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Effect of wettability on microplastic aerosolization *via* film and jet drops ejected from bursting bubbles

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Bubble bursting during oceanic breaking waves releases tiny droplets that can transport species—including sea salt, microorganisms, and microplastics—across the air–water interface. While many studies have investigated particle–bubble interactions and the role of particle wettability in particle attachment to rising bubbles, a limited number have extended this to particle aerosolization onto the ejected droplets. This study aims to experimentally investigate how wettability of microplastic (MP) particles affects their aerosolization *via* the two major droplet ejection pathways from a bursting bubble: film and jet drops. Controlled experiments are conducted with 1 μm diameter surface-modified polystyrene MPs of two contrasting wettabilities (*i.e.*, hydrophilic *vs.* hydrophobic) in ultrapure water. Film and jet drop pathways are isolated by generating two distinct bubble populations known to primarily produce each droplet type. The results show that the aerosolization factor – defined here as the air-to-water MP concentration ratio – of hydrophobic MPs is approximately one order of magnitude higher than that of hydrophilic MPs for jet drops. In contrast, no significant difference was observed for the film drop aerosolization factor, which can be attributed to a potentially complex effect that MP particles can have on bubble film stability, bursting, and enrichment dynamics. These findings highlight that MP surface properties can significantly influence their ejection into the atmosphere at the ocean surface. Given the potential for inhalation and long-range transport, this mechanism may contribute to the global dispersion of airborne MP pollutants. The results underscore the need to consider aerosolization pathways in the environmental fate and risk assessment of plastic pollution.

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Environmental significance

Microplastics (MPs) can be transported from oceans to the atmosphere *via* film and jet droplets released during bursting bubbles, but the role of their surface properties in this process remains poorly understood. Our study demonstrates that MP wettability influences their concentration in airborne droplets—but this effect is pathway-specific. Hydrophobic MPs transfer more efficiently *via* jet drops, while no clear effect was observed for film drops, suggesting more complex mechanisms at play. These findings offer insights into how MPs may contribute to atmospheric pollution, cloud formation, and long-range contaminant transport. By emphasizing the role of MP surface properties, this work underscores the need for further laboratory studies to refine MP enrichment models and improve understanding of plastic pollution transport and fate.

Introduction

Bubble bursting during oceanic breaking waves is a crucial component of the ocean–atmosphere interaction. These bursting bubbles release sea spray aerosols (SSAs) that can transport species, including sea salts, bacteria, viruses, dissolved gases, and organic matter across the air–water interface.[1–4] Microplastics (MPs) have been similarly observed to transfer across the air–water interface *via* bubble bursting.[5–8] MPs, an emerging critical environmental pollutant, have been observed in every environmental compartment on Earth,[9] including marine environments,[10] soil,[11] freshwater systems,[12,13] and the atmosphere.[5,14] Recent studies have considered oceans to be one of the major sources of atmospheric MPs.[15] Atmospheric MPs are emerging air pollutants, with potential health risks to living beings,[16–19] and can be a vector for a wide range of toxic pollutants.[20] Therefore, it is critical to constrain the oceanic source of atmospheric MPs. Physical understanding of the processes involved in (and affecting) the ocean-to-atmosphere transfer of MPs is a crucial step to constrain the ocean source.[21]

During breaking waves, large volumes of air enter the water and rise up in the form of bubbles that burst at the surface releasing SSAs *via* the two major droplet ejection pathways: film and jet drops.[22] Shattering of the surface bubble cap releases hundreds of film drops, while the collapse of the resulting cavity emits a series of jet drops vertically upwards. Due to differences in ejection mechanisms, film drops are generally in the submicron size range (<1 μm), while jet drops are generally in the

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super-micron range.[2,3,23] Bubbles rising up in MP-contaminated waters can scavenge MPs, enriching them at the bubble interface, which are eventually aerosolized *via* film and jet droplets.[24] It is well-known that contaminant species can be present in substantially higher concentration in the resulting droplets compared to that in the bulk water, thus amplifying the influence of these contaminants.[4,25,26] Additionally, water and particle properties can potentially affect the process of MP scavenging and the resulting enrichment onto droplets.

MPs are generally considered to be hydrophobic, but environmental factors such as heat, ultraviolet (UV) radiation,[27] mechanical forces, biofouling,[28] absorption of organics and chemicals such as PFAS[29] are known to significantly affect surface chemistry and/or wettability of MPs.[9,30] Photo-oxidation is the major route for polymer degradation in the environment,[31,32] and Al Harraq *et al.*[32] observed that the UVA-weathered polyethylene (PE) exhibited increased wettability and dispersibility in the water. Such dynamic changes in wettability present a challenge in predicting the fate of microplastics in the environment.[27] Additionally, once airborne, MPs bearing hydrophilic groups can potentially act as condensation nuclei of cloud ice and water, highlighting the role of MP wettability on cloud formation processes.[33–35] These studies underscore the importance of addressing the role that particle wettability plays in the process of water-to-air transfer of MPs.

Many laboratory studies have investigated the aerosolization of MP particles *via* single bubbles as well as some developed theoretical models for film and/or jet drop enrichment.[8,25,36,37] Other laboratory studies have used a number of bubble generation mechanisms, typically to "model" oceanic wave breaking processes.[6,7,38–41] These studies have generally focused on the role of (i) MP properties (polymer type, size, concentration, density, and surface composition), (ii) bubble properties (size), and (iii) water properties/composition (salinity, surfactants, and organic matter) in the transfer of MPs at the air–water interface.

Despite that, limited laboratory studies have attempted to address the uncertainties surrounding the effect of the surface properties of MPs on their selective enrichment onto film and jet drops.[24,42] It is well-known that particle hydrophobicity favors selective attachment to rising bubbles.[43] Many studies have investigated particle–bubble interactions[44–47] and the role of particle wettability in the attachment,[43,48] but only a few studies have extended this understanding to particle aerosolization *via* film and jet droplets ejected by bubbles.[22,42] Notable studies that have developed particle enrichment models in film[36] or jet drops[8,25,37] based on experiments conducted at the level of a single pure-or tap-water bubble have not considered the role of particle wettability. Furthermore, these theoretical enrichment models have been applied to estimate MP emission from the oceans *via* sea spray.[8,21] Early studies in biological aerosols[4,49–51] observed that the hydrophobicity of *Serratia marcescens* bacterial cells enhanced their aerosolization onto the topmost jet droplets. Interestingly, Masry *et al.*[6] observed that the transfer of PE particles increased substantially after they were UV-aged, which is known to reduce hydrophobicity. They attributed this to the increased aggregation of the pristine, more hydrophobic PE at the water's surface, which lowered their transfer rate. This highlights how complex interfacial behaviors, such as particle aggregation, can govern the net aerosolization rate measured in such laboratory systems,[6,7] and underscores the need for controlled experiments that can systematically isolate the role of wettability from other confounding variables.

Recently, Dubitsky *et al.*[24] used physics-based theoretical modeling to bound the effects of particle wettability in bubble-mediated MP aerosolization by considering the two extremes of particle hydrophobicity: non-scavenged hydrophilic and fully-scavenged hydrophobic particles – represented by bubble–particle attachment efficiency of 0 or 1, respectively. Their theoretical modeling results show that perfectly attaching hydrophobic particles can be selectively enriched by several orders of magnitude higher in jet drops compared to film drops, highlighting the key role particle wettability could play in sea–air transport of MPs. Furthermore, they highlight the need for laboratory investigations into the effect of MP properties in order to better inform their theoretical models.

Thus, the primary goal of this study is to conduct a controlled experimental investigation into the effect of microplastic wettability on their aerosolization *via* film and jet drops emitted *via* bubble bursting. The approach involves using two types of MPs of atmospherically relevant size that share the same base polymer but exhibit contrasting wettabilities in a multi-bubble bursting setup.

Materials and methods

Film and jet drop production in the SAPT

Based on the experimental setup put forth by Wang *et al.*,[52] a spray aerosol pathway tank (SAPT) was built (Fig. 1a) that can generate bubble populations of two distinct size distributions known to produce film and jet drops (Fig. 1b).[2,3,22,52,53] The SAPT comprises three major components: (i) a 30 cm × 30 cm × 60 cm polycarbonate tank; (ii) a glass frit with a pore size of 250–500 μm (ROBU® Glasfilter-Geraete GmbH, Hattert, Germany), and (iii) a 10 L pressure vessel connected to the tank with a 0.15 mm diameter orifice (Precision Metal Orifices, O'Keefe, Monroe, CT, USA).

Particle-free air is passed through the glass frit at a rate of 1.4 liters per minute (LPM) into the tank containing 30 L of water producing a plume of coarse bubbles (~500 to 1500 μm in radius, see Fig. 1b) that rise through a height of ~30 cm, which upon bursting, primarily produce film drops.[3,22,52] Separately, sub-100 μm radius fine bubbles (Fig. 1b) were generated by passing pressurized ultrapure water at 60 psi (~414 kPa) through the orifice into the tank. These fine bubble plumes rise ~40 cm to the surface and burst to primarily produce jet drops.[3,22,52] It is important to note that our use of the term 'jet drops' and 'film drops' in the context of SAPT refers to droplets ejected by these two distinct bubble populations, acknowledging the potential presence of a mixture – a point expanded in the next section.

Film and jet droplet size distribution

The size distribution of film and jet droplets generated in the SAPT headspace was measured using a scanning mobility

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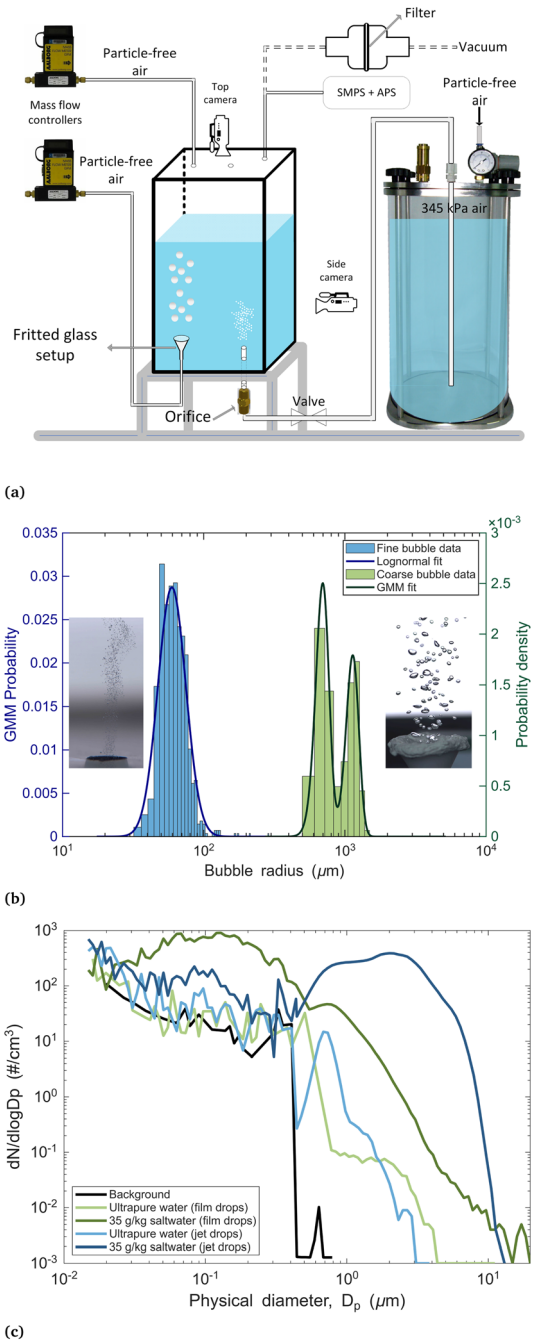

Fig. 1 (a) Schematic of the Spray Aerosol Pathway Tank (SAPT) that consists of a film drop setup producing coarse bubbles of radius between ~500 to 1500 μm, and a jet drop setup producing fine bubbles of radius < 100 μm. (b) Histogram and fitted distribution for fine (blue) and coarse (green) bubbles in the SAPT. The inset shows the photographed sub-surface fine (left inset) and coarse (right inset) bubble plume in the SAPT. (c) Steady-state aerosol size distribution (i) in the background (black), (ii) measured from coarse bubbles bursting in ultrapure (light green) and synthetic seawater (dark green) to produce predominantly film droplets, and (iii) measured from fine bubbles bursting in ultrapure (light blue) and synthetic seawater (dark blue) producing predominantly jet droplets as measured using SMPS and APS without a dryer. Synthetic seawater was produced by dissolving sea salt (Instant Ocean Sea Salt, Amazon.com, Inc., Seattle, WA, USA) in ultrapure water (~18 MΩ cm, Picopure) to achieve a mixing ratio of ~35 g kg^{-1}.

particle sizer (SMPS, Model 3936, TSI, Shoreview, Minnesota, USA) and an aerodynamic particle sizer (APS 3321, TSI, Shoreview, Minnesota, USA). The SMPS, consisting of an Electrostatic Classifier (Model 3080), a Differential Mobility Analyzer (Model 3081), and a Condensation Particle Counter (Model 3025a), measured particles with a mobility diameter (D_m) between 14 and 700 nm. The APS measured particles with an aerodynamic diameter (D_a) between 0.5 and 20 μm.

For sampling, particle-free air was continuously supplied to the SAPT headspace at 6.1 LPM to flush out aerosols. The concentration of the produced droplets in the headspace was allowed to reach a steady state (approximately 5 minutes for film drops and 15 minutes for jet drops). The droplets were then sampled through ~1 m of antistatic silicone rubber tubing (McMaster-Carr, Illinois, USA) to the instruments. The SMPS and APS operated in parallel at a scan rate of 5 minutes with sampling flow rates of 0.3 LPM and 1.0 LPM, respectively. All measurements were carried out without a dryer and the relative humidity (RH) was monitored using a HOBO UX100-011 data logger (Onset Computer Corporation, MA, USA) upstream of the aerosol sizing instruments. The RH of the film and jet droplet-laden air stabilized at around 85% before entering the aerosol sizing instruments (Fig. S1).

To obtain a single aerosol size distribution spanning the SMPS and APS measurement ranges, D_m and D_a size distributions have to be merged (May *et al.*, 2016; Stokes *et al.*, 2013) into a single physical diameter (D_p) size distribution. To convert D_m measured by the SMPS to D_p, the following relation was used under the assumption of spherical particle geometry:

$$D_m = D_p \quad (1)$$

D_a measured by the APS was converted to D_p using the following relation:

$$D_p = \frac{D_a}{\sqrt{\frac{\rho_{eff}}{\rho_0}}} \quad (2)$$

where ρ_0 is the unit density (1 g cm^{-3}) and ρ_{eff} is the effective density assigned to the particles sized by the APS. When stitching, particle bins in the overlapping size region of the SMPS and APS were removed due to undercounting of particle concentrations in this size range by both instruments.[54]

The primary experiments in this study were conducted using ultrapure water (~18 MΩ cm, Picopure). However, directly measuring sub-micron film drops (with radii of formation <1 μm) produced from ultrapure water is challenging, as they are prone to rapid evaporation and shrinkage as they adjust to the ambient RH within the SMPS. Particles of this size typically equilibrate with their surroundings in about 0.1 seconds or less.[3] To confirm that our SAPT apparatus effectively generates both film and jet drops, we performed a validation experiment using synthetic seawater, which was produced by dissolving sea salt (Instant Ocean Sea Salt, Amazon.com, Inc., Seattle, WA, USA) in ultrapure water to achieve a mixing ratio of ~35 g kg^{-1}. Unlike pure water, the evaporation of saltwater droplets leaves behind detectable salt particles.

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As shown in Fig. 1c, the seawater validation results confirm the presence of a sub-micron mode around 0.1 μm (dark green line), corresponding to film drops that can be attributed to the recently proposed flapping film mechanism for film drop production.[55] This mechanism applies to bubbles in the size range of approximately 70–1200 μm in saltwater.[23,53] A recent study by Mazzatenta *et al.*[53] – which used an intermediate-scale steady state bubble bursting setup to investigate the link between bubbles and emitted droplets – found that the theoretical size scaling and magnitude of the flapping film drops (per Jiang *et al.*[55]) matched well with their measured droplet size distribution (see their Fig. 7). Additionally, we observe a smaller mode around 1 μm that can be attributed to the well-established film centrifuge mechanism for film drop production,[56] applicable to bubbles in the size range of approximately 900–10 000 μm.[53]

Similarly, we observe a super-micron mode around 2 μm (dark blue line), which can be attributed to jet drops, as the measured droplet sizes are consistent with theoretical estimates of jet drop size (per Mazzatenta *et al.*,[53] based on Ganan-Calvo[57]) generated from our fine sub-100 μm bubbles. Additionally, we observe a smaller sub-micron mode around 0.1 μm that can be attributed to flapping film drops produced by the fraction of fine bubbles larger than about 70 μm (per Mazzatenta *et al.*,[53] based on Jiang *et al.*[55]), suggesting that our setup, as expected, does not exclusively produce film or jet drops.

Consistent with our observations in saltwater, we also observed a mode around 0.7 μm for ultrapure water (light blue line), which can be attributed to jet drops that likely shrank as they adjusted to the ambient RH within the APS. This mode cannot be attributed to film drops, as the bubble sizes responsible for producing ∼1 μm film droplets – more precisely, centrifuged film drops – are about 1–10 mm (Mazzatenta *et al.*,[53] based on Lhuissier and Villermaux[56]), which were not present in our fine bubble setup. On the other hand, a distinct sub-micron mode for film drops – more precisely, flapping film drops – was not resolved, likely due to evaporation. This is consistent with previous reports of limited or no film drop production *via* bubble bursting in pure distilled water compared to saltwater.[3,58] Similarly, we observed a small super-micron mode around 2 μm that can be attributed to shrunken centrifuged film drops generated *via* our coarse bubble setup.

Measurement of sub-surface bubble size distribution

Bubble plumes were photographed from the side of the SAPT using a Nikon D750 DSLR camera (shown in Fig. 1a) placed against the backdrop of an LED panel and a precision ruler for scale (Fig. S2). A Nikon AF-S NIKKOR 24–120 mm 1:4G ED lens was used to photograph coarse bubbles and an AF-S VR MICRO NIKKOR 105 mm 1:2.8G ED micro-lens was used for fine bubbles. The bubble radius was determined as the projected area radius, defined as the radius of a circle that has an area equal to the projected area of the bubbles. The projected areas were calculated manually from the bubble photographs using the wand tool (with an appropriate tolerance value) in the image processing software ImageJ.[59] Fig. 1b left (right) shows the histogram and the lognormal (Gaussian mixture model) fit for the fine (coarse) bubbles. Since some coalescence events were observed at the surface (Fig. S3), we note that the actual size of certain bubbles just before bursting might be slightly larger but would still tend more toward the film drop production mode.

Temporal evolution of surface bubbles

Bursting of surface bubbles was recorded from a top-down view against a dark background using a Nikon D750 DSLR camera equipped with an AF-S NIKKOR 24–120 mm 1:4G ED lens (shown in Fig. 1a), with bubbles illuminated from the sides using an LED light strip (Fig. S4). The recording also captured the display of the mass flow controller, which regulated airflow through the glass frit. This allowed us to determine the precise moment the bubble flow ceased, which was used as the reference time for the experiments. From this reference point, a four-second video segment was extracted for analysis.

To quantify the extent of surface bubbling, we estimated the percentage of the video frame area covered by bubbles. Each frame of the four-second video segment was processed using an ImageJ algorithm that applied a threshold to generate a binary image, separating the white bubbles from the dark background. The fraction of white pixels in each binary frame was used as a proxy for the area occupied by bubbles on the water surface. To ensure consistency across all video samples (15 each), each video was digitally cropped to a uniform region encompassing only the water's surface, correcting for minor variations in camera placement across recordings. This analysis provided a time-resolved estimate of surface bubble coverage, which was used as a proxy for bubble stability and compared across various experiments.

Microplastic aerosolization experiments

Fluorescent monodisperse 1 μm diameter polystyrene (PS) microspheres of two contrasting wettabilities – hydrophobic sulfate-modified PS (SuPS) and hydrophilic amine-modified PS (AmPS) microspheres (Thermo Fisher Scientific, Oregon, USA) – were suspended in ultrapure water (∼18 MΩ cm, Picopure). This medium was specifically chosen to: (i) eliminate the additional effects of salinity, (ii) establish experimental conditions comparable with previous single-bubble measurements in pure water,[8,25,36,60] and (iii) ensure consistency across all our experimental measurements, including contact angle, zeta potential, and aerosolization. The particle size of 1 μm was selected because it is atmospherically relevant and sufficiently small to be incorporated within both film and jet droplets, rather than excluded due to geometric constraints.

To establish the relative wettability of the two particle types, the apparent contact angle of these microspheres was measured by placing tiny droplets of ultrapure water on a ∼30 μm thick and dry flat surface made up of AmPS and SuPS particles. The flat surface was created by filtering a suspension of these MPs onto a 25 mm polycarbonate filter *via* a vacuum filter assembly (Sigma-Aldrich®, MO, USA). The droplet had a low contact angle of ∼58° on the AmPS layer but a high contact angle of ∼128° on the SuPS layer, as measured using a Drop Shape Analyzer

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(DSA25, Kruss, Germany) (Fig. S5). While this measurement provides an apparent value that incorporates surface roughness effects, it serves as a strong proxy for classifying the particles' overall hydrophilic or hydrophobic nature. Additionally, as a measure of surface charge, the zeta potential of SuPS and AmPS suspensions in ultrapure water was measured at a concentration of 10^{12} particles per liter (PPL) using a Zetasizer Ultra Red (Malvern Panalytical Ltd., UK) and was found to be −57.78 mV and −14.54 mV, respectively (Fig. S5). Together, these measurements confirm that the two MP types—identical in size and base polymer—differ primarily in surface functionalization. Both are negatively charged, eliminating surface charge sign as a confounding factor, and enabling us to isolate the role of wettability in modulating aerosolization behavior.

For MP aerosolization *via* film (jet) drops, three (four) experiments were conducted with ultrapure water at room temperature for each of the SuPS and AmPS suspensions, using a single representative number concentration of $\sim 10^6$ PPL,[61] as the general effect of particle concentration has already been established in our prior work.[62] Working solutions were prepared by extracting a portion of the manufacturer's stock solution using an Eppendorf pipette, diluting it in ultrapure water to achieve the target concentration, and then stirring and sonicating for 5 minutes to ensure dispersion of particles. Both SuPS and AmPS were assumed to be well-mixed in the bulk ultrapure water since their density (1.055 g cm^{-3}) was similar to the density of pure water at room temperature (1 g cm^{-3}), resulting in a high settling time on the order of months.

Each MP aerosolization experiment in the SAPT consisted of a 5 minute period of flushing the SAPT headspace with particle-free air, followed by a 5–15 minutes period of bubbling to reach steady-state aerosol concentration in the headspace. This was then followed by a 30-minute (film drop) or 60-minute (jet drop) air sampling period. Air samples – drawn from the tank headspace at a flow rate of 2 LPM – and 20 mL water samples from each experiment were passed through a Polycarbonate Track-Etch (PCTE) membrane filters (Zefon International, FL, USA) using a vacuum filter assembly (Sigma-Aldrich®, MO, USA) to collect MPs for offline analysis. To capture variations in MP number concentration in water, water samples were taken before and after each experiment. Before starting a new batch of MP experiments, the SAPT was thoroughly cleaned with soap and ultrapure water and disinfected with 70% isopropyl alcohol to eliminate residual contamination from previous experiments.

The air and water PCTE filters were scanned using a fluorescence microscope (EVOS® FL Auto Imaging System, Thermo Fisher Scientific Inc., MA, USA) (Fig. S6), and the scanned images were processed using ImageJ[59] to count the number of MPs in the sampled air and bulk water volumes. The aerosolization factor (AF) was defined as the ratio of particle number concentration (PPL) in the sampled volume of air ($C_a = N_a/V_a$) to that in the bulk water ($C_w = N_w/V_w$):[7]

$$\mathrm{AF} = \frac{C_a}{C_w} = \frac{N_a/V_a}{N_w/V_w} \quad (3)$$

Here, N_a represents the total number of aerosolized MPs captured on the PCTE filters during the sampling period, and V_a denotes the total volume of air drawn from the tank headspace through these filters. N_w represents the number of MPs present in the volume of water sampled ($V_w = 20$ mL), assuming a well-mixed condition.

We note that our AF is distinct from the enrichment factor (EF) which is commonly used in single-bubble studies to quantify particle enrichment in droplets.[24,25,36] EF is defined as the ratio of the particle number concentration in the generated film or jet droplets to that in the bulk water. We refrain from estimating EF here due to the significant uncertainties in determining the volume of pure water film or jet droplets produced in our more realistic, multi-bubble bursting setup. Instead, AF offers a more robust metric for comparing aerosolization under these conditions.

Results and discussion

Wettability-driven differences in MP aerosolization

Fig. 2 shows the empirical film and jet drop AFs of SuPS (purple) and AmPS (green), with circular markers representing the mean values and error bars representing the standard deviation. Our experimental results show that the jet drop AF of hydrophobic SuPS (mean AF_{jet}^{SuPS} of $\approx 1.8 \times 10^{-6}$) is approximately one order of magnitude higher than that of hydrophilic AmPS (mean AF_{jet}^{AmPS} of $\approx 2.2 \times 10^{-7}$). Interestingly and in contrast, no clear dependence on wettability was observed for aerosolization *via* film drops: mean AF values of SuPS (AF_{film}^{SuPS}) and AmPS (AF_{film}^{AmPS}) in film drops are $\approx 9.1 \times 10^{-7}$ and $\approx 1.6 \times 10^{-6}$, respectively. We acknowledge that the size of the MPs (1 μm) relative to the predominantly sub-micron film drops imposes

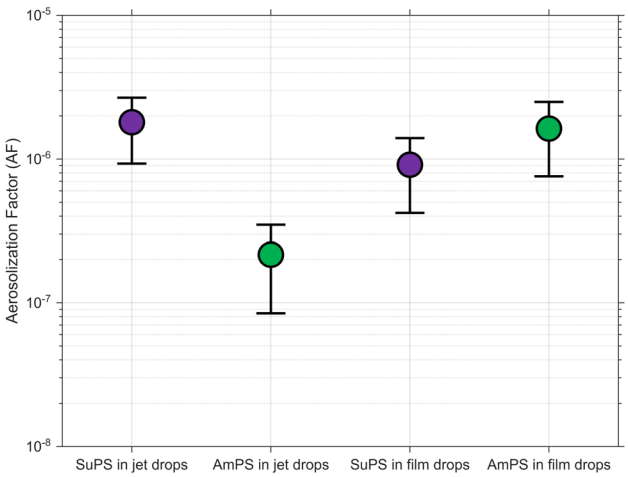

Fig. 2 Experimental aerosolization factors (AFs) of 1 μm hydrophobic (SuPS, purple) and hydrophilic (AmPS, green) microplastics in film and jet droplets generated *via* bursting ultrapure water bubbles in the Spray Aerosol Pathway Tank (SAPT). The aerosolization factor is defined as the ratio of particle number concentration in the sampled air to that in the bulk water. Circular markers represent mean values from replicate experiments ($n = 4$ for jet drops, $n = 3$ for film drops), and the error bars represent ±1 standard deviation. The y-axis is on a logarithmic scale.

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a geometric constraint that limits overall aerosolization. Consequently, the observed aerosolization is likely driven by the subset of larger, super-micron 'centrifuged' film drops.

The enhanced aerosolization of hydrophobic SuPS in jet drops aligns with the well-established role of particle hydrophobicity in favoring particle–bubble attachment, which subsequently influences particle aerosolization *via* droplets.[24,63] This finding is also consistent with historical biological aerosol studies, where the hydrophobicity of *Serratia marcescens* bacterial cells enhanced their transfer into the topmost jet droplets produced in distilled water.[4,49–51] On the other hand, Masry *et al.*[6] observed a seemingly opposite trend, where hydrophilic, UV-aged PE aerosolized significantly more than pristine, hydrophobic PE. Given that the bubbles used in their study (supposedly having radii of $\approx 340 \pm 50$ μm[6,64]) are of the size known to produce predominantly jet drops, this discrepancy can be explained by a key difference in the experimental design: they attributed their observation to the formation of aggregates of pristine hydrophobic PE at the water surface, which limited the water-to-air transfer rate – an observation consistent with our previous work.[7]

While surface roughness and charge are also known to affect particle–bubble interactions, neither of them explain our observations for the following reasons: (i) the surface morphology was visually observed to be similar between the spherical SuPS and AmPS per images taken using a JEOL IT500 scanning electron microscope (JEOL, Peabody, Massachusetts, USA), and (ii) since bubbles in pure water are negatively charged (~ -35 mV),[65,66] the stronger negative charge of SuPS (~ -58 mV) should create a greater electrostatic repulsion than for AmPS (~ -15 mV), thus hindering attachment. However, the opposite trend was observed. Therefore, aerosolization differences can be confidently attributed to wettability, indicating that hydrophobic attraction is likely the dominant mechanism.

The lack of a clear wettability effect for film drops, however, suggests a more complex mechanism. It can be assumed that hydrophobic SuPS particles are more readily scavenged by the bubbles, likely leading to a relatively higher concentration in the bubble cap film. Considering the one order of magnitude difference observed between AF_{jet}^{SuPS} and AF_{jet}^{AmPS}, it is logical to assume a correspondingly higher AF_{film}^{SuPS}. That we do not observe this implies that hydrophobicity does not enhance aerosolization *via* film drops in the same way, a point we explore further in the next section. This difference in AF can be attributed primarily to the difference in the number of SuPS or AmPS emitted *via* film drops within the sampling duration (N_a), since V_a and C_w are approximately equal for both cases in eqn (3). N_a ultimately depends on film drop formation mechanisms that determine the number and size of droplets, which are assumed to be consistent between experiments. Our unexpected result warrants revisiting this assumption.

Studies have shown that the film drop production mechanism is influenced by various environmental factors including temperature, salinity, viscosity, surface tension, and potentially the presence of insoluble contaminants such as micro- and nanoplastic pollution.[56,67] A number of studies – summarized in Table II of the study by Gupta[42] – have found that the presence

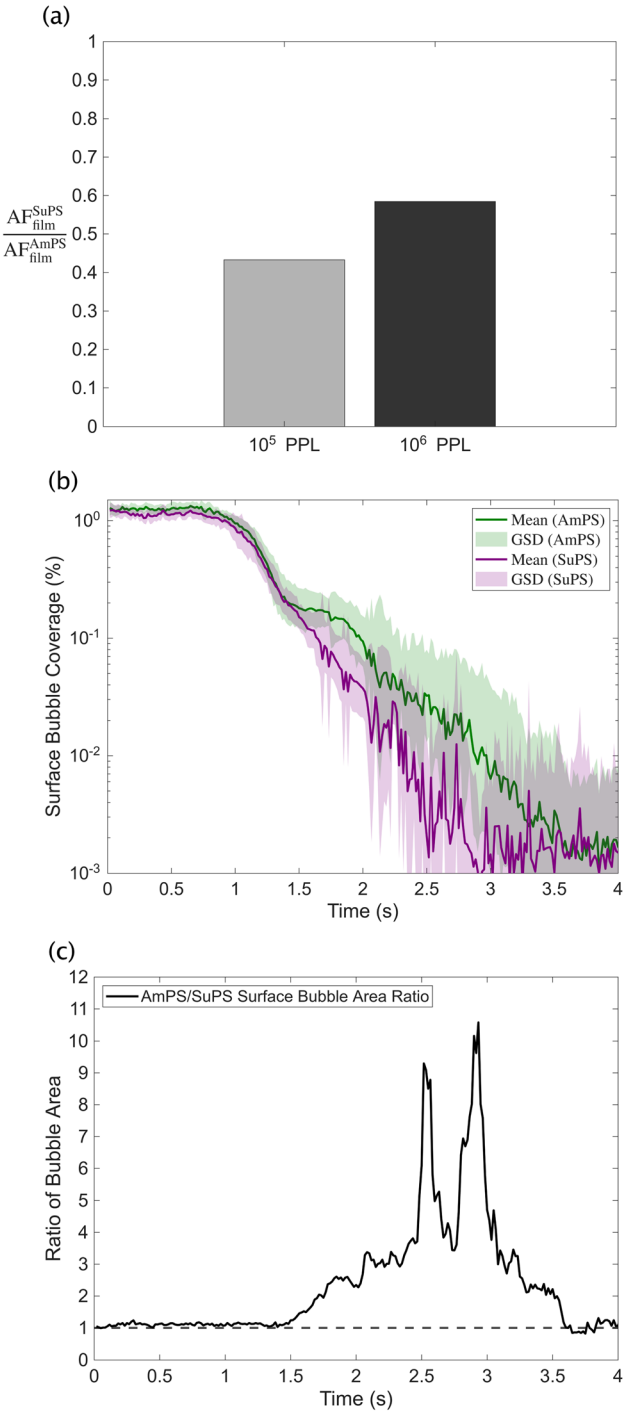

Fig. 3 (a) Ratio of film drop aerosolization factor (AF) for hydrophilic AmPS to hydrophobic SuPS at two MP number concentrations in water. (b) Evolution of the percentage of the tank surface area covered by coarse bubbles over a period of four seconds after the cessation of active bubbling for AmPS- and SuPS-containing ultrapure water ($N = 15$ each). Solid lines represent the geometric mean, and the shaded areas represent ±1 geometric standard deviation (GSD). (c) Ratio of the percentage area covered (AmPS to SuPS) shown in (b), highlighting the difference in bubble persistence.

of particulates in deionized water tends to increase bubble lifetime at the surface. As is well-established, a longer bubble lifetime generally corresponds to a thinner film, as the bubble

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has more time to drain,[67] which can return a portion of the scavenged MPs back to the bulk water.[36]

The role of particulate wettability in bubble stability and bursting dynamics appears complex. Some studies suggest that hydrophobic particles perforate films and reduce bubble lifetime,[68] while others[69] have found that hydrophobic nanoparticles increase bubble lifetime. Nano-sized particles have been shown to impact surface tension[70] and viscosity,[71] but their observed effects remain inconsistent and contradictory.[42,72,73]

Surface bubble behavior and MP concentration effects on film drop emission

Here, we hypothesize that hydrophobic SuPS and hydrophilic AmPS influence the bubble bursting mechanism in different ways by locally altering the surface tension and viscosity of the bubble cap, thus affecting the drainage time, film thickness before rupture, and ultimately governing droplet-mediated MP aerosolization.[42]

Therefore, to find preliminary evidence supporting this hypothesis, we conducted two additional sets of complementary, exploratory experiments: one examining how aerosolization varies with MP concentration in water, and another characterizing the temporal evolution of surface bubbles in SuPS- and AmPS-containing solutions.

In the first set, we examined how aerosolization behavior changes with varying MP number concentrations in water. The ratio of AF_{film}^{SuPS} to AF_{film}^{AmPS} shifted accordingly, as shown in Fig. 3a. At higher MP concentrations in water, we expect the bubble cap film to contain even more SuPS than AmPS due to bubble scavenging favoring hydrophobic particles. As a result, any differences in how SuPS and AmPS influence film drop production may have become more pronounced at higher MP concentrations – as is indeed the case.

In the second set, we examined the temporal evolution of the surface bubbles following the cessation of active coarse bubble formation in AmPS- and SuPS-containing ultrapure water. Given our limitations in directly measuring bubble thickness and the lifetime of individual bubbles, which came at the expense of our more realistic multi-bubble setup, this set of experiments was designed to provide a general statistical sense of bubble persistence and, by extension, bubble lifetime and thickness when AmPS and SuPS are added to ultrapure water. Fig. 3b shows how the percentage of the surface area covered by bubbles evolves over the span of four seconds after the cessation of active coarse bubble formation for both AmPS- and SuPS-containing water. The decay in area starts with a delay of approximately 700 ms, which is consistent with the theoretical time required for ~1 mm air bubbles to rise approximately 40 cm to the surface. The ratio of these percentages (AmPS to SuPS) – shown in Fig. 3c – is mostly greater than one, indicating that bubbles in SuPS-containing ultrapure water dissipated more rapidly. This trend is further reflected in the cumulative decay time scale, defined as the time required for the bubble-covered area to drop below 1% of its initial value. For SuPS, this time was 2314 ms (95% CI: 2140–2489 ms), noticeably shorter than the 3067 ms (95% CI: 2694–3485 ms) observed for AmPS. These results suggest that the surface bubble population in SuPS-containing ultrapure water had a shorter collective persistence, indicating a reduced bubble lifetime. While not exhaustive, these complementary experiments provide indirect evidence consistent with our hypothesis that SuPS and AmPS may have locally altered the fluid properties in different ways, thus affecting the mechanism of MP aerosolization. We emphasize that these results are intended to provide a mechanistic basis for, and motivate future detailed investigations into, the *in situ* dynamics of bubble film rupture and drainage, specifically focusing on how these processes are influenced by micro- and nanosized pollutants.[67]

Broader implications and environmental relevance

The physical principles of wettability isolated in this study are likely relevant to real-world processes in the ocean, although we acknowledge that the presence of salts, surfactants, and organics in seawater will modulate the magnitude of these enrichment effects. For example, the elevated ionic strength in seawater due to the presence of salts can compress electric double layers around particles and bubbles, thus reducing electrostatic repulsion between them.[27,47] This reduced energy barrier would likely promote attachment even for moderately hydrophilic particles, thus narrowing the contrast between hydrophobic and hydrophilic cases. While wettability likely becomes a weaker discriminating factor during the attachment phase of bubble–particle scavenging (a process involving collision, attachment, and stability[44,47]), it still likely determines the bubble–particle stability efficiency under turbulent ocean conditions.[47,74] At the same time, surface-active materials present in seawater (such as fatty acids, proteins, lipids, *etc.*) can adsorb to particle surfaces thus modifying interfacial properties such as wettability and surface charge.[25] With this context in mind, our results provide a basis for speculating on key implications for MP emission from the ocean surface – bearing in mind that such effects will be further influenced by the complex biogeochemistry of natural seawater.

Our findings suggest that hydrophobic MPs are more likely to dominate larger SSA droplets emitted by jet drops and consequently facilitate the emission of larger MPs (up to 280 μm, as observed by Shaw *et al.*[8]), indicating that wettability may act as a selection mechanism for larger MPs during aerosolization. Hydrophobic MPs can adsorb hydrophobic organic pollutants such as polycyclic aromatic hydrocarbons (PAHs) and polychlorinated biphenyls (PCBs),[75] suggesting a potential pathway for the long-range atmospheric transport of these pollutants. In contrast, both hydrophilic and hydrophobic MPs can be incorporated into the relatively smaller SSA droplets emitted by film drops. As the estimated 3400 kilotonnes of floating plastics currently in the ocean[76] undergo further solar degradation – leading to fragmentation and increased wettability – the likelihood of their aerosolization *via* film drops increases.

While jet drops may contribute more significantly to the total mass of MPs aerosolized in the atmosphere, film drops are known to dominate SSA number concentrations, accounting for

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60 to 80% of the submicrometer SSA fraction.[52] The aerosolization of hydrophilic MPs *via* smaller film drops may make them more likely to act as cloud condensation nuclei, influencing cloud microphysics and precipitation processes.[33–35] Additionally, hydrophilic MPs could undergo hygroscopic growth, potentially altering their size, optical properties, and atmospheric residence time.

Our identification of particle wettability as a governing factor in aerosolization may help refine estimates of oceanic contributions to atmospheric MPs. According to a recent study by Yang *et al.*,[21] the theoretical upper limit of the global sea–air emission flux of sub-100 μm MPs was estimated to be on the order of 10^{-2} megatons per year, based on current knowledge of oceanic MP concentration, SSA flux, and film and jet drop enrichment models developed by ref. 25, 36 and 37. These estimates are based on enrichment models developed in single-bubble bursting experiments in pure or tap water and have not considered MP wettability. Thus, they could change after factoring in the hydrophobicity of micro- and nanosized plastics present in the oceans. As discussed earlier, Dubitsky *et al.*[24] theoretically modeled the effect of MP wettability and estimated that hydrophobic MPs can be selectively enriched by several orders of magnitude higher in jet drops compared to film drops. While their film drop enrichment model does acknowledge the effect of particle size in the film bursting mechanism – by assuming that the particle will initiate bursting when the film thickness approaches the particle size – it does not incorporate the potential impact of particle wettability on bursting dynamics, as suggested by our film drop aerosolization and surface bubble area decay results. Additionally, they considered hydrophilic particles to be perfectly non-attaching – granted for the purposes of estimating emission bounds – but we demonstrate that hydrophilic particles do aerosolize *via* film and jet drops despite the lack of wettability-governed interactions, and thus can have a non-zero attachment efficiency – a metric used to quantify particle wettability in the enrichment models. We note that previous studies have revealed that hydrophilic particles can still exhibit attachment to rising bubbles even without wettability-governed interactions.[77,78]

Incorporating particle wettability into enrichment models could therefore improve the accuracy of MP emission estimates. Future studies employing such refined models will require not only accurate spatial distributions of MPs at the ocean surface but also detailed characterization of their physicochemical properties. Our observations of the absence of a clear effect of MP wettability on aerosolization *via* film drops, and limited prior work on particle enrichment onto film drops[36,79,80] – compared to numerous studies on jet drops[4,8,25,26,37,81–84] – further highlight the need for improved mechanistic understanding of bubble bursting and drop enrichment dynamics.

Finally, our demonstration of wettability as a key factor in MP aerosolization highlights the importance of conducting future laboratory studies in realistic seawater. Other surface properties – such as surface charge, particle shape, roughness, and biofilm colonization – may also play important roles in enrichment and atmospheric transport.[8,42,68] While some theoretical studies have begun to include these factors – for instance, Ji *et al.*[25] considered particle shape and Dubitsky *et al.*[24] incorporated particle wettability – there remains a pressing need for experimental validation under environmentally representative conditions.

Conclusions

The goal of this study was to conduct a controlled experiment to investigate the effect of particle wettability on aerosolization *via* film and jet drops. Droplets were attributed to either the film or jet drop pathway based on their generation from two distinct, well-characterized bubble populations known from the literature to predominantly produce their respective drop types.[53] To isolate the effect of wettability, we kept constant the properties of the bubbles (fine or coarse), water (ultrapure), and MPs (polymer type, size, shape, concentration, density, and surface charge). Our results demonstrate that wettability of MP particles significantly influences their aerosolization, but in different ways for film and jet drops. The jet drop aerosolization factor of hydrophobic SuPS was approximately one order of magnitude higher than that of hydrophilic AmPS. In contrast, no significant difference was observed in film drop aerosolization factor. We argue that this discrepancy arises from the complex and contradictory effects of particle presence and surface properties on local bubble film behavior, which in turn affect film drop production and particle transfer. Together, our findings highlight the nuanced role of particle surface properties in shaping the aerosolization of microplastics *via* bubble bursting. These results underscore the importance of interfacial processes in governing microplastic emissions to the atmosphere and motivate continued investigation across different ejection pathways and environmental conditions.

Author contributions

Nishan Pokhrel: conceptualization, data curation, formal analysis, investigation, methodology, visualization, writing – original draft, writing – review and editing. Hosein Foroutan: conceptualization, formal analysis, funding acquisition, methodology, project administration, supervision, validation, writing – review and editing.

Conflicts of interest

There are no conflicts to declare.

Data availability

Data used to plot the figures in this article are available at Figshare at https://doi.org/10.6084/m9.figshare.29906480.v1.

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