

Cite this: *RSC Adv.*, 2018, 8, 40984

Double-ratiometric fluorescence imaging of H₂Se and O₂^{•−} under hypoxia for exploring Na₂SeO₃-induced HepG2 cells' apoptosis†

Xiaonan Gao,[‡] Wenfei Guo,[‡] Lihong Ge, Fanpeng Kong, Kehua Xu* and Bo Tang[‡]

Sodium selenite (Na₂SeO₃), as an anti-tumor drug for inducing tumor cells' apoptosis, has been widely studied under normoxic conditions and has been shown to exhibit oxidative stress process-induced apoptosis. However, since the real tumor environment is hypoxic, the actual mechanism is still unclear. Hence, considering the main metabolite of Na₂SeO₃ in the metabolic process to be hydrogen selenide (H₂Se) and also that it can be converted to superoxide anion (O₂^{•−}) instantaneously in the presence of oxygen, a dual-ratiometric fluorescence imaging system for simultaneous monitoring of the changes of H₂Se and O₂^{•−} induced by Na₂SeO₃-guided tumor cell apoptosis under hypoxic conditions was constructed. Two molecular probes NIR-H₂Se and dihydroethidium were used to detect H₂Se and O₂^{•−}, respectively, whereas Rhodamine 110 was used as the reference fluorophore. Notably, H₂Se levels significantly increased under hypoxic conditions, but there was no change in the level of O₂^{•−}, which is inconsistent with the results of the previous researches. Therefore, we hypothesize that the mechanism of Na₂SeO₃-induced apoptosis for tumor cells is caused by reductive stress; also, this method can be applied for the future study of other anti-cancer selenium compounds.

Received 1st October 2018
Accepted 16th November 2018

DOI: 10.1039/c8ra08142e

rsc.li/rsc-advances

Introduction

Selenium,^{1,2} as a common protective agent against cancer, can effectively reduce the morbidity and mortality of tumors including lung cancer, colorectal cancer, lymphoma, and prostate cancer.^{3–5} The evidence from latent periods, clinical stages and epidemiology supports the chemopreventive role of selenium.^{6–8} Therefore, sodium selenite (Na₂SeO₃) has recently become the research hotspot as an anti-cancer medicine due to its high efficiency, low toxicity and cost-effectiveness. However, the precise anti-cancer mechanism of how Na₂SeO₃ can induce tumor cell apoptosis remains poorly understood. To date, research has been done on tumor cell cultures with Na₂SeO₃ under normoxic conditions. It shows that the most generally accepted mechanism for Na₂SeO₃ treatment is by cell apoptosis induced by elevating the reactive oxygen species (ROS) levels and the involvement of oxidative stress.^{9–12} Specifically, hydrogen selenide (H₂Se) is the key small molecule metabolite

of Na₂SeO₃ *in vivo*, and it reacts with O₂ to produce and accumulate a large amount of ROS (mainly the superoxide anion, O₂^{•−}), resulting in tumor cell oxidative stress, which leads to apoptosis.^{13–16} Nevertheless, since the tumor microenvironment is hypoxic (0.7–1.8% O₂)¹⁷ due to the rapid proliferation of cancer cells and imbalance of new blood vessel formation,^{18–20} it is still a great challenge to understand the anti-cancer mechanism in hypoxic conditions, which needs to be solved urgently.

To clarify the mechanism, fluorescent probes, which are promising detection tools²¹ due to their non-invasiveness, high sensitivity, selectivity, rapid response, and high spatial resolution,^{22–28} are the primary option. Unfortunately, these fluorescent probes are limited by environmental conditions, probe distribution, and instrument performance, thereby only showing changes in emission intensities.^{29,30} In contrast, ratio-metric probes, using the ratio of two different emission wavelengths as the detection signal, provide a built-in correction to the above factors for a more accurate analysis.^{31–34}

In this regard, a double-ratiometric method for simultaneous fluorescence imaging analysis to monitor the changes in H₂Se and O₂^{•−} levels in tumor cells was developed. The method studies the apoptosis process induced by pharmacological doses of Na₂SeO₃ in hypoxic conditions. Herein, we chose the fluorescence probe (NIR-H₂Se, λ_{ex}/λ_{em} = 688/735 nm) designed by our group³⁵ and the commercial fluorescence probe (dihydroethidium, λ_{ex}/λ_{em} = 370/420 nm)^{36,37} to monitor the changes in intracellular H₂Se and O₂^{•−} concentrations in both normoxic

College of Chemistry, Chemical Engineering and Materials Science, Key Laboratory of Molecular and Nano Probes, Ministry of Education, Collaborative Innovation Center of Functionalized Probes for Chemical Imaging in Universities of Shandong, Shandong Normal University, Jinan 250014, P. R. China. E-mail: Xukehua@sdsu.edu.cn; Tangb@sdsu.edu.cn

† Electronic supplementary information (ESI) available. See DOI: 10.1039/c8ra08142e

‡ X. G. and W. G. contribute equally to this work.



conditions and hypoxic conditions, respectively. Meanwhile, Rhodamine 110 ($\lambda_{\text{ex}}/\lambda_{\text{em}} = 496/532 \text{ nm}$) was chosen as a reference fluorophore. This method could potentially avoid the influences of certain external factors on detection accuracy including environmental conditions, light scattering and probe distribution, therefore improving the detection accuracy of the target molecules *in vivo*.^{38,39}

Experimental

Materials

All chemicals were analytical-reagent grade and all solvents were purified by a conventional method before use. Rhodamine 110 was purchased from J&K (China). The NIR-H₂Se probe was synthesized in our laboratory following a previous protocol.³⁵ Dihydroethidium (DHE) and KO₂ were purchased from Aladdin (China). Stock solutions (1 mM) for probes were prepared by dissolving probe in DMSO (Sigma-Aldrich). Sartorius ultrapure water (18.2 M Ω cm) was used throughout the experiments. H₂Se was prepared by reacting Al₂Se₃ with H₂O under an N₂ atmosphere for 30 min at room temperature before use every time.^{40,41} O₂^{•−} was generated from KO₂ in DMSO or xanthine oxidase. Na₂SeO₃ was purchased from Sigma-Aldrich. Dulbecco's Modified Eagle Medium (DMEM) was obtained from British Biotechnology (China). HepG2 cells were purchased from the Committee on Type Culture Collection of the Chinese Academy of Sciences.

Instruments

The fluorescence spectra measurements were performed with an FLS-920 Fluorescence Spectrometer (Edinburgh, UK). The fluorescence imaging of cells was recorded on a TCS SP5 confocal laser scanning microscope with an objective lens ($\times 40$) (Leica, Germany). The cells were cultured in a 5% CO₂/95% air incubator MCO-15 AC (Sanyo, Japan).

Fluorescence spectra measurements

Ten μM probe NIR-H₂Se was mixed with 5 μM probe DHE, 1 μM Rhodamine 110 and 10 mM PBS (pH = 7.4); then, various amounts of H₂Se (0, 1, 2, 4, 6, 8, 9, 10, 11 μM) were added. The fluorescence intensities were measured at $\lambda_{\text{ex}}/\lambda_{\text{em}} = 496/532 \text{ nm}$ and $\lambda_{\text{ex}}/\lambda_{\text{em}} = 688/735 \text{ nm}$. For a parallel study, 5 μM probe DHE was mixed with 10 μM probe NIR-H₂Se, 1 μM Rhodamine 110 and 10 mM PBS (pH = 7.4) and then, various amounts of O₂^{•−} (0, 1, 2, 3, 4, 5, 6, 7, 8 μM) were added for fluorescence measurements at $\lambda_{\text{ex}}/\lambda_{\text{em}} = 496/532 \text{ nm}$ and $\lambda_{\text{ex}}/\lambda_{\text{em}} = 488/638 \text{ nm}$.

Cell culture

HepG2 cells in the exponential phase of growth were used for the cell experiments. The concentrations of the counted cells were adapted to 1×10^6 cells per mL for confocal fluorescence imaging with a culture medium. The cells were cultured with DMEM and supplemented with 1% antibiotics (100 U mL^{−1} penicillin and 100 mg mL^{−1} streptomycin) and 10% fetal bovine serum (FBS) at 37 °C under a humidified atmosphere of 5%

CO₂/95% air (20% O₂), 75% CO₂/25% air (5% O₂) and 95% CO₂/5% air (1% O₂) for normoxic and hypoxic conditions in an MCO-15AC incubator, respectively.

Confocal fluorescence imaging

HepG2 cells were prepared in glass-bottom culture dishes and cultured for 24 h. One group of cells without treatment of Na₂SeO₃ served as the control group and the other groups were treated with various amounts of Na₂SeO₃ (2, 5, 10 μM) for 12 h under either normoxic or hypoxic conditions. All groups were further incubated in an FBS-free culture medium mixed with 10 μM probe NIR-H₂Se, 5 μM probe DHE and 1 μM Rhodamine 110 at 37 °C for 15 min and then washed with PBS (pH = 7.4, 10 mM) three times. Afterwards, the cells were imaged immediately using a confocal microscope with an objective lens ($\times 40$) and examined by CLSM with different laser transmitters. The emitted light intensities were collected with an META detector from 490 to 550 nm, 590 to 670 nm and 690 to 760 nm.

Results and discussion

Spectroscopic properties of NIR-H₂Se with H₂Se, DHE with O₂^{•−} and Rhodamine 110

The excitation and emission spectra of NIR-H₂Se with H₂Se, DHE with O₂^{•−} and Rhodamine 110 were recorded (Fig. S1†). Interestingly, the emission spectra of the reaction products of NIR-H₂Se with H₂Se, DHE plus O₂^{•−}, and Rhodamine 110 are located at 735, 638 and 532 nm, respectively. Therefore, the emission spectra of Rhodamine 110 and the two reaction products exhibit a fairly ideal separation effect with approximately 100 nm between each peak, making their simultaneous detection possible (Fig. 1).

Ratiometric detection

The fluorescence responses of DHE to H₂Se and NIR-H₂Se to O₂^{•−} were tested by using the mixture of NIR-H₂Se, DHE and Rhodamine 110 to simultaneously monitor the levels of H₂Se and O₂^{•−}. As shown in Fig. 2a, O₂^{•−} (5 μM) yielded a strong fluorescence signal with DHE (5 μM), but high concentrations

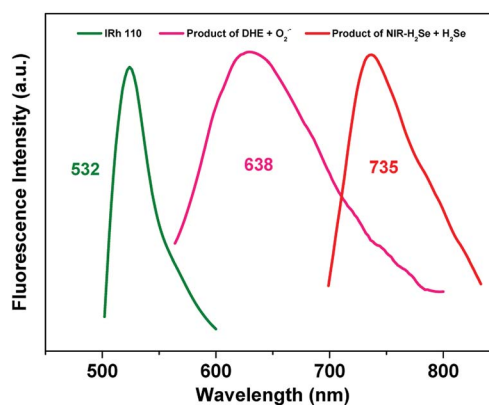


Fig. 1 Fluorescence emission spectra of (red) NIR-H₂Se with H₂Se, (pink) DHE with O₂^{•−} and (green) Rhodamine 110.



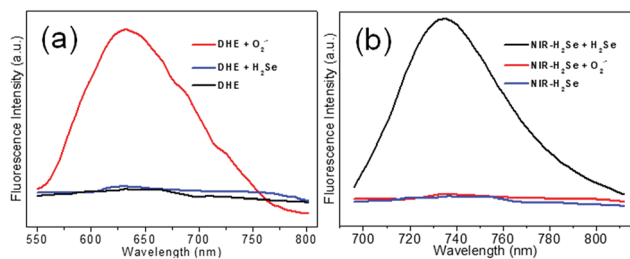


Fig. 2 Interference measurement of (a) DHE to H₂Se and (b) NIR-H₂Se to O₂^{•-}. All of the spectra were acquired at room temperature in 10 mM PBS (pH = 7.4) with (a) $\lambda_{\text{ex}}/\lambda_{\text{em}} = 488/638$ nm or (b) $\lambda_{\text{ex}}/\lambda_{\text{em}} = 688/735$ nm.

of H₂Se (15 μM) did not exhibit a fluorescence response. Similarly, the fluorescence response of NIR-H₂Se (10 μM) to O₂^{•-} (10 μM) was negligible, but a strong fluorescence signal was obtained with 10 μM H₂Se (Fig. 2b). Then, the optical properties of the probe mixtures with Rhodamine 110 were recorded with various concentrations of H₂Se in simulated physiological conditions (10 mM PBS, pH = 7.4). The fluorescence intensities increased with increasing concentrations of H₂Se at 735 nm (Fig. 3a) but hardly fluctuated at 532 nm (Fig. 3b, black curve, right Y scale). Interestingly, there is a good linear correlation between the fluorescence intensity ratio F_{735}/F_{532} and H₂Se concentrations from 0 to 11.0 μM (Fig. 3b, red curve, left Y scale). The equation of linear regression is $F_{735}/F_{532} = 0.23 + 0.17 [\text{H}_2\text{Se}] \mu\text{M}$ with the R value of 0.9937, indicating that the fluorescence intensity ratio F_{735}/F_{532} increased with increasing concentrations of H₂Se linearly. Simultaneously, similar

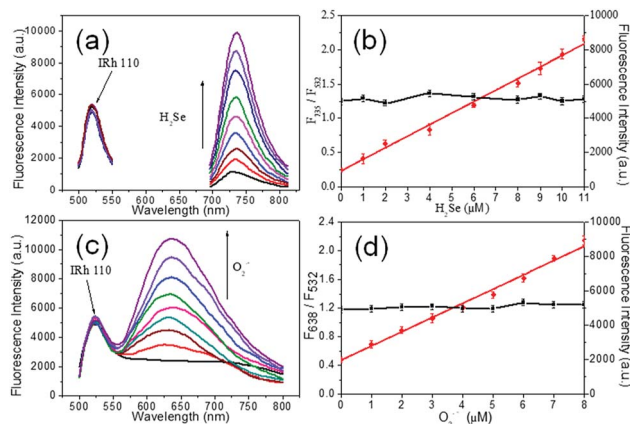


Fig. 3 Ratiometric detection with the mixture of 10 μM NIR-H₂Se, 5 μM DHE and 1 μM Rhodamine 110 toward H₂Se and O₂^{•-}. (a) Fluorescence response of the mixture to various concentrations (0 to 11.0 μM) of H₂Se. (b) The linear correlation between the fluorescence intensity ratio F_{735}/F_{532} and H₂Se concentrations (red) and fluorescence intensities changes of Rhodamine 110 to H₂Se concentrations (black). (c) Fluorescence response of the mixture to various concentrations (0 to 8.0 μM) of O₂^{•-}. (d) The linear correlation between the fluorescence intensity ratio F_{638}/F_{532} and O₂^{•-} concentrations (red) and changes in the fluorescence intensity of Rhodamine 110 to O₂^{•-} concentrations (black). The data shown are representative of repeating experiments ($n = 3$).

fluorescence responses of the mixture towards O₂^{•-} under various concentrations of O₂^{•-} in simulated physiological conditions (10 mM PBS, pH = 7.4) were also obtained (Fig. 3c); the fluorescence intensity was significantly enhanced *via* increasing O₂^{•-} concentrations at 638 nm but was maintained at 532 nm (Fig. 3d, black curve, right Y scale). Also, a linear correlation between the fluorescence intensity ratio F_{638}/F_{532} and O₂^{•-} concentrations from 0 to 8.0 μM (Fig. 3d, red circles, left Y scale) was observed. The equation of linear regression is $F_{638}/F_{532} = 0.47 + 0.20 [\text{O}_2^{\bullet-}] \mu\text{M}$ with the R value of 0.9923, indicating that the fluorescence intensity ratio of F_{638}/F_{532} increased linearly with increasing concentrations of O₂^{•-}. Overall, the results demonstrated that it is entirely feasible to use NIR-H₂Se and DHE for simultaneously monitoring the changes in H₂Se and O₂^{•-} in a complex system.

Co-staining imaging in living cells

To further test the simultaneous imaging of H₂Se and O₂^{•-} *in vitro*, co-staining experiments were carried out. Since Na₂SeO₃ metabolite H₂Se is mainly accumulated in the liver, the cell model HepG2 was selected for the following study. HepG2 cells were first incubated in FBS-free culture medium mixed with 10 μM NIR-H₂Se, 5 μM DHE and 1 μM Rhodamine 110 at 37 $^{\circ}\text{C}$ for 15 min and then, confocal fluorescence images were obtained. Fig. 4 displays the distribution of Rhodamine 110 (green), the product of DHE with native O₂^{•-} of HepG2 cells (yellow), the product of NIR-H₂Se with H₂Se (red), and the merged image. The two probes and the reference achieved excellent spectral separation in the cell as well as great biocompatibility. The two probes' reaction products and the reference molecular structure all contain two amino groups, making the degree of impact by the intracellular complex environment almost the same, which further improved the sensitivity of the detection. Hence, the mixture of NIR-H₂Se, DHE and Rhodamine 110 can be used to simultaneously recognize H₂Se and O₂^{•-} in living cells.

Double-ratiometric simultaneous detection under normoxia and hypoxia

For the next step, the probe mixture was applied to explore its application for double-ratiometric, simultaneous imaging of endogenous H₂Se and O₂^{•-} in HepG2 cells pre-treated with Na₂SeO₃ under normoxic (20% O₂) and hypoxic (5% O₂, 1% O₂)

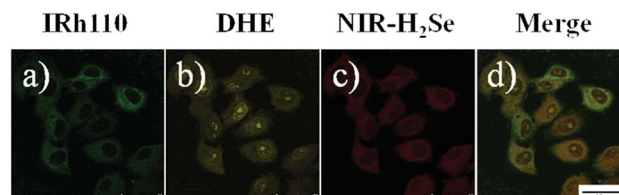


Fig. 4 Co-staining images of HepG2 cells incubated with FBS-free culture medium mixed with 1 μM Rhodamine 110, 5 μM DHE and 10 μM NIR-H₂Se at 37 $^{\circ}\text{C}$ for 15 min. The emitted light was collected with an META detector from 490 to 550 nm, 590 to 670 nm and 690 to 760 nm. (a) Green channel. (b) Yellow channel. (c) Red channel. (d) Merge of (a), (b) and (c). Scale bar = 50 μm .



conditions. One group of HepG2 cells without treatment of Na_2SeO_3 served as the control group, and the other groups were treated with Na_2SeO_3 (2, 5, 10 μM) for 12 h under either normoxia (20% O_2) or hypoxia (5% O_2 , 1% O_2). Afterwards, the cells were incubated in an FBS-free culture medium mixed with 10 μM NIR- H_2Se , 5 μM probe DHE and 1 μM Rhodamine 110 at 37 $^\circ\text{C}$ for 15 min. The ratiometric images of H_2Se were achieved by combining the images obtained in the band path of 490–550 nm with the corresponding images in the band path of 690–760 nm. Meanwhile, the concentration of $\text{O}_2^{\cdot-}$ was also obtained by combining the images obtained in the band path of 490–550 nm with the corresponding images in the band path of 590–670 nm (Fig. 5 and S2–S4†). Fig. 5a shows that under different oxygen concentrations, the concentration of H_2Se in HepG2 cells increased by the Na_2SeO_3 concentration. From the quantitative map, it can be seen that with the decrease in oxygen concentration and increase in Na_2SeO_3 concentration, the H_2Se content gradually increased. At 1% O_2 , the concentration of H_2Se in the HepG2 cells was relatively high with 10 μM Na_2SeO_3 (Fig. 5b). Consequently, under different oxygen concentrations, the content of $\text{O}_2^{\cdot-}$ in the HepG2 cells increased while increasing Na_2SeO_3 since the increase of oxygen and Na_2SeO_3 led to gradually rising $\text{O}_2^{\cdot-}$ levels (Fig. 5c). At 20% O_2 , the concentration of $\text{O}_2^{\cdot-}$ in the HepG2 cells was relatively high with 10 μM Na_2SeO_3 incubation, but the content of $\text{O}_2^{\cdot-}$ in the HepG2 cells remained unchanged under the condition of 1% O_2 (Fig. 5d).

In this regard, higher H_2Se contents were observed in hypoxic environments rather than in normoxic conditions in parallel experiments. Correspondingly, lower $\text{O}_2^{\cdot-}$ contents were maintained and they hardly changed under hypoxic environments, whereas $\text{O}_2^{\cdot-}$ contents gradually increased in the

HepG2 cells with Na_2SeO_3 in a time- and dose-dependent manner under normoxic conditions. These results indicate that the Na_2SeO_3 anticancer mechanism is not an ROS-induced apoptosis process under hypoxic conditions. Thus, the anti-cancer effect of Na_2SeO_3 in solid tumors should be reductive stress to trigger cell death of cancer cells.

Conclusions

In summary, we developed a novel double-ratiometric fluorescence probe system for simultaneous fluorescence imaging analysis of the changes in H_2Se and $\text{O}_2^{\cdot-}$ levels in the tumor cell HepG2's apoptosis process induced by pharmacological doses of Na_2SeO_3 . The system verifies that the content of $\text{O}_2^{\cdot-}$ is clearly higher in the tumor cell apoptosis process induced by Na_2SeO_3 in normoxic conditions, and H_2Se is almost completely oxidized by O_2 to produce $\text{O}_2^{\cdot-}$. In contrast, under hypoxic conditions, the $\text{O}_2^{\cdot-}$ content is extremely low, but the H_2Se content increases dramatically. Therefore, in solid tumors, the anti-cancer mechanism of Na_2SeO_3 can be ascribed to reductive stress induced by H_2Se but not oxidative stress. This finding will significantly improve the investigation of the anti-cancer mechanism of Na_2SeO_3 , and the double-ratiometric fluorescence method can be used as a suitable tool for the study of other selenium compounds in the future.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This work was supported by National Natural Science Foundation of China (21575081, 21775091, 21535004, 91753111 and 21705098) and the Key Research and Development Program of Shandong Province (2018YFJH0502).

References

- 1 J. Chen, Y. Zhu and Y. Zhang, *RSC Adv.*, 2016, **6**, 62193–62199.
- 2 Y. Nan, W. Zhao, X. Xu, C.-T. Au and R. Qiu, *RSC Adv.*, 2015, **5**, 69299–69306.
- 3 Y. Cui, S. Vogt, N. Olson, A. G. Glass and T. E. Rohan, *Cancer Epidemiol., Biomarkers Prev.*, 2007, **16**, 1682–1685.
- 4 T. Chen and Y.-S. Wong, *Int. J. Biochem. Cell Biol.*, 2009, **41**, 666–676.
- 5 I. A. Asfour, M. M. El-Tehewi, M. H. Ahmed, M. A. Abdel-Sattar, N. N. Moustafa, H. M. Hegab and O. M. Fathey, *Biol. Trace Elem. Res.*, 2009, **127**, 200–210.
- 6 R. Sinha and K. El-Bayoumy, *Curr. Cancer Drug Targets*, 2004, **4**, 13–28.
- 7 L. C. Clark, B. Dalkin, A. Krongrad, G. F. Combs Jr, B. W. Turnbull, E. H. Slate, R. Witherington, J. H. Herlong, E. Janosko, D. Carpenter, C. Borosso, S. Falk and J. Rounder, *Br. J. Urol.*, 1998, **81**, 730–734.

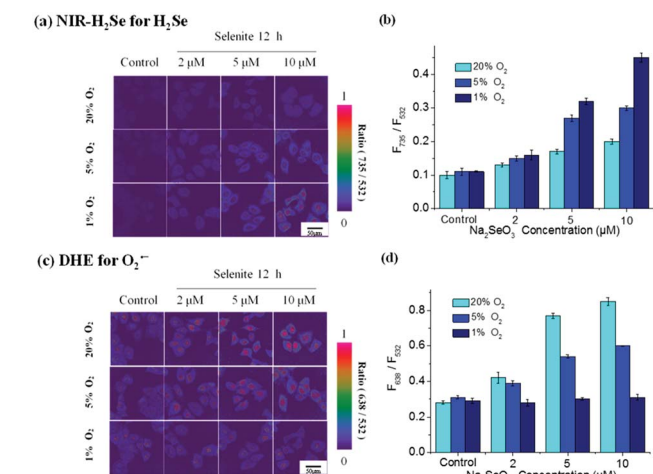


Fig. 5 Confocal fluorescence double-ratiometric images of endogenous H_2Se and $\text{O}_2^{\cdot-}$ in living HepG2 cells treated with various concentrations of Na_2SeO_3 under normoxic and hypoxic conditions. (a) The fluorescence ratio image (F_{735}/F_{532}). (b) The corresponding statistic fluorescence ratio (F_{735}/F_{532}) for (a). (c) The fluorescence ratio image (F_{638}/F_{532}). (d) The corresponding statistic fluorescence ratio (F_{638}/F_{532}) for (c). The images shown are representative of repeated experiments ($n = 3$). Scale bar = 50 μm .



- 8 Y. Xia, C. Wang, T. Xu, Y. Li, M. Guo, Z. Lin, M. Zhao and B. Zhu, *RSC Adv.*, 2018, **8**, 1917–1926.
- 9 H.-M. Shen, C.-F. Yang and C.-N. Ong, *Int. J. Cancer*, 1999, **81**, 820–828.
- 10 G. Nilsson, X. Sun, C. Nyström, A.-K. Rundlöf, A. P. Fernandes, M. Björnstedt and K. Dobra, *Free Radical Biol. Med.*, 2006, **41**, 874–885.
- 11 L. Guan, B. Han, Z. Li, F. Hua, F. Huang, W. Wei, Y. Yang and C. Xu, *Apoptosis*, 2009, **14**, 218–225.
- 12 J. J. An, K. J. Shi, W. Wei, F. Y. Hua, Y. L. Ci, Q. Jiang, F. Li, P. Wu, K. Y. Hui, Y. Yang and C. M. Xu, *Cell Death Dis.*, 2013, **4**, 973–982.
- 13 H. Luo, Y. Yang, F. Huang, F. Li, Q. Jiang, K. Shi and C. Xu, *Cancer Lett.*, 2012, **315**, 78–85.
- 14 S.-H. Park, J.-H. Kim, G. Y. Chi, G.-Y. Kim, Y.-C. Chang, S.-K. Moon, S.-W. Nam, W.-J. Kim, Y. H. Yoo and Y. H. Choi, *Toxicol. Lett.*, 2012, **212**, 252–261.
- 15 X. Liu, B. Hu, R. Cheng, F. Kong, X. Pan, K. Xu and B. Tang, *Chem. Commun.*, 2016, **52**, 6693–6696.
- 16 B. Hu, R. Cheng, X. Liu, X. Pan, F. Kong, W. Gao, K. Xu and B. Tang, *Biomaterials*, 2016, **92**, 81–89.
- 17 S. Rey, L. Schito, M. Koritzinsky and B. G. Wouters, *Adv. Drug Delivery Rev.*, 2017, **109**, 45–62.
- 18 P. Vaupel, F. Kallinowski and P. Okunieff, *Cancer Res.*, 1989, **49**, 6449–6465.
- 19 D. Hanahan and R. A. Weinberg, *Cell*, 2000, **100**, 57–70.
- 20 M. C. Hung, G. B. Mills and D. Yu, *Nat. Med.*, 2009, **15**, 246–247.
- 21 E. Zhang, P. Ju, P. Guo, X. Hou, X. Hou, H. Lv, J.-j. Wang and Y. Zhang, *RSC Adv.*, 2018, **8**, 31658–31665.
- 22 T. Ueno and T. Nagano, *Nat. Methods*, 2011, **8**, 642–645.
- 23 L. L. Zhang, H. K. Zhu, C. C. Zhao and X. F. Gu, *Chin. Chem. Lett.*, 2017, **28**, 218–221.
- 24 F. Wang, Y. Zhu, L. Zhou, L. Pan, Z. Cui, Q. Fei, S. Luo, D. Pan, Q. Huang, R. Wang, C. Zhao, H. Tian and C. Fan, *Angew. Chem., Int. Ed.*, 2015, **54**, 7349–7353.
- 25 C. Zhao, X. Zhang, K. Li, S. Zhu, Z. Guo, L. Zhang, F. Wang, Q. Fei, S. Luo, P. Shi, H. Tian and W. H. Zhu, *J. Am. Chem. Soc.*, 2015, **137**, 8490–8498.
- 26 J. Xu, J. Pan, X. Jiang, C. Qin, L. Zeng, H. Zhang and J. F. Zhang, *Biosens. Bioelectron.*, 2016, **77**, 725–732.
- 27 X. Cheng, H. Jia, J. Feng, J. Qin and Z. Li, *Sens. Actuators, B*, 2013, **184**, 274–280.
- 28 X. Gu, C. Liu, Y. C. Zhu and Y. Z. Zhu, *J. Agric. Food Chem.*, 2011, **59**, 11935–11939.
- 29 X. Liu, Q. Yang, W. Chen, L. Mo, S. Chen, J. Kang and X. Song, *Org. Biomol. Chem.*, 2015, **13**, 8663–8668.
- 30 L. Yuan, W. Lin, Y. Xie, B. Chen and J. Song, *Chem.–Eur. J.*, 2012, **18**, 2700–2706.
- 31 L. Yuan, W. Lin, J. Song and Y. Yang, *Chem. Commun.*, 2011, **47**, 12691–12693.
- 32 X. Jiang, J. Xu, Y. Zhang, H. Wang, L. Zeng and Y. Zhang, *Anal. Methods*, 2016, **8**, 1572–1576.
- 33 L. Zhu, J. Xu, Z. Sun, B. Fu, C. Qin, L. Zeng and X. Hu, *Chem. Commun.*, 2015, **51**, 1154–1156.
- 34 Z. Ye, C. Duan, R. Sheng, J. Xu, H. Wang and L. Zeng, *Talanta*, 2018, **176**, 389–396.
- 35 F. Kong, L. Ge, X. Pan, K. Xu, X. Liu and B. Tang, *Chem. Sci.*, 2016, **7**, 1051–1056.
- 36 Z. Yu, Q. Sun, W. Pan, N. Li and B. Tang, *ACS Nano*, 2015, **9**, 11064–11074.
- 37 R. M. Monaghan, R. G. Barnes, K. Fisher, T. Andreou, N. Rooney, G. B. Poulin and A. J. Whitmarsh, *Nat. Cell Biol.*, 2015, **17**, 782–792.
- 38 K. Komatsu, Y. Urano, H. Kojima and T. Nagano, *J. Am. Chem. Soc.*, 2007, **129**, 13447–13454.
- 39 D. Srikun, E. W. Miller, D. W. Domaille and C. J. Chang, *J. Am. Chem. Soc.*, 2008, **130**, 4596–4597.
- 40 C. Mealli, S. Midollini and L. Sacconi, *Inorg. Chem.*, 1978, **17**, 632–637.
- 41 V. V. Matytilsky, A. Shavel, N. Gaponik, A. Eychmüller and J. Wachtveitl, *J. Phys. Chem. C*, 2008, **112**, 2703–2710.

