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An *o*-phthalimide-based multistimuli-responsive aggregation-induced emission (AIE) system†

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Aggregation-induced emission luminogens (AIEgens) have been widely used in the development of functional materials due to their excellent luminescence properties in the solid state. The development of new AIEgens is of great significance. In this work, a novel *o*-phthalimide-based AIEgen of 2,3-diphenylquinoxaline-6,7-dicarboimide (**2**) is reported, which exhibited multifunctional sensing applications. Interestingly, compound **2** exhibited ultrafast photo-response properties upon UV light irradiation in both the dissolved state and solid state, which was attributed to a unique photo-induced rearrangement reaction. This property of **2** makes it suitable for UV sensing and photo-patterning materials with a significant fluorescence change. Meanwhile, based on the highly specific interaction between Hg(II) and the *o*-phthalimide moiety of **2**, a complex of **2**-Hg-**2** formed in the presence of 0.5 equiv. Hg(II), resulting in a strong fluorescence change of **2**. The active hydrogen in the *o*-phthalimide moiety also makes the fluorescence of **2** sensitive to the pH of aqueous solutions. Due to these characteristics, **2** was successfully used in the detection of UV light, Hg(II) and the pH of aqueous solutions, in which **2** exhibited satisfactory detection performance.

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1 Introduction

Luminogens with aggregation-induced emission (AIE) properties have attracted considerable attention because of their promising applications in the fields of chemosensors, bioimaging, organic light-emitting diodes (OLEDs), smart materials, *etc.*¹ Unlike the traditional aggregation-caused quenching (ACQ) luminogens, whose luminescence in dispersed states is much stronger than that in aggregated states, AIE luminogens (AIEgens) exhibit intense luminescence in aggregated states such as in concentrated solutions and solid states.² This characteristic makes AIEgens suitable for luminescent chemosensors because most of the detections need to be performed in aqueous solutions where organic sensors usually have poor solubility and exist in an aggregated state.³

To date, a series of AIEgen-based luminescent chemosensors have been reported with excellent performances for the

detection of different analytes such as reactive oxygen species (ROS), metal ions, toxic substances, amino acids, pH, explosives and so on, which exhibit excellent performances.⁴ However, most of the reported chemosensors exhibit only singular sensing capability, and AIEgens with multifunctional sensing applications are still rare. Meanwhile, it is noticed that most of these reported chemosensors are designed based on a limited number of classic AIEgens such as tetraphenylethene (TPE), hexaphenylsilole (HPS), tetraphenylpyrazine (TPP), salicylaldehyde azine (SAA), and (2-hydroxyphenyl)benzothiazole (HBT), *etc.*⁵ The development of new AIEgens for luminescent chemosensors is still of great interest and importance.

In this work, an *o*-phthalimide moiety was introduced for the preparation of a new AIE system and a novel AIEgen of 2,3-diphenylquinoxaline-6,7-dicarboimide (**2**) was facilely synthesized *via* two-step reactions involving commercially available materials (Scheme 1). **2** exhibits a typical AIE characteristic due to the restriction of intramolecular rotation (RIR) mechanism. The *o*-phthalimide moiety of **2** endows it with a multifunctional sensing performance. The electron-withdrawing ability properties

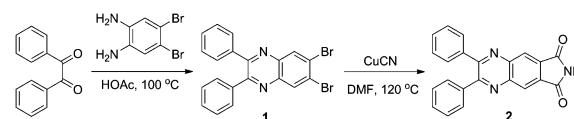
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† Electronic supplementary information (ESI) available: Selected spectra and data referred to in the paper; detailed crystal data and structure refinement for **2**. CCDC 1859465. For ESI and crystallographic data in CIF or other electronic format see DOI: 10.1039/c8qm00454d

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Scheme 1 Synthesis of **1** and **2**.

of *o*-phthalimide activated the phenyl group in **2**. Upon UV light irradiation, **2** turned into **3** *via* a unique photo-induced rearrangement reaction within 5 s, along with a remarkable fluorescence change from blue to green, which makes **2** suitable for monitoring ultraviolet radiation. Meanwhile, based on the highly specific interaction of Hg(II) with *o*-phthalimide, **2** was successfully used in the detection of Hg(II) in aqueous solutions with good selectivity and sensitivity. The active hydrogen in the *o*-phthalimide moiety also makes the fluorescence of **2** sensitive to the pH of the aqueous solution. Under alkaline conditions, the deprotonation endows **2** with stronger and blue-shifted luminescence compared to that under acidic conditions, which makes it suitable for detecting the pH of an aqueous solution.

2 Experimental section

2.1 Reagents

Unless otherwise stated, all materials used in this work are commercially available without further purification. 4,5-Dibromo-1,2-benzenediamine was purchased from AIEgen Biotech Co., Limited, Hong Kong. Benzil, copper(I) cyanide, and poly(ethyleneglycol) (M.W. 6000) were purchased from J&K Chemical Co. Beijing, China. All the other materials such as organic solvents, metal salts, acids and alkalis were purchased from Sinopharm Chemicals Beijing Co. Beijing, China. Analytical grade organic solvents and doubly distilled deionized water were used throughout the experiments. The solutions of all metal ions were prepared from their perchlorate salts or nitrate salts of analytical grade. Fe(II) solution was prepared from $(\text{NH}_4)_2\text{Fe}(\text{SO}_4)_2$. 10 mmol L⁻¹ phosphate buffer solutions (PBS) with different pH values were prepared using different ratios of phosphoric acid, sodium dihydrogen phosphate and sodium hydroxide. A 10 mmol L⁻¹ Tris solution of pH 7.0 was prepared using tris(hydroxymethyl)aminomethane and moderate perchloric acid. The pH values of the buffer solutions were detected using a pH meter at room temperature.

2.2 Apparatus

Absorption spectra were obtained on a JASCO V-750 UV-Vis spectrometer. Fluorescence spectra were obtained on a JASCO FP-8300 spectrometer. All pH values are detected using a METTLER-TOLEDO FE20/EL20 pH meter. The nuclear magnetic resonance (NMR) spectra were recorded on a Bruker 400 Avance NMR spectrometer operated at 400 MHz. ESI-MS spectra were obtained on Agilent Technologies 6420 triple quadrupole LC/MS apparatus without using the LC part. Dynamic Light Scattering (DLS) experiments were performed using a NanoPlus-3 dynamic light scattering particle size/zeta potential analyzer. Single crystal X-ray diffraction analysis was performed on a Rigaku Saturn 724 CCD diffractometer with Mo K α radiation ($\lambda = 0.71073$ Å) at room temperature. Light with different wavelengths was produced by a CEL-HXF300/CEL-HXUV300 xenon light source with different optical filters. High performance liquid chromatography (HPLC) analysis was performed using JASCO LC-4000 HPLC apparatus with a fluorescence

detector and an absorbance detector. The power density of light was detected using a CEL-NP200 light power meter. The photos and videos were taken using a Nikon D5500 camera.

2.3 Syntheses

The synthetic routes of the new compounds are shown in Scheme 1. Detailed synthetic procedures are shown below and the characterization data of the new compounds are shown in the ESI.†

2,3-Diphenyl-6,7-dibromoquinoxaline (1). 1,2-Diphenyl-1,2-ethanedione (5 mmol, 1.05 g) and benzil (5 mmol, 1.30 g) are mixed in 10 mL acetic acid. The mixture was stirred and heated to 110 °C for 3 h. After cooling to room temperature, a white precipitate was formed. The precipitate was filtrated and washed using 10 mL acetic acid three times. After being dried under reduced pressure, compound **1** was obtained as a white powder (1.85 g, yield 84%). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 8.49 (s, 1H), 7.50 (d, $J = 4.0$ Hz, 2H), 7.39 (t, $J = 4.0$ Hz, 1H), 7.34 (t, $J = 4.0$ Hz, 2H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 154.56, 140.42, 138.40, 133.23, 129.82, 129.33, 128.37, 126.43. ESI-MS spectrometry: *m/z* calculated for [M + H]⁺: 438.94; found: 438.96.

2,3-Diphenylquinoxaline-6,7-dicarboimide (2). Compound **1** (1 mmol, 0.44 g) and copper(I) cyanide (5 mmol, 0.45 g) were added to 10 mL DMF. The mixture was stirred at 170 °C under nitrogen. After 20 h, the mixture was cooled to room temperature and 20 mL ammonia was added. The resulting precipitate was filtered and purified by silica gel column chromatography (dichloromethane/methanol, 50:1). Compound **2** was finally gained as a light-yellow powder with a yield of 4.3%. ¹H NMR (400 MHz, DMSO-*d*₆) δ (ppm): 11.87 (s, 1H), 8.47 (s, 2H), 7.55–7.53 (m, 4H), 7.49–7.38 (m, 6H). ¹³C NMR (100 MHz, DMSO-*d*₆) δ (ppm): 168.31, 155.47, 143.34, 138.38, 133.03, 130.27, 129.91, 128.64, 124.90. ESI-MS spectrometry: *m/z* calculated for [M + H]⁺: 352.11; found: 352.11.

3 Results and discussion

3.1 AIE characteristics of 2

To understand the AIE characteristics of **2**, the fluorescence emission of **2** in H₂O/THF mixed solution with different water fractions (*f*_w, from 0% to 99%, v/v) was investigated. As shown in Fig. 1A, along with the increase of *f*_w, the fluorescence intensity of **2** enhanced gradually and reached the maximum when *f*_w was 99% (quantum yields enhanced from 0.35% to 7.46%). Normally, water is a poor solvent for organic molecules, which suggested that the fluorescence of **2** in aqueous solution might originate from its aggregated state, *i.e.*, an AIE emission. The aggregation of **2** in aqueous solution was supported by absorption spectra and dynamic light scattering (DLS) measurements. As shown in Fig. 1B, in the absorption spectra of **2** in aqueous solution, a level-off tail in the visible region could be clearly observed, which is believed to be due to the light scattering of aggregate suspensions.⁶ DLS measurements

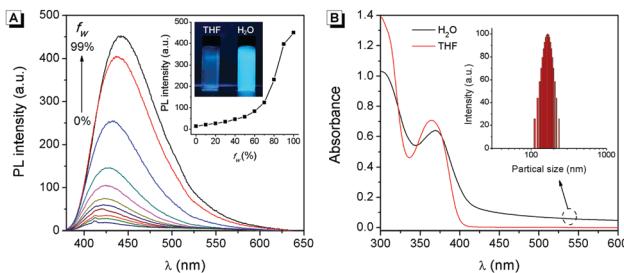


Fig. 1 (A) Fluorescence emission spectra of $50 \mu\text{mol L}^{-1}$ **2** in water/THF mixtures with different f_w . Inset: Fluorescence intensity of **2** at 445 nm as a function of f_w and fluorescence photograph of **2** in THF and H_2O (99% water/THF v/v solution). (B) Absorption spectra of $50 \mu\text{mol L}^{-1}$ **2** in THF and H_2O . Inset: DLS results of **2** in H_2O .

indicated that the particle size was around 100 nanometers (Fig. 1B inset).

The origination of the AIE characteristics of **2** was further investigated. First, a single crystal of **2** was grown from methanol/dichloromethane mixtures *via* slow solvent evaporation. The structure of the crystal was characterized by X-ray crystallography. As shown in Fig. 2A, the steric hindrances between the two phenyl groups endow **2** with a typical propeller-like structure. The dihedral angles between the phenyl groups and the quinoxaline center were 38.6° and 42.6° , respectively. This structural feature indicated that the AIE characteristics of **2** might originate from a RIR process: in the dissolved state, the phenyl groups undergo dynamic intramolecular rotations, resulting in an excited state non-radiative transition and fluorescence quenching. In contrast, the rotations of the phenyl groups are limited effectively in the aggregated state. As a result, the radiative transition became the primary pathway for the excited state electrons back to the ground state, along with intense fluorescence emission.⁷ Meanwhile, the minimum distance between the adjacent conjugate moieties of **2** was 3.95 Å (Fig. 2B), suggesting that the stacking interaction in the crystal can be ignored.⁸ The RIR mechanism for the AIE characteristics of **2** was supported by its fluorescence enhancement in solvents with high viscosity. As shown in Fig. S1 (ESI[†]), the fluorescence intensity of **2** in glycerin is 4-fold stronger than

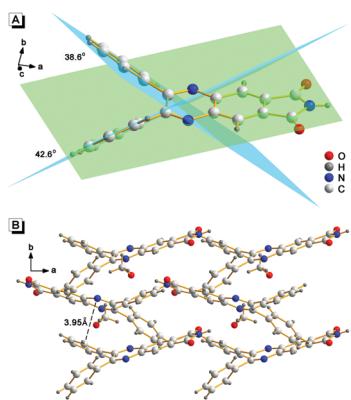


Fig. 2 Crystal structure of **2**. (A) The dihedral angles between the conjugate rigid planes. (B) The packing of **2** in the crystal and the minimum distance between the adjacent conjugate planes.

that in EtOH, which was attributed to the high viscosity solvent that restricted intramolecular rotations.

3.2 Photo-response performance of **2**

As a typical AIEgen, **2** exhibits weak fluorescence in the dissolved state. Interestingly, upon UV light irradiation, a strong green fluorescence appeared within 5 s (Fig. 3A and Video S1, ESI[†]). To understand this photo-response process, the absorption spectra of **2** upon UV light irradiation were recorded. As shown in Fig. 3B, three isobestic points could clearly be observed, suggesting the formation of new compounds after UV light irradiation.

More evidence for the formation of new compounds was obtained from HPLC analysis. As shown in Fig. 3C, in a mobile phase of MeOH, compound **2** exhibits a peak at a retention time of 3.6 min. After UV light irradiation, the peak of compound **2** decreased gradually, while a new peak with a retention time of 3.2 min appeared. These results confirmed that new compounds were formed after light irradiation. To understand the photo-response mechanism of **2**, ESI-mass spectra of the system before and after light irradiation were recorded. Interestingly, both of them exhibited a peak with a charge-to-mass ratio of 352.11 ($[2 + \text{H}]^+$, m/z calcd 352.11), which suggested that there might be a photo-induced rearrangement reaction for compound **2**. According to the reports, photoclick cycloaddition reactions could happen in some AIEgens which have adjacent aromatic rings combined with electron-withdrawing groups.⁹ As shown in Fig. 3D, compound **2** has these structural features. Thus, a possible photo-response mechanism of **2** was proposed based on the above results: upon UV light irradiation, **3** was produced through a photoclick cycloaddition reaction, and protons in the adjacent phenyl moieties transferred to the quinoxaline moiety. **3** has a large conjugate rigid plane structure and exhibits strong green fluorescence in the dissolved state.

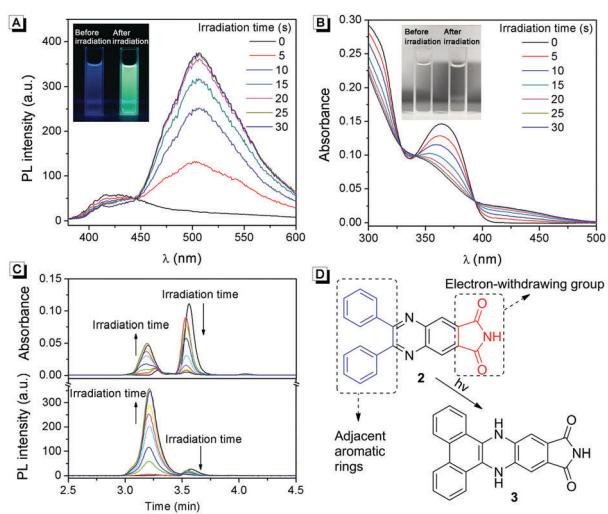


Fig. 3 (A) Fluorescence spectra and (B) absorption spectra of $10 \mu\text{mol L}^{-1}$ **2** in THF before and after UV light irradiation. (C) HPLC analysis of the photoconversion of $10 \mu\text{mol L}^{-1}$ **2** to **3** under UV light irradiation for different times. (D) Proposed mechanism for the photo-response performance of **2** upon UV light irradiation.

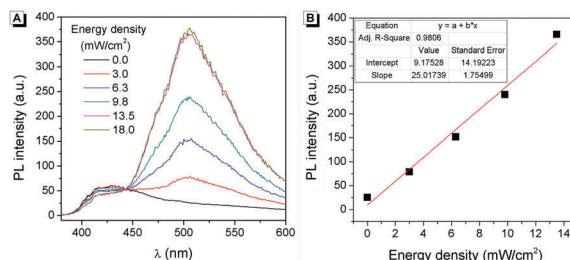


Fig. 4 (A) Fluorescence spectra of 10 $\mu\text{mol L}^{-1}$ of **2** in THF irradiated by 365 nm light with different irradiation intensities for 5 s. (B) Fluorescence intensity at 506 nm as a function of irradiation intensity.

The effect of the light irradiation wavelength on the photo-response reaction was further investigated. As shown in Table S1 (ESI[†]), light with different wavelengths was used to irradiate the sample, and only UV light (<400 nm) can promote the photo-response reaction. This wavelength range was fitted well with the absorption spectrum of **2** because there was no absorption beyond 400 nm (Fig. 1B). Then 365 nm UV light was used to understand the influence of light intensity on the photo-response reaction. As shown in Fig. 4A, after being irradiated by 365 nm UV light with different light intensities for 5 s, different fluorescence intensities can be observed. Stronger light led to more intense fluorescence and a good linear relationship was achieved (Fig. 4B). When the light intensity was higher than 13.5 mW cm⁻², the fluorescence intensity reached saturation after 5 s of irradiation. These results suggested that compound **2** can be used as a sensitive UV strength sensor.

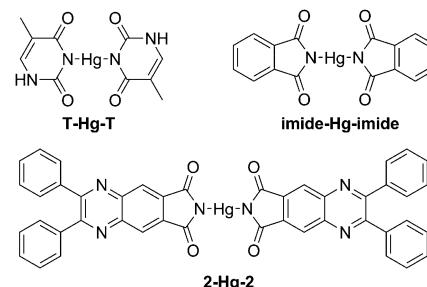
Furthermore, the photo-response property of **2** in the solid state was investigated. The fluorescence and absorption spectra of **2** in polyethylene oxide (PEO) before and after light irradiation are shown in Fig. S2 (ESI[†]). As can be seen, a ratiometric absorption change and a turn-on fluorescence change can be observed. These features indicate that **2** is capable of serving as a material for photo-printing. As shown in Fig. 5 and Video S2 (ESI[†]), letters are successfully recorded on **2** in polyethylene oxide after UV light irradiation. Under day light, the letters were very faint and hard to recognize. In contrast, the letters were clearly visualized under UV light. This tremendous contrast endows **2** with great potential for application in anti-counterfeiting materials.

3.3 Hg(II) sensing ability of **2**

Hg(II) is one of the most toxic metal ions for human health, and is widespread in environmental samples.¹⁰ The detection of Hg(II) has long been a great concern. According to the reports,



Fig. 5 Photographic images of **2** in PEO. The letters were generated upon irradiation at 365 nm. The scale bar is 2 mm.



Scheme 2 The structures of the **T-Hg-T** complex, the **imide-Hg-imide** complex and the **2-Hg-2** complex.

Hg(II) can efficiently bind with two T bases (thymine) to form a stable neutral **T-Hg-T** complex (Scheme 2). Guided by this theory, many fluorescence probes containing thymine moieties have been developed for the detection of Hg(II).¹¹ Besides thymine, other organic molecules with *N*-unsubstituted cyclic imides can also highly selectively bind to Hg(II).¹² Thus, as a luminogen with an *N*-unsubstituted cyclic imide moiety, the fluorescence response of **2** to Hg(II) was investigated.

As shown in Fig. 6A, the fluorescence intensity of **2** decreased gradually upon the addition of Hg(II) and reached the minimum when more than 0.5 equiv. of Hg(II) was added. When excess cysteine (Cys, a classic chelator for Hg(II)¹³) was added to the solution, the fluorescence intensity restored immediately (Fig. S3, ESI[†]). As shown in Fig. 6B, **2** exhibited a linear range of 0.0–2.0 $\mu\text{mol L}^{-1}$ Hg(II) with a correlation coefficient of $R_2 = 0.990$. Interestingly, there was nearly no fluorescence emission change upon the addition of other common metal ions (Na(I), K(I), Mg(II), Ca(II), Ba(II), Al(III), Cr(III), Mn(II), Fe(II), Fe(III), Co(II), Ni(II), Cu(II), Ag(I), Zn(II), Cd(II), and Pb(II)), which indicated that **2** has

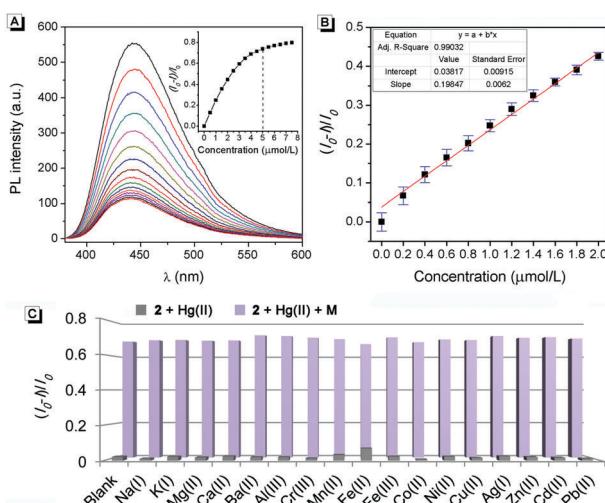


Fig. 6 (A) Fluorescence spectra of **2** upon the addition of Hg(II). Inset: The fluorescence intensity at 445 nm as a function of the Hg(II) concentration. (B) Calibration curve for the emission of **2** with increasing concentrations of Hg(II). (C) Fluorescence intensity at 445 nm in the presence of 0.5 equiv. of different metal ions. Conditions: the concentration of **2** was 10 $\mu\text{mol L}^{-1}$, 99% water/DMSO (v/v) at pH 7.0 controlled by 10 mmol L⁻¹ Tris solution.

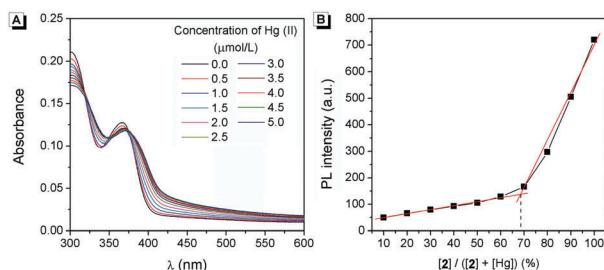


Fig. 7 (A) Absorption spectra of $10 \mu\text{mol L}^{-1}$ **2** upon the addition of Hg(II). (B) Job's plot data for evaluating the stoichiometry of the **2**-Hg-**2** complex. The total concentration of **2** and Hg(II) was kept at $20 \mu\text{mol L}^{-1}$. 99% water/DMSO (v/v) at pH 7.0 controlled by 10 mmol L⁻¹ Tris solution was used. Excitation and emission were performed at 368 nm and 445 nm, respectively.

excellent selectivity and provided more anti-interference capability to Hg(II) than other metal ions (Fig. 6C). These results suggested that **2** could be used as a selective chemosensor for the detection of Hg(II).

The mechanism of the response for **2** to Hg(II) was further investigated (Scheme 2). Firstly, three isobestic points could clearly be observed in the absorption of **2** upon the addition of Hg(II), indicating the formation of a complex between **2** and Hg(II) (Fig. 7A). Secondly, Job's plot analysis was carried out to confirm the stoichiometry of **2**-Hg-**2**. The result indicated that the binding ratio between **2** and Hg(II) was 2:1 (Fig. 7B). Moreover, the binding site of **2** to Hg(II) was verified by ¹H-NMR spectra (Fig. 8). Before the addition of Hg(II), the proton of the *o*-phthalimide moiety (proton a) could well be assigned. After the addition of Hg(II) into the mixture, the peak of "proton a" reduced significantly while other peaks showed almost no change, suggesting that **2** bound Hg(II) with its imide group.

3.4 pH sensing ability of **2**

According to the reports, hydrogen in the *o*-phthalimide moiety was very active, which could be removed under alkaline conditions.¹⁴ Therefore, the fluorescence of **2** in aqueous

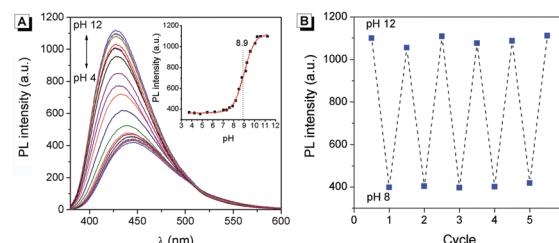


Fig. 9 (A) Fluorescence spectra of **2** in aqueous solution with different pH values. Inset: The fluorescence intensity at 426 nm as a function of pH. (B) Reversible fluorescence changes of **2** at 426 nm by adjusting its solution to pH 8 and pH 12 repeatedly.

solutions with different pHs was investigated. As shown in Fig. 9A, along with the increase of pH, the fluorescence intensity of **2** enhanced gradually and the emission wavelength blue-shifted from 445 nm to 426 nm. The titration curve as shown in the Fig. 9A inset indicated a jump ranging from about pH 8 to pH 10, and an acid dissociation constant (pK_a) of 8.9 was obtained by nonlinear fitting ($R_2 = 0.997$). Meanwhile, the fatigue resistance of **2** to the pH change was investigated. As can be seen in Fig. 9B, the fluorescence intensity of **2** stayed almost constant without apparent degradation when the pH of the solution was switched back and forth between 8.0 and 12.0 5 times. These results suggested that **2** could be used as a reversible fluorescent chemosensor for pH sensing.

To understand the pH response mechanism of **2**, the ¹H-NMR spectrum of **2** in the absence and presence of alkali was recorded. As shown in Fig. 10, the peak of the proton in the *o*-phthalimide moiety (proton a) disappeared after the addition of triethylamine (TEA). When excess trifluoroacetic acid (TFA) was further added to the solution, the peak of "proton a" recovered. These results provide powerful evidence for the deprotonation and protonation of **2** at different pH values. Furthermore, the absorption spectra and particle size distribution of **2** at different pHs were investigated. As shown in Fig. S4 (ESI†), the shapes of the absorption spectra of **2** at different pHs were very similar, while level-off tails in the visible region disappeared when the pH was higher than 9, indicating that

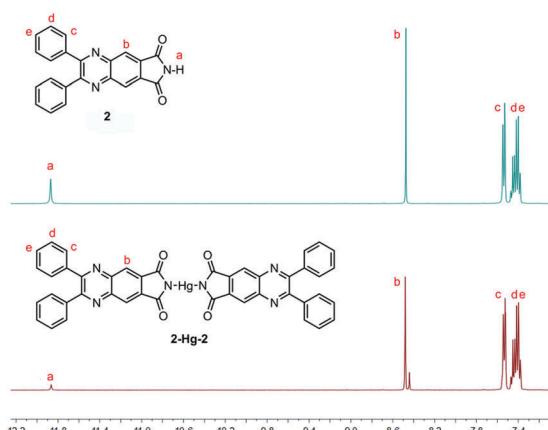


Fig. 8 ¹H-NMR spectra of **2** in DMSO-*d*₆ before (up) and after (down) the addition of 0.5 equiv. Hg(II).

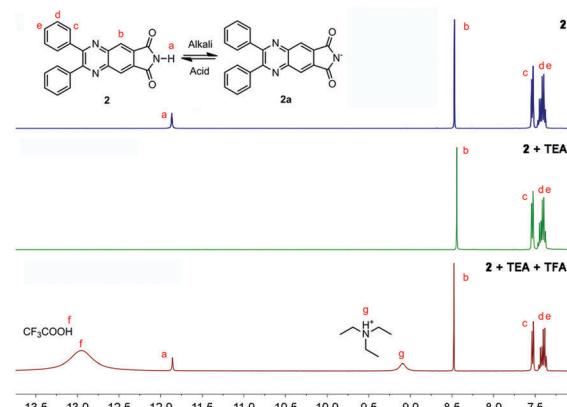


Fig. 10 ¹H-NMR spectra of **2** before and after the addition of excess TEA and TFA.

the aggregation of **2** was dissolved. Meanwhile, in the DLS test no particle could be observed for **2** at pH 12, which also confirmed the good solubility of **2a**. According to the above results, the proposed response mechanism of **2** to pH is obtained: when the pH was lower than 8, **2** existed in the form of a neutral molecule, exhibiting AIE fluorescence around 445 nm; when the pH was higher than 10, **2** underwent deprotonation, producing **2a** with stronger fluorescence around 426 nm. The negative charge of **2a** endows it with good solubility in aqueous solution.

4 Conclusions

In conclusion, an *o*-phthalimide-based AIE molecule of **2** was developed, which can be used in multifunctional sensing applications for UV light, Hg(II) and pH value. Mechanism studies showed that the AIE property of **2** originated from a typical RIR process. Upon UV light irradiation, a photoclick cycloaddition reaction occurred in **2**, producing an isomer with a large conjugate rigid plane structure and intense green fluorescence. This feature makes **2** suitable for the detection of UV light. The *o*-phthalimide moiety plays a crucial role in the detection of Hg(II) and the pH value. By forming a **2**-Hg-**2** complex with the *o*-phthalimide moiety, **2** exhibited a highly selective fluorescence response to Hg(II) with a linear range of 0.0–2.0 $\mu\text{mol L}^{-1}$. A deprotonation process takes place in the *o*-phthalimide moiety of **2** at a high pH value, resulting in an increase of fluorescence. Besides these sensing applications, **2** was also successfully used in photo-printing and anti-counterfeiting materials.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

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