

Cite this: *Dalton Trans.*, 2019, **48**,
15198Received 18th July 2019,
Accepted 6th September 2019

DOI: 10.1039/c9dt02951f

rsc.li/dalton

Mechanistic insight into novel sulfoxide containing SABRE polarisation transfer catalysts†‡

Ben. J. Tickner, ^a Jennifer S. Lewis, ^a Richard O. John, ^a
Adrian C. Whitwood ^b and Simon B. Duckett ^{*a}

Signal Amplification By Reversible Exchange (SABRE) is a hyperpolarisation technique that commonly uses $[\text{Ir}(\text{H})_2(\text{carbene})(\text{substrate})_3]\text{Cl}$ complexes to catalytically transfer magnetisation from *para*-hydrogen derived hydride ligands to coordinated substrates. Here, we explore the reactivity of a novel class of such catalysts based on sulfoxide containing $[\text{IrCl}(\text{H})_2(\text{carbene})(\text{DMSO})_2]$, which are involved in the hyperpolarisation of pyruvate using SABRE. We probe the reactivity of this species by NMR and DFT and upon reaction with sodium pyruvate establish the formation of two isomers of $[\text{Ir}(\text{H})_2(\eta^2\text{-pyruvate})(\text{DMSO})(\text{IMes})]$. Studies with related disodium oxalate yield $[\text{Ir}_2(\text{H})_4(\text{IMes})_2(\text{DMSO})_2(\eta^2\text{-}\kappa^2\text{-Oxalate})]$ that is characterised by NMR and X-ray diffraction.

Introduction

Small molecule bond activation is an important area of inorganic chemistry that finds a central role in a range of applications in biochemistry,¹ recycling,² and organic synthesis.³ In fact, many rather inert reagents such as CO_2 ⁴ and R_3CH ⁵ can be activated by light⁶ or transition metal systems.^{7,8} The activation of small homonuclear diatomic molecules such as H_2 and O_2 by oxidative addition to metal centres has been known for many years.^{9,10} It typically occurs *via* a concerted mechanism wherein the newly introduced groups are located in a *cis* relationship.¹¹ Activation of more polar molecules such as CH_3I is, however, more often complex and this difference is revealed in the *trans* relationship between the two newly introduced groups.⁹ Orbital overlap effects during H_2 addition, as revealed through density function theory (DFT), are also complex with repulsive interactions between filled orbitals on the metal and H_2 yielding a barrier to the oxidative addition process that must be overcome.¹²

Many catalytic processes exploit the oxidative addition of H_2 to a transition metal centre by enabling the subsequent transfer of the two hydrogen atoms into an unsaturated centre^{13,14} However, recent examples in main group chemistry involving frustrated Lewis pairs mean that direct addition is

also possible.¹⁵ Understanding of these reactions can be improved by exploiting a technique known as *para*-Hydrogen Induced Polarisation (PHIP) to detect reaction intermediates.^{16,17} This process incorporates the two protons of a single *para*-hydrogen (*p*- H_2) molecule into a substrate *via* a hydrogenation reaction to see the PHIP effect. *p*- H_2 is a spin isomer of H_2 and exists as a singlet state with a nuclear spin order of zero. H_2 gas can be enriched (>98%) in its *para* state by simply cooling it in the presence of a paramagnetic catalyst such as Fe_2O_3 or activated charcoal.^{18–20} If the spin orientation of *p*- H_2 survives the ensuing hydrogenation reaction the NMR properties of the product can be harvested to increase product detectability. This effect was exemplified in 1986 by Bowers and Weitekamp^{21,22} with Eisenberg and Bargon^{23,24} producing similar independent observations around the same time. As the net nuclear spin of *p*- H_2 is zero it is invisible to an NMR experiment but once the symmetry of its two protons is broken the resulting spin order can be detected. In fact, the NMR signal intensity of the now NMR visible, and hyperpolarised, product is derived from the resulting large non-Boltzmann populations that lie across its nuclear spin energy levels. Since its introduction PHIP has been used to detect low concentration analytes and true intermediates in the field of catalysis.^{22,25–27} Excitement has resulted from the hydrogenation of some unsaturated clinical agents which has led to their *in vivo* MRI detection.^{28–30}

Since 2009, the *p*- H_2 based method Signal Amplification By Reversible Exchange (SABRE) has been used to harness signal gains from *p*- H_2 without the need for the direct hydrogenation of a substrate (sub).³¹ It achieves the catalytic transfer of magnetisation into a substrate through the formation of a *J*-coupled network within the associated catalyst.³² The first

^aCenter for Hyperpolarization in Magnetic Resonance (CHyM), University of York, Heslington, York, YO10 5NY, UK. E-mail: simon.duckett@york.ac.uk

^bDepartment of Chemistry, University of York, Heslington, York, YO10 5DD, UK

† Dedicated to Prof. Robin N. Perutz on the occasion of his 70th birthday.

‡ Electronic supplementary information (ESI) available. CCDC 1941589. For ESI and crystallographic data in CIF or other electronic format see DOI: 10.1039/c9dt02951f



step in SABRE typically involves the conversion of a stable 16 electron precursor such as $[\text{IrCl}(\text{COD})(\text{IMes})]$ (**1**) (where $\text{IMes} = 1,3\text{-bis}(2,4,6\text{-trimethylphenyl})\text{imidazol-2-ylidene}$ and $\text{COD} = \text{cis,cis-1,5-cyclooctadiene}$) into an intermediate $[\text{Ir}(\text{COD})(\text{IMes})(\text{sub})]\text{Cl}$ which then undergoes hydrogen addition to form highly reactive $[\text{Ir}(\text{H})_2(\text{COD})(\text{IMes})(\text{sub})]\text{Cl}$.^{33–35} Subsequent hydrogenation of the COD ligand then leads to a SABRE catalyst such as $[\text{Ir}(\text{H})_2(\text{IMes})(\text{sub})_3]\text{Cl}$ which importantly reversibly binds both H_2 and sub. In SABRE, H_2 addition to square planar $[\text{Ir}(\text{COD})(\text{IMes})(\text{Sub})]\text{Cl}$ typically occurs over the COD-Ir-Sub axis rather than the COD-Ir-IMes axis.³⁶ This observation has been explained in terms of both reduced steric interactions with the bulky carbene ligand and electronic effects in terms of orbital interactions.³⁶ It has been suggested that H_2 addition is favoured over axis containing ligands with π acceptor orbitals which provide additional stabilising interactions as the geometry of the complex changes during the addition step.³⁷

The most common substrate molecules used in SABRE have proven to feature *N*-donor sites that readily coordinate to iridium. *N*-Heterocycles such as pyridine,^{31,38–41} nicotinamides,^{31,42,43} pyrazines,^{39,40} and pyrazoles⁴¹ reflect common examples although other *N*-functionalities found in nitriles⁴⁴ and amines⁴⁵ have been used. It has recently been reported that in the presence of a stabilising sulfoxide coligand, the reversible coordination and subsequent hyperpolarisation of oxygen ligating pyruvate can occur.⁴⁶

The direct hyperpolarisation of pyruvate was made possible in this case by the formation of the novel polarization transfer catalyst $[\text{Ir}(\text{H})_2(\eta^2\text{-pyruvate})(\text{DMSO})(\text{IMes})]$. In addition to this active catalyst, $[\text{IrCl}(\text{H})_2(\text{DMSO})_2(\text{IMes})]$, is also present in solution and is expected to be critical in improving the efficiency of this important hyperpolarisation transfer process. In this work we investigate the formation, behaviour, and ligand exchange processes exhibited by this sulfoxide containing polarisation transfer catalyst. We do this in solvents that were purchased from Sigma and used without further purification as our aim is to study reactivity without taking any special precautions. This is reflective of the most likely scenario when used by the non-specialist. We extend this method to include an examination of the behaviour of oxalate with the aim of developing the range of materials that can be hyperpolarised with sulfoxide containing SABRE polarisation transfer catalysts.

Results and discussion

Formation of $[\text{Ir}(\text{COD})(\text{IMes})(\text{OH}_2)]\text{Cl}$ (**2**) from $[\text{IrCl}(\text{COD})(\text{IMes})]$ (**1**)

When the SABRE precatalyst $[\text{IrCl}(\text{COD})(\text{IMes})]$ (**1**) is dissolved in 0.6 mL of methanol- d_4 at 298 K, ^1H NMR resonances can be observed for **1** in addition to a minor product, **2** which forms at the ~2% level. The ratio of this product increases to 13% after the addition of 50 μL H_2O , as shown in Fig. 1 thereby confirming a role of the original H_2O contaminant in metha-

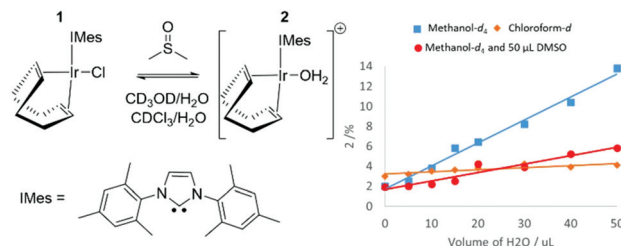


Fig. 1 Demonstrating the proportion of **2** in solution increases with the level of added water in the solvents methanol and chloroform in addition to a methanol–DMSO mixture thereby confirming its identity as $[\text{Ir}(\text{COD})(\text{IMes})(\text{OH}_2)]\text{Cl}$ as shown.

nol- d_4 . When this experiment was repeated in a solution of 0.6 mL methanol- d_4 doped with 5 μL DMSO, the ratio of **2** formed after the addition of 50 μL of H_2O proved to be just 6%. It is noteworthy that no evidence for $[\text{IrCl}(\text{COD})(\text{DMSO})(\text{IMes})]$ was apparent. A similar ^1H NMR measurement of **1** in a solution of CDCl_3 revealed the presence of **2** at the 3% level. Subsequent addition of H_2O increases this to 5% but the immiscibility of CDCl_3 and H_2O prevents the use of high water concentrations. Hence, the equilibrium position between **1** and **2** is dependent on solvent choice and water concentration. **2** is assigned as $[\text{Ir}(\text{COD})(\text{IMes})(\text{OH}_2)]\text{Cl}$ which is further confirmed by 2D NMR characterisation data (see ESI ‡) at 245 K which shows a NOE between the bound aqua ligand at δ 8.15 and the IMes backbone at δ 2.16. ^1H NMR resonances for hexa-aqua species are often found between δ 8 and δ 11.⁴⁷ The formation of **2** is not unexpected as iridium aqua complexes are known and the reactions performed throughout this work are not completed under anhydrous conditions because our aim is to achieve SABRE catalysis using an air stable precursor with minimum end-user challenge.^{48–50}

DFT calculations have been used to further confirm these product assignments, with their predicted relative energies detailed in Table 1 (see ESI ‡ for details). These calculations used full models of the complexes, $[\text{Ir}(\text{COD})(\text{IMes})(\text{X})]$, and are relative to a zero point which includes all species (indicated complex, DMSO, H_2O and CH_3OH) in an equimolar amount. They do not reflect transition state barriers and excess reagents are not accounted for. These calculations confirm that **1**, where $\text{X} = \text{Cl}$, is most stable in agreement with experiment. $[\text{Ir}(\text{COD})(\text{IMes})(\text{OH}_2)]\text{Cl}$, **2** is predicted to be more stable than the corresponding methanol or sulfoxide adducts, again supporting our NMR observations.

Table 1 Relative enthalpy (*H*) and Gibbs free energies (*G*) of a series of $[\text{Ir}(\text{COD})(\text{IMes})(\text{X})]$ complexes according to DFT calculations

X	<i>H</i> /kJ mol ⁻¹	<i>G</i> /kJ mol ⁻¹
Cl	0	0
OH_2	52.8	57.0
<i>S</i> -DMSO	42.3	68.9
HOCH_3	56.1	69.4



Formation of $[\text{IrCl}(\text{H})_2(\text{DMSO})_2(\text{IMes})]$ (**3**)

When H_2 is added to an equilibrium mixture of **1** and **2** containing 4 equivalents of DMSO in methanol- d_4 at 245 K the initial ^1H NMR spectra reveal two hydride resonances at $\delta -15.49$ and $\delta -21.51$ which share a mutual J coupling of 5.5 Hz. Low temperature NMR characterisation of the complex yielding these signals confirms it to be of the form $[\text{Ir}(\text{H})_2(\text{IMes})(\text{DMSO})_2(\text{L})]$, **3**.⁴⁶ The formation of iridium(III) sulfoxide complexes bound through sulphur for use in SABRE hyperpolarisation studies has been reported previously.⁵¹ When this hydrogenation reaction is examined at 245 K in CDCl_3 or dichloromethane- d_2 ($\text{DCM}-d_2$) hydride resonances appear at $\delta -15.50$, $\delta -21.12$ and $\delta -15.67$, $\delta -21.36$, respectively. The similarity in these signals suggests the common presence of **3** in both cases. This chemistry is complicated, however, by the fact **3** is unstable in solution over long time-scales, as detailed in the ESI.†

The identity of **3** was further confirmed by repeating this experiment using $[\text{IrBr}(\text{COD})(\text{IMes})]$, as the corresponding reaction product $[\text{IrBr}(\text{H})_2(\text{DMSO})_2(\text{IMes})]$ yields hydride signals at $\delta -15.67$ and $\delta -20.45$ in methanol- d_4 which are clearly different to those of **3**. Furthermore, when $[\text{Ir}(\text{CH}_3\text{CN})(\text{COD})(\text{IMes})]\text{PF}_6$ is used instead, $[\text{Ir}(\text{H})_2(\text{CH}_3\text{CN})(\text{DMSO})_2(\text{IMes})]\text{PF}_6$ forms which yields signals at $\delta -15.76$ and $\delta -19.83$ at 255 K; we note there was no evidence for the displacement of CH_3CN by DMSO in the associated NMR spectra. Hence, the hydride resonance in these complexes shift according to the identity of the ligand that is *trans* to it thereby confirming the indicated product identities. The identity of **3** was also studied by DFT as detailed in the results shown in Table 2. $[\text{IrCl}(\text{H})_2(\text{DMSO})_2(\text{IMes})]$ was found to be lower in energy than the corresponding species where chloride is replaced by water, methanol or sulfoxide.

When the reaction of H_2 with an equilibrium mixture of **1** and **2** in methanol- d_4 containing DMSO was monitored at 245 K (Fig. 2) no evidence for the formation of any H_2 addition products except **3** is observed. This suggests that H_2 addition is slow, and subsequent COD hydrogenation in intermediate **1-H**₂ of Fig. 2 is fast. Interestingly, the proportion of **2** remains roughly constant as this conversion proceeds thereby suggesting any equilibrations involving it are also slow. The route to **3** is therefore most likely to involve direct H_2 addition to **1** rather than **2**. DFT confirms that H_2 addition to **1** proceeds over its COD-Ir-Cl axis rather than the COD-Ir-IMes axis according to the relative energies of the corresponding products (see ESI†). This is supported by the fact that when this H_2 addition



Fig. 2 (a) Reaction steps involved in the conversion of **1** to **3** in the presence of DMSO and H_2 , (b) time course for the reaction, as determined in methanol- d_4 solution when the starting concentrations are 5 mM $[\text{IrCl}(\text{COD})(\text{IMes})]$, 1 μL DMSO and 3 bar hydrogen pressure and (c) analogous reaction time course data in CDCl_3 at 245 K.

reaction is monitored in CDCl_3 at 245 K, resonances corresponding to **1-H**₂, at $\delta -13.39$ and $\delta -18.42$, are detected in addition to those of **3**. The resonances for **1-H**₂ rapidly disappear upon warming this solution to 298 K where **3** then forms.

Ligand exchange processes of $[\text{IrCl}(\text{H})_2(\text{DMSO})_2(\text{IMes})]$ (**3**)

When **3** is formed from a methanol- d_4 solution containing 2 mg of **1**, 1 μL of DMSO and 3-bar H_2 , its hydride resonances appear very broad at 298 K (line width of ~ 120 Hz) which is consistent with rapid hydrogen loss. This deduction is confirmed upon shaking methanol- d_4 solutions of **3** with 3-bar $p\text{-H}_2$ for 10 seconds at 65 G at 298 K. Its hydride resonances exhibit PHIP signal enhancement alongside a free DMSO signal that yields a weak SABRE ^1H NMR signal enhancement of 7-fold (see ESI†). This is consistent with the reversible pairwise addition of H_2 to **3** and loss of DMSO. Product **3** actually yields four distinct ^1H NMR signals for the methyl groups of its two DMSO ligands. The DMSO ligand *cis* to carbene yields a pair of inequivalent CH_3 signals at $\delta 2.83$ and 3.12 while DMSO *trans* to carbene resonates at $\delta 3.19$ and 3.27 . Upon selective excitation of bound DMSO resonances *cis* to carbene in **3**, evidence for chemical exchange into free DMSO is observed. In contrast, we observe no exchange of the DMSO ligand that lies *trans* to carbene on this timescale although we do observe interchange of the inequivalent CH_3 groups. This suggests that loss of the DMSO ligand *cis* to IMes accounts for this scrambling of the inequivalent CH_3 resonances of the DMSO *trans* to IMes.

Measuring the change in these resonance's signal intensities as a function of time after the initial *r.f.* excitation step

Table 2 Relative enthalpy (H) and Gibbs free energies (G) of a series of $[\text{IrX}(\text{H})_2(\text{DMSO})_2(\text{IMes})]$ complexes according to DFT calculations

X	$H/\text{kJ mol}^{-1}$	$G/\text{kJ mol}^{-1}$
Cl	0	0
OH_2	24.4	34.3
S-DMSO	39.1	70.8
ODCH_3	29.3	45.4



allowed the associated ligand loss rates to be calculated. The rate of reversible hydrogen loss, k_{H_2} , was found to be $3.31 \pm 0.25 \text{ s}^{-1}$ at 263 K and is faster than that seen in many related systems.⁵² Loss, and hence exchange of the DMSO ligand *cis* to IMes, was found to occur on a similar timescale with k_{DMSO} $3.35 \pm 0.01 \text{ s}^{-1}$ at 263 K. Repeating these measurements at different temperatures allowed the transition state barriers for these two processes to be determined as $\Delta H^\ddagger_{(\text{H}_2)}$ $78 \pm 6 \text{ kJ mol}^{-1}$, $\Delta S^\ddagger_{(\text{H}_2)}$ $60 \pm 25 \text{ J K}^{-1} \text{ mol}^{-1}$ and $\Delta H^\ddagger_{(\text{DMSO})}$ $79 \pm 6 \text{ kJ mol}^{-1}$, $\Delta S^\ddagger_{(\text{DMSO})}$ $66 \pm 8 \text{ J K}^{-1} \text{ mol}^{-1}$ respectively. This enthalpic barrier to hydrogen loss is comparable to those for similar complexes, overall the entropy change suggests that a dissociative pathway is adopted.⁵³

The effects of H_2 pressure and DMSO concentration on these exchange processes were also studied in methanol- d_4 at 243 K for differing reagent concentrations. k_{H_2} proved to increase at higher H_2 concentrations before reaching a plateau as detailed in the ESI.† The rate of DMSO exchange is unaffected by increasing DMSO or H_2 concentration. We therefore propose that DMSO and H_2 loss proceed *via* formation of the common 16-electron intermediate $[\text{IrCl}(\text{H})_2(\text{DMSO})(\text{IMes})]$ in what is a dissociative first step. H_2 exchange then proceeds *via* the formation of $[\text{IrCl}(\text{H})_2(\text{H}_2)(\text{DMSO})(\text{IMes})]$; such indirect H_2 exchange, rather than direct H_2 loss, has been reported for several similar systems.^{51,54} When similar EXSY data were collected in CD_2Cl_2 , the rate of hydrogen and DMSO loss proved to be slower than those in methanol- d_4 , as summarised in Table 3.

These deductions were confirmed through further DFT calculations that revealed the products of direct H_2 loss to form a 16 electron product as being highly energetically unfavourable, as shown in Table 4. The five coordinate product formed by loss of DMSO *trans* to carbene proved highly unstable, undergoing spontaneous rearrangement to form an intermediate with a vacancy in the equatorial plane; this is consistent with the EXSY data. In contrast, the predicted ligand dissociation pathway involves the formation of five-coordinate $[\text{IrCl}(\text{H})_2(\text{DMSO})(\text{IMes})]$ *via* loss of the DMSO ligand that lies *cis* to carbene.

Even at low temperature (245–265 K), H/D exchange leading to $[\text{IrCl}(\text{D})(\text{H})(\text{DMSO})_2(\text{IMes})]$ (**3-d**) and $[\text{IrCl}(\text{D})_2(\text{DMSO})_2(\text{IMes})]$ was evident. Binding of solvent methanol to $[\text{IrCl}(\text{H})_2(\text{DMSO})(\text{IMes})]$ is an obvious route to deuterium exchange and the formation of these species alongside HD and D_2 gas. While we do not directly observe methanol bound adducts, they are suggested to form in similar systems and indeed proposed to account for the hydrogen isotope exchange

Table 4 Relative enthalpy (H) and Gibbs free energies (G) of products arising from ligand loss of **3** according to DFT calculations. These energies are relative to those of **3** and do not reflect transition state barriers

Loss of	$H/\text{kJ mol}^{-1}$	$G/\text{kJ mol}^{-1}$
H_2	89.0	58.5
Cl	39.6	3.9
DMSO <i>cis</i> carbene	32.7	-10.1

reaction that is often observed to run in parallel to SABRE.^{50,55} Experimentally, at low temperature it proved possible to reliably and selectively excite the hydride resonance of **3** or **3-d**. The selective excitation of the hydride resonances for these two species revealed their selective chemical exchange into H_2 and HD, from **3** and **3-d** respectively. Therefore, k_{H_2} and k_{HD} can be determined as previously described. These values proved to be the same within error and hence, there is little or no kinetic isotope effect which is consistent with other reports.^{56,57} We note that exchange between **3** and **3-d** is not observed in the associated EXSY data which provides confirmation that the underlying deuterium exchange processes involving methanol- d_4 are slow.

Formation of $[\text{Ir}(\text{H})_2(\eta^2\text{-pyruvate})(\text{DMSO})(\text{IMes})]$, (**4**)

When sodium pyruvate (5 equivalents relative to iridium) is added to a solution of preformed **3** in methanol- d_4 at 245 K, two new hydride ligand containing products form, which correspond to isomers **4a** and **4b** of $[\text{Ir}(\text{H})_2(\eta^2\text{-pyruvate})(\text{DMSO})(\text{IMes})]$ as detailed in Fig. 3a. Logically, a further isomer of **4a** is possible, **4c**, in which the orientation of the pyruvate is rotated but this is not observed in solution. These products are differentiated from each other by the geometry of the coordinated pyruvate ligand and their proportion is both reaction time and temperature dependent (Fig. 3b). DFT (Table 5) is used to confirm that **4a** exhibits the ligand geometry shown in Fig. 3a and that **4b** is the most stable species. Despite observation of a 10 Hz coupling between the pyruvate COOH group and the hydride *trans* to oxygen, according to DFT the pyruvate CH_3 and DMSO groups of **4a** are arranged in a *cis* fashion as shown in Fig. 3a. This deduction was further confirmed by the observation of an NOE peak between the pyruvate CH_3 group and the phenyl protons on the mesityl group of the carbene ligand.

When this solution was examined with a 32 scan ^1H NMR measurement at 298 K, unlike the 245 K data of Fig. 3b, the main hydride containing complex observed is **4b** and reso-

Table 3 Rate and thermodynamic parameters for hydrogen and DMSO exchange of **3**

Solvent	Process	$k_{(263 \text{ K})}/\text{s}^{-1}$	$\Delta H^\ddagger/\text{kJ mol}^{-1}$	$\Delta S^\ddagger/\text{J K}^{-1} \text{ mol}^{-1}$
Methanol- d_4	Hydrogen exchange (k_{H_2})	3.31 ± 0.25	78 ± 6	60 ± 25
	DMSO exchange (k_{DMSO})	3.35 ± 0.04	79 ± 6	66 ± 8
Dichloromethane- d_2	Hydrogen exchange (k_{H_2})	1.16 ± 0.04	83 ± 8	73 ± 29
	DMSO exchange (k_{DMSO})	1.56 ± 0.01	84 ± 2	78 ± 8



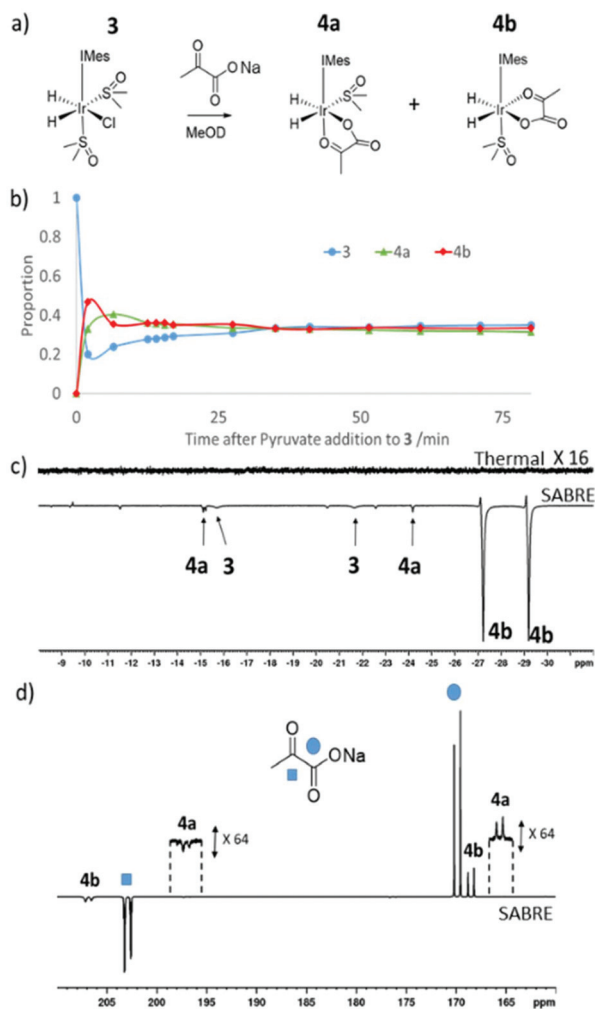


Fig. 3 (a) Reaction of **3** and pyruvate to form **4** (b) monitoring pyruvate addition to **3** using ^1H NMR spectroscopy. Pyruvate addition was made at room temperature to a solution of preformed **3** at 245 K before it was introduced into the spectrometer whose probe was at 245 K. There is therefore a rapid temperature change at the start of this data series. Upon shaking a solution of **3** and **4** with 3-bar $p\text{-H}_2$ for 10 seconds at (c) 65 G or (d) in a mu metal shield strongly hyperpolarised ^1H (c) or ^{13}C (d) resonances of **4b** are observed compared to the thermal trace.

Table 5 Relative enthalpy (H) and Gibbs free energies (G) of **4** according to DFT calculation

Complex	$H/\text{kJ mol}^{-1}$	$G/\text{kJ mol}^{-1}$
4a	8.5	14.8
4b	0	0
4c	22.6	28.1

nanances for **3** and **4a** cannot be discerned. **4a** and **4b** have previously been implicated in pyruvate polarisation using SABRE and their NMR characterisation data has been reported.⁴⁶ **4** is also detected immediately after H_2 addition to a solution of **1** containing pyruvate (5 equivalents relative to iridium) and DMSO (10 equivalents) at 298 K. When an equilibrium mixture

of **3** and **4** is shaken with 3 bar $p\text{-H}_2$ at 65 G enhanced hydride resonances are observed in the ^1H NMR spectrum as shown in Fig. 3c, which are strongest for **4b**.

If sodium-1,2-pyruvate- $^{13}\text{C}_2$ is used as the substrate and the $p\text{-H}_2$ shaking process is performed under SABRE-SHEATH conditions in a mu metal shield (see Experimental) an enhanced ^{13}C response for the free material at δ 169 and δ 203 can also be readily detected.⁴⁶ Two enhanced signals can be also seen for the bound pyruvate ligand in **4b** at δ 168 and δ 207 in these NMR spectra. It is also possible to observe extremely weak hyperpolarised signals for the bound pyruvate ligand in **4a** at δ 161 and δ 198, as shown in Fig. 4d. Interestingly, when EXSY measurements are used to probe these signals, the selective excitation of the hydride resonances of **4b** reveals no exchange into H_2 on the NMR timescale. Furthermore, when ^{13}C -EXSY is used to probe the bound signals of **4b**, no exchange is seen into free pyruvate on this timescale. Hence **4b** appears to be relatively stable in agreement with the DFT study, but this is not consistent with the strong hydride polarisation that is evident for **4b** in Fig. 3. Consequently, a role for **3** which is present and undergoes rapid H_2 exchange as discussed earlier in its formation is indicated. It is by this route that the delivery of fresh $p\text{-H}_2$ into **4** and the resulting enhancement of hydride and pyruvate ligand signals is achieved. As the NMR relaxation times of the ^{13}C resonances in pyruvate are much longer than those of ^1H , slow exchange is still commensurate with the build-up of pyruvate polarisation in solution. Related relayed $p\text{-H}_2$ enhancement effects have been described recently.^{58,59}

An equilibrium mixture containing **4** can also be formed if sodium-1,2-pyruvate- $^{13}\text{C}_2$ is added to a preformed methanol- d_4 solution of $[\text{IrBr}(\text{H})_2(\text{DMSO})_2(\text{IMes})]$ (**3-Br**) or $[\text{Ir}(\text{H})_2(\text{CH}_3\text{CN})(\text{DMSO})_2(\text{IMes})]\text{PF}_6$ (**3-Acn**). When these samples are shaken with $p\text{-H}_2$ for 10 seconds at 65 G strongly hyperpolarised hydride resonances can again be observed for **4b** in the corresponding high field NMR measurement. These resonances appear with significantly lower intensity than those achieved with **3** derived systems (35% and 31% of this signal

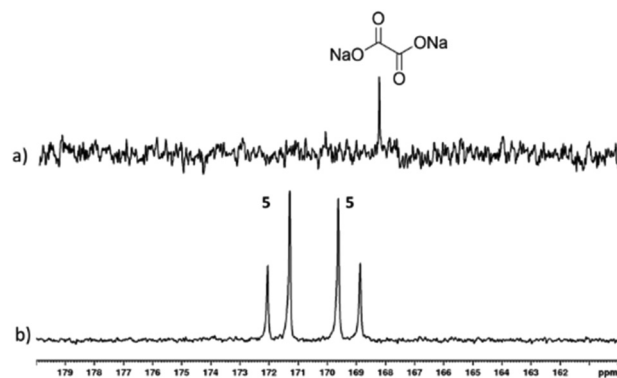


Fig. 4 (a) 128 scan ^{13}C spectrum of a solution of **1**, DMSO and sodium-1,2-oxalate- $^{13}\text{C}_2$ (1 : 5 : 5) in methanol- d_4 (b) ^{13}C NMR signals seen after shaking with 3-bar $p\text{-H}_2$ in a mu metal shield for 30 seconds.



intensity respectively). Furthermore, when the p -H₂ shaking step is repeated in a mu metal shield, hyperpolarised resonances for ¹³C-pyruvate are again visible. The corresponding ¹³C-pyruvate signal enhancements are typically 580-fold and 120-fold for the **3-Br** and **3-Acn** derived systems respectively. These are lower than those achieved for the analogous **3** system (1070-fold) which is consistent with the reduced amount of **4b** which is present at equilibrium when these precursors are used (22 and 23% respectively when compared to 92% with **3**).

We note that the linewidths of the hydride resonances of **4b** at 298 K are similar in each of these samples (43–45 Hz). Similarly, the linewidths of **3**, **3-Br** and **3-Acn** are comparable at 245 K (3–5 Hz). Therefore, we do not expect that differences in hydride polarisation levels seen for **4b** in these solutions are due to different hydrogen exchange rates in **3**, but rather the different binding strengths of the coligands that must be displaced by pyruvate to form **4** from **3**. Hence, whilst there appears to be a link between pyruvate polarisation level and **4b** concentration, the identity of L in [Ir(H)₂(DMSO)₂(IMes)₂L]_n must play a large effect on the level of pyruvate signal enhancement. These data further confirm that **3** is important in mediating efficient H₂ exchange within the 3/4 hyperpolarisation mixture.

Using [IrCl(H)₂(DMSO)₂(IMes)] (**3**) to hyperpolarise sodium-1,2-oxalate-[¹³C₂]

We have shown how **3** can be formed *in situ* and subsequently used as a precursor to form [Ir(H)₂(η²-pyruvate)(DMSO)(IMes)], (**4**) which delivers SABRE enhancement to pyruvate. This α-keto acid motif also features in oxalate, a metabolic product that binds mineral ions in the body and is found in many foods.⁶⁰ Indeed, Levitt and coworkers have reported the creation of a long lived hyperpolarised ¹³C₂ singlet spin pair for a deuterated ester derivative of oxalate using DNP.⁶¹ Consequently, we now describe tests on sodium-1,2-oxalate-[¹³C₂]. This involved shaking a mixture of **1**, DMSO, and sodium-1,2-oxalate-[¹³C₂] (1 : 5 : 5) with 3-bar p -H₂ in 0.6 mL methanol-*d*₄ for 30 seconds in a mu metal shield. Hyperpolarised ¹³C resonances were observed at δ 169.20 and δ 171.66 that share a J_{CC} coupling of 76 Hz, as shown in Fig. 4. These signals cannot be due to free sodium-1,2-oxalate-[¹³C₂] as a single resonance is expected.

While the complexation dynamics of oxalate are complex,^{62–64} based on the pyruvate observations described earlier it should be possible to form a species such as [Ir(H)₂(η²-oxalate)(DMSO)(IMes)], **5** which would account for this observation. In fact, the ¹³C₂ NMR signal profile shown in Fig. 4 is indicative of Zeeman magnetisation in such a product rather than singlet spin order. This indicates that while ¹³C polarisation transfer from a set of p -H₂ derived hydride ligands initially results this must be associated with a [AA'BB'] spin system, which converts to the [AMXS] type with inequivalent hydride ligands in high field. A product with ligand arrangement analogous to **4a** would match with this hypothesis.⁴⁶ Unfortunately, examination of the hydride region of the result-

ing ¹H NMR spectra reveal signals for **3** at δ –15.45 and δ –21.53 alongside over seven minor hydride containing complexes. None of the hydride resonances of these species correlated to these two ¹³C signals in an overnight 2D measurement at 243 K. Studying this reaction again at 315 K, proved to be of no benefit as conversion of **3** into the same range of hydride containing species results.

In contrast, when this experiment was repeated in DCM-*d*₂ the hydride resonances of **3** are again observed at δ –15.71 and δ –21.27 but no ¹³C oxalate derived polarisation is seen. There is very low solubility of sodium oxalate in this medium. As a consequence, samples of **3** in CD₂Cl₂ or CH₃OD were prepared and reacted with 50 μL D₂O solutions of sodium-1,2-oxalate-[¹³C₂]. Now a new product forms cleanly, that yields a hydride signal at δ –27.1 alongside diagnostic resonances for the IMes at δ 2.10, δ 2.34, δ 6.99, δ 7.15 and DMSO at δ 2.92 whose relative signal intensities suggest the presence of a [Ir(H)₂(IMes)₂(DMSO)(L)_n]_x species, **6**. Upon shaking with p -H₂ the hydride resonances for **6** do not enhance, however, the addition of 3 mL of degassed hexane results in the growth of single crystals. Subsequent X-ray diffraction studies confirmed that the product **6** corresponds to the dimer [Ir₂(H)₄(IMes)₂(DMSO)₂(η²-κ²-Oxalate)] with structure shown in Fig. 5 (full X-ray and NMR characterisation data are given in the in the ESI†). The ability of oxalate to form such dimers restricts its dissociation and thereby limits its use in these hyperpolarisation studies. Nonetheless, the structure of **6** reflects a common binding mode of oxalate with transition metals.^{63,65,66}

Experimental

All NMR measurements were carried out on a 400 MHz Bruker Avance III spectrometer using solutions at room temperature (298 K) unless otherwise stated. *Para*-hydrogen (p -H₂) was produced by passing hydrogen gas over a spin-exchange catalyst (Fe₂O₃) and used for all hyperpolarisation experiments. This method produces constant p -H₂ with *ca.* 93% purity. ¹H (400 MHz) and ¹³C (100.6 MHz) NMR spectra were recorded

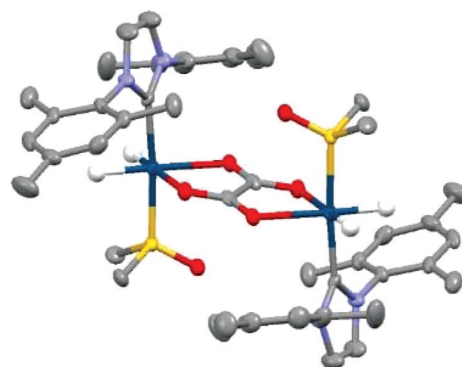


Fig. 5 Structure of [Ir₂(H)₄(IMes)₂(DMSO)₂(η²-κ²-Oxalate)], **6**, determined by X-ray crystallography. All non-hydride hydrogen atoms and solvent of crystallisation have been omitted for clarity.



with an internal deuterium lock. Chemical shifts are quoted as parts per million and referenced to the solvent. ^{13}C NMR spectra were recorded with broadband proton decoupling. Coupling constants (J) are quoted in Hertz.

Samples were prepared containing 2 mg $[\text{IrCl}(\text{COD})(\text{IMes})]$ precatalyst (where IMes = 1,3-bis(2,4,6-trimethyl-phenyl)imidazole-2-ylidene and COD = *cis,cis*-1,5-cyclooctadiene) in 0.6 mL of deuterated methanol- d_4 unless otherwise stated in a 5 mm NMR tube that was fitted with a J. Young's tap. All commercial compounds were purchased from Sigma-Aldrich, Fluorochem, or Alfa-Aesar and used as supplied. $[\text{IrCl}(\text{COD})(\text{IMes})]$ was synthesized according to a literature procedure.⁶⁷ The resulting solutions were degassed by two freeze-pump-thaw cycles before the addition of 3-bar H_2 .

The shake and drop method was employed for recording hyperpolarised NMR spectra. This involves filling NMR tubes with $p\text{-H}_2$ at 3 bar pressure and shaking them vigorously for 10 seconds in a 65 G magnetic field if ^1H NMR spectra are to be recorded at 9.4 T. A μ metal shield is used if ^{13}C NMR spectra are to be recorded. Unless otherwise stated multiple shake and drop measurements are undertaken and average signal enhancement values quoted. Signal enhancements and exchanges rates were calculated according to literature procedures.⁶⁸

Conclusions

In conclusion, we have confirmed that the complex $[\text{IrCl}(\text{H})_2(\text{DMSO})_2(\text{IMes})]$ can be used as a polarisation transfer catalyst for hyperpolarising pyruvate. Complexes of this type undergo rapid hydrogen exchange which through a mechanistic investigation involving EXSY and DFT is expected to proceed dissociatively *via* five coordinate $[\text{IrCl}(\text{H})_2(\text{DMSO})(\text{IMes})]$. When this process is completed in the presence of an α -ketoacid, such as pyruvate, reaction to form $[\text{Ir}(\text{H})_2(\eta^2\text{-pyruvate})(\text{DMSO})(\text{IMes})]$, which exists in two coordination isomers, occurs readily. Isomer **4b**, in which the hydride ligands lie *trans* to pyruvate, is implicated in strong SABRE hyperpolarisation despite it undergoing slow ligand exchange. Consequently, a role for $[\text{IrCl}(\text{H})_2(\text{DMSO})_2(\text{IMes})]$, which undergoes rapid H_2 and DMSO loss, is indicated in this process. When pyruvate is replaced by oxalate, stronger and more complex ligand binding is evident. The trapping of oxalate within the dimer **6** is expected to explain its much less efficient hyperpolarisation. Nonetheless, we predict that variations of the sulfoxide and NHC will enable future hyperpolarisation of oxalate and those of a wider range of other α -keto acids.

Conflicts of interest

B. J. T. and S. B. D. (and others) are inventors on a patent application filed by the University of York related to this work (patent no. GB1818171.9, filed 7 November 2018).

Acknowledgements

We thank Dr Peter Rayner, Hannah Kettle, and Dr Victoria Annis for synthesis of the $[\text{IrCl}(\text{COD})(\text{IMes})]$, $[\text{IrBr}(\text{COD})(\text{IMes})]$ and $[\text{Ir}(\text{CH}_3\text{CN})(\text{COD})(\text{IMes})]\text{PF}_6$ precatalysts. Dr Wissam Iali is thanked for early stage discussions. Financial support from the Wellcome Trust (Grants 092506 and 098335), the MRC (MR/M008991/1) and the EPSRC (B. J. T. studentship and Impact Accelerator Award G0025101) is gratefully acknowledged.

References

- 1 T. S. Leyh, *Crit. Rev. Biochem. Mol. Biol.*, 1993, **28**, 515–542.
- 2 X. Liu, L. He, Y.-M. Liu and Y. Cao, *Acc. Chem. Res.*, 2013, **47**, 793–804.
- 3 H. Amii and K. Uneyama, *Chem. Rev.*, 2009, **109**, 2119–2183.
- 4 D. M. Ermert and L. J. Murray, *Dalton Trans.*, 2016, **45**, 14499–14507.
- 5 R. Pretorius, M. R. Fructos, H. Müller-Bunz, R. A. Gossage, P. J. Pérez and M. Albrecht, *Dalton Trans.*, 2016, **45**, 14591–14602.
- 6 H. Huo, X. Shen, C. Wang, L. Zhang, P. Röse, L.-A. Chen, K. Harms, M. Marsch, G. Hilt and E. Meggers, *Nature*, 2014, **515**, 100.
- 7 L.-S. Wang, R. McDonald and M. Cowie, *Inorg. Chem.*, 1994, **33**, 3735–3744.
- 8 M. Álvarez, E. Álvarez, M. R. Fructos, J. Urbano and P. J. Pérez, *Dalton Trans.*, 2016, **45**, 14628–14633.
- 9 P. B. Chock and J. Halpern, *J. Am. Chem. Soc.*, 1966, **88**, 3511–3514.
- 10 W. H. Thompson and C. T. Sears, *Inorg. Chem.*, 1977, **16**, 769–774.
- 11 K. Searles, M. Pink, K. G. Caulton and D. J. Mindiola, *Dalton Trans.*, 2012, **41**, 9619–9622.
- 12 C. E. Johnson and R. Eisenberg, *J. Am. Chem. Soc.*, 1985, **107**, 3148–3160.
- 13 N. Wang, M. Wang, Y. Wang, D. Zheng, H. Han, M. r. S. G. Ahlquist and L. Sun, *J. Am. Chem. Soc.*, 2013, **135**, 13688–13691.
- 14 C. R. Landis and T. W. Brauch, *Inorg. Chim. Acta*, 1998, **270**, 285–297.
- 15 S. Grimme, H. Kruse, L. Goerigk and G. Erker, *Angew. Chem., Int. Ed.*, 2010, **49**, 1402–1405.
- 16 K. Sorochkina, V. V. Zhivonitko, K. Chernichenko, V.-V. Telkki, T. Repo and I. V. Koptyug, *J. Phys. Chem. Lett.*, 2018, **9**, 903–907.
- 17 V. V. Zhivonitko, K. Sorochkina, K. Chernichenko, B. Kótai, T. Földes, I. Pápai, V.-V. Telkki, T. Repo and I. Koptyug, *Phys. Chem. Chem. Phys.*, 2016, **18**, 27784–27795.
- 18 S. Wagner, *Magn. Reson. Mater. Phys., Biol. Med.*, 2014, **27**, 195–199.



- 19 D. Canet, C. Aroulanda, P. Mutzenhardt, S. Aime, R. Gobetto and F. Reineri, *Concepts Magn. Reson., Part A*, 2006, **28**, 321–330.
- 20 P. M. Richardson, R. O. John, A. J. Parrott, P. J. Rayner, W. Iali, A. Nordon, M. E. Halse and S. B. Duckett, *Phys. Chem. Chem. Phys.*, 2018, **20**, 26362–26371.
- 21 C. R. Bowers and D. P. Weitekamp, *Phys. Rev. Lett.*, 1986, **57**, 2645.
- 22 C. R. Bowers and D. P. Weitekamp, *J. Am. Chem. Soc.*, 1987, **109**, 5541–5542.
- 23 T. C. Eisenschmid, R. U. Kirss, P. P. Deutsch, S. I. Hommeltoft, R. Eisenberg, J. Bargon, R. G. Lawler and A. L. Balch, *J. Am. Chem. Soc.*, 1987, **109**, 8089–8091.
- 24 R. Eisenberg, *Acc. Chem. Res.*, 1991, **24**, 110–116.
- 25 T. C. Eisenschmid, R. U. Kirss, P. P. Deutsch, S. I. Hommeltoft, R. Eisenberg, J. Bargon, R. G. Lawler and A. L. Balch, *J. Am. Chem. Soc.*, 1987, **109**, 8089–8091.
- 26 S. B. Duckett, C. L. Newell and R. Eisenberg, *J. Am. Chem. Soc.*, 1994, **116**, 10548–10556.
- 27 K. V. Kovtunov, I. E. Beck, V. I. Bukhtiyarov and I. V. Koptuyug, *Angew. Chem., Int. Ed.*, 2008, **47**, 1492–1495.
- 28 P. Bhattacharya, E. Y. Chekmenev, W. H. Perman, K. C. Harris, A. P. Lin, V. A. Norton, C. T. Tan, B. D. Ross and D. P. Weitekamp, *J. Magn. Reson.*, 2007, **186**, 150–155.
- 29 N. M. Zacharias, H. R. Chan, N. Sailasuta, B. D. Ross and P. Bhattacharya, *J. Am. Chem. Soc.*, 2011, **134**, 934–943.
- 30 F. Reineri, T. Boi and S. Aime, *Nat. Commun.*, 2015, **6**, 5858.
- 31 R. W. Adams, J. A. Aguilar, K. D. Atkinson, M. J. Cowley, P. I. Elliott, S. B. Duckett, G. G. Green, I. G. Khazal, J. López-Serrano and D. C. Williamson, *Science*, 2009, **323**, 1708–1711.
- 32 R. W. Adams, S. B. Duckett, R. A. Green, D. C. Williamson and G. G. Green, *J. Chem. Phys.*, 2009, **131**, 194505.
- 33 A. J. Ruddlesden, R. E. Mewis, G. G. Green, A. C. Whitwood and S. B. Duckett, *Organometallics*, 2015, **34**, 2997–3006.
- 34 A. J. Ruddlesden and S. B. Duckett, *Chem. Commun.*, 2016, **52**, 8467–8470.
- 35 L. S. Lloyd, A. Asghar, M. J. Burns, A. Charlton, S. Coombes, M. J. Cowley, G. J. Dear, S. B. Duckett, G. R. Genov and G. G. R. Green, *Catal. Sci. Technol.*, 2014, **4**, 3544–3554.
- 36 K. M. Appleby, R. E. Mewis, A. M. Olaru, G. G. R. Green, I. J. S. Fairlamb and S. B. Duckett, *Chem. Sci.*, 2015, **6**, 3981–3993.
- 37 A. L. Sargent and M. B. Hall, *Inorg. Chem.*, 1992, **31**, 317–321.
- 38 M. Fekete, O. Bayfield, S. B. Duckett, S. Hart, R. E. Mewis, N. Pridmore, P. J. Rayner and A. Whitwood, *Inorg. Chem.*, 2013, **52**, 13453–13461.
- 39 M. J. Cowley, R. W. Adams, K. D. Atkinson, M. C. Cockett, S. B. Duckett, G. G. Green, J. A. Lohman, R. Kerssebaum, D. Kilgour and R. E. Mewis, *J. Am. Chem. Soc.*, 2011, **133**, 6134–6137.
- 40 H. Zeng, J. Xu, J. Gillen, M. T. McMahon, D. Artemov, J.-M. Tyburn, J. A. Lohman, R. E. Mewis, K. D. Atkinson and G. G. Green, *J. Magn. Reson.*, 2013, **237**, 73–78.
- 41 E. B. Dücker, L. T. Kuhn, K. Münnemann and C. Griesinger, *J. Magn. Reson.*, 2012, **214**, 159–165.
- 42 M. L. Truong, T. Theis, A. M. Coffey, R. V. Shchepin, K. W. Waddell, F. Shi, B. M. Goodson, W. S. Warren and E. Y. Chekmenev, *J. Phys. Chem. C*, 2015, **119**, 8786–8797.
- 43 P. Spanning, I. Reile, M. Emondts, P. P. Schleker, N. K. Hermkens, N. G. van der Zwaluw, B. J. van Weerdenburg, P. Tinnemans, M. Tessari and B. Blümich, *Chem. – Eur. J.*, 2016, **22**, 9277–9282.
- 44 R. E. Mewis, R. A. Green, M. C. Cockett, M. J. Cowley, S. B. Duckett, G. G. Green, R. O. John, P. J. Rayner and D. C. Williamson, *J. Phys. Chem. B*, 2015, **119**, 1416–1424.
- 45 W. Iali, P. J. Rayner, A. Alshehri, A. J. Holmes, A. J. Ruddlesden and S. B. Duckett, *Chem. Sci.*, 2018, **9**, 3677–3684.
- 46 W. Iali, S. S. Roy, B. J. Tickner, F. Ahwal, A. J. Kennerley and S. B. Duckett, *Angew. Chem.*, 2019, **131**, 10377–10381.
- 47 A. F. Oliveri, L. A. Wills, C. R. Hazlett, M. E. Carnes, I.-Y. Chang, P. H.-Y. Cheong and D. W. Johnson, *Chem. Sci.*, 2015, **6**, 4071–4085.
- 48 L. Dadci, H. Elias, U. Frey, A. Hoernig, U. Koelle, A. E. Merbach, H. Paulus and J. S. Schneider, *Inorg. Chem.*, 1995, **34**, 306–315.
- 49 S. Ogo, N. Makihara, Y. Kaneko and Y. Watanabe, *Organometallics*, 2001, **20**, 4903–4910.
- 50 S. Knecht, S. Hadjiali, D. A. Barskiy, A. Pines, G. Sauer, A. S. Kiryutin, K. L. Ivanov, A. V. Yurkovskaya and G. Buntkowsky, *J. Phys. Chem. C*, 2019, **123**, 16288–16293.
- 51 B. J. Tickner, R. O. John, S. S. Roy, S. J. Hart, A. C. Whitwood and S. B. Duckett, *Chem. Sci.*, 2019, **10**, 5235–5245.
- 52 A. M. Olaru, A. Burt, P. J. Rayner, S. J. Hart, A. C. Whitwood, G. G. Green and S. B. Duckett, *Chem. Commun.*, 2016, **52**, 14482–14485.
- 53 B. J. Tickner, W. Iali, S. S. Roy, A. C. Whitwood and S. B. Duckett, *ChemPhysChem*, 2019, **20**, 241–245.
- 54 B. E. Hauger, D. Gusev and K. G. Caulton, *J. Am. Chem. Soc.*, 1994, **116**, 208–214.
- 55 O. Semenova, P. M. Richardson, A. J. Parrott, A. Nordon, M. E. Halse and S. B. Duckett, *Anal. Chem.*, 2019, **91**, 6695–6701.
- 56 P. Zhou, A. A. Vitale, J. San Filippo Jr. and W. H. Saunders Jr., *J. Am. Chem. Soc.*, 1985, **107**, 8049–8054.
- 57 F. Abu-Hasanayn, A. S. Goldman and K. Krogh-Jespersen, *J. Phys. Chem.*, 1993, **97**, 5890–5896.
- 58 B. J. Tickner, R. O. John, S. S. Roy, S. Hart, A. C. Whitwood and S. B. Duckett, *Chem. Sci.*, 2019, **10**, 5235–5245.
- 59 S. S. Roy, K. M. Appleby, E. J. Fear and S. B. Duckett, *J. Phys. Chem. Lett.*, 2018, **9**, 1112–1117.
- 60 H. Sidhu, R. Gupta, S. K. Thind and R. Nath, *Ann. Nutr. Metab.*, 1987, **31**, 354–361.
- 61 C. Laustsen, G. Pileio, M. C. D. Tayler, L. J. Brown, R. C. D. Brown, M. H. Levitt and J. H. Ardenkjaer-Larsen, *Magn. Reson. Med.*, 2012, **68**, 1262–1265.
- 62 N. Hao, E. Shen, Y. G. Li, E. B. Wang, C. W. Hu and L. Xu, *Eur. J. Inorg. Chem.*, 2004, **2004**, 4102–4107.



- 63 B. Modec, J. V. Brenčič and J. Koller, *Eur. J. Inorg. Chem.*, 2004, **2004**, 1611–1620.
- 64 M. Gruselle, C. Train, K. Boubekeur, P. Gredin and N. Ovanesyan, *Coord. Chem. Rev.*, 2006, **250**, 2491–2500.
- 65 B. Modec, J. V. Brenčič, D. Dolenc and J. Zubieta, *Dalton Trans.*, 2002, 4582–4586.
- 66 M. E. Robinson, J. E. Mizzi, R. J. Staples and R. L. LaDuca, *Cryst. Growth Des.*, 2015, **15**, 2260–2271.
- 67 L. D. Vazquez-Serrano, B. T. Owens and J. M. Buriak, *Inorg. Chim. Acta*, 2006, **359**, 2786–2797.
- 68 B. J. Tickner, W. Iali, S. S. Roy, A. C. Whitwood and S. B. Duckett, *ChemPhysChem*, 2019, **20**, 241–245.

