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Rational design of ratiometric and lysosome-targetable AIE dots for imaging endogenous HClO in live cells†

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Rational design of novel ratiometric and targetable fluorescent probes for selective imaging of lysosomal hypochlorous acid (HClO) is conducive to investigating the function of HClO in pathophysiological processes. Herein, we for the first time combine 1-pyrenecarboxaldehyde (with a π - π stacking effect) and 2-methylquinoline derivatives to synthesize an aggregation-induced-emission (AIE) probe (**SA-C2-PCD**) for ratiometric detection of HClO in living cells. Subsequently, a kind of novel lysosome-targetable AIE dot (**AIED-Lyso**) with dual emission features is prepared via a simple co-precipitation of a lysosome-targeting surfactant MAA-CO720, an amphiphilic block copolymer PEO₁₁₃-*b*-PS₄₆ and the HClO-responsive probe (**SA-C2-PCD**). The free **AIED-Lyso** shows only strong red AIE fluorescence without addition of HClO. However, when exposed to HClO, a strong oxidizing agent, the **SA-C2-PCD** would release 1-pyrenecarboxaldehyde that aggregates through π - π stacking interaction in aqueous solutions, inducing a distinct fluorescence change from red to blue. Interestingly, the as-prepared **AIED-Lyso** displays good water dispersity, high selectivity and sensitivity (~ 70.4 nM), strong anti-interference ability, prominent long-term fluorescence stability (> 10 weeks), and low cytotoxicity, and it can be successfully applied to visually detect endogenous lysosomal HClO in live cells.

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Introduction

Hypochlorous acid (HClO) is known as an active oxygen molecule that is mainly produced by hydrogen peroxide and chloride ions under the catalysis of myeloperoxidase (MPO) and it also

plays an important role in fighting pathogenic microorganisms, resisting viruses, *etc.*^{1,2} Abnormal levels of endogenous HClO can cause a series of diseases, including Parkinson's disease, Alzheimer's disease, cancer, *etc.*^{3,4} On the other hand, lysosomes, as an acidic organelle (pH 4.5–5.5), have long been considered as a waste disposal facility in cells.⁵ Recent studies have shown that lysosomes possess various biological functions, including cell signal transduction, protein degradation, plasma membrane repair, homogenous stability, and autophagy, and are regarded as critical subcellular organelles.^{6,7} Lysosomes are one of the main HClO-producing organelles, and excess HClO can induce cell death through calpain activation and lysosomal disruption.^{8,9} Therefore, qualitative analysis and detection of HClO in lysosomes are of great significance.

The recently developed fluorescence method has the merits of simple operation, rapid real-time analysis, and high spatial and temporal resolution, and has received extensive attention and significant development.^{10–18} To date, a series of small molecule-based fluorescent probes for optically sensing HClO have been fabricated.^{19–22} However, most of them show a single fluorescence signal that often varies with other factors such as microenvironment, sensor concentration, photobleaching, and excitation intensity and thus the reliability could be low.^{23–25}

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In this study, we report on the construction of lysosome-targetable AIE dots (**AIED-Lyso**) for ratiometric imaging of lysosomal HClO. Our AIE dots are made *via* a simple co-precipitation of a lysosome-targetable surfactant MAA-CO720, an amphiphilic block copolymer PEO₁₁₃-*b*-PS₄₆ and a HClO-responsive probe (**SA-C2-PCD**), as shown in Scheme 1. In our design, in the presence of strong oxidant HClO, the double bond of **SA-C2-PCD** would break to reduce its red AIE fluorescence and also generate 1-pyrenecarboxaldehyde. This new-born chromophore would induce a progressively enhanced blue fluorescence due to intermolecular π - π stacking interaction. The as-prepared **AIED-Lyso** with good biocompatibility can mainly locate in lysosomes, and monitor endogenous HClO in lysosomes.

Materials and instruments

The strategy for the fabrication of AIED-Lyso

The figure is divided into two main panels illustrating the synthesis and application of PMA-TPP conjugates.

Top Panel: Synthesis and Application of PMA-TPP Conjugates

- Synthesis:** TPP (represented by a grey star) and PMA (represented by a green wavy line) react via **Reprecipitation** to form a red, spiky **PMA-TPP conjugate**.
- Application:** The conjugate is taken up by a cell (indicated by a dashed line). Inside the **Lysosome**, it is released (indicated by a dashed line). The conjugate interacts with **H₂O₂** and **Cl⁻** in the presence of **MPO** to generate **HClO**, which then reacts with the conjugate.
- Stimulation:** The conjugate is also shown reacting with **LPS/PMA** and being **Stimulated** by **HClO** to form a blue, spiky **PMA-TPP conjugate**.

Bottom Panel: Synthesis and Application of the PMA-TPP Conjugate

- Synthesis:** TPP (represented by a grey star) and PMA (represented by a green wavy line) react via **Aggregation** to form a red, spiky **aggregates**.
- Application:** The aggregates are taken up by a cell (indicated by a dashed line). Inside the **Lysosome**, they are released (indicated by a dashed line). The aggregates interact with **H₂O₂** and **Cl⁻** in the presence of **MPO** to generate **HClO**, which then reacts with the aggregates to form a blue, spiky **π-π stack**.

Chemical Structures:

- Top Left:** Chemical structure of a PMA-TPP conjugate. It consists of a TPP core (grey star) linked to a PMA chain (green wavy line). The PMA chain is terminated by a C₁₁H₂₃ group. The structure is labeled with **II** and **48**.
- Bottom Left:** Chemical structure of a PMA-TPP conjugate. It consists of a TPP core (grey star) linked to a PMA chain (green wavy line). The PMA chain is terminated by a C₁₁H₂₃ group. The structure is labeled with **II** and **48**.
- Bottom Right:** Chemical structure of a PMA-TPP conjugate. It consists of a TPP core (grey star) linked to a PMA chain (green wavy line). The PMA chain is terminated by a C₁₁H₂₃ group. The structure is labeled with **II** and **48**.

The synthesis of **SA-C2-PCD** and **MAA-CO720** is described in Schemes S1 and S2 (ESI[†]). All the intermediates and the final product were characterized by ¹H NMR, ¹³C NMR and mass spectroscopy (Fig. S1–S5, ESI[†]).

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Fig. 1 (A) TEM image of **AIED-Lyso**; (B) size of **AIED-Lyso** determined by DLS.

Ratiometric detection of HClO

We first tested the fluorescence of **SA-C2-PCD** under different conditions (THF/water mixture) to confirm its AIE property (Fig. S8, ESI[†]). In THF, a good solvent for **SA-C2-PCD**, no visible fluorescence was observed. When water was added to induce aggregation, the relative emission intensity (F_0/F) of **SA-C2-PCD** gradually increased at first (water fractions $f_w < 80$ vol%) and then jumped at $f_w > 80$ vol%, showing a F_0/F of 198 at $f_w = 95$ vol%, indicating a typical AIE effect.

We then investigated the responsiveness of **AIED-Lyso** to HClO. Note that **AIED-Lyso** itself exhibited an obvious absorption peak at 440 nm in the solution state. When HClO was added, the peak at 440 nm gradually declined, accompanied by the formation of an increasing new absorption peak centered at 350 nm (Fig. S9, ESI[†]). Simultaneously, as shown in Fig. 2A and B (Fig. S10, ESI[†]), obvious blue-shift from 620 nm to 475 nm was observed in the emission spectra, suggesting a reliable ratiometric detection mode. Moreover, the fluorescence intensity ratio (F_{475}/F_{620}) varies linearly with HClO concentration (0–5 μM) with a detection limit of 70.4 nM (Fig. S11, ESI[†]).



Fig. 2 (A) Fluorescence response of **AIED-Lyso** ($2.67 \mu\text{g mL}^{-1}$) towards different concentrations of HClO (0–20 μM , $\lambda_{\text{ex}} = 399$ nm); (B) the fluorescence intensity ratio (F_{475}/F_{620}) as a function of HClO concentration (0–20 μM); (C) the fluorescence intensity ratio (F_{475}/F_{620}) as a function of time; and (D) the fluorescence intensity ratio (F_{475}/F_{620}) without and with HClO (20 μM) as a function of pH.



Fig. 3 (A) Fluorescence intensity ratio (F_{475}/F_{620}) for **AIED-Lyso** ($2.67 \mu\text{g mL}^{-1}$) in the presence of 20 μM HClO or one biologically relevant molecule; (B) fluorescence intensity ratio (F_{475}/F_{620}) for **AIED-Lyso** ($2.67 \mu\text{g mL}^{-1}$) in the presence of 20 μM of HClO and one biologically relevant molecule; note: (1–18: 200 μM , 19–20: 5.0 mM, 21–25: 200 μM) 1, Na^+ ; 2, K^+ ; 3, Ca^{2+} ; 4, Zn^{2+} ; 5, Co^{2+} ; 6, Cu^{2+} ; 7, Mg^{2+} ; 8, Fe^{3+} ; 9, Fe^{2+} ; 10, Ni^{2+} ; 11, Cl^- ; 12, SO_4^{2-} ; 13, NO_3^- ; 14, H_2PO_4^- ; 15, HPO_4^{2-} ; 16, PO_4^{3-} ; 17, SO_3^{2-} ; 18, NO_2^- ; 19, Cys; 20, GSH; 21, H_2O_2 ; 22, NO; 23, $^1\text{O}_2$; 24, HO; 25, *t*-BuO. $\lambda_{\text{ex}} = 399$ nm.

Fig. 2C shows the time response of the nanoprobe toward HClO. The F_{475}/F_{620} ratio changed after HClO was added into the **AIED-Lyso** solution for only one min, implying a rapid detection feature. Fig. 2D describes the effect of pH on the fluorescence intensity ratio (F_{475}/F_{620}) in the absence and presence of HClO. Without HClO, **AIED-Lyso** exhibits good stability from pH 3.0 to 9.0. In contrast, the addition of HClO (20 μM) induces an obvious enhancement of F_{475}/F_{620} from pH 3.0 to 6.0, suggesting the wide adaptive range of **AIED-Lyso** in acidic environments including lysosomes.

To evaluate the selectivity of **AIED-Lyso**, the F_{475}/F_{620} ratio of **AIED-Lyso** solution was examined in the presence of HClO or other biologically relevant molecules including anions, reactive oxygen species (ROS) and thiols (Fig. 3A). Except for HClO, other biologically relevant molecules did not induce obvious variation in F_{475}/F_{620} . Moreover, a negligible interfering effect on the F_{475}/F_{620} ratio of **AIED-Lyso** was found when HClO and other biologically relevant molecules were added together (Fig. 3B). All these results indicated that **AIED-Lyso** can selectively detect HClO in complex biological environments.

On the basis of the fluorescent response of **SA-C2-PCD** toward HClO, we anticipated that the strongly oxidized HClO can first oxidize the double bond between the SA-C2 moiety and pyrene moiety, then gradually hydrolyze into **SA-C2-CHO** and pyrene-CHO (Scheme S3, ESI[†]). The newly produced pyrene-CHO would form excimers in aqueous solution.²⁷ Therefore, the change in the F_{475}/F_{620} ratio can be linked to the response of **SA-C2-PCD** toward HClO, as shown in Fig. 2A. This proposed response mechanism was further confirmed by MS measurements. As expected, the mass peak at 730.4 m/z which corresponds to the nanoprobe **SA-C2-PCD** was replaced by mass peaks at 531.3 m/z and 231.1 m/z that correspond to **SA-C2-CHO** and pyrene-CHO, respectively (Fig. S12A and B, ESI[†]). These results supported the proposed reaction mechanism (Scheme S3, ESI[†]).

Imaging exogenous and endogenous HClO levels in living cells

We firstly evaluated the cytotoxicity of **AIED-Lyso** with representative HeLa cells (Fig. S13, ESI[†]). Positive results indicated minimal cytotoxicity and our **AIED-Lyso** should be suitable for intracellular HClO sensing.



Fig. 4 Images of HeLa cells co-incubated with (A) **AIED-Lyso** ($2.67 \mu\text{g mL}^{-1}$; $\lambda_{\text{ex}} = 405 \text{ nm}$, and $\lambda_{\text{em}} = 575\text{--}675 \text{ nm}$) and (B) LysoTracker Green (500 nM ; $\lambda_{\text{ex}} = 468 \text{ nm}$, and $\lambda_{\text{em}} = 500\text{--}520 \text{ nm}$); (C) the merged image of (A) and (B); (D) the intensity profile of ROI across HeLa cells from the red and green channels; and (E) the correlation plot of two channels. Scale bar: $20 \mu\text{m}$.

It has been well documented that small molecule-based probes with a morpholine moiety mainly accumulate in lysosomes;^{21,49} meanwhile, the intracellular uptake of nanoparticles is generally performed by the endocytosis strategy and they are often located in lysosomes.^{50–53} To increase the lysosomal targeting ability, we introduce morpholine groups onto the surface of the nanoparticles to sense HClO in lysosomes. To conduct the colocalization studies, HeLa cells were treated with both **AIED-Lyso** and LysoTracker Green (a commercial lysosomal tracker), and the results are given in Fig. 4.



Fig. 5 Fluorescence images of HeLa cells treated with **AIED-Lyso** ($2.67 \mu\text{g mL}^{-1}$) with the addition of 0 (control), $10 \mu\text{M}$, $20 \mu\text{M}$ and $30 \mu\text{M}$ HClO (from the left column to the right column). The ratio images were all obtained as $F_{\text{blue}}/F_{\text{red}}$. $\lambda_{\text{ex}} = 405 \text{ nm}$, $\lambda_{\text{em1}} = 425\text{--}525 \text{ nm}$ (blue channel), and $\lambda_{\text{em2}} = 575\text{--}675 \text{ nm}$ (red channel), scale bar = $20 \mu\text{m}$.

The red emission of **AIED-Lyso** fully overlapped with the green fluorescence of LysoTracker Green and the Pearson's R value was calculated to be 0.96 between our nanoprobe (shown in Red channel) and LysoTracker Green (shown in Green channel), which is far beyond the threshold of >0.5 required for correlation, indicating that **AIED-Lyso** mainly localized in lysosomes and can specifically monitor HClO in lysosomes.

AIED-Lyso was subsequently used for ratiometric imaging of exogenous HClO levels in HeLa cells. As shown in Fig. 5, HeLa cells treated with only **AIED-Lyso** provide strong red fluorescence and a low ratio of emission intensity (blue/red) (Fig. 5E, I and M). When the cells pre-treated with **AIED-Lyso** were exposed to different concentrations of HClO ($10 \mu\text{M}$, $20 \mu\text{M}$ and $30 \mu\text{M}$), the blue fluorescence was obviously enhanced (Fig. 5J, K and L), accompanied by a decrease in red fluorescence (Fig. 5F, G and H). Accordingly, the ratio of emission intensity (blue/red) is also significantly enhanced (Fig. 5N, O and P). These results suggest that **AIED-Lyso** can be applied for ratiometric imaging of exogenous HClO in living cells.



Fig. 6 Fluorescence microscopy images of RAW264.7 cells. The control groups (A, D and G) are incubated with **AIED-Lyso** ($2.67 \mu\text{g mL}^{-1}$) for 3 h. The cells in groups (B, E, and H) are first incubated with LPS ($1 \mu\text{g mL}^{-1}$) for 12 h, followed by coinubation with PMA ($1 \mu\text{g mL}^{-1}$) and **AIED-Lyso** ($2.67 \mu\text{g mL}^{-1}$) for 3 h. The cells in groups (C, F, and I) are first incubated with LPS ($1 \mu\text{g mL}^{-1}$) and NAC (2 mM) for 12 h, followed by coinubation with PMA ($1 \mu\text{g mL}^{-1}$) and **AIED-Lyso** ($2.67 \mu\text{g mL}^{-1}$) for 3 h. The ratio image is obtained as the ratio of blue fluorescence intensity to red fluorescence intensity ($F_{\text{blue}}/F_{\text{red}}$) (J, K and L). $\lambda_{\text{ex}} = 405 \text{ nm}$, $\lambda_{\text{em1}} = 425\text{--}525 \text{ nm}$ (blue channel), and $\lambda_{\text{em2}} = 575\text{--}675 \text{ nm}$ (red channel), scale bar = $20 \mu\text{m}$.

It was noted that the living cells were complicated systems, and the intracellular uptake of HClO was lower than the initial concentration. In addition, the intracellular reducing substances (e.g. GSH) can consume some of the HClO. Thus, a relatively high HClO concentration (10–30 μM) was applied to the cells in order to make the nanoprobe facile for ratiometric imaging of exogenous HClO levels in HeLa cells.

Finally, we imaged endogenous HClO in living cells. It has been well documented that endogenous HClO can be produced from RAW264.7 cells by treating them with lipopolysaccharide (LPS) and phorbol myristate acetate (PMA).^{54,55} As illustrated in Fig. 6, the RAW264.7 cells incubated with only **AIED-Lyso** showed bright red fluorescence with a low ratio of emission intensity (blue/red) (Fig. 6D, G and J). After stimulating with LPS and PMA, the RAW264.7 cells treated with **AIED-Lyso** revealed an obvious decrease in red fluorescence but a significant enhancement in blue fluorescence with a high ratio of emission intensity (blue/red) (Fig. 6E, H and K). To further confirm that the intracellular fluorescence emission changes were triggered by the endogenous HClO, *N*-acetylcysteine (NAC) was added as an effective ROS scavenger to reduce the cellular HClO level.^{56,57} As shown in Fig. 6F, I and L, when the RAW264.7 cells were incubated with LPS, PMA and NAC, the blue fluorescence was suppressed, whereas the red fluorescence was recovered to some extent. The above results demonstrated that **AIED-Lyso** can serve as a ratiometric nanoprobe to visualize the endogenously generated HClO in RAW264.7 cells.

Conclusions

In summary, a novel lysosome-targetable and ratiometric nanoprobe (**AIED-Lyso**) was rationally designed and synthesized for selectively sensing exogenous and endogenous HClO levels in live cells. The results indicated that **AIED-Lyso** possessed stable water dispersity, high selectivity, excellent long-term fluorescence stability (> 10 weeks), and good biocompatibility. Importantly, with the aid of the lysosomal targeting group, **AIED-Lyso** can mainly accumulate in lysosomes and then monitor the exogenous and endogenous HClO in live cells. The design of the lysosome-targetable and ratiometric fluorescent nanoprobe presented herein should provide a new avenue for exploring the function of HClO-associated diseases.

Conflicts of interest

The authors declare no competing financial interest.

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