



Rapid screening of CO₂ capture fluids†

 Cite this: *Lab Chip*, 2025, 25, 2918

 Yaohao Guo,^{ab} Feng Li,^b Sepehr Saber,^b Mohammad Zargartalebi,^b Siyu Sonia Sun,^b Yurou Celine Xiao,^{id} Bo Bao,^{id} *^a Zhi Xu,^{id} *^a and David Sinton,^{id} *^b

The evaluation of CO₂ capture fluids is crucial for the advancement of carbon capture technologies. Recent advancements in amine-based carbon capture fluids motivate a broad search for high-performance fluids and the development of methods capable of exploring a large chemical space. Here, we present a microfluidic approach paired with automated image processing and density functional theory simulations that enables comprehensive rapid screening of capture fluids. The principle of measurement leverages the ability to monitor phase expansion and contraction in fixed-volume dead-end channels. This approach enables fast comparative assessments of reaction kinetics and thermodynamic parameters, including CO₂ absorption rate (~30 s), desorption rate (~30 s), absorption capacity (~20 min), and vapor pressure (~5 min), exceeding the speed of conventional methods by two orders of magnitude. The method is broadly applicable, effective for primary, secondary, and tertiary amine types. Rapid screening of capture fluids holds promise for the accelerated discovery of improved CO₂ capture processes and an opportunity for the microfluidics community to contribute to decarbonization efforts.

 Received 17th September 2024,
 Accepted 12th May 2025

DOI: 10.1039/d4lc00772g

rsc.li/loc

1. Introduction

Increasing emissions of CO₂ underscore the imperative need for CO₂ capture technologies as a strategy towards achieving carbon neutrality.^{1,2} Among the major types of CO₂ capture solutions developed to date – amine solutions, alkaline solutions, ionic liquids, and ammonia – amines are the most developed.^{3–5} With decades of research and development, aqueous amine solvents are now employed to capture CO₂ from a wide variety of post-combustion flue gases.^{6,7} The typical requirements for a CO₂ capture fluid are high absorption rate and capacity, selectivity, and energy efficiency in desorption, as well as exhibiting low vapor pressure, corrosiveness, and toxicity.^{8,9} However, current capture solutions do not meet these requirements in full. For instance, the widely used 30 wt% MEA aqueous solution exhibits rapid reaction kinetics but exhibits low absorption capacity and suffers high desorption energy demand.^{10,11} Recent advancements in material science, combined with in-depth investigations of CO₂ capture systems, have yielded a diverse array of novel capture fluids and hybrid systems.^{12,13} These efforts broaden the spectrum of promising chemistries and present a testing bottleneck.¹⁴ Blends of capture solutions

are the norm, and with over 150 common capture solvents in use and more under development,¹⁵ the chemical space is vast, with over 10⁷ potential combinations to explore.

Current approaches to evaluate capture fluid performance involve, in effect, smaller versions of the capture system.^{16–18} To determine absorption and desorption rates, for instance, bubble column reactors, wetted wall columns, and pack towers are used. These approaches are batch and low-throughput, with test times on the order of hours.^{19–21} Furthermore, the substantial sample volumes needed, ranging from tens of milliliters to a few liters, contribute to inefficiency and cost.²² These coupled time- and volume-requirement limitations are analogous to the biomedical laboratory processes that have been disrupted by lab-on-chip technologies over the last few decades.^{23,24} Microfluidic technology has demonstrated exceptional versatility and broad applicability in CO₂-related studies, spanning capture, conversion, and monitoring applications.^{25–27} Adapting the lab-on-chip approach to advance decarbonization technology is needed for the decades ahead.^{28,29}

Herein, we introduce a microfluidic approach for the comprehensive and high-throughput screening of CO₂ capture fluids, integrating automated image analysis and density functional theory (DFT) simulations for enhanced efficiency and accuracy. The microfluidic approach leverages dead-end channels as gas–liquid reactors to assess CO₂ absorption/desorption rates, absorption capacity, and vapor pressure simultaneously. This approach circumvents the complexities associated with dynamic gas–liquid segmented flows and provides a stable and controllable gas–liquid interface. Notably, variations in fluid properties may introduce deviations in

^a School of Chemical Engineering, East China University of Science and Technology, Shanghai, 200237, China

^b Department of Mechanical & Industrial Engineering, University of Toronto, Toronto, M5S 3G8, Canada. E-mail: sinton@mie.utoronto.ca

† Electronic supplementary information (ESI) available. See DOI: <https://doi.org/10.1039/d4lc00772g>



capture performance evaluation. The DFT simulations are used to calculate the Gibbs free energy profiles, offering theoretical insights into the intrinsic reaction kinetics between CO₂ and capture fluid. Additionally, comparative analysis with microfluidic data helps distinguish genuine performance from experimental artifacts. This combined approach enables an assessment of both the potential of a given capture fluid and how it would perform in practice, and provides an understanding of the underlying mechanisms that can inform the next test or formulation.

2. Methodology and experimental design

The microfluidic assay was based on the 1-D fixed-volume dead-end channels that have been proven robust in many lab-on-chip applications. Examples included assessing CO₂-oil interactions,^{30,31} thousands of such channels applied to assess phase properties of mixtures,³² and colloidal particle sorting.³³ The channels here were 300 μm wide and 250 μm deep, and distributed in a parallel array. These channels functioned as miniaturized gas-liquid reactors, as illustrated in Fig. 1a. Initially, CO₂ gas was introduced into these channels under a predefined pressure, followed by the infusion of the capture fluid, thus triggering the reaction under test. Additionally, we monitored the pressure during the introduction of the capture fluid to gain insights into the thermodynamic conditions within the channel. During the subsequent absorption process, the pressure was assumed to remain constant due to the small volume of trapped CO₂. The structure ensured the constancy of the gas and liquid segments' diameter, with volume variations restricted to the channels' longitudinal axis. Taking amine solutions as an example, at room temperature, the gas slug in the dead-end shrank gradually as the solvent efficiently absorbs CO₂. Conversely, elevated temperatures facilitated the reverse reaction, promoting amine regeneration and liberation of CO₂, ultimately resulting in an enlargement of the gas slug. These temperature-dependent reactions and interface dynamics served as indicators for assessing the absorption and desorption rates. The absorption capacity measurement shared similarities with that for determining CO₂ absorption rates. However, to assess capacity a finite-length absorbent slug was introduced into the test channel and then segmented by a gas phase, leading to an excess of CO₂ and a deficiency of absorbent within the microchannels (see Fig. 1a). As the absorbent within the slug becomes depleted, the final position of the slug reflected its absorption capacity. To assess vapor pressure measurements, we invoked the proportional relationship between vapor pressure and evaporation rate.³⁴ Each experiment was conducted three times, and for each run, the reported result represents the average across all channels imaged simultaneously.

Microfluidic measurements were integrated with automated image analysis and DFT simulations to expedite and enrich the evaluation of CO₂ capture fluids (Fig. 1b). Specifically, DFT simulations were applied to ascertain the Gibbs free energy changes along the reaction pathways, which — together with

adsorption and desorption measurements — indicated the potential of the capture fluid and its applicability. These experimental measurements were implemented using a microfluidic device (Fig. 1c), where real-time interfacial dynamics during CO₂ absorption/desorption cycles were tracked *via* automated imaging (Fig. 1d, ESI† Appendix section S1) and analyzed in conjunction with the gas state equation.

Fig. 2 shows the schematic diagram of the experimental setup for high-throughput screening of CO₂ capture fluids. The central component of our microfluidic screening platform is a customized microfluidic chip (75.2 mm × 35 mm × 1.5 mm). Its fabrication starts with creating patterns on a chromium mask, which are transferred onto a silicon substrate *via* UV lithography, followed by deep reactive ion etching (RIE) to achieve 250 μm channel depths. The silicon substrate is then anodically bonded to a borosilicate glass cover, forming tightly sealed microchannels that allow direct optical observation of fluidic behaviors. This silicon-glass construction provides excellent chemical resistance and can withstand high temperatures (~200 °C) and pressures (~200 bar). A high-pressure syringe pump (Teledyne ISCO 200DX) delivers CO₂ or N₂ gas to maintain experimental pressure, while a high-precision syringe pump (Harvard Apparatus, Pump 11 Elite) continuously injects the capture solution at a constant rate. Before experiments, a vacuum pump evacuates the system to ensure gas purity. Pressure sensors (Omega, PX409) monitor gas and liquid pressures at the inlet and outlet channels.

The experimental procedures for CO₂ capture performance evaluation comprise four key tests: absorption rate, desorption rate, absorption capacity, and vapor pressure comparison.

2.1. Absorption rate test

CO₂ is injected into the chip through port *a* and maintained at a target pressure (~2.15 bar), while a 3 mol L⁻¹ aqueous amine solution (35 °C) is introduced through port *c*. A T-shaped junction upstream of the testing channels diverts pre-reacted fluid to port *b*, ensuring only fresh solution enters the dead-end channels. Once unreacted fluid is confirmed at the T-junction, port *b* is closed to direct the solution into the testing channels. To ensure consistency and robustness of the reported results, we present the averaged response across multiple channels.

2.2. Desorption rate test

Following the absorption rate protocol, capture solution injection ceases when the gas-liquid interface reaches the appropriate position. N₂ is then injected through port *a* to purge residual solution from the main channel. Ports *a* and *c* are vented to atmospheric pressure, and the chip is heated to 70–80 °C to initiate CO₂ release from the amine solution.

2.3. Absorption capacity test

After replicating the absorption rate procedures, N₂/CO₂ is injected through port *a* to flush residual solution from the main channel, forming a finite liquid slug in the testing





Fig. 1 Microfluidic-driven comprehensive and fast screening of CO₂ capture fluids. (a) Operational principle and schematic representation of microfluidic measurement for CO₂ absorption rate, desorption rate, absorption capacity and vapor pressure. Different colors on the chip denote distinct temperature regions, with blue indicating the low-temperature region at 35 °C and red signifying the high-temperature region at 80 °C. The cartoon diagram illustrates the movement of the interface and the reaction process during different parameter measurements, where the intensity of absorbent color reflects the concentration distribution of reaction products. (b) Microfluidic measurements are integrated with automated image analysis and DFT simulations to establish an accelerated platform for CO₂ capture fluids screening. (c) Schematic diagram of experimental setup. The microfluidic device consists of a stainless-steel manifold that secures the silicon-glass chip, providing enclosed fluid pathways. Temperature control is achieved through a copper block and an electric heater. The chip features multiple dead-end channels arranged for the comparison of amine solutions' performance by monitoring the movement of gas-liquid interface. (d) Automated image analysis and performance parameters calculation. The CO₂-amine interface is detected *via* intensity changes using the Sobel edge detection method. Interface position and velocity are calculated along a predefined coordinate axis. The absorption rate, desorption rate, and absorption capacity are then determined through a gas equation of states.

channel. Subsequently, port *c* is closed, and N₂/CO₂ is maintained at target pressure. CO₂ occupies the slug's left

side, while N₂/CO₂ fills the right side. Tests use 1 mol L⁻¹ amine solution at 25 °C.



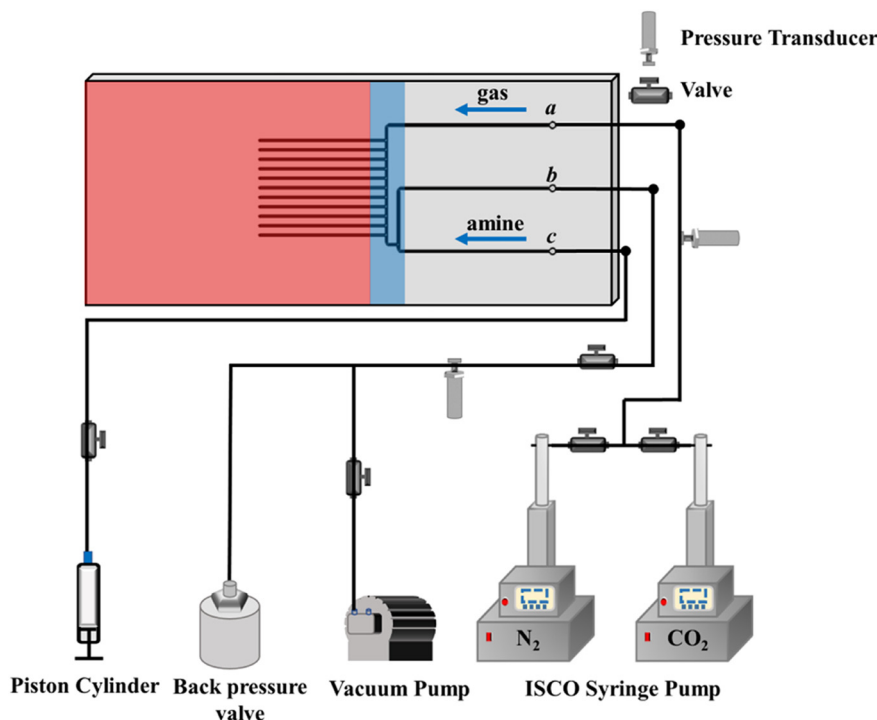


Fig. 2 Schematic diagram of experimental setup for CO₂ capture fluids screening.

2.4. Vapor pressure comparison

The microfluidic system is evacuated, followed by the injection of capture fluid to saturate the dead-end testing channels. N₂ purges residual solution from the main channel *via* port *a*. The chip is heated to 35 °C or 80 °C to initiate evaporation, with N₂ continuously supplied at ~1.1 bar to accelerate evaporation and minimize atmospheric CO₂ interference.

3. Results and discussion

3.1. Microfluidic results

We selected six different types of amines (monoethanolamine, pentylamine, diethylamine, diethanolamine, *N*-methyldiethanolamine, triethanolamine) as model CO₂ capture solutions. This set includes the most commonly used amines and covers primary, secondary, and tertiary amine types (details in Table S1 of the ESI† Appendix).

3.1.1. Absorption rate and desorption rate measurements.

Absorption and desorption rates are critical metrics of CO₂ capture fluid performance, influencing the scale and capital cost of a capture system. Initial CO₂ absorption tests were performed at 35 °C and 2.15 bar. Upon introducing amine solutions into the dead-end channels, a noticeable absorption of CO₂ is observed, as evidenced by the gradual reduction in gas slug length and the leftward shift of the gas–liquid interface, as illustrated in Fig. 3a. Initially, the rapid shrinkage of the gas slug indicates the chemical reaction rate between the gas and absorbent. As the interface-adjacent amine was depleted, a subtle deceleration was observed, indicating the onset of a mass transfer

limitation. All test fluids were confronted with this diffusion-limiting phenomenon, thus diffusive effects at later times have minimal effect on the comparison of absorption rates especially within a short period of time. Fig. 3b depicts the time-dependent evolution of the gas–liquid interface movement for various amine solutions, with shaded areas representing the standard deviation among different dead-end channels. By quantifying the changes in gas slug volume over a 10-second interval and employing the real gas equation, we ascertain the average CO₂ absorption rates, as shown in Fig. 3c. For these calculations, the gas state is modeled using the Van der Waals equation:

$$\left(P + \frac{an^2}{V^2}\right)(V - nb) = nRT \quad (1)$$

where P represents the gas pressure, V denotes its volume, n is the quantity of gas (in moles), R is the ideal gas constant, and T is the gas's absolute temperature. The constants a and b , specific to CO₂, are 3.66 bar L² mol⁻² and 0.0429 L mol⁻¹, respectively.³⁵ The observed order of absorption rates for different amine solutions generally aligns with that reported in the literature, with primary and secondary amines exhibiting higher absorption rates than tertiary amines due to their smaller steric hindrance and distinct reaction mechanisms.³⁶ Notably, our study achieves the determination of absorption rates within a matter of seconds, surpassing conventional methodologies by orders of magnitude.²¹ This approach leverages rapid heat transfer characteristic of microfluidics to minimize the effect of exothermic heat of reaction, thereby allowing the absorption tests to proceed



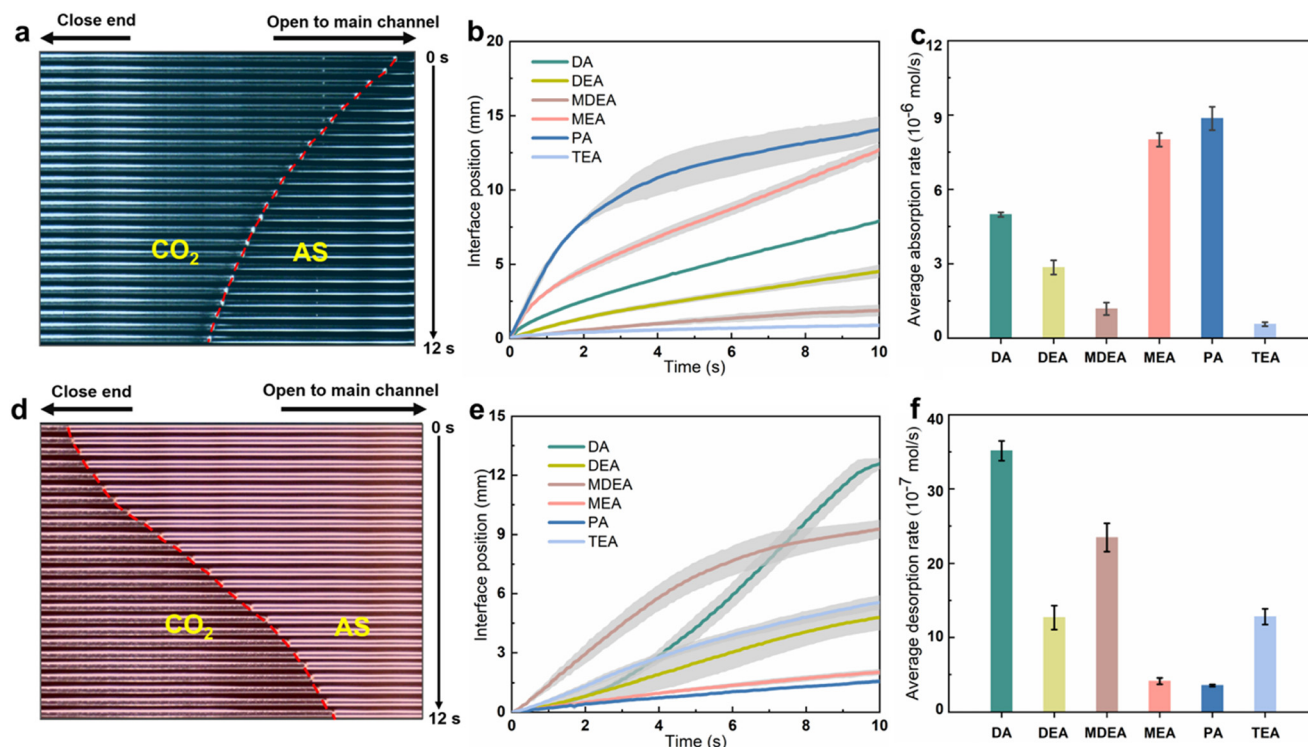


Fig. 3 CO₂ absorption and desorption tests at 35 °C and 80 °C, respectively. (a) Dynamic interface movement during CO₂ absorption: the red dashed line outlines the interface, with amine solution on the right and CO₂ on the left; (b) temporal evolution of interface displacement in absorption tests, with gray shading indicating standard deviation from at least 3 experimental sets; (c) calculation of average CO₂ absorption rate based on the real gas state equation (Van der Waals equation, $a = 3.66 \text{ L}^2 \text{ mol}^{-2}$, $b = 0.0429 \text{ L mol}^{-1}$) and interface displacement within 10s ($T = 35 \text{ °C}$, $P = 2.15 \text{ bar}$); (d) dynamic interface movement during CO₂ desorption: the red dashed line outlines the interface, with amine solution on the right and CO₂ on the left; (e) temporal evolution of interface displacement in desorption tests; (f) calculation of average CO₂ desorption rate within 10s ($T = 80 \text{ °C}$, $P = 1 \text{ bar}$).

under nearly isothermal conditions – a challenge for conventional testing setups.³⁷

The CO₂ desorption test follows the CO₂ absorption test. Here, rapid heating on the microchip is employed to contain the diffusion of reaction products within the test channels. The system can elevate the chip's temperature to $\sim 70 \text{ °C}$ within $\sim 6 \text{ s}$ and further to 80 °C after $\sim 16 \text{ s}$ (refer to section S3 of ESI† Appendix). Within a time frame of less than 100 seconds and the absence of convection, the diffusion distance remains under 1 mm (section S4 of ESI† Appendix). This limited diffusion ensures that the absorption and desorption reactions occur in close proximity to the gas–liquid interface (see Fig. 1a), approximating a closed system within the test channel. The heating of the chip instigates the release of absorbed CO₂, leading to an increase in pressure within the residual gas slug and a gradual shift of the gas–liquid interface towards the right (Fig. 3d).

Fig. 3e shows the interface movement, indicating the thermal response associated with CO₂ desorption in various amine solutions. The expansion of the gas segment is not solely influenced by CO₂ release, but also by the combined effects of thermal expansion and vapor pressure changes. With regard to vapor pressure, the capture fluids (3 mol L^{-1} aqueous solutions) possess a solute molar fraction below 0.1. According to Raoult's law, the vapor pressure of the

solution is primarily determined by water, provided that the vapor pressure of the solute is lower than that of water (See section S5 of ESI† Appendix). Consequently, the contribution of vapor pressure to the expansion of the gas slug is largely consistent across different test fluids, rendering it negligible in the comparative analysis of desorption rates. The effect of thermal expansion can be compensated for using the gas state equation. Desorption rate is defined as the average rate over 10 seconds, and detailed outcomes are presented in Fig. 3f. Notably, the desorption rate trend generally inverts that of absorption, with tertiary amines exhibiting more rapid CO₂ release than primary and secondary amines. This difference is primarily attributed to the simpler decomposition of bicarbonate (main reaction product between tertiary amines and CO₂) compared to that of carbamate (main reaction product between primary/secondary amines and CO₂).^{19,38,39} An anomalous case is observed with diethylamine (DA), which exhibits the highest desorption rate among all examined amine solutions, deviating from expected behaviour.⁴⁰ This anomaly is due to its relatively high vapor pressure at 80 °C , DA contributes a substantial partial pressure ($\sim 16 \text{ kPa}$) despite its low mole fraction, making its effect on gas slug expansion during desorption non-negligible compared to other amines (see ESI† section S5).



3.1.2. Absorption capacity measurement. In the absorption capacity tests, as the amine in the liquid slug is depleted, the final position directly indicates the absorption capacity, as shown in Fig. 4a. Absorption capacity can be quantified as M_g/M_s when the liquid slug is segmented by N_2 (or absorption capacity = $2M_g/M_s$ when the liquid slug is segmented by CO_2), where M_g represents the moles of CO_2 absorbed (determined by the reduction in gas slug size) and M_s denotes the moles of absorbent in the liquid plug. The bar chart (Fig. 4b) presents the absorption capacities of different amine solutions at a CO_2 partial pressure of 2.65 bar and includes data for monoethanolamine (MEA) at varying partial pressures (higher pressures are attainable with this system, up to ~ 20 MPa (ref. 41)). The results are consistent with established literature, exhibiting a deviation of less than 6% (see section 6 of ESI† Appendix), providing validation of the experimental approach.^{42–44} We attribute the residual error to the determination of the small volumes of capture fluid and absorbed CO_2 . Challenges also arise with certain amines, such as pentylamine (PA) and Triethanolamine (TEA), which show a strong affinity and lower contact angles towards the microchannel walls, hindering the formation of stable liquid segments. Modifications to the channel wall surface wettability could address this challenge in future.

3.1.3. Vapor pressure assessment. Vapor pressure governs the solution's volatility during the absorption and regeneration

processes of carbon capture, impacting both system efficiency and operational costs. Typically, direct measurement of a fluid's vapor pressure requires elaborate experimental procedures or sophisticated instrumentation.⁴⁵ Here we infer the vapor pressure from the evaporation rate of fluids in microchannels.³⁴ The test channel is initially filled with capture fluid, with the gas–liquid interface shifting towards the channel end (Fig. 4c). To expedite the evaporation rate while preventing the formation of salts or other reaction products with CO_2 , N_2 is introduced into the main channel at a steady pressure of 1.1 bar. The average evaporation rates of various amine solutions over a 5-minute observation period, as presented in Fig. 4d, align with the trends in vapor pressures and boiling points (sourced from PubChem, see section S7 of ESI† Appendix). Amines such as diethanolamine (DEA), *N*-methyldiethanolamine (MDEA), and triethanolamine (TEA), known to have low vapor pressures, exhibit low evaporation rates at room temperature. To accelerate testing, we employ heating (to 80 °C here, Fig. 4d, and a maximum of 200 °C generally) for comparative analysis in low-vapor-pressure fluids.

3.2. DFT simulation

The microfluidic measurements leverage the dynamics of the gas–liquid interface to evaluate the reaction kinetics and thermodynamic parameters involved in CO_2 capture. Within the

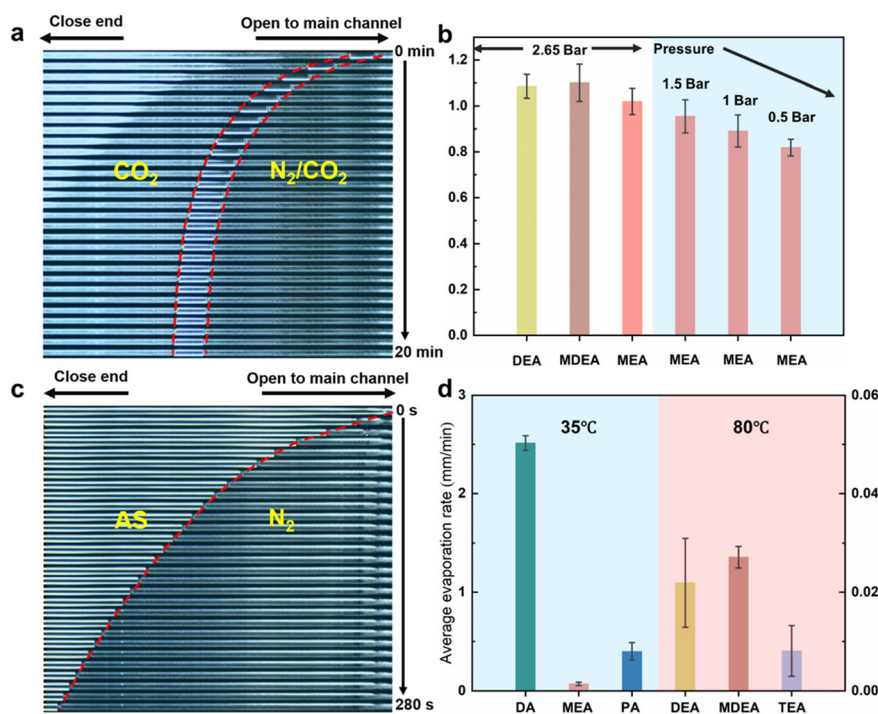


Fig. 4 CO_2 absorption capacity and vapor pressure comparison tests. (a) Dynamic movement process of absorbent slug, with the final position reflecting the absorption capacity for CO_2 ; (b) absorption capacities of DEA, MDEA and MEA at a CO_2 partial pressure of 2.65 bar, along with the variation in MEA absorption capacity under different CO_2 partial pressures; (c) dynamic interface movement in vapor pressure tests. The evaporation process of the amine occurs within an N_2 environment; (d) average evaporation rate of different amines. Experiments for amines with higher vapor pressures (*i.e.*, DA, MEA, PA) are conducted at a lower temperature (35 °C), while experiments for amines with lower vapor pressures (*i.e.*, DEA, MDEA, TEA) are conducted at a higher temperature (80 °C) to shorten the testing duration.





Fig. 5 DFT calculated energy barrier diagram of the CO₂ and amine reaction in aqueous solution. H, white; C, gray; N, blue; O, red.

quasi-closed dead-end channels, the movement of the interface is primarily governed by the absorption or desorption of CO₂. However, the interface movement it is also influenced by fluid properties such as interfacial tension and viscosity. To further corroborate that the experimental data accurately reflects the reaction rates, DFT simulations were employed to assess CO₂ and absorbent molecule interactions, with further details available in section S8 of ESI† Appendix. The changes in Gibbs free energy and transition states during CO₂ adsorption on six amine variants are illustrated in Fig. 5.

The energy barriers for CO₂ absorption and desorption are represented by $\Delta G_{\text{TS-IS}}$ and $\Delta G_{\text{TS-FS}}$, respectively. DFT calculations reveal a CO₂ absorption kinetic sequence of DA \approx PA \approx MEA $>$ DEA \approx MDEA $>$ TEA, while exhibiting an inverse trend in desorption. This ordering matches that of the experimental data. However, limitations in each methodology lead to some discrepancies. The theoretical prediction for desorption kinetics for DA is slow, in stark contrast to the experimental result showing the fastest CO₂ desorption rate. This discrepancy stems from the significant influence of DA's vapor pressure on experimental outcomes (see section S5 of ESI† Appendix). Similarly, DFT predicts comparable absorption or desorption kinetics for some capture fluids, in contrast to the differences observed in the experimental results. This may be attributed to the influence of fluid properties and mass transfer processes in the liquid phase – aspects not captured by DFT. Thus, the microfluidic experiments assess apparent reaction rates while DFT calculations reveal intrinsic kinetics. The combination enables an assessment of both the potential of a given capture fluid and how it would perform in practice.

4. Conclusion

We introduce a microfluidic-driven framework for comprehensive and rapid screening of CO₂ capture fluids, integrated with automated image processing and DFT simulations for accelerated analysis. The DFT simulations elucidate intrinsic reaction kinetics between capture fluids and CO₂, offering theoretical insights that augment the experimental findings. The reliable, and high-throughput

experimental data enriches theoretical prediction models with empirical evidence.

The microfluidic system employs dead-end channels that circumvent challenges of two-phase dynamic flow reactors. The quasi-static nature of measurement facilitates a stable and controllable interface and improves the versatility of the platform. This design enables a rapid comparative assessment of capture fluid performance in terms of CO₂ absorption rate (~ 30 s), desorption rate (~ 30 s), absorption capacity (~ 20 min), and vapor pressure (~ 5 min), achieving a speed that surpasses traditional methodologies by two orders of magnitude. The integration of microfluidic measurements with automated image processing and DFT simulation provides a means to screen, develop, and ultimately discover CO₂ capture fluids. When extending this approach to novel fluids, practical implementation may benefit from parameter optimization (*e.g.*, flow rates, pressure) under extreme conditions (such as exceptionally high viscosity or low surface tension) and supplementary characterization (*e.g.*, viscosity, vapor pressure) to contextualize performance deviations, while the demonstrated success with diverse amine solutions confirms its foundational reliability for scalable CO₂ capture fluids discovery.

Data availability

The data and code supporting this article have been included as part of the ESI†

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

The authors would like to express their sincere gratitude for the financial backing provided by the Natural Sciences and Engineering Research Council of Canada (NSERC), and the Canada Research Chairs Program (CRC-2021-00316). D. Sinton holds a Canada Research Chair in Fluids and Energy.

References

- 1 N. MacDowell, N. Florin, A. Buchard, J. Hallett, A. Galindo, G. Jackson, C. S. Adjiman, C. K. Williams, N. Shah and P. Fennell, *Energy Environ. Sci.*, 2010, **3**, 1645–1669.
- 2 D. Y. Shu, S. Deutz, B. A. Winter, N. Baumgärtner, L. Leenders and A. Bardow, *Renewable Sustainable Energy Rev.*, 2023, **178**, 113246.
- 3 F. Shakerian, K.-H. Kim, J. E. Szulejko and J.-W. Park, *Appl. Energy*, 2015, **148**, 10–22.
- 4 T. N. Borhani and M. Wang, *Renewable Sustainable Energy Rev.*, 2019, **114**, 109299.
- 5 F. O. Ochedi, J. Yu, H. Yu, Y. Liu and A. Hussain, *Environ. Chem. Lett.*, 2021, **19**, 77–109.



- 6 F. Meng, Y. Meng, T. Ju, S. Han, L. Lin and J. Jiang, *Renewable Sustainable Energy Rev.*, 2022, **168**, 112902.
- 7 S. Laribi, L. Dubois, G. De Weireld and D. Thomas, *Int. J. Greenhouse Gas Control*, 2019, **90**, 102799.
- 8 O. Aschenbrenner and P. Styring, *Energy Environ. Sci.*, 2010, **3**, 1106–1113.
- 9 A. Gautam and M. K. Mondal, *Fuel*, 2023, **334**, 126616.
- 10 L.-C. Lin, A. H. Berger, R. L. Martin, J. Kim, J. A. Swisher, K. Jariwala, C. H. Rycroft, A. S. Bhowm, M. W. Deem and M. Haranczyk, *Nat. Mater.*, 2012, **11**, 633–641.
- 11 B. Aghel, S. Sahraie and E. Heidaryan, *Energy*, 2020, **201**, 117618.
- 12 W. Yu, T. Wang, A.-H. A. Park and M. Fang, *Nanoscale*, 2019, **11**, 17137–17156.
- 13 P. K. Chhotaray, S. K. Biswal and S. Pandey, *J. Mol. Liq.*, 2020, **312**, 113477.
- 14 X. Zhang, X. Zhang, H. Dong, Z. Zhao, S. Zhang and Y. Huang, *Energy Environ. Sci.*, 2012, **5**, 6668–6681.
- 15 J. L. McDonagh, S. Zavitsanou, A. Harrison, D. Zubarev, T. van Kessel, B. H. Wunsch and F. Cipcigan, *Digital Discovery*, 2024, **3**, 528–543.
- 16 J.-G. Lu, A.-C. Hua, L.-L. Bao, S.-X. Liu, H. Zhang and Z.-W. Xu, *Sep. Purif. Technol.*, 2011, **82**, 87–92.
- 17 H. Karlsson and H. Svensson, *Energy Procedia*, 2017, **114**, 2009–2023.
- 18 C. Nwaoha, P. Tontiwachwuthikul and A. Benamor, *Int. J. Greenhouse Gas Control*, 2019, **82**, 218–228.
- 19 B. Lv, B. Guo, Z. Zhou and G. Jing, *Environ. Sci. Technol.*, 2015, **49**, 10728–10735.
- 20 I. M. Bernhardsen and H. K. Knuutila, *Int. J. Greenhouse Gas Control*, 2017, **61**, 27–48.
- 21 R. Rivera-Tinoco and C. Bouallou, *J. Cleaner Prod.*, 2010, **18**, 875–880.
- 22 G. Puxty, R. Rowland and M. Attalla, *Chem. Eng. Sci.*, 2010, **65**, 915–922.
- 23 J. Ma, S. M.-Y. Lee, C. Yi and C.-W. Li, *Lab Chip*, 2017, **17**, 209–226.
- 24 H. Shi, K. Nie, B. Dong, M. Long, H. Xu and Z. Liu, *Chem. Eng. J.*, 2019, **361**, 635–650.
- 25 T. Cubaud, M. Sauzade and R. Sun, *Biomicrofluidics*, 2012, **6**(2), 022002.
- 26 D. Voicu, M. Abolhasani, R. Choueiri, G. Lestari, C. Seiler, G. Menard, J. Greener, A. Guenther, D. W. Stephan and E. Kumacheva, *J. Am. Chem. Soc.*, 2014, **136**, 3875–3880.
- 27 M. Abolhasani, A. Günther and E. Kumacheva, *Angew. Chem., Int. Ed.*, 2014, **53**, 7992–8002.
- 28 S. S. Datta, I. Battiato, M. A. Fernø, R. Juanes, S. Parsa, V. Prigiobbe, E. Santanach-Carreras, W. Song, S. L. Biswal and D. Sinton, *Lab Chip*, 2023, **23**, 1358–1375.
- 29 T.-H. M. Ho, J. Yang and P. A. Tsai, *Lab Chip*, 2021, **21**, 3942–3951.
- 30 A. Sharbatian, A. Abedini, Z. Qi and D. Sinton, *Anal. Chem.*, 2018, **90**, 2461–2467.
- 31 Y. Guo, F. Liu, J. Qiu, Z. Xu and B. Bao, *Energy*, 2022, **256**, 124524.
- 32 Y. Xu, J. Riordon, X. Cheng, B. Bao and D. Sinton, *Am. Ethnol.*, 2017, **129**, 14150–14155.
- 33 M. Nooryani, A. M. Benneker and G. Natale, *Lab Chip*, 2023, **23**, 2122–2130.
- 34 D. Mackay and I. van Wesenbeeck, *Environ. Sci. Technol.*, 2014, **48**, 10259–10263.
- 35 S. P. De Visser, *J. Phys. Chem. B*, 2011, **115**, 4709–4717.
- 36 G. Sartori and D. W. Savage, *Ind. Eng. Chem. Fundam.*, 1983, **22**, 239–249.
- 37 J. Gomes, S. Santos and J. Bordado, *Environ. Technol.*, 2015, **36**, 19–25.
- 38 M. Kumari, F. Vega, L. M. G. Fernández, K. P. Shadangi and N. Kumar, *J. Mol. Liq.*, 2023, 122288.
- 39 Z. Wang, M. Fang, Y. Pan, S. Yan and Z. Luo, *Chem. Eng. Sci.*, 2013, **93**, 238–249.
- 40 F. A. Chowdhury, H. Yamada, T. Higashii, K. Goto and M. Onoda, *Ind. Eng. Chem. Res.*, 2013, **52**, 8323–8331.
- 41 T. W. de Haas, B. Bao, H. Acosta Ramirez, A. Abedini and D. Sinton, *Energy Fuels*, 2021, **35**, 7866–7873.
- 42 J. I. Lee, F. D. Otto and A. E. Mather, *J. Chem. Eng. Data*, 1972, **17**, 465–468.
- 43 A. M. Bhairi, Experimental equilibrium between acid gases and ethanalamine solutions, *PhD*, Oklahoma State University, 1984.
- 44 J. I. Lee, F. D. Otto and A. E. Mather, *J. Appl. Chem. Biotechnol.*, 1976, **26**, 541–549.
- 45 W. Chen, A. J. Haslam, A. Macey, U. V. Shah and C. Brechtelsbauer, *J. Chem. Educ.*, 2016, **93**, 920–926.

