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# Exploring the influence of H-bonding and ligand constraints on thiolate ligated non-heme iron mediated dioxygen activation†

Maike N. Lundahl,<sup>a</sup> Maria B. Greiner,<sup>a</sup> Marc C. Piquette,<sup>b</sup> Paige M. Gannon,<sup>a</sup> Werner Kaminsky<sup>‡</sup> and Julie A. Kovacs<sup>‡\*</sup>

Converting triplet dioxygen into a powerful oxidant is fundamentally important to life. The study reported herein quantitatively examines the formation of a well-characterized, reactive, O<sub>2</sub>-derived thiolate ligated Fe<sup>III</sup>-superoxo using low-temperature stopped-flow kinetics. Comparison of the kinetic barriers to the formation of this species via two routes, involving either the addition of (a) O<sub>2</sub> to [Fe<sup>II</sup>(S<sub>2</sub><sup>Me2</sup>N<sub>3</sub>(Pr,Pr))] (1) or (b) superoxide to [Fe<sup>III</sup>(S<sub>2</sub><sup>Me2</sup>N<sub>3</sub>(Pr,Pr))] (3) is shown to provide insight into the mechanism of O<sub>2</sub> activation. Route (b) was shown to be significantly slower, and the kinetic barrier 14.9 kJ mol<sup>-1</sup> higher than route (a), implying that dioxygen activation involves inner-sphere, as opposed to outer sphere, electron transfer from Fe(II). H-bond donors and ligand constraints are shown to dramatically influence O<sub>2</sub> binding kinetics and reversibility. Dioxygen binds irreversibly to [Fe<sup>II</sup>(S<sub>2</sub><sup>Me2</sup>N<sub>3</sub>(Pr,Pr))] (1) in tetrahydrofuran, but reversibly in methanol. Hydrogen bonding decreases the ability of the thiolate sulfur to stabilize the transition state and the Fe<sup>III</sup>-superoxo, as shown by the 10 kJ mol<sup>-1</sup> increase in the kinetic barrier to O<sub>2</sub> binding in methanol vs. tetrahydrofuran. Dioxygen release from [Fe<sup>III</sup>(S<sub>2</sub><sup>Me2</sup>N<sub>3</sub>(Pr,Pr)) O<sub>2</sub>] (2) is shown to be 24 kJ mol<sup>-1</sup> higher relative to previously reported [Fe<sup>III</sup>(S<sup>Me2</sup>N<sub>4</sub>(tren))(O<sub>2</sub>)]<sup>+</sup> (5), the latter of which contains a more flexible ligand. These kinetic results afford an experimentally determined reaction coordinate that illustrates the influence of H-bonding and ligand constraints on the kinetic barrier to dioxygen activation an essential step in biosynthetic pathways critical to life.

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## Introduction

Understanding dioxygen binding and activation by Fe centers in metalloenzymes and synthetic complexes can provide insights into how to perform challenging oxidative reactions with abundant and environmentally friendly components.<sup>1–4</sup> Foundational studies evaluating nitric oxide and O<sub>2</sub> binding to non-heme Fe in enzymes found that the charge donation by the thiolate (RS<sup>-</sup>), and a highly covalent Fe(III)–SR bond<sup>5</sup> produces an energetically favorable Fe<sup>III</sup>-superoxo species, thereby driving the formation of an Fe(III)–O<sub>2</sub><sup>•-</sup>.<sup>6</sup> These findings strongly suggest that the incorporation of a thiolate (RS<sup>-</sup>) in the coordination sphere would likely lower the activation barrier to O<sub>2</sub> binding. Understanding how thiolates provide favorable conditions for O<sub>2</sub> binding should provide important insight

into enzyme reaction mechanisms and bio-inspired catalyst design especially since thiolates have been experimentally shown to increase the reactivity of iron-peroxo and -oxo intermediates.<sup>7–10</sup>

Studying O<sub>2</sub> binding is challenging given the complex nature of the initial interaction between triplet O<sub>2</sub> and the high-spin (S = 2) Fe<sup>II</sup> center as well as the transient nature of the electronically ambiguous intermediate, which could either be described as an Fe<sup>II</sup>–O<sub>2</sub> adduct or an Fe<sup>III</sup>-superoxo species. The mechanism of its formation could involve either an outer or inner sphere electron transfer and requires changes to the spin multiplicity and geometry to accommodate the O<sub>2</sub> molecule. Direct kinetic evidence for this reaction in thiolate-ligated non-heme iron metalloenzymes such as cysteine dioxygenase (CDO) and isopenicillin N synthase (IPNS), is lacking. In part, this is because the O<sub>2</sub> binding event occurs too quickly for a precise determination of the activation barrier.<sup>11,12</sup> Very few model systems have looked at the mechanistic details of O<sub>2</sub> binding to non-heme Fe.<sup>13–15</sup> One example comes from Lacy and coworkers who carried out the first kinetic study evaluating the inner vs. outer sphere reduction of O<sub>2</sub> by a tunable model system lacking a thiolate ligand, finding that the system preferred an outer-sphere mechanism.<sup>13</sup> We recently reported a five-coordinate thiolate-ligated ferrous complex, [Fe<sup>II</sup>(S<sup>Me2</sup>N<sub>4</sub>(tren))] (4,

<sup>a</sup>Department of Chemistry, University of Washington, Campus Box 351700, Seattle, WA 98195, USA. E-mail: kovacs@uw.edu<sup>b</sup>Department of Chemistry, Tufts University, 62 Talbot Avenue, Medford, Massachusetts 02155, USA† Electronic supplementary information (ESI) available: Full derivation of the rate expression and explanation. CCDC 2212311. For ESI and crystallographic data in CIF or other electronic format see DOI: <https://doi.org/10.1039/d4sc02787f>

‡ University of Washington staff crystallographer





**Scheme 1** Pathways for the formation of  $[\text{Fe}^{\text{III}}(\text{S}_2\text{Me}_2\text{N}_3(\text{Pr},\text{Pr}))(\text{O}_2)]$  (2) (top), and reversible dioxygen binding to  $[\text{Fe}^{\text{II}}(\text{S}^{\text{Me}_2}\text{N}_4(\text{tren}))]^+$  (4) to form a putative superoxo intermediate  $[\text{Fe}^{\text{III}}(\text{S}^{\text{Me}_2}\text{N}_4(\text{tren}))(\text{O}_2)]^+$  (5) (bottom).

(Scheme 1)), that binds O<sub>2</sub> at diffusion-controlled rates ( $k(233.15\text{ K}) = 5.82(3) \times 10^7\text{ M}^{-1}\text{ s}^{-1}$ ) to afford a fleeting superoxo intermediate  $[\text{Fe}^{\text{III}}(\text{S}^{\text{Me}_2}\text{N}_4(\text{tren}))(\text{O}_2)]^+$  (5) only observable ( $\lambda_{\text{max}} = 490\text{ nm}$ ) on the millisecond timescale with limiting [O<sub>2</sub>] at  $-40\text{ }^\circ\text{C}$ .<sup>14,16</sup> Its fleeting nature prevented detailed characterization of superoxo 5 (Scheme 1) or a more thorough analysis of the factors involved in the low barrier to its formation.

Deciphering how the coordination environment, thiolate ligand, solvent and even counterions influence the reaction mechanism of dioxygen binding is best carried out with systems where the initial Fe(III)-O<sub>2</sub><sup>•-</sup> intermediate is well-characterized. However, isolated Fe(III)-O<sub>2</sub><sup>•-</sup> model compounds are rare<sup>17-21</sup> and only two well-characterized thiolate-ligated RS-Fe(III)-O<sub>2</sub><sup>•-</sup> compounds have been reported.<sup>17,22</sup> This includes our reported five-coordinate, alkyl thiolate-ligated Fe(II) complex,  $[\text{Fe}^{\text{II}}(\text{S}_2\text{Me}_2\text{N}_3(\text{Pr},\text{Pr}))]$  (1) that reacts with O<sub>2</sub> (Scheme 1) to form an observable metastable ferric superoxo species,  $[\text{Fe}^{\text{III}}(\text{S}_2\text{Me}_2\text{N}_3(\text{Pr},\text{Pr}))(\text{O}_2)]$  (2).<sup>22</sup> Vibrational (resonance Raman) data, computational studies, electron paramagnetic resonance, and <sup>1</sup>H NMR spectroscopic data established that the electronic structure of 2 consists of a low-spin ( $S = \frac{1}{2}$ ) Fe<sup>III</sup> ion strongly coupled antiferromagnetically ( $J^{\text{calc}} = -450\text{ cm}^{-1}$ ) to an  $S = \frac{1}{2}$  superoxo radical,<sup>22</sup> indicating that electron transfer takes place during its formation (*i.e.*,  $\text{Fe}^{\text{II}}(1) + \text{O}_2 \rightarrow \text{Fe}^{\text{III}}\text{-O}_2^{\bullet-}(2)$ ).<sup>22</sup>

The study herein explores the mechanism of formation of our well-characterized iron-superoxo compound,  $[\text{Fe}^{\text{III}}(\text{S}_2\text{Me}_2\text{N}_3(\text{Pr},\text{Pr}))(\text{O}_2)]$  (2),<sup>22</sup> *via* two distinct pathways, using low temperature stopped-flow kinetics. We will experimentally demonstrate that an inner-sphere, as opposed to outer-sphere, electron transfer mechanism is involved. In addition, hydrogen-bond donors are shown to modulate the thiolate ligand's influence,<sup>23-28</sup> and ligand constraints are shown to dramatically influence the activation barrier to O<sub>2</sub> binding.

## Results and discussion

In order to quantitatively determine the barrier to, and reversibility of, O<sub>2</sub> binding to five-coordinate  $[\text{Fe}^{\text{II}}(\text{S}_2\text{Me}_2\text{N}_3(\text{Pr},\text{Pr}))]$  (1,



**Fig. 1** Time-resolved spectral changes obtained upon mixing THF solutions of five-coordinate  $[\text{Fe}^{\text{II}}(\text{S}_2\text{Me}_2\text{N}_3(\text{Pr},\text{Pr}))]$  (1, 0.25 mM) and O<sub>2</sub> (3.95 mM) at  $-40\text{ }^\circ\text{C}$ . Inset: kinetic trace ( $\lambda = 523\text{ nm}$ ) showing the formation of the Fe-O<sub>2</sub> intermediate 2. All reported concentrations are after mixing in the stopped-flow cell.

Scheme 1) and the likely mechanism of formation of the corresponding superoxo compound,  $[\text{Fe}^{\text{III}}(\text{S}_2\text{Me}_2\text{N}_3(\text{Pr},\text{Pr}))(\text{O}_2)]$  (2),<sup>22</sup> we investigate herein variable temperature stopped-flow kinetics. Time-resolved electronic absorption spectroscopy was used to monitor the reaction between 1 and O<sub>2</sub> using a TgK cryogenic stopped-flow instrument. The growth of spectral features (Fig. 1) associated with 2 were monitored in two different solvents, THF and MeOH. Kinetics were monitored at  $\lambda = 523\text{ nm}$  under pseudo first order conditions with excess O<sub>2</sub>. As illustrated in the time-resolved absorption spectrum and kinetic trace of Fig. 1, superoxo 2 forms in less than 20 s at  $-40\text{ }^\circ\text{C}$



**Fig. 2** Temperature-dependent rate constants  $k_{\text{obs}}$  for the formation of superoxo 2 in the reaction between 1 (0.25 mM) and O<sub>2</sub> in THF plotted against [O<sub>2</sub>]. The intercept of approximately 0.0 would be consistent with irreversible O<sub>2</sub> binding.





Fig. 3 Temperature-dependent rate constants  $k_{\text{obs}}$  for the formation of superoxo **2** in the reaction between **1** (0.1 mM) and  $\text{O}_2$  in MeOH, plotted against  $[\text{O}_2]$ . The non-zero intercepts would be consistent with reversible  $\text{O}_2$  binding in MeOH.

C in THF. The kinetic trace (Fig. 1, insert) can be fit to the single exponential of eqn (1) with very small residuals (Fig. S1†).

$$A_t = A_\infty - (A_\infty - A_0)e^{-k_{\text{obs}}t} \quad (1)$$

### Low temperature kinetics for the reaction between $\text{O}_2$ and **1**

Under pseudo first-order conditions with excess  $\text{O}_2$ ,  $k_{\text{obs}}$  values obtained from fits to eqn (1) (Tables S1 and S2†) were found to be independent of the concentration of **1** in both THF (Fig. S2†) and MeOH (Fig. S3†), confirming that the reaction is first-order overall with respect to **1**. This would be consistent with the formation of a 1 : 1 dioxygen adduct. Observed rate constants,  $k_{\text{obs}}$ , were found to increase linearly with increasing  $\text{O}_2$  concentration in both THF (Fig. 2) and MeOH (Fig. 3) consistent with first-order dependence on  $\text{O}_2$  ( $k_{\text{obs}} = k_{\text{on}}[\text{O}_2] + k_{\text{off}}$ , eqn

(S10)†). Second order rate constants (Table 1),  $k_{\text{on}}$ , were obtained from the slope of the  $k_{\text{obs}}$  versus  $[\text{O}_2]$  plot (Fig. 2 and 3) over the temperature range  $-40$  °C to  $0$  °C in THF, and  $-30$  °C to  $0$  °C in MeOH, in  $5$  °C increments. In MeOH,  $\text{O}_2$  binds to **1**, and is released from **2**, too slowly to collect data at temperatures below  $-30$  °C using a stopped-flow instrument. In THF, the intercept of these plots was found to be approximately zero over the entire temperature range examined (Fig. 2), indicating that  $\text{O}_2$  binds irreversibly to **1** (eqn (S10)†) in this solvent. This is supported by the fact that no trend in  $k_{\text{off}}$  vs. temperature is observed in THF (Fig. S4†). In MeOH, the non-zero intercept (Fig. 3) was found to increase with increasing temperature (Fig. S5†) consistent with reversible  $\text{O}_2$  binding in this solvent. Dioxygen dissociation rate constants in MeOH,  $k_{\text{off}}$ , obtained from the intercept of the  $k_{\text{obs}}$  versus  $[\text{O}_2]$  plot of Fig. 3, are assembled in Table 1.

### Kinetic barrier to $\text{O}_2$ binding

Comparison of the second order rate constants,  $k_{\text{on}}$ , in THF versus MeOH (Table 1) indicates that  $\text{O}_2$  binds roughly two orders of magnitude more slowly to **1** in MeOH. Activation parameters for  $\text{O}_2$  binding were obtained from Eyring (Fig. 4) and Arrhenius plots (Fig. S6 and S7†) and are assembled in Table 1. The negative entropy of activation ( $\Delta S^\ddagger = -127(4)$  J mol $^{-1}$  K $^{-1}$  in THF;  $\Delta S^\ddagger = -123(7)$  J mol $^{-1}$  K $^{-1}$  in MeOH) is consistent with an associative mechanism, as opposed to one that involves initial solvent dissociation, consistent with a five-coordinate structure in both solvents. If, however, the entropy change associated with solvent release is less than that of  $\text{O}_2$  binding, then it would be possible that solvent release is involved. However, we have no evidence for the formation of six-coordinate ferrous derivatives **1-solv** (solv = coordinated solvent molecule) in any solvent, despite crystallization from coordinating solvents MeCN or MeOH, which reproducibly affords a five-coordinate complex.

Table 1 Temperature-dependent rate constants for irreversible  $\text{O}_2$  binding to  $[\text{Fe}^{\text{II}}(\text{S}_2^{\text{Me}_2}\text{N}_3(\text{Pr},\text{Pr}))]$  (**1**) in THF, reversible  $\text{O}_2$  binding to **1** in MeOH and  $\text{KO}_2$  binding to  $[\text{Fe}^{\text{III}}(\text{S}_2^{\text{Me}_2}\text{N}_3(\text{Pr},\text{Pr}))]^+$  (**3**) under constant ionic strength (0.01 M) maintained via the addition of  $\text{Bu}_4\text{NPF}_6$

Temperature (°C)	$k_{\text{on}}$ ( $\text{M}^{-1} \text{s}^{-1}$ ) in THF	$k_{\text{on}}$ ( $\text{M}^{-1} \text{s}^{-1}$ ) in MeOH	$k_{\text{off}}$ ( $\text{s}^{-1}$ ) in MeOH	$k_{\text{superoxide}}$ ( $\text{M}^{-1} \text{s}^{-1}$ ) in THF
0	$2.66(28) \times 10^2$	4.4(5)	$4(2) \times 10^{-3}$	N/A
-5	$2.31(19) \times 10^2$	3.4(2)	$3.4(8) \times 10^{-3}$	N/A
-10	$1.86(3) \times 10^2$	2.3(1)	$2.5(4) \times 10^{-3}$	N/A
-15	$1.59(16) \times 10^2$	1.7(1)	$2.4(3) \times 10^{-3}$	N/A
-20	$1.23(13) \times 10^2$	1.2(1)	$1.9(4) \times 10^{-3}$	$2.98(11) \times 10^2$
-25	$1.17(6) \times 10^2$	1.0(1)	$1.0(4) \times 10^{-3}$	$2.35(24) \times 10^2$
-30	$8.2(8) \times 10^1$	0.79(6)	$6(2) \times 10^{-4}$	$1.42(17) \times 10^2$
-35	$7.28(4) \times 10^1$	N/A	N/A	$9.5(1.2) \times 10^1$
-40	$5.02(5) \times 10^1$	N/A	N/A	$5.8(3) \times 10^1$
-45	N/A	N/A	N/A	$4.6(7) \times 10^1$
-50	N/A	N/A	N/A	$2.9(2) \times 10^1$
-55	N/A	N/A	N/A	$2.0(2) \times 10^1$
-60	N/A	N/A	N/A	$1.16(5) \times 10^1$
$\Delta H^\ddagger$ (kJ mol $^{-1}$ )	19.3(8)	30(2)	32(4)	34.3(9)
$\Delta S^\ddagger$ (J mol $^{-1}$ K $^{-1}$ )	-127(4)	-123(7)	-170(20)	-60(4)
$E_a$ (kJ mol $^{-1}$ )	21.4(8)	32(2)	34(4)	36.3(9)
$\Delta G^\ddagger$ (243 K, kJ mol $^{-1}$ )	50(1)	60(3)	73(4)	N/A





**Fig. 4** Eyring plots from which the activation parameters reported were obtained for the following reactions. (A)  $\text{O}_2$  binding to **1** in THF and (B)  $\text{O}_2$  binding to **1** in MeOH, which use the second order rate constants,  $k_{\text{onr}}$ , obtained from the slope of  $k_{\text{obs}}$  vs.  $[\text{O}_2]$  plots (Fig. 2 and 3) with  $[\text{Fe}^{\text{II}}] = 0.25$  mM and  $0.1$  mM respectively. (C) The reaction of **3** and  $\text{KO}_2$  (solubilized with KryptoFix) in THF plotted with second order rate constants,  $k_{\text{onr}}$ , obtained from the slope of  $k_{\text{obs}}$  vs.  $[\text{KO}_2]$  plots.  $[\text{Fe}^{\text{III}}] = 0.1$  mM, after mixing. (D) The release of  $\text{O}_2$  from **2** in MeOH which uses first order rate constants,  $k_{\text{off}}$ , obtained from the intercept of  $k_{\text{obs}}$  versus  $[\text{O}_2]$  plots (Fig. 3).

### Low temperature kinetics for the reaction between $\text{KO}_2$ and **3**

As shown in Scheme 1, superoxo intermediate **2** forms both *via* the addition of  $\text{O}_2$  to  $\text{Fe}(\text{II})$ -**1** as well as *via* the addition of  $\text{KO}_2$  to  $[\text{Fe}^{\text{III}}(\text{S}_2^{\text{Me}2}\text{N}_3(\text{Pr},\text{Pr}))\text{PF}_6]$  (**3**).<sup>22</sup> Evidence for this was described



**Fig. 5** Time-resolved spectral changes observed in the reaction between **3** ( $0.1$  mM) and  $\text{KO}_2$  ( $5.0$  mM; solubilized with 222-KryptoFix) in THF at  $-40$  °C. Inset: kinetic trace ( $\lambda = 523$  nm) showing the formation of **2**. All reported concentrations are after mixing in the stopped-flow cell.

in a recent publication,<sup>22</sup> where we showed that an electronic absorption spectrum identical to that of Fig. 1 is observed in the reaction between **3** and  $\text{KO}_2$  (Fig. 5). Here we examine the variable temperature kinetics for the reaction between  $\text{KO}_2$  and  $\text{Fe}(\text{III})$ -**3**, in order to determine whether the activation barrier is lower or higher than that for  $\text{Fe}(\text{II}) + \text{O}_2$ , and thus whether an inner- or outer-sphere electron transfer mechanism is involved. Observed pseudo first order rate constants ( $k_{\text{obs}}$ ), obtained from fits to eqn (1) (Fig. S8†), assembled in Table S3,† increase linearly as  $[\text{KO}_2]$  is increased (Fig. 6) under pseudo first-order conditions with excess  $\text{KO}_2$ , and are independent of the concentration of **3** (Fig. S9†). The latter indicates a 1st-order dependence on superoxide ( $\text{O}_2^{\cdot-}$ ) was determined by varying the concentration of  $\text{KO}_2$  (Fig. 6) over the range  $1.0$  mM to  $5.0$  mM. Second-order rate constants,  $k_{\text{superoxide}}$ , were obtained from the slope of the  $k_{\text{obs}}$  vs.  $[\text{KO}_2]$  plot of Fig. 6 and are assembled in Table 1. Activation parameters for  $\text{O}_2^{\cdot-}$  binding to **3** in THF ( $E_a = 36.3(9)$  kJ mol<sup>-1</sup>; Table 1) under constant ionic strength conditions ( $0.01$  M; maintained with  $\text{Bu}_4\text{NPF}_6$ ) were obtained from Eyring (Fig. 4) and Arrhenius (Fig. S10†) plots. The Arrhenius parameter,  $E_a$ , for  $\text{KO}_2$  binding to **3** (Table 1) is  $14.9$  kJ mol<sup>-1</sup> greater than the barrier ( $E_a = 21.4(8)$  kJ mol<sup>-1</sup>) to  $\text{O}_2$  binding to **1** (Scheme 1 and Table 1) in the same solvent. This has implications about the mechanism of the reaction between **1** and  $\text{O}_2$  (*vide infra*). The negative entropy of activation,  $\Delta S^\ddagger$ , (Table 1) is consistent with an associative process involving  $\text{KO}_2$





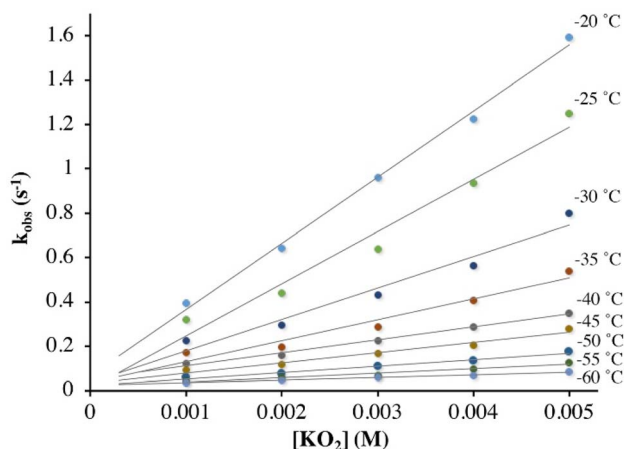


Fig. 6 Temperature-dependent rate constants,  $k_{\text{obs}}$ , for the formation of superoxo **2** in the reaction between **3** and  $\text{KO}_2$  (solubilized with KryptoFix) in THF, plotted against  $[\text{KO}_2]$ . The  $\sim 0$  intercepts would be consistent with irreversible  $\text{O}_2^{\cdot-}$  binding.  $[\text{Fe}(\text{III})] = 0.1 \text{ mM}$ . Concentrations listed correspond to after mixing in the stopped-flow cell.

binding to **3**. The smaller magnitude of  $\Delta S^\ddagger$  for the  $\text{KO}_2$  reaction relative to the  $\text{O}_2$  reaction (Table 1) likely reflects the smaller ligand reorganization required to convert  $\text{Fe}(\text{III})$ -**3** to **2** relative to that required to convert  $\text{Fe}(\text{II})$ -**1** to **2** (*vide infra*).

Possible mechanisms (Fig. 7) for the formation of **2** include (a) outer-sphere electron transfer (ET) from  $\text{Fe}(\text{II})$  to  $\text{O}_2$  to afford oxidized  $\text{Fe}(\text{III})$ -**3** and superoxide ( $\text{O}_2^{\cdot-}$ ), followed by  $\text{O}_2^{\cdot-}$  binding to **3** to afford superoxo **2**, (b) dioxygen binding to **1** to afford an  $\text{Fe}(\text{II})$ - $\text{O}_2$  dioxygen intermediate followed by inner-sphere electron transfer (ET) to afford superoxo **2**, or, (c) a concerted mechanism across the diagonal. The latter two are indistinguishable. The fact that the activation barrier ( $E_a$ ) to  $\text{O}_2^{\cdot-}$  binding to **3** was experimentally determined to be  $14.9 \text{ kJ mol}^{-1}$  higher than  $E_a$  for dioxygen binding to **1** (Table 1) implies that mechanism (b) or (c), involving inner sphere ET, are the most likely mechanisms. The unfavorable

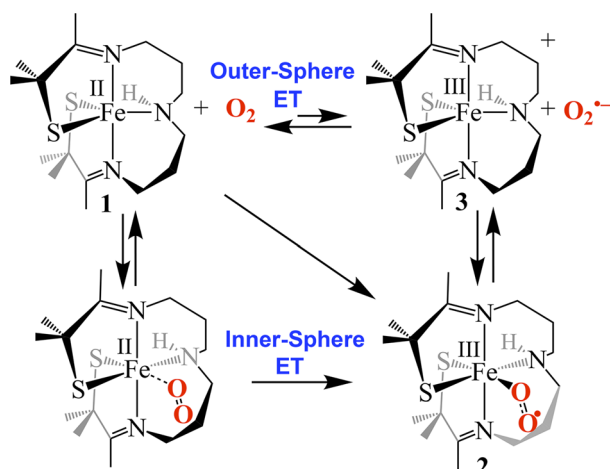


Fig. 7 Possible mechanisms for the reaction between **1** and  $\text{O}_2$  involving either outer-sphere electron transfer (ET) followed by superoxide ( $\text{O}_2^{\cdot-}$ ) binding to oxidized **3**,  $\text{O}_2$  binding followed by inner-sphere ET, or a concerted mechanism (diagonal).

thermodynamics of electron transfer from **1** to  $\text{O}_2$  ( $K_{\text{eq}} = 4.66 \times 10^{-12} \text{ M}^{-1}$  calculated using  $E_{1/2}(\text{Fe}(\text{III})\text{-3}/\text{Fe}(\text{II})\text{-1}) = -0.40 \text{ V}$  and  $E_{1/2}(\text{O}_2/\text{O}_2^{\cdot-}) = -1.07 \text{ V vs. SCE}$  in THF),<sup>29</sup> provides additional support for this conclusion.

### Kinetic barrier for $\text{O}_2$ release from **2**

In MeOH,  $\text{O}_2$  binds reversibly to  $[\text{Fe}^{\text{II}}(\text{S}_2^{\text{Me}_2}\text{N}_3(\text{Pr},\text{Pr}))]$  (**1**), in contrast to the irreversible binding in THF. Activation parameters for  $\text{O}_2$  release from  $[\text{Fe}^{\text{III}}(\text{S}_2^{\text{Me}_2}\text{N}_3(\text{Pr},\text{Pr}))(\text{O}_2)]$  (**2**) in MeOH ( $\Delta H^\ddagger = 32(4) \text{ kJ mol}^{-1}$ ,  $\Delta S^\ddagger = -170(20) \text{ J mol}^{-1} \text{ K}^{-1}$ ,  $E_a = 34(4) \text{ kJ mol}^{-1}$ ) were obtained from Eyring (Fig. 4) and Arrhenius plots (Fig. S11†) and are assembled in Table 1. The negative entropy of activation for  $\text{O}_2$  release is unexpected given its dissociative nature. A negative entropy has also been observed for  $\text{O}_2$  release from thiolate-ligated  $[\text{Fe}^{\text{III}}(\text{S}^{\text{Me}_2}\text{N}_4(\text{tren}))(\text{O}_2)]^+$  (**4**) in MeOH as well as  $[\text{Mn}^{\text{III}}(\text{L}^{\text{Quino}})(\text{O}_2)]^+$ .<sup>14,30</sup> These complexes both contain flexible ligands for which bond rearrangement during the oxidation state change (*vide infra*) is proposed to contribute to a negative entropy of activation large enough to offset  $\text{O}_2$  release.<sup>14,30</sup> To understand the negative entropy of activation for  $\text{O}_2$  release from **2**, we evaluated the requisite spin-state and oxidation state changes that require changes to the  $\text{Fe-L}$  bond lengths. Because **1** is high-spin  $S = 2$  ( $\mu_{\text{eff}} = 4.69 \text{ BM}$ ), whereas **2** contains a low-spin  $S = \frac{1}{2}$   $\text{Fe}(\text{III})$  ion, a spin-state change must take place during  $\text{O}_2$  release. To prepare for this electron transfer step a more ordered state involving adjusted  $\text{Fe-L}$  bond lengths is therefore required. In addition to the spin-state change, oxidation state-dependent H-bonding between MeOH and the thiolate sulfurs of **1** relative to **2** (*vide infra*) influences the entropy changes involved in  $\text{O}_2$  release. Both factors are examined in greater detail in the context of crystallographically determined structures, electronic absorption spectra and DFT calculations, below.

### Influence of H-bonds on kinetic barriers

An observed blue-shift (by  $2160 \text{ cm}^{-1}$ ) of the dominant feature in the electronic absorption spectrum from  $420 \text{ nm}$  (23

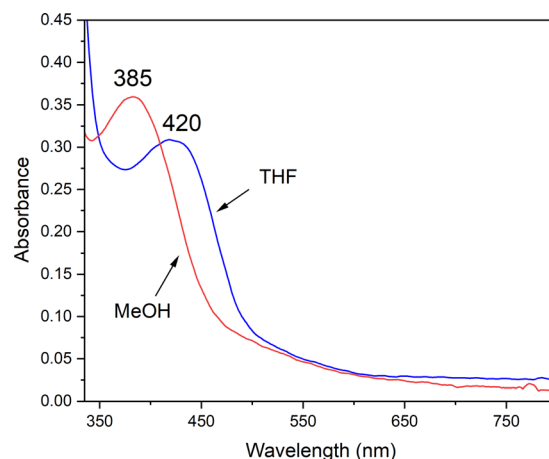


Fig. 8 H-bonding to MeOH causes the  $\text{RS} \rightarrow \text{Fe}$  band of reduced **1** ( $0.238 \text{ mM}$ ) to blue-shift relative to its energy in THF.



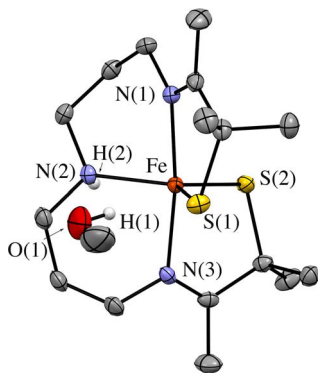


Fig. 9 ORTEP diagram of **1** crystallized from MeOH showing the MeOH that is H-bonded to one of the thiolate sulfurs, S(1). Shown with 50% probability ellipsoids and numbering scheme. All hydrogen atoms excluding H(2) and H(1) have been deleted for clarity.

800  $\text{cm}^{-1}$ ) in THF to 385  $\text{nm}$  (26 000  $\text{cm}^{-1}$ ) in MeOH provides experimental evidence for the influence of either the dielectric constant ((THF) = 7 vs.  $\epsilon(\text{MeOH}) = 33$ ), or H-bonding on the orbitals involved in the transition (Fig. 8). The crystallographically determined structure of **1** (Fig. 9 and Tables S5–S9<sup>†</sup>), contains a MeOH nestled in the  $\text{O}_2$  binding pocket and the MeOH proton H(1), points towards S(1) with an H(1)⋯S(1) distance of 2.318 Å, providing evidence for H-bonding in MeOH. The  $\text{O}_2$  binding site was deduced from previous structures that show exogenous ligands such as  $\text{N}_3^-$  and NO bind to Fe in the pocket that lies opposite to the secondary amine proton,<sup>31,32</sup> H(2), in the more open N(2)–Fe–S(1) angle. Therefore at a minimum there would be release of the MeOH that is in proximity to the binding site upon  $\text{O}_2$  binding which would contribute a favorable positive  $\Delta S^\ddagger(\text{solvent release})$  term that slightly offsets the unfavorable negative  $\Delta S^\ddagger(\text{O}_2 \text{ binding})$  term. Conversely,  $\text{O}_2$  release from **2** would involve the addition of at least one H-bonded MeOH to **1** (Fig. 9) offsetting the favorable entropy of activation associated with  $\text{O}_2$  release.

To evaluate the properties likely to affect H-bonding in **1** relative to **2** we pursued density functional theory (DFT) calculations. The DFT calculated structure of **1** (Fig. S12<sup>†</sup>), shows that the thiolate sulfurs of reduced **1**-Fe(II) have an average Mulliken charge of  $-0.670$  (Table S4<sup>†</sup>), whereas the thiolate sulfurs of oxidized superoxo **2** have Mulliken charges of  $-0.519$  and  $-0.482$ , for the *trans*  $\text{RS}^-$  and *cis*  $\text{RS}^-$ , respectively (Table S4<sup>†</sup>). Hydrogen-bonding to the thiolates would therefore be less likely, and/or involve fewer MeOH molecules, with oxidized **2**, relative to **1**. The electronic absorption spectra of **2** in THF vs. MeOH shows a distinct, but less amplified blue-shift relative to **1** ( $\Delta E = 721 \text{ cm}^{-1}$  for **2** (Fig. S13<sup>†</sup>) vs.  $\Delta E = 2114 \text{ cm}^{-1}$  for **1**).

In the absence of a more polar H-bonding solvent, the barrier to  $\text{O}_2$  binding was experimentally determined to be lower (Fig. 10). The blue-shift in the electronic absorption spectrum (Fig. 8) and insights from DFT calculations indicate this is in part caused by a stabilization of the Fe(II)–SR molecular orbitals involved in the  $\text{RS}^- \rightarrow \text{Fe}$  charge transfer transition in protic MeOH solvent (Fig. S14 and S15<sup>†</sup>).<sup>33</sup> This has a noticeable effect on the kinetic barrier,  $\Delta G^\ddagger$ , to  $\text{O}_2$  binding and release. If solvent polarity alone were responsible, then one would expect the

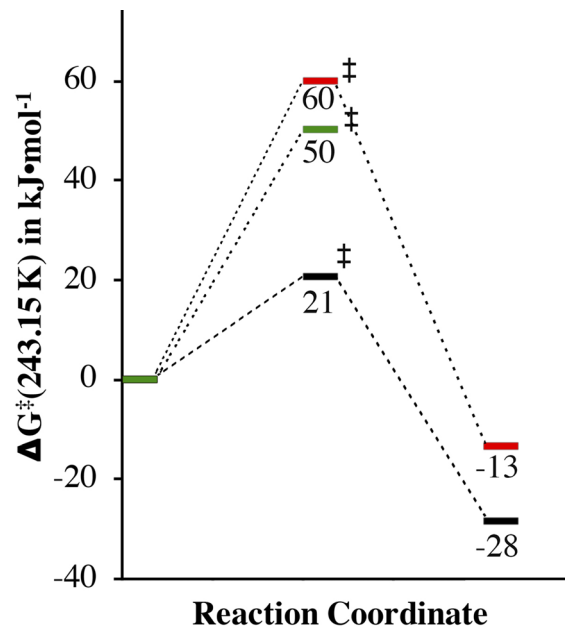


Fig. 10 Comparison of the experimentally determined barrier to  $\text{O}_2$  binding to **1** in THF (green) and MeOH (red) and **4** in MeOH (black), as well as the release of  $\text{O}_2$  from **2** and **5** in MeOH.

kinetic barrier to be lower in MeOH, as opposed to higher as is experimentally observed. This is because charge separation increases as neutral **1** and  $\text{O}_2$  convert to  $2\text{-O}_2^{\cdot-}$ .

An explanation as to why H-bonding to thiolate sulfurs would influence the barrier to  $\text{O}_2$  binding is that it would tie up lone pairs that would otherwise be involved in  $\pi$ -bonding to the metal ion. Thiolates have been shown by our group to form highly covalent Fe(III)–SR bonds<sup>5</sup> with partial Fe=S double bond character. This stabilizes oxidized products such as **2**,<sup>14,22</sup> and any transition states with developing Fe(III)–SR character. If, on the other hand, a lone pair on the thiolate sulfur is tied up in H-bonding to a protic solvent (*e.g.*, MeOH),<sup>33</sup> then the incipient Fe(III)–SR bond of the transition-state would have less double bond character, and would thus be less stable. This, in addition, to the stabilization of the Fe(II) precursor provided by H-bonding would result in slower  $\text{O}_2$  binding to **1** in MeOH, as is experimentally observed (Table 1). The experimentally measured free energy of activation,  $\Delta G^\ddagger$ , for dioxygen binding to **1** in MeOH (60(3)  $\text{kJ mol}^{-1}$ ) is 10  $\text{kJ mol}^{-1}$  higher than in THF (50(1)  $\text{kJ mol}^{-1}$ ) at 243.15 K (Fig. 10 and Table 1), consistent with this.

#### Influence of ligand constraints on kinetic barriers

Comparison of the free energy of activation,  $\Delta G^\ddagger$  (243.15 K), for  $\text{O}_2$  release from **2** (73(4)  $\text{kJ mol}^{-1}$ ) in MeOH at 243.15 K (Table 1) relative to that for previously reported  $[\text{Fe}^{\text{III}}(\text{S}^{\text{Me}_2}\text{N}_4(\text{tren}))(\text{O}_2)]^+$  (**5**, Scheme 1, 49(7)  $\text{kJ mol}^{-1}$ )<sup>14</sup> under the same conditions, shows that the barrier to  $\text{O}_2$  release is 24  $\text{kJ mol}^{-1}$  higher for **2**. This likely reflects ligand constraints<sup>31,34,35</sup> and the ligand rearrangement required to form the transition-state necessary for  $\text{O}_2$  release. The multidentate ligand of **1** and **2** consists of



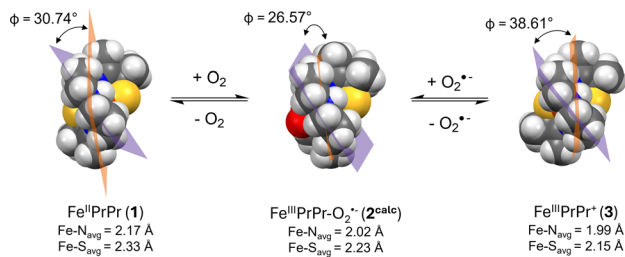


Fig. 11 Space-filling models generated from the crystallographically-determined structure of **1** (left) and **3** (left), and the DFT-optimized geometry of **2<sup>calc</sup>** (middle) displaying differences in the helical wrapping angle,  $\Phi$ , and the larger amount of structural rearrangement required for  $O_2^{\cdot-}$  binding to **3** than  $O_2$  binding to **1**. The least squared planes of N(1)FeN(2)N(3) and C(6)N(2)C(7)Fe shown in orange and purple respectively.

a single chain that wraps around the metal ion in a helical fashion (Fig. 11).<sup>35</sup> Even minor angle changes at the metal ion in preparation for  $O_2$  release translate into significant movement of the thiolate containing arms that terminate the single chain ligand backbone (Fig. 11). An experimental demonstration of this can be seen when comparing the crystallographically determined structures of Fe(II) structures **4** versus **1**, relative to the corresponding Fe(III)- $N_3$  structures (Fig. 12).<sup>22,31,36,37</sup> The geometry of each 5-coordinate Fe(II) structure, **4** and **1** (Scheme 1) is best described as distorted trigonal-bipyramidal  $\tau = 0.63$  and 0.79, respectively. Structure **4** (Scheme 1) is closer to square pyramidal, and thus requires less geometric rearrangement upon  $N_3^-$  binding (Fig. 12, left). In contrast, as the entire S(R) arm of **1** swings away from the binding site (Fig. 12, right), the thiolate is pushed 1.8 Å opening the binding site by 46°. This reorganizational barrier would be mostly entropic in nature. The entropy of activation (Table 1) for  $O_2$  binding to **1** in MeOH ( $\Delta S^\ddagger = -123(7) \text{ J mol}^{-1} \text{ K}^{-1}$ ), is less favorable (by  $178 \text{ J mol}^{-1} \text{ K}^{-1}$ ) than for  $O_2$  binding to **4** ( $+55(68) \text{ J mol}^{-1} \text{ K}^{-1}$ ).<sup>14</sup> This factor dominates and is partially responsible for the large differences in  $\Delta G^\ddagger$  (Fig. 10). The lower barrier for  $O_2$  release from **5** relative to **2** (Scheme 1), likely reflects its less constraining tripodal

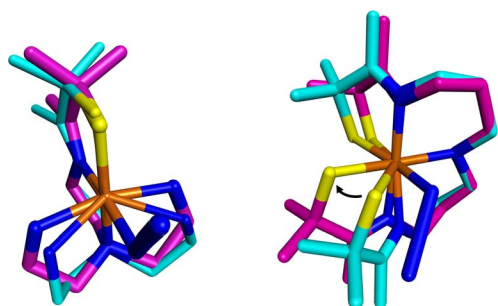


Fig. 12 Visualization of experimentally obtained structural changes that occur upon oxidation and binding of a sixth ligand, azide. Left:  $[Fe^{II}(S^{Me_2}N_4(tren))]^+$  (**4**, cyan)<sup>36</sup> to  $[Fe^{III}(S^{Me_2}N_4(tren))N_3]$  (magenta).<sup>37</sup> Right:  $[Fe^{II}(S_2^{Me_2}N_3(Pr,Pr))]$  (**1**, cyan)<sup>22</sup> and  $[Fe^{III}(S_2^{Me_2}N_3(Pr,Pr))(N_3)]$  (magenta).<sup>31</sup> Depiction utilized previously reported crystallographically determined structures.

ligand with its three independent arms (Fig. 12). The less constraining ligand of **4** also results in a lower free energy of activation for dioxygen binding to **4** ( $\Delta G^\ddagger$  (243.15 K) =  $21(2) \text{ kJ mol}^{-1}$ ) relative to **1** ( $\Delta G^\ddagger$  (243.15 K) =  $60(3) \text{ kJ mol}^{-1}$ ) in the same solvent (MeOH).<sup>14</sup>

### Understanding why the kinetic barrier to $O_2^{\cdot-}$ binding is higher than the kinetic barrier to $O_2$ binding

Given that oxidized  $[Fe^{III}(S_2^{Me_2}N_3(Pr,Pr))]^+$  (**3**) is cationic, one might expect electrostatic attraction to lower the barrier to  $O_2^{\cdot-}$  binding to **3**, relative to  $O_2$  binding to neutral **1**. This likely would be the case, if it weren't for the oxidation state-dependent ligand constraints imposed by the single chain helical ligand.<sup>31,35</sup> With five-coordinate **3**, the helical twist is "tighter" relative to five-coordinate **1** (Fig. 11) due to the shorter Fe(III)-L bond lengths. The bond lengths of **3** expand upon binding a sixth ligand, e.g.  $O_2^{\cdot-}$ .<sup>31</sup> This difference can be quantified by comparing the helical twist angle,  $\Phi$ , of crystallographically characterized **1**, **3** and the DFT geometry optimized structure of **2** (Fig. 11).<sup>35</sup> The angle  $\Phi$  is defined as the angle separating the least squared plane containing the Fe center, the two carbons flanking the equatorial nitrogen, N(2), and the equatorial nitrogen (C(6)N(2)C(7)Fe) from the mean plane containing N(1)FeN(2)N(3). One can see in Fig. 11 that the twist angle decreases (from  $30.74^\circ$  to  $26.57^\circ$ ) when  $O_2$  binds to Fe(II)-**1** to form to Fe(III)- $O_2^{\cdot-}$  **2**, whereas a larger decrease (from  $38.61^\circ$  to  $26.57^\circ$ ) is required when  $O_2^{\cdot-}$  binds to Fe(III)-**3**. This helps to explain the larger barrier to superoxide binding to **3**, relative to  $O_2$  binding to **1**.

### Relevance to metalloenzymes and transition-metal $O_2$ chemistry

The contribution of ligand constraints and H-bond donors on the reactivity reported herein is reminiscent of the influence of protein constraints and H-bond donors on metalloenzymes. Protein constraints have been shown to have a dramatic effect on electronic structure and reactivity.<sup>38-42</sup> For example, the redox potential and electron transfer kinetics blue copper proteins is controlled by protein constraints.<sup>43</sup> Hydrogen-bond donors can also influence the stability of metastable enzyme intermediates and influence reaction pathways.<sup>44-47</sup>

If we compare the activation barriers and rate constants associated with  $O_2$  binding to the Fe(II) complex, **1**, reported herein, with those reported in the literature for Fe(II), both heme and non-heme, Co(II), Mn(II), and Cu(I) complexes,<sup>30,48-58</sup> (Table 2), we see that rates are faster for porphyrin-ligated systems,<sup>59,60</sup> both synthetic and biological.<sup>61-65</sup> This can be attributed to the smaller rearrangement required for ligands to bind to the more readily accessible open face of a porphyrin-ligated metal ion—a property that would be important to facilitate  $O_2$  release in  $O_2$  transport proteins.

Irreversible  $O_2$  binding, such as that observed with **1** in THF, is important if the enzyme function involves the metal-superoxo species, as is the case with isopenicillin N-synthase (IPNS)<sup>11</sup> and cysteine dioxygenase (CDO).<sup>12</sup> With thiolate-ligated systems like **1**, and IPNS, one can see from this study (Table 2) that the



**Table 2** Comparison of kinetic parameters for dioxygen binding to mononuclear Fe(II), Mn(II), Co(II), and Cu(II) model complexes<sup>g</sup> and biological active sites

	Solvent (T)	$k_{\text{on}}$ ( $\text{M}^{-1} \text{s}^{-1}$ )	$\Delta H^\ddagger$ ( $\text{kJ mol}^{-1}$ )	$\Delta S^\ddagger$ ( $\text{J mol}^{-1} \text{K}^{-1}$ )	Ref.
3-, 4-, and 5-Coordinate complexes					
[Fe <sup>II</sup> (S <sub>2</sub> <sup>Me2</sup> N <sub>3</sub> (Pr,Pr))] (1)	THF (−40 °C)	$5.02(5) \times 10^1$	19.3(8)	−127(4)	This work
	THF (0 °C)	$2.66(28) \times 10^2$			This work
	MeOH (−30 °C)	0.79(6)	30(2)	−123(7)	This work
	MeOH (0 °C)	4.4(5)			This work
[Fe <sup>II</sup> (S <sup>Me2</sup> N <sub>4</sub> (tren))] (4)	MeOH (−40 °C)	$5.82(3) \times 10^7$	34(15)	55(68)	14
[Mn <sup>II</sup> (S <sup>Me2</sup> (6-MeDPEN))] (4)	MeCN (−40 °C)	$7.17(45) \times 10^2$	26(2)	−76(7)	30 and 48
[Cu <sup>I</sup> (TPA)(THF)] <sup>+</sup>	THF (25 °C)	$1.3 \times 10^9$	7.62	−45.1	49
[ <sup>NMe2</sup> LCu <sup>I</sup> ] <sup>a</sup>	THF (−80 °C)	$6.90(2) \times 10^7$	32.1	80.1	50
<sup>Im</sup> LCu <sup>I</sup> (MeCN) <sup>b</sup>	THF (−80 °C)	$1.80(3) \times 10^8$	23.4	35.1	50
L <sup>1</sup> Cu <sup>I</sup> (MeCN) <sup>c</sup>	THF (−50 °C) <sup>d</sup>	$1.56(2) \times 10^3$	18(2)	−100(10)	51
(Me <sub>6</sub> tren)Cu <sup>I</sup> (EtCN)	EtCN (−50 °C)	$8.7(4) \times 10^5$	17.1(6)	−52(3)	52
(TMPA)Cu <sup>I</sup> (EtCN)	EtCN (−50 °C)	$5.0(3) \times 10^5$	31.6(5)	10(3)	53
Co <sup>II</sup> (acacMeDPT)	Acetone (−50 °C)	$1.0(1) \times 10^6$	5.0	−109	54
Co <sup>II</sup> (SalMeDPT)	Acetone (−50 °C)	$2.10(4) \times 10^5$	12.6	−80	54 and 56
Co <sup>II</sup> (pXyacacMeDPT)	Acetone (−50 °C)	$1.74(5) \times 10^6$	28	−13	54
V[N <sup>+</sup> ( <sup>t</sup> Bu)Ar] <sub>3</sub>	Toluene (−53 °C)	$40(3) \times 10^3$	13.8(8)	−92(4)	55
[Pd(IPr) <sub>2</sub> ] <sup>e</sup>	Toluene (−80 °C)	$6.1(1) \times 10^3$	18(3)	−54(4)	57
[Pd(IPr)(P( <i>p</i> -tolyl) <sub>3</sub> )] <sup>e</sup>	THF (−80 °C)	$1.2 \times 10^3$	9.2	−117	58
Five coordinate porphyrin complexes					
[Fe(Piv <sub>3</sub> 5CIm)]	Toluene (25 °C)	$4.3 \times 10^8$	0.75(33)	−117(3)	59
[Fe <sup>II</sup> (Piv <sub>2</sub> C <sub>9</sub> )(RS)] <sup>1−</sup>	Toluene (20 °C)	$3.5 \times 10^6$			60
Biological systems					
Cyt bo <sub>3</sub> ubiquinol oxidase	H <sub>2</sub> O (25 °C)	$3.8 \times 10^7$			62
H-NOX <sup>f</sup>	H <sub>2</sub> O (25 °C)	$2.5(3) \times 10^7$			63
Hemoglobin (α-human)	H <sub>2</sub> O (25 °C)	$2.8 \times 10^7$			64
Fe-Mb (horse)	H <sub>2</sub> O (22 °C)	$9.8 \times 10^7$	23	−30	65

<sup>a</sup> NMe<sub>2</sub>L = DMAE of Fig. S16. <sup>b</sup> ImL = impy1 of Fig. S16. <sup>c</sup> L<sup>1</sup> = β-diketiminato (R = Me). <sup>d</sup> These values were obtained using flash photolysis methods.

<sup>e</sup> IPr = 1,3-bis(diisopropyl)phenylimidazol-2-ylidene which is an *N*-heterocyclic carbene. <sup>f</sup> H-NOX = Heme nitric oxide/oxygen binding. <sup>g</sup> See ESI, Fig. S16 for drawings of the ligands.

reversibility of O<sub>2</sub> binding can be controlled *via* the removal of H-bond donors in the vicinity of the thiolate sulfur. The influence of steric effects on activation enthalpy,  $\Delta H^\ddagger$ , has been previously observed for O<sub>2</sub> binding to a series of five-coordinate Co<sup>II</sup> complexes containing pentadentate ligands, in which the bulkiest ligand was shown to have a considerably larger (by 28 kJ mol<sup>−1</sup>)  $\Delta H^\ddagger$  value.<sup>54</sup> Complexes containing more flexible ligands with independent arms (such as TPA, TMPA, tren, Tp; Fig. S16<sup>†</sup>), or those that favor a planar geometry (such as salen, porphyrins,<sup>59</sup> or acac; Fig. S16<sup>†</sup>) bind O<sub>2</sub> more readily than **1** (Table 2).

## Conclusions

Low-temperature stopped-flow kinetics have been used to determine the most likely mechanism, and reversibility of O<sub>2</sub> binding to [Fe<sup>II</sup>(S<sub>2</sub><sup>Me2</sup>N<sub>3</sub>(Pr,Pr))] (**1**) to afford a well-characterized, reactive, thiolate-ligated ferric superoxo compound, [Fe<sup>III</sup>(S<sub>2</sub><sup>Me2</sup>N<sub>3</sub>(Pr,Pr)(O<sub>2</sub>)] (**2**).<sup>22</sup> Comparison of the kinetics of formation of **2** *via* the addition of KO<sub>2</sub> to oxidized [Fe<sup>III</sup>(S<sub>2</sub><sup>Me2</sup>N<sub>3</sub>(Pr,Pr))] (3) *versus* the addition of O<sub>2</sub> to **1** provided evidence for a mechanism involving inner-sphere electron transfer from Fe(II) to O<sub>2</sub> upon binding, as opposed to an outer-

sphere electron transfer process to afford O<sub>2</sub><sup>•−</sup> and Fe(III) followed by O<sub>2</sub><sup>•−</sup> binding to the oxidized metal ion.

H-bonding and structural constraints were shown to have a noticeable effect on the O<sub>2</sub> binding and release. The kinetic barrier to O<sub>2</sub> binding was shown to be 10 kJ mol<sup>−1</sup> higher in MeOH relative to THF, indicating that the transition-state is destabilized in the presence of H-bond donors. Thiolates have been shown by our group to form highly covalent Fe(III)–SR bonds with partial Fe=S double bond character. An explanation as to why H-bonding would influence the barrier to O<sub>2</sub> binding is that any transition states with developing Fe(III)–SR character would have less Fe=S double bond character if the thiolate sulfur lone pairs were engaged in RS⋯H–OME H-bonding. Hydrogen bonding would also, for the same reasons, destabilize the Fe(III)–superoxo intermediate in MeOH relative to THF thereby decreasing the barrier to O<sub>2</sub> release in MeOH. We cannot experimentally verify this, since irreversible binding in THF prevents us from determining the barrier to O<sub>2</sub> release in THF from an Fe(III)–superoxo that is presumably more stable.

In addition, the constraints of the single chain ligand system of **1** was shown to significantly increase the barrier to O<sub>2</sub> binding and release relative to the less constraining tripodal ligand system of **4**. From this study, H-bonding as well as ligand





flexibility are shown to have substantial impact on not only O<sub>2</sub> binding but also its release.

The experimentally determined kinetic parameters provided herein for O<sub>2</sub> binding to **1** and O<sub>2</sub> release from **2**, both of which require spin-state changes and electron transfer steps, will provide a point of reference for future in-depth computational studies of transition states for thiolate containing compounds. Further knowledge of this critical first step of O<sub>2</sub> activation has relevance to understanding enzymes important for our health as well as for the design of new environmentally friendly bio-inspired catalysts.

## Data availability

Crystallographic data for **1**·MeOH has been deposited at the Cambridge Crystallographic Data Centre (CCDC) under deposition number 2212311.†

## Author contributions

JAK raised the funding and conceptualized the project with contributions from MNL. Experimental work which included collecting and analysing the stopped flow electronic absorption spectra was carried out by MNL, MBG, and MCP. MNL and MBG performed the DFT calculations. PMG and WK collected and refined the crystallographic data. JAK and MNL wrote the manuscript.

## Conflicts of interest

There are no conflicts to declare.

## Acknowledgements

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## Notes and references

- 1 E. I. Solomon, D. E. DeWeese and J. T. Babicz Jr, Mechanisms of O<sub>2</sub> activation by mononuclear non-heme iron enzymes, *Biochemistry*, 2021, **60**, 3497–3506.
- 2 M. Gennari and C. Duboc, Bio-inspired, multifunctional metal–thiolate motif: from electron transfer to sulfur reactivity and small-molecule activation, *Acc. Chem. Res.*, 2020, **53**, 2753–2761.
- 3 D. Jeong, J. Selverstone Valentine and J. Cho, Bio-inspired mononuclear nonheme metal peroxo complexes: synthesis, structures and mechanistic studies toward understanding enzymatic reactions, *Coordin. Chem. Rev.*, 2023, **480**, 215021.
- 4 F. Nistri, M. Chino, O. Maglio, A. Bhagi-Damodaran, Y. Lu and A. Lombardi, Design and engineering of artificial oxygen-activating metalloenzymes, *Chem. Soc. Rev.*, 2016, **45**, 5020–5054.
- 5 P. Kennepohl, F. Neese, D. Schweitzer, H. L. Jackson, J. A. Kovacs and E. I. Solomon, Spectroscopy of non-heme iron thiolate complexes: insight into the electronic structure of the low-spin active site of nitrile hydratase, *Inorg. Chem.*, 2005, **44**, 1826–1836.
- 6 C. D. Brown, M. L. Neidig, M. B. Neibergall, J. D. Lipscomb and E. I. Solomon, VTVH-MCD and DFT studies of thiolate bonding to {FeNO}<sub>7</sub>/{FeO<sub>2</sub>}<sub>8</sub> complexes of isopenicillin N synthase: substrate determination of oxidase versus oxygenase activity in nonheme Fe enzymes, *J. Am. Chem. Soc.*, 2007, **129**, 7427–7438.
- 7 T. H. Yosca, J. Rittle, C. M. Krest, E. L. Onderko, A. Silakov, J. C. Calixto, R. K. Behan and M. T. Green, Iron(IV) hydroxide pK<sub>a</sub> and the role of thiolate ligation in C–H bond activation by cytochrome P450, *Science*, 2013, **342**, 825–829.
- 8 J. Rittle and M. T. Green, Cytochrome P450 compound I: capture, characterization, and C–H bond activation kinetics, *Science*, 2010, **330**, 933–937.
- 9 M. T. Green, C–H bond activation in heme proteins: the role of thiolate ligation in cytochrome P450, *Curr. Opin. Chem. Biol.*, 2009, **13**, 84–88.
- 10 C. M. Krest, E. L. Onderko, T. H. Yosca, J. C. Calixto, R. F. Karp, J. Livada, J. Rittle and M. T. Green, Reactive intermediates in cytochrome P450 catalysis, *J. Biol. Chem.*, 2013, **288**, 17074–17081.
- 11 E. Y. Tamanaha, B. Zhang, Y. Guo, W.-C. Chang, J. M. Bollinger Jr and C. Krebs, Spectroscopic evidence for the two C–H-cleaving intermediates of *Aspergillus nidulans* isopenicillin N synthase, *J. Am. Chem. Soc.*, 2016, **138**, 8862–8874.
- 12 E. P. F. Tchesnokov, A. S. Faponle, C. G. Davies, M. G. Quesne, R. Turner, M. Fellner, R. J. Souness, S. M. Wilbanks, S. P. deVisser and G. N. L. Jameson, An iron–oxygen intermediate formed during the catalytic cycle of cysteine dioxygenase, *Chem. Comm.*, 2016, **52**, 8814–8817.
- 13 R. Surendhran, A. A. D'Arpino, B. Y. Sciscent, A. F. Cannella, A. E. Friedman, S. N. MacMillan, R. Gupta and D. C. Lacy, Deciphering the mechanism of O<sub>2</sub> reduction with electronically tunable non-heme iron enzyme model complexes, *Chem. Sci.*, 2018, **9**, 5773–5780.
- 14 M. A. Dedushko, M. B. Greiner, A. N. Downing, M. K. Coggins and J. A. Kovacs, Electronic structure and reactivity of dioxygen-derived thiolate-ligated Fe-peroxo and Fe(IV) oxo compounds, *J. Am. Chem. Soc.*, 2022, **144**, 8515–8528.
- 15 S. V. Kryatov, E. V. Rybak-Akimova and S. Schindler, Kinetics and mechanisms of formation and reactivity of non-heme iron oxygen intermediates, *Chem. Rev.*, 2005, **105**, 2175–2226.
- 16 M. A. Dedushko, PhD thesis, University of Washington, 2020.
- 17 A. A. Fischer, S. V. Lindeman and A. T. Fiedler, A synthetic model of the nonheme iron–superoxo intermediate of cysteine dioxygenase, *Chem. Comm.*, 2018, **54**, 11344–11347.
- 18 S. Hong, K. D. Sutherlin, J. Park, E. Kwon, M. A. Siegler, E. I. Solomon and W. Nam, Crystallographic and



- spectroscopic characterization and reactivities of a mononuclear non-haem iron(III)-superoxo complex, *Nat. Commun.*, 2014, **5**, 5440–5446.
- 19 C.-W. Chiang, S. T. Kleepsies, H. D. Stout, K. K. Meier, P.-Y. Li, E. L. Bominaar, L. Que Jr, E. Munck and W.-Z. Lee, Characterization of a paramagnetic mononuclear nonheme iron-superoxo complex, *J. Am. Chem. Soc.*, 2014, **136**, 10846–10849.
- 20 F. O'ddon, Y. Chiba, J. Nakazawa, T. Ohta, T. Ogura and S. Hikichi, Characterization of mononuclear non-heme iron(III)-superoxo complex with a five-azole ligand set, *Angew. Chem., Int. Ed.*, 2015, **54**, 7336–7339.
- 21 C. Winslow, H. B. Lee, M. J. Field, S. J. Teat and J. Rittle, Structure and reactivity of a high-spin, nonheme iron(III)-superoxo complex supported by phosphinimide ligands, *J. Am. Chem. Soc.*, 2021, **143**, 13686–13693.
- 22 M. N. Blakely, M. A. Dedushko, P. C. Y. Poon, G. Villar-Acevedo and J. A. Kovacs, Formation of a reactive, alkyl thiolate-ligated Fe(III)-superoxo intermediate derived from dioxygen, *J. Am. Chem. Soc.*, 2019, **141**, 1867–1870.
- 23 M. Couture, S. Adak, D. J. Stuehr and D. L. Rousseau, Regulation of the properties of the Heme-NO complexes in nitric-oxide synthase by hydrogen bonding to the proximal cysteine\*, *J. Biol. Chem.*, 2001, **276**, 38280–38288.
- 24 C. M. Krest, A. Silakov, J. Rittle, T. H. Yosca, E. L. Onderko, J. C. Calixto and M. T. Green, Significantly shorter Fe-S bond in cytochrome P450-I is consistent with greater reactivity relative to chloroperoxidase, *Nat. Chem.*, 2015, **7**, 696–702.
- 25 P. K. Das, S. Chatterjee, S. Samanta and A. Dey, EPR, resonance raman, and DFT calculations on thiolate- and imidazole-bound iron(III) porphyrin complexes: role of the axial ligand in tuning the electronic structure, *Inorg. Chem.*, 2012, **51**, 10704–10714.
- 26 N. Ueyama, N. Nishikawa, Y. Yamada, T.-a. Okamura, S. Oka, H. Sakurai and A. Nakamura, Synthesis and properties of octaethylporphyrinato(arenethiolato)iron(III) complexes with intramolecular NH...S hydrogen bond: chemical function of the hydrogen bond, *Inorg. Chem.*, 1998, **37**, 2415–2421.
- 27 R. Davydov, S. Im, M. Shanmugam, W. A. Gunderson, N. M. Pearl, B. M. Hoffman and L. Waskell, Role of the proximal cysteine hydrogen bonding interaction in cytochrome P450 2B4 studied by cryoreduction, electron paramagnetic resonance, and electron-nuclear double resonance spectroscopy, *Biochemistry*, 2016, **55**, 869–883.
- 28 E. I. Solomon, S. I. Gorelsky and A. Dey, Metal-thiolate bonds in bioinorganic chemistry, *J. Comput. Chem.*, 2006, **27**, 1415–1428.
- 29 M. A. Dedushko, J. Pikul and J. A. Kovacs, Superoxide oxidation by a thiolate-ligated iron complex and anion inhibition, *Inorg. Chem.*, 2021, **60**, 7250–7261.
- 30 P. C. Y. Poon, M. A. Dedushko, X. Sun, G. Yang, S. Toledo, E. C. Hayes, A. Johansen, M. C. Piquette, J. A. Rees, S. Stoll, E. Rybek-Akimova and J. A. Kovacs, How metal ion lewis acidity and steric properties influence the barrier to dioxygen binding, peroxy O-O bond cleavage, and reactivity, *J. Am. Chem. Soc.*, 2019, **141**, 15046–15057.
- 31 J. J. Ellison, A. Nienstedt, S. C. Shoner, D. Barnhart, J. A. Cowen and J. A. Kovacs, Reactivity of five-coordinate models for the thiolate-ligated Fe site of nitrile hydratase, *J. Am. Chem. Soc.*, 1998, **120**, 5691–5700.
- 32 D. Schweitzer, J. J. Ellison, S. C. Shoner, S. Lovell and J. A. Kovacs, A synthetic model for the NO-inactivated form of nitrile hydratase, *J. Am. Chem. Soc.*, 1998, **120**, 10996–10997.
- 33 H. L. Jackson, S. C. Shoner, D. Rittenberg, J. A. Cowen, S. Lovell, D. Barnhart and J. A. Kovacs, Probing the influence of local coordination environment on the properties of Fe-type nitrile hydratase model complexes, *Inorg. Chem.*, 2001, **40**, 1646–1653.
- 34 D. Schweitzer, J. Shearer, D. K. Rittenberg, S. C. Shoner, J. J. Ellison, R. Loloee, S. Lovell, D. Barnhart and J. A. Kovacs, Enhancing reactivity via structural distortion, *Inorg. Chem.*, 2002, **41**, 3128–3136.
- 35 S. C. N. Shoner, A. Nienstedt, J. J. Ellison, I. Kung, D. Barnhart and J. A. Kovacs, Structural comparison of thiolate-ligated M(II) = Fe(II), Co(II), Ni(II), and Zn(II) ions wrapped in a chiral helical ligand, *Inorg. Chem.*, 1998, **37**, 5721–5725.
- 36 J. Shearer, J. Nehring, S. Lovell, W. Kaminsky and J. A. Kovacs, Modeling the reactivity of superoxide reducing metalloenzymes with a nitrogen and sulfur coordinated iron complex, *Inorg. Chem.*, 2001, **40**, 5483–5484.
- 37 J. Shearer, S. B. Fitch, W. Kaminsky, J. Benedict, R. C. Scarrow and J. A. Kovacs, How does cyanide inhibit superoxide reductase? Insight from synthetic Fe<sup>III</sup>N<sub>4</sub>S model complexes, *Proc. Natl. Acad. Sci. U. S. A.*, 2003, **100**, 3671–3676.
- 38 C. Guo, J. Yu, J. R. Horsley, M. Sheves, D. Cahen and A. D. Abell, Backbone-constrained peptides: temperature and secondary structure affect solid-state electron transport, *J. Phys. Chem. B*, 2019, **123**, 10951–10958.
- 39 J. S. Kavanaugh, P. H. Rogers and A. Arnone, Crystallographic evidence for a new ensemble of ligand-induced allosteric transitions in hemoglobin: the T-to-T<sub>H</sub> high quaternary transitions, *Biochemistry*, 2005, **44**, 6101–6121.
- 40 K. M. Clark, Y. Yu, N. M. Marshall, N. A. Sieracki, M. J. Nilges, N. J. Blackburn, W. A. van der Donk and Y. Lu, Transforming a blue copper into a red copper protein: engineering cysteine and homocysteine into the axial position of azurin using site-directed mutagenesis and expressed protein ligation, *J. Am. Chem. Soc.*, 2010, **132**, 10093–10101.
- 41 E. L. Hegg, S. H. Mortimore, C. L. Cheung, J. E. Huyett, D. R. Powell and J. N. Burstyn, Structure-reactivity studies in copper(II)-catalyzed phosphodiester hydrolysis, *Inorg. Chem.*, 1999, **38**, 2961–2968.
- 42 J. S. Plegaria, M. Duca, C. Tard, T. J. Friedlander, A. Deb, J. E. Penner-Hahn and V. L. Pecoraro, De novo design and characterization of copper metalloproteins inspired by native cupredoxins, *Inorg. Chem.*, 2015, **54**, 9470–9482.
- 43 P. M. Colman, H. C. Freeman, J. M. Guss, M. Murata, V. A. Norris, J. A. M. Ramshaw and M. P. Venkatappa, Blue copper structure & entatic state, *Nature*, 1978, **272**, 319.



- 44 S. I. Mann, T. Heinisch, T. R. Ward and A. S. Borovik, Peroxide activation regulated by hydrogen bonds within artificial Cu proteins, *J. Am. Chem. Soc.*, 2017, **139**, 17289–17292.
- 45 A. S. Borovik, Bioinspired hydrogen bond motifs in ligand design: the role of noncovalent interactions in metal ion mediated activation of dioxygen, *Acc. Chem. Res.*, 2005, **38**, 54–61.
- 46 S. A. B. Cook and A. S. Borovik, Molecular designs for controlling the local environments around metal ions, *Acc. Chem. Res.*, 2015, **48**, 2407–2414.
- 47 S. A. H. Cook, E. A. Hill and A. S. Borovik, Lessons from Nature: a bio-inspired approach to molecular design, *Biochemistry*, 2015, **54**, 4167–4180.
- 48 M. K. Coggins, X. Sun, Y. Kwak, E. I. Solomon, E. Rybak-Akimova and J. A. Kovacs, Characterization of metastable intermediates formed in the reaction between a Mn(II) complex and dioxygen, including a crystallographic structure of a binuclear Mn(III)–peroxo species, *J. Am. Chem. Soc.*, 2013, **135**, 5631–5640.
- 49 H. C. S. Fry, D. V. Scaltrito, K. D. Karlin and G. J. Meyer, The Rate of O<sub>2</sub> and CO Binding to a Copper Complex, Determined by a “Flash-and-Trap” Technique, Exceeds that for Hemes, *J. Am. Chem. Soc.*, 2003, **125**, 11866–11871.
- 50 H. R. Lucas, G. J. Meyer and K. D. Karlin, CO and O<sub>2</sub> Binding to Pseudo-tetradentate Ligand–Copper(I) Complexes with a Variable N-Donor Moiety: Kinetic/Thermodynamic Investigation Reveals Ligand-Induced Changes in Reaction Mechanism, *J. Am. Chem. Soc.*, 2010, **132**, 12927–12940.
- 51 N. W. Aboeella, S. V. Kryatov, B. F. Gherman, W. W. Brennessel, V. G. Young, R. Sarangi, E. V. Rybak-Akimova, K. O. Hodgson, B. Hedman, E. I. Solomon, C. J. Cramer and W. B. Tolman, Dioxygen activation at a single copper site: structure, bonding, and mechanism of formation of 1:1 Cu–O<sub>2</sub> adducts, *J. Am. Chem. Soc.*, 2004, **126**, 16896–16911.
- 52 M. Weitzer, S. Schindler, G. Brehm, E. Hörmann, B. Jung, S. Kaderli and A. Zuberbühler, Reversible Binding of Dioxygen by the Copper(I) Complex with Tris(2-dimethylaminoethyl)amine (Me<sub>6</sub>tren) Ligand, *Inorg. Chem.*, 2003, **42**, 1800–1806.
- 53 C. X. Zhang, S. Kaderli, M. Costas, E. Kim, Y.-M. Neuhold, K. D. Karlin and A. D. Zuberbühler, Copper(I)–dioxygen reactivity of [(L)CuI]<sup>+</sup> (L = tris(2-pyridylmethyl)amine): kinetic/thermodynamic and spectroscopic studies concerning the formation of Cu–O<sub>2</sub> and Cu<sub>2</sub>–O<sub>2</sub> adducts as a function of solvent medium and 4-pyridyl ligand substituent variations, *Inorg. Chem.*, 2003, **42**, 1807–1824.
- 54 E. V. Rybak-Akimova, W. Otto, P. Dearth, R. Roesner and D. H. Busch, Kinetics and equilibrium of dioxygen binding to a vacant site in cobalt(II) complexes with pentadentate ligands, *Inorg. Chem.*, 1997, **36**, 2746–2753.
- 55 A. F. Cozzolino, D. Tofan, C. C. Cummins, M. Temprado, T. D. Palluccio, E. V. Rybak-Akimova, S. Majumdar, X. Cai, B. Captain and C. D. Hoff, Two-step binding of O<sub>2</sub> to a vanadium(III) trisanilide complex to form a non-vanadyl vanadium(V) peroxo complex, *J. Am. Chem. Soc.*, 2012, **134**, 18249–18252.
- 56 R. Cini and P. Orioli, Crystal and molecular structure of two oxygen-inactive forms of [N,N'-(3,3'-dipropylmethylamine) bis(salicylideneiminato) cobalt(II)], *Inorg. Chim. Acta*, 1982, **63**, 243.
- 57 T. D. Palluccio, X. Cai, S. Majumdar, L. F. Sarafim, N. C. Tomson, K. Wieghardt, C. S. J. Cazin, S. P. Nolan, A. F. Cozzolino, E. V. Rybak-Akimova, M. A. Fernandez-Gonzalez, M. Temprado, B. Captain and C. D. Hoff, Ligand-directed reactivity in dioxygen and water binding to *cis*-[Pd(NHC)<sub>2</sub>(η<sub>2</sub>-O<sub>2</sub>)], *J. Am. Chem. Soc.*, 2018, **140**, 264–276.
- 58 X. Cai, S. Majumdar, G. C. Fortman, C. S. J. Cazin, A. M. Z. Slawin, C. Lhermitte, R. Prabhakar, M. E. Germain, T. D. Palluccio, S. P. Nolan, E. V. Rybak-Akimova, M. Temprado, B. Captain and C. D. Hoff, Oxygen binding to [Pd(L)(L0)] (L = NHC, L0 = NHC or PR<sub>3</sub>, NHC = N-heterocyclic carbene) synthesis and structure of a paramagnetic *trans*-[Pd(NHC)<sub>2</sub>(η<sub>1</sub>-O<sub>2</sub>)<sub>2</sub>] complex, *J. Am. Chem. Soc.*, 2011, **133**, 1290–1293.
- 59 J. P. Collman, J. I. Brauman, B. L. Iverson, J. L. Sessler, R. M. Morris and Q. H. Gibson, O<sub>2</sub> and CO binding to iron(II) porphyrins: a comparison of the “picket fence” and “pocket” porphyrins, *J. Am. Chem. Soc.*, 1983, **105**, 3052–3064.
- 60 M. R. Momenteau and C. A. Reed, Synthetic heme dioxygen complexes, *Chem. Rev.*, 1994, **94**, 659–698.
- 61 M. M. Purdy, L. S. Koo, P. R. Ortiz de Montellano and J. P. Klinman, Mechanism of O<sub>2</sub> Activation by cytochrome P450cam Studied by isotope effects and transient state kinetics, *Biochemistry*, 2006, **45**, 15793–15806.
- 62 I. Szundi, C. Kittredge, S. K. Choi, W. McDonald, J. Ray, R. B. Gennis and O. Einarsdottir, Kinetics and intermediates of the reaction of fully reduced *Escherichia coli* bo<sub>3</sub> ubiquinol oxidase with O<sub>2</sub>, *Biochemistry*, 2014, **53**, 5393–5404.
- 63 E. E. Weinert, C. M. Phillips-Piro, R. Tran, R. A. Mathies and M. A. Marletta, Controlling conformational flexibility of an O<sub>2</sub>-binding H-NOX domain, *Biochemistry*, 2011, **50**, 6832–6840.
- 64 B. A. Springer, S. G. Sligar, J. S. Olson and G. N. Phillips Jr, Mechanisms of ligand recognition in myoglobin, *Chem. Rev.*, 1994, **94**, 699–714.
- 65 E. Antonini and M. Brunori, *Hemoglobin and Myoglobin in Their Reactions with Ligands*, North-Holland, Amsterdam, 1971.

