

Cite this: *Chem. Sci.*, 2019, 10, 1873

All publication charges for this article have been paid for by the Royal Society of Chemistry

# Fluorogenic hydrogen sulfide (H<sub>2</sub>S) donors based on sulfenyl thiocarbonates enable H<sub>2</sub>S tracking and quantification†

Yu Zhao,  Matthew M. Cerda and Michael D. Pluth \*

Hydrogen sulfide (H<sub>2</sub>S) is an important cellular signaling molecule that exhibits promising protective effects. Although a number of triggerable H<sub>2</sub>S donors have been developed, spatiotemporal feedback from H<sub>2</sub>S release in biological systems remains a key challenge in H<sub>2</sub>S donor development. Herein we report the synthesis, evaluation, and application of caged sulfenyl thiocarbonates as new fluorescent H<sub>2</sub>S donors. These molecules rely on thiol cleavage of sulfenyl thiocarbonates to release carbonyl sulfide (COS), which is quickly converted to H<sub>2</sub>S by carbonic anhydrase (CA). This approach is a new strategy in H<sub>2</sub>S release and does not release electrophilic byproducts common from COS-based H<sub>2</sub>S releasing motifs. Importantly, the release of COS/H<sub>2</sub>S is accompanied by the release of a fluorescent reporter, which enables the real-time tracking of H<sub>2</sub>S by fluorescence spectroscopy or microscopy. Dependent on the choice of fluorophore, either one or two equivalents of H<sub>2</sub>S can be released, thus allowing for the dynamic range of the fluorescent donors to be tuned. We demonstrate that the fluorescence response correlates directly with quantified H<sub>2</sub>S release and also demonstrate the live-cell compatibility of these donors. Furthermore, these fluorescent donors exhibit anti-inflammatory effects in RAW 264.7 cells, indicating their potential application as new H<sub>2</sub>S-releasing therapeutics. Taken together, sulfenyl thiocarbonates provide a new platform for H<sub>2</sub>S donation and readily enable fluorescent tracking of H<sub>2</sub>S delivery in complex environments.

Received 21st November 2018  
Accepted 10th December 2018

DOI: 10.1039/c8sc05200j

rsc.li/chemical-science

## Introduction

Hydrogen sulfide (H<sub>2</sub>S) is an important gaseous molecule that plays critical roles in living systems.<sup>1–3</sup> Endogenous H<sub>2</sub>S is primarily produced from cysteine (Cys) and homocysteine (Hcy) by four main enzymes, including cystathionine β-synthase (CBS), cystathionine γ-lyase (CSE), 3-mercaptopyruvate sulfur transferase (3-MST), and cysteine aminotransferase (CAT).<sup>4–8</sup> Due to its bioregulatory and protective roles, H<sub>2</sub>S is considered as an important cellular signaling molecule, much like nitric oxide (NO) and carbon monoxide (CO).<sup>8–11</sup>

Although many H<sub>2</sub>S-related biological functions have been discovered in the past two decades, many investigations have been limited due to the lack of controllable and refined H<sub>2</sub>S delivery systems (H<sub>2</sub>S donors). Inorganic sulfide salts, such as sodium sulfide (Na<sub>2</sub>S) and sodium hydrosulfide (NaHS), are widely used in H<sub>2</sub>S investigations, but they release H<sub>2</sub>S quickly in aqueous media, thus failing to mimic the slow and well-regulated *in vivo* H<sub>2</sub>S generation.<sup>12</sup> By contrast, the H<sub>2</sub>S release

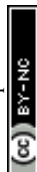
from GYY4137, a phosphorodithioate-based H<sub>2</sub>S donor, is slow in aqueous buffer, and the low H<sub>2</sub>S releasing efficiency remains as a major concern when applying it to the living systems.<sup>13</sup> Building from these donor scaffolds, a series of synthetic H<sub>2</sub>S donors have been developed in the last decade.<sup>14–19</sup> These donors are activated by different triggers, such as enzymes,<sup>20,21</sup> cellular thiols,<sup>22–27</sup> pH modulation,<sup>28,29</sup> and photo activation,<sup>30–33</sup> and the released H<sub>2</sub>S exhibits promising activities in different physiological and pathological processes.<sup>17,18</sup>

To expand the library of H<sub>2</sub>S donors, we recently developed the carbonyl sulfide (COS)-based platform for H<sub>2</sub>S donation. In this approach, caged-thiocarbamates are activated and the resultant intermediates undergo a cascade reaction to release COS, which is quickly converted to H<sub>2</sub>S by the ubiquitous enzyme carbonic anhydrase (CA) (Fig. 1a).<sup>34</sup> Expanding on this initial report, we, as well as others, have applied the similar caged-COS systems to include a series of triggerable COS-based H<sub>2</sub>S donors which can be activated by different mechanisms, such as reactive oxygen species (ROS),<sup>35–37</sup> esterase,<sup>38,39</sup> cellular nucleophiles,<sup>40</sup> click chemistry,<sup>41</sup> light,<sup>42–44</sup> and Cys,<sup>45</sup> to release H<sub>2</sub>S under various conditions.<sup>45,46</sup>

Although H<sub>2</sub>S release from these donors has been measured using different methods, the real-time tracking of donor activation and H<sub>2</sub>S delivery in living systems remains a key challenge due to the inherent limitations of current H<sub>2</sub>S detection

Department of Chemistry and Biochemistry, Institute of Molecular Biology, Materials Science Institute, University of Oregon, Eugene, OR 97403, USA. E-mail: pluth@uoregon.edu

† Electronic supplementary information (ESI) available: H<sub>2</sub>S release curves, cytotoxicity data, NMR spectra. See DOI: 10.1039/c8sc05200j



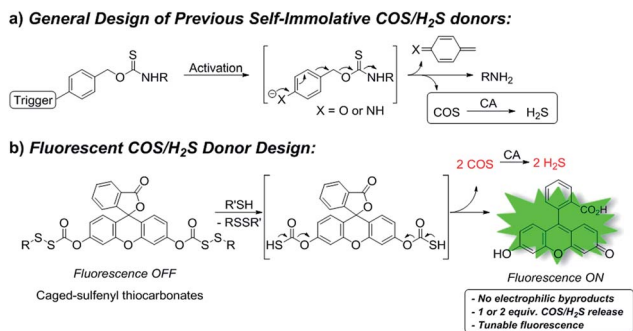


Fig. 1 (a) General design of previous self-immolative COS/H<sub>2</sub>S donors. (b) COS/H<sub>2</sub>S release from fluorogenic caged-sulfenyl thiocarbonate donor motifs.

methods. For example, the colorimetric methylene blue (MB) assay has been widely used to measure H<sub>2</sub>S levels, but requires strongly acidic conditions, which may trigger H<sub>2</sub>S release from acid labile sulfide pools. Additionally, MB is an end-point assay which destroys experimental samples, thus making it not feasible for *in vivo* H<sub>2</sub>S detection.<sup>47</sup> Similarly, H<sub>2</sub>S-selective electrodes are most often used in bulk measurements rather than non-homogenized biological samples.<sup>47</sup> By contrast, H<sub>2</sub>S fluorescent probes have attracted much attentions due to their high sensitivities and have been used to sense and visualize H<sub>2</sub>S in biological samples.<sup>48</sup> Albeit promising, H<sub>2</sub>S fluorescent probes are prone to react with reactive cellular species, such as Cys or glutathione (GSH), which results in either probe consumption or false positive signals. In addition, all of the above methods consume the H<sub>2</sub>S being measured. Recently, we reported a series of  $\gamma$ -ketothiocarbamates ( $\gamma$ -KetoTCM) that release *p*-nitroaniline as a colorimetric indicator; however, although the colorimetric response enables monitoring of COS/H<sub>2</sub>S release in cuvettes, the direct tracking in live-cell environments remains an un-met need.<sup>49</sup> Taken together, a key advancement compatible with live-cell and tissue experiments would be the development of H<sub>2</sub>S donors that deliver H<sub>2</sub>S with a concomitant fluorescence response to enable tracking of H<sub>2</sub>S delivery by common microscopy techniques.

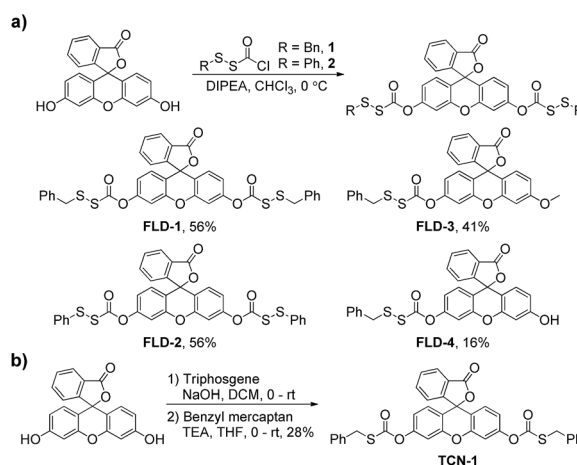
Aligned with these needs, we report here the development of a new H<sub>2</sub>S-releasing strategy based on caged sulfenyl thiocarbonates and apply this donor platform to access fluorescent turn-on H<sub>2</sub>S donors. In this approach, cellular thiols (*i.e.* Cys and GSH) activate the sulfenyl thiocarbonates through thiol-mediated disulfide reduction to release COS, which is quickly converted to H<sub>2</sub>S by CA (Fig. 1b). To the best of our knowledge, the sulfenyl thiocarbonate reduction strategy provides a new activation pathway that has not been used to trigger COS/H<sub>2</sub>S release from donor platforms. Unlike currently-available donors that function through initial COS release, the sulfenyl thiocarbonate system does not generate reactive electrophile byproducts upon activation, which provides a significant advance in the field. In addition, we leverage this new donor strategy to access systems in which a concomitant fluorescence turn-on occurs upon donor activation, thus allowing for real-time monitoring and quantification of H<sub>2</sub>S release using fluorescence spectroscopy.

## Results and discussion

To test our hypothesis that caged-sulfenyl thiocarbonates could serve as thiol-triggered fluorescent COS/H<sub>2</sub>S donors, four donors (**FLD-1-4**) were prepared by reacting fluorescein or 3-O-methylfluorescein with ((benzyl)dithio)carbonyl chloride **1** or ((phenol)dithio)carbonyl chloride **2** in the presence of DIPEA (Scheme 1a). A caged thiocarbonate (**TCN-1**) was synthesized as a triggerless control compound, which is expected to be stable toward thiol activation due to the lack of a disulfide functional group (Scheme 1b).

With these compounds in hand, we first evaluated their spectroscopic properties in PBS buffer (pH 7.4, 10 mM). As expected, **FLD-1-3** and **TCN-1** are not absorptive in the visible region and are all nonfluorescent because the fluorescein unit is locked in the closed lactone form. By contrast, **FLD-4** shows a prominent absorbance band in the visible region ( $\lambda_{\text{max}} = 449 \text{ nm}$ ,  $\epsilon = 27\,300 \pm 2500 \text{ M}^{-1} \text{ cm}^{-1}$ ) with measurable fluorescence ( $\lambda_{\text{em}} = 514 \text{ nm}$ ,  $\Phi = 0.11 \pm 0.01$ ) due to the free hydroxyl group (Table S1†). Based on the promising spectroscopic properties, large dynamic range, and efficient release of two equivalents of COS/H<sub>2</sub>S, we chose to use **FLD-1** as the model donor for initial reactivity and selectivity evaluations.

To test the reactivity of **FLD-1** towards Cys-induced activation, **FLD-1** (10  $\mu\text{M}$ ) was incubated with Cys (100  $\mu\text{M}$ ) in PBS buffer (pH 7.4, 10 mM) containing physiologically-relevant concentrations of CA (25  $\mu\text{g mL}^{-1}$ ), and the fluorescence intensity was measured using a fluorescence spectrometer. As expected, Cys successfully activated **FLD-1** and resulted in a 500-fold fluorescence turn on over 2 h, demonstrating the release of the fluorescein upon **FLD-1** activation (Fig. 2a). Fluorescein formation was also confirmed by UV-vis spectroscopy under the identical conditions (Fig. S1†). A Cys-dependent fluorescence enhancement was observed when treating **FLD-1** (10  $\mu\text{M}$ ) with increasing concentrations of Cys (1–20 equiv.), indicating a high sensitivity of **FLD-1** towards Cys. No fluorescent signal was observed in the absence of Cys, suggesting that **FLD-1** is stable in aqueous buffer, and that it is not hydrolyzed to provide false



Scheme 1 Synthesis of fluorescent COS/H<sub>2</sub>S donors (a) and thiocarbonate control compound (b).



signals (Fig. 2b). In addition, to confirm the H<sub>2</sub>S delivery from **FLD-1**, we treated **FLD-1** (10 μM) with Cys (100 μM) in PBS containing CA (25 μg mL<sup>-1</sup>) and quantified H<sub>2</sub>S release using the MB assay. In this experiment, 15 μM of H<sub>2</sub>S was detected (75% releasing efficiency), which is consistent with our hypothesis that 2 equivalents of COS/H<sub>2</sub>S would be released upon **FLD-1** activation (Fig. S2†). Taken together, these experiments demonstrate that **FLD-1** is highly responsive to Cys activation.

To determine whether H<sub>2</sub>S release correlated directly with the observed fluorescence response, we measured the fluorescent response from **FLD-3** in the presence of Cys and CA and quantified H<sub>2</sub>S release using the MB assay. We chose to use **FLD-3** as the model compound for these investigations because it only contains one sulfenyl thiocarbonate moiety, and therefore should simplify the reaction kinetics. In comparison, **FLD-1** contains two sulfenyl thiocarbonate groups, and the cleavage of one sulfenyl thiocarbonate would generate **FLD-4** as a reaction intermediate, which exhibits moderate fluorescence (see Fig. 4). Incubation of **FLD-3** (10 μM) with Cys (100 μM) resulted in a rapid fluorescence response with 96% of the H<sub>2</sub>S release measured by MB. At extended time points, we observed a slight decrease in measured H<sub>2</sub>S, possibly due to volatilization of H<sub>2</sub>S in the headspace of the closed system or adventitious oxidation of released H<sub>2</sub>S. Negligible H<sub>2</sub>S was detected in the absence of CA, indicating that Cys-triggered H<sub>2</sub>S delivery from **FLD-3** proceeds through intermediate COS formation (Fig. 3a). Importantly, the strong linear correlation between the measured fluorescence and H<sub>2</sub>S measured from the MB method detection (first 25 min, R<sup>2</sup> = 0.988) demonstrates that fluorescent readouts can serve as reliable optical tools to track COS/H<sub>2</sub>S release from **FLD** donors with temporal resolution (Fig. 3b). Moreover, this linear correlation suggests that choice of other fluorophores with different brightnesses and photophysical properties could be used to access different dynamic ranges of H<sub>2</sub>S release, thus enabling this approach to be translated to different types of experimental designs.

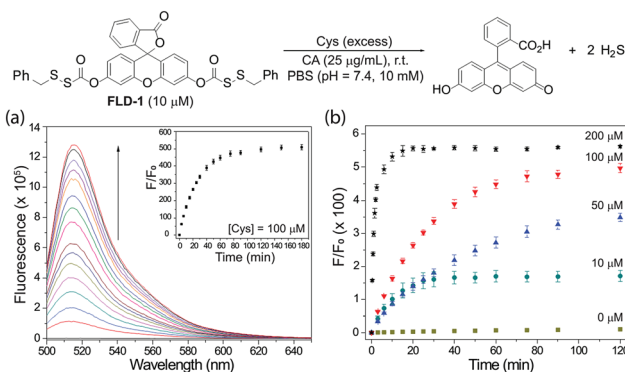


Fig. 2 (a) Time-dependent fluorescence spectra of **FLD-1** (10 μM) in PBS (pH 7.4, 10 mM) containing Cys (100 μM) and CA (25 μg mL<sup>-1</sup>). (b) Cys-dependent (0–200 μM) fluorescence turn on of **FLD-1** (10 μM) in PBS. λ<sub>ex</sub> = 490 nm, λ<sub>em</sub> = 500–650 nm, and slit width = 0.3 nm. The experiments were performed in triplicate and results are expressed as mean ± SD (n = 3).

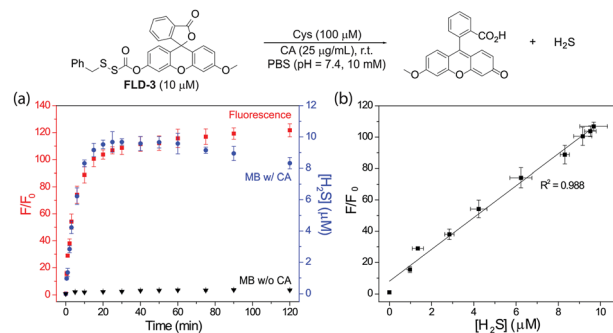


Fig. 3 (a) Time-dependent fluorescence turn on (red) and H<sub>2</sub>S release (blue) upon **FLD-3** (10 μM) activation in PBS (pH 7.4, 10 mM) containing Cys (100 μM) and CA (25 μg mL<sup>-1</sup>). No H<sub>2</sub>S was detected in the absence of CA (black). λ<sub>ex</sub> = 454 nm, λ<sub>em</sub> = 500–650 nm, and slit width = 0.3 nm. (b) Correlation between fluorescence measurement and MB detection. The experiments were performed in triplicate. The results are expressed as mean ± SD (n = 3).

Having confirmed the efficiency of Cys-triggered fluorescence turn on and COS/H<sub>2</sub>S release, we next evaluated the reactivity of other **FLD** donors towards Cys activation. Treating **FLD-1–3** (10 μM) with Cys (100 μM) in PBS buffer (pH 7.4, 10 mM) resulted in a 120–500-fold fluorescence turn on over 2 h. We attribute the faster response of **FLD-2** to the more electrophilic phenyl sulfenyl thiocarbonate in comparison to the less electrophilic benzyl sulfenyl thiocarbonate in **FLD-1**. **FLD-4**, however, provided minimal fluorescence enhancement due to its strong background fluorescence. No fluorescence response from **TCN-1** (10 μM) was observed under the identical conditions, which demonstrates the stability of the thiocarbonate group in the presence of Cys (Fig. 4).

To further support our proposed activation mechanism in Fig. 1b, as well as the activation of the sulfenyl thiocarbonate group by other thiols, we incubated **FLD-1** (10 μM) in PBS

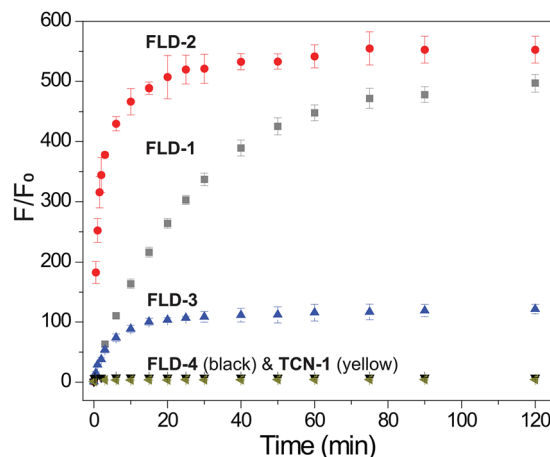


Fig. 4 Fluorescence turn on of **FLD-1** (gray), **FLD-2** (red), **FLD-3** (blue), **FLD-4** (black), and **TCN-1** (dark yellow) (10 μM) in PBS (pH 7.4, 10 mM) containing Cys (100 μM). λ<sub>ex</sub> = 490 nm for **FLD-1**, 2, 4, and **TCN-1**, λ<sub>ex</sub> = 454 nm for **FLD-3**, λ<sub>em</sub> = 500–650 nm, and slit width = 0.3 nm. The experiments were performed in triplicate. The results are expressed as mean ± SD (n = 3).



(pH 7.4, 10 mM) with 10 equivalents of benzyl mercaptan (100  $\mu\text{M}$ ) for 1 h and analyzed the reaction products by HPLC (Fig. S3†). Consistent with our proposed activation mechanism, we observed **FLD-1** consumption and the formation of both benzyl disulfide and fluorescein, which supports the proposed mechanism.

In addition to Cys and BnSH, other cellular thiols and biological nucleophiles were tested to determine whether they resulted in donor activation. In these experiments, **FLD-1** (10  $\mu\text{M}$ ) was incubated with GSH, homocysteine (Hcy), *N*-acetyl cysteine (NAC), penicillamine (PEN), or bovine serum albumin (BSA) (100  $\mu\text{M}$ ), in PBS buffer (pH 7.4, 10 mM) containing CA (25  $\mu\text{g mL}^{-1}$ ) and the fluorescence intensity was measured after 2 h. As expected, **FLD-1** is stable in PBS at physiological pH in the absence of thiols (Fig. 5 bar 1). Incubation of **FLD-1** with Cys, NAC, GSH, and Hcy, however, led to a significant fluorescence enhancement, indicating successful donor activation and COS/H<sub>2</sub>S release (Fig. 5 bars 2–5 and S4†). In addition, these results demonstrate that the sulfenyl thiocarbonate group is responsive to different types of thiols. In comparison, PEN resulted in only minimal fluorescence response and BSA failed to activate the donor presumably due to the bulkiness of these two thiol species, which hindered their reactions with **FLD-1** (Fig. 5 bars 6 and 7). Additionally, *N*-ethylmaleimide (NEM) pretreatment of Cys samples significantly reduced the fluorescence enhancement from **FLD-1**, confirming the necessity of the thiol-induced reduction for the donor activation (Fig. 5 bar 8).

We also tested the response of the **FLD** donors in the presence of other cellular reactive sulfur, oxygen, and nitrogen species (RSONs). **FLD-1** (10  $\mu\text{M}$ ) was incubated with RSONs (100  $\mu\text{M}$ ), such as hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>), hyperchlorite (ClO<sup>-</sup>), superoxide (O<sub>2</sub><sup>-</sup>), *tert*-butyl hydroperoxide (TBHP), serine (Ser), lysine (Lys), glycine (Gly), oxidized glutathione (GSSG), and *S*-nitrosoglutathione (GSNO), and COS/H<sub>2</sub>S release was monitored by tracking the fluorescein formation. As expected, minimal fluorescence was observed in these

experiments, confirming the stability of **FLD-1** to common RSONs (Fig. 5 bars 9–17). Because the carbonate functional group may be sensitive to esterase-catalyzed hydrolysis, we also tested the esterase stability of the sulfenyl thiocarbonate group by incubating **FLD-1** (10  $\mu\text{M}$ ) with porcine liver esterase (PLE, 1 U mL<sup>-1</sup>). Although a slight fluorescence turn on was observed after a 2 h incubation, the observed response was much lower than that from thiol activation (Fig. 5 bar 18). Taken together, these studies demonstrate that **FLD-1** is highly responsive and selective to thiol activation and common cellular RSONs do not trigger **FLD-1** to release COS/H<sub>2</sub>S.

After confirming the thiol-triggered COS/H<sub>2</sub>S release from **FLD-1** in aqueous buffer, we next investigated the H<sub>2</sub>S delivering capacity of **FLD-1** in cellular environment. In these experiments, HeLa cells were treated with **FLD-1** (50  $\mu\text{M}$ ) or **TCN-1** (50  $\mu\text{M}$ ) and H<sub>2</sub>S release was monitored using C7-Az, a H<sub>2</sub>S-responsive fluorescent probe.<sup>50,51</sup> Incubation of HeLa cells with C7-Az (50  $\mu\text{M}$ ) alone resulted in a negligible fluorescence response, indicating minimal endogenous H<sub>2</sub>S (Fig. 6 top row). Treatment with **TCN-1** also failed to provide a fluorescence signal, suggesting that the thiocarbonate group was stable and did not decompose to generate false signals in cellular environments (Fig. 6 middle row). By contrast, a strong C7-Az fluorescent signal was observed when incubating HeLa cells with **FLD-1**, suggesting that H<sub>2</sub>S release was successfully triggered by endogenous thiols. In addition, a strong fluorescence signal was also observed from activated **FLD-1** in the fluorescein channel, confirming the fluorescence response upon donor activation (Fig. 6 bottom row). Taken together, these results demonstrate that the **FLD** donors not only function as efficacious H<sub>2</sub>S donors in live cells, but also provide a fluorescence signal that enables observing H<sub>2</sub>S release.

To further demonstrate the H<sub>2</sub>S-releasing fidelity of the **FLD** donors we also investigated the anti-inflammatory activities of **FLD-1**. We pretreated macrophage RAW 264.7 cells with **FLD-1** (0–25  $\mu\text{M}$ ) for 2 h, followed by a 24 h incubation with lipopolysaccharide (LPS, 0.5  $\mu\text{g mL}^{-1}$ ) to trigger the inflammatory

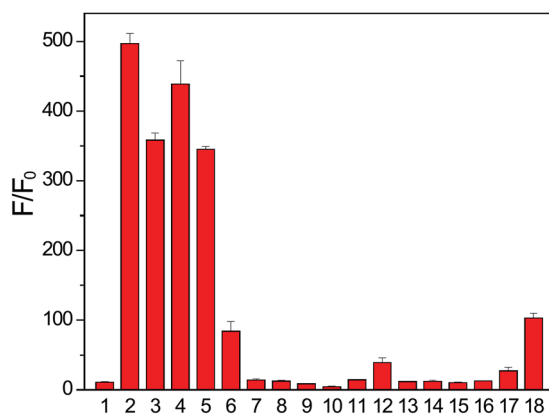


Fig. 5 Fluorescence turn on of **FLD-1** (10  $\mu\text{M}$ ) in the presence of cellular RSONs (100  $\mu\text{M}$ ): (1) **FLD-1** only, (2) Cys, (3) NAC, (4) GSH, (5) Hcy, (6) PEN, (7) BSA, (8) Cys + NEM, (9) H<sub>2</sub>O<sub>2</sub>, (10) ClO<sup>-</sup>, (11) O<sub>2</sub><sup>-</sup>, (12) TBHP, (13) Ser, (14) Lys, (15) Gly, (16) GSSG, (17) GSNO, and (18) PLE (1 U mL<sup>-1</sup>). Fluorescence intensity was measured after 2 h incubation. The experiments were performed in triplicate. The results were expressed as mean  $\pm$  SD ( $n = 3$ ).

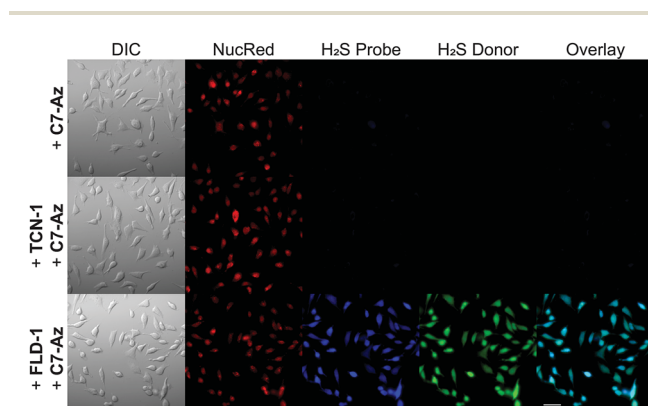


Fig. 6 H<sub>2</sub>S delivery from **FLD-1** in HeLa cells. HeLa cells were treated with NucRed nuclear dye and C7-Az (50  $\mu\text{M}$ ) in DMEM or DMEM containing **TCN-1** (50  $\mu\text{M}$ ) (middle row) or **FLD-1** (50  $\mu\text{M}$ ) (bottom row) for 30 min. Cells were then washed with PBS and cell images were taken in PBS using a fluorescent microscope. Bar scale: 50  $\mu\text{m}$ .





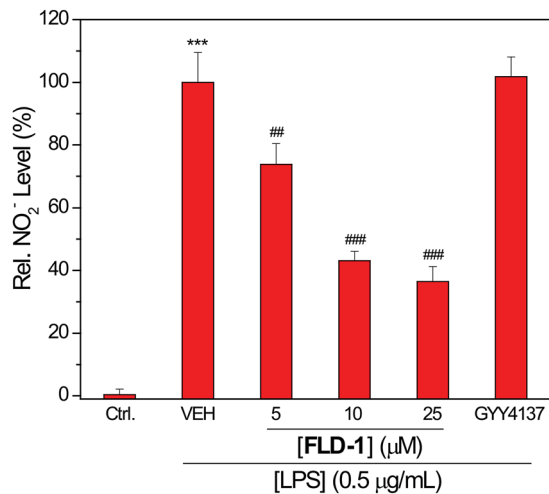


Fig. 7 Effects of FLD-1 on LPS-induced NO<sub>2</sub><sup>-</sup> accumulation. RAW 264.7 cells were pretreated with FLD-1 (5–25 μM) or GYY4137 (25 μM) for 2 h, followed by a 24 h treatment of LPS (0.5 μg mL<sup>-1</sup>). Results are expressed as mean ± SD (n = 4). \*\*\*P < 0.001 vs. the control group; ##P < 0.01 vs. vehicle-treated group; ###P < 0.001 vs. vehicle-treated group.

response. The inflammation event usually results in the NO generation, which can be monitored by measuring nitrite (NO<sub>2</sub><sup>-</sup>) accumulation. We chose to use concentrations of FLD-1 up to 25 μM because these concentrations did not induce cytotoxicity (Fig. S6†).

As expected, the pretreatment of RAW 264.7 cells with FLD-1 showed a dose-dependent inhibition of NO<sub>2</sub><sup>-</sup> accumulation, indicating anti-inflammatory activity from FLD-1. Although GYY4137 has shown anti-inflammatory effects at higher concentration and longer incubation time (*i.e.* 100–1000 μM and 24 h incubation),<sup>52</sup> such cytoprotection was not observed at the 25 μM concentration used for comparison, highlighting the efficacious H<sub>2</sub>S release from FLD-1 in the cellular environment (Fig. 7). To further confirm that the observed effects were due to H<sub>2</sub>S rather than other components of donor activation, we treated cells with 25 μM of TCN-1, fluorescein or benzyl mercaptan and measured NO<sub>2</sub><sup>-</sup> production. None of these compounds attenuated NO<sub>2</sub><sup>-</sup> generation, confirming that the anti-inflammatory activities of FLD-1 is due to H<sub>2</sub>S release (Fig. S7†). Overall, these studies demonstrate that FLD-1 releases COS/H<sub>2</sub>S in complex cellular environment and exhibits promising anti-inflammatory protections, indicating potential applications of FLD-1 as H<sub>2</sub>S-releasing therapeutics.

## Conclusions

In conclusion, we prepared and evaluated a class of caged sulfenyl thiocarbonates as new controllable and fluorescent COS/H<sub>2</sub>S donors. Thiol-triggered COS/H<sub>2</sub>S release from these molecules has been detected and visualized in both aqueous buffer and in live cells. Importantly, the concomitant release of a fluorescein reporter after H<sub>2</sub>S release enables the real-time monitoring of H<sub>2</sub>S release dynamics. In addition, we

demonstrate that FLD-1 exhibits a dose-dependent inhibition of the LPS-induced NO formation, which is consistent with anti-inflammatory activities of H<sub>2</sub>S. Taken together, caged-sulfenyl thiocarbonates are promising new class of COS/H<sub>2</sub>S donors with high potential for accessing H<sub>2</sub>S-related protective activities, and the developed fluorescent donors provide new methods for monitoring H<sub>2</sub>S release in real-time in complex environments.

## Conflicts of interest

There are no conflicts to declare.

## Acknowledgements

Research reported in this publication was supported by the NIH (R01GM113030) and Dreyfus Foundation. NMR, fluorescence microscopy, and MS instrumentation in the UO CAMCOR facility is supported by the NSF (CHE-1427987, CHE-1531189, and CHE-1625529).

## Notes and references

- 1 C. Szabo, *Nat. Rev. Drug Discovery*, 2007, **6**, 917–935.
- 2 R. Wang, *Physiol. Rev.*, 2012, **92**, 791–896.
- 3 M. R. Filipovic, J. Zivanovic, B. Alvarez and R. Banerjee, *Chem. Rev.*, 2018, **118**, 1253–1337.
- 4 O. Kabil and R. Banerjee, *Antioxid. Redox Signaling*, 2014, **20**, 770–782.
- 5 S. Bruce King, *Free Radical Biol. Med.*, 2013, **55**, 1–7.
- 6 L. F. Hu, M. Lu, P. T. Hon Wong and J. S. Bian, *Antioxid. Redox Signaling*, 2011, **15**, 405–419.
- 7 B. D. Paul and S. H. Snyder, *Nat. Rev. Mol. Cell Biol.*, 2012, **13**, 499–507.
- 8 J. M. Fukuto, S. J. Carrington, D. J. Tantillo, J. G. Harrison, L. J. Ignarro, B. A. Freeman, A. Chen and D. A. Wink, *Chem. Res. Toxicol.*, 2012, **25**, 769–793.
- 9 K. R. Olson, *Antioxid. Redox Signaling*, 2012, **17**, 32–44.
- 10 C. Szabo, *Antioxid. Redox Signaling*, 2012, **17**, 68–80.
- 11 L. Li, P. Rose and P. K. Moore, *Annu. Rev. Pharmacol. Toxicol.*, 2011, **51**, 169–187.
- 12 E. R. DeLeon, G. F. Stoy and K. R. Olson, *Anal. Biochem.*, 2012, **421**, 203–207.
- 13 L. Li, M. Whiteman, Y. Y. Guan, K. L. Neo, Y. Cheng, S. W. Lee, Y. Zhao, R. Baskar, C. H. Tan and P. K. Moore, *Circulation*, 2008, **117**, 2351–2360.
- 14 M. D. Pluth, T. S. Bailey, M. D. Hammers, M. D. Hartle, H. A. Henthorn and A. K. Steiger, *Synlett*, 2015, **26**, 2633–2643.
- 15 C. Szabo and A. Papapetropoulos, *Pharmacol. Rev.*, 2017, **69**, 497–564.
- 16 M. D. Hartle and M. D. Pluth, *Chem. Soc. Rev.*, 2016, **45**, 6108–6117.
- 17 Y. Zhao, T. D. Biggs and M. Xian, *Chem. Commun.*, 2014, **50**, 11788–11805.
- 18 Y. Zhao, A. Pacheco and M. Xian, *Handb. Exp. Pharmacol.*, 2015, **230**, 365–388.



- 19 C. R. Powell, K. M. Dillon and J. B. Matson, *Biochem. Pharmacol.*, 2018, **149**, 110–123.
- 20 P. Shukla, V. S. Khodade, M. SharathChandra, P. Chauhan, S. Mishra, S. Siddaramappa, B. E. Pradeep, A. Singh and H. Chakrapani, *Chem. Sci.*, 2017, **8**, 4967–4972.
- 21 Y. Zheng, B. Yu, K. Ji, Z. Pan, V. Chittavong and B. Wang, *Angew. Chem., Int. Ed.*, 2016, **55**, 4514–4518.
- 22 J. C. Foster, C. R. Powell, S. C. Radzinski and J. B. Matson, *Org. Lett.*, 2014, **16**, 1558–1561.
- 23 Y. Zhao, S. Bhushan, C. Yang, H. Otsuka, J. D. Stein, A. Pacheco, B. Peng, N. O. Devarie-Baez, H. C. Aguilar, D. J. Lefer and M. Xian, *ACS Chem. Biol.*, 2013, **8**, 1283–1290.
- 24 Y. Zhao, H. Wang and M. Xian, *J. Am. Chem. Soc.*, 2011, **133**, 15–17.
- 25 Y. Zhao, C. Yang, C. Organ, Z. Li, S. Bhushan, H. Otsuka, A. Pacheco, J. Kang, H. C. Aguilar, D. J. Lefer and M. Xian, *J. Med. Chem.*, 2015, **58**, 7501–7511.
- 26 M. M. Cerda, M. D. Hammers, M. S. Earp, L. N. Zakharov and M. D. Pluth, *Org. Lett.*, 2017, **19**, 2314–2317.
- 27 Y. Zhao, J. Kang, C. M. Park, P. E. Bagdon, B. Peng and M. Xian, *Org. Lett.*, 2014, **16**, 4536–4539.
- 28 J. Kang, Z. Li, C. L. Organ, C. M. Park, C. T. Yang, A. Pacheco, D. Wang, D. J. Lefer and M. Xian, *J. Am. Chem. Soc.*, 2016, **138**, 6336–6339.
- 29 J. Wu, Y. Li, C. He, J. Kang, J. Ye, Z. Xiao, J. Zhu, A. Chen, S. Feng, X. Li, J. Xiao, M. Xian and Q. Wang, *ACS Appl. Mater. Interfaces*, 2016, **8**, 27474–27481.
- 30 N. O. Devarie-Baez, P. E. Bagdon, B. Peng, Y. Zhao, C. M. Park and M. Xian, *Org. Lett.*, 2013, **15**, 2786–2789.
- 31 N. Fukushima, N. Ieda, K. Sasakura, T. Nagano, K. Hanaoka, T. Suzuki, N. Miyata and H. Nakagawa, *Chem. Commun.*, 2014, **50**, 587–589.
- 32 Y. Venkatesh, J. Das, A. Chaudhuri, A. Karmakar, T. K. Maiti and N. D. P. Singh, *Chem. Commun.*, 2018, **54**, 3106–3109.
- 33 Z. Y. Xiao, T. Bonnard, A. Shakouri-Motlagh, R. A. L. Wylie, J. Collins, J. White, D. E. Heath, C. E. Hagemeyer and L. A. Connal, *Chem.–Eur. J.*, 2017, **23**, 11294–11300.
- 34 A. K. Steiger, S. Pardue, C. G. Kevil and M. D. Pluth, *J. Am. Chem. Soc.*, 2016, **138**, 7256–7259.
- 35 Y. Zhao and M. D. Pluth, *Angew. Chem., Int. Ed.*, 2016, **55**, 14638–14642.
- 36 Y. Zhao, H. A. Henthorn and M. D. Pluth, *J. Am. Chem. Soc.*, 2017, **139**, 16365–16376.
- 37 P. Chauhan, S. Jos and H. Chakrapani, *Org. Lett.*, 2018, **20**, 3766–3770.
- 38 P. Chauhan, P. Bora, G. Ravikumar, S. Jos and H. Chakrapani, *Org. Lett.*, 2017, **19**, 62–65.
- 39 A. K. Steiger, M. Marcatti, C. Szabo, B. Szczesny and M. D. Pluth, *ACS Chem. Biol.*, 2017, **12**, 2117–2123.
- 40 C. R. Powell, J. C. Foster, B. Okyere, M. H. Theus and J. B. Matson, *J. Am. Chem. Soc.*, 2016, **138**, 13477–13480.
- 41 A. K. Steiger, Y. Yang, M. Royzen and M. D. Pluth, *Chem. Commun.*, 2017, **53**, 1378–1380.
- 42 Y. Zhao, S. G. Bolton and M. D. Pluth, *Org. Lett.*, 2017, **19**, 2278–2281.
- 43 A. K. Sharma, M. Nair, P. Chauhan, K. Gupta, D. K. Saini and H. Chakrapani, *Org. Lett.*, 2017, **19**, 4822–4825.
- 44 P. Štacko, L. Muchová, L. Vítek and P. Klán, *Org. Lett.*, 2018, **20**, 4907–4911.
- 45 Y. Zhao, A. K. Steiger and M. D. Pluth, *Chem. Commun.*, 2018, **54**, 4951–4954.
- 46 P. Bora, P. Chauhan, K. A. Pardeshi and H. Chakrapani, *RSC Adv.*, 2018, **8**, 27359–27374.
- 47 B. Peng and M. Xian, *Asian J. Org. Chem.*, 2014, **3**, 914–924.
- 48 V. S. Lin, W. Chen, M. Xian and C. J. Chang, *Chem. Soc. Rev.*, 2015, **44**, 4596–4618.
- 49 Y. Zhao, A. K. Steiger and M. D. Pluth, *Angew. Chem., Int. Ed.*, 2018, **57**, 13101–13105.
- 50 B. Chen, W. Li, C. Lv, M. Zhao, H. Jin, J. Du, L. Zhang and X. Tang, *Analyst*, 2013, **138**, 946–951.
- 51 M. K. Thorson, T. Majtan, J. P. Kraus and A. M. Barrios, *Angew. Chem., Int. Ed.*, 2013, **52**, 4641–4644.
- 52 M. Whiteman, L. Li, P. Rose, C. H. Tan, D. B. Parkinson and P. K. Moore, *Antioxid. Redox Signaling*, 2010, **12**, 1147–1154.

