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Metallic nanostructure-based aptasensors for robust detection of proteins

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There is a significant need for fast, cost-effective, and highly sensitive protein target detection, particularly in the fields of food, environmental monitoring, and healthcare. The integration of high-affinity aptamers with metal-based nanomaterials has played a crucial role in advancing the development of innovative aptasensors tailored for the precise detection of specific proteins. Aptamers offer several advantages over commonly used molecular recognition methods, such as antibodies. Recently, a variety of metal-based aptasensors have been established. These metallic nanomaterials encompass noble metal nanoparticles, metal oxides, metal-carbon nanotubes, carbon quantum dots, graphene-conjugated metallic nanostructures, as well as their nanocomposites, metal-organic frameworks (MOFs), and MXenes. In general, these materials provide enhanced sensitivity through signal amplification and transduction mechanisms. This review primarily focuses on the advancement of aptasensors based on metallic materials for the highly sensitive detection of protein targets, including enzymes and growth

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factors. Additionally, it sheds light on the challenges encountered in this field and outlines future prospects. We firmly believe that this review will offer a comprehensive overview and fresh insights into metallic nanomaterials-based aptasensors and their capabilities, paving the way for the development of innovative point-of-care (POC) diagnostic devices.

Introduction

The development of appropriate methods to detect specific proteins in biological samples with great sensitivity and precision is very important since their quantification can be complex, and time-consuming, and the information is crucial for biosensing applications. Biosensors are greatly sensitive and

accurate diagnostic tools and they are relatively cheap to produce, highly portable, and can be mass-produced.¹ For many years, biosensors have been developed and commercialized for various applications including measuring blood glucose, thrombin, lactate, *etc.* However, the sensitivity, accuracy, stability, and design of biosensors need to be improved considering the broad commercial potential. Towards this, metallic nanoparticle-based aptamer sensors (metallic aptasensors) could offer great potential.

Aptamers are short single-strand DNA or RNA molecules with a great affinity to their targets because of their capability to form three-dimensional structures in solution.² Aptamers, often termed chemical antibodies, can be used as an alternative to antibodies, and possess some obvious advantages over antibodies including high stability, freedom to incorporate multiple chemical modifications without changing their binding properties, long shelf life, and easy laboratory-based development and production in large amounts.² In addition, aptamers can discriminate between chiral molecules and can identify a certain epitope of a target molecule. Aptamers are formed by a reiterative selection process *in vitro* mentioned as the Systematic Evolution of Ligands by EXponential enrichment (SELEX), and aptamers can be selected for almost any desired



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continued his postdoctoral career under the supervision of Prof. Jesper Wengel at the Nucleic Acid Center, University of Southern Denmark in the field of nucleic acid chemical biology. Later in 2009, he was appointed as a Research Associate Professor within the Nucleic Acid Center. He then returned to the University of Queensland in mid-2010 and established functional nucleic acid therapeutics research group. His current research is focused on developing novel synthetic gene targeting nucleic acid therapeutics and nucleic acid diagnostics against a range of rare and acquired diseases. In addition, he is also one of the co-founders and the director of SynGenis Pty Ltd and ProGenis Pty Ltd.



target ranging from small molecules to complex proteins (Fig. 1).³ Aptamers are generally used for several applications, such as diagnostics development, drug development, and drug delivery.^{4–12} The biosensing potential of aptamers (aptasensors) can be strengthened by incorporating them with various metal-based nanomaterials.^{13,14} The use of metallic nanomaterials can increase the optical, thermal, and electrochemical properties of the transducers, resulting in enhanced aptamer–ligand interactions. In fact, the coupling of nanomaterials with aptamers is a quickly emerging field where various aptamer–material conjugates *via* outstanding sensing efficacies have been developed.¹⁵ Metal-based nanomaterials, including carbon-based nanomaterials, metal nanoparticles (NPs), metal–organic frameworks (MOFs), and MXenes, have been developed to fabricate biosensing systems according to their exclusive properties.^{16,17} Besides the above-mentioned properties, metallic nanomaterials might be used as aptamer carriers in biosensing systems.

Since the aptamer is immobilized onto the transducer surface, using an effective transducer is paramount for the performance of aptasensors. Therefore, transducers have a crucial role in the aptasensors development to record the signal and exchange it for a detectable output in different plans.¹⁸ In fact, based on the type of transducer, sensors can be classified into electrochemical, optical, and thermal biosensors.¹⁹ Optical aptasensors comprise aptamers labeled with NPs, luminophores, and fluorophores (Fig. 2A), or label-free

aptamers as observed in surface plasmon resonance (SPR) (Fig. 2B) and surface-enhanced Raman spectroscopy (SERS) sensors. Most targets can be optically identified by using fluorescence, chemiluminescence, or SPR.²⁰ In electrochemical aptasensors (ECAs), the surface of the electrode is used as a substrate to immobilize the aptamer, and changes in the electrochemical current are investigated. High sensitivity and low synthesis cost are the advantages of the electrochemical signaling.²¹ ECAs include labeled and label-free systems.^{22,23} Electrochemical signal changes can be analyzed qualitatively and quantitatively to investigate target molecules. Labels can be electroactive materials, enzymes, and some NPs such as quantum dots (QDs).²³ In label-free systems, the detection of analytes is performed by the recognition of the analytes by the specific aptamers *via* electrostatic interaction.²² Detection with the high specificity of proteins, such as enzymes, growth factors, *etc.*, can be done by this type of aptasensors.

Several studies related to the role of metallic nanomaterials-based aptasensors in high-sensitivity detection of protein targets have been published in recent years, which reflects a higher importance of metal NPs than other nanomaterials in this field. Fig. 3 displays the status of research articles published in the area of metal nanomaterials-based aptasensors for protein detection over the last dozen years. According to the figure, there is a gradual increase in the number of studies during this period, indicating an increasing awareness of the importance of metallic nanomaterials for the design of



Fig. 1 Schematic illustration of the SELEX process. This figure was drawn by using <https://BioRender.com>.





Fig. 2 Labeled or label-free aptamers are used in optical aptasensors. (A) Emission spectra of common fluorophores used in different optical (bio)sensor systems; and (B) a schematic illustration of the SPR method. This figure was drawn in <https://BioRender.com>.

aptasensors. Although different latest review articles have been published on the role of metal-based nanomaterials in aptasensors, none has provided a thorough investigation of these aptasensors in detecting proteins.^{24,25} In our estimation, there is a certain need for a comprehensive review of this topic. Thus, we herein review the relevant studies and finally present the challenges and prospects for the future development of metallic nanomaterials-based aptasensors.

The commercialization of aptamer-based biosensors (*i.e.*, aptasensors) has the potential to generate significant economic benefits for various industries. The global biosensors market was valued at US\$21.2 billion in 2020 and is anticipated to develop at a compound annual growth rate (CAGR) of 8.8% from 2021 to 2028, driven by the demand for accurate and reliable diagnostic tools. The healthcare sector is the largest consumer of biosensors, with a market share of over 40%. The market for aptasensors is expected to grow rapidly, mainly

owing to their great specificity and sensitivity, and low production cost. According to a study conducted by the National Institutes of Health (NIH), the cost of developing a biosensor can vary between US\$100 000 and US\$1 million, depending on the level of innovation and the required regulatory approvals. Although costly, once commercialized, the potential revenue generated by a biosensor on the market can far exceed the initial investment. In fact, several government agencies and private organizations have been providing funding to maintain the development and commercialization of sensors. For instance, the National Science Foundation (NSF) offers grants of up to US\$1.5 million for biosensor research and development (R&D) projects. The NIH also offers funding opportunities for biosensor research, including Small Business Innovation Research (SBIR) and Small Business Technology Transfer (STTR) grants. In addition, private investors and venture capitalists are interested in funding biosensor startups as well, with



Table 1 List of different metal-based nanomaterials used in aptasensors, their synthesis, and their properties

Metal-based materials	Synthesis methods	Desired physical properties
Noble metal/metal oxide	<ul style="list-style-type: none"> • Hydrothermal • Electrochemical • <i>In situ</i> reduction 	<ul style="list-style-type: none"> • High optical and electrical conductivity • Easy synthesis method • Non-toxicity
Carbon materials	<ul style="list-style-type: none"> • Chemical vapor deposition (CVD) • Laser ablation 	<ul style="list-style-type: none"> • Large surface area • Excellent electrical and mechanical properties • High stretchability
Metal organic framework (MOFs)	<ul style="list-style-type: none"> • Surfactant-assisted synthesis • Exfoliation • Ultrasonication • Liquid exfoliation 	<ul style="list-style-type: none"> • Porosity • Stability • Synthetic tunability
MXenes	<ul style="list-style-type: none"> • Hydrofluoric acid etching • Fluoride-free etching • Molten salt etching 	<ul style="list-style-type: none"> • High electrical conductivity • High biocompatibility • Excellent electrical properties

“A” group metal layer from the $M_{n+1}AX_n$ (MAX) phase structure, wherever M refers to transition metals. Although bottom-up approaches have been applied to synthesize MXenes, they are often formed by top-down approaches, with their structure and composition originating from their nitride precursors.⁵⁰ These top-down approaches are based on hydrofluoric (HF) etching of the MAX phase.⁴⁹ In general, the properties of MXenes can be engineered during the synthesis process. They are predisposed by various parameters, such as the precursor MAX phase, temperature, and the etchant. However, regulating these parameters is challenging to acquire suitably designed MXenes with wanted properties.⁵¹ Table 1 shows different metal-based nanomaterials used in aptasensors, their synthesis, and their physical properties.

Metal-based aptasensors for protein detection

Noble metal/metal oxide-based aptasensors

Metal-based nanomaterials with different sizes have interesting physicochemical and optoelectronic properties.^{52,53} They are being progressively used as transducing materials in biosensors for the analysis of biomolecules. Noble metal NPs are beneficial over other nanomaterials due to the simplicity of chemical modifications, great surface area, exclusive optical, redox, and luminescence properties, and their capability to respond optically and electrochemically to external stimuli.⁵⁴ With the enhancement of the surface area in metal nanomaterials, the optical, electronic, chemical, and thermal properties of the materials are altered. The choice of synthesis approach and compounds and conditions used for the synthesis are key to the formation of structures with the appropriate sizes and shapes.

Noble metal nanomaterials with exclusive physicochemical properties can increase the sensitivity and selectivity of biosensors. Utilizing aptamers as the recognition elements, these sensing platforms hold great promise in achieving economic, quick, and highly sensitive detection of proteins.¹³ The key roles of metal nanomaterials in biosensors are immobilizing receptors, catalyzing bioreactions, electron transfer, and amplifying mass change.⁵⁵

Au nanostructures have been extensively used in studies including the construction of biosensors, which can be attributed to their great compatibility, simplicity in their synthesis, and stability in the synthesis procedures.⁵⁶ To greatly increase the sensitivity of aptamer-based biosensors in protein detection, an efficient signal amplification method is needed. Oligonucleotide-modified AuNPs have been broadly used as signal-amplifying tags in new sensing systems since they can connect biological molecules and have excellent electrochemical performance. Each AuNP can be loaded within DNA strands.⁵⁷ In contrast, aptamers can provide a high level of control over the interaction between the aptamer and protein in AuNPs-based aptasensors. One of the important questions is, what effect will aptamer density have on AuNPs. In a study, the kinetics of interaction among the IFN- γ and AuNPs-aptamer is investigated through fluorescence spectroscopy. The connection of IFN- γ with AuNPs-aptamer alters the construction of aptamer and enhances the fluorescence signal. Furthermore, the number of aptamers on the surface of AuNPs affects the binding reaction between aptamer and protein.⁵⁸ The AuNPs-aptamer system with the greatest surface coverage is advantageous in biosensing systems due to its high sensitivity. These outcomes expand our knowledge of the aptamer-protein interaction on the AuNPs surface, which is critical to enhancing the application of AuNPs-aptamer-based biosensors.

Thrombin is one of the important proteins in blood coagulation, as well as tissue repair. Thrombin can control platelet aggregation and another crucial responses in the vascular system.⁵⁹ A photoelectrochemical (PEC) aptasensor developed to detect different concentrations of thrombin. The transducer is a glassy carbon electrode (GCE), and a layer of AuNPs was deposited on its surface (Fig. 4A and B). This electrode was functionalized with perylene tetracarboxylic acid (PTCA), and a new layer of AuNPs was deposited on the surface of the transducer. With the immobilization of amine-modified thrombin-specific aptamer on the surface of AuNPs, a structure comprising GCE/AuNPs/PTCA/AuNPs/aptamer was formed. Subsequently, a sandwich structure was formed based on the stabilization of the analyte from both sides through the aptamer on the transducer surface. Glutaraldehyde was added as a cross-linking agent, while $C_3N_4@C_{60}$ core-shell was added as



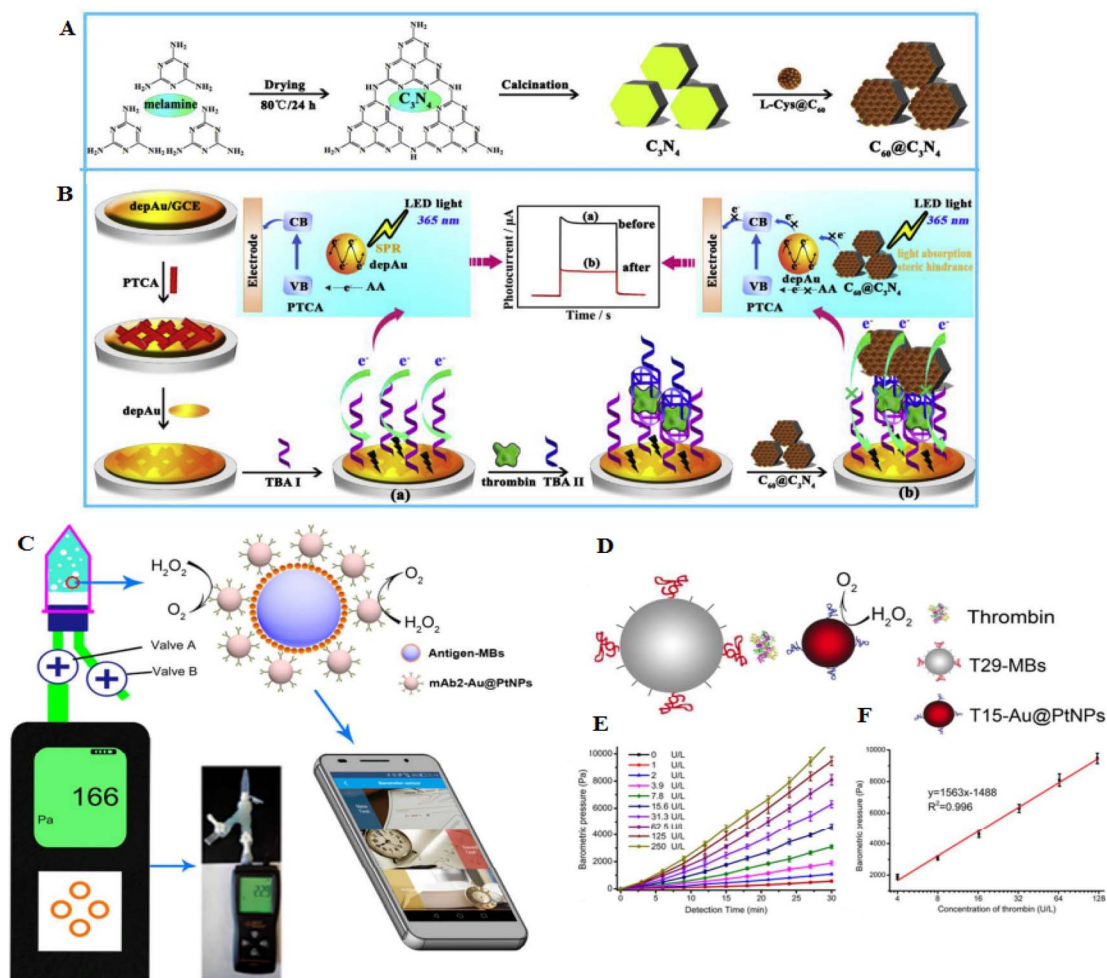


Fig. 4 An electrochemical sensor for recognition of thrombin by using C₆₀@C₃N₄ nanocomposites-based electrodes. (A) The formation of C₆₀@C₃N₄ composites. (B) C₆₀@C₃N₄ nanocomposites stimulate signal-off PEC aptasensor. Reproduced from ref. 60 with permission from Elsevier, Copyright 2019. (C) The barometer-based sensor is based on the measurement of oxygen production using a precise barometer. (D) Diagnosis of thrombin using the barometer-based aptasensor. (E) Concentration-dependent barometric pressures to thrombin detection. (F) The calibration curve of the thrombin, reproduced from ref. 61 with permission from American Chemical Society, Copyright 2017.

a quencher, and the PEC signal yielded by the aptasensor was measured under different analyte concentrations. Here, a competition between C₃N₄ and PTCA for light absorption in the existence of the electron-donating ascorbic acid delivered various photocurrents when the sandwich conformation existed or the thrombin was absent. This quenched signal of the PEC aptasensor could sensitively detect the mentioned biomarker with an LOD of 1.5 fM.⁶⁰ Therefore, the important point is to choose a signal amplification strategy to obtain a lower LOD in electrochemical systems. Another study showed an electrochemical sandwich aptasensor containing two thrombin aptamers, AuNPs, and [Ru(NH₃)₆]³⁺ to signal alteration. In the presence of thrombin, aptamers connect to the thrombin, and their complexes lead to the fabrication of a sandwich structure, causing more [Ru(NH₃)₆]³⁺ adsorbed on the phosphate backbone of the aptamers, and enhancing the sensitivity for pulse voltammetric measurements. The LOD of this detection was 0.14 fM. This aptasensor showed good selectivity and was used to detect thrombin.⁵⁷

Au@PtNPs were developed as probes since they showed catalytic stability for the decomposition of H₂O₂ and were highly sensitive in the diagnosis of carcinoembryonic antigen (CEA) and ractopamine (RAC) in urine samples (Fig. 4C). To detect thrombin, aptamer 1 (T29) was conjugated with molecular beacons (MBs), and aptamer 2 (T15) was labeled with Au@PtNPs. With the binding of T15-Au@PtNPs to the T29-MBs, a high concentration of thrombin was detected (LOD of 0.025 ng mL⁻¹) (Fig. 4D–F). A smartphone was applied to detect and transmit the results. Moreover, the aptasensor showed great specificity to CEA, RAC, and thrombin in serum, urine, and serum samples, respectively.⁶¹

Tumor necrosis factor α (TNF α) is one of the most important proteins involved in the signaling pathways, which have an important role in inflammation as well as inhibition of tumor progression. An electrochemical aptasensor was established for the recognition of TNF- α . At first, the electrode modification print with graphite plate by cobalt hexacyanoferrate (CoHCF) was done. Its modification was also done with colloidal gold



NPs. Then, a thiolated aptamer was immobilized on the electrode surface modified with CoHCF/AuNPs, and the aptamer was connected to the AuNPs surface by Au-S bonds. TNF- α was captured through aptamer on one side, and an antibody absorbed TNF- α on the other side. This platform is capable of quantifying TNF- α . Also, the LOD of the said aptasensor was found to be about 0.52 $\mu\text{g mL}^{-1}$.⁶²

Silver nanomaterials have high electrical conductivity, which can immobilize molecules through an Ag-S bond. Therefore, silver nanowires (AgNWs) and AgNPs can be recognized as good substrate materials for biosensor fabrication.⁶³ Using AgNPs increases the film's conductivity and raises the surface area of the GCE. Instead, AgNPs were fixed in a polymer matrix and showed significant antimicrobial activity.⁶⁴ AgNWs can provide

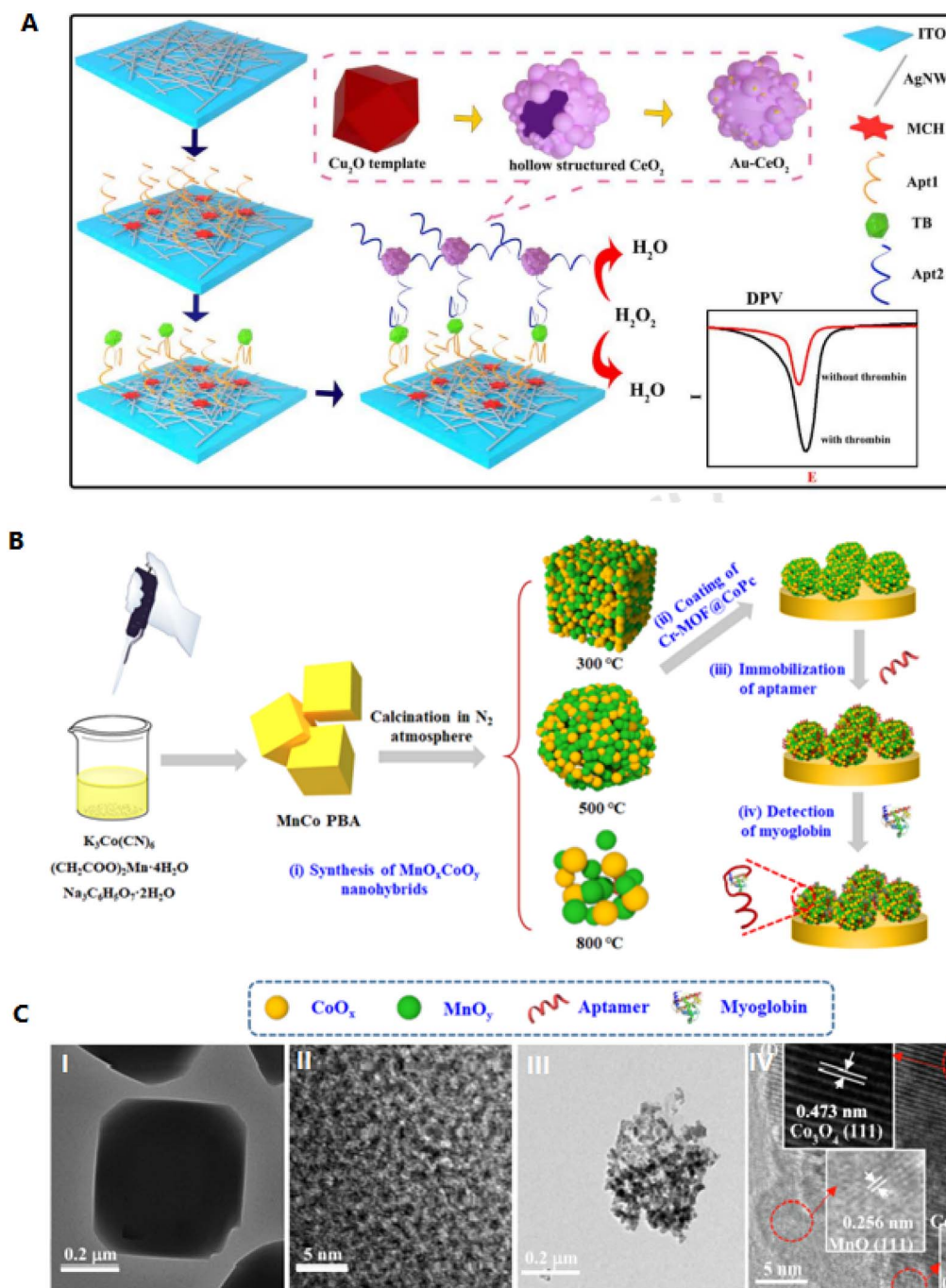


Fig. 5 Designing a new type of aptasensor for highly efficient detection of myoglobin. (A) Schematic illustration of decorated AgNWs on the cleaned indium tin oxide (ITO) surface. These hierarchical networks cause electron transfer and act as an excellent conducting substrate and a free electrochemical sensor. Reproduced from ref. 63 with permission from Elsevier, Copyright 2021. (B) Fabrication of MnO_xCoO_y-based aptasensor. (C) Transmission electron microscopy (TEM) image of the (I, II) MnCo PBA as well as (III, IV) MnO_xCoO_y. Reproduced from ref. 67 with permission from Elsevier, Copyright 2020.



high diffusion ways to electrons, which can intensely increase the electrochemical activity of the biosensors. Fig. 5A shows a sensitive electrochemical aptasensor based on AgNWs electrode and signal amplification of ZnFe_2O_4 for thrombin detection. Here, AgNWs@PCs are applied as the electrode and substrate to bind aptamer. Deposited AgPCs can effectively increase the soldering of AgNWs, which improves the conductivity of the electrode. This type of sensor has a LOD of 0.01 pM, which shows suitable performance in selectivity and reproducibility and is effective in identifying proteins.⁶³

A study showed that polydopamine (PDA) films act as an intermediate to inhibit the interaction of sulfhydryl-polyethylene glycol (SH-PEG) to AgNPs, as it results in the delamination of the silver layer.⁶⁴ PDA-AgNPs can be applied as perfect electrode substrates to electrochemical biosensors. Here, deposition of PDA has occurred on the GCE through self-polymerization, and the thrombin-specific aptamer was grafted onto the PDA-modified GCE. In the presence of the thrombin, the electrode's electrochemical impedance is progressively enhanced. The results showed a LOD of 36 fM in human samples. The selective method detects thrombin in diluted human serum.⁶⁵ Despite the signal from AgNPs, the AgNPs aggregates offer a highly amplified electrochemical signal three times and display high merits in the electrochemical biosensing.⁶⁶ An aptasensor based on amperometric assay using AgNPs on a gold electrode is designed for glycoprotein detection. Aggregation is performed by adding 4-mercaptophenylboronic acid (MPBA) according to the production of Ag-S bonds. A thiolated aptamer was connected to the electrode to absorb the glycoprotein. As a result, after absorption, the prespore-specific antigen (PSA) glycoprotein reacts with MPBA by creating boronated esters. Subsequently, the electrochemical signal is amplified through the synthesis of an AgNPs network that acts as a redox reporter. The LOD of PSA is 0.2 pg mL^{-1} , which is used as a strong aptamer to detect all types of glycoproteins.⁶⁶

One of the metal oxides is manganese oxides (MnO_x), including MnO_2 , Mn_3O_4 , Mn_2O_3 , etc. MnO_x can be used as active electrodes according to their great precise capacitances *via* the structures controlled in growth procedures. The electron transfer of MnO_x can be appropriated for different applications, such as biosensing systems.⁶⁷ MnO_xCoO_y were formed from MnCo-based Prussian blue analog through calcination at various temperatures, and this impedimetric aptasensor showed precise detection of myoglobin (Myo) (Fig. 5B). Under a high temperature of 500 °C, the homogeneously dispersed MnO_x and CoO_y NPs were attained, increasing the electrochemical conductivity. MnO_xCoO_y showed high sensitivity than the MnCo-phenylboronic acid (PBA)-based aptasensor in the detection of Myo, with a LOD of 0.56 fg mL^{-1} . This approach creates a critical system for building an aptasensor that is possible for Myo recognition in a real human serum sample.⁶⁷ Co_3O_4 nanomaterial, as a semiconductor, was applied to detect the cardiac troponin. The diagnosis of cardiac troponin T is performed *via* immobilizing its DNA aptamer. The LOD of the biosensor was $0.1 \mu\text{g mL}^{-1}$, which was enhanced by about 250% compared with pure Co_3O_4 nanorods (NRs). However, the limitation of this aptasensor is its LOD ability, which can be

improved by changing the fabrication procedure. This DNA aptasensor is new and can be established in a combined and high-throughput system.⁶⁸ Table 2 shows the role of noble metal/metal oxide nanomaterials applied as transducers to detect proteins.

An invention presents a portable system for detecting cardiac biomarkers in a patient's bodily fluids using impedimetric techniques. The system is composed of a conductive material interface that includes an epoxy substrate and a vertically aligned platinum wires. A sensor agent is used to the surface of the conductive material interface, which consists of an immobilization agent and one aptamer that binds to the cardiac biomarkers of interest. The aptamer generates an electrochemical impedance signal when it binds to the biomarkers, which is indicative of their presence or absence in the bodily fluid sample. The platinum wires work as the point of contact in order to transducing the electrochemical impedance signals into an interpretable output. The system can detect multiple cardiac biomarkers simultaneously, including C-reactive protein, troponin T, myoglobin, brain natriuretic peptide, and D-dimer. To detect cardiac biomarkers using the system, a fluid sample is obtained from the patient and is then brought into contact with the detection device. The detection device is formed by casting the platinum wires in the epoxy substrate and polishing the surface of the conductive material interface. Then, the electrochemical impedance signal generated by the aptamer binding to the cardiac biomarkers is transduced and displayed using a hand-held device. Alternatively, the detection system can take the form of a test strip, and the method involves correlating visual changes in the test strip with a chart or key to determine the presence or absence of cardiac biomarkers. The surface of the conductive material interface is polished to provide a surface roughness inside the range of about 320 to 2400 grit.⁷⁵

Magnetic-based aptasensors

Magnetic NPs show paramagnetic properties, which are highly capable of separating target molecules from other mixtures through a magnetic field, causing them gorgeous materials to manufacture sensors.^{76,77} One of the applications of this type of material is as an electrode modifier to increase electron conductivity (an inherent feature of metal oxide NPs). Also, their combination with aptamers can separate biomolecules and increase the signal.⁷⁸ Binary transition metal oxides (BTMOs) provide different benefits than single metal oxides. They can offer a high potential, greater conductivity, extra active sites, and good stability, which are critical to increasing biosensor activity. MFe_2O_4 ($\text{M} = \text{Ni}, \text{Co}, \text{Zn}, \text{or Cd}$) are a group of BTMOs. In addition to the magnetic characteristics, spinel ferrites such as ZnFe_2O_4 are critical in proposing high redox reaction electrochemistry. An ZnFe_2O_4 NPs-based aptasensor using laser-scribed graphene (LSG) electrodes as a polyimide substrate was developed. LSG- ZnFe_2O_4 electrodes showed an increased sensitivity and electrocatalytic activity of about 30% higher than the unmodified electrode. ZnFe_2O_4 is conjugated with aptamer by Zn-S and Fe-S bonds, which can produce a greatly stabilized



Table 2 List of different types of noble metal/metal oxide nanomaterials applied as transducers for the detection of protein^a

Nanomaterials	Aptamer	Biosensing principle	Target protein	Properties	LOD	Ref.
AuHCF/AuNPs	TGGTGGATGGCGCCAGTCGGCGACAA	EL	Tumor necrosis factor alpha (TNF- α)	High sensitivity detection even at very low concentrations	5.5 pg mL ⁻¹	69
Fern leaves-like gold nanostructure (FLGN)	TTTTTTTTTTGGGAUUGAGCUCGCGUACCUUUAACGGUAAG GCCUGUCUUCGUUUGACAGCGGCUUUGAGACCCUCACAC UUUGUACCUUGCUGCCAAACUCGACGGCAUGCAAGCUUGG	EL	Amyloid beta (A β)	Administration of Alzheimer's disease at early stages	0.4 pg mL ⁻¹	70
AuNPs	ATACCAGCTTATTCAATTACAGTAGTGAGG GGTCCGTCGTGGGGT/AGTTGGGTCTGGAGATAGTAAGTGCAATCT	LSPR	Retinol-binding protein 4 (RBP4)	Rapid, high sensitivity	90.7 nM	
AgNCs	CAGGCTACGGCCAGTAGAGCATCACCATGATCC TGATTAGTTCCCTCAGCATCAGG(P1) CCCACCTCCCACTGATGCTGAGGGT(P2)	FL	Platelet-derived growth factor B-chain homodimer (PDGF-BB)	High selectivity toward specific protein	0.37 nM	71
ZrCo-MOF	CCACGATACCAGCTTATTCAATTCTGTGG	EL	CEA	High selectivity, good reproducibility, high stability	0.35 fg mL ⁻¹	72
QDs	N/A	EL	p24-HIV protein	Early detection of HIV infection with a wide linear range, high selectivity	51.7 pg mL ⁻¹	73
CdSe/CdS/ZnS QDs	GCCTGTGTGGGGGGGTGCG (A β O) GCGGAGCGTGGCAGG (Tau)	PL	A β O and tau protein	High flexibility, high sensitivity	A β O: 50 pM, Tau: 20 pM	74

^a PL: photoluminescence. EL: electrochemical. FL: fluorescence. LSPR: localized surface plasmon resonance.





electron transfer bonding way. This aptasensor can detect the cardiac troponin I (cTn-I) with a LOD of 0.001 ng mL⁻¹. The fabricated aptasensor has greater to detecting cTn-I and slight cross-reactivity with other biomolecules. Fig. 6A shows the mechanism of the LSG-ZnFe₂O₄ electrochemical aptasensor to detect cTn-I.⁷⁸ Another magnetic composite consists of Fe₃O₄@Au nanomaterials, magnetic field-focused self-assembly, and selected aptamer with great specificity to thrombin. Here, in the existence of protein, aptamer connected with thrombin and showed complementary strands; thus the electrical signal was enhanced (Fig. 6B). Consequently, the electrochemical signal increased by three times through the catalyzed cleavage of surface modification by Pb²⁺-related DNAzyme.⁷⁹

Developing a label-free electrochemical aptasensor based on a modified glassy carbon electrode (MGCE) modified with Fe₃O₄/Fe₂O₃@AuNPs was used to detect VEGF165 highly. Fe₃O₄/Fe₂O₃@AuNPs act as the signal amplifier, which is critical to detecting biological analytes. The aptamers' binding to the nanomaterial was performed by Au-S bonds and is precisely known as VEGF165. Under optimal detection conditions, a virtuous linear association was attained with the aptasensor. The LOD was 0.01 pg mL⁻¹, and this aptasensor showed high stability. The outcomes exhibited that the aptasensor was hopeful to offer an effective, precise, and cost-effective detection assay to the early investigation of VEGF165-related diseases, which showed significant biomedical applications for the study of medicine accuracy.⁸⁰

Due to the establishment of magnetic Fe₃O₄ NPs to reduce the inherent peroxidase-like activity, colorimetric detection has appealed to enhance the consideration of recycling and clearance from the system with progress in the magnetic separation.⁸¹ Au@Fe₃O₄ NPs were synthesized by the solvothermal method. This platform showed great peroxidase-like activity according to the synergistic effect. Besides, the hydroxyl groups (-OH) on the NPs enable their binding with aptamer.⁸¹ A new colorimetric sensor was synthesized through the connection of peroxidase-mimicking DNAzyme with magnetic NPs for high-sensitivity detection of target proteins. This complex was used as the peroxidase-mimicking DNAzyme, wherein hemin was conjugated to G-quadruplex (G4)-DNA to double into a catalytic structure and perform as an enzyme. These aptasensors are

formed from magnetic NPs, cDNA modified with biotin, as well as ssDNA comprising the aptamer and DNAzyme. The trivalent DNAzyme has high catalytic activity, which catalyzed the H₂O₂-related oxidation of 2,2'-azino-bis(3-ethylbenzthiazoline-6-sulfonic acid) (ABTS). Besides, this aptasensor was used to detect mucin 1 (MUC1), which showed LOD of 5.08 and 5.60 nM. This aptasensor with high selectivity and good anti-interference capability has high potential in disease diagnoses.⁸² Table 3 shows the role of noble magnetic nanomaterials applied as transducers to detect proteins.

Carbon-based aptasensors

Carbon nanomaterials appeal to attention according to their exclusive chemical, and electrical properties. They are broadly applied in photovoltaic, electronic, optical, and sensing systems. These exceptional properties considerably rely on the functional structures of carbon nanomaterials but how they interact with another materials, including AuNPs, SiO₂, *etc.*⁸⁸ Besides, virtuous electrical conductivity, thermal properties, and great electrocatalytic activity permit carbon nanomaterials to be promising carriers for aptamers.⁸⁹ Carbon nanomaterials for fabricating ECAs include carbon nanotubes (CNTs), graphene oxide (GO), carbon nanocomposites, *etc.*⁹⁰

CNTs are one-dimensional nanomaterials made of hybridized carbon atoms connecting *via* C-C interaction. Commonly, CNTs include SWCNTs and MWCNTs. CNTs have exclusive thermal, mechanical and electrochemical properties, which have an important role in the electron transfer of various electroactive materials.^{91,92} CNTs have good adjustable properties that can be functionalized with various chemical groups according to specific requirements. Thus excellent characteristics of CNTs would be turned up, such as compatibility and solubility.⁸⁹ SWCNTs can selectively adsorb aptamers. ssDNA can be adsorbed, although dsDNA cannot. A study showed that SWCNTs-based electrochemical sensors are designed based on the use of ssDNA.⁹³ Progress of an electrochemical biosensing system by SWCNTs-screen-printed electrodes (SWCNT-SPEs) modified with a redox-tagged DNA aptamer can be connected to the SARS-CoV-2 spike protein S1. It controls a folding-based apparatus that outcomes in major changes in the amperometric under the connecting of the S1 protein to the aptamer (Fig. 7). The aptasensor can detect S1 protein with a LOD of

Table 3 List of different types of magnetic nanomaterials applied as transducers for the detection of proteins^a

Nanomaterials	Aptamer	Biosensing principle	Target protein	Properties	LOD	Ref.
Fe ₃ O ₄ @SiO ₂ @Ag	N/A	SERS	cTnI	High specificity and sensitivity	10 ng mL ⁻¹	83
Magnetic NPs	GGGATGGGTTTAGTCCGTGGTAGG GCAGGTTGGGGTGA CTTTACTGCC TGACTTTCCTTTTTGGGTTGGG	Optical	Thrombin	Sensitive detection of thrombin in complex samples	0.5 nM	84
Fe ₃ O ₄	N/A	ECL	HBsAg	Rapid magnetic separation, highly sensitive detection	0.05 ng mL ⁻¹	85
Magnetic NPs	N/A	FL	CD63 exosome	High specificity	100 particles per mL	86
AuMNPs	GGTGGTGTGGTTGG	EL	Thrombin	High sensitivity detection	5.5 fM	87

^a SERS: surface-enhanced Raman spectroscopy. ECL: electrochemiluminescence. FL: fluorescence. EL: electrochemical.



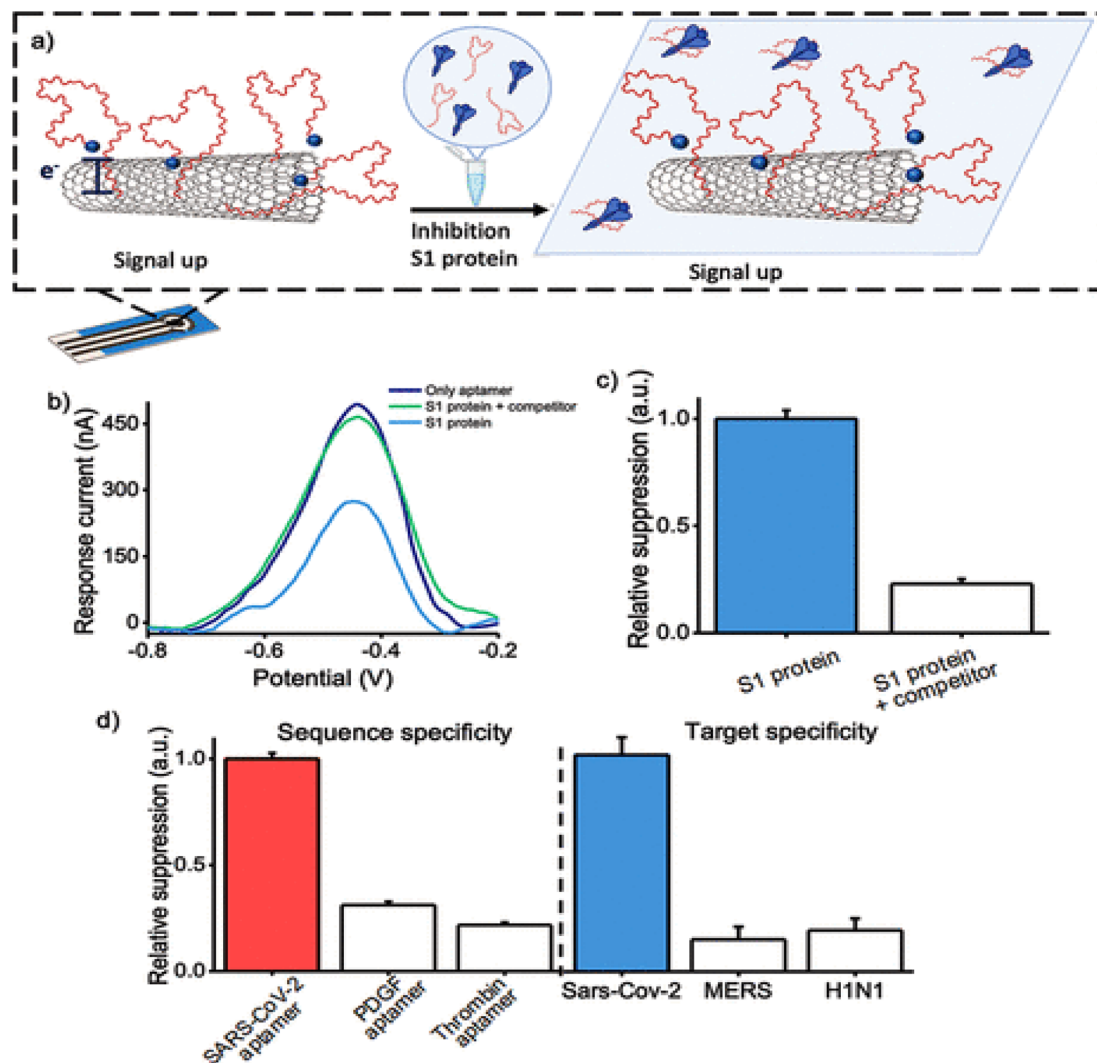


Fig. 7 Inhibition of S1 protein with its mechanism on the surface of the MWCNTs, as a hopeful approach for the recognition of coronavirus via a novel aptasensor. (a) The aptasensor activity when treating the S1 protein with the COVID-19 aptamer. (b) DPV voltammograms without any target protein, in the existence of S1 protein, threatened with increased participant aptamer. (c) Signal suppression was gained when measuring the amperometric in the existence of the S1 protein. (d) The specificity of the biosensor was assessed by using various aptamer sequences immobilized onto the electrode surface in the existence of the S1 protein, reproduced from ref. 94 with permission from American Chemical Society, Copyright 2022.

7 nM. This protein is specifically detectable in a buffer solution, and a synthetic viral transport medium is broadly applied for collecting nasopharyngeal swabs. This electrochemical aptasensor based on SWCNT-SPE can detect other proteins.⁹⁴

The development of metal oxide/carbon-based aptasensors based on CNTs showed more benefits than other CNTs-based biosensors. For example, a biosensor based on integrating a composite of titanium dioxide (TiO₂), chitosan, MWCNT, and a synthetic Schiff base is suggested by a hydrothermal assay and applied for thrombin detection. This aptasensor has important advantages to biomolecule immobilization, according to the great surface area, π -conjugation structure, high conductivity produced by MWCNT, a porous structure of TiO₂ nanocrystals, as well as great amounts of amino/hydroxyl groups, and positively charged chitosan. If thrombin is present, the aptamer is

placed on the absorbent layer, preventing electrons from flowing. This leads to the inhibition of electron transfer and increases the impedance signals of TiO₂-MWCNT/chitosan-Schiff base (CHIT-SB). This aptasensor works exclusively and does not include other proteins. It also has a LOD of 1.0 fmol L⁻¹ for thrombin. Also, the clinical application was carried out by examining thrombin levels in the blood using an aptasensor and commercial enzyme-linked immunosorbent assay (ELISA) kit, and the outcomes show a promising system for the clinical analysis of the thrombin.⁹⁵

In addition to CNTs, carbon quantum dots (CQDs) can also enhance the immobilization of aptamer on the surfaces of the materials due to their good stability, hydrophilicity, biodegradability, non-toxicity, and high conductivity.⁹⁶ An MWCNT/poly(diallyldimethylammonium chloride) (PDDA)/CQD



nanocomposite is an efficient electrochemical biosensing used to detect lysozyme. The lysozyme-aptamers immobilized on the composite through covalent binding among the amino groups of the aptamer and the carboxy groups of the composite. This electrode was considered through electrochemical impedance spectroscopy and cyclic voltammetry. Increased sensitivity and enhancement of the electrochemical signal were obtained by this aptasensor (LOD of 12.9 fmol L^{-1}). This aptasensor is different from other ECAs for aptamers immobilization, such as: (1) the presence of high amounts of amino and carboxy groups in CQD; (2) high surface area in the existence of CQDs and MWCNTs; and (3) quick electron-transfer potential in the presence of MWCNTs.⁹⁷ The limitations of this study are the difficulties and expensive synthesis of NPs and CQDs; however, its high sensitivity and chemical stability make it useful for developing and improving ECAs.

Graphene, as a new carbon-based material, has a significant role in biosensors with outstanding optical, electrochemical as well as magnetic properties.⁹⁸ Graphene oxide (GO) has exclusive properties such as virtuous aqueous solubility, great surface area, and good fluorescence quenching capability. GO is considered a good energy receptor in fluorescence resonance energy transfer (FRET), which causes GO to have an extensive use in fluorescence-based aptasensor. Furthermore, GO can connect to aptamers through π - π stacking interactions.^{99,100} A non-label fluorescence aptasensor based on GO showed highly sensitive thrombin detection. The aptamer can be bound to its target and be separated from GO, and robust fluorescence was observed (LOD of 0.37 nM).¹⁰¹ A fluorescent aptasensor based on a GO can be used to detect extremely sensitive troponin I (cTnI). Thus, GO connects to the anti-cTnI aptamer and reduces its fluorescence. In fact, in the existence of cTnI, the anti-cTnI aptamer greeneries the GO combines with cTnI and then renovates the fluorescence of the aptamer. This aptasensor showed highly sensitive analytical measurement with a LOD of 0.07 ng mL^{-1} . The formed diagnosis platform suggested a high-potential instrument for cTnI detection in serum samples.¹⁰²

A novel hydrothermal assay was applied for the production of electrochemiluminescence (ECL) nanographene oxide covering TiO_2 . The NGO@TiO_2 NLPs showed a core shell shape. This platform was applied to manufacture the ECL to target recognition, ECL signal generation, and amplification. If cTnI is present, the aptamer remains from the electrode surface due to its severity, which leads to a decrease in the charge transfer resistance of the electrode and an increase in the ratio of the two ECL signals of the NGO@TiO_2 nanoluminophore (NLP) (Fig. 8A–C). As a result, according to the increase in the ECL ratio, cTnI can be detected by the aptasensor with a detection limit of $4 \times 10^{-14} \text{ mol L}^{-1}$. This method has more advantages than most of the reported electrochemical methods. This ECL approach displayed high potential in clinical diagnosis with its self-regulation ability and quick analytical performance.¹⁰³ The synthesis of noble NPs, such as AgNPs decorated GO, can provide novel electrocatalytic properties according to their inherent catalytic activities. This type of composite can increase the catalytic activities of metal NPs and attain various applications. For example, AgNP-GO was formed through a chemical

reduction assay, and one anti-thrombin aptamer (TBA1), was immobilized onto AgNP-GO. Besides, the second anti-thrombin aptamer (TBA2), keeping a thiol group at the 3' ends, was self-assembled onto the electrode surface. The formed sandwich conjugate runs to a robust oxidation signal of Ag. Then, this voltammetry signal of AgNPs was documented to display the thrombin (LOD of 0.03 nmol L^{-1}). From the characteristics of this aptasensor, the use of catalytic signal amplification caused by AgNP-GO, high-efficiency electrode, and also high sensitivity in detection by the aptamer-thrombin sandwich binding model, a sensitive electrochemical technique to the recognition of thrombin in samples was established¹⁰⁴ (Fig. 8D).

Conductive polymers, such as poly(3,4-ethylenedioxythiophene) (PEDOT), have great electrical conductivity and high electrochemical stability and are deliberated as gorgeous sensing materials according to their biocompatibility.¹⁰⁵ Graphene-based ternary nanocomposite comprises conductive polymers and displays greater activity over binary composites. The presence of conductive polymers, according to their high electrical conductivity and stability, can make them important in developing great-performance biosensors.¹⁰⁶ AuNPs/GO-doped PEDOT was deposited onto the surface of fluorine tin oxide glass at a potential of 1.0 V . MUC1 is a transmembrane glycoprotein, which is known as a biomarker that has a prognostic value in breast cancer. MUC1 aptamer was immobilized onto the composite film *via* a biotin-avidin interaction approach. This biosensor displayed a LOD of 1 fg mL^{-1} . This aptasensor also recovered about 85–93% during the detection of MUC1 (ref. 106) (Fig. 9A).

Graphite-like carbon nitride ($\text{g-C}_3\text{N}_4$) is beneficial in photocatalysts and biosensors according to the large surface and cost-effectiveness.^{107,108} $\text{g-C}_3\text{N}_4$, a 2D semiconductor, holds an exclusive electronic band structure, high compatibility, and great thermal stability.¹⁰⁹ They have uniform nitrogen, making them easily decorated with metal NPs. However, one of their challenges is the poor conductivity of $\text{g-C}_3\text{N}_4$, which restricts them from electrochemical biosensing. It remains a limitation to increase the use of the $\text{g-C}_3\text{N}_4$ in electrochemical biosensors. Many strategies can overcome these limitations, including modification with metal NPs and polymers. $\text{g-C}_3\text{N}_4$ can be modified with AgNPs, which can be considered a photocatalyst. An electrochemical aptasensor, Ag- $\text{g-C}_3\text{N}_4$, showed a great surface and high compatibility. A glass carbon electrode (GCE) modified with Ag- $\text{g-C}_3\text{N}_4$ can immobilize many thrombin-binding aptamers. This aptasensor displays an LOD of 38 fM . The assay was used to detect thrombin, and the recoveries changed from 97–103% (ref. 110) (Fig. 9B–D). Table 4 shows the role of carbon-based nanomaterials-based aptasensors in protein detection.

Metal-organic framework (MOF)-based aptasensors

Metal-organic frameworks (MOFs) were fabricated from metal ions and organic linkers, have established high consideration, and have been extensively applied in catalysis and separation according to their flexible structures, and exclusive porosity.¹¹³ MOFs-based materials can be applied to design different



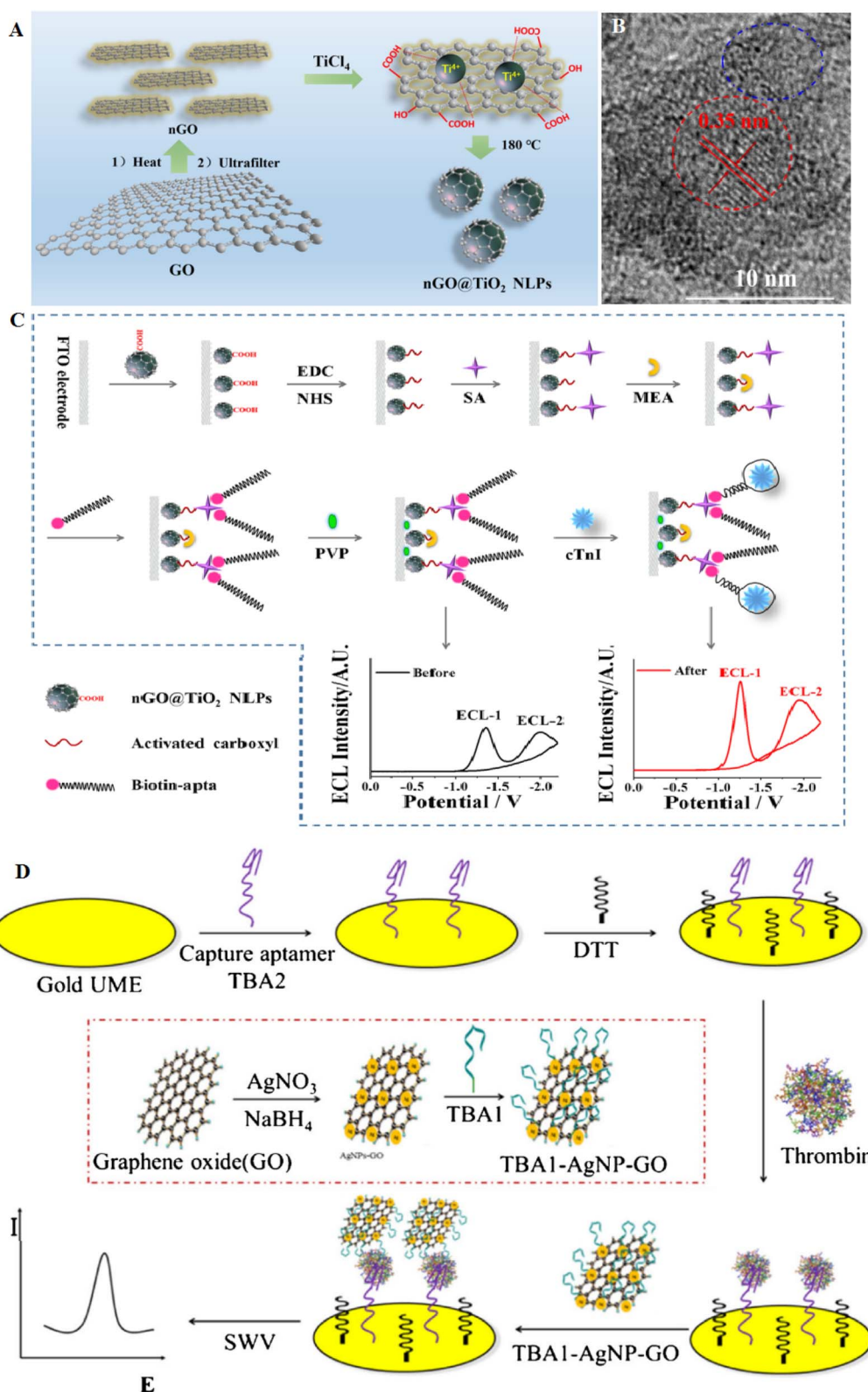


Fig. 8 A carbon-based aptasensor for highly efficient recognition of thrombin. (A) The formation of nGO@TiO₂ NLPs via hydrothermal assay. (B) TEM image of the nGO@TiO₂ NLPs with the size of 6 nm. (C) Schematic of the ratiometric ECL aptasensor to cTnI detection based on nGO@TiO₂ NLPs, reproduced from ref. 103 with permission from American Chemical Society, Copyright 2019. (D) Scheme of the electrochemical aptasensor for thrombin at gold microelectrode using TBA1-AgNP-GO as signal probe, reproduced from ref. 104 with permission from Elsevier, Copyright 2018.





Fig. 9 A novel electrochemical aptasensor platform for detection of a wide range of analytes, including thrombin, with a new mechanism and recoverable electrodes. (A) Schematic illustration of the formation of APT/SA/AuNPs-GO-PEDOT coated FTO electrode in order to breast cancer diagnosis, reproduced from ref. 106 with permission from Elsevier, Copyright 2018. (B) Schematic of the aptasensor construction process. (C) EIS signal of the aptasensor incubated with variable thrombin concentrations, from 0.1 pM to 20 nM. (D) Charge-transfer resistance changes of various concentrations of thrombin, reproduced from ref. 110 with permission from Elsevier, Copyright 2020.



Table 4 List of different types of carbon nanomaterials applied as transducers for the detection of protein^a

Nanomaterials	Aptamer	Biosensing principle	Target protein	Properties	LOD	Ref.
PEI-rGO	ATCCGTCACACCTGCTCTTAAT TACAGGCAGTTCCACTTAGACAGA CACACGAATGGTGGTCCCGTAT	EL	Cardiac myoglobin (cMb)	High specificity, high reproducibility	0.97 pg mL ⁻¹	22
g-C ₃ N ₄	GGT TGG TGT GGT TGG	EL	Thrombin	Wide linear range, high selectivity, reproducibility	38 fM	110
rGO/GCE	N/A	EL	Carcinoembryonic antigen (CEA)	Without immobilization of bioprobe, high sensitivity	80 pg mL ⁻¹	111
MWCNTs	CCCTCCTTTCTTCGACGCTAGA TCTGCTGCGTTGTCCGA	EL	Myoglobin	Greatly specific recognition capability of the aptamer	2.2 pM	112

^a EL: electrochemical.

electrochemical sensing systems, such as electrochemical and photoelectrochemical biosensors.¹¹⁴ MOFs-MIL-53(Al) showed high thermal and chemical stability.¹¹⁵ Researchers have described that Pd NPs coated MIL-53 were used to create an electrochemiluminescence immunosensor for the diagnosis of amyloid- β proteins.¹¹⁶ The combination of aptamers with MOFs causes the formation of nanostructure with properties of both aptamer and MOFs.^{117,118} MOFs with large surface areas permit an extensive range of ssDNA or RNA to adhere to the surface. Additionally, MOFs with adjustable pore diameters permit various nucleic acids to be immobilized on their surface. According to these properties, different types of MOFs-based aptasensors have been established, each with advantages such as great sensitivity and visual detection activity.¹¹⁹ Currently, there is no clear roadmap to select desirable MOFs for different biosensor applications; however, with the current trends in the literature, it is vividly predictable that in the near future, a pattern to choose the desired MOF for specific biosensor applications is possible.

The fabrication of an electrochemical aptasensor based on the MIL-53(Al) decorated with Au@Pt NPs can be used to identify SARS-CoV-2 nucleocapsid protein through the catalysis of the nanomaterials. First, the immobilization of two thiol-modified aptamers on the surface of the gold electrode occurred; at that time, the Au@Pt/MIL-53 (Al) was decorated through horseradish peroxidase (HRP) and hemin/G-quadruplex DNAzyme as signal nanoprobe. This aptasensor has a high potential for detecting nCoV-NP, demonstrating the results with an extensive linear range from 0.02–50 ng mL⁻¹ and the LOD of 8.3 pg mL⁻¹ for SARS-CoV-2. The MOFs-based aptasensor showed high potential in the early detection of selectivity markers associated to the virus (Fig. 8).¹¹⁴

Magnetic MOFs are a class of MOFs with magnetic ability, which can be refined and separated throughout the synthesis by presenting an external magnetic field for protein separation.¹²⁰ Sensitive detection of cTnI was performed by a voltammetric aptasensor based on dual-aptamer (dAPT) attached to nanotetrahedron (NTH) DNA.¹²¹ A meaningful increase in cTnI detection was observed according to the great binding affinity among the aptamer and cTnI. Owing to the advantage of the great surface area of MOFs, the nanoprobles can immobilize

DNAzyme, HRP enzyme, and Au@PtNPs, intensifying the electrochemical signal. This aptasensor creates an excellent activity in the detection of cTnI with great specificity, high stability, and LOD of about 5 pg mL⁻¹. Since this aptasensor usage still has several limitations, including the time-consuming fabrication procedure, new technologies such as microfluidics is needed to improve its performance in clinical diagnosis.

Developing a dual-signal sensing system is critical in bioanalysis according to the likelihood of using both simultaneously. Each method has benefits and limitations that can complement each other's abilities in two-manner detection. As a result, being stronger and more resistant is observed in the case of sensors with dual-mode detection features. For example, a biosensor was formed to detect C-reactive protein (CRP) using the two modes of the optical transducer, fluorometry and colorimetry. In this aptasensor, the combination of RNA on Cu-MOF was done. Cu-MOF can exhibit fluorescence and peroxidase enzymatic activity, which can be applied as signal transduction. CRP binding RNA as a greatly sensitive recognition element immobilized on the Cu-MOF (Fig. 10). Besides, with the addition of CRP to the RNA/Cu-MOF, the deliver of RNA from the Cu-MOF surface has occurred. Here, the LOD of this aptasensor was 40 pg mL⁻¹ in fluorescence sensing and LOD of 240 pg mL⁻¹ in the colorimetry approach. This LOD is lesser than the LOD of nephelometric methods applied in clinical applications and CRP assays with 1 μ g mL⁻¹. The MOF-aptasensor was effectively used to diagnose CRP in COVID-19 patients.¹²²

The objective of the disclosed method (US Patent) was to provide a low-cost, and greatly sensitive way of preparing a ratiometric electrochemical aptasensor for vanillin. The method involved several steps, including dissolving 2-methylimidazole (Hmim) in ethanol, adding ferrocene and Ketjenblack, stirring well, adding zinc nitrate, and then transferring to a high-pressure reactor to form a ferrocene-Ketjenblack-ZIF-8 composite. The nanocomposite was applied to modify a glassy carbon electrode, which is immersed in a buffer electrolyte solution and chloroauric acid to generate gold NPs *in situ* by one-step electrodeposition, thereby obtaining a gold NP-deposited ZIF-8 nanocomposite. Then, the aptamer DNA of vanillin was connected to the surface of the gold NP-deposited ZIF-8 nanocomposite to obtain an aptamer-modified Au NP-





Fig. 10 MOF-based electrodes for the detection of coronavirus *via* aptamer-surface decoration. (A) (I) manufacture procedure of the nanoprobe (II) and the aptasensor to recognition of COVID-19. The Au@Pt NPs were connected to the MOFs $\text{NH}_2\text{-MIL-53(Al)}$ NPs *via* amino groups. The catalysis of HRP and GQH DNAzyme nanoprobe after the decorated HRP onto the NPs, reproduced from ref. 114 with permission from Elsevier, Copyright 2021. (B) The procedures for CRP aptasensor based on Cu-MOF. (C) FESEM of Cu-MOF, the materials have polydisperse decahedral crystal forms. (D) Fluorescence spectra of MOF-RNA probe treated with various concentrations of CRP, reproduced from ref. 122 with permission from Elsevier, Copyright 2022.

ZIF-8 composite-modified electrode. The modified electrode was then applied as a working electrode, and the electrochemical square-wave voltammetry curves are measured in the existence of various vanillin concentrations. The ratios of the peak intensities of ferrocene and vanillin were applied as a reference signal and a response signal, respectively, to construct the ratiometric electrochemical aptasensor for

vanillin. The method had several parameters that can be adjusted for optimal results for commercializations. In step (1), the reaction temperature, reaction time, and mass concentration ratio of Hmim, ferrocene, Ketjenblack to zinc nitrate can be varied. In step (2), the dosage of Nafion, mass concentration of the ZIF-8 nanocomposite, the concentration of chloroauric acid, potential range, scanning rate, and scanning number can be



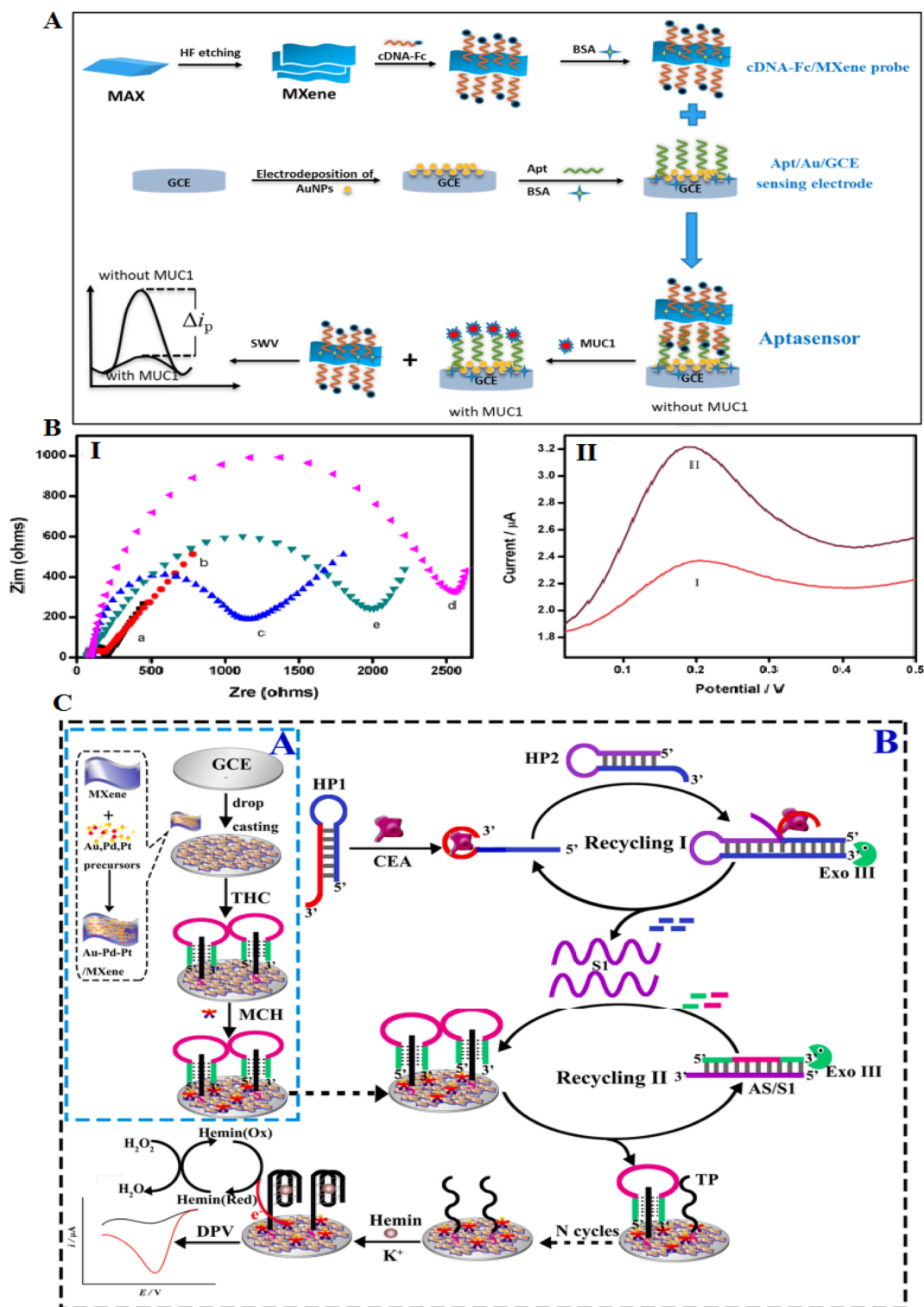


Fig. 11 MXene-based aptasensors for recognition of low concentrations of CEA. (A) Schematic of the formation process of the competitive electrochemical aptasensor. (B) (I) EIS of various modified electrodes in 5 mM $\text{K}_3[\text{Fe}(\text{CN})_6]/\text{K}_4[\text{Fe}(\text{CN})_6]$ solution. (II) SWV of various electrodes in PBS buffer (0.1 M): (I) cDNA-Fc/Apt/Au/GCE, (II) cDNA-Fc/MXene/Apt/Au/GCE, reproduced from ref. 126 with permission from Elsevier, Copyright 2020. (C) (A) The synthesis assay for formed nanocomposite. (C) The Exo III-related dual recycling amplifications to catalytic CEA detection, reproduced from ref. 132 with permission from Elsevier, Copyright 2022.



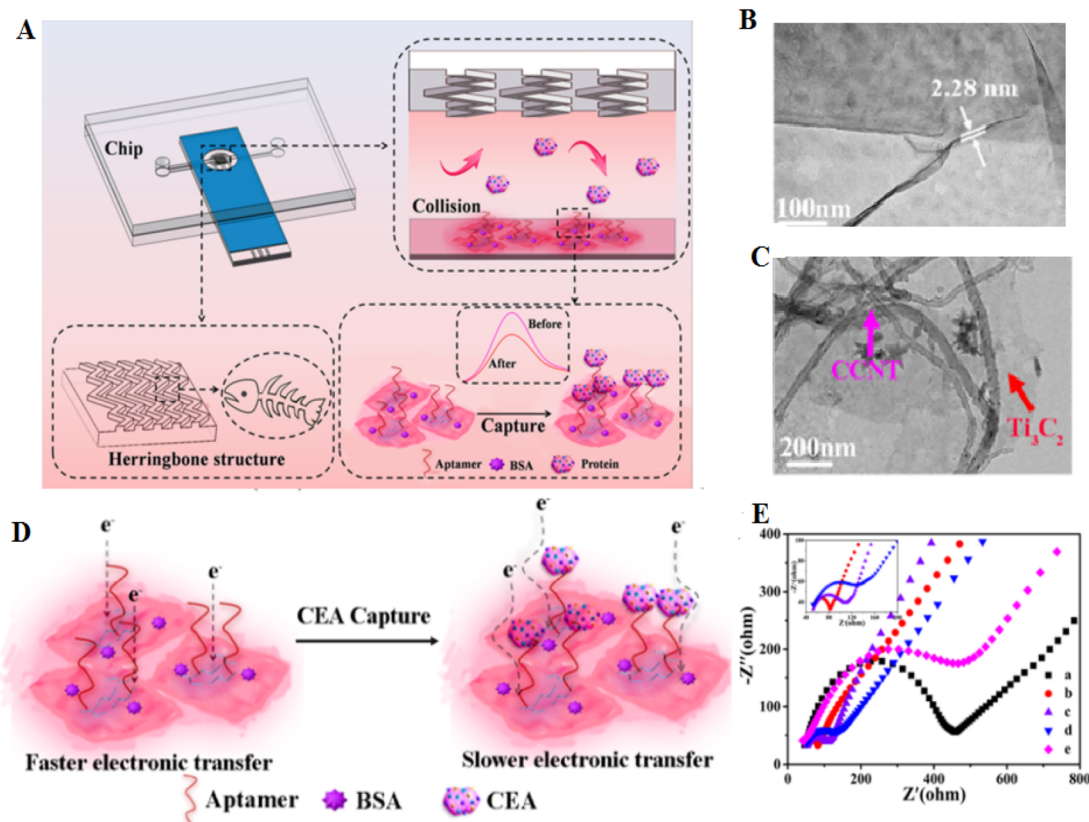


Fig. 12 This is a novel system for capturing and detection of CEA, based on microfluidics and aptamer. (A) Microfluidic electrochemical system for detection of CEA. (B) FESEM image of Ti_3C_2 nanosheets and (C) $\text{He@CCNT}/\text{Ti}_3\text{C}_2$. (D) Diagram of electron transfer before and after CEA capture. (E) The decreases of the R_{et} value of $\text{He@CCNT}/\text{Ti}_3\text{C}_2/\text{SPCE}$ (curve b) compared with that of bare SPCE (curve a), endorsing the good conductivity of $\text{He@CCNT}/\text{Ti}_3\text{C}_2$, reproduced from ref. 133 with permission from American Chemical Society, Copyright 2021.

Metallic nanostructures-based aptasensors: the road of commercialization

Aptasensors have revealed immense potential as a diagnostic tool for a wide range of diseases and disorders, such as cancer, infectious diseases, and autoimmune disorders. One of the most promising types of aptasensors is the metallic nanostructures-based aptasensor. These sensors are built using metallic NPs such as gold, silver, and platinum, and can detect a variety of target analytes with high sensitivity and specificity. Despite their potential, there are several challenges that must be addressed before metallic nanostructures-based aptasensors can be successfully commercialized.

(1) **Stability and reproducibility:** one of the biggest challenges facing metallic nanostructures-based aptasensors is achieving stability and reproducibility. Metallic NPs are known to be sensitive to changes in their environment, such as pH, temperature, and ionic strength. These changes can cause the NPs to aggregate or change shape, which can affect their optical and electrical properties, and thus, their ability to detect target analytes. In addition, the synthesis of metallic NPs can be variable, leading to differences in their size, shape, and surface chemistry, which can also affect their performance as

aptasensors. To address these challenges, researchers have developed methods to stabilize metallic NPs and improve their reproducibility. For example, the use of biocompatible coatings such as polyethylene glycol (PEG) or silica can prevent aggregation and stabilize the NPs in solution. In addition, the use of template-assisted synthesis or microfluidic techniques can control the size and shape of the NPs, leading to more reproducible synthesis.

(2) **Sensitivity and selectivity:** another challenge facing metallic nanostructures-based aptasensors is achieving high sensitivity and selectivity. Metallic NPs can enhance the detection of target analytes through localized surface plasmon resonance (LSPR) and surface-enhanced Raman scattering (SERS). However, the performance of these sensors can be affected by factors such as the size and shape of the NPs, the distance between the NPs, and the type and density of functional groups on the surface of the NPs. To address these challenges, researchers have developed strategies to optimize the performance of metallic nanostructures-based aptasensors. For example, the use of different shapes of NPs such as nanospheres, nanorods, and nanostars can enhance the LSPR effect and improve the sensitivity of the sensor. In addition, the use of functionalized ligands such as thiolated DNA can improve the selectivity of the sensor by specifically binding to the target analyte.



Table 5 List of various types of MXene nanomaterials applied as transducers for the detection of proteins^a

Nanomaterials	Aptamer	Biosensing principle	Target protein	Properties	LOD	Ref.
Ti ₃ C ₂ T _x /ZIF-8/GCE/Apt	AGTCAGTGTGGAAAAATCTCTAGC	ECL	HIV protein	Good ECL response, improved electrical conductivity	0.3 fM	135
AuNPs/Ti ₃ C ₂ T _x /Apt	N/A	ECL	CD63 protein	High sensitivity and selectivity to exosome and their proteins, accurate detection	30 particle per μ L	136
Ti ₃ C ₂ /PAMAM-Au NPs/CD63-Apt/GCE	TTTTTTCACCCACCTCGCTCCC GTGACACTAATGCTA	EC	CD63 protein	High specificity even in serum samples, excellent conductivity	229 particle per μ L	137
Cy3-CD63-Apt/Ti ₃ C ₂	CACCCACCTCGCTCCC TGACACTAATGCTA	FRET	CD63 protein	Detection of multiple biomarkers on the exosome surface	1.4×10^3 particles per mL	138
Apt/He@CCNT/Ti ₃ C ₂ /SPCE	N/A	EC	GEA	Increases the movability of the sensing platform	2.88 pg mL ⁻¹	133

^a ECL: electrochemiluminescence. EC: electrochemical. FRET: fluorescence resonance energy transfer.

(3) Manufacturing and scalability: another challenge facing metallic nanostructures-based aptasensors is manufacturing and scalability. The synthesis of metallic NPs is often carried out in small batches using specialized equipment, which can be time-consuming and expensive. In addition, the functionalization of the NPs with aptamers requires careful optimization to ensure their specificity and stability. To address these challenges, researchers have developed methods for large-scale synthesis and functionalization of metallic NPs. For example, the use of microwave-assisted synthesis or continuous flow reactors can increase the yield and reduce the time required for synthesis. In addition, the use of automated functionalization platforms can improve the consistency and reproducibility of aptamer immobilization on the surface of the NPs.

(4) Regulatory approval: finally, one of the most significant challenges facing metallic nanostructures-based aptasensors is obtaining regulatory approval for commercialization. In many countries, medical devices such as aptasensors must undergo rigorous testing and regulatory approval before they can be marketed to the public. This process can be time-consuming and expensive, requiring extensive clinical trials and documentation of safety and efficacy. To address these challenges, researchers and manufacturers must work closely with regulatory agencies to ensure that their aptasensors meet the required standards for safety and effectiveness. This may involve consulting with regulatory experts, conducting pre-clinical and clinical trials, and providing detailed documentation of the device's performance and safety. In addition to regulatory approval, the marketing and distribution of aptasensors also face challenges related to intellectual property and market competition. Companies must protect their intellectual property and secure patents for their technology to prevent infringement by competitors. They must also develop effective marketing strategies to promote their products and differentiate them from other similar devices in the market.

Apart from the challenges mentioned above, there are several other obstacles that need to be overcome for the commercialization of metallic nanostructures-based aptasensors. One such issue is the reproducibility of the sensor response. The variability in the size, shape, and composition of metallic NPs used in the synthesis of the sensor can lead to variations in the signal response. Therefore, to achieve reproducible results, standardized synthesis and fabrication protocols need to be established. Another challenge is the stability of the sensor over time. The metallic NPs used in the sensor fabrication can undergo aggregation or oxidation, leading to a decrease in the sensor's sensitivity and stability. Moreover, the aptamers used in the sensor can undergo denaturation or degradation, leading to a reduction in their affinity and specificity towards the target analyte. Therefore, the development of strategies to increase the stability of both the NPs and aptamers is crucial for the successful commercialization of the sensor. Another critical issue is the cost-effectiveness of the sensor. The synthesis and fabrication of metallic NPs are relatively expensive compared to other materials used in sensor fabrication. Moreover, the use of sophisticated instrumentation for the detection of the signal response can also add to the overall cost of the sensor. Therefore, efforts should be

made to reduce the cost of the sensor without compromising its performance. Despite these challenges, metallic nanostructures-based aptasensors keep high potential for the commercialization of novel sensing technologies. The ability of these sensors to detect analytes with high sensitivity, selectivity, and accuracy makes them an attractive option for a wide range of applications, such as clinical diagnostics, and environmental monitoring. Moreover, the versatility of the aptamer sequences allows the development of sensors for a wide range of analytes, including small biomolecules, proteins, and cells. In conclusion, the commercialization of aptasensors presents several challenges that require to be overcome for their effective integration into the market. The development of reliable, stable, and cost-effective sensors with high selectivity and sensitivity towards the target analyte is crucial for their acceptance by end-users. The advancement of technologies for the synthesis and fabrication of metallic nanostructures, as well as the optimization of signal detection methods, can pave the way for the successful commercialization of aptasensors. With the current pace of research and development in this field, it is expected that aptasensors will become an integral part of the diagnostic and monitoring tools used in healthcare and other industries in the near future.

Conclusion and future perspective

This review discussed recent progress in developing aptasensors based on metal nanomaterials as a transducer for the specific detection of proteins. The integration of metal nanomaterials indicates a remarkable improvement in analytical activity in aptasensors, in line with progress in biosensors based on metal nanomaterials. Nanomaterials add to the sensitivity of the biosensors by providing a great surface area for aptamer immobilization and by deliberating their physical and electrochemical properties to the biosensor. Metal nanomaterials-based aptasensors seem highly beneficial due to their detection sensitivity ranging from picomolar to femtomolar, which is greater than common methods, such as chromatography. Noble metal/metal oxide nanomaterials, such as AgNPs, AuNPs, QDs, *etc.*, have unique properties for accurate and sensitive biosensors. The use of composite nanomaterials and the mixture use of two or more nanomaterials shows the current tendency toward synergetic effects on sensitivity and stability enhancement in aptasensors. MXene, with its exclusive properties, functionalization, and conductivity, shows greater electrochemical properties than other materials; therefore, ample functional groups on their surface are vital for developing increased sensors.

Despite these benefits, aptasensors are still in their beginning stages compared to other biosensing techniques. However, aptasensors based on metal nanomaterials could be suitable when critical challenges are addressed:

(1) Aptamers have a short half-life, which is a serious problem for the aptasensor industry. As a result, there is a need for modification and changes in aptamers to increase their half-life, such as the modification of the aptamer with 2'-O-methylation, which may considerably enhance the aptamer's half-life, without compromising their affinity.¹³⁹

(2) Mass production and additional costs, including the scalability of manufacturing processes, can limit aptasensors based on metal NPs, including MOF and MXene. This type of aptasensors was made for laboratory-scale production and cannot be used for mass construction. Thus, aptasensors must be marketed on a large scale, and manufacturing techniques must be developed to build a large supply of aptasensors that are produced using inexpensive cost-effective precursors.

(3) One of the important challenges is the oxidation of metal nanomaterials, such as noble metal nanomaterials and MXenes in aptasensors, which affects the stability of sensors in hot environments; thus, the investigation at the molecular level, as well as the interactions among different metal NPs and aptamer is essential. One suggestion would be the use of natural products and leaf extracts as capping agents and surface decoration agents.

(4) It is vital to investigate the toxicity and compatibility of metal nanomaterials-based aptasensor systems, especially in cells or *in vivo*. However, selecting flexible metal NPs composites allows researchers to build customizable aptasensors with the desired functionality. For instance, since fluorine is toxic, fluorine-free MXene $Ti_3C_2T_x$ is better than HF-etched MXene when used in *in vivo* detection.¹⁴⁰ Since these theories need to be tested widely, we encourage researchers to focus on these topics to address the current challenges.

(5) A more comprehensive study of aptamer immobilization approaches on different metal NP-based aptasensors is needed for cost-effective sensing of multiple molecules, especially for POC systems.

Commercialization requires inventions to confirm that these techniques can be applied regularly. Continuous benefits in all these metal nanomaterials-autosensing are possible by joining multiple methods. Thus, future hard work should be focused on the building of less erudite technologies that can be employed *via* non-professional people. The assessment of metal nanomaterials-based aptasensors and future propensity are predicted to further discover prospects for biomedical sensing developments, especially for protein detection.

Author contributions

N. R., S. A., K. R., S. C., and R. N. V. wrote the main manuscript, conceptualized and finalized the manuscript. All authors reviewed and approved the manuscript.

Conflicts of interest

The authors declare they have no conflict of interest.

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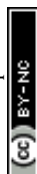
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