



Cite this: *RSC Adv.*, 2024, 14, 21174

Received 16th April 2024  
Accepted 23rd June 2024

DOI: 10.1039/d4ra02833c

rsc.li/rsc-advances

# An efficient alternative to DBU in the oxathiaphospholane (OTP) method for the solid phase synthesis of P-stereodefined phosphorothioate analogs†

Katarzyna Jastrzębska \*

This study presents a modified (extended) 1,3,2-oxathiaphospholane (OTP) method for the synthesis of P-stereodefined phosphorothioate analogs in the presence of previously unused organic bases. TBD (5,7-triazabicyclo[4.4.0]dec-5-ene) and Verkade's proazaphosphatane (2,8,9-trimethyl-2,5,8,9-tetraaza-1-phosphabicyclo[3.3.3]undecane) are herein used for the first time as efficient organic bases compared to DBU, which is commonly used in the OTP approach.

## Introduction

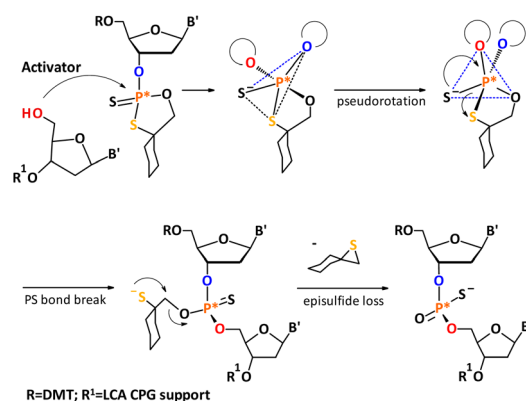
Only a few methods have been developed for stereocontrolled synthesis of phosphorothioate oligonucleotides, including methods that utilize *N*-acyl oxazaphospholidine<sup>1</sup> (Beaucage *et al.*), oxazaphosphorinane<sup>2</sup> (Just *et al.*) and bicyclic oxazaphospholidine<sup>3</sup> (Agrawal *et al.*) derivatives. Wada and co-workers have developed a methodology to synthesize stereochemically regular PS-ODNs using diastereopure nucleoside 3'-*O*-(1,3,2-oxazaphospholidine) monomers (originally proposed by Agrawal *et al.*).<sup>4</sup> The first method for stereocontrolled chemical synthesis of phosphorothioate oligonucleotides, which was developed by Stec *et al.*, is based on a chemistry employing P-diastereomerically pure monomers possessing the 2-thio-1,3,2-oxathiaphospholane moiety.<sup>5</sup> Their applicability and limitations have been summarized in several reports.<sup>6</sup> Recently, Baran *et al.* described a method using a citrus-derived PSI reagent.<sup>7</sup>

## General mechanistic principle of the OTP method

In the OTP method, the P-chiral nucleoside 3'-*O*-(2-thio-1,3,2-oxathiaphospholane) monomers are prepared as a mixture of both P-isomers, which can be separated by silica gel column chromatography. Separated P-diastereomers may be used for the synthesis of diastereomerically pure fragments of phosphorothioate oligonucleotides. In this synthesis, the

oxathiaphospholane reacts with the 5'-OH group of a nucleoside (or a growing oligonucleotide attached to the solid support) in the presence of 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU). This leads to an intermediate, which undergoes pseudorotation (permutational isomerization), followed by the elimination of the episulfide to form an internucleotide phosphorothioate linkage (Scheme 1).<sup>8</sup> In the synthesis of PS-DNA oligomers, repetitive yields in the range of 92–94% are usually observed, making the synthesis of oligomers longer than 12–14 nucleotides problematic; although successful but very low yields of 23-mers have also been reported.<sup>9</sup>

The OTP approach promoted by DBU has been successfully used for the stereocontrolled synthesis of phosphorothioate oligonucleotides and their analogues. For instance, in our laboratory, the OTP method was also used for the stereocontrolled synthesis of PS-LNA analogs (Locked Nucleic Acids)<sup>10</sup>



**Scheme 1** Synthesis of P-stereodefined phosphorothioate analogs developed by Stec *et al.* A mechanism of DBU-promoted condensation step.

Centre of Molecular and Macromolecular Studies, Polish Academy of Sciences, Department of Bioorganic Chemistry, Sienkiewicza 112, 90-363 Łódź, Poland.  
E-mail: katarzyna.jastrzebska@cbmm.lodz.pl

† Electronic supplementary information (ESI) available. See DOI: <https://doi.org/10.1039/d4ra02833c>



and PS-GNA analogs (Glycol Nucleic Acids).<sup>11</sup> Recently, there has been successful contribution to research in the field of P-diastereomerically pure PS-RNA and PS-(2'-OMe) RNA derivatives.<sup>12</sup>

The constantly increasing number of approved nucleic acid therapeutics shows their great potential.<sup>13</sup> It is necessary to provide more detailed information on the influence of the absolute configuration of the phosphorus atoms in P-stereodefined oligonucleotides. The studies carried out so far have shown their potential. It has been demonstrated that the configuration of the phosphorus atom has an influence on biological activity when interacting with biomolecules.  $R_P$ -PS linkages are preferred for duplex formation with complementary ORN strands,<sup>14</sup> while  $S_P$ -PS linkages are chosen to increase stability towards the 3'-exonuclease present in the human plasma.<sup>15</sup> PS DNA in which both the  $R_P$  and  $S_P$ -PS linkages are properly arranged would therefore be better drug candidates than those synthesized in a non-stereocontrolled manner. Therefore, the stereoselective synthesis of such compounds is an important goal to achieve. In view of the growing interest of molecular biologists, biochemists, as well as pharmaceutical companies in the synthesis of modified oligonucleotides and their therapeutic application,<sup>16</sup> more efficient organic bases must be considered.

## Results and discussion

### Novel and efficient organic bases in OTP method

In the search for new organic bases, we focused on strong, yet non-nucleophilic bases. We found bases consisting of nitrogenated bases and proazaphosphatranes (Fig. 1).

Here we discovered that TBD (1,5,7-triazabicyclo[4.4.0]dec-5-ene) and one of the Verkade bases, 2,8,9-trimethyl-2,5,8,9-tetraaza-1-phosphabicyclo[3.3.3]undecane, can be used as alternatives and efficient bases for the 1,3,2-oxathiaphospholane ring-opening condensation (Scheme 2).

TBD is a strong bicyclic guanidine base ( $pK_a$  in acetonitrile: 26.0)<sup>18</sup> with a steric hindrance and the ability, when protonated, to delocalize the positive charge between amino groups and the

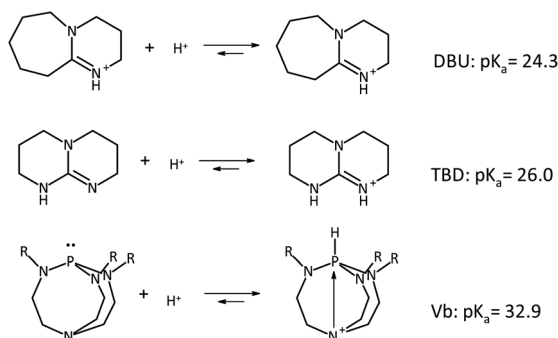
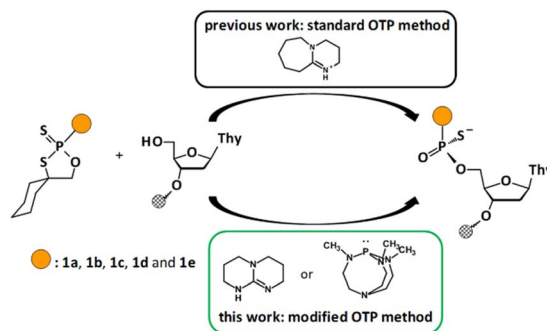


Fig. 1 Non-nucleophilic bases used in this study. DBU (1,8-diazabicyclo[5.4.0]undec-7-ene), TBD (1,5,7-triazabicyclo[4.4.0]dec-5-ene), Verkade base (Vb, in this study: R = methyl group, 2,8,9-trimethyl-2,5,8,9-tetraaza-1-phosphabicyclo[3.3.3]undecane), their protonated molecules and  $pK_a$ s in acetonitrile.<sup>17</sup>



Scheme 2 Synthesis of dinucleoside phosphorothioate  $1_{p5}T$  by the oxathiaphospholane approach.

double bond. Because of their strong basicity and relatively weak nucleophilicity, proazaphosphatranes have also attracted attention. Proazaphosphatranes bases have been used in multiple types of catalytic processes for organic synthesis.<sup>19</sup> Pioneered by Verkade *et al.*,<sup>20</sup> the 'molecular football' shaped compound is commonly known as Verkade's superbase (in this paper labeled as Vb).

The term 'superbase' is given to organic compounds whose basicity is greater than that of 1,8-bis(dimethylamino)naphthalene (proton sponge),<sup>21</sup> with  $pK_a$  of 18.6 (conjugate acid form) in acetonitrile. The  $pK_a$  of the protonated form of proazaphosphatranes ranges from 31.6 to 33.6 in acetonitrile, depending on the substituents attached to the equatorial nitrogen atoms R (in this study R = Me). The conjugate acid  $[\text{HP}(\text{MeNCH}_2\text{CH}_2)_3\text{N}]^+$  has a  $pK_a$  of 32.9 in acetonitrile;<sup>22</sup> thus, this base is by eight orders of magnitude stronger than DBU.<sup>23</sup>

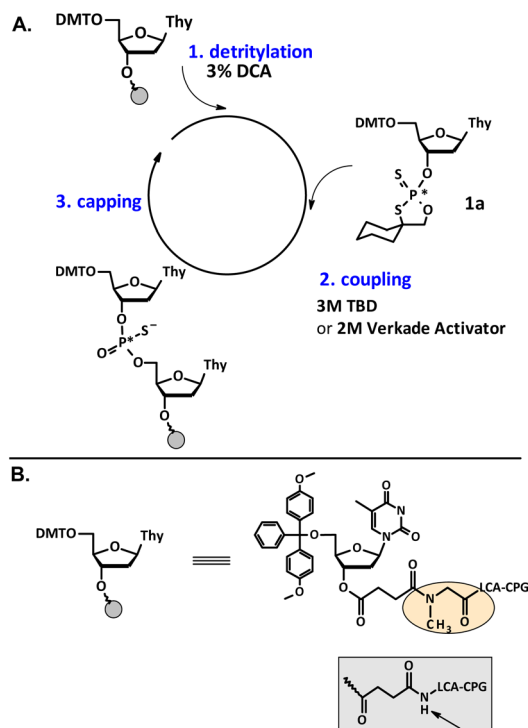
### General protocol of the OTP method

Synthesis of P-stereodefined oligonucleotides by oxathiaphospholane method was carried out manually at a 1  $\mu\text{mol}$  scale, according to the previously published protocols. This method uses a CPG-support (controlled pore glass, LCAA CPG 500 Å, ChemGenes, USA), to which a 5'-O-DMT-nucleoside unit (in this study: loading 35  $\mu\text{mol g}^{-1}$ ) is attached with a succinyl-sarcosinyl type linker (Scheme 3). To prevent the DBU-promoted intramolecular cleavage of the standard LCAA (long chain alkylamino) linker, a modified sarcosine linker  $[-\text{COCH}_2\text{CH}_2\text{CON}(\text{CH}_3)\text{CH}_2\text{CO-LCAA-CPG}]$  is used.<sup>24</sup> Coupling steps are carried out using 20 mg of 3'-O-OTP-derivatives of the standard 5'-O-DMT-protected nucleosides in the presence of 1 M DBU in acetonitrile. Manual preparation (without the synthesizer) is considered demanding in terms of the preparation of reagents and technical skills during the oligonucleotide synthesis. Our previous observations indicate that some parameters, such as amount of monomer, coupling time, and base concentration play a major role in determining the coupling yield during synthesis.

### UV monitoring of the solid-phase synthesis efficacy

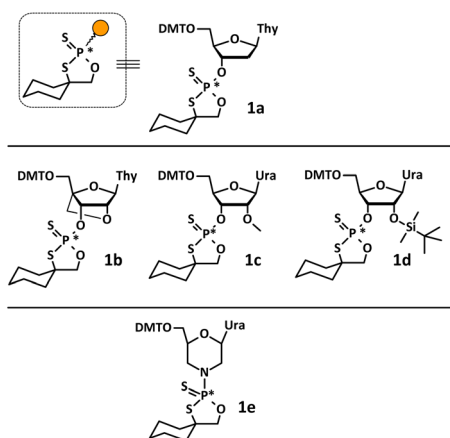
The efficiency of solid phase synthesis was studied by condensation of **1a-e** with thymidine (attached to the solid support).





**Scheme 3** (A) Synthetic cycle for the preparation of oligonucleotide fragment from **1a**; (B) structures of sarcosine linker and linker with no sarcosine are highlighted in light orange and grey, respectively.

We analyzed the coupling efficiency of the set of oxathiaphospholane monomers **1** (as a mixture of P-diastereomers) in the presence of novel organic non-nucleophilic bases. The experimental protocol involves use of previously tested 5'-O-DMT-protected 3'-O-(2-thio-“spiro”-4,4-pentamethylene-1,3,2-oxathiaphospholane) derivatives, *i.e.* 2'-deoxynucleoside (**1a**), locked nucleic acid (**1b**), 2'-O-methyl ribonucleoside (**1c**), and 2'-O-TBDMS-ribonucleoside (**1d**) monomers. It should be noted that previously unknown oxathiaphospholane derivatives of



**Fig. 2** All OTP monomers: DNA (**1a**), LNA (**1b**), 2'-OMe (**1c**), 2'-OTBDMS (**1d**), morpholino (**1e**) as a mixture of P-diastereomers used in this study.

morpholino nucleosides (**1e**)<sup>25</sup> were also synthesized and used in this modified OTP protocol (Fig. 2).

Detailed procedure for synthesis of mU-OTP is presented in the ESI.† For all synthesized products the same support (which possessed the thymidine 3'-O-attached to LCA CPG linker with loading 35  $\mu\text{mol g}^{-1}$ ) was used to eliminate any possible effect of support variability. The coupling efficacy is controlled by measurement of absorption at 504 nm coming from the released DMT<sup>+</sup> cation.

The efficiency of monomer DNA condensation in the standard OTP procedure using 1 M DBU (50-fold molar excess) is up to 95%. Attempts to use the same concentrations of novel bases did not lead to satisfactory results. When using 3 M TBD (150-fold molar excess) and 2 M Verkade base (25-fold molar excess), the efficacy was 93% (Table 1 and Fig. S4–18, ESI†). Even with double coupling, we did not observe a significantly increased yield of condensation. This result suggests that TBD and Verkade base can be successfully used alternatively.

Working earlier with LNA-derived OTP monomers using the standard DBU-assisted OTP method, we observed that in the cycles where LNA-OTP monomers were incorporated, double coupling was necessary (20 mg + 20 mg, ~25-fold molar excess) and in both steps the coupling time was extended to 20 min (Table 1 and Fig. S1, ESI†). Although the repetitive average yield of the coupling (88%, based on the DMT<sup>+</sup> cation assay) was relatively low as compared with DNA OTP (**1a**), it was considered to be satisfactory enough to attempt synthesis of chimeric P-stereodefined PS-(DNA/LNA). In the case of 3 M TBD solution in acetonitrile, the yield of a single coupling step of LNA monomer was approximately 65%. Thus, it was necessary to use double coupling (20 mg + 20 mg of OTP monomer **1b**) and the coupling time was extended to 20 minutes. Repetitive yield, calculated from the DMT<sup>+</sup> cation assay, was 86%. It is noted that the repetitive yield of condensation reduced when using 3 M TBD and 2 M Vb (86%, assessed by the DMT<sup>+</sup> cation assay) despite the use of double coupling. In the case of synthesizing LNA oligonucleotides, 1 M DBU is still the best option. In the next step, the 2'-OMe monomer (**1c**) was investigated. In standard protocol with 1 M DBU, only *ca.* 84% repetitive yield was achieved (based on a DMT<sup>+</sup> cation assay), even with double coupling. Additional experiments using 3 M TBD revealed

**Table 1** Coupling yields with OTP monomers in solid phase synthesis of the dimers calculated from the DMT<sup>+</sup> decay assay

OTP monomer	Standard OTP method	Modified OTP method	
	DBU <sup>a</sup>	TBD <sup>b</sup>	Verkade <sup>c</sup>
DNA ( <b>1a</b> )	95	93	93
LNA ( <b>1b</b> )	88 <sup>d</sup>	86 <sup>d</sup>	86 <sup>d</sup>
2'-OMe ( <b>1c</b> )	84 <sup>d</sup>	86 <sup>d</sup>	83 <sup>d</sup>
2'-OTBDMS ( <b>1d</b> )	91 <sup>d</sup>	92 <sup>d</sup>	92
mU ( <b>1e</b> )	61 <sup>d</sup>	93 <sup>d</sup>	95

<sup>a</sup> 1 M DBU solution in anhydrous acetonitrile. <sup>b</sup> 2 M TBD solution in anhydrous acetonitrile. <sup>c</sup> 3 M Verkade solution in anhydrous acetonitrile. <sup>d</sup> Double coupling.



slightly higher repetitive yield. The dimer was obtained at 86% yield as assessed from the DMT<sup>+</sup> cation assay. The coupling yield for **1c** using the Verkade base was unsatisfactory. Even with double coupling, only *ca.* 83% repetitive yield was achieved (based on a DMT<sup>+</sup> cation assay). Attempts to increase the yield by extension of the coupling time or by increasing Verkade base concentration gave similar results and were unsuccessful. Therefore, only the 3 M TBD can be used interchangeably with DBU in the 2'-OMe monomer condensation. Next, a derivative of 2'-OTBDMS (**1d**) was used and resulted in a 91% repetitive yield (based on a DMT<sup>+</sup> cation assay) using the standard OTP method. A significant increase of repetitive yield of condensation for double coupling using TBD (92%, assessed by the DMT<sup>+</sup> cation assay) was observed. The coupling of **1d** monomer using Verkade base was also achieved in a higher yield (92% based on a DMT<sup>+</sup> cation assay) than that of 1 M DBU.

The most interesting results obtained were those for the morpholino monomer (**1e**). At first, the standard protocol with 1 M DBU was used, and double coupling at the condensation steps led to a significant drop in coupling efficiency and a 61% repetitive yield was noted (based on a DMT<sup>+</sup> cation assay). There was no improvement with a longer reaction time. Attempts to increase the yields by extending the coupling time or increasing the DBU concentration were also unsuccessful. In the case of 3 M TBD solution in acetonitrile, the yield of a single coupling step was approximately 85%; therefore, a double coupling (20 mg + 20 mg of the OTP monomer **1e**) was required and the coupling time was extended to 20 min. Repetitive yield, calculated from the DMT<sup>+</sup> cation assay, was 93%.

We noted a significantly increased repetitive yield of condensation for 2 M Vb (95%, assessed by the DMT<sup>+</sup> cation assay). It should be emphasized that a single coupling is sufficient in this case. The data in Table 1 show that the initial condensations with a 2'-deoxy nucleoside bound to a solid support proceeded with the required 90+% efficacy only for monomers **1a**, **1d** and **1e**, while **1b** and **1c** offered a lower but acceptable coupling yield.

Next, longer analogs were synthesized. The yield of the first coupling step of 5T\_T and 5T\_V (as a mixture of P-diastereomers) with the 5'-OH group of a deoxyribonucleoside attached to a solid support was acceptable (93%) and the repetitive yields associated with the subsequent coupling steps were equally high (Table 2, Fig. S19 and 20, ESI†).

**Table 2** Average yield of the four consecutive coupling steps (%) in synthesis of pentamers T<sub>PS</sub>T<sub>PS</sub>T<sub>PS</sub>T<sub>PS</sub>T 5T\_T and 5T\_V using 3 M TBD and 2 M Verkade, respectively (calculated from the DMT<sup>+</sup> decay assay)

The coupling of OTP monomer DNA ( <b>1a</b> )	Modified otp method	
	5T_T 3 M TBD	5T_V 2 M Verkade
1st coupling	93	93
2nd coupling	93	92
3rd coupling	92	91
4th coupling	93	90

The experiments with the application of new bases to the synthesis of P-stereodefined phosphorothioate dinucleosides were performed. The pure diastereomer, fast-eluted of **1a**, reacted with thymidine (attached to the solid support) to yield the diastereomerically pure 5'-OH-T<sub>PS</sub>T-OH, and the diastereomeric purity was confirmed by HPLC analysis (Fig. S21–25, ESI†). The above experiments confirm that the tested bases can be used successfully for the highly stereoselective synthesis of P-chiral phosphorothioate analogs.

## Conclusions

In conclusion, our studies highlight a new route for the development of the OTP method by applying novel bases. We have discovered TBD (1,5,7-triazabicyclo[4.4.0]dec-5-ene) and the simplest Verkade base, *i.e.* 2,8,9-trimethyl-2,5,8,9-tetraaza-1-phosphabicyclo[3.3.3]undecane, which have been shown to be efficient bases in the 1,3,2-oxathiaphospholane ring-opening condensation. Optimization of solid-phase synthesis of phosphorothioate dinucleotides was performed by condensation of monomers **1a–e** (as a mixture of P-diastereomers) with 5'-OH-thymidine in the presence of various non-nucleophilic bases: DBU, TBD and Vb. Based on experiments, the optimal concentrations of TBD and Verkade base required to achieve maximum yields in solid phase synthesis were determined to be 3 and 2 M, respectively. Oligonucleotide synthesis with the application of TBD as a base is an economical alternative. This choice is mainly due to the inexpensiveness of TBD. However, it should be noted that a double coupling (increased consumption of handmade monomers) must be included in the synthetic protocol in this case. In the second procedure for the synthesis of P-stereodefined phosphorothioate analogs using Verkade solution, we observed a 93% yield with a single coupling using OTP. However, the Verkade base-activated coupling is more expensive. From the above-mentioned results, it may be concluded that the formation of internucleotide phosphorothioate linkage involving derivative **1** *via* the oxathiaphospholane route with novel bases is efficient, stereocontrolled, and with stereoselectivity.

Further studies are underway to adapt the modified oxathiaphospholane (OTP) approach to the synthesis of P-stereodefined phosphorothioate oligonucleotides.

## Author contributions

K. J. supervised the project, prepared and edited the manuscript, as well as performed all the chemical synthesis and data analysis.

## Data availability

The data supporting this article have been included as part of the ESI.†

## Conflicts of interest

There are no conflicts of interest to declare.





## Acknowledgements

This work was financially supported by National Centre of Science, Poland, grant UMO-2021/43/D/ST4/02433 (to K. J.). An Avance Neo 400 NMR spectrometer was purchased using funds provided by the EU Regional Operational Program of the Lodz Region RPLD.01.01.00-10-0008/18.

## Notes and references

- 1 A. Wilk, A. Grajkowski, L. R. Phillips and S. L. Beaucage, Deoxyribonucleoside Cyclic N-Acylphosphoramidites as a New Class of Monomers for the Stereocontrolled Synthesis of Oligothymidylyl- and Oligodeoxycytidylyl-Phosphorothioates, *J. Am. Chem. Soc.*, 2000, **122**, 2149.
- 2 (a) Y. Jin, G. Biancotto and G. Just, A stereoselective synthesis of dinucleotide phosphorothioates, using chiral phosphoramidites as intermediates, *Tetrahedron Lett.*, 1996, **37**, 973–976; (b) J.-C. Wang and G. Just, A stereoselective synthesis of dinucleotide phosphorothioates, using chiral indol-oxazaphosphorine intermediates, *Tetrahedron Lett.*, 1997, **38**, 3797–3800.
- 3 M. Guo, D. Yu, R. P. Iyer and S. Agrawal, Solid-phase stereoselective synthesis of 2'-O-methyloligoribonucleoside phosphorothioates using nucleoside bicyclic oxazaphospholidines, *Bioorg. Med. Chem. Lett.*, 1998, 2539–2544.
- 4 (a) N. Oka and T. Wada, Stereocontrolled synthesis of oligonucleotide analogs containing chiral internucleotidic phosphorus atoms, *Chem. Soc. Rev.*, 2011, **40**, 5829–5843; (b) N. Oka, T. Wada and K. Saigo, An Oxazaphospholidine Approach for the Stereocontrolled Synthesis of Oligonucleoside Phosphorothioates, *J. Am. Chem. Soc.*, 2003, **125**, 8307–8317; (c) N. Oka, M. Yamamoto, T. Sato and T. Wada, Solid-Phase Synthesis of Stereoregular Oligodeoxyribonucleoside Phosphorothioates Using Bicyclic Oxazaphospholidine Derivatives as Monomer Units, *J. Am. Chem. Soc.*, 2008, **130**, 16031–16037.
- 5 (a) W. J. Stec, A. Grajkowski, M. Koziolkiewicz and B. Uznański, Novel route to oligo(deoxyribonucleoside phosphorothioates). Stereocontrolled synthesis of P-chiral oligo(deoxyribonucleoside phosphorothioates), *Nucleic Acids Res.*, 1991, **19**, 5883–5888; (b) W. J. Stec, B. Karwowski, M. Boczkowska, P. Guga, M. Koziolkiewicz, M. Sochacki, M. W. Wieczorek and J. Błaszczyk, Deoxyribonucleoside 3'-O-(2-Thio- and 2-Oxo-“spiro”-4,4-pentamethylene-1,3,2-oxathiaphospholane)s: Monomers for Stereocontrolled Synthesis of Oligo(deoxyribonucleoside phosphorothioate)s and Chimeric PS/PO Oligonucleotide, *J. Am. Chem. Soc.*, 1998, **120**, 7156–7167.
- 6 (a) P. Guga, A. Okruszek and W. J. Stec, Recent Advances in Stereocontrolled Synthesis of P-Chiral Analogues of Biophosphates, *Top. Curr. Chem.*, 2002, **220**, 169–200; (b) B. Nawrot, B. Rębowska, B. Michalak, M. Bulkowski, D. Błaziak, P. Guga and W. J. Stec, 1,3,2-Oxathiaphospholane approach to the synthesis of P-chiral stereodefined analogs of oligonucleotides and biologically relevant nucleoside polyphosphates, *Pure Appl. Chem.*, 2008, **80**, 1859–1871.
- 7 (a) K. W. Knouse, J. N. deGruyter, M. A. Schmidt, B. Zheng, J. C. Vantourout, C. Kingston, S. E. Mercer, I. M. McDonald, R. E. Olson, Y. Zhu, Ch. Hang, J. Zhu, Ch. Yuan, Q. Wang, P. Park, M. D. Eastgate and P. S. Baran, Unlocking P(V): Reagents for chiral phosphorothioate synthesis, *Science*, 2018, **361**, 1234–1238; (b) Y. Huang, K. W. Knouse, S. Qiu, W. Hao, N. M. Padial, J. C. Vantourout, B. Zheng, S. E. Mercer, J. Lopez-Ogalla, R. Narayan, R. E. Olson, D. G. Blackmond, M. D. Eastgate, M. A. Schmidt, I. M. McDonald and P. S. Baran, A P(V) platform for oligonucleotide synthesis, *Science*, 2021, **373**, 1265–1270.
- 8 (a) T. Uchimar, W. J. Stec and K. Taira, Mechanism of the Chemoselective and Stereoselective Ring Opening of Oxathiaphospholanes: An *Ab Initio* Study, *J. Org. Chem.*, 1997, **62**, 5793–5800; (b) K. Misiura, D. Szymanowicz, M. Olesiak and W. J. Stec, DBU-assisted 1,3,2-oxathiaphospholane ring-opening condensation with selected O-, S-, N- and C-nucleophiles, *Tetrahedron Lett.*, 2004, **45**, 4301–4305.
- 9 T. Inagawa, H. Naksima, B. Karwowski, P. Guga, W. J. Stec, H. Takeuchi and H. Takaku, Inhibition of human immunodeficiency virus type 1 replication by P-stereodefined oligo(nucleoside phosphorothioate)s in a long-term infection model, *FEBS Lett.*, 2002, **528**, 48–52.
- 10 (a) K. Jastrzębska, A. Maciaszek, R. Dolot, G. Bujacz and P. Guga, Thermal Stability and Conformation of Antiparallel Duplexes Formed by P-Stereodefined Phosphorothioate DNA/LNA Chimeric Oligomers with DNA and RNA Matrices, *Org. Biomol. Chem.*, 2015, **13**, 10032–10040; (b) K. Jastrzębska, B. Mikołajczyk and P. Guga, LNA units present in [R<sub>p</sub>-PS]-(DNA#LNA) chimeras enhance the thermal stability of parallel duplexes and triplexes formed with (2'-OMe)-RNA strands, *RSC Adv.*, 2020, **10**, 22370–22376.
- 11 A. Tomaszewska-Antczak, K. Jastrzębska, A. Maciaszek, B. Mikołajczyk and P. Guga, P-Stereodefined phosphorothioate analogs of glycol nucleic acids – synthesis and structural properties, *RSC Adv.*, 2018, **8**, 24942–24952.
- 12 K. Jastrzębska, A. Maciaszek, R. Dolot, A. Tomaszewska-Antczak, B. Mikołajczyk and P. Guga, Synthesis and hybridizing properties of P-stereodefined chimeric [PS]-{DNA:RNA} and [PS]-{DNA:(2'-OMe)-RNA} oligomers, *RSC Adv.*, 2022, **12**, 26815–26824.
- 13 C. A. Stein and D. Castanotto, FDA-Approved Oligonucleotide Therapies in 2017, *Mol. Ther.*, 2017, **25**, 1069–1075.
- 14 M. Koziolkiewicz, A. Krakowiak, M. Kwinkowski, M. Boczkowska and W. J. Stec, Stereodifferentiation – the effect of P chirality of oligo(nucleoside phosphorothioates) on the activity of bacterial RNase H, *Nucleic Acids Res.*, 1995, **23**, 5000–5005.
- 15 M. Koziolkiewicz, M. Wójcik, A. Kobylańska, B. Karwowski, B. Rębowska, P. Guga and W. J. Stec, Stability of stereoregular oligo(nucleoside phosphorothioate)s in



- human plasma: diastereoselectivity of plasma 3'-exonuclease, *Antisense Nucleic Acid Drug Dev.*, 1997, **7**, 43–48.
- 16 (a) D. Collotta, I. Bertocchi, E. Chiapello and M. Collino, Antisense oligonucleotides: a novel Frontier in pharmacological strategy, *Front. Pharmacol.*, 2023, **14**, 1304342; (b) J. A. Kulkarni, D. Witzigmann, S. B. Thomson, S. Chen, B. R. Leavitt, P. R. Cullis and R. van der Meel, The current landscape of nucleic acid therapeutics, *Nat. Nanotechnol.*, 2021, **16**, 630–643; (c) B. T. Le, S. Paul, K. Jastrzebska, H. Langer, M. H. Caruthers and R. N. Veedu, Thiomorpholino oligonucleotides as a robust class of next generation platforms for alternate mRNA splicing, *Proc. Natl. Acad. Sci. U. S. A.*, 2022, **119**, e2207956119.
- 17 J. E. Taylor, S. D. Bull and J. M. Williams, Amidines, isothioureas, and guanidines as nucleophilic catalysts, *Chem Soc Rev*, 2012, **41**, 2109.
- 18 E. Fritz-Langhals, Unique Superbase TBD (1,5,7-Triazabicyclo[4.4.0]dec-5-ene): From Catalytic Activity and One-Pot Synthesis to Broader Application in Industrial Chemistry, *Org. Process Res. Dev.*, 2022, **26**(11), 3015–3023.
- 19 J. G. Verkade and P. B. Kisanga, Proazaphosphatranes: a synthesis methodology trip from their discovery to vitamin A, *Tetrahedron*, 2003, **59**, 7819–7858.
- 20 C. Lensink, S. K. Xi, L. M. Daniels and J. G. Verkade, The unusually robust phosphorus-hydrogen bond in the novel cation [cyclic] HP(NMeCH<sub>2</sub>CH<sub>2</sub>)<sub>3</sub>N<sup>+</sup>, *J. Am. Chem. Soc.*, 1989, **111**, 3478–3479.
- 21 T. R. Puleo, S. J. Sujansky, S. E. Wright and J. S. Bandar, Organic Superbases in Recent Synthetic Methodology Research, *Chem.-Eur. J.*, 2021, **13**, 4216–4229.
- 22 P. B. Kisanga, J. G. Verkade and R. Schwesinger, pK<sub>a</sub> Measurements of P(RNCH<sub>2</sub>CH<sub>3</sub>)<sub>3</sub>N, *J. Org. Chem.*, 2000, **65**, 5431–5432.
- 23 J. Gopalakrishnan, Aminophosphines: their chemistry and role as ligands and synthons, *Appl. Organomet. Chem.*, 2009, **23**, 291–318.
- 24 T. Brown, C. E. Pritchard, G. Turner and S. A. Salisbury, A new base-stable linker for solid-phase oligonucleotide synthesis, *J. Chem. Soc. Chem. Commun.*, 1989, **14**, 891–893.
- 25 paper in preparation.

