ChemComm



COMMUNICATION

View Article Online



Cite this: Chem. Commun., 2020, 56 14829

Received 14th August 2020, Accepted 30th October 2020

DOI: 10.1039/d0cc05539e

rsc.li/chemcomm



Hui Min Tay, Da Aditya Rawalb and Carol Hua D*

Chirality is a key aspect of amino acids and is essential for life. Here, a chiral metal-organic framework, S-Mg2dobpdc, is used to determine the chirality of three BOC protected amino acids (alanine, valine and proline) by ¹³C solid-state NMR with chemical shift differences of up to 1.3 ppm observed between enantiomers. The chiral sensitivity persists upon in situ deprotection of the amino acids by thermolysis of the BOC group.

Amino acids are the building blocks of proteins and an important component in chemical and biological systems. The different enantiomers of amino acids can lead to vastly different physiological and metabolic outcomes. The growing use of amino acid derivatives in pharmacology, chemical science and biochemistry highlights the need for efficient methods to determine chiral purity. The elucidation of chirality in amino acids can be challenging due to their lack of strong chromophoric groups, leading to a weak chiro-optical response in circular dichroism spectroscopy, 1 or the need for multiple protection, derivation and deprotection steps. The majority of chiral elucidation methods involve the use of chromatographic techniques, including HPLC and GC,² or optical methods, including fluorescence and luminescence.³ Solid-state NMR offers a direct means of identifying and quantifying the enantiopurity of a sample. The use of solid-state NMR for the quantification of chirality is underdeveloped, with the main focus in this area concerning the use of liquid crystals as a chiral matrix.8,9

Coordination Polymers (CPs) and Metal Organic Frameworks (MOFs) are crystalline solid materials comprising of metal ions bridged by organic ligands in 1D, 2D or 3D to create a scaffold containing pores. 4 Chiral MOFs and CPs are typically

used as stationary chiral phases for HPLC⁵ or as heterogenous asymmetric catalysts.6 Considerably less attention has been focussed on the use of MOFs and CPs for the detection of chiral purity. Developments in this area with MOFs and CPs primarily utilise optical spectroscopic techniques, including fluorescence and UV-Vis.7 CPs and MOFs can potentially act as solid-state chiral solvating agents due to their tunable pores, which can form strong intermolecular interactions with chiral guests, resulting in a diastereomeric complex observable by NMR. While solid-state NMR spectra can have large peak widths, making the small chemical shifts between diastereomers difficult to resolve, MOFs and CPs are crystalline materials that typically have narrow peak widths and sharp NMR resonances.

While three previous reports have used MOFs for chiral elucidation, none of these were able to differentiate between enantiomers from a racemic mixture. 10-12 Only small chemical shift differences of up to 0.4 ppm were observed in the ¹³C NMR spectra of a chiral analogue of the UMCM-1 framework¹¹ upon loading with enantiopure 1-phenyl-2,2,2-trifluoroethanol (TFPE), or when alanine and leucine were loaded into [Zn₂(L-Phe)(bpe)₂]_n (L-Phe = L-phenylalanine, bpe = bis(4-pyridyl)ethylene). 12

Long et al. recently reported the synthesis of a chiral form of Mg_2 dobpdc (H_4 dobpdc = 4,4'-dioxidobiphenyl-3,3'-dicarboxylic acid). 13 The axial chirality of Mg₂dobpdc results from the rotation of the two phenyl rings in dobpdc4- relative to each other, yielding enantiomerically pure domains of solely right-handed or left-handed helices. The grafting of chiral 1,2-diaminocyclohexane enabled the solid-state NMR characterisation of the ammonium carbamate chains formed upon the adsorption of CO₂ to yield chemical shift differences of up to 2.0 ppm. Mg₂(dobpdc) (and its Zn^{II} analogue) have been extensively studied by solid-state NMR, ¹⁴⁻¹⁹ including in experiments that probe molecular dynamics using 15N NMR20 and investigations of local disorder by ²⁵Mg NMR. ^{21,22} The breadth of information concerning the behaviour of Mg₂(dobpdc) makes it a valuable target for the study of chiral elucidation by solid-state NMR.

Herein, we present the use of S-Mg₂dobpdc for the chiral elucidation of amino acids using 13C solid-state NMR. S-Mg2dobpdc

^a School of Chemistry, The University of Melbourne, Parkville, Victoria, 3010, Australia. E-mail: carol.hua@unimelb.edu.au

^b Mark Wainwright Analytical Centre, University of New South Wales, Kensington, New South Wales, 2052, Australia

[†] Electronic supplementary information (ESI) available: Thermal gravimetric analysis (TGA) traces, powder X-ray diffraction (PXRD) patterns, ATR infrared spectra (IR), solution state NMR spectra and additional 13C CPMAS spectra. See DOI: 10.1039/d0cc05539e

Communication ChemComm

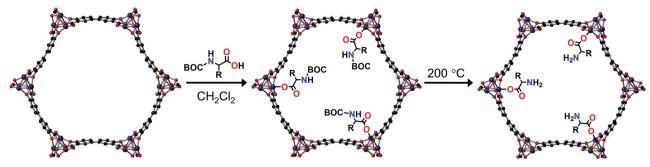


Fig. 1 Schematic of the appendage of BOC protected amino acids to the open Mg^{II} metal sites on $S-Mg_2dobpdc$ to yield $S-Mg_2dobpdc$ BOC-Xxx (Xxx = Ala, Val, Val, Val) where Val (Val) where Val (Val) where Val (Val) where Val) where Val (Val) where Val (Val) where Val) where Val (Val) where Val (Va

was grafted post-synthetically with enantiopure BOC alanine, valine and proline to the ${\rm Mg^{II}}$ metal site. Unprotected amino acids were obtained by thermal deprotection of the BOC groups post-grafting. The $^{13}{\rm C}$ NMR chemical shift differences between the L- and D-enantiomers of the BOC protected and free amino acids as well as the S-Mg₂dobpdc framework were determined and the mode of binding discussed.

S-Mg₂(dobpdc) was made according to the procedure reported by Long et al.¹³ Enantiomerically pure di-tert-butyl dicarbonate (BOC) protected alanine, valine and proline were then grafted onto the framework to yield S-Mg₂(dobpdc)@BOC-L-Xxx (Xxx = Ala, Val, Pro) and S-Mg₂(dobpdc)@BOC-D-Xxx (Xxx = Ala, Val, Pro), as white powders (Fig. 1). The BOC group ensured that coordination to the Mg^{II} metal centre only occurred through the carboxylic acid instead of the primary amine of the amino acid. Alanine, valine and proline were chosen for incorporation to minimise the steric bulk present, ensuring that a high percentage loading and strong intermolecular interactions to the framework could be achieved. Amino acids with aliphatic side chains enabled easy identification from the aromatic dobpdc4- ligand in NMR studies. The PXRD of S-Mg₂(dobpdc)@BOC-Xxx (Xxx = Ala, Val, Pro) verified that the crystallinity and structure of the S-Mg₂(dobpdc) framework had been retained (ESI,† Fig. S1-S4). The incorporation of the BOC amino acid guest into S-Mg2dobpdc was verified by the BOC carbonyl peak in the FT-IR spectrum at 1670 cm⁻¹ (ESI,† Fig. S5-S7) and via solution-state ¹H NMR spectroscopy of the digested samples of S-Mg₂(dobpdc)@BOC-Xxx (Xxx = Ala, Val, Pro) (ESI,† Fig. S8-S13).

The thermal gravimetric analyses (TGA) of S-Mg₂(dobpdc) @BOC-Xxx (Xxx = Ala, Val, Pro) displayed three stepwise mass losses (ESI,† Fig. S14–S22). The first mass loss below 100 °C was attributed to the loss of water adsorbed from the atmosphere. The second mass loss between 100 and 200 °C was due to the thermal deprotection of the BOC group from the appended BOC amino acids. The post-synthetic deprotection of the BOC group from proline within a MOF has previously been reported. ^{23–25} The third mass loss between 220 and 280 °C is due to thermal decomposition of the amino acids into gaseous products and can be correlated to the size of the side chain, with S-Mg₂(dobpdc)@BOC-Val having the largest mass loss. ^{26,27}

The thermal BOC deprotection of the amino acids was confirmed by FT-IR measurements after heating the *S*-Mg₂-(dobpdc)@BOC-Xxx (Xxx = Ala, Val, Pro) samples at 200 °C for 30 mins to yield *S*-Mg₂(dobpdc)@Xxx, where the BOC carbonyl peak at 1670 cm⁻¹ disappeared (ESI,† Fig. S23–S25). The conversion of BOC-Ala to Ala resulted in red shifts of the FT-IR absorption bands to wavelengths similar to those for the Mg₂dobpdc framework. Remarkably, PXRD measurements (ESI,† Fig. S26–S28) confirmed that crystallinity was retained after BOC deprotection. In the TGA of *S*-Mg₂(dobpdc)@Xxx (Xxx = Ala, Val, Pro), the mass loss between 100 and 200 °C was absent, confirming the successful deprotection of the BOC group (ESI,† Fig. S29–S31).

As the removal of the BOC group can be directly observed from the mass loss in the TGA traces, this was used to determine the percentage incorporation of the amino acid, which was verified by elemental analysis. The highest percentage loading was achieved with BOC-Ala (57%), followed by BOC-Pro (55%) and BOC-Val (39%). It is likely that both the degree of steric bulk and the conformation of the amino acid had an effect on the amount of amino acid incorporated into *S*-Mg₂(dobpdc). DMF solvent molecules remained coordinated to the Mg^{II} metal sites that have no amino acid appended, even after heating the samples at 200 °C, as observed in the ¹³C CPMAS spectra (*vide infra*).

The *S*-Mg₂dobpdc framework and the amino acid appended samples were analysed by ¹³C solid-state NMR (Fig. 2). The ¹³C CPMAS spectrum of *S*-Mg₂dobpdc contained well-defined spectral lines with widths at half height of 0.7 ppm, enabling all the ¹³C resonances arising from the dobpdc⁴ ligand to be assigned (ESI,† Fig. S33). A ¹³C non-quaternary suppression (NQS) experiment was used to assign the quaternary carbon resonances (ESI,† Fig. S34–S36).

The 13 C CPMAS spectra of S-Mg₂(dobpdc)@BOC-Xxx (Xxx = Ala, Val, Pro) showed distinct chemical shift differences of up to 0.6 ppm in specific 13 C NMR resonances between the L- and D-amino acids (Fig. 2a–c and Fig. S37–S42, ESI†). The largest chemical shift occurred at C2, C3, C4, C5 and C_e, suggesting that the methyl groups from the BOC group are involved in CH– π intermolecular interactions with the phenyl

ChemComm

(b) (c) L: 28.2 ppm DL: 28.5 ppm D: 28.6 ppm L: 125.8 ppm DL: 125.7 ppm L: 51.8 ppm DL: 51.4 ppm L: 20.1, 17.6 ppm DL: 20.1, 18.2 ppm D: 20.1, 17.6 ppm L: 31.5 ppm DL: 31.5 ppm D: 31.8 pr **C**5 DMF D: 117.0 ppm COO D: 36 9 ppn 60 140 105 55 45 200 180 160 140 120 100 80 60 40 0 145 135 130 125 120 110 40 15 ¹³C NMR Chemical Shift (ppm) ¹³C NMR Chemical Shift (ppm) ¹³C NMR Chemical Shift (ppm) (e) L: 130.6 ppm DL: 130.6 ppm (f) L: 50.6 ppm DL: 50.6 ppm DL: 125.4 ppm D: 130.4 ppm D: 50.7 ppm DL: 35.5 ppm DL: 29.6 ppm D: 36.4 ppm D: 30.9 ppm C2 C4 D: 20.3 ppm

Fig. 2 13C solid state CPMAS spectra of Mg2dobpdc@BOC-L-Ala (black), Mg2dobpdc@BOC-D-Ala (red) and Mg2dobpdc@BOC-DL-Ala (blue) acquired at 175 MHz showing (a) the full spectrum between 200-0 ppm, (b) zoomed in section between 145-105 ppm, and (c) zoomed in section between 60-10 ppm. The 13 C solid state CPMAS spectra of Mg₂dobpdc@L-Ala (black), Mg₂dobpdc@D-Ala (red) and Mg₂dobpdc@DL-Ala (blue) acquired at 175 MHz showing (d) the full spectrum between 200-0 ppm, (e) zoomed in section between 145-105 ppm, and (f) zoomed in section between 45-10 ppm. All spectra are scaled to the same maximum intensity.

¹³C NMR Chemical Shift (ppm)

rings of the dobpdc⁴⁻ ligand. Remarkably, the C2-C5 signals of dobpdc4- can be used to discriminate between the L and D enantiomers of BOC-Ala by the ¹³C NMR chemical shift. Mg2dobpdc@BOC-DL-Ala (comprised of 60% BOC-L-Ala, 40% BOC-D-Ala) has a chemical shift of 130.9 ppm, which is intermediate between that of Mg2dobpdc@BOC-L-Ala (131.2 ppm) and Mg₂dobpdc@BOC-p-Ala (130.6 ppm) (Fig. 2b). ¹³C chemical shift differences also occur for C3 and C_{β} . It is surprising that no changes in the chemical shift were observed for C_{α} , given this is where the stereocentre is located, however, chiral elucidation in S-Mg₂dobpdc likely involves the different orientations of the amino acid relative to the aromatic rings of dobpdc⁴⁻. A similar interaction was observed for UMCM where the 13C resonances of the framework, instead of the stereocentre, changed upon the incorporation of chiral guest molecules.¹¹

160 140 120 100

80 60 40 20

¹³C NMR Chemical Shift (ppm)

0 145

140 135 130 125 120 115 110 105 60

The largest chemical shift changes were observed for alanine, followed by proline (ESI,† Fig. S37-S39), then valine (ESI,† Fig. S40-S42). This trend is attributed to the size of the amino acid side chain, with the small methyl side chain in alanine enabling the amino acid to effectively interact with S-Mg₂dobpdc. The valine isopropyl side chain has greater steric bulk, precluding the formation of strong intermolecular interactions required for effective chiral discrimination. The orientation of the side chain relative to the carboxylate and BOC groups of the amino acid is also an important consideration. The restricted movement of the five membered pyrrolidene ring in proline likely helps to yield a geometry that allows enhanced interaction with the dobpdc⁴⁻ ligand.

The steric effect of the BOC group on chiral discrimination by ¹³C solid state NMR was additionally investigated by analysis of the deprotected amino acid samples of S-Mg2dobpdc@Ala. The ¹³C CPMAS spectra of S-Mg₂(dobpdc)@L-Ala and S-Mg₂(dobpdc)@D-Ala displayed enhanced ¹³C chemical shift differences of up to 1.3 ppm when compared with the BOCprotected samples, with the largest chemical shift differences observed for the CH3 groups of DMF, followed by that of C1 and C_{β} (Fig. 2d-f). The removal of the bulky BOC group enables a stronger interaction between the amino acid and the framework components, thereby enhancing chiral elucidation. The broad C_{β} peak of alanine becomes narrower when the BOC group is removed, indicating that the BOC group may cause some conformational disorder. When the framework was loaded with DL-Ala, the 13C chemical shifts more closely resembled that of Mg₂(dobpdc)@L-Ala than Mg₂(dobpdc)@ D-Ala. This is consistent with the composition of DL-Ala, which consists of a 60:40 ratio of L- to D-Ala.

50

¹³C NMR Chemical Shift (ppm)

The linewidths in the NMR spectra of the amino acid loaded frameworks were broader than for S-Mg₂(dobpdc), with linewidths at half height of 1.8 ppm. This can be attributed to the local disorder of the Mg^{II} environment upon guest loading. Previous studies of ²⁵Mg NMR on rac-Mg₂dobpdc and Mg₂ dobdc ($dobdc^{4-} = 2,5$ -dihydroxyterephthalate) have shown that although long-range ordering is maintained upon dehydration and guest loading, slight changes in the orientation of the ligands result in small changes in the Mg-O bond length and O-Mg-O bond angle to yield local disorder. 21,22

Communication ChemComm

The porous and robust S-Mg2dobpdc framework was shown to be highly effective for chiral elucidation by ¹³C solid state NMR with BOC-alanine, BOC-valine, BOC-proline and alanine with up to a 1.3 ppm difference. The chemical shift changes between the L- and D-amino acids were attributed to intermolecular interactions between the amino acid and the aromatic protons of S-Mg₂dobpdc. The largest changes in the ¹³C chemical shift between the L- and D-enantiomers occur in alanine-loaded frameworks due to the low steric bulk of the side chain which enables strong intermolecular interactions to be formed with S-Mg₂dobpdc. It is noted that S-Mg₂dobpdc is locally sensitive to the chirality of the amino acid which is detected by ¹³C NMR, but is unable to be detected by PXRD, which measures long range ordering. Future studies include the analysis of other amino acids, including those containing aromatic side chains and additional Hydrogen bonding sites, as well as computational calculations to determine the origin of the ¹³C chemical shift difference between enantiomers. This study has demonstrated that solid-state NMR spectroscopy together with chiral MOFs is a versatile and elegant approach to overcoming the challenges of chiral elucidation for amino acids.

We thank Dr Doug Lawes for assistance with ¹H solution state NMR, Dr Marc-Antione Sani for preliminary solid-state NMR experiments, Ms Lisa Hua for experimental assistance and A/Prof. Brendan Abrahams for support during this work. H. M. T. thanks the Australian Commonwealth Government and the University of Melbourne for a Research Training Programme Scholarship, Rowden White Scholarship and an Elizabeth and Vernon Puzey Scholarship. C. H. gratefully acknowledges the University of Melbourne for a McKenzie Fellowship.

Conflicts of interest

There are no conflicts to declare.

Notes and references

- 1 C. Wolf and K. W. Bentley, Chem. Soc. Rev., 2013, 42, 5408-5424.
- 2 M. Dołowy and A. Pyka, Biomed. Chromatogr., 2014, 28, 84-101.
- 3 J. Wang, H.-B. Liu, Z. Tong and C. S. Ha, Coord. Chem. Rev., 2015, 303, 139-184.
- 4 H. Furukawa, K. E. Cordova, M. O'Keeffe and O. M. Yaghi, Science, 2013, 341, 1230444.

- 5 T. Duerinck and J. F. M. Denayer, Chem. Eng. Sci., 2015, 124, 179-187.
- 6 C. Wang, M. Zheng and W. Lin, J. Phys. Chem. Lett., 2011, 2, 1701-1709.
- 7 X. Zhang, J. Yin and J. Yoon, Chem. Rev., 2014, 114, 4918-4959.
- 8 T. J. Wenzel, Discrimination of Chiral Compounds Using NMR Spectroscopy, John Wiley & Sons, Hoboken, New Jersey, 2007.
- 9 A. Kumar and N. Suryaprakash, Two-Dimensional NMR of Molecules Oriented in Liquid Crystals-Recent Developments, John Wiley & Sons, Ltd. 2007.
- 10 C. Kutzscher, H. C. Hoffmann, S. Krause, U. Stoeck, I. Senkovska, E. Brunner and S. Kaskel, Inorg. Chem., 2015, 54, 1003-1009.
- C. Hoffmann, S. Paasch, P. Müller, I. Senkovska, Padmanaban, F. Glorius, S. Kaskel and E. Brunner, Chem. Commun., 2012, 48, 10484-10486.
- 12 X. Ma, Y. Zhang, Y. Gao, X. Li, C. Wang, H. Yuan, A. Yu, S. Zhang and Y. Cui, Chem. Commun., 2020, 56, 1034-1037.
- 13 J. D. Martell, L. B. Porter-Zasada, A. C. Forse, R. L. Siegelman, M. I. Gonzalez, J. Oktawiec, T. Runčevski, J. Xu, M. Srebro-Hooper, P. J. Milner, K. A. Colwell, J. Autschbach, J. A. Reimer and J. R. Long, J. Am. Chem. Soc., 2017, 139, 16000-16012.
- 14 A. C. Forse, S. A. Altobelli, S. Benders, M. S. Conradi and J. A. Reimer, J. Phys. Chem. C, 2018, 122, 15344-15351.
- 15 A. C. Forse, M. I. Gonzalez, R. L. Siegelman, V. J. Witherspoon, S. Jawahery, R. Mercado, P. J. Milner, J. D. Martell, B. Smit, B. Blümich, J. R. Long and J. A. Reimer, J. Am. Chem. Soc., 2018, 140, 1663-1673.
- 16 A. C. Forse, K. A. Colwell, M. I. Gonzalez, S. Benders, R. M. Torres-Gavosto, B. Blümich, J. A. Reimer and J. R. Long, Chem. Mater., 2020, 32, 3570-3576.
- 17 P. J. Milner, R. L. Siegelman, A. C. Forse, M. I. Gonzalez, T. Runčevski, J. D. Martell, J. A. Reimer and J. R. Long, J. Am. Chem. Soc., 2017, 139, 13541-13553.
- 18 A. C. Forse, P. J. Milner, J. H. Lee, H. N. Redfearn, J. Oktawiec, R. L. Siegelman, J. D. Martell, B. Dinakar, L. B. Porter-Zasada, M. I. Gonzalez, J. B. Neaton, J. R. Long and J. A. Reimer, J. Am. Chem. Soc., 2018, 140, 18016-18031.
- 19 R. L. Siegelman, P. J. Milner, A. C. Forse, J. H. Lee, K. A. Colwell, J. B. Neaton, J. A. Reimer, S. C. Weston and J. R. Long, J. Am. Chem. Soc., 2019, 141, 13171-13186.
- 20 J. Xu, Y. M. Liu, A. S. Lipton, J. Ye, G. L. Hoatson, P. J. Milner, T. M. McDonald, R. L. Siegelman, A. C. Forse, B. Smit, J. R. Long and J. A. Reimer, J. Phys. Chem. Lett., 2019, 10, 7044-7049.
- 21 J. Xu, V. V. Terskikh and Y. Huang, J. Phys. Chem. Lett., 2013, 4, 7-11.
- 22 J. Xu, E. S. M. Blaakmeer, A. S. Lipton, T. M. McDonald, Y. M. Liu, B. Smit, J. R. Long, A. P. M. Kentgens and J. A. Reimer, J. Phys. Chem. C, 2017, 121, 19938-19945.
- 23 X. Zhou, F. Xu, Z. Wu, H. Li and S. Yang, ACS Omega, 2019, 4, 8588-8597.
- 24 C. Kutzscher, G. Nickerl, I. Senkovska, V. Bon and S. Kaskel, Chem. Mater., 2016, 28, 2573-2580.
- 25 D. J. Lun, G. I. N. Waterhouse and S. G. Telfer, J. Am. Chem. Soc., 2011, 133, 5806-5809.
- 26 Y. C. Lien and W. W. Nawar, J. Food Sci., 1974, 39, 911-913.
- 27 Y. C. Lien and W. W. Nawar, J. Food Sci., 1974, 39, 914-916.