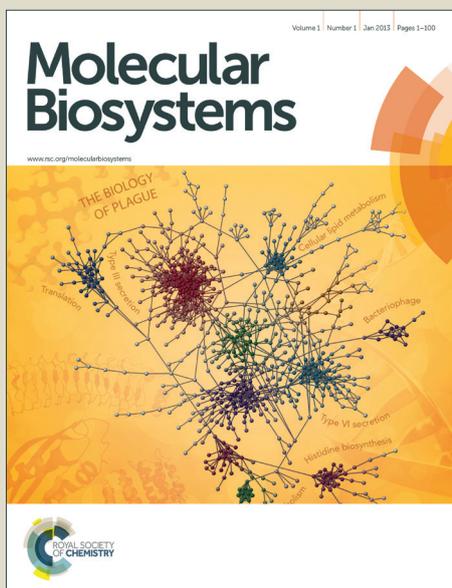


# Molecular BioSystems

Accepted Manuscript



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

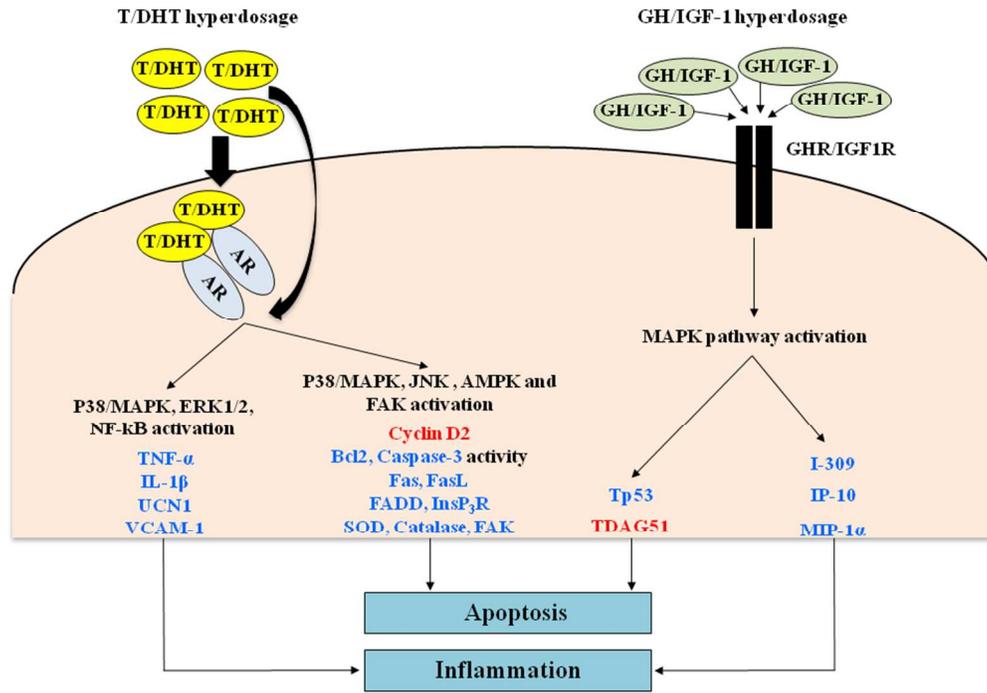
*Accepted Manuscripts* are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this *Accepted Manuscript* with the edited and formatted *Advance Article* as soon as it is available.

You can find more information about *Accepted Manuscripts* in the [Information for Authors](#).

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard [Terms & Conditions](#) and the [Ethical guidelines](#) still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.



[www.rsc.org/molecularbiosystems](http://www.rsc.org/molecularbiosystems)



Supraphysiological doses of doping agents, such as T/DHT and GH/IGF-1, affect cellular pathways associated with apoptosis and inflammation.

254x190mm (96 x 96 DPI)

## Molecular effects of supraphysiological doses of doping agents on health

Esther Imperlini<sup>a§c</sup>, Annamaria Mancini<sup>b,c§</sup>, Andreina Alfieri<sup>b,c</sup>, Domenico Martone<sup>b</sup>, Marianna Caterino<sup>c</sup>, Stefania Orrù<sup>b,c\*</sup> and Pasqualina Buono<sup>a,b\*</sup>

<sup>a</sup>IRCCS SDN, Naples, Italy;

<sup>b</sup>Dipartimento di Scienze Motorie e del Benessere, Università “Parthenope” di Napoli, Naples, Italy;

<sup>c</sup>CEINGE Biotecnologie Avanzate s.c. a r.l., Naples, Italy.

\*Corresponding authors: Proff. Stefania Orrù and Pasqualina Buono, Dipartimento di Scienze Motorie e del Benessere, Università “Parthenope” di Napoli, via Medina 40, 80133 Naples, Italy; email: [orru@uniparthenope.it](mailto:orru@uniparthenope.it); [buono@uniparthenope.it](mailto:buono@uniparthenope.it)

<sup>§</sup>These authors contributed equally

<sup>¶</sup>Present address: CEINGE Biotecnologie Avanzate s.c. a r.l., Naples, Italy.

Keywords: Testosterone, Dihydrotestosterone, Growth Hormone, Insulin-like Growth Factor-1, doping

Abbreviations: anabolic androgenic steroid (AAS), cardiovascular disease (CVD), cerebellar granulososa cell (CGC), dihydrotestosterone (DHT), growth hormone (GH), GH binding protein (GHBP), GH deficiency (GHD), GH receptor (GHR), human aortic endothelial cell line (HAEC), human hepatocyte cell line (HEPG2), human monocyte-derived macrophages (HMDM), human neuroblastoma cell line (SH-SY5Y), human peripheral blood lymphocyte (PBL), human proximal tubular epithelial cell line (HK-2), human umbilical vein endothelial cell line (HUVEC), high-density lipoprotein (HDL), insulin-like growth factor-1 (IGF-1), IGF-1 binding protein (IGFBP), IGF-1 receptor (IGF1R), low-density lipoprotein (LDL), Performance- enhancing drug (PED), primary human proximal tubular epithelial cells (PTEC), rat pheochromocytoma cell line (PC12), sex hormone binding protein (SHBP), testosterone (T), total cholesterol (TC), world anti-doping agency (WADA).

**1 Abstract**

2 Performance-enhancing drugs (PEDs) gained a wide popularity not only among sportsmen but also  
3 among specific subsets of population, such as adolescents. Apart from their claimed effects on  
4 athletic performance, they are very appealing due to the body shaping effect exerted on fat mass  
5 and fat-free mass. Beside the “underestimated” massive misuse of PEDs, the short- as well as long-  
6 term consequences of such habits remain largely unrecognized. They have been strictly associated  
7 with serious adverse effects, but molecular mechanisms are far to be elucidated. Here, we analyze  
8 the current understanding about the molecular effects of supraphysiological doses of doping  
9 agents in healthy biological systems, at genomic and proteomic level, in order to define the  
10 molecular sensors of organ/tissue impairment, determined by their misuse. The focus is put on the  
11 anabolic androgenic steroids (AASs), specifically testosterone (T) and its most potent derivative  
12 dihydrotestosterone (DHT), and on the peptide hormones, specifically the growth hormone (GH)  
13 and the insulin-like growth factor-1 (IGF-1). A map of molecular targets is defined and the risk  
14 incidence for human health is taken into account.

15

16

17

18

## 1 Introduction

2 The use of performance-enhancing drugs (PEDs), commonly referred as doping agents, is no longer  
3 restricted to sport, but affects also the general population. There is an increasing public-health  
4 concern about the widespread misuse of illicit drugs among non-competing amateurs and high-  
5 school students in several European countries and in the USA<sup>1,2</sup>. Based on anonymous  
6 questionnaires, it has been estimated that the consumers of each country represent about 1% of  
7 their respective populations<sup>1</sup>. However, one major problem related to the under- or over-  
8 interpretation of these estimates is the sparse information coming from reliable studies of PED  
9 abuse in healthy subjects. There is a substantial under-reporting of the numerous side effects of  
10 doping agents. The long-term consequences of their misuse remain largely unknown, and, on the  
11 other hand, the chronic toxicity from past long-term abusers must be considered nowadays a  
12 growing public health problem.

13 Apart from their claimed role in athletic performance, PED misuse is also strictly associated with  
14 serious adverse effects on health, such as cardiovascular diseases and cancer<sup>3-5</sup>. Clinical studies  
15 demonstrated that acute myocardial infarction is the most common event among PED abusers<sup>6,7</sup>  
16 and that left ventricular hypertrophy may even persist after abuse cessation<sup>8</sup>. The entity of these  
17 side effects depends on sex, dose, duration of treatment, fitness condition and on genetic factors.  
18 The aim of this review is to analyze the current understanding about the molecular effects of  
19 supraphysiological doses of doping agents in healthy biological systems in order to define the  
20 molecular sensors of organ/tissue impairment, at both mRNA and protein level, determined by  
21 PED misuse. Molecular targets are then discussed in relation to the risk of incidence of specific  
22 pathological outcomes. Data concerning the effects of supraphysiological doses of doping agents  
23 carried out on unhealthy subjects were not considered in this study. The focus was put on two  
24 main categories of doping agents, prohibited by the code of World Anti-Doping Agency (WADA):  
25 the anabolic androgenic steroids (AASs), specifically testosterone (T) and its most potent  
26 derivative dihydrotestosterone (DHT), and the peptide hormones, specifically the growth hormone  
27 (GH) and the insulin-like growth factor-1 (IGF-1).

28

29

30

## 1 **Testosterone and dyhydrotestosterone**

2 AASs are synthetic derivatives of T, the main male sexual hormone. Endogenous T is produced by  
3 the Leydig cells in the testes; a small amount can be also secreted by adrenal cortex or obtained by  
4 the peripheral conversion of androstenedione<sup>9</sup>. In serum most of circulating T is bound to carrier  
5 proteins: sex hormone binding globulin (SHBG) and albumin. Its lipophilic structure allows T to  
6 freely cross the plasma membrane of target cells and to bind in the cytoplasm to Androgen  
7 Receptor (AR), a 110-kDa member of the nuclear receptor superfamily of ligand-activated  
8 transcription factors<sup>10</sup>. By means of 5 $\alpha$ -reductase enzymatic activity, in androgen-responsive  
9 tissues, such as muscles, internalized T can be rapidly converted into DHT, the most potent  
10 derivative, able to bind AR with higher affinity<sup>11</sup>. Ligand-activated AR translocates to the nucleus  
11 where it is able to regulate gene expression directly<sup>12</sup> or indirectly through the interaction with  
12 some transcription factors, such as c-Jun, Foxa2, Oct1, GATA1, AP-1, p53, RelA, SHP and others<sup>13-</sup>  
13 <sup>14</sup>. Genes, regulated by nuclear ligand-activated AR, encode muscle-specific transcription factors,  
14 enzymes and structural proteins<sup>15</sup>; recently, it has been shown that the genomic regulation of  
15 ligand-activated AR is exerted also on miRNAs<sup>16-19</sup>.

16 AASs can also regulate cell-specific molecular pathways. In fact, besides to the genomic action,  
17 which occurs within an hour, T/DHT show also a rapid (seconds to minutes) non-genomic activity  
18 that modulates several signal transduction pathways, including IGF-1 signaling<sup>20-21</sup>. In this context,  
19 T/DHT activate second messenger pathways, apart from the classical transcriptional activity, by  
20 establishing a crosstalk with signaling molecules, in two main ways: they activate the tyrosine  
21 kinase c-Src<sup>22-24</sup> and two members of the MAPK signaling cascade (Raf1 and ERK-2)<sup>23</sup> in an AR-  
22 mediated fashion; moreover, T and DHT can also activate cAMP and PKA through the SHBG  
23 receptor<sup>21</sup>.

24 Testosterone can also be irreversibly converted by aromatase enzyme to estradiol (E2), the female  
25 sexual hormone, equally able to act through genomic and non-genomic mechanisms<sup>25</sup>. The  
26 relative amounts between T and E2 is cell specific and their functions, determined by a complex  
27 interaction between genomic and non-genomic activities, are distinct, if not conflicting, in  
28 different cellular types<sup>9</sup>.

29 AASs gained a wide popularity among sport players due to specific anabolic effects, such as the  
30 increase of lean body mass, decrease of fat mass, increase of strength and enhancement of  
31 athletic performance; moreover, it cannot be forget that the younger abusers are mainly  
32 influenced by aesthetic purposes<sup>26,27</sup>. Several AASs are included in the Prohibited List, published by

1 WADA yearly.

2 Such an abuse among professional athletes and amateurs is in contrast with well-documented  
3 adverse effects associated to AAS consumption: AAS abuse is reported to be strictly associated to  
4 risk increase in cardiovascular disease, liver diseases and reproductive system alterations and  
5 changes in behaviour<sup>28-33</sup>. These numerous side-effects as well as the chronic toxicity from past  
6 long-term abuse in now middle-aged men must be considered, hence, a growing public health  
7 problem.

8 Timing and protocols of assumption are extremely variable and in general consumers assume  
9 multiple drugs for a total androgen dose ranging between 10- and 100-fold above physiological  
10 concentration<sup>34</sup>. The average physiological level of T and/or DHT in male serum is 10 nM<sup>35</sup>. Several  
11 reports define 100 nM as a supraphysiological concentration (see Table S1).

12 The improvements in muscle strength observed in response to T administration have been widely  
13 described previously<sup>9</sup> and are out of scope of this review.

14 The aim of the present review is to highlight the molecular effects (genes, proteins) mediated by  
15 T/DHT administration on different cells/systems and strictly associated to increased risk for human  
16 health.

17

18

19

## 1 **Growth hormone and Insulin-like Growth factor-1**

2 Growth Hormone (GH) is a single-chain polypeptide hormone produced and secreted by anterior  
3 pituitary gland<sup>36,37</sup>. GH secretion is regulated by two hypothalamic factors, the Growth Hormone  
4 Relasing Hormone (GHRH) and somatostatin, the first stimulating and the last inhibiting the  
5 process<sup>38</sup>. GH secretion takes place in a pulsatile manner with major peaks occurring at the onset  
6 of slow-wave sleep and few hours after the meal. Many factors affect GH secretion: gender, age,  
7 adiposity, sleep, diet, exercise and other<sup>39</sup>. The levels of circulating hormone are maximal at  
8 puberty<sup>39</sup> and decline during adulthood<sup>40</sup>; in aged men, GH levels are 5- to 20-fold lower than  
9 young adults, and they are associated to a reduction in GHRH and an increment in somatostatin<sup>38</sup>.

10 Insuline-like Growth Factor 1 (IGF-1), a peptide hormone and a tissue growth factor, is produced  
11 by the liver in response to GH action and circulates at nanomolar concentrations. However, GH  
12 modulates IGF-1 production in a paracrine/autocrine fashion in healthy individuals in many other  
13 GH-responsive tissues<sup>41,42</sup>. IGF-1 mediates many of GH actions, included anabolic functions and  
14 growth promoting effects, and exhibits mitogenic and insulin-like metabolic activities<sup>42</sup>. There are  
15 two isoforms in humans: IGF-1-liver type and IGF1Ec, mainly produced by the skeletal muscle and  
16 known as Mechano Growth-Factor<sup>43</sup>.

17 Circulating GH and IGF-1 are associated to GH- and IGF-binding proteins (GHBPs and IGFbps,  
18 respectively), which regulate hormone half-life and receptor interaction<sup>44-46</sup>. In fact, both  
19 hormones activate transduction signaling in target tissues through their membrane receptors: the  
20 GH receptor (GHR), a plasma membrane-resident receptor of the cytokine receptor class I  
21 superfamily<sup>37</sup>, and the IGF-1 receptor (IGF1R), a tyrosine kinase membrane receptor homologous  
22 to oncogenes of tyrosine kinase class, along the insulin receptor (IR)<sup>47</sup>.

23 At cellular level, GH/IGF-1 activate the JAK2-Stat5b, the Akt and the MAPK intracellular signalling  
24 pathway, particularly important for GH growth-promoting activity<sup>42,48</sup>. Both hormones promote  
25 proliferation and survival of a wide range of cell types<sup>49-52</sup> and increase differentiation of cells  
26 including myoblasts<sup>53,54</sup>.

27 IGF-1 level in the serum are stable in healthy individuals and its administration inhibit GH release.  
28 Subjects with IGF-1 deficiency shows severe growth and mental retardation<sup>55</sup>; on the other hand,  
29 higher circulating levels of IGF-1, within the physiological range, are associated to better overall  
30 survival compared to subjects with lower physiological levels<sup>56</sup>.

31 The GH/IGF-1 complex signaling network regulates growth, development and differentiation in  
32 several tissues<sup>38</sup>, and also carbohydrate and lipid metabolism<sup>57-59</sup>.

1 Muscles, along with bones, represent the main target tissue of the GH/IGF-1 axis<sup>60</sup>. Acute  
2 administration of GH regulates muscle mass and metabolism by switching fuel utilization toward  
3 fat oxidation<sup>61</sup>. In fact, GH stimulates lipolysis, both at resting and during physical activity,  
4 determining a rise in plasma levels of FA<sup>62,63</sup> and reducing carbohydrate utilization in healthy and  
5 in subjects affected by GH deficiency (GHD)<sup>64,65</sup>. In GHD, lean body mass and muscle mass are  
6 reduced<sup>66</sup> but GH treatment ameliorates muscle protein balance by shifting amino acids from  
7 oxidative towards synthetic pathways<sup>67-69</sup>; similarly, in healthy subjects, GH supplementation  
8 determines anabolic effects, mediated by IGF-1<sup>70,71</sup>, not only by reducing amino acids oxidation  
9 but also increasing protein synthesis<sup>72,73</sup>. The GH-mediated changes in body protein metabolism is  
10 time-dependent with a return to basal protein turn-over within few weeks<sup>74</sup>.

11 The GH/IGF-1 axes contributes also to age-dependent sarcopenia: in muscles, aging is associated  
12 to a decrement in IGF1R expression and phosphorylation<sup>75</sup>, and to a reduced GHR mRNA  
13 expression and to a rise in myostatin expression<sup>76</sup>; in fibroblasts, aging determines a reduction of  
14 DNA synthesis and of cell proliferation<sup>77</sup>.

15 Controversial results have been reported regarding GH/IGF-1 role in increasing muscle strength,  
16 muscle protein synthesis, fatty acid availability and in sparing of glycogen stores<sup>50,63,78</sup>, due to the  
17 lack of convincing evidences supporting a direct effect on skeletal muscle. Nevertheless, GH  
18 and/or IGF-1 are assumed, at supraphysiological concentrations, alone or in combination with  
19 AASs as doping agents<sup>79</sup>; both GH and IGF-1 are included in the WADA list of banned drugs.

20 Little is known about adverse effects of long-term misuse of GH/IGF-1 at supraphysiological  
21 concentrations. Chronic administration for longer periods in healthy athletes may lead to the  
22 clinical features associated with acromegaly<sup>79</sup>. Many studies reported that high serum levels of  
23 GH/IGF-1 play a key role in CVD risk<sup>80</sup>. A recent study conducted on a large population of middle-  
24 aged healthy subjects showed a significant association between higher fasting serum GH-levels  
25 and CVD mortality and morbidity risk in man<sup>81</sup>. Moreover, a positive correlation was also observed  
26 between slight increments in circulating IGF-1 and the incidence of prostate and colorectal  
27 cancers<sup>42,82</sup>.

28 In order to define a clear map of the fuzzy network of actions determined by supraphysiological  
29 exogenous administration of GH/IGF-1 on human health, we review the current understanding of  
30 their molecular effects, considering recent works focused only on supraphysiological treatments  
31 and their consequences at mRNA and protein level (Table S2).

32

## 1 **Molecular effects of supraphysiological doses of T and DHT**

2

### 3 ***Vascular effects***

4 Testosterone is a vasoactive hormone that predominantly has vasodilatory actions on several  
5 vascular beds<sup>83,84</sup> in a variety of species<sup>85-89</sup>. The proposed molecular mechanism, underlying this  
6 action, involve either Ca<sup>2+</sup> channels and K<sup>+</sup> channels<sup>90</sup>.

7 Hoppe and co-authors observed a distinct chronic vs acute supraphysiological T effects on single  
8 cardiac T-type Ca<sup>2+</sup> channels from neonatal rat ventricular cardiomyocytes<sup>91</sup>. In particular, the  
9 chronic supraphysiological treatment (100 nM and 10 μM for 24-30 h) determined an increase in  
10 the whole cell T-type calcium current (I<sub>Ca,T</sub>) density and in the beating frequencies, supported by  
11 an increased expression of pore-forming subunits Ca<sub>v</sub>3.1 and Ca<sub>v</sub>3.2, both at protein and mRNA  
12 levels. Conversely, the administration of acute supraphysiological doses (10 μM) determined a  
13 decrease in the I<sub>Ca,T</sub> current density. Interestingly, the non-genomic acute administration of T on T-  
14 type Ca<sup>2+</sup> channel antagonized the genomic-dependent chronic effect. Such findings were also  
15 confirmed in adult rat ventricular cardiomyocytes<sup>92</sup>, where similar effects were observed on the L-  
16 type Ca<sup>2+</sup> channels in the chronic (100 nM) and in the acute (100 nM) treatment. Hoppe and co-  
17 authors speculate that the non genomic action of T could be explained by its lipophilic nature: in  
18 fact, they propose that T might act through the lipid phase of the membrane close to the T- and/or  
19 L-type Ca<sup>2+</sup> channels, similarly to other antagonists of calcium channels.

20 A different effect of T on voltage-dependent Ca<sup>2+</sup> channels was reported by Peers and co-authors  
21 <sup>93</sup> They evaluated the effects of a physiological (1nM) vs supraphysiological (IC<sub>50</sub>=275±0.7 nM) T  
22 administration on a rat aortic smooth muscle cell line, A7r5. These authors observed that, at 1 nM,  
23 T inhibited L-type Ca<sup>2+</sup> channels, whereas, at high doses, also T-type Ca<sup>2+</sup> channels were  
24 inhibited<sup>93</sup>. Such results differed from Hoppe's group, probably due to the different muscle tissue  
25 analyzed in their respective studies (striated muscle vs smooth muscle); anyway, both agreed in  
26 suggesting a direct interaction of T with pore-forming Ca<sup>2+</sup> channel subunits in muscle tissues<sup>94</sup>.

27 In general, the deregulation of calcium homeostasis is often a signal of adverse events and,  
28 recently, it has been linked to all cancer hallmarks<sup>95,96</sup>. Hence, we can speculate that  
29 supraphysiological T levels might trigger a detrimental cascade of molecular events in muscle  
30 tissues, by interfering with calcium current density, eventually leading to major risks of cancer  
31 development.

1 Regarding the effects mediated by T on  $K^+$  channels regulation, Hoppe and co-authors<sup>97</sup>  
2 demonstrated a cytoprotective effect of T from ischemic cell death in rat ventricular myocytes.  
3 The authors reported that supraphysiological doses of T (10  $\mu$ M) protected cardiomyocytes  
4 against ischemic injury by opening mitoK<sub>ATP</sub> channels and allowing the oxidation of mitochondrial  
5 flavoproteins. Similarly, at supraphysiological concentrations (up to 300  $\mu$ M), T causes potent and  
6 rapid vasorelaxation by activating K<sub>ATP</sub> channels in the human radial artery<sup>98</sup> and by inhibiting Ca<sup>2+</sup>  
7 influx in rat aorta<sup>99</sup>, leading to the hypothesis that T stimulates the K<sub>V</sub> and K<sub>Ca</sub> channels in large  
8 conductance vessels and, conversely, the K<sub>ATP</sub> channel, in small resistance vessels<sup>98</sup>.

9 Vasodilation affecting K<sup>+</sup>- and/or Ca<sup>2+</sup> channels is an endothelium-independent process; on the  
10 other hand, this phenomenon can occur also through an endothelium-dependent mechanism that  
11 includes the production of endothelium-derived vasoactive peptides. At supraphysiological level,  
12 T/DHT plays a role also in this context. Infact, T (up to 3,5  $\mu$ M) induces an increase both in  
13 adrenomedullin (ADM)-secreting endothelial cells and in ADM mRNA expression in a  
14 concentration-dependent manner in human aortic endothelial cells<sup>100</sup> (HAECs). On the other hand,  
15 high levels of T determines an increase in endothelin-1 (EDN1)-secreting endothelial cells and in  
16 the correspondent EDN1 mRNA expression<sup>101</sup>. ADM and EDN1 have potentially contrasting actions  
17 on vascular smooth cells, being the first a potent vasodilator<sup>102,103</sup> and the latter a vasoconstrictor  
18 peptide<sup>104</sup>; nevertheless, they are similarly regulated by high levels of T in HAECs. Taking in  
19 account such findings, it can be considered that, unlike the physiological levels, supraphysiological  
20 doses of T affect vascular activity in a complex manner, and it is not easy to predict any specific  
21 vascular effect.

22 An altered vascular responsiveness to hormonal stimuli is considered an hallmark of  
23 atherosclerosis. The genesis of an atherosclerotic plaque is a complex process that involves several  
24 factors (genetic, enviromental and/or pathophysiological)<sup>105</sup>. A specific set of key risk biomarkers,  
25 such as central abdominal obesity, high levels of triglycerides, elevated low-density lipoproteins  
26 (LDL) and reduced high-density lipoproteins (HDL), defines the main features predisposing to  
27 atherosclerotic plaque formation, a process closely interconnected with the action of pro-  
28 inflammatory cytokines.

29 Androgens have long been considered major contributors to the risk of atherosclerosis. For  
30 example, when rhesus monkeys were submitted to repeated injection of T (bimonthly  
31 administration; 50 mg/inj; 32 months), to increase serum levels of into supraphysiological range,  
32 LDL increased from the 12<sup>th</sup> month, whereas HDL decreased constantly during the treatment in

1 treated animals<sup>106</sup>. At the same time plasma glutamate oxaloacetate transaminase (SGOT) and  
2 plasma glutamate pyruvate transaminase (SGPT) levels increased and remained elevated up to the  
3 end of treatment.

4 Bhasin and co-authors monitored T effects on eugonadal healthy men (18-35 yrs) by weekly  
5 injections varying from low physiological (25 mg/inj/wk; 20 wks) to supraphysiological (600  
6 mg/inj/wk; 20 wks) concentrations. Participants received monthly injections of a long-acting  
7 GnRH agonist to suppress endogenous T production. Bhasin demonstrated that only at high doses  
8 T induced a decrease of HDL and apolipoprotein A1<sup>107</sup>. Similarly, Herbst and coll administered T  
9 (600 mg) weekly for 3 weeks to elderly, obese, eugonadal men and observed a reduction of HDL  
10 and SHBG together with an increase of LDL density and Hepatic Lipase (HL) activity. The rise in HL  
11 activity was responsible for the conversion of HDL<sub>2</sub> to the denser HDL<sub>3</sub>, leading to the reduction of  
12 HDL<sup>108</sup>.

13 Despite such findings, new emerging evidences are pointing out that T may play a protective role  
14 in vascular health, as antagonist of the atherosclerotic process. Indeed, clinical and  
15 epidemiological studies confirm that low plasma T levels are positively associated to  
16 atherosclerosis<sup>105,109,110</sup>. Furthermore, Androgen Deprivation Therapy (ADT) in prostate cancer  
17 patients determines an increase of total cholesterol (TC), LDL and triglycerides in serum<sup>111,112</sup>;  
18 conversely, T replacement therapy in hypogonadal men induces a decrement of serum TC, LDL and  
19 triglycerides and promotes an increase in HDL<sup>113</sup>. Nevertheless, only few reports addressed the  
20 role of supraphysiological doses of T on atherosclerosis in healthy subjects. Langer and co-  
21 authors<sup>114</sup> demonstrated that T, both at physiological (10 ng/ml) and at supraphysiological  
22 concentrations (100 ng/ml), increases the expression of the scavenger receptor B1 (SR-B1) mRNA  
23 and protein in human hepatocyte cell line (HEPG2) and in human monocyte-derived macrophages  
24 (HMMDM) cells in a dose-dependent manner. SR-B1 is known to mediate selective uptake of HDL-  
25 derived cholesterol and cholesteryl ester into the liver and in steroidogenic tissues<sup>115</sup>. Authors  
26 hypothesized that T plays a protective role in that context as it intensifies reverse cholesterol  
27 transport, by facilitating the transport of excess cholesterol from atherosclerotic plaques of  
28 arterial wall to the liver<sup>114</sup>. Likewise, an anti-inflammatory role for T was hypothesized by Corcoran  
29 and co-authors<sup>116</sup>: they observed that supraphysiological doses of T significantly reduced the  
30 expression of the pro-inflammatory cytokines TNF- $\alpha$  and IL-1 $\beta$  in primary HMMDM cell cultures  
31 treated with moderately oxidized LDL (50 mg/ml, 48 hours).

1 Recently, Wan and co-authors<sup>117</sup> reported that supraphysiological levels of DHT (1  $\mu$ M)  
2 determined a converse effect on human umbilical vein endothelial cells (HUVEC) in presence or  
3 absence of lipopolysaccharide (LPS), a powerful bacterial virulence proinflammatory factor. It was  
4 observed that, in absence of LPS, DHT induced a significantly downregulation of urocortin (UCN1)  
5 mRNA and protein expression through an AR-dependent mechanism; conversely, in presence of  
6 LPS, UCN1 mRNA and protein expression increased in HUVEC through an AR-independent  
7 mechanism, involving p38/MAPK, ERK1/2 and NF- $\kappa$ B activation. UCN1 is a neuropeptide belonging  
8 to the corticotropin-releasing factor (CRF), up-regulated by inflammatory cytokines<sup>118</sup>, and  
9 involved in the vascular inflammatory process. Hence, such opposing experimental evidences  
10 suggest that supraphysiological DHT levels exert on vascular cells a differential action, based on  
11 the inflammatory status: T/DHT administration would not induce inflammation *per se*, but, it could  
12 be able to amplify the pro-inflammatory effect of LPS. Also Annibalini and co-authors confirmed  
13 recently that the role on inflammation of sex steroids (T, DHT and E2) is dependent on the  
14 inflammatory status of the system under investigation<sup>119</sup>. In fact, they demonstrated that the  
15 inflammatory action of TNF- $\alpha$  is magnified by co-administration of supraphysiological doses of T  
16 (up to 1  $\mu$ M) or DHT (100 nM), by increasing the TNF- $\alpha$ -induced vascular cell adhesion molecule 1  
17 (VCAM-1) gene expression; conversely, in absence of TNF- $\alpha$  stimulation, T was unable to modify  
18 significantly the expression pattern of VCAM-1 gene. These results are in contrast with  
19 Hatakeyama's findings<sup>120</sup>. This latter observed that a similar supraphysiological T treatment (100  
20 nM and 1  $\mu$ M) on HAECs determined a reduction of TNF- $\alpha$ -induced VCAM-1 expression. The  
21 controversial results can be ascribed to the different endothelial system (HUVEC<sup>119</sup> vs HAEC<sup>120</sup>)  
22 and to the different TNF- $\alpha$  concentration (1 ng/ml<sup>119</sup> vs 20 ng/ml<sup>120</sup>), used by the two research  
23 groups. Anyway, such opposing data underline the complexity of the cross-talk among biologically  
24 active species, above all when supraphysiological concentrations of hormones are taken into  
25 account.

26

### 27 **Apoptosis**

28 Apoptotic damage of vascular endothelial cells is a key event in atherogenesis. Testosterone and  
29 its metabolites are able to shift the balance toward cell survival or apoptosis through highly  
30 orchestrated mechanisms, not completely elucidated yet.

31 Ling and co-authors reported that hyperandrogenic states (up to 100 nM) induced apoptosis in  
32 serum free HUVECs, through the detection of multiple apoptosis-associated determinants, such as

1 the reduction of DNA synthesis and Bcl-2 expression, and, on the other hand, the increase in the  
2 number of apoptotic cells and in genomic-DNA fragments<sup>121</sup>. Supraphysiological T doses (100 nM)  
3 on HUVEC determined an alteration of endothelial cell growth with a strong anti-proliferative  
4 effect, leading to apoptosis and affecting intracellular Ca<sup>2+</sup> levels<sup>122</sup>. Similarly, Powazniak and co-  
5 authors on HUVECs confirmed that supraphysiological T concentrations (0.1-9.6 μM) promoted  
6 the activation of JNK and p38/MAPK pathways, causing apoptotic cell death<sup>123</sup>. Also Kayampilly  
7 and co-authors observed that hyperandrogenic DHT states reduced cyclin D2 mRNA expression  
8 and inhibited granulosa cell proliferation, all events being mediated by AMPK activation<sup>124</sup>.  
9 Similarly, the same authors showed that DHT activates AMPK in a time and dose-dependent  
10 manner and reduces FSH-mediated mitogenic signal, leading to the inhibition of granulosa cell  
11 proliferation<sup>125</sup>. Furthermore, Verzola and co-authors<sup>126</sup> confirmed a pro-apoptotic behavior of  
12 supraphysiological doses of T (up to 1 μM) on immortalized human proximal tubular epithelial cell  
13 line (HK-2) and in primary human proximal tubular epithelial cells (PTECs), through upregulation of  
14 Fas, FasL and FADD and activation of caspase-dependent apoptotic pathway.

15 In neurons, T acts as a neurosteroid determining both neuroprotection and neurodegeneration<sup>127-</sup>  
16 <sup>129</sup>. Also in these cell systems T can affect intracellular Ca<sup>2+</sup> concentration<sup>130</sup>, and it is known that  
17 prolonged elevated cytosolic calcium concentrations can initiate the apoptotic program in many  
18 cell types<sup>131,132</sup>. Estrada and co-authors evaluated the effect of acute supraphysiological T  
19 treatments (100 nM, 1 μM, 10 μM) in a human neuroblastoma cell line (SH-SY5Y).  
20 Hyperandrogenic states in neuroblastoma cells induced a decrease in cell viability, an increase in  
21 DNA fragmentation and the activation of caspase, triggering apoptotic cell death<sup>133</sup>; moreover an  
22 activation of inositol 1,4,5-triphosphate receptor (InsP<sub>3</sub>R) was also observed<sup>130</sup>.

23 Conversely, to such an amount of experimental evidence stating the proapoptotic action of T, a  
24 parallel literature convincingly demonstrates its ability to suppress cell death and promote cell  
25 survival. Erkkila and co-authors reported that high T levels (100 nM, 1 μM) suppressed apoptosis  
26 in seminiferi tubules from human testis tissues *in vitro*, indicating that T play a critical role in germ  
27 cell survival<sup>134</sup>. Ahlbom and co-authors showed that cerebellar granule cells (CGCs) obtained from  
28 7-day old rat pups, pretreated *in vivo* with T (500 μg/0.05 ml injection), are selectively protected *in*  
29 *vitro* from apoptosis induced by oxidative stress (H<sub>2</sub>O<sub>2</sub> or S-nitrosocysteine)<sup>135</sup>; these observations  
30 were associated to an increased activity of two of the major antioxidant enzymes, SOD and  
31 catalase. Similarly, CGCs, treated *in vitro* with supraphysiological doses of T (1 μM), were less  
32 susceptible to oxidative challenges showing up-regulation of cellular antioxidant defences through

1 an AR-dependent mechanism<sup>136</sup>. Likewise, Pike<sup>137</sup> demonstrated that T or DHT (up to 100 nM)  
2 conferred neuroprotection from cell death induced by  $\beta$ -amiloid peptide into PC12 cells and into  
3 primary hippocampal neurons cultures from Sprague-Dawley rat pups. These results can partly  
4 explain the increased vulnerability of an aged brain to neurodegenerative disorders, such as  
5 Alzheimer's disease (AD), and the age-related decline of circulating T levels in elderly men<sup>137</sup>.  
6 Nowadays, it is clear that T increases neuronal resilience against AD-related injuries and it is used  
7 as anti-aging drug<sup>138</sup>, but more studies are needed to define the molecular mechanisms involved  
8 and to optimize the hormone therapy<sup>139</sup>.  
9 More recently, Imperlini and co-authors found out that chronic supraphysiological DHT treatment  
10 (0.7  $\mu$ g/ml total concentration in three doses) on primary human peripheral blood lymphocytes  
11 (PBLs) from male healthy donors, induced an anti-apoptotic effect 7d after the first treatment<sup>140</sup>.  
12 In fact, it was observed an over-expression of the pro-survival factor Bcl-2 and a reduced  
13 activation of pro-apoptotic caspase-3 in the treated cells compared to the untreated PBLs. Indeed  
14 that proteomic study pointed out that the steroid treatment affected the expression profile of  
15 more than 30 protein species, half of which were related to apoptosis. Similarly, the same anti-  
16 apoptotic effect was registered in human PBLs when DHT treatment was associated to a single  
17 IGF-1 supraphysiological administration<sup>141</sup>. Such a double treatment mimicks the ability of  
18 supraphysiological T treatments in increasing serum IGF-1 levels in healthy young men<sup>142</sup> and  
19 indicates that DHT+IGF-1 hyperstimulation affects cell adhesion, migration and survival through  
20 both downregulation of cytokines and paxillin signaling-related proteins, and activation of several  
21 pathways downstream FAK.

22

### 23 **Biomarkers of T/DHT doping**

24 Supraphysiological doses of T and/or DHT are assumed by athletes and amateurs to enhance sport  
25 performance and/or to obtain a better body shape. Up to now it is difficult to find convincing  
26 reports describing genomic effects of T/DHT chronic treatment on the health of young sportsmen;  
27 sometimes, few gene information can be gained by studies planned with different aims, even if  
28 those data need to be confirmed in specific validation assays. But recently, Mancini and co-authors  
29 performed a chronic supraphysiological DHT treatment (0.7  $\mu$ g/ml total concentration in three  
30 doses) on human PBLs and analysed the differentially expressed genes by a transcriptomic  
31 approach in order to define a putative set of biomarkers of steroid doping<sup>143</sup>. In this study, authors  
32 reported that 275 genes (210 up-regulated, 65 down-regulated) were differentially expressed 7

1 days after the first treatment, most of them matching significantly the “Skeletal and Muscular  
2 Disorder” category according to the Ingenuity Pathway Analysis database. The most upregulated  
3 genes in this dataset were IDO1, CXCL13, CCL1, GZMB, VDR and IL2RA.

4

5 Several factors concur in shaping the effects of supraphysiological doses of T/DHT: i)the age, ii)the  
6 gender, iii)the organ system (tissue and source), status and enzymatic profile, iv)the type, dose  
7 and duration of a T/DHT treatment, v)the endogeneous T/DHT levels and vi) the putative  
8 interactions with other biologically active molecules (es.: hormones, cytokines) or endocrine  
9 tissues (es.: adipose tissue). Each parameter modify heavily the outcome of any single treatment;  
10 such a multiparameter context explains the extreme variability in published reports. Such a  
11 variability gives rise to the present confounding information about the effect of supraphysiological  
12 doses of T/DHT, able both to trigger and to inhibit pivotal cellular pathways/functions  
13 (vasoactivity, calcium homeostasis, atherosclerosis, inflammation, apoptosis; Fig. 1). Indeed more  
14 molecular studies are needed to better define the complex array of effects determined by T/DHT  
15 on several organs and apparatus.

16

17

## 1 **Molecular effects of supraphysiological doses of GH and IGF-1**

### 3 **Apoptosis**

4 As a growth factor, IGF-1 controls proliferation and differentiation, and protects cells against  
5 apoptosis, as demonstrated in several *in vitro* and *in vivo* systems<sup>144,145</sup>. Velazquez and co-authors  
6 investigated the effects of physiological and supraphysiological IGF-1 levels on preimplantation  
7 bovine embryos, with a particular focus on polycystic ovary syndrome (PCS), characterized by high  
8 levels of serum IGF-1. These authors found out that at physiological levels (from 50 to 150 ng/ml),  
9 IGF-1 did not affect apoptosis. Conversely, at supraphysiological concentrations (from 950 to 1500  
10 ng/ml), IGF-1 induced several biological/biochemical effects: a) increased apoptosis; b) decreased  
11 TP53 protein expression; c) increased number of cells and IGF1R protein expression in the inner  
12 cell mass (ICM)<sup>145</sup>. Such findings do not correlate with previous studies, where a downregulation  
13 of IGF1R was observed in blastocysts or mouse embryos treated with high IGF-1 levels<sup>146-148</sup>.

14 As for anti-apoptotic effects, it has been reported that treatments with 50nM of IGF-1, up to 48 h,  
15 increased T cell death-associated gene 51 (TDAG51) expression, at gene and protein level, through  
16 activation of IGF-1R and p38 MAPK pathway, in mouse embryo fibroblasts<sup>149</sup>. In particular, authors  
17 demonstrated that TDAG51 plays a regulative role in the anti-apoptotic effects of IGF-1.

18 Nevertheless, the anti-apoptotic effects of IGF-1 are dose- and system-dependent. In fact, high  
19 IGF-1 levels did not determine any apoptotic effects in human PBLs (Orrù personal  
20 communication). In this cell system, 6d after a single IGF-1 hyperstimulation, the MAPK signaling  
21 pathway was still active; in particular, p70S6K Tyr229, Tyr389 and Tyr421/Ser424 were found all  
22 phosphorylated, thus indicating that the acute *in vitro* treatment generated several sustained  
23 signaling, including those related to protein synthesis processes<sup>150</sup>. Such proteomic study showed  
24 also a consistent cytoskeletal reorganization mediated by Stat-1 and an overproduction of  
25 cytokines positively related to immune response and inflammation. All together these data  
26 indicated that, following IGF-1 hyperdosage, circulating PBLs could be more prone to  
27 transendothelial migration<sup>150</sup>.

28 In the same experimental model, an acute supraphysiological IGF-1 treatment determined the  
29 overexpression of 102 genes, involved in skeletal muscle disorders, as well as in cell-mediated  
30 immunological response<sup>143</sup>. Among these genes, the most upregulated species are fibronectin 1  
31 (FN1), involved in cell adhesion and migration processes, including host defense and metastasis<sup>151</sup>,  
32 and RAB31, an oncogene key regulator of intracellular membrane trafficking and associated to

1 breast cancer<sup>152, 153</sup>. The transcriptomic approach was also adopted by Mitchell and co-authors<sup>154</sup>  
2 to evaluate the molecular effects induced by GH abuse in PBLs isolated from male and female  
3 recreational athletes. GH treatment (2 mg/inj/die for 8 wks) induced an approximately 2-fold  
4 increase in serum IGF-1; RTqPCR validation assays confirmed an upregulation of HSPC159, ITGB3,  
5 OLFM4 and TUBB1 genes only in females.

6

### 7 **Cancer**

8 The system GH/IGF-1 has been recognized for decades for its role in tumorigenesis and  
9 growth<sup>42,155</sup>. IGF-1 plays a key role in tumour formation and proliferation. Several evidences from  
10 both humans and animal models demonstrate a link between GH/IGF-1 levels and cancer risk<sup>155</sup>.  
11 Transgenic GH overexpressing mice, characterized by elevated circulating IGF-1 levels, exhibit  
12 hepatomegaly due to hypertrophy and hyperplasia<sup>156</sup>. The cellular morphological modifications,  
13 so-called pre-neoplastic lesions, observed in the liver of GH overexpressing mice are similar to that  
14 observed in humans at high risk of liver cancer development. Hence, GH overexpression induces  
15 tumorigenesis in the liver of transgenic GH overexpressing mice by stimulating tumor cell  
16 proliferation<sup>157</sup>. Recently, Miquet JG et al. investigated the molecular pathogenesis and the signal  
17 transduction pathways related to the pro-oncogenic liver pathology induced by prolonged  
18 exposure to elevated hepatic GH levels in transgenic mice model<sup>158,159</sup>. In particular, the authors  
19 evaluated the mRNA and protein expression and the activation of several signaling mediators and  
20 effectors involved in cell growth, proliferation and survival, such as Akt2, NFκB, GSK3β, β-catenin,  
21 cyclin D1, cyclin E, c-myc, c-jun and c-fos<sup>158,159</sup>. These studies indicate that prolonged exposure to  
22 GH leads to a liver dysregulation of several oncogenic pathways similar to that observed in many  
23 human tumors.

24

### 25 **Biomarkers of GH/IGF-1 doping**

26 In the last decade, in order to clarify the doping action of the GH/IGF-1 system, few studies were  
27 based on gene doping animal models.

28 In this context, a gene doping model of GH-overexpressing rats has been recently used to evaluate  
29 both the molecular effects of GH abuse in healthy animals by using a transcriptome approach and  
30 to identify putative biomarkers for the detection of unauthorized GH gene therapy in humans. In  
31 particular, a gene expression profile was identified on PBLs from rats subjected to long-term GH  
32 gene therapy and sacrificed 24 weeks after the injections<sup>160</sup>. Sixty one genes were found

1 differentially expressed in GH gene-treated rats 24 weeks after GH gene therapy. These genes  
2 were mainly involved in processes as angiogenesis, oncogenesis, apoptosis, cardiac hypertrophy,  
3 immune networks, signaling pathways, adipocytokines, arachidonic acid metabolism, CAMs and  
4 cytokine-cytokine receptor interaction. Eight differentially expressed genes were selected as  
5 candidate biomarkers for the detection of GH abuse, after RT-qPCR validation experiments. Some  
6 of the differentially expressed genes are involved in inflammation and immunity, such as: *Pla2g2a*,  
7 a PLA2 group IIa secreted phospholipase A2 involved in many human diseases, including coronary  
8 artery disease, colon cancer and inflammation; *Rap1B*, a small GTPase involved in the platelet  
9 activation; and *Nfkbia*, the NF-kappa B inhibitor alpha, involved in the inflammatory response<sup>160</sup>.  
10 Following the same rationale, Macedo A et al. performed a proteomic study to characterize the  
11 molecular effects in transgenic IGF-1 overexpressing mice<sup>161</sup>. By delivering the IGF-1-cDNA into  
12 multiple muscles of adult animals using adeno-associated virus (AAV) vectors, the muscle whole-  
13 proteome changes were analyzed after 15 and 30 days, and they were correlated with  
14 morphological and functional modifications. The AAV-IGF-1-injected mice can be properly  
15 considered a mouse model of doping, since the measured levels of transduced IGF-1 exceed by  
16 more 100- and 10-fold those of the endogenous mouse mRNA and protein, respectively. This  
17 supraphysiological condition determined, at morphological and structure levels, a marked muscle  
18 hypertrophy, neovascularization and a fiber switch from fast to slow type. These cellular  
19 alterations are finely supported by proteomic analysis outputs: in IGF-1-transduced muscles,  
20 structural proteins involved in muscle hypertrophy and slow fiber-specific proteins were  
21 overexpressed, fast type-ones were underexpressed, and the key proteins controlling energy  
22 metabolism were upregulated. In particular, the authors suggest that following IGF-1 delivery, a  
23 transition from an anaerobic to an aerobic metabolism might occur in muscles, since some of  
24 specific enzymes, belonging to both type of energy metabolisms, are concomitantly induced, but  
25 not at the same levels. Such a novelty could have been more convincing if time points longer than  
26 30 days would have been considered.

27 Although the clinical effects of supraphysiological IGF-1 treatment are well documented, a  
28 comparable production of molecular studies on short- and long-term effects on healthy subjects is  
29 still missing. In this field, the published papers are mainly aimed to discover new biomarkers for  
30 detecting GH/IGF-1 doping. In this context, the unique data, at molecular level, are restricted to  
31 the study of effects on the serum levels of IGFBP; for example, it has been reported that IGFBP-4

1 and IGFBP-5 are increased in healthy adults during one month's treatment with supraphysiological  
2 GH doses<sup>162</sup>.

3

4 Many questions still remain to be clarified before accurate and reliable methods for doping  
5 detection are found. Certainly, in this field, further studies with a large number of subjects are  
6 needed. At moment, a very limited number of studies assess the molecular modifications induced  
7 by high doses of GH/IGF-1 over time in healthy in vitro and in vivo systems. However, those  
8 studies often describe GH/IGF-1 effects in pathological rather than healthy condition, at  
9 physiological and/or pharmacological concentrations, values that are lower compared to the doses  
10 assumed by abusers. Despite this, the current understanding of the molecular effects of GH/IGF-1  
11 abuse shows that supraphysiological doses affect cell function such as apoptosis and cytoskeletal  
12 reorganization, and they have implications on the inflammation response and on the skeletal  
13 muscle system (Fig. 2).

14

15

## 1 **Future perspective**

2

3 The extreme heterogeneity of data published so far, regarding the effects of doping agents on  
4 health, mirrors the scanty knowledge of PED administration protocols as well as the sparse  
5 information on their biological activity at supraphysiological concentrations. There is still a long  
6 way to solve such a puzzling tangle, and in this scenario metabonomics studies can provide the  
7 missing piece that will allow to gain complementary information to genomics and proteomics. In  
8 fact, metabonomics, as defined by Nicholson in 1999, represents “the quantitative measurement  
9 of the time-related multiparametric metabolic response of living systems to patho-physiological  
10 stimuli or genetic modification”<sup>163</sup>. This approach uses biofluids (urine, plasma, seminal fluid,  
11 cerebrospinal fluid, saliva and others)<sup>164</sup> and is mainly based on NMR spectroscopy and/or mass  
12 spectrometry procedures<sup>165</sup>.

13 Over the past few years, metabonomic protocols were also applied to the doping field. Dumas and  
14 co-authors were the first to demonstrate the potentiality of metabonomics as a tool for the  
15 analysis of metabolic perturbations induced by doping agents<sup>166</sup>; they revealed the biological  
16 signature of AASs in cattle by using NMR spectroscopy, and, in particular, they found out that the  
17 urinary biomarkers, at supraphysiological level, are associated to nitrogen and energy metabolism.  
18 Afterwards, advances in mass spectrometry (MS) and bioinformatics contributed to the  
19 development of untargeted metabonomics to investigate the urine profiling following doping  
20 agents administration in animals. In this context, Kieken and co-authors developed a  
21 metabonomic approach based on liquid chromatography-electrospray-high resolution mass  
22 spectrometry (LC-ESI-HRMS) to compare horse urine fingerprints collected before and after  
23 treatment with recombinant equine growth hormone (reGH)<sup>167</sup>. About 20 metabolites detected by  
24 different mass/charge values and retention times were selected as potential biomarkers of GH  
25 abuse; interestingly, the results of this preliminary study showed a long-term effect of reGH,  
26 demonstrating global modifications of horse urine metabolome mostly 25 days after the first  
27 administration. However, this pioneering study remains mainly technical, since the identity of the  
28 metabolites responsible for the discrimination between treated and non-treated animals are  
29 nowadays unknown. A similar approach based on ultraperformance liquid chromatography in  
30 combination with time-of-flight accurate mass spectrometry (UPLC-TOFMS) was used to analyze  
31 urine profiles of bovine treated with AASs<sup>168</sup>. In this paper, metabolites, differentially regulated  
32 following doping agents administration, were partially identified by accurate mass data and

1 retention time comparison with commercially available standards or by LC-LTQ-Orbitrap tandem  
2 MS.

3 However, the purpose of these early studies was to set up and assess MS-based metabonomic  
4 strategies as new screening tools for doping agents abuse, thus demonstrating the feasibility of  
5 such approaches<sup>169,170</sup>.

6 Recently it has been described a semi-automated strategy for the annotation (based on  
7 experimental masses and retention times) of metabolites in global fingerprints acquired from  
8 untargeted metabonomics approach from tissue samples of bovine treated with AASs<sup>171</sup>; this  
9 implementation, in fact, requires specific softwares and is essential for metabolite identification,  
10 thus representing the major challenge for the feasibility of metabonomic approach.

11 The application of metabonomics to the doping in humans is mainly focused on the technological  
12 advancements and on the search for the most sensitive protocols serum<sup>172,173</sup>; few reports are  
13 focused on the potential risk of PED consumption for human health. One example comes from  
14 West and co-authors, who assessed the risk of human exposure to endocrine active compounds,  
15 such as T, in human germ-like cells (GLCs)<sup>174</sup>. At the highest dose tested (100  $\mu$ M), all the steroid  
16 hormones determined a decrease in intracellular level of amino acids and an increase in  
17 metabolites related to cellular energetics and metabolism, such as glucose and lactate. Moreover,  
18 a decrement in the degree of fatty acid saturation and in C14-C20 fatty acids was observed; finally,  
19 at 100  $\mu$ M T caused a reduction of cholesterol and cholesterol-derivatives.

20 The low number of metabonomic studies evaluating the molecular effects of doping agents on  
21 health is related both to ethical concerns and to the great effort in setting up the experimental  
22 design. In fact, to perform a metabonomic investigation, whatever the aim pursued, tools,  
23 procedures and methods need to be finely scheduled, ranging from the sample collection and  
24 preparation to the generation of metabolic profiles/fingerprints, from the raw data processing to  
25 the bioinformatic/statistical analysis for metabolite identification.

26 In the field of doping, a successful experimental design depends on the type of sample (plasma or  
27 urine) and on the sample collection from PED abusers and from a control-non doped population.  
28 Urine samples are, of course, most feasible and easy to collect rather than plasma, especially in a  
29 context of amateur or professional sporting event. However, both types of samples are  
30 informative about metabolic changes resulting from doping agents abuse: urine, in fact, is a rich  
31 source of hydrophilic metabolites; the plasma composition, on the other hand, is more stable and  
32 representative of other kind of molecules.

1 The sample collection represents a challenging issue in a metabonomic study applied to the  
2 analysis of doping agents molecular effects in healthy subjects. In fact, metabolic profiles are  
3 influenced by fitness condition besides to sex, food consumption, environmental context as well as  
4 by individual genetic profile; as a consequence, the critical step of such experimental designs is  
5 related to the definition of exclusion/inclusion criteria for volunteers enrollment in both groups,  
6 doped and control. A prerequisite of a good study design is having strong matched-groups in order  
7 to observe exclusively doping-dependent phenomena and, hence, to identify putative biomarkers  
8 in anti-doping analysis.

9

10

## 1 **Conclusions**

2

3 AASs and growth factors constitute the most popular prohibited substances among abusers, and it  
4 is now increasingly frequent to run into administration protocols that mix molecules belonging to  
5 both classes. The consequences on human health determined by a short and/or long-term PED  
6 misuse are partly known but the molecular mechanisms underlying such adverse events are still  
7 unclear, and sometime confounding. Here, we reviewed the supraphysiological effects of T/DHT or  
8 GH/IGF-1 on different cellular pathways/functions, and discussed the consequences of both  
9 treatments strictly associated to inflammation processes and apoptosis. In general, the growth  
10 factors, at supraphysiological concentrations, exert mainly anti-apoptotic and/or pro-inflammatory  
11 effects in different biological systems; conversely, the actions produced by hyperandrogenic states  
12 are less defined and sometimes confounding. Surely, several endogeneous and exogeneous  
13 factors have to be considered in order to clarify the molecular mechanisms responsible for health  
14 risk factors related to PED abuse; further investigations, including metabonomic studies, are  
15 needed to define new biomarkers related to the emergent issue of doping-related-dysfunctions.

16

1

2 **Acknowledgement**

3 This work was supported by a Fondazione SDN-IRCCS grant to SO and PB, by Ministero della Salute  
4 (Roma) prot. DGRST. 8/9560/I92c/05/P to PB.

5 The authors have declared no conflicts of interest.

6

7

1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32

## References

1. F. Sjöqvist, M. Garle and A. Rane, *Lancet*, 2008, **371**, 1872.
2. N. Guha, A. Dashwood, N. J. Thomas, A. J. Skingle, P. H. Sönksen and R. I. Holt, *Curr. Drug Abuse Rev.*, 2009, **2**, 263.
3. D. M. Kelly and T. H. Jones, *Front. Horm. Res.*, 2014, **43**, 1.
4. L. Klotz, *Nat. Rev. Urol.*, 2015, **12**, 48.
5. J. K. Perry, D. X. Liu, Z. S. Wu, T. Zhu and P. E. Lobie, *Curr. Opin. Endocrinol. Diabetes Obes.*, 2013, **20**, 307.
6. J. H. Nascimento and E. Medei, *Mini Rev. Med. Chem.*, 2011, **11**, 425.
7. C. R. Palmeiro, R. Anand, I. K. Dardi, N. Balasubramaniyam, M. D. Schwarcz and I. A. Weiss, *Cardiol Rev.*, 2012, **20**, 197.
8. S. Achar, A. Rostamian and S. M. Narayan, *Am. J. Cardiol.*, 2010, **106**, 893.
9. F. Kadi, *Br. J. Pharmacol.*, 2008, **154**, 522.
10. W. Gao, C. E. Bohl and J. T. Dalton, *Chem. Rev.*, 2005, **105**, 3352.
11. M. Oettel, *Aging Male*, 2003, **6**, 230.
12. E. C. Bolton, A. Y. So, C. Chaivorapol, C. M. Haqq, H. Li and K. R. Yamamoto, *Genes development* 2007, **21**, 2005.
13. M. Beato and J. Klug, *Hum. Reprod. Update* 2000, **6**, 225.
14. H. V. Heemers and D. J. Tindall, *Endocrine Reviews* 2007, **28**, 778.
15. V. Dubois, M. Laurent, S. Boonen, D. Vanderschueren and F. Claessens, *Cell Mol. Life Sci.* 2012, **69**, 1651.
16. W. L. Wang, N. Chatterjee, S. V. Chittur, J. Welsh and M. P. Tenniswood, *Mol. Cancer*, 2011, **10**, 58.
17. D. Delić, C. Grosser, M. Dkhil, S. Al-Quraishy and F. Wunderlich, *Steroids*, 2010, **75**, 998.
18. S. Panneerdoss, Y. F. Chang, K. C. Buddavarapu, H. I. Chen, G. Shetty, H. Wang, Y. Chen, T. R. Kumar and M. K. Rao, *PLoS One*, 2012, **7**, e41146.
19. S. Nielsen, T. Hvid, M. Kelly, B. Lindegaard, C. Dethlefsen, K. Winding, N. Mathur, C. Scheele, B. K. Pedersen and M. J. Laye, *Front. Physiol.*, 2014, **4**, 394.
20. G. Pandini, R. Mineo, F. Frasca, C. T. Roberts, M. Marcelli, R. Vigneri and A. Belfiore, *Cancer Res.* 2005, **1**, 1849.

- 1 21. C. D. Foradori, M. Weiser and R. J. Handa, *Front. Neuroendocrinol.* 2008, **29**, 169.
- 2 22. Migliaccio, G. Castoria, M. Di Domenico, A. de Falco, A. Bilancio, M. Lombardi, M. V.
- 3 Barone, D. Ametrano, M. S. Zannini, C. Abbondanza and F. Auricchio, *Embo J.*, 2000, **19**,
- 4 5406.
- 5 23. S. Kousteni, T. Bellido, L. I. Plotkin, C. A. O'Brien, D. L. Bodenner, L. Han, K. Han, G. B.
- 6 DiGregorio, J. A. Katzenellenbogen, B. S. Katzenellenbogen, P. K. Roberson, R. S. Weinstein,
- 7 R. L. Jilka and S. C. Manolagas, *Cell*, 2001, **104**, 719.
- 8 24. J. Cheng, S. C. Watkins and W. H. Walker, *Endocrinology*, 2007, **148**, 2066.
- 9 25. L. Bjornstrom and M. Sjoberg, *Molecular Endocrinology* 2005, **19**, 833.
- 10 26. C. E. Yesalis (ed), (1993). *Anabolic Steroids in Sport and Exercise*, Human Kinetics,
- 11 Champaign, IL, USA, pp 49.
- 12 27. F. Hartgens and H. Kuipers, *Sports Med.*, 2004, **34**, 513.
- 13 28. I. Velazquez and B. P. Alter, *Am. J. Hematol.* 2004, **77**, 257.
- 14 29. R. Modlinski and K. B. Fields, *Curr. Sports Med. Rep.*, 2006, **5**, 104.
- 15 30. van Amsterdam, A. Opperhuizen and F. Hartgens, *Regul. Toxicol. Pharmacol.* 2010, **57**, 117.
- 16 31. G. L. de Souza and J. Hallak, *BJU Int.*, 2011, **108**, 1860.
- 17 32. S. Scaccianoce, A. Caruso, J. Miele, R. Nisticò and F. Nicoletti, *J. Biol. Regul Homeost.*
- 18 *Agents*, 2013, **27**, 107.
- 19 33. M. A. Santos, C. V. Oliveira and A. S. Silva, *Subst Use Misuse*, 2014, **49**, 1132.
- 20 34. D. Wilson, *Endocr. Rev.*, 1988, **9**, 181.
- 21 35. S. Pathak, R. Singh, R. D. Verschoyle and P. Greaves, P. B. Farmer, W. P. Steward, J. K.
- 22 Mellon, A. J. Gescher and R. Sharma. *Cancer Lett.*, 2008, **261**, 74.
- 23 36. M. B. Davidson, *Endocrine Rev.*, 2010, **205**, 201.
- 24 37. J. Brooks and M. J. Waters, *Nat. Rev. Endocrinol.*, 2010, **6**, 515.
- 25 38. Giustina, G. Mazziotti and E. Canalis, *Endocrine Rev.*, 2008, **29**, 535.
- 26 39. Moran, D. R. J. Jacobs, J. Steinberger, P. Cohen, C. Hong, R. Prineas and A. R. Sinaiko, *J. Clin.*
- 27 *Endocrinol. Metab.*, 2002, **87**, 4817.
- 28 40. E. Corpas, S. M. Harman, M. A. Pineyro, R. Roberson and M. R. Blackman, *J. Clin. Endocrinol.*
- 29 *Metab.*, 1993, **76**, 134.
- 30 41. L. Laviola, A. Natalicchio and F. Giorgino. *Curr. Pharm. Des.*, 2007, **13**, 663.
- 31 42. M. Pollak, *Nat. Rev. Cancer*, 2012, **12**, 159.
- 32 43. G. Goldspink, *J. Musculoskelet. Neuronal. Interact.*, 2004, **10**, 397.

- 1 44. S. M. Firth and R. C. Baxter, *Endocr. Rev.*, 2002, **23**, 824.
- 2 45. L. A. Bach, S. J. Headey, and R. S. Norton, *Trends Endocrinol. Metab.*, 2005, **16**, 228.
- 3 46. R. C. Baxter, *Nature Reviews Cancer*, 2014, **14**, 329.
- 4 47. J. Brooks, J. W. Wooh, K. A. Tunny and M. J. Waters, *Int. J. Biochem. Cell Biol.*, 2008, **40**,
- 5 1984.
- 6 48. T. P. J. Garrett, N. M. McKern, M. Z. Lou, M. J. Frenkel, J. D. Bentley, G. O. Lovrecz, T. C.
- 7 Elleman, L. J. Cosgrove and C. W. Ward, *Nature*, 1998, **394**, 395.
- 8 49. N. J. Lanning and C. Carter-Su, *Rev Endocr. Metab. Disord.*, 2006, **7**, 225.
- 9 50. G. P. Baumann, *Endocr. Rev.*, 2012, **33**, 155.
- 10 51. A. Denley, L. J. Cosgrove, G. W. Booker, J. C. Wallace and B. E. Forbes, *Cytokine Growth*
- 11 *Factor Rev.*, 2005, **16**, 421.
- 12 52. A. Denley, J. M. Carroll, G. V. Brierley and L. Cosgrove, *Mol. Cell. Biol.*, 2007, **27**, 3569.
- 13 53. D. R. Clemmons, *Trends Endocrinol Metab.*, 2009, **20**, 349.
- 14 54. J. Tureckova, E. M. Wilson, J. L. Cappalonga and P. Rotwein, *J. Biol. Chem.*, 2001, **276**,
- 15 39264.
- 16 55. X. L. Chen, K. Lee, D. L. Hartzell, R. G. Dean, G. J. Hausman, R. A. McGraw, M. A. Della-Fera
- 17 and C. A. Baile, *Biochem. Biophys. Res. Commun.*, 2001, **283**, 933.
- 18 56. M. P. Brugts, M. B. Ranke, L. J. Hofland, K. van der Wansen, K. Weber, J. Frystyk, S. W. J.
- 19 Lamberts and J. A. M. I. L. Janssen, *J. Clin. Endocrinol. Metab.*, 2008, **93**, 2539.
- 20 57. S. Perrini, M. C. Carreira, A. Conserva, L. Laviola and F. Giorgino, *J. Endocrinol. Invest.*, 2008,
- 21 **31**, 79.
- 22 58. S. Perrini, A. Natalicchio, L. Laviola, A. Cignarelli, M. Melchiorre, F. De Stefano, C.
- 23 Caccioppoli, A. Leonardini, S. Martemucci, G. Belsanti, S. Miccoli, A. Ciampolillo, A. Corrado,
- 24 F. P. Cantatore, R. Giorgino and F. Giorgino, *Endocrinology*, 2008, **149**, 1302.
- 25 59. N. Moller and J. O. L. Jorgensen, *Endocrine Rev.*, 2009, **30**, 152.
- 26 60. S. Perrini, L. Laviola, M. C. Carreira, A. Cignarelli, A. Natalicchio and F. Giorgino, *J.*
- 27 *Endocrinol.*, 2010, **205**, 201.
- 28 61. K. R. Short, N. Moller, M. L. Bigelow, J. Coenen-Schimke and K. S. Nair, *J. Clin. Endocrinol.*
- 29 *Metab.*, 2008, **93**, 597.
- 30 62. M. L. Healy, J. Gibney, D. L. Russell-Jones, C. Pentecost, P. Croos, P. H. Sönksen, A. M.
- 31 Umpleby, *J. Clin. Endocrinol. Metab.*, 2003, **88**, 5221.
- 32 63. T. K. Hansen, C. H. Gravholt, H. Ørskov, M. H. Rasmussen, J. S. Christiansen, J. O. Jørgensen,

- 1 *J. Clin. Endocrinol. Metab.*, 2002, **87**, 4691.
- 2 64. M. B. Krag, L. C. Gormsen, Z. Guo, J. S. Christiansen, M. D. Jensen, S. Nielsen and J. O.  
3 Jørgensen JO, *Am. J. Physiol. Endocrinol. Metab.*, 2007, **292**, E920.
- 4 65. J. Gibney, T. Wolthers, G. Johannsson, A. M. Umpleby and K. K. Ho, *Am. J. Physiol.*  
5 *Endocrinol. Metab.*, 2005, **289**, E266.
- 6 66. L. J. Woodhouse, A. Mukherjee, S. M. Shalet and S. Ezzat, *Endocrine Rev.*, 2006, **27**, 287.
- 7 67. N. Mauras, K. O. O'Brien, S. Welch, A. Rini, K. Helgeson, N. E. Vieira and A. L. Yergey, *J. Clin.*  
8 *Endocrinol. Metab.*, 2000, **85**, 1686.
- 9 68. J. Shi, R. V. Sekhar, A. Balasubramanyam, K. Ellis, P. J. Reeds, F. Jahoor and M. D. Sharma, *J.*  
10 *Clin. Endocrinol. Metab.*, 2003, **88**, 5827.
- 11 69. P. Lucidi, S. Laureti, S. Santoni, M. Lauteri, N. Busciantella-Ricci, G. Angeletti, F. Santeusano  
12 and P. De Feo, *Clin. Endocrinol.* 2000, **52**, 173.
- 13 70. D. Le Roith, C. Bondy, S. Yakar, J. L. Liu and A. Butler, *Endocrine Rev.*, 2001, **22**, 53.
- 14 71. G. R. Adams, *Clin Orthop Relat Res.*, 2002, **403S**, S188.
- 15 72. K. C. Copeland and K. S. Nair, *J. Clin. Endocrinol. Metab.*, 1994, **78**, 1040.
- 16 73. F. F. Horber and M. W. Haymond, *J. Clin. Invest.*, 1990, **86**, 265.
- 17 74. M. G. Burt, J. Gibney, D. M. Hoffman, A. M. Umpleby and K. K. Ho, *Growth Horm. IGF Res.*,  
18 2008, **18**, 55.
- 19 75. M. Li, C. Li and W. S. Parkhouse, *Mech. Ageing Dev.*, 2003, **124**, 771.
- 20 76. T. J. Marcell, S. M. Harman, R. J. Urban, D. D. Metz, B. D. Rodgers and M. R. Blackman, *Am.*  
21 *J. Physiol. Endocrinol. Metab.*, 2001, **281**, E1159.
- 22 77. C. Sell, A. Ptaznik, C. D. Chang, J. Swantek, V. J. Cristofalo and R. Baserga, *Biochem. Biophys.*  
23 *Res. Commun.*, 1993, **194**, 259.
- 24 78. V. Chikani and K. K. Y. Ho, *J. Mol. Endocrinol.*, 2014, **52**, R107.
- 25 79. N. Guha, A. Dashwood, N. J. Thomas, A. J. Skingle, A. J., et al., *Curr. Drug Abuse Rev.*, 2009,  
26 **2**, 263.
- 27 80. R. Palmeiro, R. Anand, I. K. Dardi, N. Balasubramaniyam, *Cardiol. Rev.*, 2012, **20**, 197.
- 28 81. Hallengren, P. Almgren, G. Engström, B. Hedblad, M. Persson, J. Suhr, A. Bergmann and O.  
29 Melander, *J. Am. Coll. Cardiol.*, 2014, **64**, 1452.
- 30 82. R. K. Nam, J. Trachtenberg, M. A. Jewett, A. Toi, et al., *Cancer Epidemiol. Biomarkers Prev.*,  
31 2005, **14**, 1270.

- 1 83. K. M. English, R. D. Jones, T. H. Jones, A. H. Morice and K. S. Channer, *J. Endocrinol Invest.*  
2 2002, **25**, 455.
- 3 84. D. M. Kelly and T. H. Jones, *J. Endocrinol*, 2013, **217**, R47.
- 4 85. M. Perusquia, R. Hernandez, M. A. Morales, M. G. Campos and C. M. Villalon, *Gen.*  
5 *Pharmacol.*, 1996, **27**, 181.
- 6 86. H. Honda, T. Unemoto, H. Kogo, *Hypertension*, 1999, **34**, 1232.
- 7 87. A. Q. Ding and J. N. Stallone, *J. Appl. Physiol.*, 2001, **91**, 2742.
- 8 88. P. Tep-areenan, D. A. Kendall and M. D. Randall, *Br. J. Pharmacol.*, 2002, **135**, 735.
- 9 89. R. D. Jones, K. M. English, P. J. Pugh, A. H. Morice, T. H. Jones and K. S. Channer, *J.*  
10 *Cardiovasc. Pharmacol.*, 2002, **39**, 814.
- 11 90. R. D. Jones, P. J. Pugh, T. H. Jones and K. S. Channer, *Br. J. Pharmacol.*, 2003, **138**, 733.
- 12 91. G. Michels, F. Er, M. Eicks, S. Herzig and U. C. Hoppe. *Endocrinol.*, 2006, **147**, 5160.
- 13 92. F. Er, G. Michels, M. C. Brandt, I. Khan, H. Haase, M. Eicks, M. Lindner and U. C. Hoppe, *Cell*  
14 *calcium*, 2007, **41**, 467.
- 15 93. J. L. Scragg, R. D. Jones, K. S. Channer, H. Jones and C. Peers, *Biochem. Biophys. Res.*  
16 *Commun.*, 2004, **318**, 503.
- 17 94. J. L. Scragg, M. I. Dallas and C. Peers, *Cell calcium* 2007, **42**, 11.
- 18 95. T. Stewart, K. T. D. S. Yapa and G. R. Monteith, *BBA*, 2014, doi: 10.1016/j.bbamem.  
19 2014.08.016.
- 20 96. W. Li, S. L. Zhang, N. Wang, B. B. Zhang and M. Li, *Cancer Invest.*, 2011, **29**, 339.
- 21 97. F. Er, G. Michels, N. Gassanov, F. Rivero and U. C. Hoppe, *Circul.*, 2004, **110**, 3100.
- 22 98. M. Seyrek, O. Yildiz, H. B. Ulusoy and V. Yildirim, *J. Pharmacol. Sci.*, 2007, **103**, 309.
- 23 99. P. Tep-areenan, D. A. Kendall and M. D. Randal, *Eur. J. Pharmacol.*, 2003, **465**, 125.
- 24 100. L. J. Pearson, C. Rait, M. G. Nicholls, T. G. Yandle and J. J. Evans, *J. Endocrinol.* 2006,  
25 **191**, 171
- 26 101. L. J. Pearson, T. G. Yandle, M. G. Nicholls and J. J. Evans, *Peptides*, 2008, **29**, 1057.
- 27 102. H. K. Wong, T. T. Cheung B. M. Cheung, *J. R. Soc. Med. Cardiovasc. Dis.*, 2012, **1**, doi:  
28 10.1258/cvd.2012.012003.
- 29 103. J. Kato, T. Tsuruda, T. Kita, K. Kitamura, T. Eto, *Arterioscler. Thromb. Vasc. Biol.*,  
30 2005, **25**, 2480.
- 31 104. B. L. Stauffer, C. M. Westby, C. A. DeSouza, *Curr. Opin. Cardiol.* 2008, **23**, 350.
- 32 105. T. H. Jones and F. Saad, *Atherosclerosis*, 2009, **207**, 318.

- 1 106. Tyagi, M. Rajalakshmi, D. A. Jeyaraj, R. S. Sharma and J. S. Bajaj, *Int. J. Androl.*, 1999,  
2 **22**, 347.
- 3 107. B. Singh, S. Hsia, P. Alaupovic, I. Sinha-Hikim, L. Woodhouse, T. A. Buchanan, R.  
4 Shen, R. Bross, N. Berman and S. Bhasin, *J. Clin. Endocrinol. Metab.*, 2002, **87**, 136.
- 5 108. K. L. Herbst, J. K. Amory, J. D. Brunzell, H. A. Chansky and W. J. Bremner, *Am. J.*  
6 *Physiol. Endocrinol. Metab.* 2003, **284**, E1112.
- 7 109. M. Traish, R. Abdou, K. E. Kypreos, *Vascul. Pharmacol.*, 2009, **51**, 303.
- 8 110. M. Traish and K. E. Kypreos, *Atherosclerosis*, 2011, **214**, 244.
- 9 111. M. R. Smith, J. S. Finkelstein, F. J. McGovern, A. L. Zietman, M. A. Fallon, D. A.  
10 Schoenfeld, P. W. Kantoff, *J. Clin. Endocrinol. Metab.*, 2002, **87**, 599.
- 11 112. M. R. Smith, H. Lee, D. M. Nathan, *J. Clin. Endocrinol. Metab.* 2006, **91**, 1305.
- 12 113. F. Saad, L. J. Gooren, A. Haider and A. Yassin, *J. Androl.*, 2008, **29**, 102.
- 13 114. C. Langer, B. Gansz, C. Goepfert, T. Engel, Y. Uehara, G. von Dehn, H. Jansen, G.  
14 Assmann and A. von Eckardstein, *Biochem. Biophys. Res. Commun.*, 2002, **296**, 1051.
- 15 115. G. Valacchi, C. Sticozzi, Y. Lim and A. Pecorelli, *Ann. N. Y. Acad. Sci.*, 2011, **1229**, E1.
- 16 116. M. P. Corcoran, M. Meydani, A. H. Lichtenstein, E. J. Schaefer, A. Dillard, S. Lamon-  
17 Fava, *J. Endocrinol.*, 2010, **206**, 217.
- 18 117. R. Wan, C. Zhu, R. Guo, L. Jin, Y. Liu, L. Li, H. Zhang and L. Shengnan, *J. Endocrinol.*,  
19 2013, **218**, 321.
- 20 118. M. Kohno, Y. Kawahito, Y. Tsubouchi, A. Hashiramoto, R. Yamada, K. I. Inoue, Y.  
21 Kusaka, T. Kubo, I. J. Elenkov, G. P. Chrousos GP, M. Kondo and H. Sano, *J. Clin. Endocrinol.*  
22 *Metab.*, 2001, **86**, 4344.
- 23 119. G. Annibalini, D. Agostini, C. Calcabrini, C. Martinelli, E. Colombo, M. Guescini, P.  
24 Tibollo, V. Stocchi and P. Sestili, *J Endocrinol Invest.*, 2014, **37**, 861.
- 25 120. H. Hatakeyama, M. Nishizawa, A. Nakagawa, S. Nakano, T. Kigoshi and K. Uchida,  
26 *FEBS Lett.*, 2002, **530**, 129.
- 27 121. S. Ling, A. Dai, M. R. Williams, K. Myles, R. J. Dilley, P. A. Komesaroff and K. Sudhir,  
28 *Endocrinology*, 2002, **143**, 1119.
- 29 122. S. D'Ascenzo, D. Millimaggi, C. Di Massimo, G. Sacconi-Jotti, F. Botrè, G. Carta, M. G.  
30 Tozzi-Ciancarelli, A. Pavan and V. Dolo, *Toxicol. Lett.*, 2007, **169**, 129.
- 31 123. Y. Powazniak, A. C. Kempfer, M. de la Paz Dominguez, C. Farias, L. Keller, J. C.  
32 Calderazzo and M. A. Lazzari, *Mol. Med. Rep.*, 2009, **2**, 441.

- 1 124. P. P. Kayampilly, X. Li, H. Peegel and K. M. Menon, *Endocrinology*, 2002, **143**, 2930.
- 2 125. P. P. Kayampilly and K. M. Menon, *Endocrinology*, 2012, **153**, 2831.
- 3 126. D. Verzola, M. T. Gandolfo, F. Salvatore, B. Villaggio, F. Gianiorio, P. Traverso, G.
- 4 Deferrari and G. Garibotto, *Kidney Int.*, 2004, **65**, 1252.
- 5 127. M. Aragno, S. Parola, E. Brignardello, A. Mauro, E. Tamagno, R. Manti, O. Danni and
- 6 G. Boccuzzi, *Diabetes*, 2000, **49**, 1924.
- 7 128. N. A. Compagnone and S. H. Mellon, *Proc.Natl.Acad.Sci.U. S. A.*, 1998, **95**, 4678.
- 8 129. L. M. Freeman, N. V. Watson and S. M. Breedlove, *Horm. Behav.*, 1996, **30**, 424.
- 9 130. M. Estrada, P. Uhlen and B. E. Ehrlich, *J. Cell Sci.*, 2005, **119**, 733.
- 10 131. G. Hajnoczky, E. Davies and M. Madesh, *Biochem. Biophys. Res. Commun.*, 2003,
- 11 **304**, 445.
- 12 132. S. Orrenius, B. Zhivotovsky and P. Nicotera, *Nat. Rev. Mol. Cell. Biol.*, 2003, **4**, 552.
- 13 133. M. Estrada, A. Varshney and B. E. Ehrlich, *J. Biol. Cell.*, 2006, **281**, 25492.
- 14 134. K. Erkkilä, K. Henriksén, V. Hirvonen, S. Rannikko, J. Salo, M. Parvonen and L. Dunkel,
- 15 *J. Clin. Endocrinol. Metab.*, 1997, **82**, 2314.
- 16 135. E. Ahlbom, L. Grandison, E. Bonfoco, B. Zhivotovsky and S. Ceccatelli, *Eur. J.*
- 17 *Neurosci.*, 1999, **11**, 1285.
- 18 136. E. Ahlbom, G. S. Prins and S. Ceccatelli, *Brain Res.*, 2001, **892**, 255.
- 19 137. J. Pike, *Brain Res.*, 2001, **919**, 160.
- 20 138. N. S. Kamel, J. Gammack, O. Cepeda and J. M. Flaherty, *Cleve Clin. J. Med.*, 2006, **73**,
- 21 1049.
- 22 139. J. Pike, J. C. Carroll, E. R. Rosario and A. M. Barron. *Front Neuroendocrinol.*, 2009, **30**,
- 23 239.
- 24 140. Imperlini, A. Mancini, S. Spaziani, D. Martone, A. Alfieri, M. Gemei, L. Del Vecchio, P.
- 25 Buono and S. Orrù, *Proteomics*, 2010, **10**, 3165.
- 26 141. Imperlini, S. Spaziani, A. Mancini, M. Caterino, P. Buono and S. Orrù, *Proteomics*,
- 27 2015, doi: 10.1002/pmic.201400242.
- 28 142. L. Magliano, C. Dzekov, J. Dzekov, R. Bross, J. Phillips, I. Sinha-Hikim, R. Shen, T. W.
- 29 Storer, *Am. J. Physiol. Endocrinol. Metab.*, 2001, **281**, E1172.
- 30 143. Mancini, E. Imperlini, A. Alfieri, S. Spaziani, D. Martone, A. Parisi, S. Orrù and P.
- 31 Buono, *J. Biol. Reg. Homeos. Ag.*, 2013, **27**, 757
- 32 144. M. Vincent and E. L. Feldman, *Growth Horm. IGF Res.*, 2002, **12**, 193.

- 1 145. M. A. Velazquez, D. Hermann, W. A. Kues and H. Niemann, *Reproduction*, 2011, **142**,  
2 41.
- 3 146. M. M. Chi, A. L. Schlein and K. H. Moley, *Endocrinology*, 2000, **141**, 4784.
- 4 147. B. Pinto, A. L. Schlein AL and K. H. Moley, *Human Reproduction*, 2002, **17**, 457.
- 5 148. G. S. Eng, R. A. Sheridan, A. Wyman, M. M. Y. Chi, K. P. Bibee, E. S. Jungheim and K.  
6 H. Moley, *Diabetes*, 2007, **56**, 2228.
- 7 149. Y. Toyoshima, M. Karas, S. Yakar, J. Dupont, L. Helman and D. LeRoith, *J. Biol. Chem.*  
8 2004, **279**, 25898.
- 9 150. S. Spaziani, E. Imperlini, A. Mancini, M. Caterino, P. Buono and S. Orrù, *Proteomics*,  
10 2014, **14**, 1623.
- 11 151. S. Steffens, A. J. Schrader, G. Vetter, H. Eggers, H. Blasig, J. Becker, M. A. Kuczyk and  
12 J. Serth, *Oncol Lett.*, 2012, **3**, 787.
- 13 152. Grismayer, S. Sölch, B. Seubert, T. Kirchner, S. Schäfer, G. Baretton, M. Schmitt, T.  
14 Luther, A. Krüger, M. Kotsch and V. Magdolen, *Mol. Cancer.*, 2012, **11**, 62.
- 15 153. E. Chua and B. L. Tang, *J. Cell. Mol. Med.*, 2015, **19**, 1.
- 16 154. J. Mitchell, A. E. Nelson, M. J. Cowley, W. Kaplan, G. Stone, S. K. Sutton, A. Lau, C. M.  
17 Lee and K. K. Ho, *J Clin Endocrinol Metab.*, 2009, **94**, 4703.
- 18 155. P. J. Jenkins, A. Mukherjee and S. M. Shalet, *Growth Horm IGF Res.*, 2006, **16**, 277.
- 19 156. J. J. Kopchick, L. L. Bellush, K. T. and Coschigano, *Annu Rev Nutr* 1999.
- 20 157. K. J. Snibson, P. S. Bhathal and T. E. Adams, *Liver*, 2001, **21**, 149.
- 21 158. J. G. Miquet, T. Freund, C. S. Martinez, L. González, M. E. Díaz, G. P. Micucci, E.  
22 Zotta, R. K. Boparai, A. Bartke, D. Turyn and A. I. Sotelo, *Cell Cycle*, 2013, **12**, 1042.
- 23 159. J. G. Miquet, L. González, M. N. Matos, C. E. Hansen, A. Louis and A. Bartke, *J*  
24 *Endocrinol.*, 2008, **198**, 317.
- 25 160. Y. Qin and Y. P. Tan, *Cellular and molecular biology letters*, 2010, **15**, 177.
- 26 161. Macedo, M. Moriggi, M. Vasso, S. De Palma, M. Sturnega, G. Friso, C. Gelfi, M.  
27 Giacca and S. Zacchigna, *Hum. Gene Ther.*, 2012, **23**, 146.
- 28 162. Ehrnborg, C. Ohlsson, S. Mohan, B. A. Bengtsson and T. Rosen, *Growth Hormone &*  
29 *IGF Research*, 2007, **17**, 234.
- 30 163. J. K. Nicholson, J. C. Lindon, E. Holmes, *Xenobiotica*, 1999, **29**, 1181.
- 31 164. M. E. Bollard, E. G: Stanley, J. C. Lindon, J. K. Nicholson and E. Holmes, *NMR*  
32 *Biomed.*, 2005, **18**, 143.

- 1 165. O. Beckonert, H. C. Keun, T. M. Ebbels, J. Bundy, E. Holmes, J. C. Lindon and J. K.  
2 Nicholson, *Nat. Protoc.*, 2007, **2**, 2692.
- 3 166. M. E. Dumas, C. Canlet, J. Vercauteren, F. André and A. Paris, *J Proteome Res.*,  
4 2005, **4**, 1493.
- 5 167. F. Kieken, G. Pinel, J. P. Antignac, F. Monteau, A. Christelle Paris, M. A. Popot, Y.  
6 Bonnaire and B. Le Bizec, *Anal. Bioanal. Chem.*, 2009, **394**, 2119.
- 7 168. J. C. Rijk, A. Lommen, M. L. Essers, M. J. Groot, J. M. Van Hende, T. G. Doeswijk and  
8 M. W. Nielen, *Anal Chem.*, 2009, **81**, 6879.
- 9 169. G. Dervilly-Pinel, S. Weigel, A. Lommen, S. Chereau, L. Rambaud, M. Essers, J. P.  
10 Antignac, M. W. Nielen and B. Le Bizec, *Anal. Chim. Acta*, 2011, **700**, 144.
- 11 170. A. Vonaparti, E. Lyris, Y. S. Angelis, I. Panderi, M. Koupparis, A. Tsantili-Kakoulidou,  
12 R. J. Peters, M. W. Nielen, C. Georgakopoulos, *Rapid Commun. Mass Spectrom.*, 2010, **24**,  
13 1595.
- 14 171. F. Courant, A. L. Royer, S. Chéreau, M. L. Morvan, F. Monteau, J. P. Antignac And B  
15 Le Bizec, *Analyst.*, 2012, **137**, 4958.
- 16 172. C. J. Broccardo, K. L. Schauer, W. M. Kohrt, R. S. Schwartz, J. P. Murphy and J. E.  
17 Prenni, *J. Chromatogr. B*, 2013, **934**, 16.
- 18 173. H-W. Liao, G-Y. Chen, M-S. Wu, W-C. Liao, I-L. Tsai and C-H. Kuo, *J. Chromatogr. A*,  
19 2015, **1375**, 62.
- 20 174. F. D. West, W. M. Henderson. P. Yu, J-Y Yang, S. L. Stice and M. A. Smith. *Toxicol.*  
21 *Sci.*, 2012, **129**, 9.
- 22  
23  
24

1 **Legend to Figures**

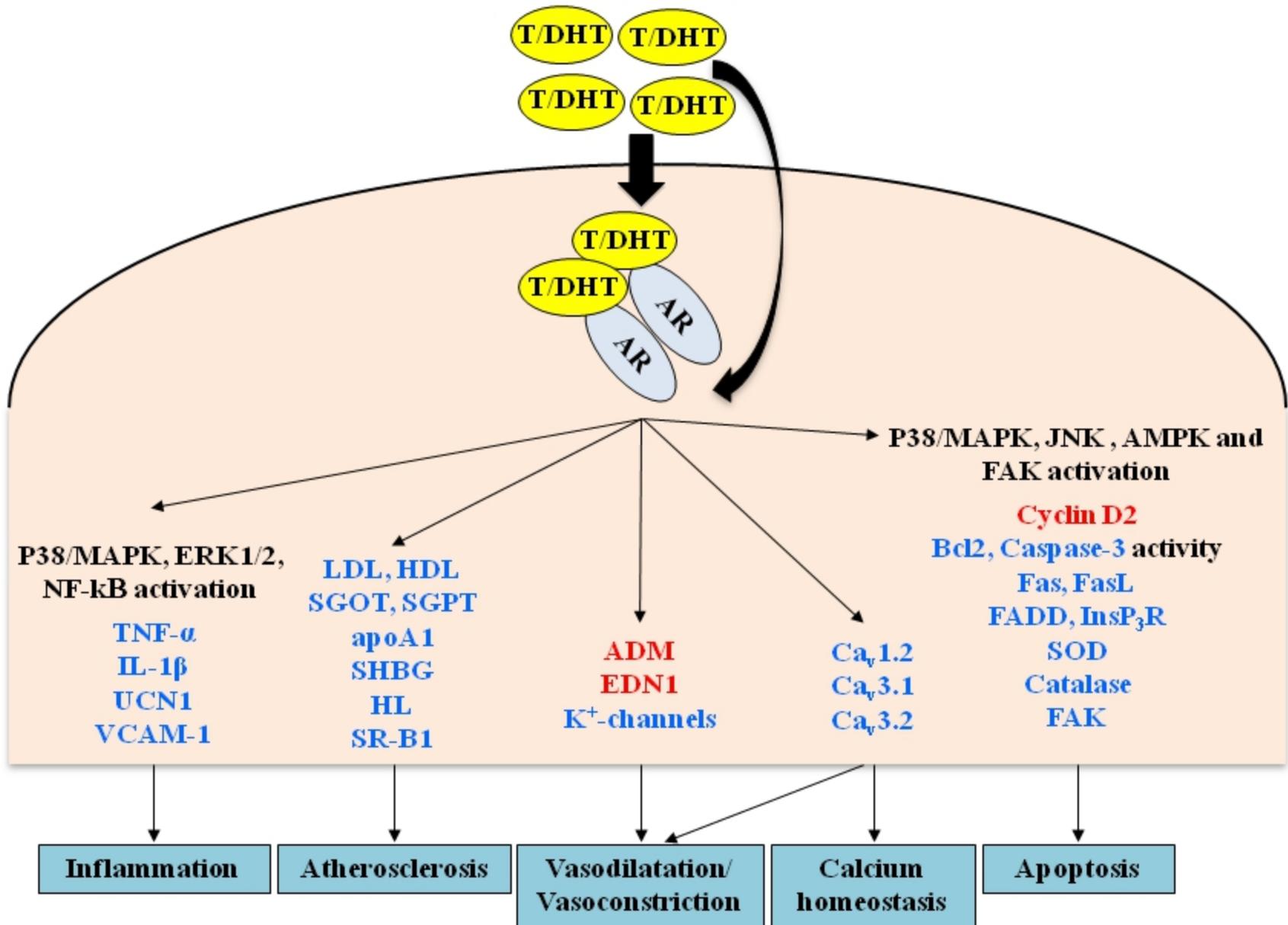
2

3 **Figure 1:** Schematic representation of the effects induced by supraphysiological doses of T/DHT in  
4 several *in vitro* and *in vivo* systems. The main molecular targets (genes in red, proteins in blue) are  
5 shown; their main downstream effects are framed.

6

7 **Figure 2:** Schematic representation of the effects induced by supraphysiological doses of GH/IGF-1  
8 in several *in vitro* and *in vivo* systems. The main molecular targets (genes in red, proteins in blue)  
9 are shown; their main downstream effects are framed.

10



Molecular BioSystems Accepted Manuscript

Figure 1

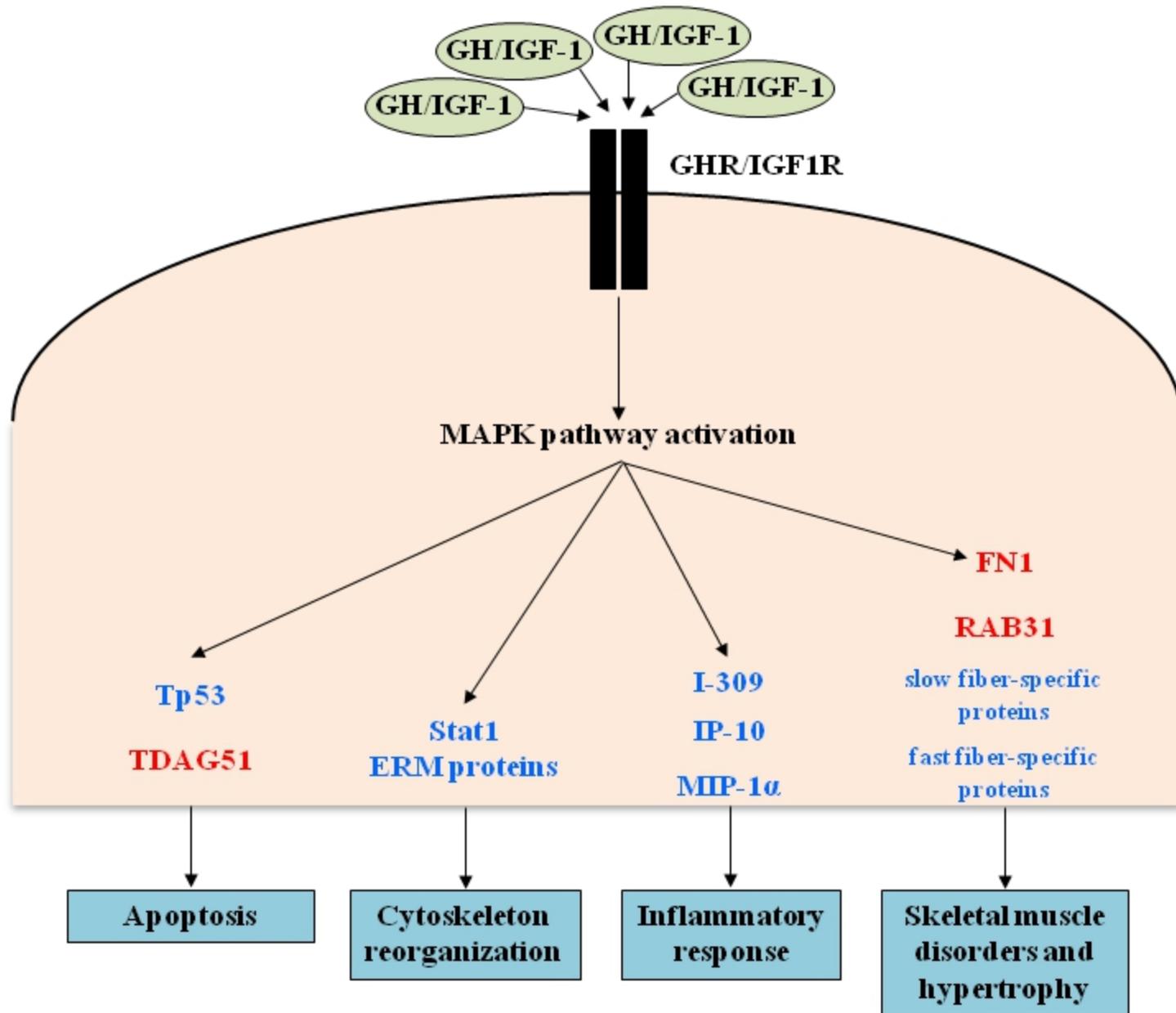


Figure 2