

Cite this: *Polym. Chem.*, 2025, **16**, 3092

Adjusting the electroosmotic flow for CE separation of proteins by using poly(α -L-lysine)-based mixed polycationic/polyzwitterionic multilayer coatings†

Henry Frick,^{a,b} Laura Dhellemmes,^c Alisa Höchsmann,^a Laurent Leclercq,^{id} ^c Christian Neusüß,^{id} ^a Hervé Cottet,^{id} ^c and Norbert Schaschke,^{id} ^{*a}

Capillary electrophoresis (CE) is a robust, selective and highly efficient technique for the analysis of peptides and (intact) proteins. The anionic and/or hydrophilic character of the fused silica surface generally leads to protein adsorption by electrostatic interaction or H-bonding. Successive multiple ionic-polymer layer (SMIL) coatings are often used to limit adsorption and to improve the repeatability of migration times. Besides these adsorption phenomena, the electroosmotic flow (EOF) also has a strong influence on the resolution. Here, the frequently used and efficient cationic SMIL coating agent poly(α -L-lysine) (α -PLL) is modified for the systematic modulation of the EOF. In particular, by converting stepwise the ϵ -amino functions of this polycation into carboxamides, the total number of positive charges decreases, leading to reduced EOF. To simultaneously keep the analyte-surface interactions as small as possible, carboxylic acids bearing a zwitterionic functionality based structurally on a sulfobetaine motif were developed. Using the water-soluble and highly hydrolysis-resistant condensation agent 4-(4,6-dimethoxy-1,3,5-triazin-2-yl)-4-methylmorpholinium chloride (DMTMM), the degree of functionalization could be adjusted between 8 and 99%, depending on the applied stoichiometry. The obtained set of mixed polycationic/polyzwitterionic polymers based on the α -PLL scaffold were investigated as the outermost layers of the SMIL coatings, clearly showing that the EOF decreases depending on the degree of functionalization while maintaining high efficiency.

Received 6th February 2025,
Accepted 28th May 2025

DOI: 10.1039/d5py00122f

rsc.li/polymers

Introduction

For the analysis of complex mixtures of peptides or proteins, capillary electrophoresis (CE) is a frequently used separation technique. CE has a variety of distinct advantages compared to liquid chromatography: high separation efficiencies, small sample volumes and fast analysis times.^{1,2} The separation mechanism in CE is orthogonal to reversed phase-high performance liquid chromatography (RP-HPLC). While the separation in RP-HPLC is primarily based on the hydrophobic interaction between the analyte and the stationary phase, in CE, the different electrophoretic mobilities of the analytes are exploited for separation.³ CE is also an environmentally

friendly technique. Only small amounts of organic chemicals and reagents are required. In particular, the coupling of CE with mass spectrometry (MS) analysis makes it a powerful tool in the field of protein analytics, which has been highlighted in a series of studies.⁴⁻⁶

The anionic and/or hydrophilic character of the fused silica surface generally leads to protein adsorption by electrostatic interaction or H-bonding.⁷ Today, the best way to circumvent the protein adsorption for CE-MS analysis is the coating of the capillary. Based on the mode of attachment, one can distinguish between dynamic and permanent capillary wall coatings.^{8,9} Besides neutral coatings,¹⁰ mostly cationic coatings as a single layer¹¹⁻¹⁶ or in multiple ionic polymer layers (SMILs)^{17,18} have been used for highly efficient separation of peptides and proteins.

Katayama *et al.* have developed SMIL coatings in 1998.¹⁹ First, the silanol groups are deprotonated with NaOH. Then, a polycation is adhered to the inner capillary surface, followed by a polyanion, which is adsorbed by electrostatic interaction with the cationic layer. These steps can be repeated several times. Thus, SMIL coatings are generated by alternating poly-

^aFaculty of Chemistry, Aalen University, Beethovenstraße 1, D-73430 Aalen, Germany. E-mail: norbert.schaschke@hs-aalen.de

^bDepartment of Chemistry, Bielefeld University, Bielefeld, Germany

^cIBMM, University of Montpellier, CNRS, ENSCM, Montpellier, France

† Electronic supplementary information (ESI) available: NMR and HR-MS spectra of compounds 4-9 and 1H-1H NOESY NMR spectrum of polymer 12e. See DOI:

<https://doi.org/10.1039/d5py00122f>

cationic and polyanionic layers with either a negatively or positively charged surface as the outermost layer. A broad range of polycations have been developed, such as poly(diallyldimethylammonium) chloride (PDADMAC),²⁰ polyarginine,²¹ chitosan,^{22,23} ϵ -poly(L-lysine) (ϵ -PLL)¹⁸ and α -PLL.²⁴ Among the possible polyanions, dextran sulfate (DS)²⁵ and poly(sodium-4-styrenesulfonate) (PSS)¹⁸ are often used. Acidic separation conditions are often preferred in CE-MS to facilitate ionization under positive electrospray conditions. In this case, the outermost layer should be positively charged, typically applying three or five homopolymer layers.^{26,27} In the last few years, different parameters, such as the number and the thickness of the polyelectrolyte layers, were investigated for CE and CE-MS separation of proteins using SMIL coated capillaries.^{26,28–32} Besides reduced adsorption, the coating is important to control the electroosmotic flow (EOF) in order to achieve the best resolution by counterbalancing the analyte mobility.³³

In this work, we present novel mixed polycationic/polyzwitterionic polymers used as the outermost layer interacting with the protein analytes (Fig. 1). Sulfobetaine moieties with their anti-adsorptive properties have a net charge of zero and are introduced *via* an amide bond into the polycation. Thus, the adjustment of the EOF is achieved by replacing a defined number of positive charges of the polycation by introducing zwitterionic building blocks. The best polycations for SMIL coatings, however, are mostly polymers containing quaternary or tertiary amino groups and thus cannot be derivatized with the typical coupling reagents. Therefore, this work is focused on the functionalisation of α -PLL, one of the few polymers with primary amino groups and good properties as a cationic layer for SMIL coatings.^{18,24,26}



Fig. 1 Schematic representation of the migration of a peptide or protein with a positive net charge in a CE-MS setup. (A) Standard 5-layer SMIL capillary coating. (B) SMIL coating with an α -PLL-based outermost cationic layer. The strength of the EOF can be adjusted by reducing the number of positive charges using the ϵ -amino function for functionalization with a zwitterionic sulfobetaine motif. The thus-resulting improved resolution is due to a decreased apparent mobility of the peptide or protein towards the anode. The length of the arrows represents the strength of the EOF (light blue), the magnitude of analyte mobility (orange), and the magnitude of the resulting apparent mobility (light gray).

Experimental

Materials

N,N-Dimethylglycine ethyl ester, 4-(dimethylamino)butyric acid hydrochloride, *N*-methylmorpholine (NMM), and DMTMM were purchased from Tokyo Chemical Industry (TCI). Methyl 3-(dimethylamino)propionate was purchased from Sigma-Aldrich. 1,3-Propanesultone was purchased from Thermo Fisher Scientific. α -PLL hydrochloride (average $M_w = 66,000$ Da) with a polydispersity index (PDI) between 1.0 and 1.2 was purchased from Alamanda Polymers. 2-[4-(2-Hydroxyethyl)piperazine-1-yl]ethane sulfonic acid (HEPES) and acetic acid were purchased from Sigma-Aldrich (Saint-Quentin Fallavier, France). The model proteins, carbonic anhydrase I from human erythrocytes (CA, purity not indicated by the supplier), myoglobin from equine skeletal muscle (Myo, purity $\geq 95\%$), ribonuclease A from bovine pancreas (RNase A, purity $\geq 60\%$), β -lactoglobulin A from bovine milk (β -lac A, purity $\geq 90\%$), and lysozyme from chicken egg white (Lyz, purity $\geq 90\%$) were purchased from Sigma-Aldrich (Saint-Quentin Fallavier, France). Ultrapure water was obtained using a MilliQ system from Millipore (Molsheim, France). PDADMAC ($M_r 4 \times 10^5 - 5 \times 10^5$, 20% w/w in water) was purchased from Sigma-Aldrich (Lyon, France). PSS ($M_r 7 \times 10^4$) was purchased from Acros Organics (Geel, Belgium). All other solvents and chemicals were obtained from commercial sources and used as received unless otherwise noted. Dialysis tubes (D-Tube™ Dialyzers, MWCO 3.5 kDa) were purchased from Merck KGaA. LoBind® tubes were purchased from Eppendorf.

NMR spectroscopy

NMR spectra were recorded using a Bruker DRX 500 spectrometer at room temperature. Chemical shifts are reported in parts per million (ppm) calibrated using a residual non-deuterated solvent as the internal reference. All samples were dissolved in D_2O (99.95% deuteration).

CE-MS analysis of zwitterions 4–9

CE-MS analysis was performed using an Agilent 7100 CE coupled to a compact mass spectrometer from Bruker. Separation conditions are provided in the ESI.†

Size exclusion chromatography

SEC analyses were performed using an Agilent 1260 quaternary pump at a flow rate of 0.5 mL min^{-1} . 150 mM phosphate buffer (pH 6.8) with 750 mM NaCl was used as the mobile phase and eluted using a $7.8 \times 300 \text{ mm}$ Agilent AdvanceBio SEC column (with a guard), and the elution of the polymers was monitored with a refractive index detector (RID). The temperature of the column and the RID was set at $35 \text{ }^\circ\text{C}$.

Capillary electrophoresis

Electrophoretic separations were performed using an Agilent 7100 CE system. Fused silica capillaries of $50 \text{ }\mu\text{m}$ in diameter and 60 cm total length (51.5 cm to the detector) were used. For most of the coatings, the electric fields applied during the sep-

arations were between -30 kV and -10 kV. Negative polarity was used due to the strong anodic (reversed) EOF and counter-electroosmotic migration of the positively charged proteins. Positive polarity was used for the coatings with a low EOF magnitude compared to protein electrophoretic mobilities. The capillary was flushed for 5 min with the BGE between each run. First, 0.001% DMF in 2 M acetic acid was injected for 3 s at 30 mbar. DMF was used as a neutral marker for EOF determination. The EOF is given as an average obtained on 25 runs \pm one standard deviation. Then, the protein mixture was injected for 6 s at 30 mbar. The cassette and tray temperatures were set at 25 °C and the detection wavelength was 214 nm. Plate numbers per meter N/l were calculated using CEval software available at [<https://echmet.natur.cuni.cz/>]³⁴ based on the following equation:

$$N/l = \frac{5.54}{l} \left(\frac{t_m}{\delta} \right)^2 \quad (1)$$

where l is the capillary length to the detector, N is the plate number, t_m the migration time, and δ the width at half height of each peak.

Capillary coating procedure

The background electrolyte (BGE) was 2 M acetic acid (pH 2.2), while the construction buffer was made from 20 mM HEPES and 10 mM NaOH (pH 7.4). PDADMAC, PSS and the polyzwitterion solutions were made by dissolving each coating agent in 3 g L⁻¹ of HEPES buffer at least one night before the first use. The polycation and zwitterion solutions were kept in the freezer while the polyanions could be kept in the fridge. The protein mixture was prepared from the stock solutions of proteins stored individually in water at a higher concentration. 100 μ L aliquots were prepared by mixing 10 μ L of each protein solution with 50 μ L of 2 M acetic acid so that each protein was at a final concentration of 0.2 g L⁻¹ in BGE. The protein sample was placed in an oven for 30 min at 37 °C in order to reach more stable protein conformations.³⁵ Before coating, the silica capillary was preconditioned by flushing with 1 M NaOH for 10 min and then with water for 5 min and HEPES for 10 min. Next, PDADMAC was flushed for 7 min, HEPES for 3 min, PSS for 7 min, HEPES for 3 min, and so on, until 4 layers were formed. The polyzwitterion was flushed for 7 min as the outermost layer. Then, HEPES was flushed for 3 min, and after 5 min, ultrapure water was flushed for 3 min and BGE for 10 min. Analyses were performed after standing for 10 min. All flushes were performed at 930 mbar. SMILs are designated as (A/B)_{*i*}, where A refers to the polycation, B refers to the polyanion, and *i* refers to the number of bilayers.

Synthesis of zwitterions

Methyl 4-(dimethylamino)butanoate (3). Purified thionyl chloride (13 mL, 178 mmol) was added to dry methanol (50 mL) over a period of 10 minutes at -10 °C. Then 4-(dimethylamino)butyric acid hydrochloride (8.4 g, 50 mmol) was added and stirred at room temperature for 24 h for complete

dissolution. The solvent was evaporated under reduced pressure and dry methanol (25 mL) was added to completely dissolve the crude reaction product. MTBE (125 mL) was added to the solution and the product precipitated as a white solid. The product was filtered, washed with MTBE and dried under vacuum. Yield: 8.7 g (97%); ¹H NMR (500 MHz, D₂O): δ = 1.91–1.95 (m, 2 H), 2.41 (t, J = 7.0 Hz, 3 H), 2.79 (s, 6 H), 3.08 (t, J = 7.6 Hz, 2 H), 3.60 (s, 3 H); ¹³C NMR (125 MHz, D₂O): 18.9, 29.9, 42.3, 51.9, 56.3, 174.8.

To generate the free amine, 8.0 g of hydrochloride was partitioned between EtOAc and 2 M NaOH and extracted again with a copious amount of EtOAc. The combined organic layers were washed with brine and dried over MgSO₄. The solvent was removed under reduced pressure to give a colourless liquid which was used without purification (yield: 3.7 g, 57%).

General procedure for the synthesis of zwitterionic esters 4, 5 and 6. In a 100 mL round-bottom flask with a stir bar, the methyl or ethyl ester-protected ω -dimethylamino carboxylic acid (4.0 g) was dissolved in acetone (75 mL). To this stirred solution of 1,3-propanesultone (1.1 eq.) was added. The mixture was heated to 50 °C overnight. The product was obtained as a white precipitate, filtered off, washed with a small amount of acetone, and dried under vacuum.

3-[(2-Ethoxy-2-oxoethyl)(dimethyl)azaniumyl]propane-1-sulfonate (4). Yield: 7.4 g (94%); ¹H NMR (500 MHz, D₂O): δ = 1.19 (t, J = 7.2 Hz, 3 H), 2.14 (m, 2 H), 2.87 (t, J = 7.1 Hz, 2 H), 3.21 (s, 6 H), 3.63 (m, 2 H), 4.20 (q, J = 7.2 Hz, 4 H); ¹³C NMR (125 MHz, D₂O): 12.5, 17.7, 46.6, 51.2, 61.0, 62.9, 63.2, 164.5; CE-MS (ESI) m/z calcd for [M + H]⁺ 254.098, found [M + H]⁺ 254.107; m/z calcd for [2M + H]⁺ 507.196, found [2M + H]⁺ 507.205; m/z calcd for [3M + Na]⁺ 782.283, found [3M + Na]⁺ 782.282.

3-[(3-Methoxy-3-oxopropyl)dimethylamino]propane-1-sulfonate (5). Yield: 7.5 g (93%); ¹H NMR (500 MHz, D₂O): δ = 2.12 (m, 2 H), 2.88 (m, 4 H), 3.02 (s, 6 H), 3.38 (m, 2 H), 3.59 (t, J = 7.5 Hz, 2 H), 3.63 (s, 3 H); ¹³C NMR (125 MHz, D₂O): 17.6, 27.0, 46.6, 50.1, 52.3, 58.7, 62.0, 171.3; CE-MS (ESI) m/z calcd for [M + H]⁺ 254.098, found [M + H]⁺ 254.107; m/z calcd for [2M + H]⁺ 507.196, found [2M + H]⁺ 507.205; m/z calcd for [3M + Na]⁺ 782.283, found [3M + Na]⁺ 782.282.

3-[(4-Methoxy-4-oxobutyl)dimethylammonio]propane-1-sulfonate (6). Yield: 7.4 g (91%); ¹H NMR (500 MHz, D₂O): δ = 1.96 (m, 2 H), 2.10 (m, 2 H), 2.40 (t, J = 7.0 Hz, 2H), 2.86 (t, J = 7.1 Hz, 2 H), 3.00 (s, 6 H), 3.25 (m, 2 H), 3.36 (m, 2 H), 3.59 (s, 3 H); ¹³C NMR (125 MHz, D₂O): 16.9, 17.5, 29.3, 46.7, 50.1, 51.8, 61.6, 62.3, 174.4; CE-MS (ESI) m/z calcd for [M + H]⁺ 268.121, found [M + H]⁺ 268.123; m/z calcd for [2M + H]⁺ 535.243, found [2M + H]⁺ 535.235; m/z calcd for [3M + Na]⁺ 824.364, found [3M + Na]⁺ 824.328.

General procedure for the synthesis of zwitterionic building blocks 7, 8, and 9. In a 100 mL round bottom flask, the methyl or ethyl ester-protected ω -dimethylamino carboxylic acid (4.0 g) was dissolved in 6 M HCl (60 mL) and heated to 70 °C. After 24 h, the reaction was completed and the solvent was evaporated under reduced pressure to give a white solid and it was used without purification.

3-((Carboxymethyl)dimethylammonio)propane-1-sulfonate (7). Yield: 4.5 g (96%); ^1H NMR (500 MHz, D_2O): δ = 2.12 (m, 2 H), 2.86 (t, J = 7.1 Hz, 2 H), 3.18 (s, 6 H), 3.61 (m, 2 H), 4.11 (s, 2 H); ^{13}C NMR (125 MHz, D_2O): 17.7, 46.7, 50.9, 61.3, 63.0, 166.6; CE-MS (ESI) m/z calcd for $[\text{M} + \text{H}]^+$ 226.074, found $[\text{M} + \text{H}]^+$ 226.076; m/z calcd for $[2\text{M} + \text{H}]^+$ 451.148, found $[2\text{M} + \text{H}]^+$ 451.143.

3-((2-Carboxyethyl)dimethylammonio)propane-1-sulfonate (8). Yield: 3.6 g (94%); ^1H NMR (500 MHz, D_2O): δ = 2.13 (m, 2 H), 2.86 (m, 4 H), 3.02 (s, 6 H), 3.39 (m, 2 H), 3.57 (t, J = 7.2 Hz, 2 H); ^{13}C NMR (125 MHz, D_2O): 17.6, 27.1, 46.6, 50.1, 58.9, 62.0, 172.6; CE-MS (ESI) m/z calcd for $[\text{M} + \text{H}]^+$ 240.090, found $[\text{M} + \text{H}]^+$ 240.091; m/z calcd for $[2\text{M} + \text{H}]^+$ 479.180, found $[2\text{M} + \text{H}]^+$ 479.172; m/z calcd for $[3\text{M} + \text{H}]^+$ 718.270, found $[3\text{M} + \text{H}]^+$ 718.251.

3-((3-Carboxypropyl)dimethylammonio)propane-1-sulfonate (9). Yield: 3.3 g (86%); ^1H NMR (500 MHz, D_2O): δ = 1.97 (m, 2 H), 2.12 (m, 2 H), 2.39 (t, J = 7.0 Hz, 2 H), 2.87 (t, J = 7.1 Hz, 2 H), 3.01 (s, 6 H), 3.26 (m, 2 H), 3.38 (m, 2 H); ^{13}C NMR (125 MHz, D_2O): 16.9, 17.5, 29.4, 46.7, 50.1, 61.7, 62.4, 175.8; CE-MS (ESI) m/z calcd for $[\text{M} + \text{H}]^+$ 254.212, found $[\text{M} + \text{H}]^+$ 254.107; m/z calcd for $[2\text{M} + \text{H}]^+$ 507.196, found $[2\text{M} + \text{H}]^+$ 507.204; m/z calcd for $[3\text{M} + \text{H}]^+$ 760.636, found $[3\text{M} + \text{H}]^+$ 760.318.

General procedure for the functionalization of α -PLL

In a 5 mL LoBind® tube, α -PLL hydrochloride (10 mg) was dissolved in $\text{H}_2\text{O}/\text{MeOH}$ (800 μL , 1:1, v:v). In another 2 mL LoBind® tube, the zwitterion was dissolved in $\text{H}_2\text{O}/\text{MeOH}$ (200 μL , 1:1, v:v). After the addition of 2 equivalents of NMM, the reaction mixture was stirred for 5 min at room temperature. The deprotonated zwitterion and the coupling reagent DMTMM were added to the α -PLL solution in the desired stoichiometric ratio (see Table 1) under vigorous stirring. The reaction was allowed to proceed overnight at room temperature, followed by dialysis against 0.5 M sodium perchlorate for 48 h. The dialyzed polymer was lyophilized for 48 h.

Results and discussion

Synthesis of zwitterionic building blocks

The zwitterionic motif consists of a quaternary amine and a sulfonic acid function (Fig. 2). The difference between these molecules lies in the length of the alkyl chain between the carboxylic acid and the quaternary amino function. Such molecules with zwitterionic characters have been known for a long time in biochemistry as buffer agents.³⁶ Also for SMIL coatings, polyzwitterionic polymers are used to reduce the adsorption of proteins onto the wall of fused silica capillaries.^{37–39}

Sultones, cyclic esters of sulfonic acids, are versatile substances that react under mild conditions with a variety of nucleophiles under ring-opening, yielding the corresponding alkylsulfonic acids.⁴⁰ We took advantage of this particular reactivity of sultones for the synthesis of zwitterions 7–9 (Scheme 1). Due to severe solubility problems of the carboxylic

Table 1 Mixed polycationic/polyzwitterionic polymers **10a–g**, **11a–c**, and **12a–e** obtained by the functionalization of the α -PLL scaffold

Entry	Polymer ^a	Applied equiv. of zwitterion/DMTMM/NMM (1 : 1 : 2)	Degree of functionalization ^b (%)	N/I^c (10^3 plates per m)
1	10a	0.5	8	316
2	10ba	1.0	17	—
3	10bb	1.0	19	193
4	10c	1.5	29	202
5	10d	1.75	38	234
6	10e	2.0	50	182
7	10f	3.0	71	Not applicable
8	10g	4.0	88	104
9	11a	0.5	17	—
10	11b	1.0	25	173
11	11c	2.0	25	—
12	12a	0.25	11	316
13	12b	0.5	31	188
14	12c	0.75	51	293
15	12da	1.0	60	—
16	12db	1.0	63	49
17	12e	2.0	99	44
18	PDADMAC	—	—	367

^a Reactions were performed twice, yielding almost the same degree of functionalization, respectively. Exemplary, this is shown for polymers **10b** (a low degree of functionalization) and **12d** (a high degree of functionalization). ^b Determined by ^1H NMR (500 MHz) using D_2O as a solvent. ^c Separation efficiencies of Lyz obtained on PDADMAC/PSS SMILs with different final layers. Data are shown for -10 kV runs except for polymer **10g**, which is shown for -20 kV.



Fig. 2 Chemical structures of zwitterions 7–9 used for the functionalization of α -PLL (7: n = 1, 8: n = 2, and 9: n = 3).

acid salts of these ω -dimethylamino acids in acetone and, furthermore, to avoid side reactions originating from the nucleophilic character of the carboxylate function during the quaternization reaction,⁴¹ the corresponding methyl or ethyl esters 1–3 were utilized as nucleophiles. The ring-opening reactions of 1,3-propanesultone with nucleophiles 1–3 result directly in the formation of *tert*-alkylammonium sulfonates 4–6, respectively. Furthermore, it turned out that these *tert*-alkylammonium sulfonates 4–6 precipitate from acetone during the reaction and, thus, can be easily isolated by filtration as pure compounds in excellent yields.

The needed methyl ester 3 was prepared from 4-(dimethylamino)butyric acid hydrochloride. For this, the amino acid was quantitatively converted into its corresponding methyl ester in the presence of methanol and an excess amount of SOCl_2 .⁴² Eventually, by treating the thus obtained hydrochloride with 2 M NaOH, the free amine 3 could be isolated by extracting it into ethyl acetate.

The three synthesized zwitterionic molecules 4–6 are protected as esters. Thus, to generate free carboxylic acid func-



Scheme 1 Synthesis of zwitterions 7–9. Reagents and conditions: (a) acetone, 50 °C, overnight, 4: 94%, 5: 93%, 6: 91%; (b): 6 M HCl, 70 °C, 24 h, 7: 96%, 8: 94%, 9: 86%.

tions, first of all, saponification with NaOH was investigated. Due to serious difficulties in isolating fully water-soluble acids 7–9 free from salts after the saponification process, we decided to employ acid hydrolysis (Scheme 1). Using 6 M HCl at 70 °C, cleavage of the esters proceeded smoothly and the zwitterionic building blocks 7–9 could be isolated free from by-products without any problems as confirmed by means of CE-MS and NMR spectroscopy. After the successful synthesis of the zwitterionic molecules 7–9, we could proceed with the preparation of mixed cationic/zwitterionic polymers by derivatization of the ϵ -amino functions of the lysine residues of α -PLL.

Functionalization of α -PLL with zwitterions 7–9

Due to the fact that both α -PLL and zwitterions 7–9 are almost exclusively water soluble, it was necessary to find a coupling chemistry for the functionalization of the polymer that shows a high degree of water tolerance. Among the coupling reagents typically used for amide bond formation that combine both water solubility and a high extent of hydrolysis resistance, the recently developed 4-(4,6-dimethoxy-1,3,5-triazin-2-yl)-4-methylmorpholinium chloride (DMTMM) is most suitable.^{43–45} Compared to the 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide-hydrochloride (EDC)/*N*-hydroxysuccinimide (NHS) or *N*-hydroxysulfo-succinimide (sulfo-NHS) system, which is frequently used for labelling or crosslinking peptides, proteins or biomolecular probes in water,^{46–48} DMTMM has significantly better stability in water. While EDC has a half-life of 3.9 h in water at a pH of 5.0,⁴⁹ DMTMM is 24 h stable in water without any decomposition by demethylation or hydrolysis.⁴³ Moreover, DMTMM is more efficient and easier to handle than the EDC/NHS system.^{45,50,51}

Initially, for the functionalization of α -PLL, a pre-activation approach was chosen. For this, the zwitterions 7–9 were transformed with DMTMM into the corresponding triazine esters and then reacted with the completely deprotonated α -PLL, respectively. It turned out that under these reaction con-

ditions, a gel-like solid was formed immediately after the addition of the active ester, preventing the formation of any condensation product. This behaviour of α -PLL in basic aqueous solutions at higher concentrations is in good agreement with previous observations made by Hull *et al.*⁵² and later confirmed by Dos *et al.*,⁵³ both of which investigated α -PLL by ¹⁵N NMR spectroscopy. They found that α -PLL forms a gel-like solid which is built up by α -helices that assemble into higher aggregates in the pH range from 9.2 to 11.4.

To circumvent these severe solubility problems with α -PLL arising under basic aqueous conditions, a stepwise liberation of the ϵ -amino function in terms of a titration approach was selected (Scheme 2). For this, the condensing agent DMTMM provides the prerequisites. In this one-pot approach, the carboxylates of zwitterions 7–9 which were obtained upon treatment with two equivalents of NMM and α -PLL hydrochloride were dissolved in MeOH/H₂O (v/v, 1 : 1) in the desired stoichiometric ratio to adjust the degree of functionalization, followed by DMTMM. Under these conditions, first of all, the carboxylates of zwitterions 7–9 are readily converted by DMTMM into their corresponding triazine esters. Along the formation of this active ester, NMM acts as a leaving group and deprotonates *in situ* in a stepwise manner the ϵ -amino functions of α -PLL hydrochloride, which then react immediately with the active ester to give carboxamides. Thus, under these one-pot reaction conditions, the coupling reaction with DMTMM proceeds completely under neutral conditions and no additional base is necessary to deprotonate the ϵ -amino function of α -PLL hydrochloride. Therefore, one does not run into aggregation problems caused by the helix formation of α -PLL. The initial attempts to purify the thus obtained mixed polycationic/polyzwitterionic polymers by dialysis were not successful. Even after 48 h by dialysis against 0.001 M hydrochloric acid or 1 M acetate buffer, the excess of zwitterion could not be removed completely. To break up the strong ionic interactions between the non-functionalized protonated ϵ -amino function of α -PLL and the sulfonate residues of the excess zwitterions, only dialysis for 48 h against sodium perchlorate was successful.

This one-pot condensation approach between the ϵ -amino groups of α -PLL and the carboxyl groups of zwitterionic molecules 7–9 yields a set of mixed polycationic/polyzwitterionic polymers with different degrees of functionalization (10a–g, 11a–c, and 12a–e), which are summarized in Table 1. The degree of functionalization was determined from the ratio of the integrals of the signals of the α -CH protons of the lysine residues of α -PLL and the α -CH₂ protons of the respective spacer amino acid (degree of functionalization = $[I(\alpha\text{-CH}_2\text{ X}) / (I(\alpha\text{-CH Lys}) \times 2)] \times 100\%$, X = Gly, β -alanine, and γ -aminobutyric acid) by ¹H NMR spectroscopy. The obtained data clearly show that the achieved degree of functionalization depends on both the applied excess of the activated zwitterion for the coupling reaction and the length of the respective spacer amino acid. In particular, in the case of zwitterion 7, the degree of functionalization increases stepwise from 8% (Table 1, entry 1, polymer 10a) to 88% (Table 1, entry 8, polymer 10g). Similar results were obtained with the zwitterion



Scheme 2 Functionalization of α -PLL. Reagents and conditions: (a): Zwitterions 7–9/DMTMM/NMM (1 : 1 : 2; 0.25–4 equiv.); $\text{H}_2\text{O}/\text{MeOH}$ (1 : 1, v : v), rt, overnight and (b): dialysis against NaClO_4 , 48 h.

9 and to a lesser extent with the zwitterion 8. The expected influence of the length of the spacer amino acid on the functionalization efficiency shows the comparison of zwitterions 7 (spacer: glycine) and 9 (spacer: γ -aminobutyric acid) clearly. While 4.0 equiv. of activated zwitterion 7 lead only to a functionalization of 88% (Table 1, entry 8), 2.0 equiv. of activated zwitterion 9 are completely sufficient for an almost complete functionalization of α -PLL (Table 1, entry 17). Surprisingly, the activated zwitterion 8 (spacer: β -alanine) shows in comparison with the activated zwitterions 7 and 9 a different functionalization behaviour. It turned out that the degree of functionalization stops at 25% (Table 1, entry 10 vs. entry 11).

Characterization of the α -PLL-based mixed polycationic/poly-zwitterionic polymers by NMR spectroscopy and SEC

Due to the fact that an almost complete coverage of the degree of side chain functionalization of α -PLL was obtained using the zwitterion 9 for the derivatisation step, we chose this series of polymers (compounds 12a–12e) to investigate in detail the impact of the functionalization on the structure of the α -PLL scaffold by ^1H NMR spectroscopy. For this, we selected α -PLLs with 31% (12b), 63% (12db) and 99% (12e) side-chain functionalizations as prominent representatives. Fig. 3 shows the ^1H NMR spectra of these polymers. Based on a combination of COSY and NOESY spectra, all signals in the ^1H NMR spectra could be assigned. First of all, the inspection of the spectra shows that the complexity reflecting the fact that the functionalization occurs randomly decreases with increasing degree of the functionalization of the ϵ - NH_2 functions of the side chains of α -PLL from 12b over 12db to 12e as the end

point which represents a completely functionalized species. A measure for the increasing degree of functionalization are the ϵ - CH_2 protons of the lysine side chains with a chemical shift at 3.23 ppm. The intensity of this signal decreases from 12b to 12db and has disappeared completely in the case of 12e. Upon transformation of the ϵ - NH_2 function into the corresponding carboxamide, the signal of the ϵ - CH_2 protons of the lysine side chains shifts towards 3.35 ppm. Expectedly, for the complete functionalization, the signal of ϵ - CH_2 near the free amino function at 3.23 ppm disappears completely and only the new signal at 3.35 ppm was observed.

Because of the difficulties in removing the excess amount of 9 used in the functionalization step (*vide supra*) and thus to completely exclude non-covalent interactions between the ϵ - NH_2 functions of the side chains of α -PLL and the sulfonate residues of 9 and to corroborate the covalent amide linkage, a 2D-NOESY experiment of 12e was carried out. As expected for an amide linkage, the NOE between the α - CH_2 GABA protons at 2.50 ppm of the zwitterionic segment and the ϵ - CH_2 protons of the lysine side chain at 3.34 ppm could be detected (see the ESI†).

In addition to the characterization of the functionalized α -PLLs 12b, 12db, and 12e by 1D and 2D ^1H NMR, size exclusion chromatography (SEC) was also performed. We used both aqueous acetate buffer (1 M, pH 4.65) and aqueous phosphate buffer (150 mM, pH 6.8) as eluents with different concentrations of sodium chloride as additive which were demonstrated in the literature as suitable eluents for the SEC analysis of α -PLL.^{54,55} Surprisingly, when acetate buffer (1 M, pH 4.65) was used for the elution of the functionalized polymers, no peaks were detected. Based on these findings, phosphate



Fig. 3 ^1H NMR spectra (500 MHz, D_2O) of α -PLL functionalized with zwitterion 9. (A) Structures of polymers 12a–12e, $X = 44$ –396. (B) 31%, (C) 63%, and (D) 99% functionalization.

buffer (150 mM, pH 6.8) with different concentrations of sodium chloride was used. Interestingly, with low concentrations of sodium chloride (250 mM and 400 mM), only α -PLLs with a high degree of side chain substitutions (12db: 63% and 12e: 99%) could be eluted as very broad peaks, probably due to their strong interactions with the SEC phase. With higher salt concentrations (750 mM), the interaction with the SEC matrix could be further reduced and polymer 12b with 31% functionalization could also be eluted (Fig. 4). It turned out that the elution time decreases from 11.8 min in the case of 12b to 11.2 min in the case of 12e with increasing degree of functionalization.

Polyzwitterionic coatings in CE

Previous work has shown that very high separation efficiencies could be obtained with $(\text{PDADMAC}/\text{PSS})_{2.5}$ coatings under optimized conditions.³⁵ Nevertheless, the plate height still appeared to increase slightly with the electric field, an effect that has been attributed to analyte adsorption and coating inhomogeneity.⁵⁶ Furthermore, the best resolution in CE is obtained when the EOF is close to the mobility of the proteins of interest along with minimized protein adsorption. Therefore, we applied the novel mixed zwitterionic/cationic polymers as a final layer in a 5-layer SMIL approach, as shown in Fig. 1.

Protein separations were carried out on each modified SMIL, leading to the plate numbers provided in Table 1. The classical $(\text{PDADMAC}/\text{PSS})_{2.5}$ coating led to the most efficient separations, followed closely by polymers 10a and 12a. Overall, the polymers with low percentages of derivatization lead to satisfactory efficiencies, while the ones that are highly derivatized (above 60%) result in much poorer performance. Fig. 5 shows the impact of the percentage of α -PLL derivatization on the EOF. The EOF decreases as the amount of zwitterion present in the coating increases, which is coherent since the overall

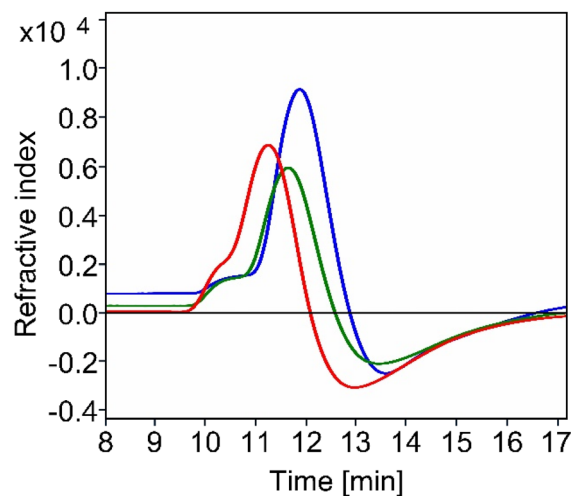


Fig. 4 SEC analysis of the functionalized α -PLLs 12b (●), 12db (●), and 12e (●). Eluent: 150 mM phosphate buffer with 750 mM NaCl at pH = 6.8; flow rate: 0.5 mL min^{-1} ; temperature: $35 \text{ }^\circ\text{C}$.



Fig. 5 Impact of % derivatization of polymers used as the outermost layer of SMIL coatings on the EOF magnitude. Error bars are \pm one standard deviation on 25 runs (except for % derivatization above 50%, which were on 5 runs). Experimental conditions: (PDADMAC/PSS)₂ coatings terminating with different polyelectrolytes. Capillary: 60 cm (51.5 cm to the detector) \times 50 μ m I.D. BGE: 2 M acetic acid, pH 2.2. Flushed before each run: BGE 1 bar, 5 min. Hydrodynamic injection: 30 mbar, 6 s. Sample mixture: 0.2 g L⁻¹ of each protein in BGE. Hydrodynamic co-injection of 0.01% DMF in BGE: 30 mbar, 3 s. Temperature: 25 °C. The EOF magnitude is given as the absolute value of the EOF mobility expressed in Tiselius units (10⁻⁹ m² V⁻¹ s⁻¹). For the coating procedure, see the Experimental section.

surface charge decreases. The type of polymer, **10** to **12**, does not seem to have much influence on both the separation efficiency and the magnitude of the EOF. The 71% derivatized polymer, **10f**, was not able to analyze any proteins because it generated a too low EOF to transport the proteins to the detector. Similarly, the two most derivatized polymers, **10g** and **12e**, led to EOF magnitudes that were lower than the proteins' electrophoretic mobilities, and so the separations had to be carried out with a positive polarity, while all the other coatings were used with a negative polarity.

Coatings containing inner layers of polyelectrolytes instead of PDADMAC (as in (polyelectrolyte/PSS)_{2.5}) were also tested but resulted in an unstable film and poor migration time repeatability (data not shown). When the polyelectrolyte was used in two layers with PDADMAC as the first layer (as in PDADMAC₁(PSS/polyelectrolyte)₂), the stability improved and the separation performance was similar to those of (PDADMAC/PSS)₂polyelectrolyte₁ coatings, with small modifications in the EOF. This is coherent with studies that find that the impact of the innermost and outermost layers is more important than that of the intermediate layers of a SMIL.⁵⁷ These coatings were not further investigated and only the results obtained with SMILs terminating with a layer of polyelectrolyte were included in this work.

Therefore, introducing polymers with different percentages of derivatization allows for the precise control of the EOF. A similar approach using polycations modified with poly(ethyl-

ene glycol) (PEG) as the outermost layer of SMIL coatings was recently presented, enabling highly suppressed EOF as low as 2 TU and good separation efficiency (200 \times 10³ plates per m for Lyz).²⁴ The polyelectrolytic coating with up to 50% derivatization could reduce the EOF by just 4 TU but led to more efficient separation (Table 1). Nevertheless, this is already interesting for certain applications, namely in CE-MS, where better resolution can be achieved with even a slightly lower EOF.⁵⁸ Polycations leading to reduced EOFs such as a quaternized diethylaminoethyl dextran (DEAEDq) are sought after for use in CE-MS applications. Indeed, a DEAEDq/poly(L-lysine citramide) (PLC) SMIL generating an EOF of -43.3 TU was able to separate glycoforms of ribonuclease B.¹⁸ Hence, SMILs modified with polyelectrolytes offer an interesting alternative to these methods.

Conclusions

Based on α -PLL, which is frequently used as the outermost cationic layer in SMIL coatings to suppress analyte-surface interactions in CE separations of proteins, a series of mixed polycations/polyelectrolytes were synthesized. The zwitterionic functionalities structurally based on sulfobetaines were grafted by amide linkages using the ϵ -amino functions of the polymer as attachment points. DMTMM as a water-soluble and highly hydrolysis resistant condensation agent was the key for the successful functionalization of α -PLL in water/methanol mixtures as solvents, which also completely suppresses the undesired formation of aggregates. The degree of functionalization can be fully controlled by the applied stoichiometry for each polymer. Using these novel mixed polycationic/polyelectrolytic polymers as the outermost layer in SMIL coatings, efficient separation is obtained. Moreover, it could be clearly demonstrated that the higher the degree of zwitterionic functionalization, the more the EOF decreases. These findings should give access to tailor-made conditions for the challenging separation of charge variants of therapeutic antibodies as well as any other separation issues of closely related proteins/proteoforms.

Author contributions

Henry Frick: conceptualization, data curation, formal analysis, investigation, validation, and writing – original draft. Laura Dhellemmes: conceptualization, data curation, formal analysis, investigation, methodology, software, validation, and writing – original draft. Alisa Höchsmann: investigation, data curation, formal analysis, and writing – review & editing. Laurent Leclercq: conceptualization, funding acquisition, methodology, project administration, resources, supervision, validation, and writing – review & editing. Christian Neusüß: funding acquisition, methodology, project administration, supervision, validation, and writing – review & editing. Hervé Cottet: conceptualization, funding acquisition, methodology,

project administration, supervision, validation, and writing – review & editing. Norbert Schaschke: conceptualization, funding acquisition, methodology, project administration, supervision, validation, and writing – review & editing.

Data availability

All data are available either in the manuscript or in its ESI.†

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This study was supported by the DFG (grant: SCHA 1012/4-1).

References

- J. S. Creamer, N. J. Oborny and S. M. Lunte, *Anal. Methods*, 2014, **6**, 5427–5449.
- G. K. E. Scriba and A. Psurek, *Methods Mol. Biol.*, 2008, **384**, 483–506.
- M. Pelzing and C. Neusüss, *Electrophoresis*, 2005, **26**, 2717–2728.
- A. Staub, D. Guillaume, J. Schappler, J.-L. Veuthey and S. Rudaz, *J. Pharm. Biomed. Anal.*, 2011, **55**, 810–822.
- S. S. Zhao and D. D. Y. Chen, *Electrophoresis*, 2014, **35**, 96–108.
- J. Hernández-Borges, C. Neusüss, A. Cifuentes and M. Pelzing, *Electrophoresis*, 2004, **25**, 2257–2281.
- T. Wehr, R. Rodríguez-Díaz and M. Zhu, *Capillary electrophoresis of proteins*, Marcel Dekker, New York, 1999.
- J. Horvath and V. Dolník, *Electrophoresis*, 2001, **22**, 644–655.
- V. Dolník, *Electrophoresis*, 2004, **25**, 3589–3601.
- F. H. Navarro, J. E. Gómez, J. H. Espinal and J. E. Sandoval, *Anal. Chim. Acta*, 2016, **948**, 104–112.
- E. Córdova, J. Gao and G. M. Whitesides, *Anal. Chem.*, 1997, **69**, 1370–1379.
- J. Znalezniona, D. Drahoňovský, B. Drahoš, J. Ševčík and V. Maier, *J. Sep. Sci.*, 2016, **39**, 2406–2412.
- J. P. Quirino, *J. Sep. Sci.*, 2017, **40**, 4835–4838.
- R. Sebastiano, M. E. Mendieta, N. Contiello, A. Citterio and P. G. Righetti, *Electrophoresis*, 2009, **30**, 2313–2320.
- S. Ullsten, A. Zuberovic and J. Bergquist, *Methods Mol. Biol.*, 2008, **384**, 631–646.
- N. Hamidli, M. Andrasi, C. Nagy and A. Gaspar, *J. Chromatogr. A*, 2021, **1654**, 462448.
- F. Kitagawa, M. Kamiya, Y. Okamoto, H. Taji, S. Onoue, Y. Tsuda and K. Otsuka, *Anal. Bioanal. Chem.*, 2006, **386**, 594–601.
- L. Leclercq, M. Morvan, J. Koch, C. Neusüss and H. Cottet, *Anal. Chim. Acta*, 2019, **1057**, 152–161.
- H. Katayama, Y. Ishihama and N. Asakawa, *Anal. Chem.*, 1998, **70**, 2254–2260.
- Q. Liu, F. Lin and R. A. Hartwick, *J. Chromatogr. Sci.*, 1997, **35**, 126–130.
- R. W. Chiu, J. C. Jimenez and C. A. Monnig, *Anal. Chim. Acta*, 1995, **307**, 193–201.
- X. Huang, Q. Wang and B. Huang, *Talanta*, 2006, **69**, 463–468.
- L. Vitali, F. Della Betta, A. C. O. Costa, F. A. S. Vaz, M. A. L. Oliveira, J. P. Vistuba, V. T. Fávere and G. A. Micke, *Talanta*, 2014, **123**, 45–53.
- S. Roca, L. Leclercq, P. Gonzalez, L. Dhellemmes, L. Boiteau, G. Rydzek and H. Cottet, *J. Chromatogr. A*, 2023, **1692**, 463837.
- R. Haselberg, G. J. de Jong and G. W. Somsen, *Anal. Chim. Acta*, 2010, **678**, 128–134.
- S. Bekri, L. Leclercq and H. Cottet, *J. Chromatogr. A*, 2015, **1399**, 80–87.
- R. Haselberg, G. J. de Jong and G. W. Somsen, *J. Sep. Sci.*, 2009, **32**, 2408–2415.
- R. Nehmé, C. Perrin, H. Cottet, M.-D. Blanchin and H. Fabre, *Electrophoresis*, 2009, **30**, 1888–1898.
- T. W. Graul and J. B. Schlenoff, *Anal. Chem.*, 1999, **71**, 4007–4013.
- S. T. Dubas and J. B. Schlenoff, *Macromolecules*, 1999, **32**, 8153–8160.
- S. L. Clark, M. F. Montague and P. T. Hammond, *Macromolecules*, 1997, **30**, 7237–7244.
- S. T. Dubas and J. B. Schlenoff, *Macromolecules*, 2001, **34**, 3736–3740.
- A. Höchsmann, L. Dhellemmes, L. Leclercq, H. Cottet and C. Neusüss, *Electrophoresis*, 2025, **46**, 279–289.
- Web.natur.cuni.cz/gas, ECHMET, <https://echmet.natur.cuni.cz/>, (accessed 14 December 2023).
- L. Dhellemmes, L. Leclercq, A. Höchsmann, C. Neusüss, J.-P. Biron, S. Roca and H. Cottet, *J. Chromatogr. A*, 2023, **1695**, 463912.
- N. E. Good, G. D. Winget, W. Winter, T. N. Connolly, S. Izawa and R. M. Singh, *Biochemistry*, 1966, **5**, 467–477.
- W.-H. Kuo, M.-J. Wang, H.-W. Chien, T.-C. Wei, C. Lee and W.-B. Tsai, *Biomacromolecules*, 2011, **12**, 4348–4356.
- F. Cao, L. Tan, L. Xiang, S. Liu and Y. Wang, *J. Biomater. Sci., Polym. Ed.*, 2013, **24**, 2058–2070.
- L. Chen, L. Tan, S. Liu, L. Bai and Y. Wang, *J. Biomater. Sci., Polym. Ed.*, 2014, **25**, 766–785.
- S. Mondal, *Chem. Rev.*, 2012, **112**, 5339–5355.
- M. Borzenkov, N. Mitina, V. Lobaz and O. Hevus, *J. Surfactants Deterg.*, 2015, **18**, 133–144.
- St. Guttman and R. A. Boissonnas, *Helv. Chim. Acta*, 1958, **41**, 1852–1867.
- M. Kunishima, C. Kawachi, J. Monta, K. Terao, F. Iwasaki and S. Tani, *Tetrahedron*, 1999, **55**, 13159–13170.
- A. Falchi, G. Giacomelli, A. Porcheddu and M. Taddei, *Synlett*, 2000, 275–277.

- 45 M. Kunishima, C. Kawachi, K. Hioki, K. Terao and S. Tani, *Tetrahedron*, 2001, **57**, 1551–1558.
- 46 A. Samavati, Z. Samavati, M. Velashjerdi, A. F. Ismail, M. H. D. Othman, G. Eisaabadi B, M. S. Abdullah, M. Bolurian and M. Bolurian, *Chem. Eng. J.*, 2021, **420**, 127655.
- 47 N. Olsson, P. James, C. A. K. Borrebaeck and C. Wingren, *Mol. Cell. Proteomics*, 2012, **11**, 342–354.
- 48 A. K. Bergfeld, O. M. T. Pearce, S. L. Diaz, T. Pham and A. Varki, *J. Biol. Chem.*, 2012, **287**, 28865–28881.
- 49 M. A. Gilles, A. Q. Hudson and C. L. Borders, *Anal. Biochem.*, 1990, **184**, 244–248.
- 50 A. Li, Y. Ye, P. Gong, B. Xiao and B. Jiang, *J. Appl. Polym. Sci.*, 2023, **140**, e53318.
- 51 M. D'Este, D. Eglin and M. Alini, *Carbohydr. Polym.*, 2014, **108**, 239–246.
- 52 W. E. Hull, H. R. Kricheldorf and M. Fehrle, *Biopolymers*, 1978, **17**, 2427–2443.
- 53 A. Dos, V. Schimming, S. Tosoni and H.-H. Limbach, *J. Phys. Chem. B*, 2008, **112**, 15604–15615.
- 54 W. Vayaboury, O. Giani, H. Collet, A. Commeyras and F. Schué, *Amino Acids*, 2004, **27**, 161–167.
- 55 K. Sato, C. E. Farquhar, J. Rodriguez and B. L. Pentelute, *J. Am. Chem. Soc.*, 2023, **145**, 12992–12997.
- 56 L. Dhellemmes, L. Leclercq, L. Lichtenauer, A. Höchsmann, M. Leitner, A. Ebner, M. Martin, C. Neusüß and H. Cottet, *Anal. Chem.*, 2024, **96**, 11172–11180.
- 57 L. Dhellemmes, L. Leclercq, H. Frick, A. Höchsmann, N. Schaschke, C. Neusüß and H. Cottet, *J. Chromatogr. A*, 2024, **1720**, 464802.
- 58 C. Huhn, R. Ramautar, M. Wuhrer and G. W. Somsen, *Anal. Bioanal. Chem.*, 2010, **396**, 297–314.