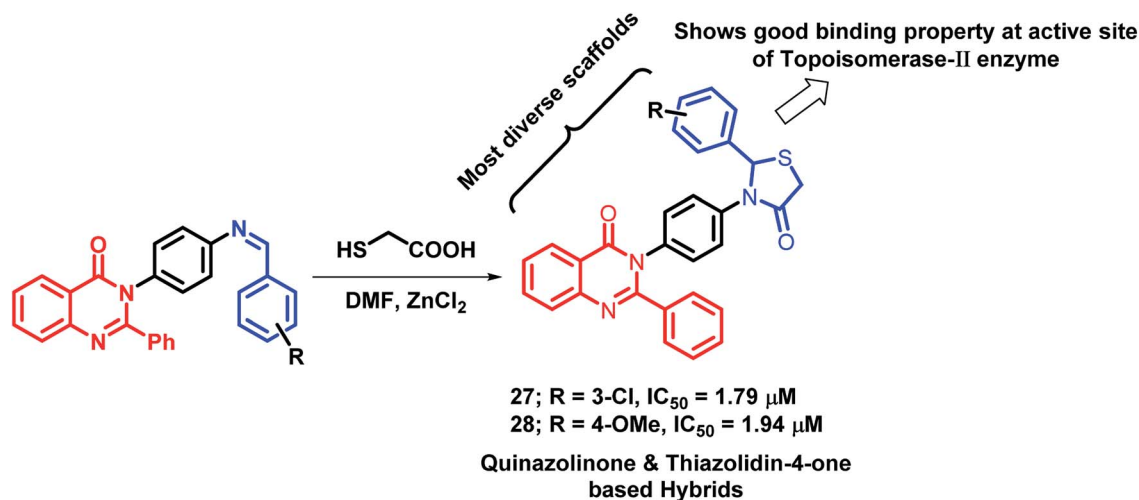
proliferative screening against 3 cancer cell lines (PC3, MCF-7, HT-29), compound 17 was found to be most active with IC_{50} values of 10, 10, and $12 \mu\text{M}$ against PC3, MCF-7, and HT-29 cells, respectively (Scheme 4).

Hassanzadeh *et al.* have designed novel hybrids using 2-substituted quinazolinones and 2-phenyloxadiazole.³⁴ As shown in Scheme 5, quinazolinone and oxadiazole derivatives show

anti-proliferative activity through various mechanisms. Amongst the synthesized hybrid analogues, compound 18 exhibited potent anti-cancer activity when tested on HeLa cell lines with an IC_{50} of $7.52 \mu\text{M}$. Substitution at the 2nd position of quinazolinone plays an important role in the anti-cancer activity as analogues with propyl substitution (compound 18) were found to be potent as compared with other analogues.



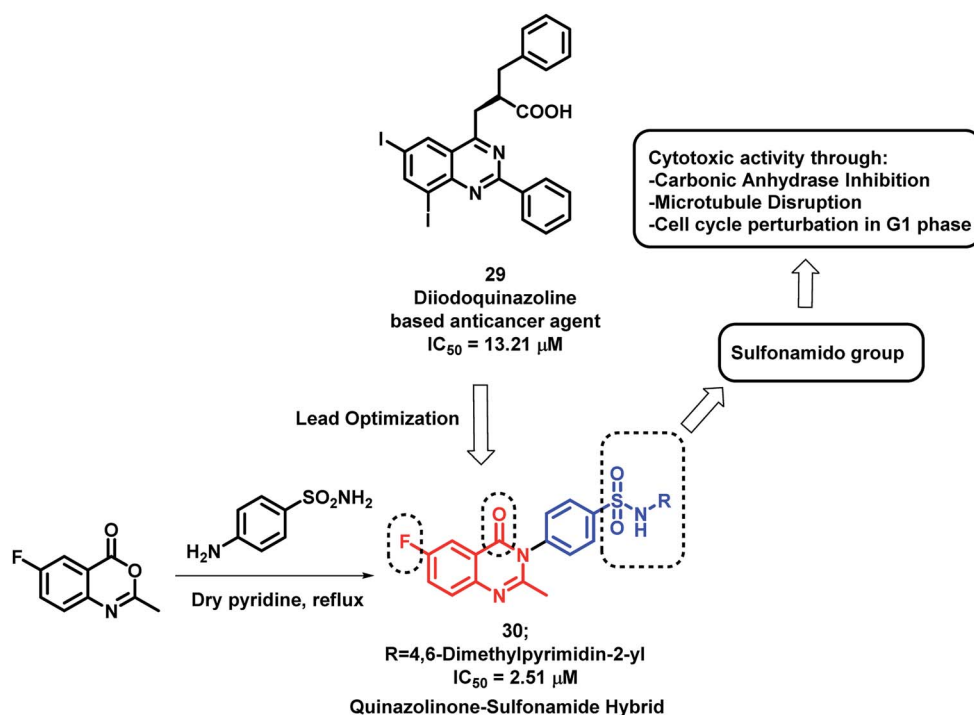
Scheme 8 Synthetic and design strategy for the Schiff base hybrids of quinazolinone as PDE-4 inhibitors.



Scheme 9 Synthetic scheme for quinazolinone- and thiazolidinone-based hybrids.

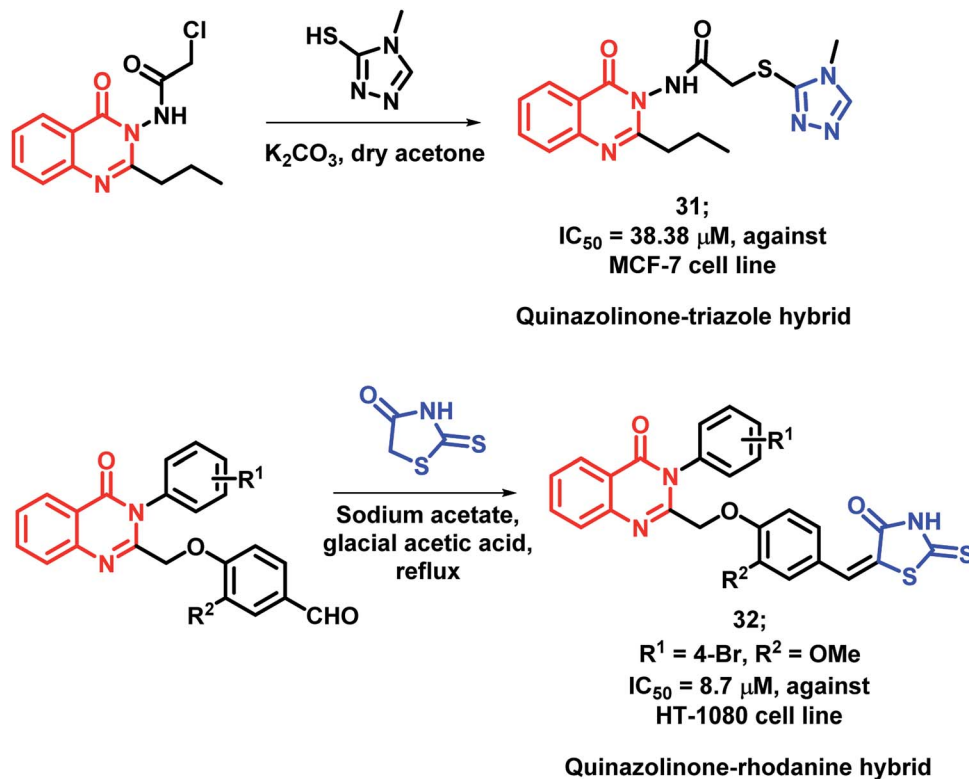
Wu *et al.* designed quinazolinone- and indolin-2-one-based hybrids by linking 2-methylquinazolin-4(3H)-one with indolin-2-one through an imine linkage.³⁵ These hybrid analogues were synthesized by the condensation of 6-amino-2-methylquinazolin-4(3H)-one with indolin-2,3-dione or *N*-substituted indolin-2,3-dione followed by *in vitro* anti-cancer screening against 5 human cancer cell lines (A549, MCF-7, HeLa, HT-29, HCT-116). It was observed that *N*-substitution on indolin-2-one is essential for the anti-proliferative activity. Amongst both the series, compound **19** was found to be most potent with % inhibition in the range of 32–63% against above-mentioned cell lines (Scheme 6).

Benzoxazinone, which is a widely active heterocycle,^{36,37} was combined with allylquinazolinone through molecular hybridization by Bollu *et al.*³⁸ Pd-catalyzed Heck coupling was utilized in order to club 1,4-benzoxazinone and quinazolinone to produce novel 1,4-benzoxazinone-acetylphenylallylquinazolin-4(3H)-one hybrids. All the hybrids showed moderate to good anti-proliferative activity against 3 cancer cell lines (A549, HeLa, MDA-MB-231), with GI_{50} in the range of 0.32–13.4 μ M. Among these, compounds **20** (GI_{50} = 0.58 μ M; MDA-MB-231 cell line), **21** (GI_{50} = 0.32 μ M; A549 cell line), and **22** (GI_{50} = 0.37 μ M; HeLa cell line) were exceptionally good in the anti-proliferative activity against the particular cell lines (Scheme 7).



Scheme 10 Synthetic and design strategy for quinazolinone-sulfonamide hybrids.





Scheme 11 Synthetic schemes for quinazolinone-based rhodanine (31) and triazole (32) hybrids.

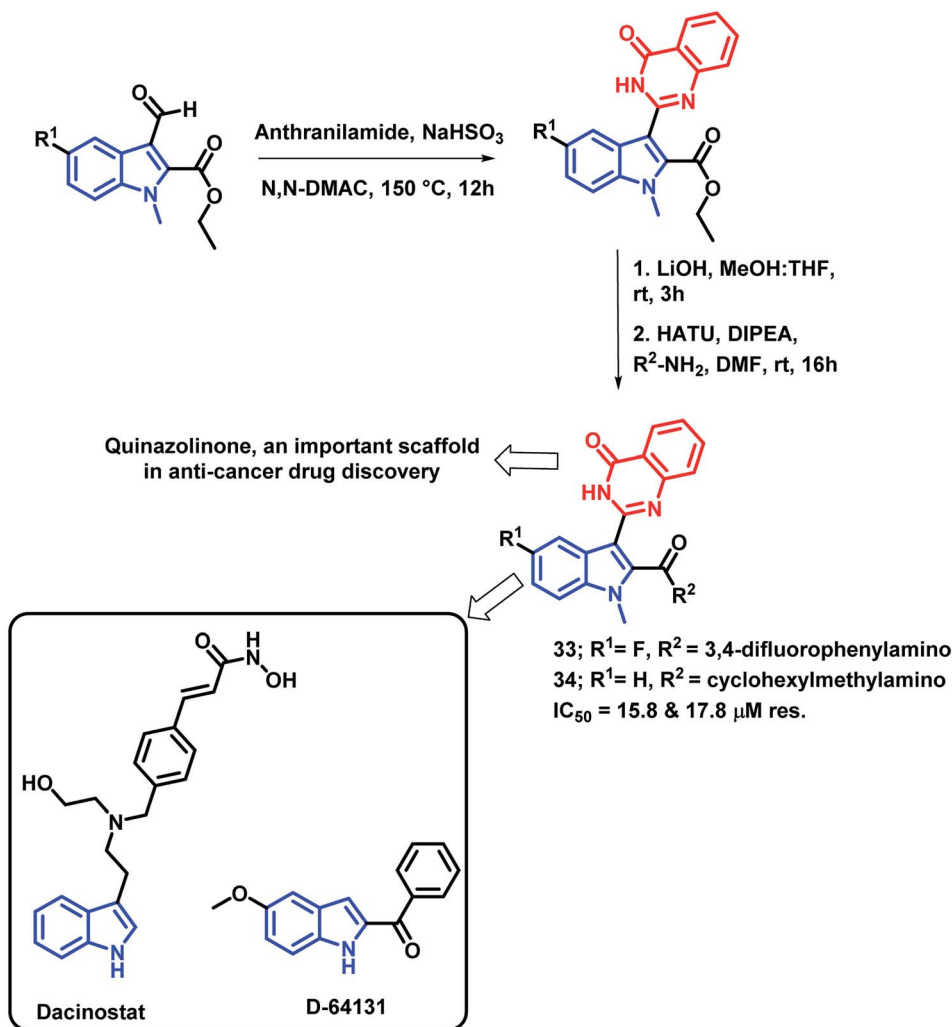
Phosphodiesterase-4 (PDE-4) is known to play a vital role in the proliferation, angiogenesis, and motility of many cancerous cells;^{39–41} hence, PDE-4 inhibition will lead to anti-proliferative activity. *N*-acylhydrazones are potent inhibitors of PDE-4 (Compound 23; Scheme 8); therefore, in previous reports, quinazoline was combined with *N*-acylhydrazone to obtain the quinazoline-hydrazone hybrid with an IC_{50} of 190 nM.^{42,43} Further to enhance the activity of such hybrids, Rahman *et al.* performed lead optimization by replacing quinazoline with quinazolin-4(3*H*)one.⁴⁴ These quinazolin-4(3*H*)one-hydrazone hybrids were designed, synthesized, and screened for PDE-4 inhibition. The condensation of 3-amino-2-phenylquinazolin-4(3*H*)one with different substituted aromatic aldehydes afforded the desired arylmethylidenehydrazides. These synthesized Schiff base hybrids were evaluated for the *in vitro* inhibition of the PDE-4(B) enzyme by taking rolipram as a positive control. Many compounds were found to be potent PDE-4(B) inhibitors. Further, these active compounds were screened for the *in vitro* anti-cancer activity against 3 human cancer cell lines (MDA-MB-231, A549, HT-29). Amongst these compounds, compound 26 has shown more anti-proliferative activity with an IC_{50} of 140, 80, and 320 nM against MDA-MB-231, A549, and HT-29 cell lines, respectively (Scheme 8).

Thiazolidin-4-one, a saturated 4-oxo form of thiazole, occupies an important place in medicinal chemistry. Due to its diverse biological activities, Thakral *et al.* combined thiazolidin-4-one with quinazolinone scaffold *via* molecular hybridization approach.⁴⁵ For the synthesis of the desired hybrids, 3-(4-

aminophenyl)-2-phenylquinazolin-4(3*H*)one was treated with aromatic aldehydes to produce Schiff bases of quinazolin-4(3*H*)one. Further, cyclocondensation of the obtained products (Schiff's base) with thioglycolic acid in DMF resulted in desired hybrid derivatives. Hybrid compounds 27 ($IC_{50} = 1.79 \mu M$) and 28 ($IC_{50} = 1.94 \mu M$) showed good anti-cancer activity against Hep-G2 and MCF-7 cell lines, respectively (Scheme 9), as compared with doxorubicin ($IC_{50} = 0.09 \mu M$ against Hep-G2 and MCF-7 cell lines), used as a standard. Moreover, the results of the molecular docking study of the designed hybrids with topoisomerase-II enzyme (PDB code: 1ZXM) have shown positive correlation of topoisomerase-II inhibition with the anti-proliferative activity.

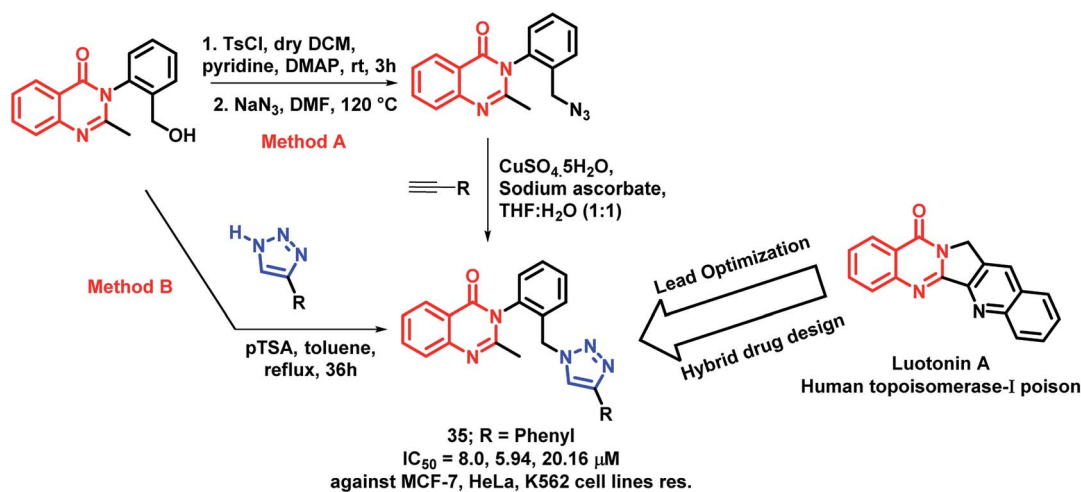
Sulphonamide derivatives have shown anti-cancer activity through various mechanisms including the disruption of microtubule assembly, carbonic anhydrase (CA) inhibition, and cell cycle perturbation.^{46–48} By considering such points, Zayed *et al.* optimized previously designed potent quinazoline-based analogues (29)⁴⁹ by attaching the benzenesulfonamido moiety on quinazolinone backbone⁵⁰ (Scheme 10). The target compounds were synthesized by the simple fusion of various *N*-substituted 4-Aminobenzene sulphonamides with benzoxazin-4-one in pyridine. Amongst the synthesized analogues, compound 30 was found to be potent as a cytotoxic agent with an IC_{50} of 2.51 μM against NCI cell line.

As triazole was found to increase the existing activity of many bioactive scaffolds,⁴ Hassanzadeh *et al.* have designed hybrids with the structural features of triazole and quinazolinone.⁵¹



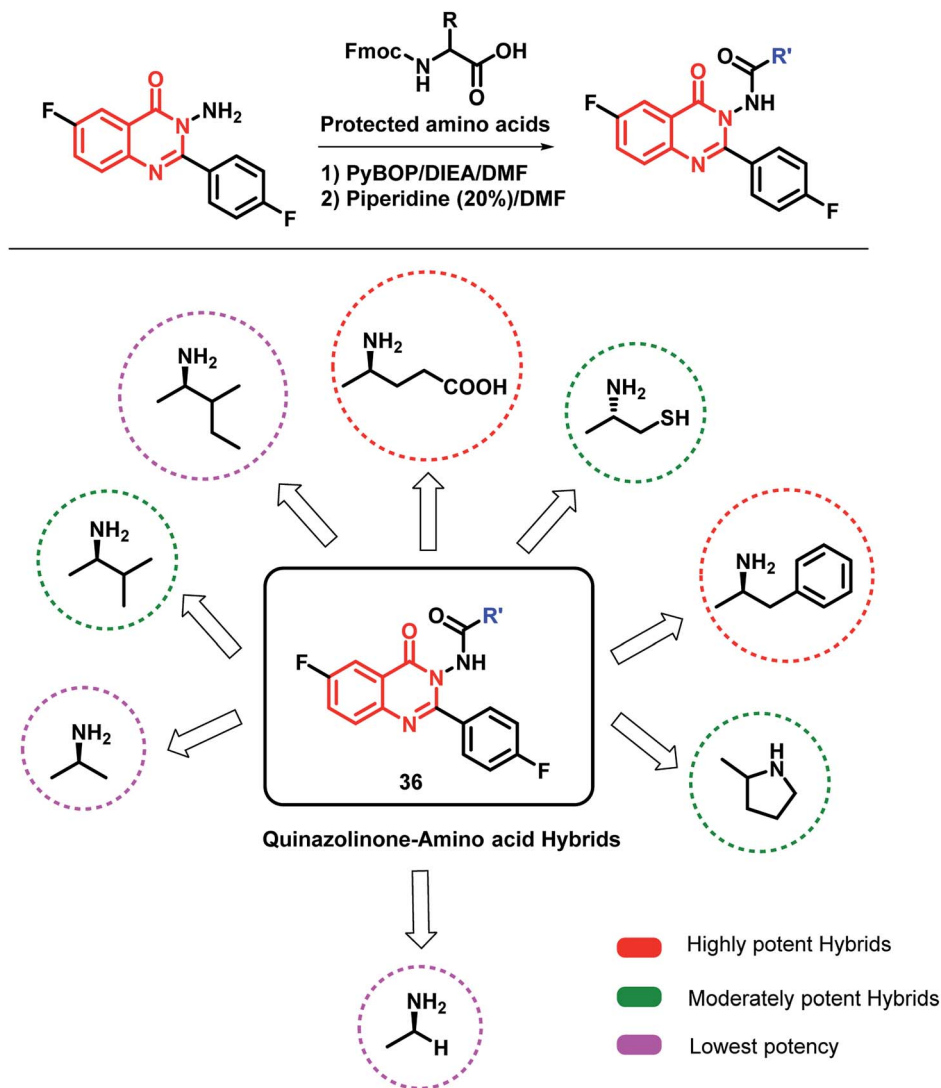
2&3 Substituted Indole analogues as anti-cancer agents

Scheme 12 Synthetic and design strategy for indole- and quinazolinone-based hybrids for anti-cancer potential.



Scheme 13 Synthetic scheme for luotonin A-inspired quinazolinone-triazole hybrids.



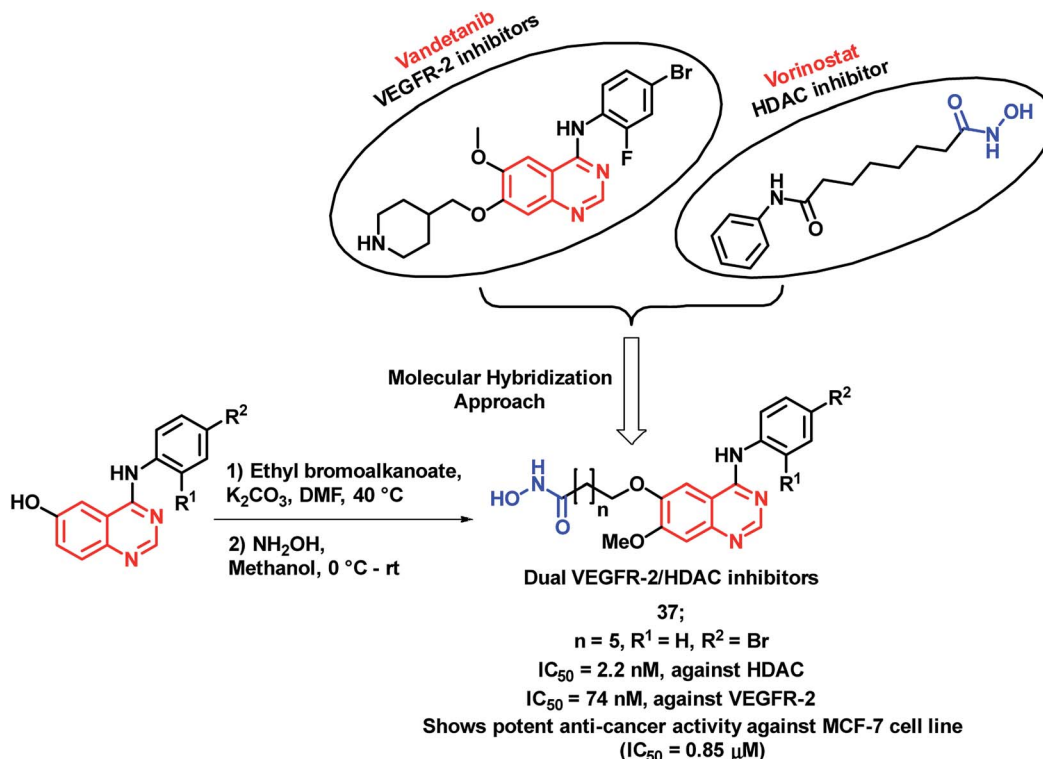


Scheme 14 Synthetic scheme for amino acid-linked quinazolinone hybrids.

These triazole and quinazolinone scaffolds were linked through a 2-thioacetamido linkage and synthesized *via* the substitution reaction of 2-chloro-*N*-(4-oxo-2-quinazolin-3(1*H*)-yl)acetamide derivatives with 4-methyl-4-*H*-1,2,4-triazole-3-thiol. These synthesized hybrids were tested against MCF-7 and HeLa cell lines and MCF-7 cell line was found to be susceptible to compound 31 with an IC_{50} of 38.38 μ M (Scheme 11).

Rhodanine derivatives were found to induce apoptosis through the modulation of the Bcl-2 family of proteins. Hence, Sayed *et al.* linked rhodanines with quinazolinones in order to design rhodanine-based quinazolinone hybrids.⁵² All the synthesized compounds were found to be active with an IC_{50} value in the range of 10–60 μ M, when screened against the HT-1080 cell line. The SAR study revealed that bulky, hydrophobic, and electron-withdrawing substituents at the *para* position of the phenyl ring linked at the 3rd position of quinazolinone moiety is essential for anti-proliferative activity. Compound 32 (IC_{50} = 38.38 μ M) was the most potent analogue, having desired substituents (Scheme 11).

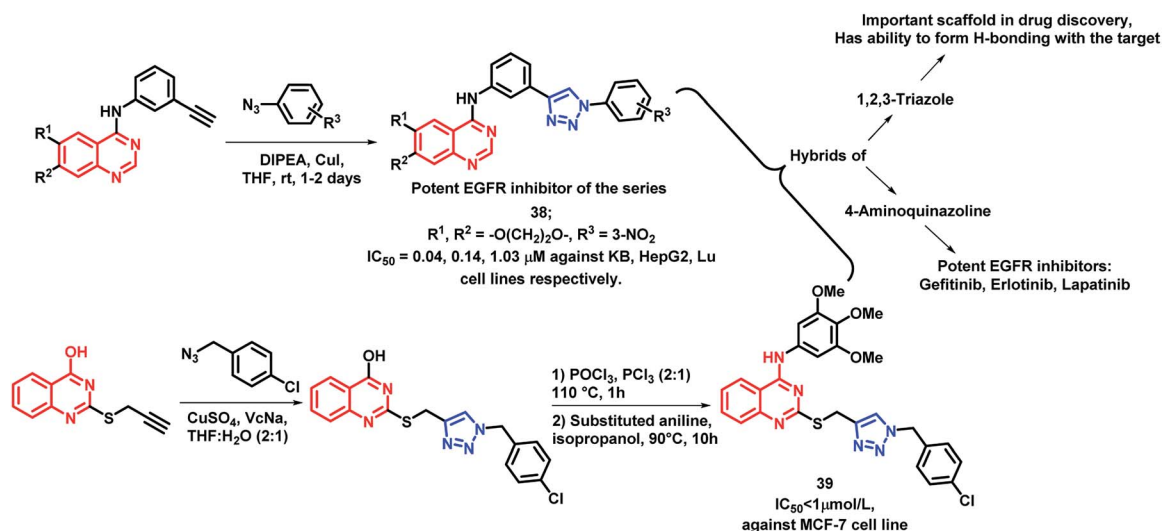
Gokhale *et al.* have designed indole and quinazolinone-based amides for anti-cancer activity⁵³ by considering the apoptosis inducing property of various indole-based amides.⁵⁴ Also, the introduction of amide at the 2nd position of indole leads to cytotoxicity through histone deacetylase (HDAC) inhibition. Based on such observations, quinazolinone-based indolyl amides were designed (Scheme 12) and synthesized, followed by the screening of their anti-cancer activity. It was observed that the substituent at the phenyl ring attached to the amide nitrogen is essential for the anti-proliferative activity. Various substituents such as cyclopropyl, cyclohexylmethyl, fluoro, electron releasing groups were found to be active against the cancer cell lines (HepG2 and MCF-7). Compounds 33 and 34 had potential cytotoxicity (IC_{50} of 15.8 and 17.8 μ M, respectively) against the HepG2 cell line as compared with doxorubicin and 5-fluorouracil as positive controls (IC_{50} of 6.5 and 18.8 μ M, respectively). All the synthesized compounds were less toxic against the non-cancerous Vero cell line, which confirms the selectivity of the compounds towards the cancerous cells.



Scheme 15 Synthetic and design strategy for quinazoline-hydroxamic acid hybrids.

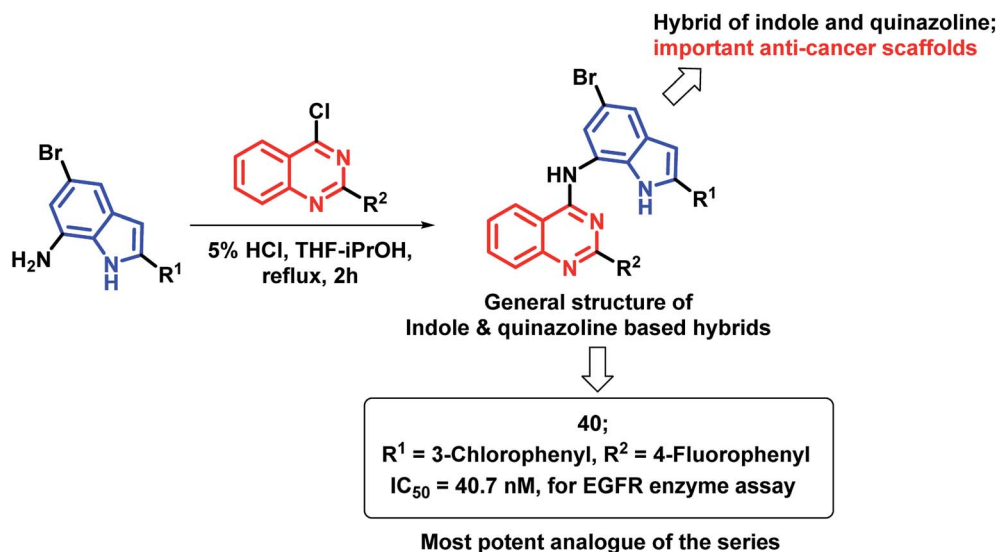
Luotonin A (pyrroloquinazolinoquinoline alkaloid) possesses anti-cancer activity through the inhibition of human topoisomerase-I. By taking such facts into consideration, Venkatesh *et al.* attempted to develop luotonin A- and triazole-based hybrid analogues by targeting HDAC and enhancer of zeste homolog 2 (EZH2) proteins.⁵⁵ The designed quinazolinone hybrid analogues were synthesized by two methods from 3-(2-(hydroxymethyl)phenyl)-2-methylquinazolin-4(3H)-ones. In the first method, click chemistry approach was applied by using

various acetylenes. The second method was applied by the direct attachment of substituted triazoles in the presence of *para*-toluenesulfonic acid (*pTSA*). Amongst all, compound 35 was found to be active with IC_{50} values of 8.0, 5.94, 20.16, and 256 μM against the MCF-7, HeLa, K562, and HEK cell lines, respectively (Scheme 13). Further, the effect of compound 35 on the cell cycle was identified by flow cytometry, showing G1 phase arrest on the HeLa cell line.



Scheme 16 Synthetic scheme for 4-aminoquinazoline and triazole hybrids.





Scheme 17 Synthetic scheme for indole and 4-aminoquinazoline hybrids.

Zayed *et al.* modified the quinazolinone scaffold by attaching desired fragments that may be helpful in EGFR binding.⁵⁶ In this way, various amino acid-linked quinazolinone hybrids were synthesized by using various amino acids (Scheme 14). The amino acids were supposed to provide additional H-bonding interactions at the target site. Amongst the synthesized hybrids, phenylalanine- and glutamine-based hybrid analogues were potent against the MCF-7 cell line (IC₅₀ of 0.44 and 0.43 μ M, respectively) compared to that of the reference drug erlotinib (IC₅₀ of 1.14 μ M). The docking study was also performed by taking the EGFR enzyme (PDB: 1M17) and tubulin (PDB: 1SA0), followed by EGFR inhibition and tubulin polymerization inhibition assays. The results of both the docking and enzyme inhibition studies were correlated with the results of the cell-based cytotoxicity study. Hence, the authors concluded that the anti-cancer activity of these hybrid analogues was possibly due to EGFR and tubulin polymerization inhibition.

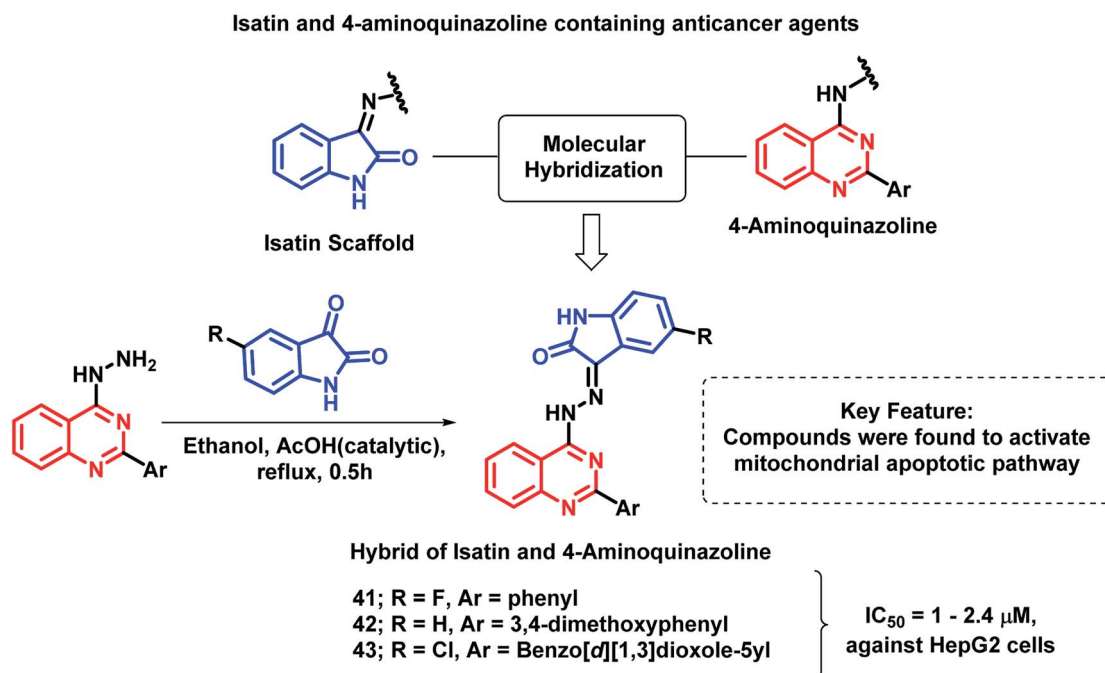
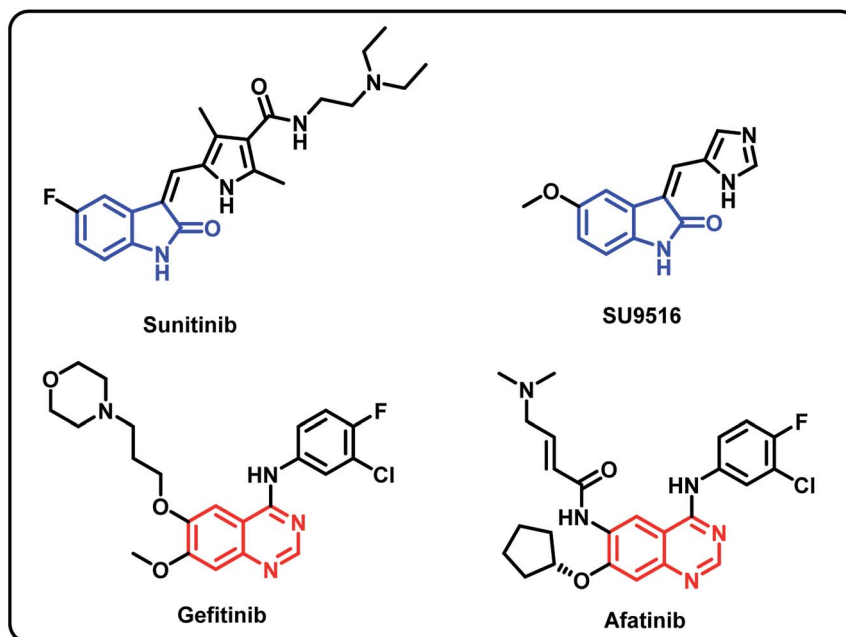
The inhibition of tumor angiogenesis is one of the targets for cancer treatment and the VEGFR-2 receptor is one of the regulators of tumoral angiogenesis.^{57,58} Vandetanib is one of the drugs used as an anti-cancer agent, which acts through VEGFR-2 inhibition. Peng *et al.* designed molecular hybrids having desired features for VEGFR-2 and HDAC inhibition.⁵⁹ The hybrid analogues were designed by considering the structural features of vandetanib (VEGFR-2 inhibitor) and vorinostat (HDAC inhibitor). As shown in Scheme 15, 4-aminoquinazoline (pharmacophore of vandetanib) was linked with hydroxamic acid (for HDAC inhibition) *via* a long alkyl chain. Those designed analogues have shown proper binding at the active site of VEGFR-2 kinase (PDB: 2QU5) and Histone Deacetylase-Like Protein (HDLP) (PDB: 1C3S). These synthesized compounds were tested for HDAC and VEGFR-2 inhibition, followed by anti-cancer screening against the MCF-7 cell line. Compound 37 was found to have potent inhibition of both VEGFR-2 and HDAC, with IC₅₀ values of 74 and 2.2 nM,

respectively. Also, it was found to be cytotoxic against the MCF-7 cell line with an IC₅₀ of 0.85 μ M.

1,2,3-Triazole is one of the important fragments in medicinal chemistry and can be used for the synthesis of a variety of bioactive heterocycles.^{60–64} Also, various triazole-containing hybrids were synthesized recently with optimum activities. By considering such factors, Le-Nhat-Thuy *et al.* designed triazole linked 4-aminoquinazoline-based hybrids (Scheme 16), followed by synthesis and *in vitro* anti-cancer screening.⁶⁵ Two structural requirements were found to be essential for cytotoxic activity: (a) oxygenated substituents on quinazoline, as derivatives with fused deoxygenated ring were effective as anti-cancer agents, (b) substituents on the phenyl ring, attached to the triazole moiety. With such structural features, compound 38 was found to be most potent with IC₅₀ of 0.04, 0.14, and 1.03 μ M against KB, HepG2, and Lu cell lines, respectively.

Quinazoline and triazole were linked through thioether linkage *via* the click chemistry approach by Song *et al.* and the synthesized analogues were screened for *in vitro* anti-cancer activity.⁶⁶ Compound 39 was found to be highly active as compared with other analogues, having an IC₅₀ value less than 1 μ mol L⁻¹ (Scheme 16).

The indole nucleus was found to be prevalent in many bioactive heterocycles, with anti-microbial, anti-oxidant, and anti-cancer activities.^{67–70} Due to such diverse properties of the indole nucleus, Mphahlele *et al.* designed hybrids of indole and 4-aminoquinazoline (Scheme 17) for increased anti-cancer potential.⁷¹ Those hybrids were found to bind at the ATP binding region of EGFR (PDB code: 1M17), with similar interactions as that of erlotinib. Compound 40 has shown potential EGFR inhibition with an IC₅₀ of 40.7 nM, as compared with the standard drug gefitinib (IC₅₀ = 38.9 nM). Many compounds of the series were found to be active in the *in vitro* cytotoxicity screening against various cancer cell lines (A549, Caco-2, C3A, MCF7, and HeLa).

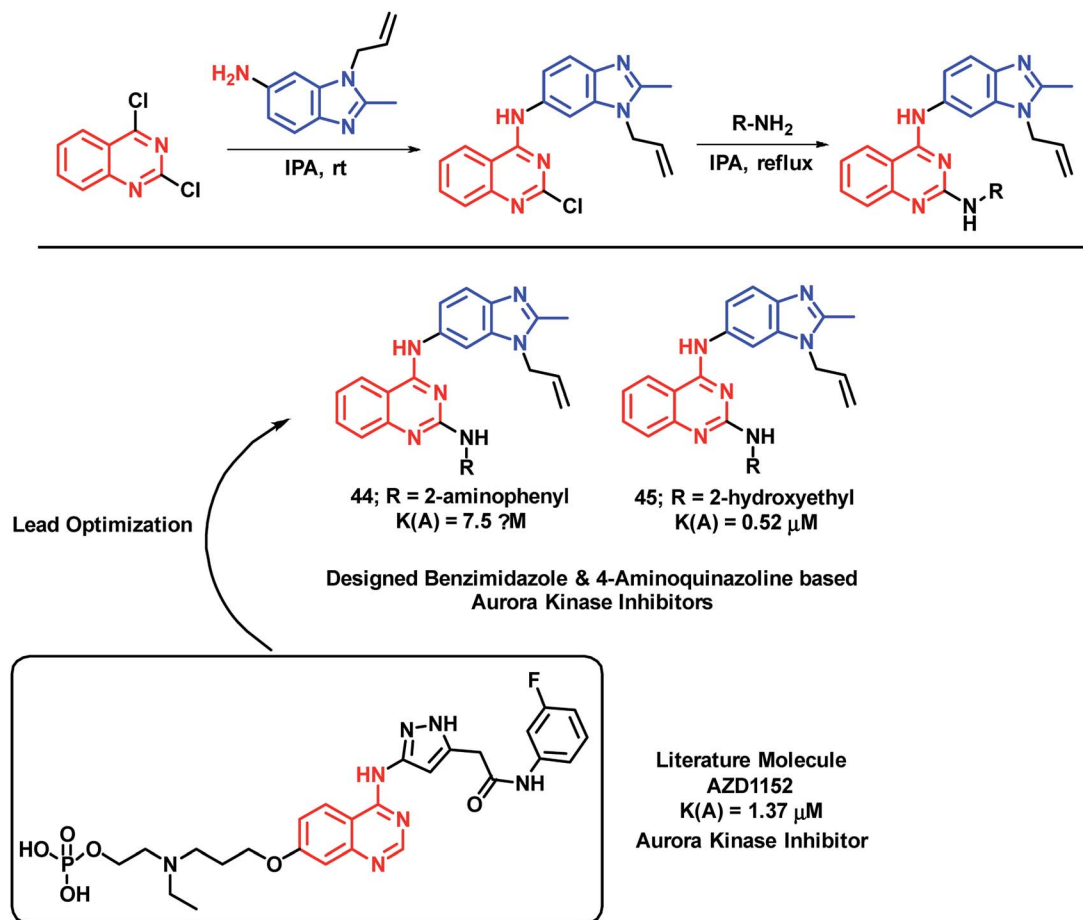


Scheme 18 Synthetic and design strategy for isatin and 4-aminoquinazoline hybrids.

Another indole derivative, *i.e.*, 1*H*-indole-2,3-dione (isatin), was linked with 4-aminoquinazoline scaffold *via* the molecular hybridization approach (Scheme 18) by Fares *et al.*⁷² Imine linkage was used as it is present in many of the isatin-containing EGFR inhibitors such as sunitinib. Compounds **41**, **42**, and **43** were the most potent anti-cancer agents in the series, with IC₅₀ values in the range of 1–2.4 μM against the HepG2 cell line. These active compounds were found to induce the activation of caspase-3 along with a reduction in the Bcl-2/Bax ratio in the HepG2 cells. These results revealed that the anti-cancer activity of these hybrid analogues is due to the activation of the mitochondrial apoptotic pathway.

Aurora kinase is one of the enzymes responsible for the proliferation of cancer cells. Hence, by targeting such enzymes, cancerous cells can be inhibited from proliferation. Considering the biological importance of the benzimidazole heterocycle, Luxami *et al.* have designed hybrid analogues having the structural features of benzimidazole and 4-aminoquinazoline (important scaffold with anti-cancer potential).⁷³ Amongst the synthesized analogues, compounds **44** and **45** have shown aurora kinase inhibition with IC₅₀ values of 7.50 and 0.53 μM, respectively (Scheme 19). Along with aurora kinase inhibition, these hybrids were also found to be useful as ratiometric chemosensors for the estimation of lead and cyanide concentration

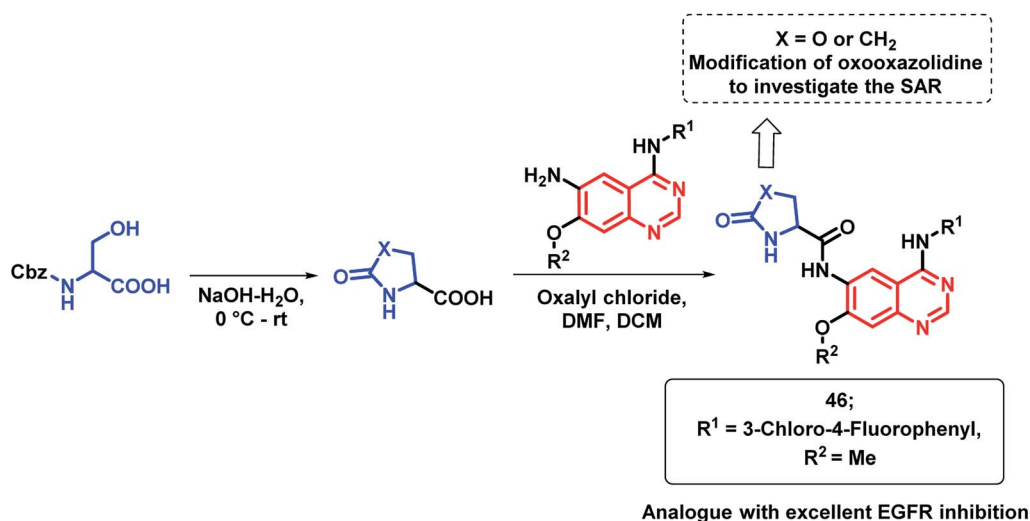




Scheme 19 Synthetic and design strategy for benzimidazole and 4-aminoquinazoline hybrids.

in the given sample. Heavy metals are found to be toxic to the environment as well as the human health; hence, chemosensors will be effective for the detection of such ions. Pb^{2+} and CN^- binding property was investigated *via* UV-Vis spectroscopy,

fluorescence spectroscopy, ^1H NMR titration experiment, and DFT calculations. Compound **44** was used to estimate CN^- (2–500 μM) and compound **45** was used to estimate the Pb^{2+} ion (0.05–1500 μM) ratiometrically.



Scheme 20 Synthesis of oxazolidinone and 4-aminoquinazolinone hybrids.

EGFR inhibitors containing 4-aminoquinazoline such as gefitinib and erlotinib are beneficial for anti-cancer activity but they lack clinical efficiency due to the point mutation of the EGFR enzyme. To address such problems, Shao *et al.* prepared hybrid analogues consisting of oxoxazolidine and 4-aminoquinazoline.⁷⁴ As shown in Scheme 20, these hybrids were designed as per the structural requirements for EGFR inhibition. Many of the synthesized hybrids exhibited potent EGFR kinase inhibitory activity against the mutant forms of EGFR (EGFR^{T790M} and EGFR^{L858R}) but amongst these hybrid analogues, compound **46** was highly potent. Through Western blotting analysis, it was found that compound **46** reversibly inhibited EGF-mediated phosphorylation of EGFR in A431 cells.

Sharma *et al.* synthesized novel regioisomeric quinazoline/benzimidazole hybrids and evaluated the *in vitro* anti-tumor effects in 60 cell lines.⁷⁵ The hybrid analogues were designed for increased lipophilicity (*via* the substitution of alkyl/allyl groups in benzimidazole and aryl groups in the quinazoline moiety) and for electron donor and acceptor effects (by substitution of secondary amines). The thrust effort is focused to evaluate the effects of substitution at the 2nd position of quinazoline for potential anti-tumor effect with desired selectivity. The anti-tumor activity of the synthesized compounds was evaluated against nine tumor cell subpanels such as leukemia, non-small cell lung, colon, CNS, melanoma, ovarian, renal,

prostate, and breast cancer cells. As shown in Fig. 4, compound **47** exhibited selectivity towards leukaemia cancer cells, colon cancer cells, melanoma cancer cells, and breast cancer cell lines (GI value 45.0–98%, 76.6–94.3%, 97.5%, and 58%, respectively).

Banerji *et al.* synthesized a series of triazole-substituted quinazoline hybrid analogues and evaluated their anti-cancer effect against HCT116 (human colorectal cancer cell line), HEPG2 (human liver cancer cell line), MCF-7 (human breast cancer cell line), and PC-3 (human prostate cancer cell line).⁷⁶ The preliminary SAR of erlotinib and lapatinib (EGFR inhibitors) revealed that the presence of 5-aminoquinazoline moiety was helpful for EGFR inhibition. The triazole scaffold has also been previously reported due to its anti-cancer activity *via* various mechanisms. Hence, an attempt was made for a triazole-substituted quinazoline hybrid analogues as anti-cancer agents (Scheme 21). The designed molecules were expected to exhibit anti-cancer activity by inhibiting EGFR, induction of oxidative stress, and thereby, excessive ROS generation by mitochondrial dysfunction. In anti-cancer evaluation, erlotinib was used as the positive control, which exhibited an activity of 11.57 to 26.87 μM . From the results, it can be concluded that MCF-7 is more susceptible to the synthesized compounds than the remaining cell lines. Compound **53** exhibited the highest cytotoxicity (20.71 μM). Further, the activity of the compounds highlighted that the substitution of -Cl at the 2nd position and

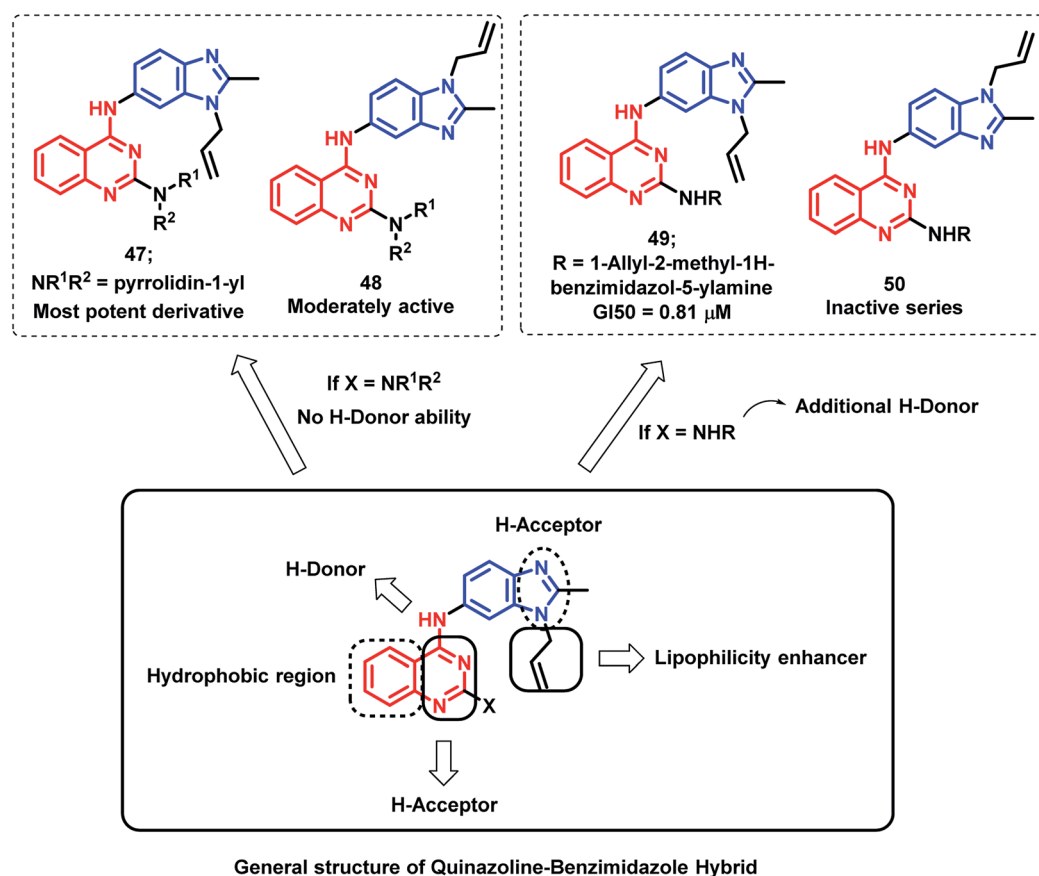
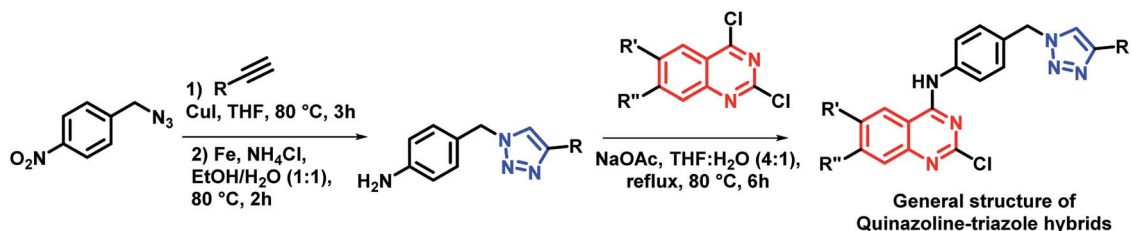


Fig. 4 Designing strategy for benzimidazole- and 4-aminoquinazoline-based hybrids.

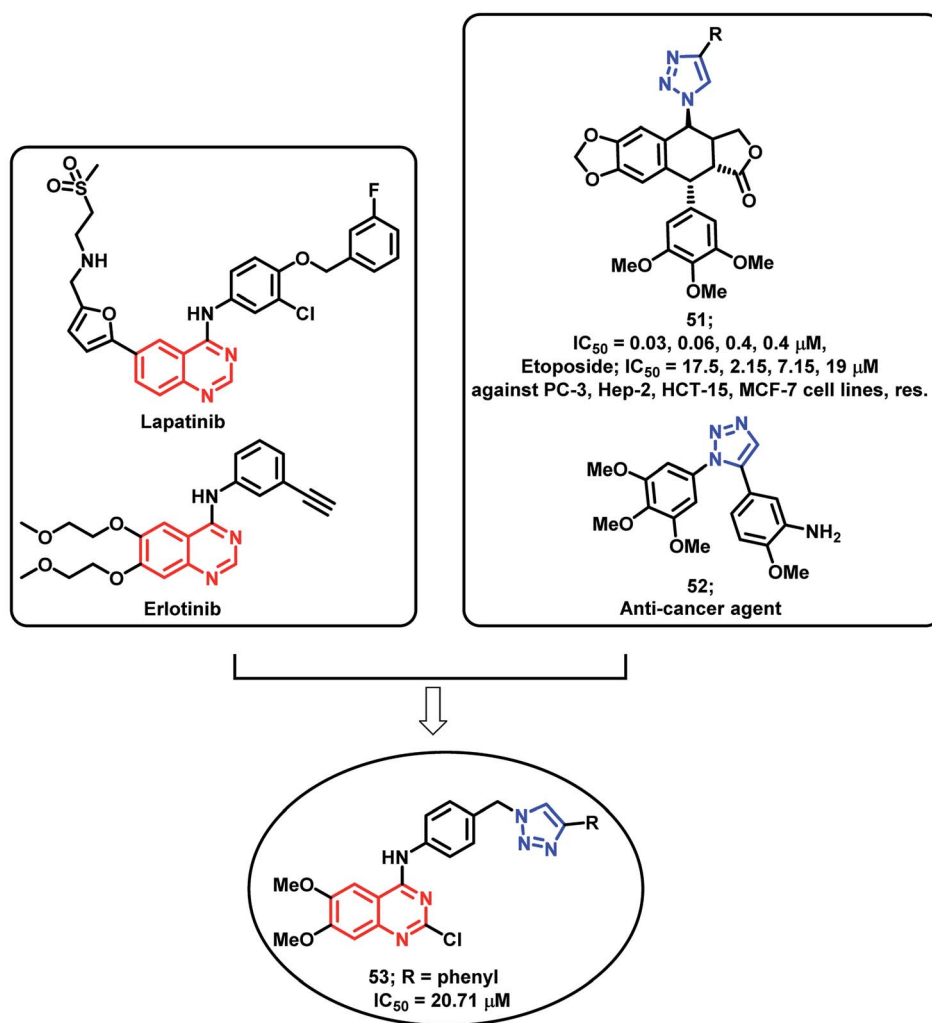


the $-\text{OCH}_3$ groups at the 6 and 7 positions of quinazoline are essential for the cytotoxicity activity. The effects of compound **53** on the EGFR pathway in the MCF-7 cell lines were identified by using Western blot analysis. The levels of EGFR and *p*-EGFR were decreased by the treatment of various concentrations of compound **53**. Hence, it is concluded that the cytotoxicity effects are mainly attributed to the decrease in EGFR and phosphorylation of EGFR and its downstream process. The effects of ROS-mediated pathway on apoptosis are confirmed by using flow cytometric analysis. The study highlighted that the

mean intensity of fluorescein isothiocyanate (FITC) was lesser in the control cells than the compound **53**-treated cells. These results also suggested the ROS-mediated pathways in the apoptosis of the MCF-7 cell lines. Fluorescence-activated cell sorting (FACS) was performed on the MCF-7 cell lines treated with compound **5b** at different time intervals for identifying the apoptosis effects of compound **53**. At the initial stage of apoptosis, phosphatidyl serine was exposed from inside the cell membrane to the outside, which can bind with annexin V. It was observed that early apoptosis rates increased from 5.9 to 24.6%

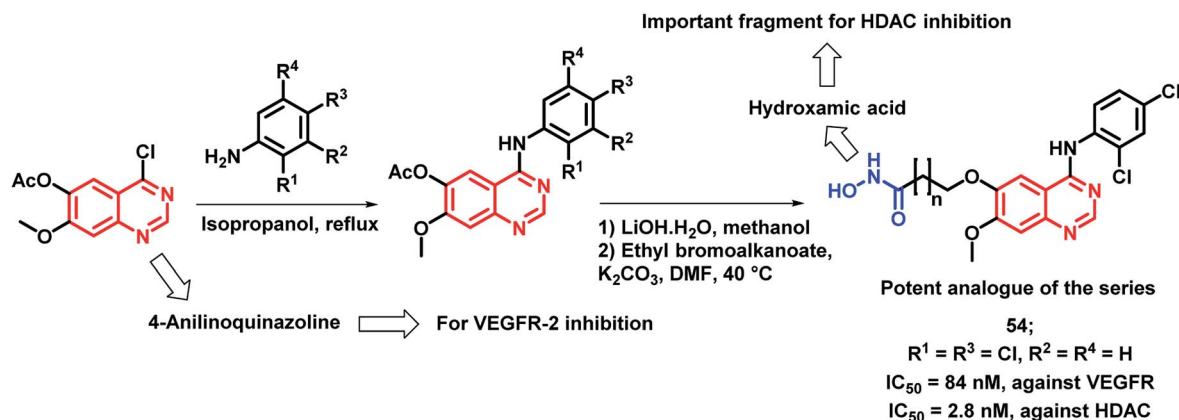


General synthetic scheme for synthesis of Quinazoline-triazole hybrids



Designing strategy for Quinazoline-triazole hybrids

Scheme 21 Synthetic and design strategy for triazole-linked 4-aminoquinazoline hybrids.

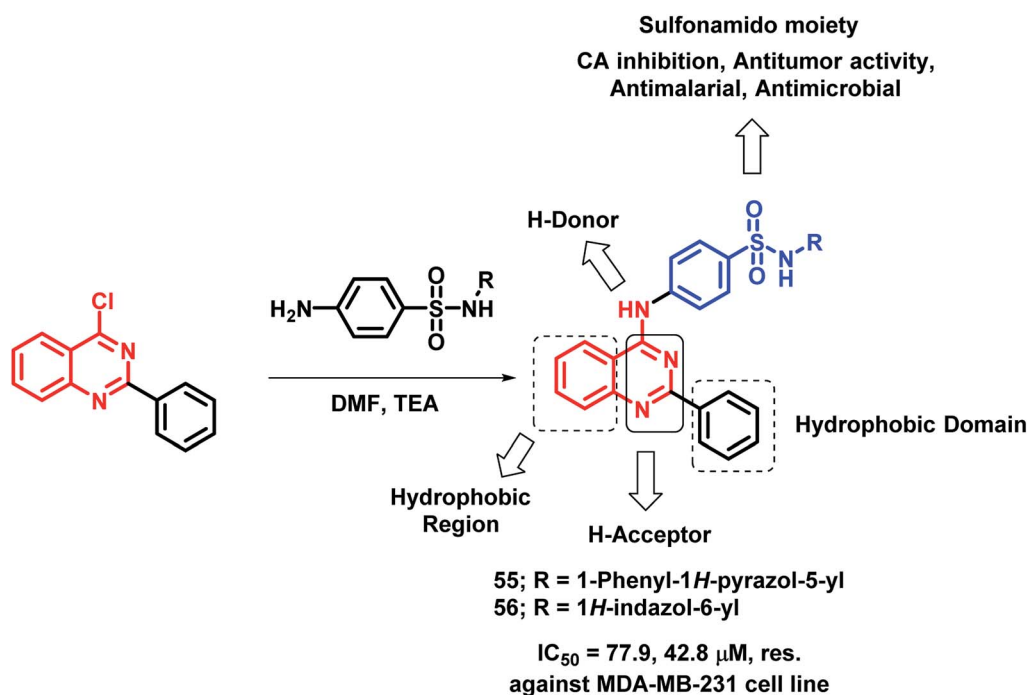


Scheme 22 Synthetic scheme for hydroxamic acid and 4-aminoquinazoline-based hybrids for HDAC and VEGFR inhibition.

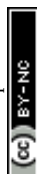
and the late apoptosis rates increased from 0.7 to 14.2%. Time-dependent cellular apoptosis was shown by compound 53. These results indicate that compound 53 might induce apoptosis by the generation of ROS *via* the mitochondrial pathway as it leads to the lowering of the mitochondrial membrane potential.

Peng *et al.* synthesized a series of 4-anilinoquinazoline and hydroxamic acid hybrids as dual inhibitors for vascular endothelial growth factor receptor-2 (VEGFR-2) and histone deacetylase (HDAC).⁷⁷ HDAC inhibitors are claimed to possess minimal toxicities toward the normal cells; hence, their combinations with other anti-cancer agents could be proved to be a successful application. As shown in Scheme 22, the synthesized analogues exhibited moderate VEGFR-2 inhibitory

activities, wherein compound 54 exhibited a comparable activity (84 nM) to that of the positive drug, VEGFR-2 TKI vandetanib ($\text{IC}_{50} = 62 \text{ nM}$). Further, an increased activity was observed with the increment in the length of the hydroxamic acid side chain. The most active VEGFR-2 in the series, *i.e.*, compound 54 exhibited maximum HDAC inhibition ($\text{IC}_{50} = 2.8 \text{ nM}$) with 5 times higher potency than the reference compound vorinostat ($\text{IC}_{50} = 12 \text{ nM}$). The isoform selectivity was evaluated by conducting the HDAC inhibition assay in HDAC1, HDAC2, HDAC6, and HDAC8 ($\text{IC}_{50} = 3.5 \text{ nM}$, 6.8 nM , 25.5 nM , and 5.2 nM , respectively). Further, the anti-proliferative activity against four types of human cancer cell lines, including a human breast cancer cell line (MCF-7), a human liver cancer cell line (HepG2), a human lung cancer cell line (A549), and



Scheme 23 Synthetic scheme for sulfonamide linked 4-aminoquinazoline hybrids.



a human colon cancer cell line (HCT-116) revealed a potent effect (IC_{50} ranges in 1.2 to 5.4 μM).

Sulfonamide-containing drugs were found to be biologically active, with various activities such as carbonic anhydrase (CA) inhibition, anti-malarial, and anti-microbial activities.^{78–80} Hence, Ghorab *et al.* designed sulfonamido-linked 4-amino-quinazoline hybrids as anti-cancer agents.⁸¹ The screening of the synthesized analogues was done against human cancer cell lines (A549, HeLa, LoVo, MDA-MB-231). Most of the compounds were found to be inactive against the HeLa cell line. Compounds **55** and **56** were found to be cytotoxic against the above-mentioned cell lines (Scheme 23).

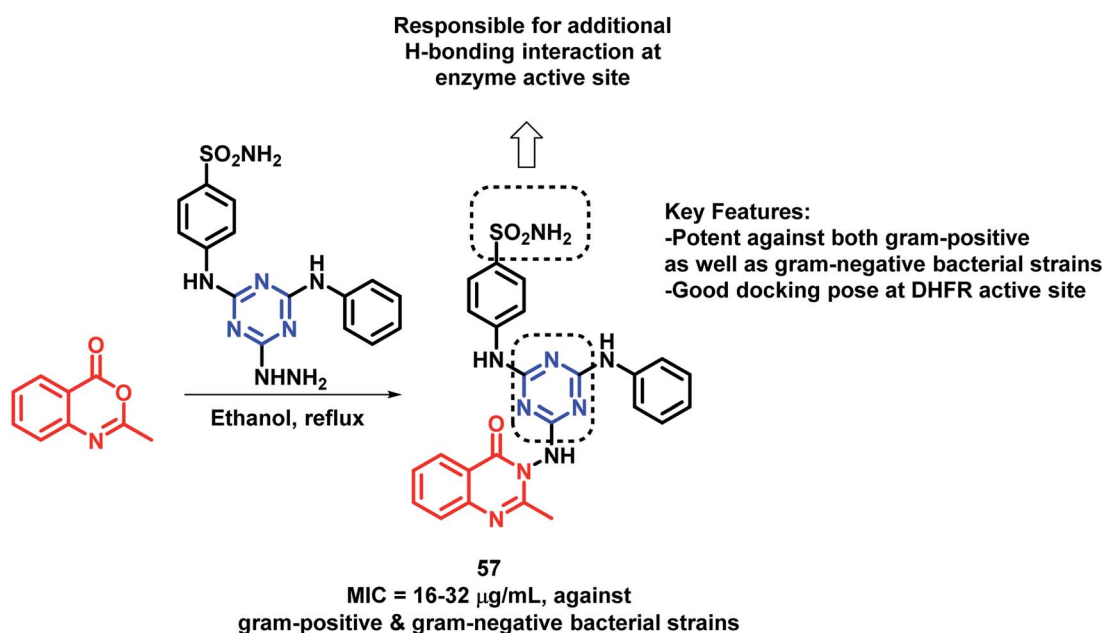
3. Anti-microbial hybrids

By considering the anti-microbial potential of both triazine and quinazolinone moieties, Dinari *et al.* designed a novel chemical framework by combining both the heterocycles to get potential anti-microbial agents.⁸² The synthesized hybrid analogues were screened for anti-bacterial and anti-fungal potential by using microplate alamar blue assay (MABA) against Gram-negative (*Salmonella enterica*, *Pseudomonas aeruginosa*, *Escherichia coli*), Gram-positive bacteria (*Bacillus subtilis*, *Staphylococcus aureus*, *Listeria monocitogenes*), and *Candida albicans*, a yeast-like fungi, by taking ciprofloxacin and ketoconazole as the positive controls. Compound **57** was found to have strong anti-bacterial property against four bacterial strains with low minimum inhibitory concentration (MICs) in the range of 16–32 $\mu\text{g mL}^{-1}$ (Scheme 24). The docking study of all the synthesized hybrids shows better binding in the pocket of *S. aureus* dihydrofolate reductase (DHFR) enzyme (PDB; 2W9S). The docking study revealed that the most potent analogue **57** showed the best *in vitro* anti-bacterial activity due to the presence of the

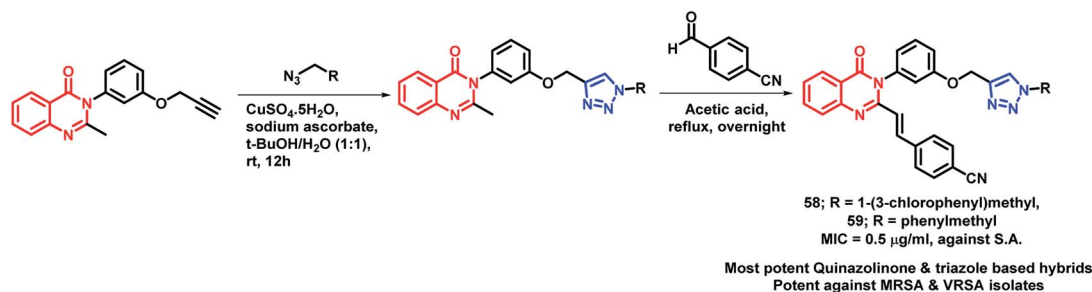
sulfonamido group, which provides additional H-bonding interactions with Ser 49, Ileu 50, Gln 19, and Phe 92 at the binding site.

Several triazole containing molecules are reported to have many biological activities; also, β -lactam antibiotic cefatrizine and the β -lactamase inhibitor tazobactam contain 1,2,3-triazole as a structural feature.⁸³ Based on such observations, Gatadi *et al.* synthesized hybrids by a combination of structural features of triazole and quinazolinone moieties.⁸⁴ Triazole was attached to the quinazolinone scaffold *via* the click chemistry approach. After the anti-bacterial evaluation of the synthesized compounds against the ESKAP panel of bacteria, many of the synthesized compounds were found to be active as anti-bacterial agents against *Staphylococcus aureus*. Compounds **58** and **59** were highly potent as anti-bacterial agents against *S. aureus*, with an of MIC 0.5 $\mu\text{g mL}^{-1}$ (Scheme 25). These compounds were inactive against *E. coli* (Gram-negative bacteria) and were also found to be non-toxic against the Vero cell line. Compounds **58** and **59** also displayed inhibitory activity against 8 MRSA strains (IC_{50} in the range of 0.5–2 $\mu\text{g mL}^{-1}$) & 3 VRSA strains (IC_{50} in the range of 0.5–32 $\mu\text{g mL}^{-1}$), indicating the ability of these compounds to overcome the drug resistance mechanism.

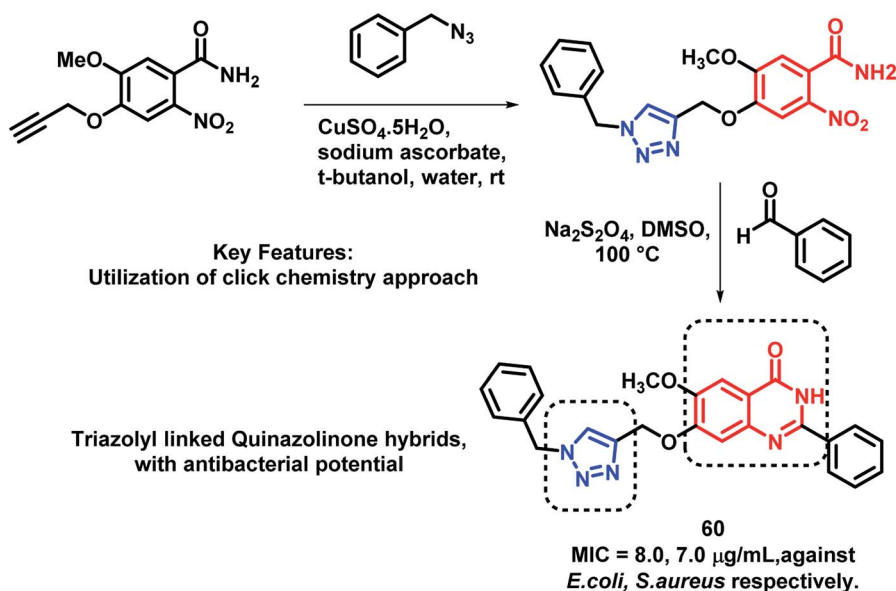
In another report, Veeramreddy *et al.* synthesized triazolyl-quinazolinone hybrids *via* the click chemistry approach.⁸⁵ The paper disc method was utilized for the anti-bacterial screening of these analogues against *Escherichia coli* (MTCC 443) and *Staphylococcus aureus* (MTCC 096). All the compounds have shown moderate to good anti-bacterial activity with MIC in the range of 7–30 $\mu\text{g mL}^{-1}$, as compared with Ciprofloxacin (MIC of 20 and 21 $\mu\text{g mL}^{-1}$ against *E. coli* and *S. aureus*, respectively), used as the positive control. Compound **60** was the most potent analogue (Scheme 26).



Scheme 24 Synthetic scheme for quinazolinone- and triazole-based hybrids, showing its key features.



Scheme 25 Synthetic scheme for quinazolinone- and triazole-based hybrids.



Scheme 26 Synthetic strategy for quinazolinone-thiazole hybrids.

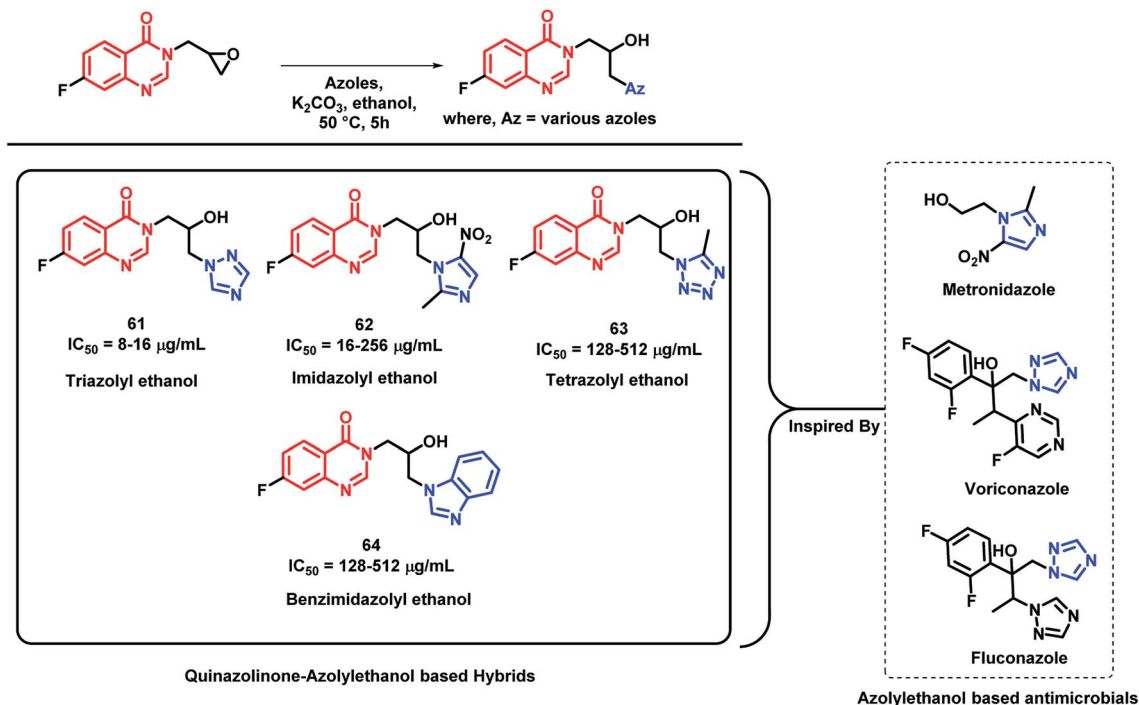
Azolyl ethanol fragment is widely present in various biologically-active compounds.⁸⁶ Many azolyl ethanol have been successfully developed and are used to treat clinical infections (fluconazole and voriconazole, which are triazole type of azolyl ethanol, are used as first line drugs to treat fungal infections).⁸⁷ Various azolyl ethanol fragments such as imidazolyl, triazolyl, tetrazolyl, benzoimidazolyl, and benzotriazolyl were incorporated at the 3rd position of quinazolin-4(3H)one by Peng *et al.* to get a series of hybrid analogues as potent antimicrobial agents.⁸⁸ These synthesized analogues were screened for *in vitro* anti-bacterial activity against Gram-positive (*Micrococcus luteus* ATCC4698, MRSA, *Staphylococcus aureus* ATCC25923, and *Bacillus subtilis* ATCC6633) and Gram-negative strains of bacteria (*Pseudomonas aeruginosa* ATCC27853, *Escherichia coli* DH52, *Bacillus proteus* ATCC13315, and *Eberthella typhosa* ATCC14028). Amongst the synthesized hybrid analogues, triazolyl ethanol hybrids were found to effectively inhibit all the bacterial strains, except for *P. aeruginosa*, and their activities were comparable or even superior than that of norfloxacin (MIC of 8 µg mL⁻¹) and chloromycin (MIC of 16 µg mL⁻¹) against MRSA. Analogue **61** (MIC of 8 µg mL⁻¹) was found to be potent in the series. Further, the docking study of

analogue **61** has shown good interaction with MRSA DNA (PDB; 2XCS), which further supports the *in vitro* results. Imidazolyl ethanol hybrids has shown potential inhibitory activity against *E. coli* DH52 (MIC of 16 µg mL⁻¹), while nitroimidazolyl analogues are potent against *M. luteus* and *P. aeruginosa* with MIC values between 16 and 32 µg mL⁻¹. The most potent analogues of each series (**61**, **62**, **63**, and **64**), along with their MIC values, are depicted in Scheme 27.

Desai *et al.* have synthesized novel 5-arylidine derivatives of thiazolidin-4-one and quinazolin-4(3H)one.⁸⁹ No derivative was found to be exceptionally potent in anti-microbial screening. Amongst them, compound **65a** was found to be potent, with the MIC value of 25 µg mL⁻¹ against *E. coli*.

In another study, in order to improve the anti-bacterial activity of previously synthesized quinazolinone and thiazolidin-4-one hybrids, the hydroxyl group at the *para* position of the phenyl ring directly attached to thiazolidinone was replaced by fluorine by Desai *et al.*⁹⁰ The concept of increasing the hydrophobicity and the permeability of the compounds was effective, which resulted in potent fluorinated analogues. As compared with hybrid analogue **65a** (MIC = 25, 100 µg mL⁻¹ against *E. coli* and *P. aeruginosa*, respectively), analogue **66** was

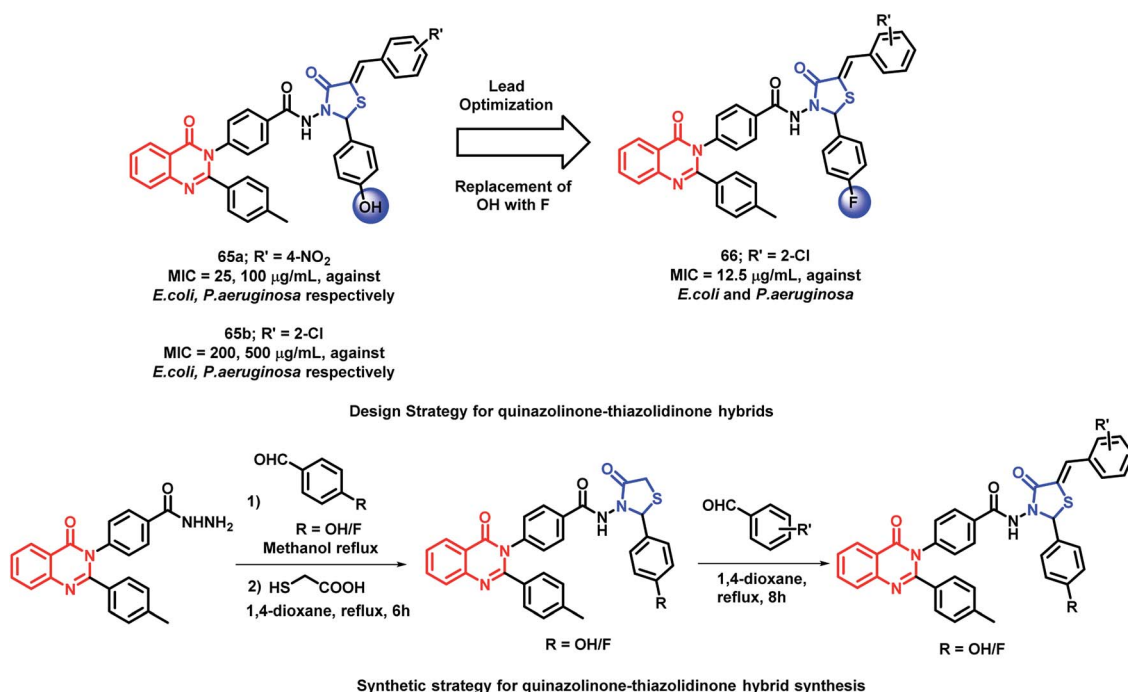




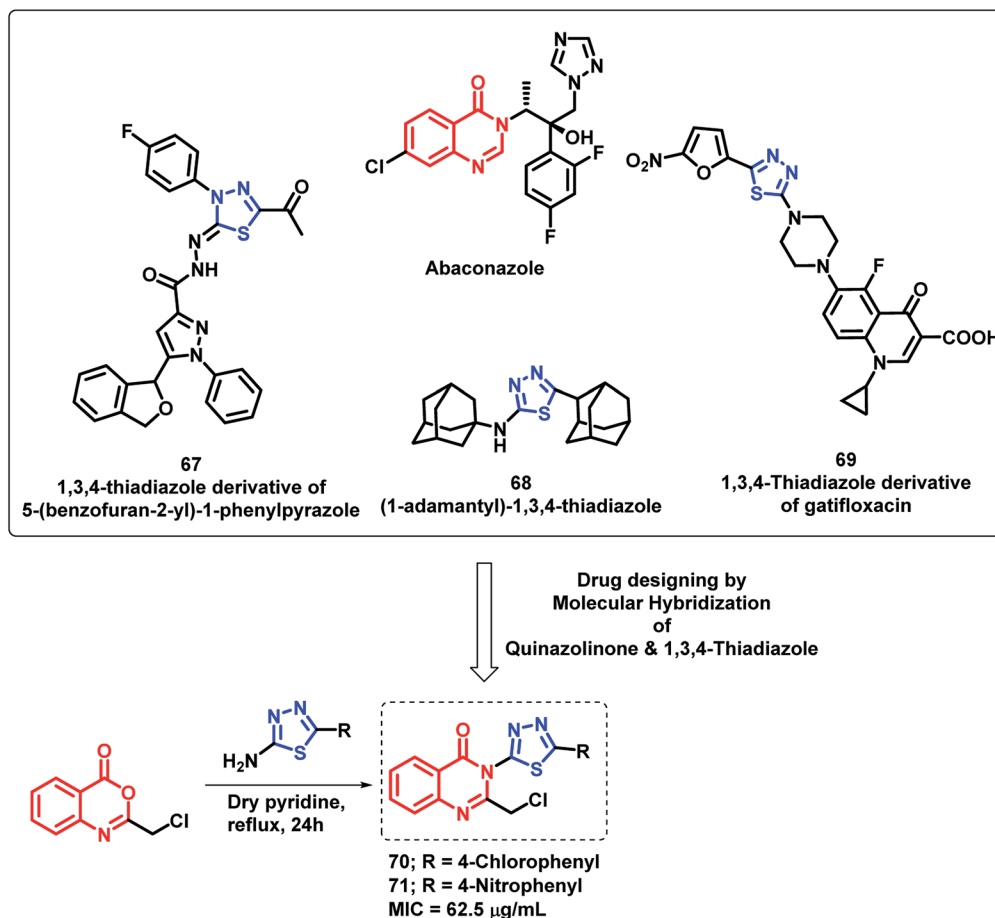
Scheme 27 Synthetic scheme for various azolyl ethanol-based quinazolinone hybrids.

most potent with an MIC of $12.5 \mu g mL^{-1}$ against both *E. coli* and *P. aeruginosa*. The presence of electron withdrawing groups such as chloro, bromo, fluoro, and nitro at the arylidine ring are effective for anti-bacterial activity. The comparison of compound **65b** and **66** is shown in Scheme 28.

By considering various thiadiazole-containing anti-microbial agents (**67**, **68**, **69**), Patel *et al.* synthesized novel quinazolino-thiadiazole hybrids and evaluated their anti-bacterial and anti-fungal potential.⁹¹ The MIC values of all the synthesized compounds were found in the range of $62.5-250 \mu g mL^{-1}$



Scheme 28 Synthesis and design strategy for quinazolinone-thiazolidinone hybrids, showing the importance of fluorine in the anti-microbial potential.

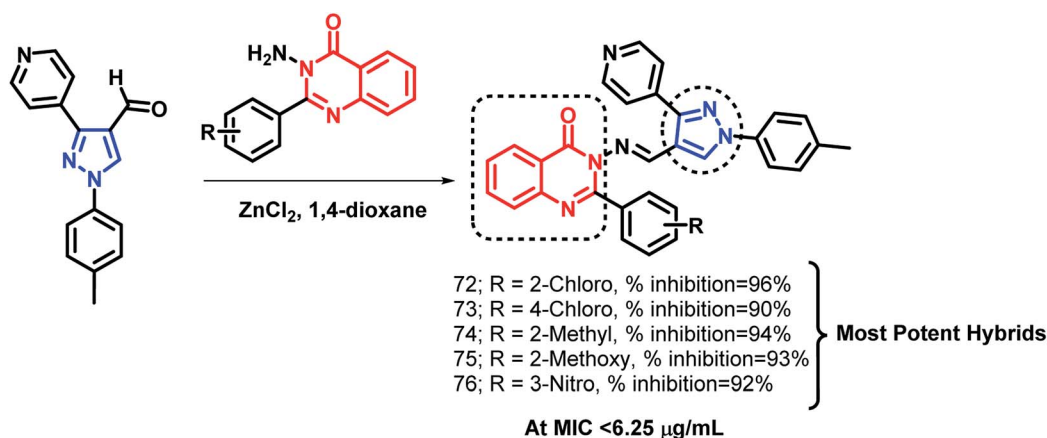


Scheme 29 Synthesis and design strategy for quinazolinone-thiadiazole hybrids.

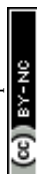
against all the evaluated strains. From the SAR study of the synthesized analogues, it was found that the electron withdrawing groups at the phenyl ring are important for the antimicrobial activity. Compounds **70** and **71** with Cl and NO₂ groups at the *para* position of the phenyl ring attached at C-2 of thiadiazole were most active with an MIC value of 62.5 µg mL⁻¹ against Gram-negative bacteria (*S. aureus*, *B. subtilis*, *E. coli*, *P. vulgaris*) and fungal strains (*C. albicans*, *A. niger*) (Scheme 29).

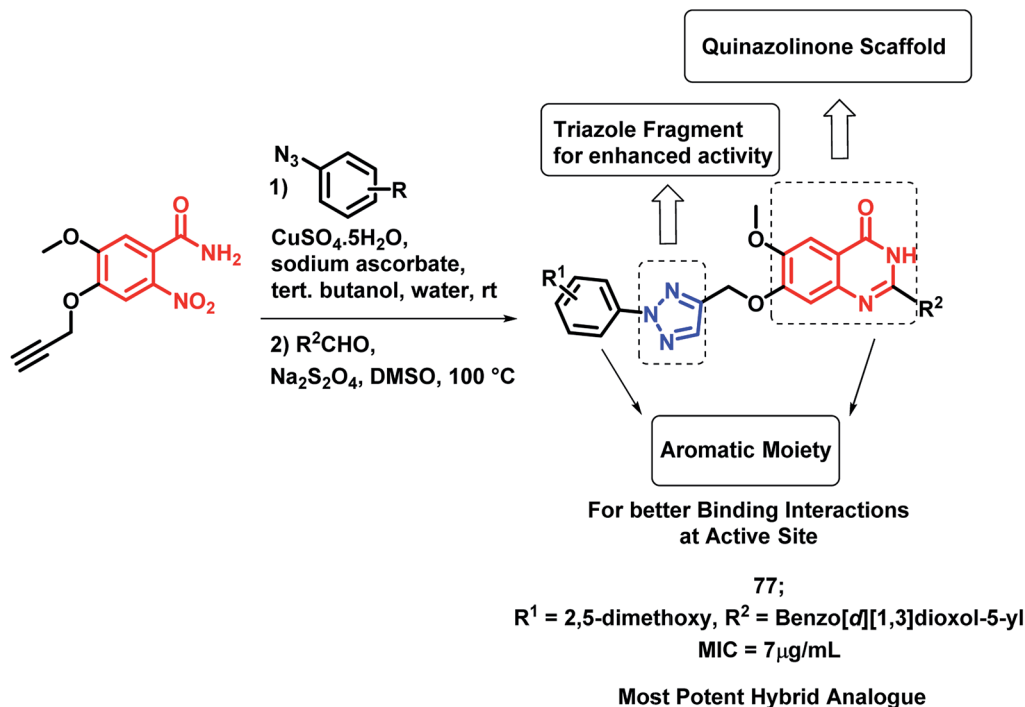
4. Anti-tubercular hybrids

Pandit *et al.* designed hybrid analogues based on pyrazole and quinazolinone for targeting *M. tuberculosis*.⁹² Radiometric BACTEC and broth dilution assay methods were utilized for the *in vitro* screening of the synthesized analogues against *M. tuberculosis* H37Rv. Isoniazid was taken as a standard with % inhibition of 99% at an MIC < 6.5 µg mL⁻¹. Some of the



Scheme 30 Synthetic scheme for pyrazole-linked quinazolinone hybrids.

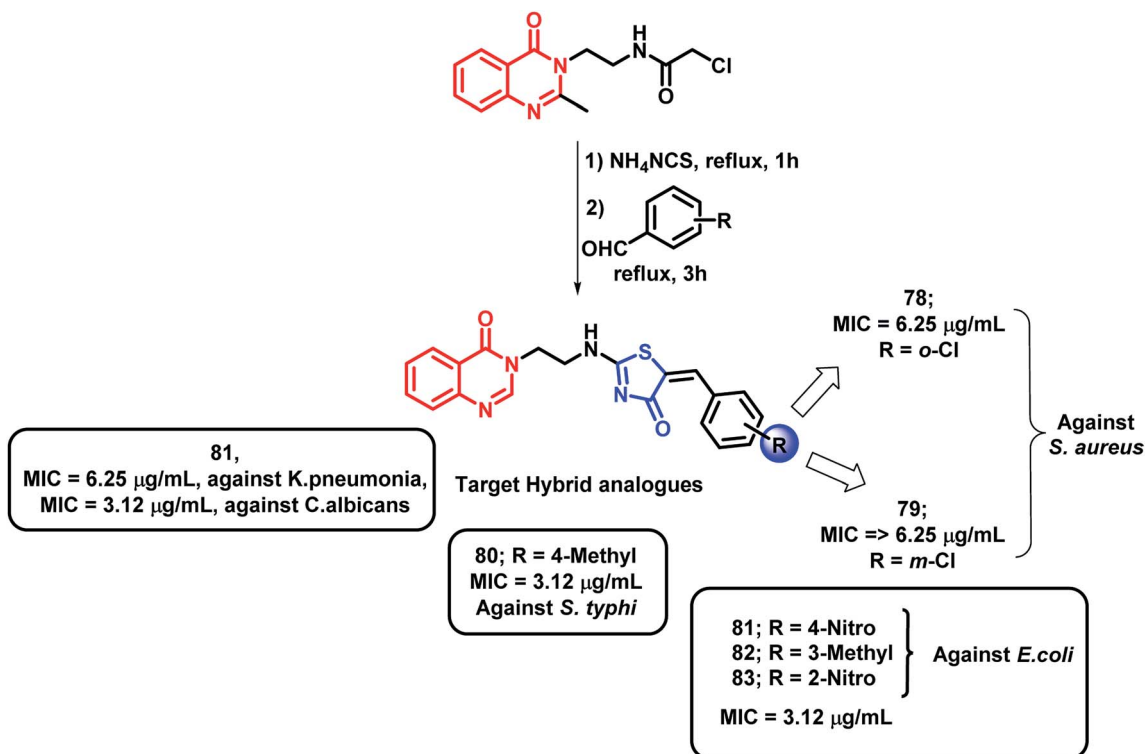




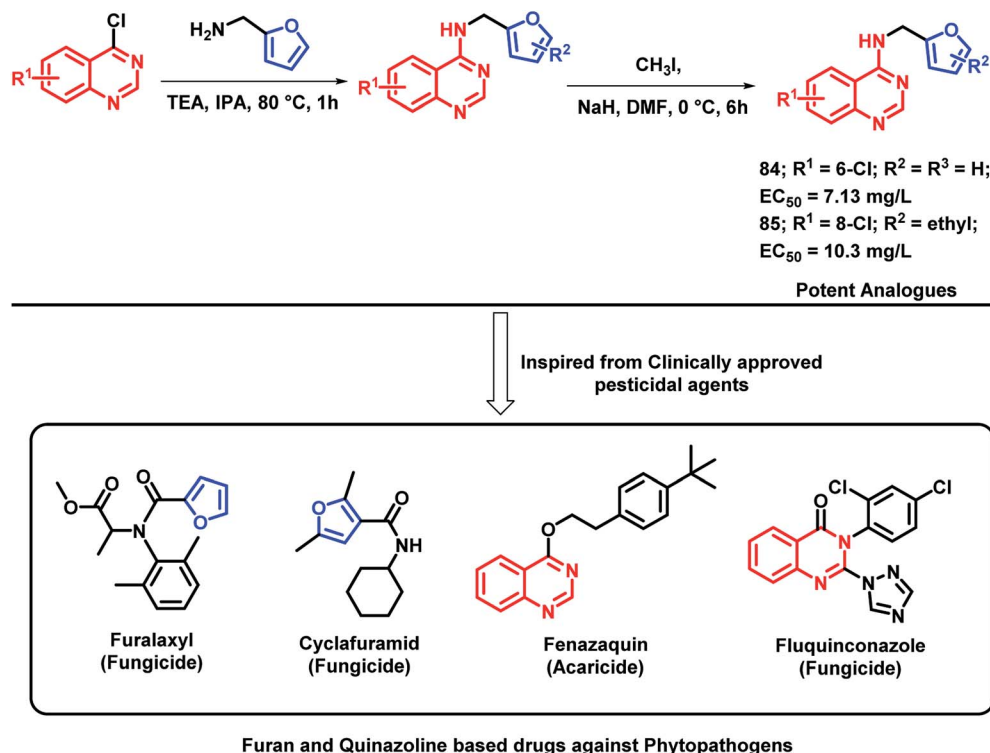
Scheme 31 Synthetic scheme for quinazolinone-triazole hybrids, showing the importance of each structural unit.

compounds (72, 73, 74, 75, 76) exhibited excellent anti-tubercular activity with % inhibition of 96, 90, 94, 93, and 92, respectively, at an MIC of less than $6.25\text{ }\mu\text{g mL}^{-1}$ (Scheme 30). According to the SAR study, the electron donating groups (Cl,

CH_3 , OCH_3) at the *ortho* position (compounds 72, 74, 75) and the electron withdrawing groups at the *meta* position (compound 76) are responsible for the increased anti-tubercular activity.



Scheme 32 Synthetic scheme for quinazolinone- and thiazolidinone-based hybrids.



Scheme 33 Synthesis and design strategy for furan-linked quinazoline hybrids.

In the study by Maddali *et al.*, triazole- and quinazolinone-linked hybrid analogues were designed through the molecular modelling study, targeting pantothenate synthase protein (PDB code: 3IVX).⁹³ The effects of substitutions on the anti-tubercular activity was studied, with respect to the quinazolinone core, 1,2,3-triazole moiety, and phenyl ring attached to triazole. Through the SAR study, it was found that electron donating groups on the phenyl ring, electron withdrawing groups on triazole, and electron donating groups on the quinazolinone ring enhanced the anti-tubercular activity. Compound 77 was the most potent hybrid analogue with an MIC of $7 \mu\text{g mL}^{-1}$ against *M. tuberculosis* H₃₇Rv (Scheme 31), using Rifampicin as a standard drug.

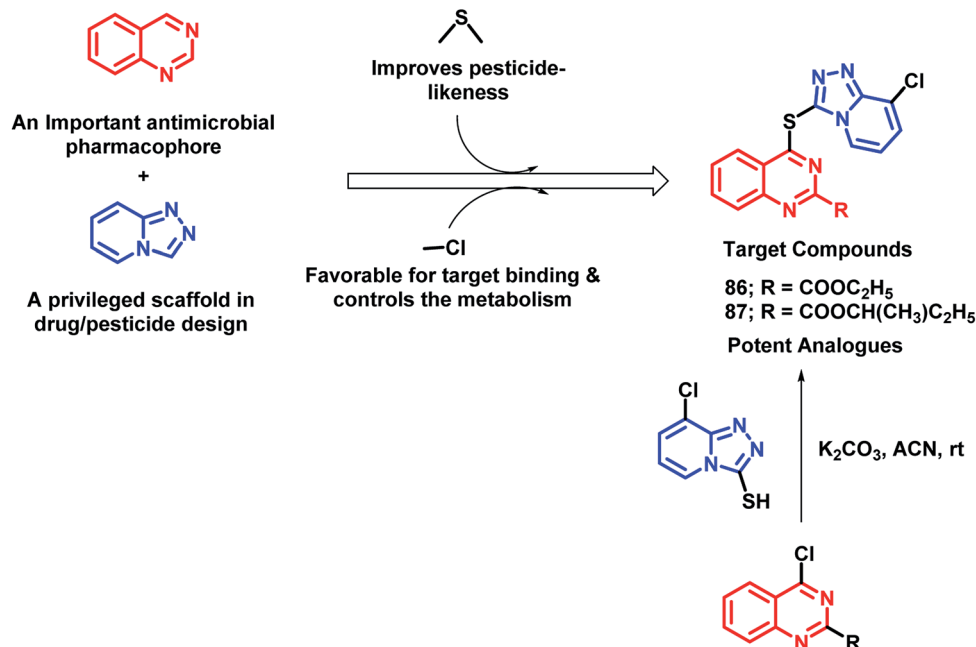
Hybrids of quinazolinones and thiazolidinones were designed by Shah *et al.* and evaluated for their anti-tubercular as well as the anti-microbial potential against *M. tuberculosis* H37Rv, fungal strains, and Gram-positive and Gram-negative bacteria.⁹⁴ Compounds with Cl and NO₂ group at the 2nd position (compounds 78 and 83, respectively) were found to be active (MIC of 6.25 and $3.12 \mu\text{g mL}^{-1}$, respectively) against Gram-positive *S. aureus*. Compounds 81, 82, and 83 showed good activity against Gram-negative *E. coli* (MIC of $3.12 \mu\text{g mL}^{-1}$). The *S. typhi* strain was found to be inhibited by 80 and 81 (MIC of $3.12 \mu\text{g mL}^{-1}$); 81 also showed excellent inhibition of *K. Pneumonia* (MIC of $6.25 \mu\text{g mL}^{-1}$). Ciprofloxacin was used as a standard for the *in vitro* anti-bacterial screening. As shown in Scheme 32, compounds 80 and 81, with NO₂ and CH₃ groups attached, were found to be active against *M. tuberculosis* H37Rv, with % inhibition of 99% at the MIC of $6.25 \mu\text{g mL}^{-1}$. The

fungal strain *C. albicans* was inhibited by compound 81 with an MIC of $3.12 \mu\text{g mL}^{-1}$ as compared with the standard drug ketoconazole (MIC = $1.56 \mu\text{g mL}^{-1}$). All the anti-tubercular results were compared with isoniazid, rifampicin, ethambutol, and pyrazinamide standards (MIC = 0.20, 0.25, 3.12, and 6.25 $\mu\text{g mL}^{-1}$, respectively).

Plant pathogenic bacteria are widely studied because of their threats towards agricultural production. Molecular hybridization technology can be used in order to inhibit such plant pathogens in the era of drug resistance. Quinazoline is a biologically-diverse scaffold, including some clinically approved pesticides, for *e.g.*, fluquinconazole (fungicide), fenazaquin (acaricide), and pyrifluquinazon (insecticide). Also, by taking into consideration the various furan-based pesticides (cyclofuramid and furamizole), Long *et al.* designed quinazoline-furan-based hybrids.⁹⁵ The designed furan-functionalized quinazolin-4-amines were synthesized and tested for their anti-bacterial activity. Compounds 84 and 85 were found to be effective in inhibiting the growth of *Xanthomonas oryzae* pv *oryzae* and *X. axonopodis* pv. *citri*, with EC₅₀ values of 7.13 and 10.3 mg L⁻¹, respectively (Scheme 33).

In continuation of the development of anti-microbial agents against phytopathogens, the hybrid drug approach was utilized by Fan *et al.*⁹⁶ In an attempt to design such hybrids, quinazoline and 1,2,4-triazolo[4,3-*a*]pyridine (privileged scaffold for the discovery of drug/pesticide leads) were linked through a thioether linkage (Scheme 34). Thus, all the hybrids were designed, synthesized, and evaluated as anti-microbial agents. Amongst all the analogues, 86 and 87 were found to be potent with EC₅₀



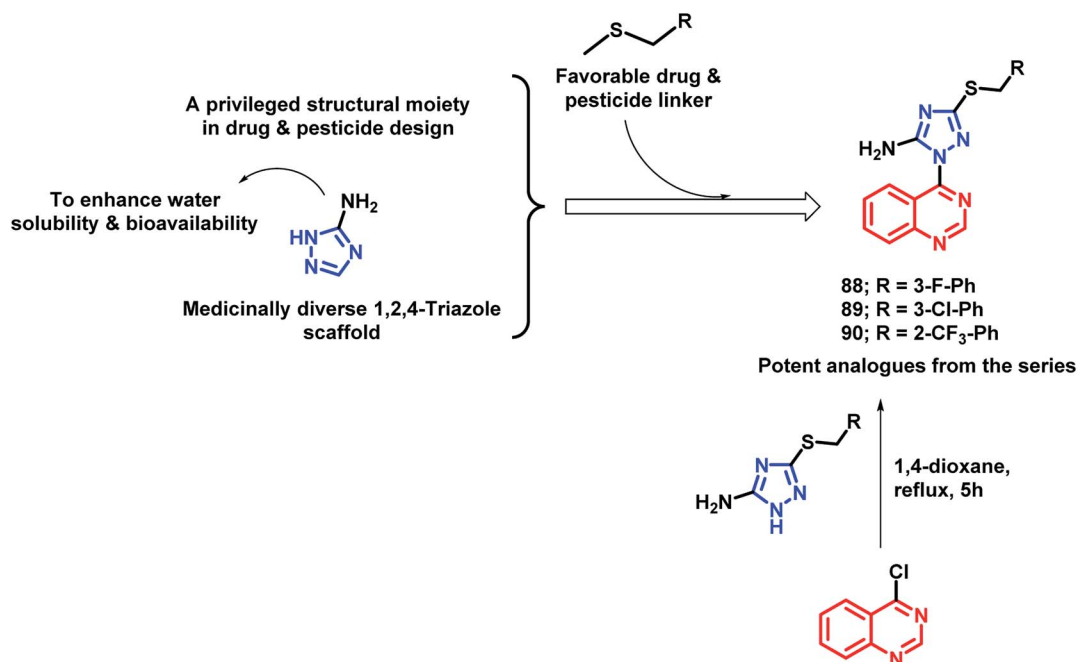


Scheme 34 Synthesis and design strategy for quinazoline-linked 1,2,4-triazolopyridine hybrids.

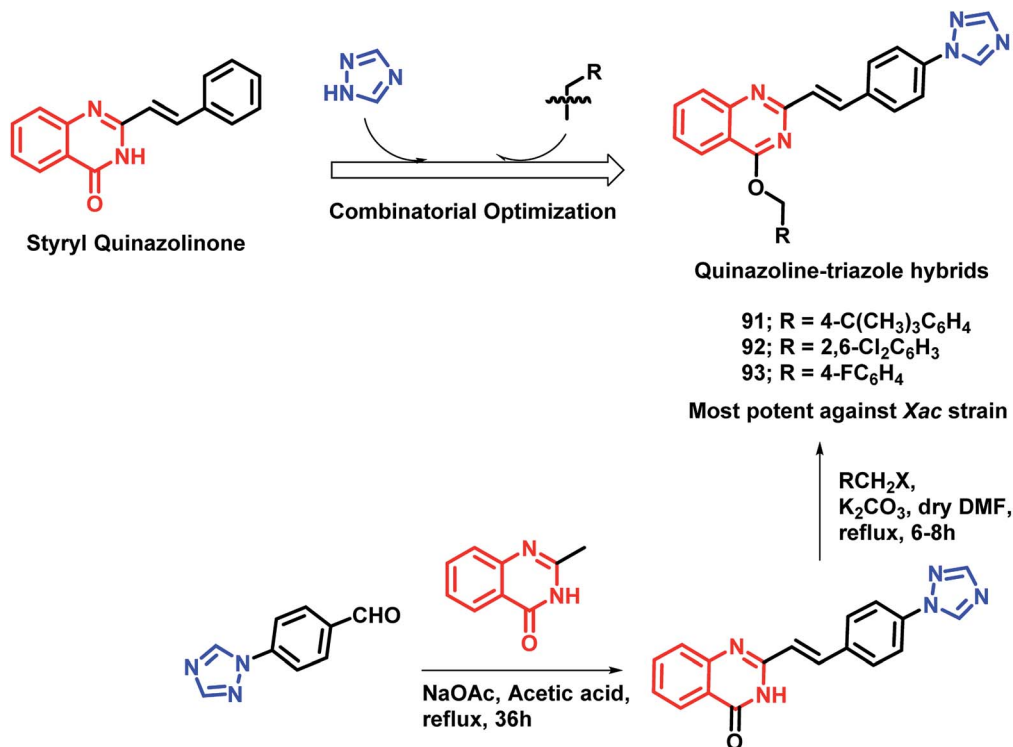
of 10.0 and $24.7 \mu\text{g mL}^{-1}$, respectively. The SAR study revealed some pharmacophoric features: (a) carboxylic acid ester at the 2nd position of quinazoline is essential for the anti-bacterial activity; (b) amongst the halogenated compounds, fluorine-containing molecules were found to have greater potential, probably due to the improved lipophilicity, bioavailability, and metabolic stability imparted by fluorine.

In another study by Fan *et al.*, quinazoline was linked with 5-amino-1,2,4-triazole by the pharmacophore hybrid approach to

obtain anti-microbial agents against plant pathogens.⁹⁷ Along with quinazoline and triazole that have pesticidal effects, two additional features were incorporated into the designed molecules: (a) an amino group was placed on triazole in order to increase the water solubility and bioavailability, (b) thioether linkage for H-bond interaction and decreasing lipophilicity. Anti-bacterial activities of all the tested analogues were tested against 3 pathogenic bacteria, namely, *Xanthomonas axonopodis* pv. *citri* (Xac), *Ralstonia solanacearum* (Rs), and *Xanthomonas*



Scheme 35 Synthesis and design strategy for quinazoline and 5-aminotriazole hybrids.



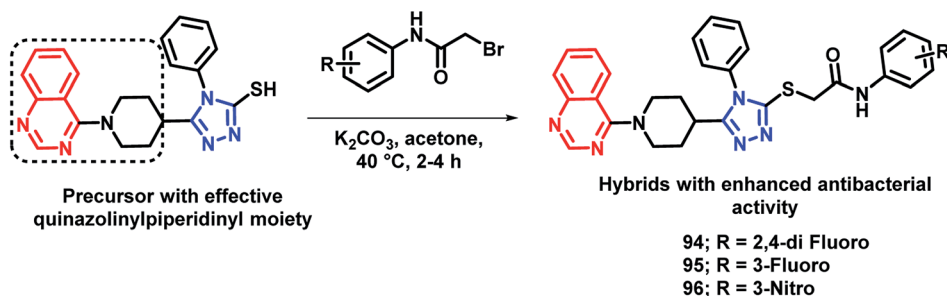
Scheme 36 Synthesis and design strategy for triazole- and styryl quinazoline-based hybrids.

oryzae pv. *oryzae* (*Xoo*), by taking Bismethiazol (BMT) and Thiodiazole-copper (TDC) as the control agents. Compounds **88**, **89**, and **90** were found to be potent with EC₅₀ of 46.9, 47.8, and 43.2 $\mu\text{g mL}^{-1}$, respectively, against *Xac* (Scheme 35). These analogues were found to be more potent as compared with the commercial agrobactericide bismethiazol (56.9 $\mu\text{g mL}^{-1}$).

As quinazoline and triazole are well-explored as anti-microbial agents,^{98–105} Yang *et al.* designed triazolyl-linked quinazoline hybrids.¹⁰⁶ As many styryl quinazolines have been reported to have anti-microbial and anti-mycobacterial activities,^{107,108} hence, styryl quinazolines were considered over simple ones. Also, various alkyl (aryl) methyleneoxy units were incorporated on the quinazoline backbone in order to study its effect on the anti-microbial activity. Turbidometric assay was used for the anti-bacterial screening of the synthesized analogues against phytopathogenic bacteria (*Xoo*, *Xac*, *Rs*). Bismethiazol (BMT), which is a commercial bactericide, was

used as a positive control. After the bioassay, compounds **91**, **92**, and **93** (% inhibition of 57.3, 66.6, and 57.8, respectively; at a concentration of 100 $\mu\text{g mL}^{-1}$) were found to be potent (Scheme 36), comparable or even better than that of BMT (% inhibition of 58.0 at a concentration of 100 $\mu\text{g mL}^{-1}$ against the *Xac* strain).

By considering the diverse range of activities of quinazoline and triazole scaffolds, Yang *et al.* designed hybrid analogues by linking these motifs together.¹⁰⁹ Molecules were designed with the following rationale: (a) 1,2,4-triazole-based analogues are well reported with anti-bacterial activities,^{105,110,111} with Triadimefon, Triadimenol, Diniconazole, Flusilazole, and Difenoconazole, which are clinically used agrofungicides. (b) The quinazoline moiety is found in many pesticidal drugs such as Fluquinconazole and Pyrifluquinazon. (c) Acetamide is an important linkage as it is a part of a fungicide, Oxathiapiprolin. (d) Quinazoline with the piperidinyllinker is important to



Scheme 37 Synthesis of quinazoline- and triazole-linked hybrids.



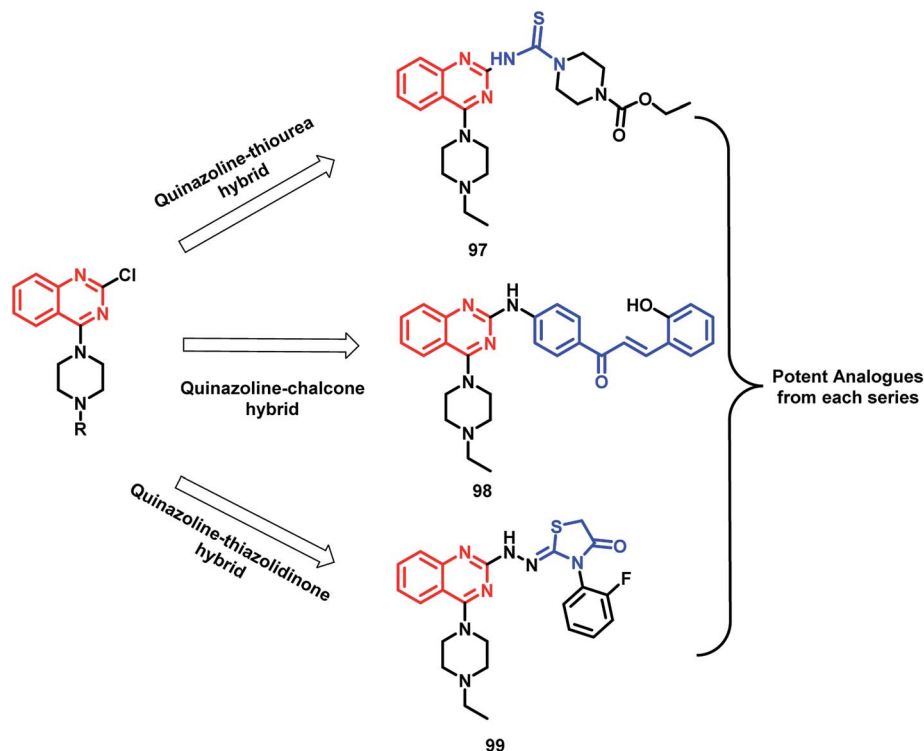
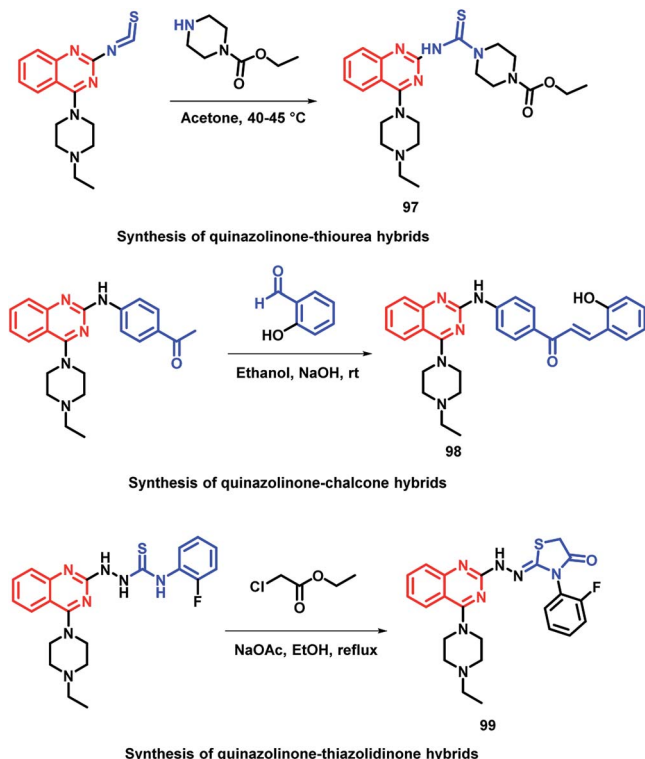


Fig. 5 Design strategy for quinazoline-linked thiourea (97), chalcone (98), and thiazolidinone (99) hybrids.



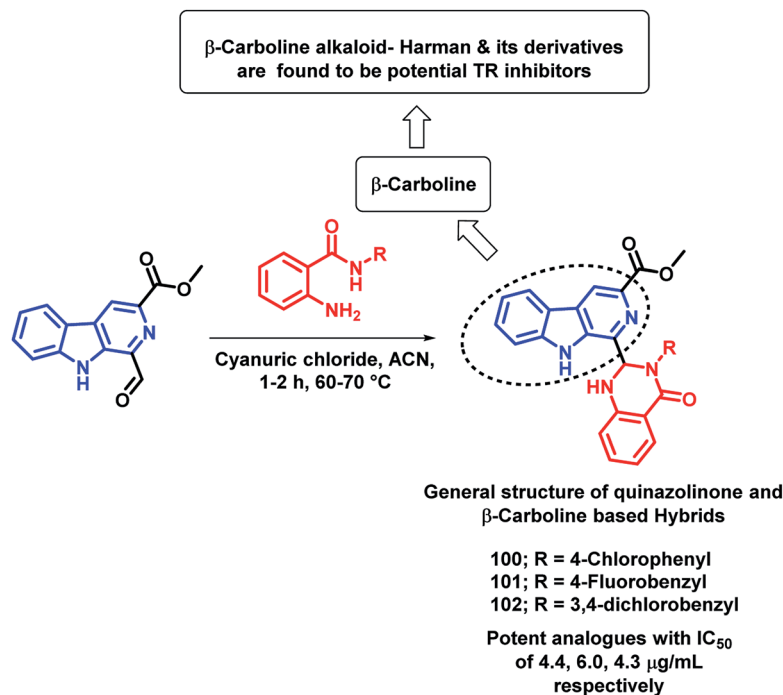
Scheme 38 Synthesis of quinazoline-linked thiourea (97), chalcone (98), and thiazolidinone (99) hybrids.

achieve effective anti-bacterial properties. By taking the above points in consideration, various *N*-(substituted phenyl)acetamide groups were linked with the quinazolinylpiperidinyl-modified 1,2,4-triazole ring (Scheme 37). These hybrid molecules were synthesized and screened for anti-bacterial activities. Compounds **94**, **95**, and **96** were found to be more potent, with EC_{50} in the range of 34.5–39.0 $\mu\text{g mL}^{-1}$ as compared with that of the standard BMT (EC_{50} of 85.6 $\mu\text{g mL}^{-1}$) against the *Xoo* pathogenic strain of bacteria.

In the study by Shah *et al.*, various hybrids were synthesized by fixing the quinazoline moiety and linking various other pharmacophores on it in order to study their effect on the activity.¹¹² Quinazoline-thiourea, quinazoline-chalcone, and quinazoline-thiazolidinone hybrids were designed (Fig. 5, Scheme 38), followed by *in vitro* anti-bacterial and anti-fungal screening. Compound **97** was potent from the thiourea series. Compound **98**, a chalcone hybrid, was found to be the most potent amongst all the series of compounds. From the last thiazolidinone series, compound **99** was found to be potent as both an anti-bacterial and anti-fungal agent.

5. Anti-leishmanial hybrids

Destruction of the metabolic pathways that are essential for the survival of the parasite in the host is found to be a major target for the development of anti-leishmanial agents. Trypanothione reductase (TR) is one of the essential enzymes responsible for the anti-oxidant defence of the parasite. Previously, various anti-leishmanial agents, (with β -carboline, dihydro- β -carboline, and

Scheme 39 Synthetic scheme for β -carboline-quinazolinone hybrids.

quinazolinone) were synthesized with potent activity towards each series. By taking such factors into consideration, Chauhan *et al.* synthesized quinazolinone and β -carboline-based hybrids, followed by their *in vitro* screening for anti-leishmanial

activity.¹¹³ Many of the compounds were active in inhibiting the LdTR enzyme after *in vitro* enzyme inhibition performed against the LdTR enzyme. Compounds with aromatic groups on quinazolinone were found to be as active as the aliphatic

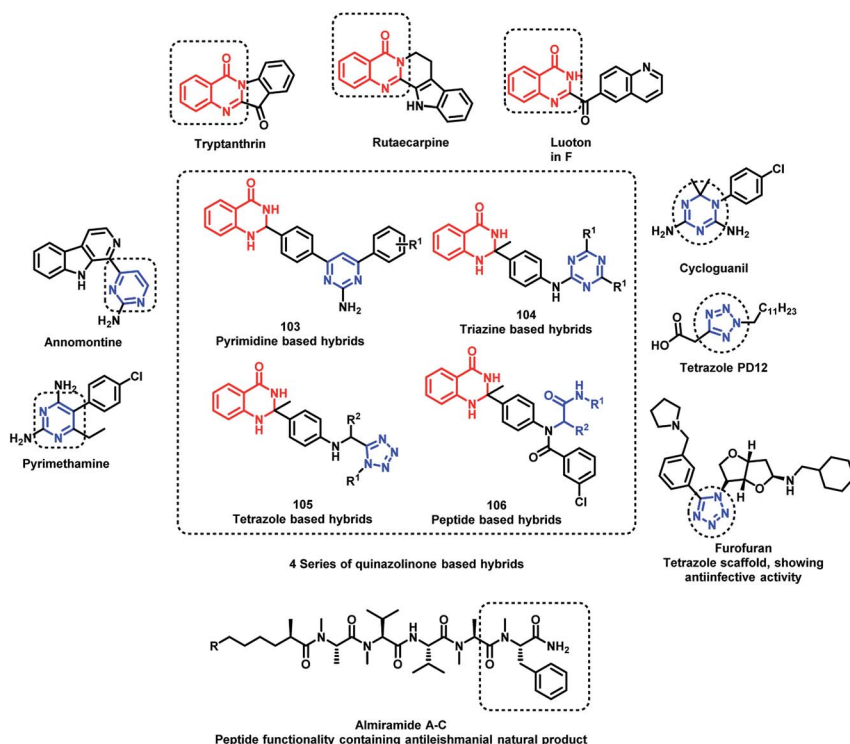
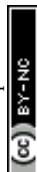
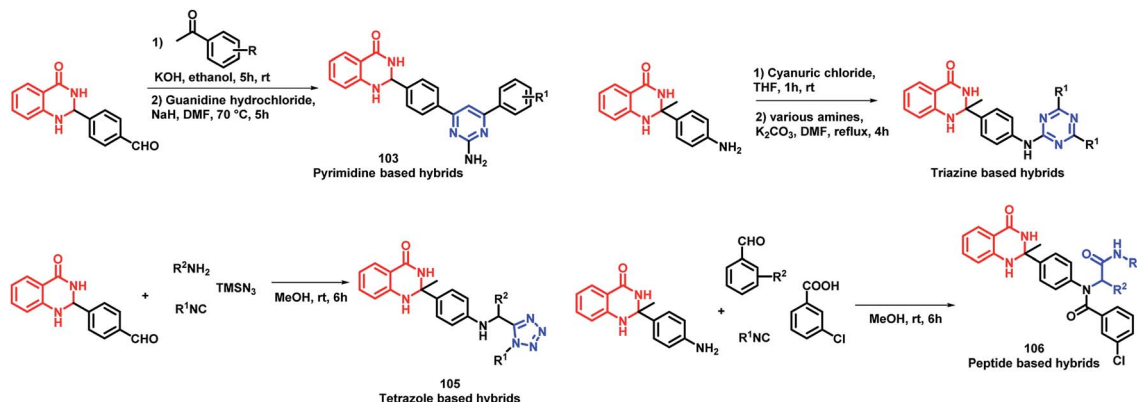


Fig. 6 Design strategy for quinazolinone hybrids with pyrimidine, triazine, tetrazole, and peptide.





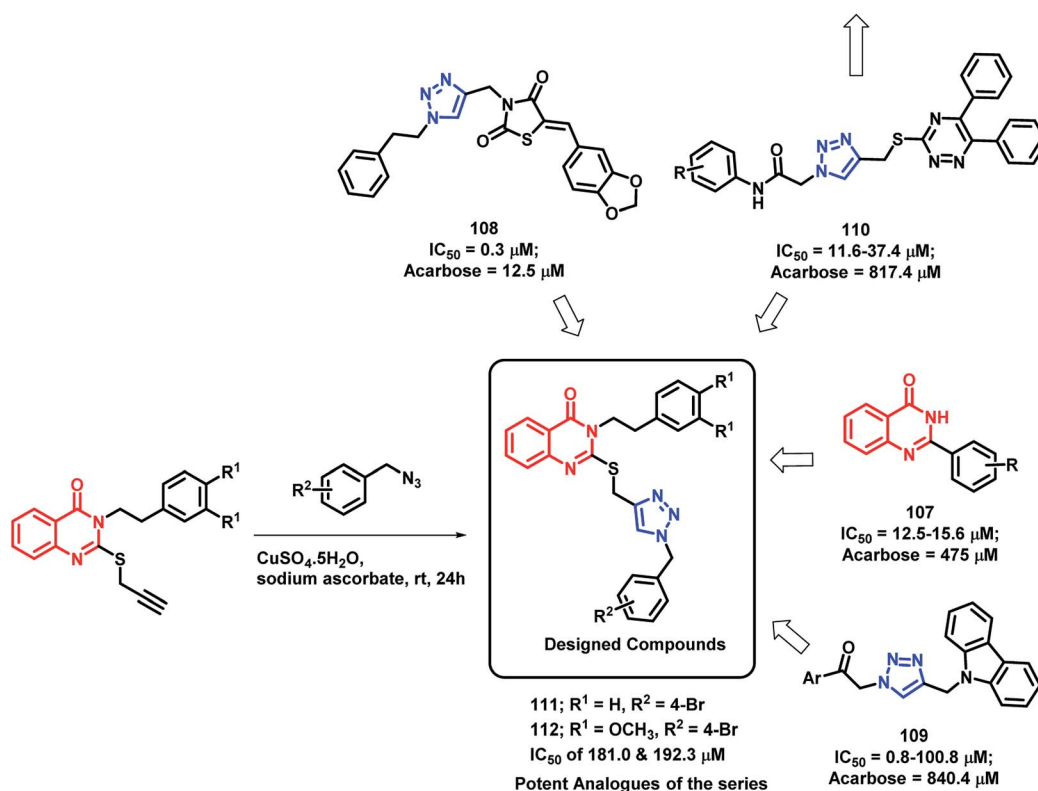
Scheme 40 Synthetic scheme for quinazolinone hybrids with pyrimidine, triazine, tetrazole, and peptide.

substituents. Aromatic groups with electron withdrawing substituents (compounds **100**, **101**, and **102**) show better inhibition as compared with the unsubstituted phenyl or phenyl ring with electron donating substituents (Scheme 39). All the enzyme inhibitors have shown competitive inhibition. Anti-leishmanial activity was also performed and the results of the enzyme assay were correlated with the anti-leishmanial activity. Compounds **100**, **101**, and **102** were found to be the most potent in the series with IC_{50} of 4.4, 6.0, and 4.3 $\mu\text{g mL}^{-1}$, respectively, against intracellular amastigotes of *L. donovani* by taking Mil-tefosine (IC_{50} of 8.1 $\mu\text{g mL}^{-1}$) and SSG (IC_{50} of 54.4 $\mu\text{g mL}^{-1}$) as the standards.

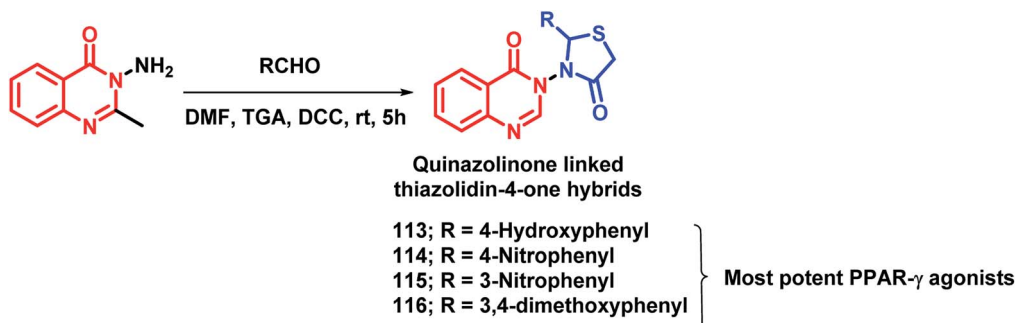
Sharma *et al.* designed 4 series of quinazolinone-based hybrids, namely, (a) quinazolinone-pyrimidine hybrids, (b) quinazolinone-triazine hybrids, (c) quinazolinone-peptide hybrids, and (d) quinazolinone-tetrazole hybrids (Fig. 6 and Scheme 40).¹¹⁴ After the screening of anti-leishmanial activity, all the series of compounds were found to be active except for the tetrazole series.

6. Anti-diabetic hybrids

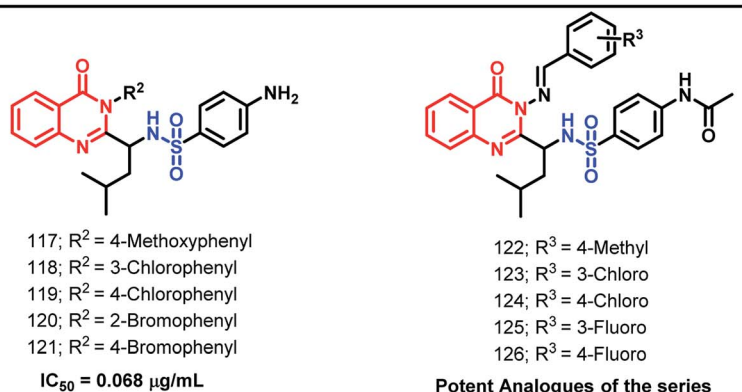
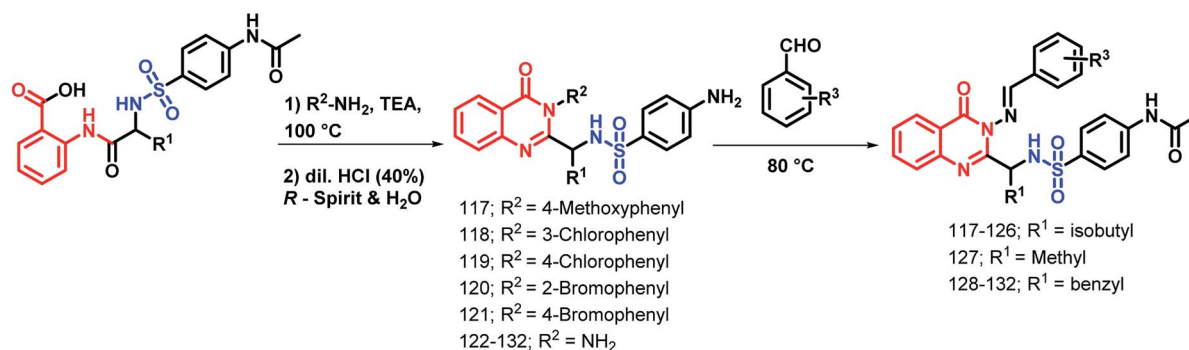
α -Glucosidase is an important enzyme responsible for the digestion of carbohydrates, which leads to the absorption of glucose and other monosaccharides. Inhibition of this enzyme reduces the postprandial blood glucose level, which results in



Scheme 41 Synthesis and design strategy for triazole-linked quinazolinone hybrids, targeting α -glucosidase.

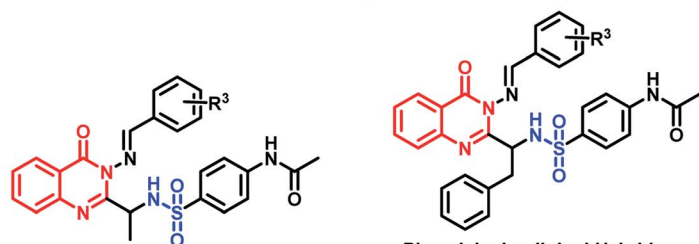


Scheme 42 Synthesis of quinazolinone-thiazolidinone hybrids.



Potent Analogues of the series
 IC₅₀ in the range of 0.068-0.092 μ g/mL

Leucine Linked Hybrids



Alanine linked Hybrids

Phenylalanine linked Hybrids

Scheme 43 Synthetic scheme for various amino acid-linked quinazolinone hybrids.



decreased risk of postprandial hyperglycemia. Previous reports have depicted the importance of quinazolinone^{115–117} as well as triazole^{118–120} in α -glucosidase inhibition. Taking such points into consideration, Saeedi *et al.* designed quinazolinone- and triazole-based hybrids and evaluated their α -glucosidase inhibition.¹²¹ All the synthesized hybrid analogues were found to inhibit the α -glucosidase enzyme through competitive inhibition with IC_{50} value in the range of 181–474.5 μ M, using acarbose (IC_{50} of 750 μ M) as the standard. Compounds **111** and **112** were the most potent with an IC_{50} of 181.0 and 192.3 μ M, respectively (Scheme 41).

By considering the importance of thiazolidinediones as PPAR- γ agonists (rosiglitazone, pioglitazone, troglitazone), it was hybridized with the medicinally important quinazolinone scaffold by Jangam *et al.*¹²² The docking study of all the designed compounds showed good binding at the active site of the PPAR- γ receptor (PDB: 4PRG). The anti-diabetic activity of the synthesized compounds was tested against Streptozotocin-induced diabetic rats and compounds **113**, **114**, **115**, **116** showed significant anti-diabetic potential (Scheme 42). All the tested compounds were tested for various biochemical parameters such as alkaline phosphatase (ALP), alanine transaminase (ALT), aspartate transaminase (AST), insulin, glycated haemoglobin (HbA1C), and lipid profile such as high density lipoprotein (HDL), low density lipoprotein (LDL), cholesterol, triglycerides, as well as total protein level, and all were found to be significant.

7. Anti-malarial hybrids

Patel *et al.* performed the *N*-heterocyclization of leucine-linked sulphonamides in order to synthesize quinazolinone- and sulphonamide-based hybrids *via* modified Grimm's method (Scheme 43).¹²³

4-Amino-*N*-[(4-oxo-3-substituted aryl-3,4-dihydroquinazolin-2-yl)alkyl]benzenesulfonamide derivatives were synthesized and tested for anti-malarial activity against the *P. falciparum* strain. Compounds **117**, **118**, **119**, **120**, and **121** have shown comparable anti-malarial activity with an IC_{50} of 0.068 μ g mL⁻¹ as compared with that of Chloroquine, Pyrimethamine, and

Trimethoprine standards, with IC_{50} of 0.020, 0.25, and 0.38 μ g mL⁻¹, respectively.

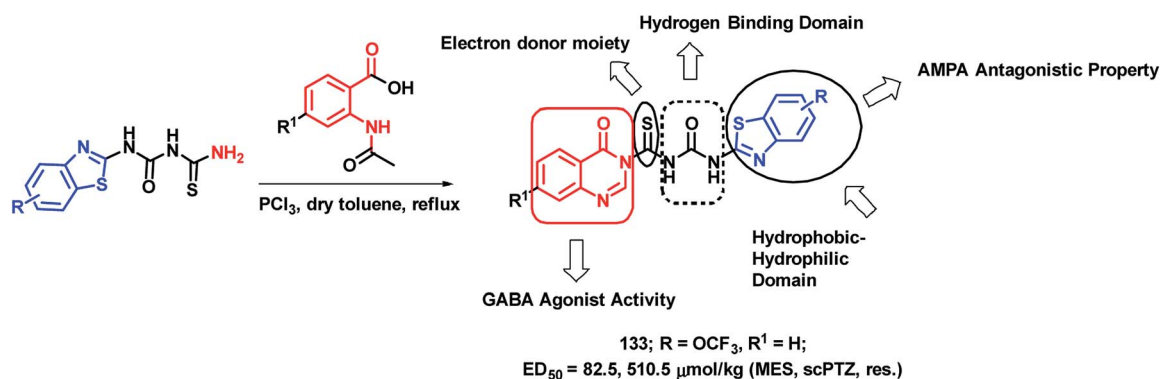
The further optimization of these compounds (4-amino-*N*-[(4-oxo-3-substituted aryl-3,4-dihydroquinazolin-2-yl)alkyl]benzenesulfonamide derivatives) was performed by the replacement of the phenyl ring at the 3rd position of quinazolinone with the benzylideneamino group.¹²⁴ The activity of such novel lead optimized analogues was tested and amongst them, compounds **122**, **123**, **124**, **125**, and **126** were found to be active as anti-malarial agents when screened against *P. falciparum*.

Patel *et al.* synthesized alanine-linked quinazolinone-sulfonamide hybrids by *N*-heterocyclization of the alanine-linked sulphonamides.¹²⁵ Compound **127** was found to be potent against anti-malarial screening (IC_{50} = 0.068 μ g mL⁻¹) as well as the DHFR inhibition assay (IC_{50} = 0.068 μ g mL⁻¹).

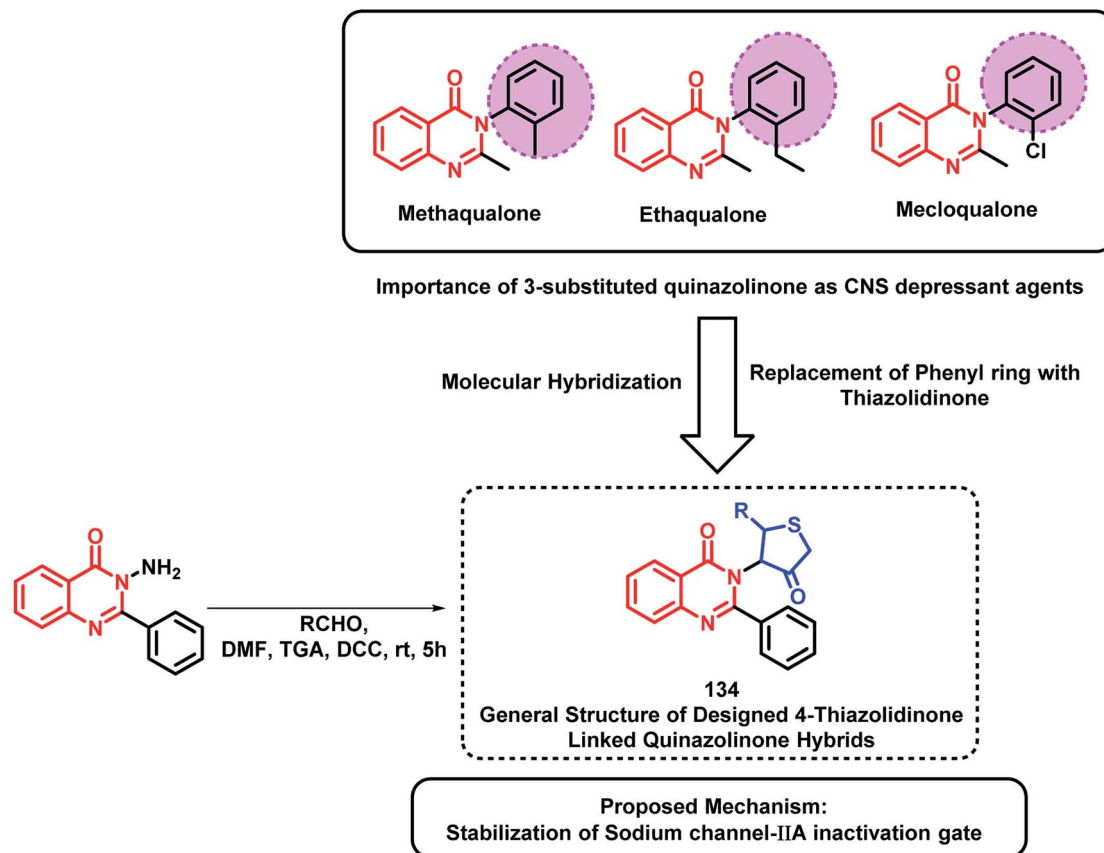
Also, the utility of phenylalanine was checked by the synthesis and anti-malarial screening of phenylalanine-linked quinazolinone-sulfonamide hybrids by Patel *et al.*¹²⁶ Compounds **128**, **129**, **130**, **131**, and **132** were found to be potent for anti-malarial efficacy against the *P. falciparum* strain with IC_{50} in the range of 0.068–0.092 μ g mL⁻¹.

8. Anti-convulsant hybrids

Epilepsy is a group of disorders, characterized by neuronal hyper excitability with episodes of sensory, motor, or autonomic phenomenon with or without the loss of consciousness. Quinazolinone derivatives are found to be potential in the treatment of epilepsy, with many clinically approved drugs such as Methaqualone, Afloqualone, and Mecloqualone. Derivatives of benzothiazole are also gaining attention for their anti-convulsant effects, mainly due to AMPA antagonism (*e.g.*, Riluzole). Hence, Malik *et al.* designed some hybrid analogues by combining GABA agonistic and AMPA antagonistic features of quinazolinones and benzothiazoles, respectively, in one hybrid scaffold.¹²⁷ The pharmacophoric features, which are listed below, were incorporated into the molecule for enhancing the anti-convulsant potential of the designed analogues. Such features are (a) hydrophobic domain, (b) hydrogen bonding domain, (c) electron donor moiety, and (d) distal hydrophobic domain. In this way, quinazolinone- and benzothiazole-based



Scheme 44 Synthetic strategy for quinazolinone-benzothiazole hybrids, showing the structural importance.



Scheme 45 Synthesis and design strategy for 4-oxothiazolidine and quinazolinone hybrids.

hybrids were designed (Scheme 44) and synthesized. Maximal electroshock (MES) and subcutaneous pentylenetetrazole (scPTZ) seizure models were developed in mice for the anti-convulsant screening of the synthesized hybrids. Compound **133** with ED_{50} values of $82.5 \mu\text{mol kg}^{-1}$ (MES) and $510.5 \mu\text{mol kg}^{-1}$ (scPTZ) was found to be more potent than phenytoin ($ED_{50} = 92 \mu\text{mol kg}^{-1}$ (MES) and $> 3540 \mu\text{mol kg}^{-1}$ (scPTZ)) and ethosuximide ($ED_{50} \geq 3540 \mu\text{mol kg}^{-1}$ (MES) and $1182 \mu\text{mol kg}^{-1}$ (scPTZ)), which were used as the positive control.

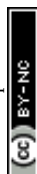
Based on quinazolinone-containing anti-convulsant analogues (methaqualone, ethaqualone, mecloqualone), it can be concluded that the aromatic ring substitution at the 3rd position of quinazolinone is essential for CNS depressant and anti-convulsant activities. Many researchers have tried to substitute several aromatic ring substituents to obtain potent CNS acting drugs. By taking such facts into consideration, Jangam *et al.* designed quinazolinone-based hybrids by linking 4-oxothiazolidine at the 3rd position of the quinazolinone backbone (Scheme 45).¹²⁸ Many of the synthesized compounds have shown moderate to good anti-convulsant potential when screened using MES-induced mice model. The SAR study showed the importance of electron withdrawing groups at the *ortho*, *meta*, and *para* position of the phenyl ring attached at the 2nd position of thiazolidine for anti-convulsant activity. These designed analogues were docked at the active site of the sodium channel IIA inactivation gate (PDB ID: 1BYY) and their binding

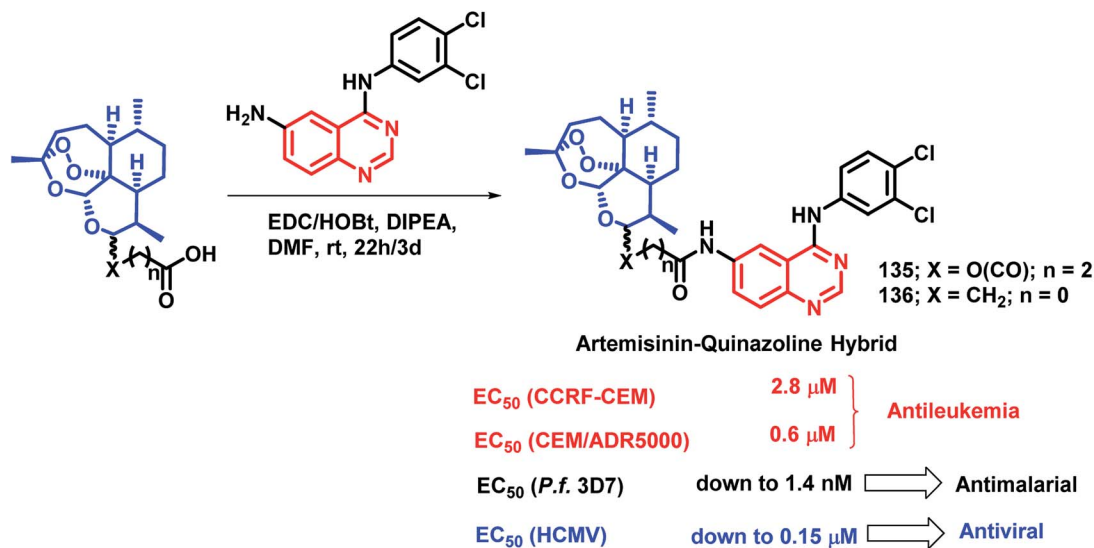
energy was in the range from -5.15 to $-6.13 \text{ kcal mol}^{-1}$, which shows proper binding at the receptor active site.

9. Hybrids with dual inhibition

Quinazoline derivatives are found to have diverse pharmacological properties including anti-malarial property (febrifugine, a well known anti-malarial drug). Hence, the quinazoline scaffold was combined with artemisinin, a known anti-malarial agent, by Frohlich *et al.* to form a hybrid pharmacophore.¹²⁹ Five novel quinazoline-artemisinin hybrids were designed, synthesized, and screened for their inhibitory potency against *Plasmodium falciparum* 3D7 strain, leukemia cell lines CCRF-CEM and CEM/ADR5000, and human cytomegalovirus (HCMV). For anti-malarial activity against *P. falciparum*, all the hybrids were found to be active with EC_{50} in the range of 1.4 – 39.9 nM , comparable to that of artesunic acid ($EC_{50} = 9.7 \text{ nM}$). Against HCMV replication in primary cell cultures, compounds **135** and **136** were found to be the most potent inhibitors, surpassing the anti-viral activity of ganciclovir by about 12–17 times. Compound **135** also showed anti-leukemia effect, with an IC_{50} of $0.6 \mu\text{M}$ against the multidrug-resistant CEM/ADR5000 cells (Scheme 46).

Cancer and diabetes are common diseases with tremendous impact on the health of human beings. It is observed that people with diabetes are at a greater risk of developing various



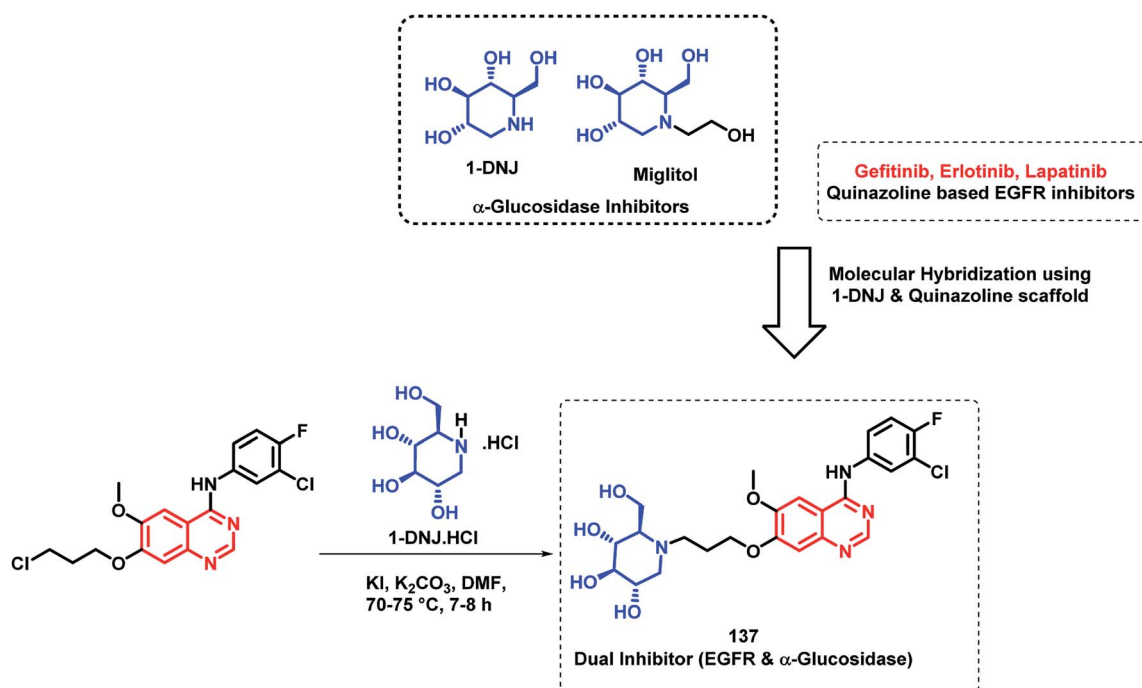


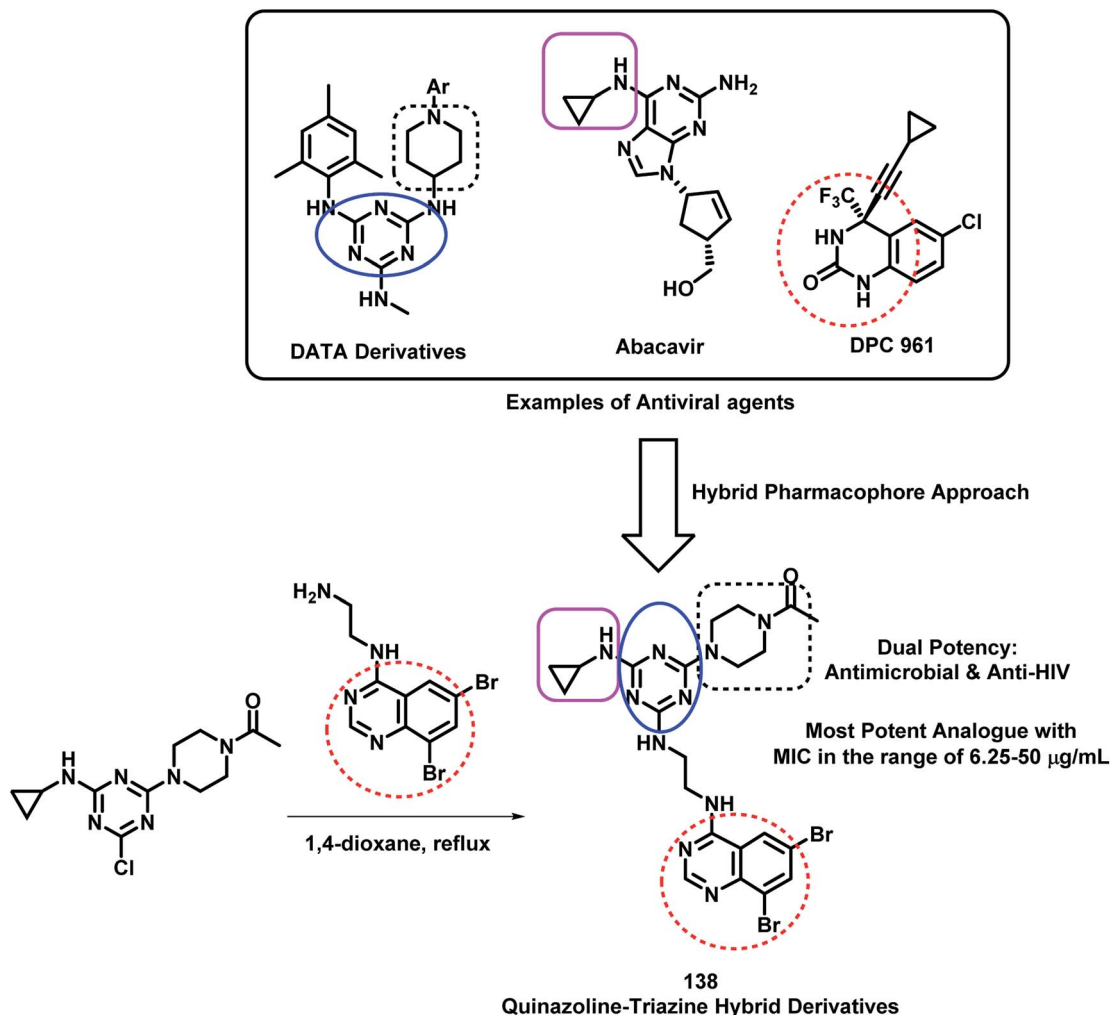
Scheme 46 Synthetic scheme for artemisinin and 4-aminoquinazoline hybrids.

forms of cancer.^{130,131} 1-Deoxynojirimycin (1-DNJ) is a naturally occurring aza sugar, having anti-cancer as well as α -glucosidase inhibitory potential. In the way of making drugs for the treatment of diabetes as well as cancer, Zhang *et al.* linked quinazoline (having anti-cancer potential) with an anti-diabetic, 1-DNJ, with α -glucosidase inhibitory potential (Scheme 47).¹³² In such a way, dual inhibitors, targeting EGFR and α -glucosidase, were designed and synthesized, followed by their *in vitro* evaluation. EGFR inhibition was evaluated by the ELISA method against recombinant EGFR^{wt} tyrosine kinase, with gefitinib and

lapatinib as the reference compounds. For the α -glucosidase inhibition assay, 1-DNJ and miglitol were selected as the standards. Many compounds were active against individual enzymes but compound 137 was found to be a dual inhibitor with high inhibition against EGFR^{wt} tyrosine kinase (IC₅₀ = 1.79 nM) and α -glucosidase (IC₅₀ = 0.39 μM).

Modh *et al.* designed some analogues as dual inhibitors for HIV and microbial infections by combining triazine, quinazoline, and piperazine moieties in one hybrid structure.¹³³ These hybrids were designed by joining various fragments found in

Scheme 47 Synthesis and design strategy for 1-deoxynojirimycin and 4-aminoquinazoline hybrids as dual inhibitors of EGFR and α -glucosidase enzymes.



Scheme 48 Synthesis and design strategy for quinazoline-triazine hybrids as anti-viral and anti-microbial agents.

clinically approved drugs, such as triazine in **R106168**, quinazoline in **DPC 961**, and piperazine type linkage in **Abacavir**. With this rationale, they synthesized novel ethylenediamine-linked quinazoline-triazine derivatives (Scheme 48), followed by their anti-microbial and anti-HIV screening. All the compounds were moderately active against HIV infection but were highly potent as anti-bacterial agents. Hence, it was found that quinazoline with ethylenediamine linkage is important for anti-microbial activity. Amongst all, compound **138** was found to be the most potent from the series, with MIC in the range of 6.25–50 $\mu\text{g mL}^{-1}$ against *S. aureus*, *B. cereus*, *P. aeruginosa*, *K. pneumonia*, *A. clavatus*, and *C. albicans*.

10. Current status, challenges, and future directions

Research interest in the synthesis and pharmacological evaluation of quinazoline/quinazolinone hybrids has increased over the years due to their diversified biological activities. Though hybrid analogues have shown to possess enhanced efficacy compared to the parent scaffold or their combinations, to the

best of our knowledge, none of the hybrid analogues in the present review have reached the clinical phase. However, a detailed literature search (<https://clinicaltrials.gov/>) revealed the following quinazoline scaffold-containing analogues in clinical trials (Table 1).

The present review summarizes the various synthetic strategies of quinazoline/quinazolinone hybrids. However, in many reports, it was observed that the hybrid analogues were designed without proper support of *in silico* studies. Most of the research in such cases has relied on the mere combination of the active functionalities from the potential agents. Nevertheless, these hybrid analogues possess good activity and proper substitution/structural modification strategy using *in silico* approaches will further help to achieve a potent activity profile.

Numerous quinazoline/quinazolinone hybrid analogues discussed above have exhibited similar or better pharmacological activity compared to that of the standard drugs. For instance, Peng *et al.* designed a hybrid of quinazoline-hydroxamic acid as a dual inhibitor for VEGFR and HDAC enzymes.³⁹ The study resulted in compound **37**, which exhibited a potential HDAC inhibition ($\text{IC}_{50} = 2.2 \text{ nM}$) and VEGFR 2

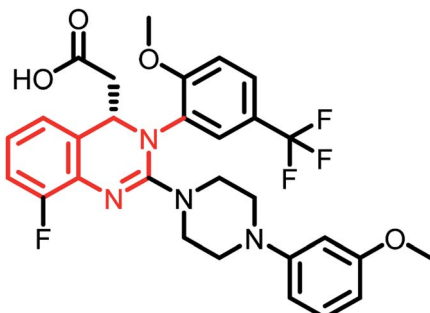


Table 1 Quinazoline-based drugs in clinical trials

Name	Status	Structure	Indication
Afatinib dimaleate	Phase II		Targeted treatment in cancers with HER2 genetic changes ¹³⁴
RXDX-105	Phase I/Ib		Non-small cell lung cancer (NSCLC) ¹³⁵
Icotinib	Phase II		Neurofibromatosis type 2 (NF2) ¹³⁶
Dacomitinib	Phase II		Advanced non-small cell lung cancer ¹³⁷
Doxazosin mesylate	Phase II		Nightmares, sleep disturbance, and non-nightmare clinical symptoms in PTSD ¹³⁸



Table 1 (Contd.)

Name	Status	Structure	Indication
Letermovir (MK-8228)	Phase II		Prevention of cytomegalovirus reactivation in patients with haematological malignancies treated with alemtuzumab ¹³⁹

inhibition ($IC_{50} = 74$ nM) as compared to the standard drugs [vorinostat ($IC_{50} = 15$ nM) and vandetanib ($IC_{50} = 54$ nM) respectively]. Apart from that, Saeedi *et al.* synthesized novel quinazolinone-1,2,3-triazole hybrids as novel anti-diabetic agents (acts *via* the inhibition of α -glucosidase).¹²¹ All the synthesized derivatives exhibited potential inhibition ($IC_{50} = 181$ to 474 μ M) compared to acarbose (standard drug) with an IC_{50} of 750 μ M. However, these studies are mainly focused on the evaluation of their *in vitro* potential towards the corresponding targets. As no further pre-clinical studies have been reported for a majority of such analogues, their clinical evaluation data could not be found in the literature.

A few studies have been reported the *in vivo* pharmacological efficacy of the quinazoline/quinazolinone hybrid. For instance, Jangam *et al.* evaluated the anti-diabetic potential of 4-thiazolidinone and 4(3*H*)-quinazolinone hybrids against streptozotocin-induced diabetic rats.¹²² The synthesized compounds exhibited a comparable anti-diabetic activity to that of pioglitazone. Also, Malik *et al.* synthesized benzotriazole-quinazolinone hybrids and evaluated the anti-convulsant activities in Wistar rats.¹²⁷ Compound 133 exhibited potential activity in maximal electroshock (MES) induced convulsion model and subcutaneous pentylenetetrazole seizure test ($ED_{50} = 82.5$ and 510.5 μ mol kg^{-1} , respectively). Phenytoin exhibited an ED_{50} of 92.0 and >3450 μ mol kg^{-1} , respectively, for the above tests. However, no reports are available for the clinical studies of these molecules and no further information is available for their current status.

In addition to the above challenges, there are certain other points to be addressed, including the reliance on the synthesis of such hybrid analogues on traditional methodologies. Efforts are required for the synthesis of these analogues in a greener, cost-effective, eco-friendly manner along with the utility of multicomponent reactions. In addition, there is a need to explore the dual inhibition ability of these molecules as they contain two scaffolds. Although some dual inhibitory activities of the hybrids are reported, such analogues exhibited non-selectivity towards the targets. Further, there are no reports on the pharmacokinetic properties of the active analogues. Studies are required in these directions for solving the same.

However, so far, the reported quinazoline/quinazolinone hybrids are still limited and their structural diversity needs further refinement for finding potential drug candidates. In the near future, continuous efforts will provide further vital discoveries as clinical drugs with quinazoline/quinazolinone hybrid scaffolds.

11. Conclusion

The present review is comprised of the rationale and various techniques involved in the design of hybrids of quinazoline/quinazolinone with other bioactive scaffolds, targeting individual or multiple targets. Major diseases, such as cancer, microbial infections, tuberculosis, leishmania, diabetes, and epilepsy, are covered in this review. Quinazoline and quinazolinone scaffolds are diverse as far as their various biological activities are concerned. Hence, many research groups have carried out extensive research for enhancing the activity of such scaffolds by hybridization with another bioactive moiety. Hybrid drug design approach has been found to be very attractive and efficient, especially in the research area of cancer and infectious diseases. The triazole moiety was present in most of the quinazoline/quinazolinone hybrids against a myriad of disease conditions. In the overall study, we found that the linking approach was used most prominently instead of framework integration for designing molecular hybrids. Overall, there is much scope of molecular hybridization approach in order to make various hybrids for better selectivity, mixed mechanism of action, overcoming the drug resistance, and so on.

Conflicts of interest

There is no conflict of interest among the authors.

Acknowledgements

The authors acknowledge Birla Institute of Technology and Science, Pilani (BITS Pilani), Pilani Campus, Rajasthan (India) for providing necessary infrastructural support. The authors



acknowledge the financial support received from DST-SERB (Grant No. CRG/2018/002608). Mr Ginson George thankfully acknowledges the Council for Scientific and Industrial Research (CSIR), New Delhi, India, for providing a fellowship (SRF-file no: 09/719 (0100)/2019-EMR-I).

References

- 1 F. Prati, E. Uliassi and M. Bolognesi, *RSC Med. Chem.*, 2014, **5**, 853–861.
- 2 M. L. Bolognesi and A. Cavalli, *ChemMedChem*, 2016, **11**, 1190–1192.
- 3 V. Ivasiv, C. Albertini, A. E. Gonçalves, M. Rossi and M. L. Bolognesi, *Curr. Top. Med. Chem.*, 2019, **19**, 1694–1711.
- 4 V. Abbot, P. Sharma, S. Dhiman, M. N. Noolvi, H. M. Patel and V. Bhardwaj, *RSC Adv.*, 2017, **7**, 28313–28349.
- 5 S. Sandhu, Y. Bansal, O. Silakari and G. Bansal, *Bioorg. Med. Chem.*, 2014, **22**, 3806–3814.
- 6 N. Kerru, P. Singh, N. Koorbanally, R. Raj and V. Kumar, *Eur. J. Med. Chem.*, 2017, **142**, 179–212.
- 7 A. Hameed, M. Al-Rashida, M. Uroos, S. A. Ali, Arshia, M. Ishtiaq and K. M. Khan, *Expert Opin. Ther. Pat.*, 2018, **28**, 281–297.
- 8 L. F. Kuyper, D. P. Baccanari, M. L. Jones, R. N. Hunter, R. L. Tansik, S. S. Joyner, C. M. Boytos, S. K. Rudolph, V. Knick and H. R. Wilson, *J. Med. Chem.*, 1996, **39**, 892–903.
- 9 H. Georgey, N. Abdel-Gawad and S. Abbas, *Molecules*, 2008, **13**, 2557–2569.
- 10 V. Srivastava and A. Kumar, *Eur. J. Med. Chem.*, 2002, **37**, 873–882.
- 11 P. M. Chandrika, T. Yakaiah, A. R. R. Rao, B. Narsaiah, N. C. Reddy, V. Sridhar and J. V. Rao, *Eur. J. Med. Chem.*, 2008, **43**, 846–852.
- 12 P. Verhaeghe, N. Azas, M. Gasquet, S. Hutter, C. Ducros, M. Laget, S. Rault, P. Rathelot and P. Vanelle, *Bioorg. Med. Chem. Lett.*, 2008, **18**, 396–401.
- 13 M. A. Ismail, S. Barker, D. A. Abou El Ella, K. A. Abouzid, R. A. Toubar and M. H. Todd, *J. Med. Chem.*, 2006, **49**, 1526–1535.
- 14 V. Alagarsamy, S. Meena, K. Ramaseshu, V. R. Solomon, T. D. A. Kumar and K. Thirumurugan, *Chem. Biol. Drug Des.*, 2007, **70**, 254–260.
- 15 M. S. Malamas and J. Millen, *J. Med. Chem.*, 1991, **34**, 1492–1503.
- 16 N. M. A. Gawad, H. H. Georgey, R. M. Youssef and N. A. El-Sayed, *Eur. J. Med. Chem.*, 2010, **45**, 6058–6067.
- 17 M. Decker, *Eur. J. Med. Chem.*, 2005, **40**, 305–313.
- 18 A. Rosowsky, J. E. Wright, C. M. Vaidya and R. A. Forsch, *Pharmacol. Therapeut.*, 2000, **85**, 191–205.
- 19 D. W. Fry, A. J. Kraker, A. McMichael, L. A. Ambroso, J. M. Nelson, W. R. Leopold, R. W. Connors and A. J. Bridges, *Science*, 1994, **265**, 1093–1095.
- 20 A. Garofalo, L. Goossens, A. Lemoine, S. Ravez, P. Six, M. Howsam, A. Farce and P. Depreux, *RSC Med. Chem.*, 2011, **2**, 65–72.
- 21 I. Khan, S. Zaib, S. Batool, N. Abbas, Z. Ashraf, J. Iqbal and A. Saeed, *Bioorg. Med. Chem.*, 2016, **24**, 2361–2381.
- 22 J. D. Palem, G. R. Alugubelli, R. Bantu, L. Nagarapu, S. Polepalli, S. N. Jain, R. Bathini and V. Manga, *Bioorg. Med. Chem. Lett.*, 2016, **26**, 3014–3018.
- 23 R. Venkatesh, S. Kasaboina, N. Jain, S. Janardhan, U. D. Holagunda and L. Nagarapu, *J. Mol. Struct.*, 2019, **1181**, 403–411.
- 24 A. Kamal, E. V. Bharathi, J. S. Reddy, M. J. Ramaiah, D. Dastagiri, M. K. Reddy, A. Viswanath, T. L. Reddy, T. B. Shaik and S. Pushpavalli, *Eur. J. Med. Chem.*, 2011, **46**, 691–703.
- 25 E. Hamel, C. M. Lin, J. Plowman, H.-K. Wang, K.-H. Lee and K. D. Paull, *Biochem. Pharmacol.*, 1996, **51**, 53–59.
- 26 H. L. Yale and M. Kalkstein, *J. Med. Chem.*, 1967, **10**, 334–336.
- 27 D. Simoni, G. Grisolia, G. Giannini, M. Roberti, R. Rondanin, L. Piccagli, R. Baruchello, M. Rossi, R. Romagnoli and F. P. Invidiata, *J. Med. Chem.*, 2005, **48**, 723–736.
- 28 J. Kaffy, R. Pontikis, D. Carrez, A. Croisy, C. Monneret and J.-C. Florent, *Bioorg. Med. Chem.*, 2006, **14**, 4067–4077.
- 29 S. T. Al-Rashood, G. S. Hassan, S. M. El-Messery, M. N. Nagi, E.-S. E. Habib, F. A. Al-Omary and H. I. El-Subbagh, *Bioorg. Med. Chem. Lett.*, 2014, **24**, 4557–4567.
- 30 S. T. Al-Rashood, I. A. Aboldahab, M. N. Nagi, L. A. Abouzeid, A. A. Abdel-Aziz, S. G. Abdel-Hamide, K. M. Youssef, A. M. Al-Obaid and H. I. El-Subbagh, *Bioorg. Med. Chem.*, 2006, **14**, 8608–8621.
- 31 F. A. Al-Omary, G. S. Hassan, S. M. El-Messery, M. N. Nagi, E.-S. E. Habib and H. I. El-Subbagh, *Eur. J. Med. Chem.*, 2013, **63**, 33–45.
- 32 P. Singla, V. Luxami and K. Paul, *J. Photochem. Photobiol., B*, 2017, **168**, 156–164.
- 33 L. Hosseinzadeh, A. Aliabadi, M. Rahnama, H. M. M. Sadeghi and M. R. Khajouei, *Res. Pharm. Sci.*, 2017, **12**, 290.
- 34 F. Hassanzadeh, H. Sadeghi-Aliabadi, E. Jafari, A. Sharifzadeh and N. Dana, *Res. Pharm. Sci.*, 2019, **14**, 408.
- 35 W.-Y. Wu, S.-L. Cao, B.-B. Mao, J. Liao, Z.-F. Li, H.-B. Song and X. Xu, *Lett. Drug Des. Discovery*, 2013, **10**, 61–66.
- 36 B. Parrino, C. Ciancimino, A. Carbone, V. Spano, A. Montalbano, P. Barraja, G. Cirrincione and P. Diana, *Tetrahedron*, 2015, **71**, 7332–7338.
- 37 J.-H. Wu, F.-R. Chang, K.-I. Hayashi, H. Shiraki, C.-C. Liaw, Y. Nakanishi, K. F. Bastow, D. Yu, I.-S. Chen and K.-H. Lee, *Bioorg. Med. Chem. Lett.*, 2003, **13**, 2223–2225.
- 38 R. Bollu, S. Banu, S. Kasaboina, R. Bantu, L. Nagarapu, S. Polepalli and N. Jain, *Bioorg. Med. Chem. Lett.*, 2017, **27**, 5481–5484.
- 39 S. Pullamsetti, G. Banat, A. Schmall, M. Szibor, D. Pomagruk, J. Hânze, E. Kolosionek, J. Wilhelm, T. Braun and F. Grimminger, *Oncogene*, 2013, **32**, 1121–1134.
- 40 M. Narita, T. Murata, K. Shimizu, T. Nakagawa, T. Sugiyama, M. Inui, K. Hiramoto and T. Tagawa, *Oncol. Rep.*, 2007, **17**, 1133–1139.



- 41 K. Murata, T. Sudo, M. Kameyama, H. Fukuoka, M. Mukai, Y. Doki, Y. Sasaki, O. Ishikawa, Y. Kimura and S. Imaoka, *Clin. Exp. Metastasis*, 2000, **18**, 599–604.
- 42 A. E. Kümmerle, M. Schmitt, S. V. Cardozo, C. Lugnier, P. Villa, A. B. Lopes, N. C. Romeiro, H. L. N. Justiniano, M. A. Martins and C. A. Fraga, *J. Med. Chem.*, 2012, **55**, 7525–7545.
- 43 A. E. Kümmerle, M. M. Vieira, M. Schmitt, A. L. Miranda, C. A. Fraga, J.-J. Bourguignon and E. J. Barreiro, *Bioorg. Med. Chem. Lett.*, 2009, **19**, 4963–4966.
- 44 H. M. Abdel-Rahman, M. Abdel-Aziz, J. C. Canzoneri, B. D. Gary and G. A. Piazza, *Arch. Pharmazie*, 2014, **347**, 650–657.
- 45 S. Thakral, D. Saini, A. Kumar, N. Jain and S. Jain, *Med. Chem. Res.*, 2017, **26**, 1595–1604.
- 46 J. Ślawiński, K. Szafranski, D. Vullo and C. T. Supuran, *Eur. J. Med. Chem.*, 2013, **69**, 701–710.
- 47 F. Xu, H. Xu, X. Wang, L. Zhang, Q. Wen, Y. Zhang and W. Xu, *Bioorg. Med. Chem.*, 2014, **22**, 1487–1495.
- 48 C. T. Supuran, F. Briganti, S. Tilli, W. R. Chegwidden and A. Scozzafava, *Bioorg. Med. Chem.*, 2001, **9**, 703–714.
- 49 M. Zayed and M. Hassan, *Drug Res.*, 2013, **63**, 210–215.
- 50 M. F. Zayed, H. E. Ahmed, S. Ihmaid, A.-S. M. Omar and A. S. Abdelrahim, *J. Taibah Univ. Medical Sci.*, 2015, **10**, 333–339.
- 51 F. Hassanzadeh, H. Sadeghi-Aliabadi, S. Nikooei, E. Jafari and G. Vaseghi, *Res. Pharm. Sci.*, 2019, **14**, 130.
- 52 S. El-Sayed, K. Metwally, A. A. El-Shanawani, L. M. Abdel-Aziz, H. Pratsinis and D. Kletsas, *Chem. Cent. J.*, 2017, **11**, 1–10.
- 53 N. Gokhale, N. Panathur, U. Dalimba, P. G. Nayak and K. S. R. Pai, *J. Heterocycl. Chem.*, 2016, **53**, 513–524.
- 54 A. Imramovský, R. Jorda, K. Pauk, E. Řezníčková, J. Dušek, J. Hanusek and V. Kryštof, *Eur. J. Med. Chem.*, 2013, **68**, 253–259.
- 55 R. Venkatesh, M. J. Ramaiah, H. K. Gaikwad, S. Janardhan, R. Bantu, L. Nagarapu, G. N. Sastry, A. R. Ganesh and M. Bhadra, *Eur. J. Med. Chem.*, 2015, **94**, 87–101.
- 56 M. F. Zayed, H. S. Rateb, S. Ahmed, O. A. Khaled and S. R. Ibrahim, *Molecules*, 2018, **23**, 1699.
- 57 S. Shinkaruk, M. Bayle, G. Lain and G. Deleris, *Curr. Med. Chem. Anti Canc. Agents*, 2003, **3**, 95–117.
- 58 T. Veikkola, M. Karkkainen, L. Claesson-Welsh and K. Alitalo, *Cancer Res.*, 2000, **60**, 203–212.
- 59 F.-W. Peng, J. Xuan, T.-T. Wu, J.-Y. Xue, Z.-W. Ren, D.-K. Liu, X.-Q. Wang, X.-H. Chen, J.-W. Zhang and Y.-G. Xu, *Eur. J. Med. Chem.*, 2016, **109**, 1–12.
- 60 X. Kang, J. Hu, Z. Gao, Y. Ju and C. Xu, *RSC Med. Chem.*, 2012, **3**, 1245–1249.
- 61 R. Majeed, P. L. Sangwan, P. K. Chinthakindi, I. Khan, N. A. Dangroo, N. Thota, A. Hamid, P. R. Sharma, A. K. Saxena and S. Koul, *Eur. J. Med. Chem.*, 2013, **63**, 782–792.
- 62 A. Worachartcheewan, N. Songtawee, S. Siri Wong, S. Prachayasittikul, C. Nantasenamat and V. Prachayasittikul, *Med. Chem.*, 2019, **15**, 328–340.
- 63 M. J. Genin, D. A. Allwine, D. J. Anderson, M. R. Barbachyn, D. E. Emmert, S. A. Garmon, D. R. Graber, K. C. Grega, J. B. Hester and D. K. Hutchinson, *J. Med. Chem.*, 2000, **43**, 953–970.
- 64 N. G. Aher, V. S. Pore, N. N. Mishra, A. Kumar, P. K. Shukla, A. Sharma and M. K. Bhat, *Bioorg. Med. Chem. Lett.*, 2009, **19**, 759–763.
- 65 G. Le-Nhat-Thuy, T. Van Dinh, H. Pham-The, H. N. Quang, N. N. Thi, T. a. D. Thi, P. H. Thi, T. A. Le Thi, H. T. Nguyen and P. N. Thanh, *Bioorg. Med. Chem. Lett.*, 2018, **28**, 3741–3747.
- 66 P. Song, F. Cui, N. Li, J. Xin, Q. Ma, X. Meng, C. Wang, Q. Cao, Y. Gu and Y. Ke, *Chin. J. Chem.*, 2017, **35**, 1633–1639.
- 67 R. Gastpar, M. Goldbrunner, D. Marko and E. Von Angerer, *J. Med. Chem.*, 1998, **41**, 4965–4972.
- 68 N. K. Kaushik, N. Kaushik, P. Attri, N. Kumar, C. H. Kim, A. K. Verma and E. H. Choi, *Molecules*, 2013, **18**, 6620–6662.
- 69 H. Hu, J. Wu, M. Ao, H. Wang, T. Zhou, Y. Xue, Y. Qiu, M. Fang and Z. Wu, *Chem. Biol. Drug Des.*, 2016, **88**, 766–778.
- 70 S. H. Watterson, T. M. Dhar, S. K. Ballentine, Z. Shen, J. C. Barrish, D. Cheney, C. A. Fleener, K. A. Rouleau, R. Townsend and D. L. Hollenbaugh, *Bioorg. Med. Chem. Lett.*, 2003, **13**, 1273–1276.
- 71 M. J. Mphahlele, M. M. Mmonwa, A. Aro, L. J. McGaw and Y. S. Choong, *Int. J. Mol. Sci.*, 2018, **19**, 2232.
- 72 M. Fares, W. M. Eldehna, S. M. Abou-Seri, H. A. Abdel-Aziz, M. H. Aly and M. F. Tolba, *Arch. Pharmazie*, 2015, **348**, 144–154.
- 73 V. Luxami, R. Rani, A. Sharma and K. Paul, *J. Photochem. Photobiol., A*, 2015, **311**, 68–75.
- 74 J. Shao, E. Chen, K. Shu, W. Chen, G. Zhang and Y. Yu, *Bioorg. Med. Chem.*, 2016, **24**, 3359–3370.
- 75 A. Sharma, V. Luxami and K. Paul, *Bioorg. Med. Chem. Lett.*, 2013, **23**, 3288–3294.
- 76 B. Banerji, K. Chandrasekhar, K. Sreenath, S. Roy, S. Nag and K. D. Saha, *ACS Omega*, 2018, **3**, 16134–16142.
- 77 F.-W. Peng, T.-T. Wu, Z.-W. Ren, J.-Y. Xue and L. Shi, *Bioorg. Med. Chem. Lett.*, 2015, **25**, 5137–5141.
- 78 J. Drews, *Science*, 2000, **287**, 1960–1964.
- 79 C. T. Supuran, A. Casini, A. Mastrolorenzo and A. Scozzafava, *Mini Rev. Med. Chem.*, 2004, **4**, 625–632.
- 80 F. Abbate, A. Casini, T. Owa, A. Scozzafava and C. T. Supuran, *Bioorg. Med. Chem. Lett.*, 2004, **14**, 217–223.
- 81 M. M. Ghorab, M. S. Alsaid, M. S. Al-Dosari, M. G. El-Gazzar and M. K. Parvez, *Molecules*, 2016, **21**, 189.
- 82 M. Dinari, F. Gharahi and P. Asadi, *J. Mol. Struct.*, 2018, **1156**, 43–50.
- 83 R. Romagnoli, P. G. Baraldi, F. Prencipe, J. Balzarini, S. Liekens and F. Estévez, *Eur. J. Med. Chem.*, 2015, **101**, 205–217.
- 84 S. Gatadi, J. Gour, M. Shukla, G. Kaul, S. Das, A. Dasgupta, S. Malasala, R. S. Borra, Y. Madhavi and S. Chopra, *Eur. J. Med. Chem.*, 2018, **157**, 1056–1067.
- 85 V. Veeramreddy, T. Allaka, J. Anireddy and R. Varala, *Org. Commun.*, 2015, **8**, 98.



- 86 X.-M. Peng, K. V. Kumar, G. L. Damu and C.-H. Zhou, *Sci. China Chem.*, 2016, **59**, 878–894.
- 87 C. H. Zhou and Y. Wang, *Curr. Med. Chem.*, 2012, **19**, 239–280.
- 88 X.-M. Peng, L.-P. Peng, S. Li, S. R. Avula, V. K. Kannekanti, S.-L. Zhang, K. Y. Tam and C.-H. Zhou, *Future Med. Chem.*, 2016, **8**, 1927–1940.
- 89 N. Desai, A. M. Dodiya and P. N. Shihora, *Med. Chem. Res.*, 2012, **21**, 1577–1586.
- 90 N. Desai, H. Vaghani and P. Shihora, *J. Fluorine Chem.*, 2013, **153**, 39–47.
- 91 H. Patel, A. Shirkhedkar, S. Bari, K. Patil, A. Arambhi, C. Pardeshi, A. Kulkarni, and S. Surana, *Bulletin of Faculty of Pharmacy*, Cairo University, 2018, vol. 56, pp. 83–90.
- 92 U. Pandit and A. Dodiya, *Med. Chem. Res.*, 2013, **22**, 3364–3371.
- 93 N. K. Maddali, I. K. Viswanath, Y. Murthy, R. Bera, M. Takhi, N. S. Rao and V. Gudla, *Med. Chem. Res.*, 2019, **28**, 559–570.
- 94 D. R. Shah, R. P. Modh, D. D. Desai and K. H. Chikhalia, *Indian J. Chem., Sect. B: Org. Chem. Incl. Med. Chem.*, 2014, **53B**, 1169–1177.
- 95 Q.-S. Long, L.-W. Liu, Y.-L. Zhao, P.-Y. Wang, B. Chen, Z. Li and S. Yang, *J. Agric. Food Chem.*, 2019, **67**, 11005–11017.
- 96 Z. Fan, J. Shi, N. Luo, M. Ding and X. Bao, *J. Agric. Food Chem.*, 2019, **67**, 11598–11606.
- 97 Z. Fan, J. Shi and X. Bao, *Mol. Divers.*, 2018, **22**, 657–667.
- 98 X. Zhu, K. S. Van Horn, M. M. Barber, S. Yang, M. Z. Wang, R. Manetsch and K. A. Werbovetz, *Bioorg. Med. Chem.*, 2015, **23**, 5182–5189.
- 99 Q. Ji, D. Yang, X. Wang, C. Chen, Q. Deng, Z. Ge, L. Yuan, X. Yang and F. Liao, *Bioorg. Med. Chem.*, 2014, **22**, 3405–3413.
- 100 P. M. Bedi, V. Kumar and M. P. Mahajan, *Bioorg. Med. Chem. Lett.*, 2004, **14**, 5211–5213.
- 101 A. Kumar, P. Sharma, P. Kumari and B. L. Kalal, *Bioorg. Med. Chem. Lett.*, 2011, **21**, 4353–4357.
- 102 P. Zoumpoulakis, C. Camoutsis, G. Pairas, M. Soković, J. Glamočlija, C. Potamitis and A. Pitsas, *Bioorg. Med. Chem.*, 2012, **20**, 1569–1583.
- 103 S. M. Hashemi, H. Badali, H. Irannejad, M. Shokrzadeh and S. Emami, *Bioorg. Med. Chem.*, 2015, **23**, 1481–1491.
- 104 Y. Jiang, J. Zhang, Y. Cao, X. Chai, Y. Zou, Q. Wu, D. Zhang, Y. Jiang and Q. Sun, *Bioorg. Med. Chem. Lett.*, 2011, **21**, 4471–4475.
- 105 Y. Wang, G. L. Damu, J.-S. Lv, R.-X. Geng, D.-C. Yang and C.-H. Zhou, *Bioorg. Med. Chem. Lett.*, 2012, **22**, 5363–5366.
- 106 L. Yang, S. Ge, J. Huang and X. Bao, *Mol. Divers.*, 2018, **22**, 71–82.
- 107 V. Gupta, S. K. Kashaw, V. Jatav and P. Mishra, *Med. Chem. Res.*, 2008, **17**, 205–211.
- 108 J. Jampilek, R. Musiol, J. Finster, M. Pesko, J. Carroll, K. Kralova, M. Vejsova, J. O'mahony, A. Coffey and J. Dohnal, *Molecules*, 2009, **14**, 4246–4265.
- 109 L. Yang and X.-P. Bao, *RSC Adv.*, 2017, **7**, 34005–34011.
- 110 C. Li, J.-C. Liu, Y.-R. Li, C. Gou, M.-L. Zhang, H.-Y. Liu, X.-Z. Li, C.-J. Zheng and H.-R. Piao, *Bioorg. Med. Chem. Lett.*, 2015, **25**, 3052–3056.
- 111 A. Vijesh, A. M. Isloor, P. Shetty, S. Sundershan and H. K. Fun, *Eur. J. Med. Chem.*, 2013, **62**, 410–415.
- 112 D. Shah, H. Lakum and K. Chikhalia, *Russ. J. Bioorg. Chem.*, 2015, **41**, 209–222.
- 113 S. S. Chauhan, S. Pandey, R. Shivahare, K. Ramalingam, S. Krishna, P. Vishwakarma, M. Siddiqi, S. Gupta, N. Goyal and P. M. Chauhan, *RSC Med. Chem.*, 2015, **6**, 351–356.
- 114 M. Sharma, K. Chauhan, R. Shivahare, P. Vishwakarma, M. K. Suthar, A. Sharma, S. Gupta, J. K. Saxena, J. Lal and P. Chandra, *J. Med. Chem.*, 2013, **56**, 4374–4392.
- 115 V. Gurram, R. Garlapati, C. Thulluri, N. Madala, K. S. Kasani, P. K. Machiraju, R. Doddapalla, U. Addepally, R. Gundla and B. Patro, *Med. Chem. Res.*, 2015, **24**, 2227–2237.
- 116 K. Javaid, S. M. Saad, S. Rasheed, S. T. Moin, N. Syed, I. Fatima, U. Salar, K. M. Khan, S. Perveen and M. I. Choudhary, *Bioorg. Med. Chem.*, 2015, **23**, 7417–7421.
- 117 M. Wei, W.-M. Chai, R. Wang, Q. Yang, Z. Deng and Y. Peng, *Bioorg. Med. Chem.*, 2017, **25**, 1303–1308.
- 118 F. Jabeen, S. A. Shehzadi, M. Q. Fatmi, S. Shaheen, L. Iqbal, N. Afza, S. S. Panda and F. L. Ansari, *Bioorg. Med. Chem. Lett.*, 2016, **26**, 1029–1038.
- 119 Y. Chinthala, A. K. Domatti, A. Sarfaraz, S. P. Singh, N. K. Arigari, N. Gupta, S. K. Satya, J. K. Kumar, F. Khan and A. K. Tiwari, *Eur. J. Med. Chem.*, 2013, **70**, 308–314.
- 120 S. Iqbal, M. A. Khan, K. Javaid, R. Sadiq, S. Fazal-Ur-Rehman, M. I. Choudhary and F. Z. Basha, *Bioorg. Chem.*, 2017, **74**, 72–81.
- 121 M. Saeedi, M. Mohammadi-Khanaposhtani, P. Pourrabia, N. Razzaghi, R. Ghadimi, S. Imanparast, M. A. Faramarzi, F. Bandarian, E. N. Esfahani and M. Safavi, *Bioorg. Chem.*, 2019, **83**, 161–169.
- 122 S. Jangam and S. Wankhede, *Russ. J. Gen. Chem.*, 2019, **89**, 1029–1041.
- 123 T. S. Patel, S. F. Vanparia, U. H. Patel, R. B. Dixit, C. J. Chudasama, B. D. Patel and B. C. Dixit, *Eur. J. Med. Chem.*, 2017, **129**, 251–265.
- 124 T. S. Patel, J. D. Bhatt, R. B. Dixit, C. J. Chudasama, B. D. Patel and B. C. Dixit, *Arch. Pharmazie*, 2019, **352**, 1900099.
- 125 T. S. Patel, J. D. Bhatt, S. F. Vanparia, U. H. Patel, R. B. Dixit, C. J. Chudasama, B. D. Patel and B. C. Dixit, *Bioorg. Med. Chem.*, 2017, **25**, 6635–6646.
- 126 T. S. Patel, J. D. Bhatt, R. B. Dixit, C. J. Chudasama, B. D. Patel and B. C. Dixit, *Bioorg. Med. Chem.*, 2019, **27**, 3574–3586.
- 127 S. Malik, R. S. Bahare and S. A. Khan, *Eur. J. Med. Chem.*, 2013, **67**, 1–13.
- 128 S. S. Jangam, S. B. Wankhede and S. S. Chitlange, *Res. Chem. Intermed.*, 2019, **45**, 471–486.
- 129 T. Fröhlich, C. Reiter, M. M. Ibrahim, J. Beutel, C. Hutterer, I. Zeiträger, H. Bahsi, M. Leidenberger, O. Friedrich and B. Kappes, *ACS Omega*, 2017, **2**, 2422–2431.
- 130 E. Giovannucci, D. M. Harlan, M. C. Archer, R. M. Bergenstal, S. M. Gapstur, L. A. Habel, M. Pollak,



- J. G. Regensteiner and D. Yee, *Ca-Cancer J. Clin.*, 2010, **60**, 207–221.
- 131 M.-Y. Lee, K.-D. Lin, P.-J. Hsiao and S.-J. Shin, *Metabolism*, 2012, **61**, 242–249.
- 132 Y. Zhang, H. Gao, R. Liu, J. Liu, L. Chen, X. Li, L. Zhao, W. Wang and B. Li, *Bioorg. Med. Chem. Lett.*, 2017, **27**, 4309–4313.
- 133 R. P. Modh, E. De Clercq, C. Pannecouque and K. H. Chikhalia, *J. Enzym. Inhib. Med. Chem.*, 2014, **29**, 100–108.
- 134 G. D. Goss, E. Felip, M. Cobo, S. Lu, K. Syrigos, K. H. Lee, E. Göker, V. Georgoulas, W. Li, S. Guclu, D. Isla, Y. J. Min, A. Morabito, A. Ardizzoni, S. M. Gadgeel, A. Fölöp, C. Böhnemann, N. Gibson, N. Krämer, F. Solca, A. Cseh, E. Ehrnrooth and J. C. Soria, *JAMA Oncol.*, 2018, **4**, 1189–1197.
- 135 A. Drilon, S. Fu, M. R. Patel, M. Fakih, D. Wang, A. J. Olszanski, D. Morgensztern, S. V. Liu, B. C. Cho, L. Bazhenova, C. P. Rodriguez, R. C. Doebele, A. Wozniak, K. L. Reckamp, T. Seery, P. Nikolinakos, Z. Hu, J. W. Oliver, D. Trone, K. McArthur, R. Patel, P. S. Multani and M. J. Ahn, *Cancer Discovery*, 2019, **9**, 384–395.
- 136 J. Qu, Y. N. Wang, P. Xu, D. X. Xiang, R. Yang, W. Wei and Q. Qu, *Oncotarget*, 2017, **8**, 33961–33971.
- 137 T. S. Mok, Y. Cheng, X. Zhou, K. H. Lee, K. Nakagawa, S. Niho, M. Lee, R. Linke, R. Rosell, J. Corral, M. R. Migliorino, A. Pluzanski, E. I. Sbar, T. Wang, J. L. White and Y. L. Wu, *J. Clin. Oncol.*, 2018, **36**, 2244–2250.
- 138 S. Pallesen, H. S. Hamre, N. Lang and B. Bjorvatn, *SAGE Open Med. Case Rep.*, 2020, **8**, 2050313X2093607.
- 139 P. Lischka, D. Michel and H. Zimmermann, *J. Infect. Dis.*, 2016, **213**, 23–30.

