

Cite this: *Chem. Sci.*, 2023, 14, 14100

All publication charges for this article have been paid for by the Royal Society of Chemistry

Received 7th August 2023

Accepted 22nd November 2023

DOI: 10.1039/d3sc04116f

rsc.li/chemical-science

A single phosphorylation mechanism in early metabolism – the case of phosphoenolpyruvate†

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Phosphorylation is thought to be one of the fundamental reactions for the emergence of metabolism. Nearly all enzymatic phosphorylation reactions in the anabolic core of microbial metabolism act on carboxylates to give acyl phosphates, with a notable exception – the phosphorylation of pyruvate to phosphoenolpyruvate (PEP), which involves an enolate. We wondered whether an ancestral mechanism for the phosphorylation of pyruvate to PEP could also have involved carboxylate phosphorylation rather than the modern enzymatic form. The phosphorylation of pyruvate with P₄O₁₀ as a model phosphorylating agent was found to indeed occur *via* carboxylate phosphorylation, as verified by mechanistic studies using model substrates, time course experiments, liquid and solid-state NMR spectroscopy, and DFT calculations. The *in situ* generated acyl phosphate subsequently undergoes an intramolecular phosphoryl transfer to yield PEP. A single phosphorylation mechanism acting on carboxylates appears sufficient to initiate metabolic networks that include PEP, strengthening the case that metabolism emerged from self-organized chemistry.

Introduction

Dynamic self-organized systems exhibiting complex behavior are generally thought to arise when a small number of particular rules or mechanisms simultaneously act on a set of substrates. As the core of microbial metabolism features a small number of repeating reaction mechanisms,¹ it has been suggested that it may have initiated through chemical self-organization.^{2–9} Several experimental reports support the idea that core metabolic pathways, including substantial segments of the central (r)TCA cycle and related pathways, could have initiated in self-organized chemistry, possibly as early as in prebiotic chemistry.^{2,10–22} However, regardless of whether metabolism emerged enzymatically or nonenzymatically, there is no doubt that phosphorylation is one of the crucial mechanisms within these biological pathways. In addition to introducing phosphoryl groups into the molecular backbone of sugars, phosphorylation can activate functional groups toward reactions that would otherwise be energetically unfeasible.^{23,24}

The most common type of phosphorylation in the core pathways of microbial anabolism (the acetyl CoA pathway, the rTCA cycle, and gluconeogenesis) is the phosphorylation of carboxylates to acyl phosphates,^{25–30} in which nucleotide

triphosphates such as adenosine triphosphate (ATP) act as the phosphoryl group donor (Fig. 1). However, the second type of phosphorylation in the anabolic core is quite unique – the conversion of pyruvate or oxaloacetate to form phosphoenolpyruvate (PEP), the metabolite with the most energetic phosphate-bond (ΔG^0 of hydrolysis = -61.9 kJ mol⁻¹).³¹ Unlike the other reactions in Fig. 1, this involves the phosphorylation of an enolate (Fig. 1, red box), rather than a carboxylate. In the enzymatic mechanism, the enolate of pyruvate is generated within the active site of the enzyme (stabilized by Mg²⁺), which subsequently undergoes phosphoryl transfer from a phosphohistidine intermediate to yield PEP.^{32–34} In the following steps of anabolic pathways, PEP provides all carbon and phosphoryl groups for the sugar-phosphate intermediates of gluconeogenesis and the pentose phosphate pathway, including ribose-5-phosphate, the building block for ribonucleotide biosynthesis. Thus, enzymatic phosphorylation serves not only a bioenergetic role but also a fundamental structural role in biochemistry through PEP. This dual function suggests that phosphorylation reactions and PEP were both present in the earliest metabolic networks that supported the emergence of life.³ However, if metabolism was the result of chemical self-organization, one would expect that a single phosphorylation mechanism would be sufficient to account for the emergence of the core metabolic network, including the formation of PEP.

PEP is kinetically stable toward hydrolysis at room temperature but readily hydrolyses at elevated temperatures (pH 1–7) or in the presence of metal ions.^{35,36} Interestingly, studies on the nonenzymatic hydrolysis of PEP, the reverse reaction of phosphorylation, have concluded that the mechanism proceeds

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† Electronic supplementary information (ESI) available. See DOI: <https://doi.org/10.1039/d3sc04116f>



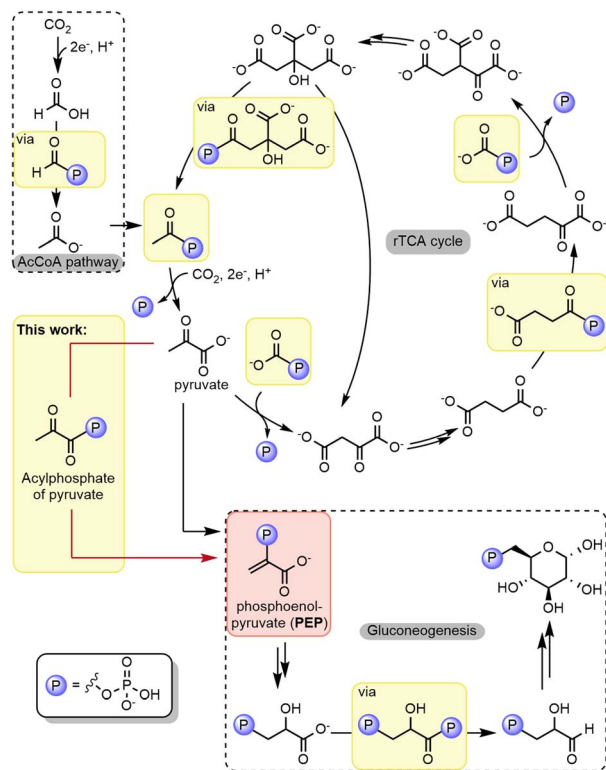


Fig. 1 Phosphorylation reactions in core metabolism. Examples of acyl phosphates (highlighted in yellow boxes) involved in different chemical transformations and PEP (highlighted in the red box).

through an acyl phosphate intermediate (Fig. 2).^{35–37} Initially, the enol phosphate (PEP) was suggested to undergo intramolecular nucleophilic attack by the carboxy group, forming a cyclic pentacoordinate phosphorus intermediate (cPEP·H₂O) in equilibrium with the cyclic tetravalent intermediate (cPEP) and the acyl phosphate (AcP-Pyr). Finally, irreversible hydrolysis of the acyclic product leads to the formation of pyruvic acid (Pyr) and inorganic phosphate. It therefore seems plausible that, under the right conditions, phosphorylation of pyruvate to PEP might occur in the reverse way, offering a unified mechanism for all the phosphorylations in core anabolism.

We hypothesized that PEP formation *via* an enolate is the result of biological evolution and that an ancient pathway, whether enzymatic or nonenzymatic, yielded PEP through the direct phosphorylation of the carboxylate of pyruvate followed

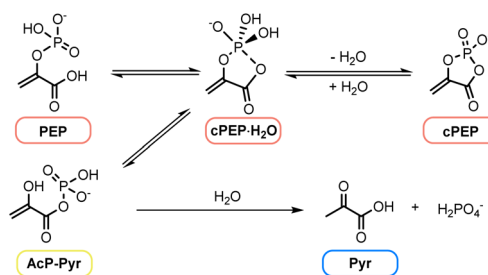


Fig. 2 Nonenzymatic hydrolysis of PEP.³⁷

by intramolecular transfer of the phosphoryl group (Fig. 1, yellow box). Although syntheses of PEP have been reported in various contexts,^{38–40} the only direct nonenzymatic phosphorylation of pyruvate to PEP was reported by Kiessling using POCl₃ in quinoline as a solvent, which, even after further improvements, yielded a maximum of 9% PEP.^{41,42} However, the mechanism of this reaction is unknown and has never been studied. Considering the energetically highly favorable hydrolysis of PEP, phosphorylation of pyruvate to PEP *via* initial carboxylate phosphorylation would be unlikely to happen in bulk water. Indeed, biological phosphorylation and many potentially prebiotic phosphorylation reactions occur in environments where water is largely excluded.^{43–45} We, therefore, elected to study the mechanism of the Kiessling-type reaction as a model for the potential ancestral enzymatic or nonenzymatic phosphorylation of pyruvate to PEP. Our goal is not to identify prebiotically plausible conditions for such a reaction but simply to determine whether a single phosphorylation mechanism might account for the phosphorylation reactions in core metabolism, strengthening the case for a self-organized metabolic origin.

Results and discussion

Nonenzymatic phosphorylation of pyruvate to PEP, reaction progress, and intermediates

Considering that the free energy of hydrolysis of PEP is 61.9 kJ mol⁻¹,³¹ the reverse reaction requires a strong phosphate donor to overcome this energy barrier. Although POCl₃ was reported to phosphorylate pyruvate,⁴¹ we wondered whether simpler oxygen-based phosphorylating agents would be able to achieve the phosphorylation reaction of pyruvate to PEP. After some screening, we found that when Pyr (0.06 mmol) was mixed with one equiv. of P₄O₁₀ in paste conditions at 60 °C for 90 minutes, the characteristic ¹H NMR signals of PEP were observed after solubilization of the paste in H₂O, but only in trace quantities (Table S1, entry 1 and p. S11; also confirmed by mass spectroscopy, see ESI Section VI.A.3 and p. S74†). The pH after solubilization proved to be highly acidic due to the formation of H₃PO₄ (roughly pH 1). In line with the previous reports of Kiessling and analogous to the formation of the phosphohistidine intermediate found in the biological pathway,^{32–34} the addition of nitrogenous bases improved the yield of PEP (Table S2 and p. S13†).⁴⁶ The highest PEP yield (7%) was obtained with the following conditions: P₄O₁₀ (1 equiv.), pyridine (2 equiv.), paste conditions, 60 °C, and a reaction time of 1–2 h (Table S3 and p. S23†). P₄O₁₀ is known to decompose to various activated phosphate species,⁴⁷ and to react with pyridine at 70 °C to form bipyridinium phosphates.^{46,48} Therefore, a series of further phosphorylating agents were evaluated as potential reactive intermediates (Table S4 and p. S24†). Among them, bipyridinium phosphate P₂O₅L₂ (L = pyridine) was found to form PEP in 3% yield after 90 min at 60 °C (Table S4,† entry 9). Polyphosphoric acid gave only traces of PEP (Table S4,† entry 8), and all other polyphosphates were unreactive. The phosphorylation might, therefore, involve a combination of P₄O₁₀ and *N*-phosphorylated adducts as active species.



To gain further insight into the mass balance and mechanism, a series of identical reactions were quenched at different time points and examined by ^1H and ^{31}P NMR. The three main compounds observed after quenching are shown in Fig. 3A, with a ^1H NMR spectrum of the reaction quenched after 3 min depicted in Fig. 3B. The qualitative evolution of the reaction (Table S5 and p. S24[†]) is shown in Fig. 3C. Immediately after setting up the reaction at 60 °C and quenching, a phosphorylated compound **P1-pyr-py** is detected prior to significant **PEP** formation (3 : 1 ratio of **P1-pyr-py**/**PEP** at $t = 1$ min). Based on the evolution of their relative concentration, **P1-pyr-py** is likely the key intermediate in the formation of **PEP**, as it can afford **PEP** after elimination of HPy^+ .

To determine the structure of **P1-pyr-py**, its formation was maximized by scaling up and quenching after 2–5 min. A combination of NMR experiments using through-bond (HMBC HSQC, HMQC) and through-space (NOESY, HOESY) correlation techniques, as well as mass spectrometry, allowed **P1-pyr-py** to be assigned as a phosphorylated covalent adduct of pyruvate and pyridine (ESI, Section VI. A.2 and p. S69[†]).⁴⁹ A higher yield of the adduct was observed when 1-methylimidazole (**Im**) was used as base instead of pyridine (**Py**), allowing easier characterization (ESI, Section VI. A.1 and p. S61[†]). Here, **P1** refers to

the general structure of the phosphorylated covalent adducts formed from a ketoacid and a heterocyclic base (Fig. 3A), whereas **P1-pyr-py** and **P1-pyr-im** refer to the specific adducts formed from pyruvate and pyridine (**Py**) or 1-methylimidazole (**Im**), respectively. In solution, **P1-pyr-py** is stable below pH 5, while **P1-pyr-im** is also stable at basic pH (Table S6 and p. S27[†]).

Furthermore, the side-product **P3-pyr** was also identified, which might be formed through the addition of an enol at an activated carboxylic group. To determine the structure of **P3-pyr**, a similar NMR analysis was performed after suspending the paste in CD_3CN , as **P3-pyr** was observed to be highly soluble under these conditions. The structure of **P3-pyr** was assigned as a partially enolized dimer of pyruvate (Fig. 3A, ESI, Section VI.A.4 and p. S75[†]). In water, **P3-pyr** proved to be stable under acidic conditions but hydrolyzed at elevated temperatures (Table S6 and p. S27[†]).

Finally, by ^1H NMR, pyruvic acid, its hydrate, and acetic acid were also detected (ESI, Section VI.A.5 and p. S79[†]). Several hydrolysis products of P_4O_{10} were identified by ^{31}P NMR, along with two signals near -5.3 and -4.5 ppm, which were assigned to **P1-pyr-py** and **PEP**, respectively (Table S5 and p. S26[†]).

Paste composition

To differentiate which products are formed directly within the paste *versus* those that are formed only upon hydrolysis during quenching of the paste with water, solid-state NMR was used to characterize the paste (for details, see ESI, Section VI.B and p. S126[†]). Magic Angle Spinning NMR (MAS-NMR) allowed the characterization of the paste (phosphorylation conditions with pyridine, 10 min, 60 °C). One-dimensional ^1H MAS NMR showed the characteristic ^1H NMR signals of **PEP** and **P3-pyr** (ESI, Section VI.B.1, p. S127 and S128[†]), whereas 2D NMR (^{31}P - ^1H) showed some polyphosphates and pyridinium phosphate (Fig. 4A). Next, the paste (phosphorylation conditions with 1-methylimidazole, 10 min, 60 °C) was analyzed by HR-MAS-NMR spectroscopy. One-dimensional ^1H HR-MAS NMR spectra also showed the characteristic ^1H NMR signals of **PEP** and **P3-pyr** (ESI, Section VI.B.2 and p. S130[†]).

The 2D NMR (^1H , ^1H RFDR) analysis⁵⁰ confirmed the presence of **PEP** and **P3-pyr** in the paste (Fig. 4B). Additionally, the



Fig. 3 (A) Intermediates and side products in the phosphorylation of pyruvate to **PEP** with P_4O_{10} . (B) Typical ^1H NMR spectrum ($\text{H}_2\text{O}/\text{D}_2\text{O}$ 9 : 1 mixture, 400 MHz) of the quenched reaction mixture after 3 min at 60 °C. (C) Qualitative evolution of the main species in the reaction (for details, see Table S5 and p. S24[†]).

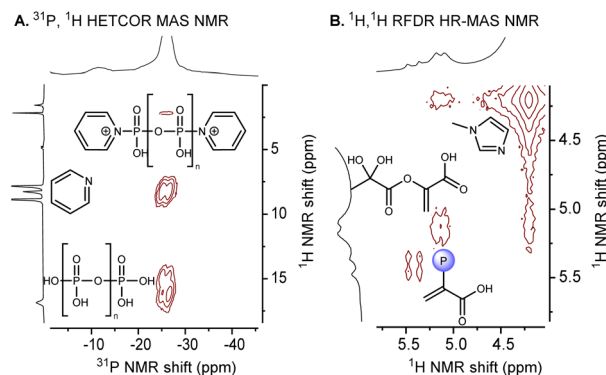


Fig. 4 (A) Characterization of the paste by ^{31}P , ^1H HETCOR MAS NMR. (B) Characterization of the paste by ^1H , ^1H RFDR HR-MAS NMR.



paste was dissolved in various anhydrous solvents to verify the presence of **PEP** (Table S13 and p. S131†). In general, the paste was poorly soluble in organic solvents (acetone- d_3 , CD_3CN , $CDCl_3$) and partially soluble in DMSO- d_6 . Among the different solvents tested, **PEP** could be detected in CD_3CN and DMSO- d_6 along with **P3-pyr**, whereas **P1-pyr-py** was not detected (Table S13,† entries 2 and 4). Thus, **PEP** and **P3-pyr** are formed under the paste conditions and not only after hydrolysis.

Reaction with further substrates

To obtain insights into the mechanistic requirements of the reaction, experiments with additional substrates were performed (Fig. 5, see also Tables S8, S9 and p. S30–S44†). First, reactions of pyruvate derivatives were studied under the established paste conditions after 90 min at 60 °C. The expected phosphoenol products were obtained when one or two $-CH_3$ groups were introduced at the α -position, yielding 9% of **P2-1Me** and 3% of **P2-2Me** (21% without base), respectively. Similarly, phenylpyruvic acid was phosphorylated to **P2-Ph** in 15% yield. Next, α -ketoacids bearing an additional carboxy group were tested. For α -ketoglutarate, **P2-glu** was only obtained in 0.6% yield; the main phosphorylated product was assigned as the cyclic species **P2'-glu** (Fig. 6A, 3%). Concerning oxaloacetic acid, the phosphoenol product was obtained in 3% yield (**P2-ox**, Fig. 5) under base-free conditions in order to avoid decarboxylation of the substrate to pyruvate. Under standard conditions, **PEP** is also obtained from oxaloacetic acid (Table S9, entry 4 and p. S44†).

To gain insight into whether the carboxylate or enol components of pyruvate are initially phosphorylated, we first submitted two representative model substrates to the reaction conditions. Acetic acid, which features a free carboxylic acid, was phosphorylated to acetyl phosphate **AcP** (1% yield). In contrast, acetylacetone, which exists to a large extent in its enol

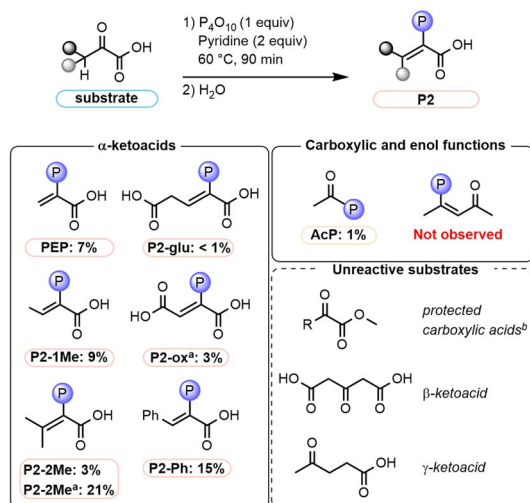


Fig. 5 Substrate scope (Table S9†). ^aThe reaction was run without pyridine. ^bThe methyl or ethyl esters of pyruvate, phenylpyruvate, oxaloacetate, and α -ketoglutarate were tested (for details, see Table S8, entries 11–14 and p. S31†).

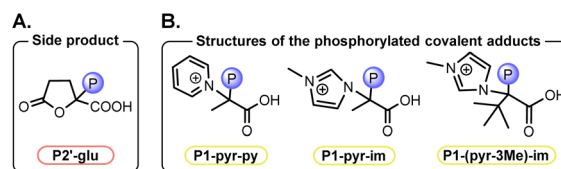


Fig. 6 Structures of intermediates and side products observed in various phosphorylation reactions. (A) Side product arising from α -ketoglutarate. (B) Structures of observed **P1**-type adducts arising from the reactions of pyruvate (**Pyr**) with pyridine (**Py**), **Pyr** with 1-methylimidazole (**Im**), and trimethylpyruvic acid (**Pyr-3Me**) with **Im**.

form (Table S14, entry 4, p. S135 and S136†), did not produce any enol phosphate under the same reaction conditions. We next investigated the importance of the free carboxylic acid function of α -ketoacids for the phosphorylation reaction. When the methyl or ethyl esters of various ketoacids evaluated in Fig. 5 (pyruvate, phenylpyruvate, oxaloacetate, and α -ketoglutarate) were subjected to the reaction conditions, no phosphorylated products were observed, highlighting the importance of a free carboxylic acid group (Table S8, entries 11–14, p. S31†). As the methyl ester of phenylpyruvate predominantly exists in the enol form (Table S14, entry 5, p. S135 and S137†), this result further suggests that the enol is not phosphorylated directly. Moreover, the influence of the distance between the keto and carboxylic acid moieties was investigated. No products were observed with the β -ketoacid 1,3-acetonedicarboxylic acid or with the γ -ketoacid levulinic acid (Table S8, entries 6–8 and p. S31†), hinting at an intramolecular phosphoryl transfer.

To rationalize the differences in reactivity between α -dimethylpyruvic acid (21% yield of **P2-2Me** in base-free conditions) and pyruvic acid (traces of **PEP** in base-free conditions), we compared the reaction mixtures of each substrate after quenching at a short reaction time (3–5 min). Unlike pyruvic acid, even in the presence of pyridine, α -dimethylpyruvic acid did not produce any detectable **P1**-type adducts (Table S8,† entry 2). Instead, the enol phosphate product was immediately observed. The lack of **P1** correlates with the ketoacid's propensity to enolize, with α -dimethylpyruvic acid being more acidic at the α -position.⁵¹ Similarly, phenylpyruvic acid produced **P2-Ph** without any detectable **P1**-type intermediate (Table S10 and p. S45†). The nucleophilic addition of the base on the ketone is, therefore, less pronounced for substrates that are readily enolizable.

We next explored the influence of the base on the formation and the stability of the **P1** adducts. When the reaction was run using **Py**, **P1-pyr-py** was formed immediately (Fig. 6B), and its transformation into **PEP** was almost complete after 1 h (*cf.* Fig. 3C, Table S5 and p. S24†). In contrast, when **Im** was used as base, **P1-pyr-im** was observed in 4% yield (Fig. 6B), but only trace amounts of **PEP** were observed over the same 1 h period (Table S7, entries 1–6 and p. S29†).

Thus, the phosphorylated adducts generated with different bases (*i.e.*, **P1-pyr-py** and **P1-pyr-im**) display different propensities to eliminate, with the formation of **PEP** from **P1-pyr-py** being more favorable than from **P1-pyr-im**. Indeed, increasing



the amount of **Im** used in the reaction (from 2 to 4 equiv.), increased the yield of **P1-pyr-im** and **PEP** to 13% and 1.5%, respectively (Table S7, entry 5 and p. S29[†]), whereas increasing the amount of **Py** (from 2 to 5 to 10) decreased the yields of both compounds (Table S3, entries 7–8, p. S23[†]). Even when a stronger base like triethylamine **TEA** was added to the system 10 min after the formation of **P1-pyr-im**, the elimination of **Im** from **P1-pyr-im** to **PEP** did not occur (Table S7, entry 11). Overall, **Im** appears to be a poorer leaving group than **Py** for subsequent elimination to the enol phosphate and consequently forms a more stable **P1** adduct. Additionally, when the reaction was run with trimethylpyruvic acid **Pyr-3Me**, a sterically hindered nonenolizable pyruvic acid derivative, the formation of a **P1** adduct was only observed when **Im** was used as base (Fig. 6B, **P1-(pyr-3Me)-im**; Table S8, entry 3).

Finally, the reaction of **Pyr** with P_4O_{10} was studied using triethylamine **TEA**, a stronger base. In this case, the formation of the phosphorylated intermediate **P1** was not observed even at the beginning of the reaction, but **PEP** was detected directly (Table S7, entries 15–18). Consequently, the reaction with **TEA** might not occur through the same mechanism as with pyridine and 1-methylimidazole.

Proposed mechanism

Based on the above observations, a simplified reaction scheme was constructed, which involves two possible pathways for the phosphorylation of α -ketoacids (Fig. 7). Pathway I involves the phosphorylation of the carboxy group and the addition of the base to the ketone to generate the acyl phosphate intermediate **Int. A** (step I₁), which features a free hydroxy group. Subsequent intramolecular phosphoryl transfer to the hydroxy group generates **P1** (step I₂). **P2** is generated by elimination of the base, as seen in the case of pyridine (step I₃). The formation of side product **P3-pyr** can result from the attack of the enol tautomer of pyruvate on the acyl phosphate intermediate (e.g., **Int. A**). Pathway II involves phosphorylation of the carboxy group to generate the acyl-phosphate intermediate **Int. B** (step II₁), which features a free enol group.

Subsequently, intramolecular phosphoryl transfer to the enol group generates **P2** (step II₂). This pathway might be operating in cases where the enol phosphate is formed in the absence of a base. It also seems plausible for α -ketoacids whose enolization is facile, or for reactions run with bases that are

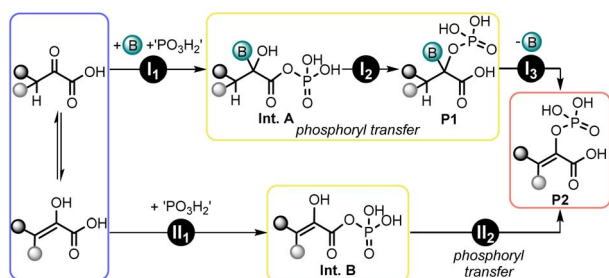


Fig. 7 Simplified reaction scheme for the phosphorylation of α -ketoacids to products **P2**.

unable to add to the keto group (e.g., **TEA**). The possibility of producing enol phosphates through two different pathways can explain why higher yields are observed for readily enolizable substrates. To verify whether ketoacids with higher enol content are more rapidly phosphorylated, a competition experiment was performed between pyruvic acid and phenylpyruvic acid (Table S8, entry 5 and p. S30[†]), the latter being more easily enolizable.^{52,53} Starting from a 1 : 1 mixture of the two substrates in the presence of pyridine, a product ratio of 20 : 1 was observed in favor of the enol-phosphate derived from phenylpyruvic acid when the reaction was quenched after 3–5 min. So far, the acyl phosphate intermediate (**Int. A** or **Int. B**) has been assumed to be formed *in situ*, but no direct proof for their formation could be obtained.

Phosphoryl transfer mechanism – alternative synthesis of **PEP** in CD_3CN

To investigate the proposed intramolecular phosphoryl transfer mechanism, we set out to study conditions favoring the formation of the acyl phosphate of pyruvate (**AcP-Pyr**). To this end, pyruvoyl chloride (**Pyr-Cl**), featuring an activated carbonyl group, was used instead of **Pyr** with the corresponding phosphate nucleophile H_3PO_4 . Importantly, in the presence of **Py**, **PEP**, **P1-pyr-py**, and **P3-pyr** were all observed after 5 min at 60 °C in neat conditions, indicating the similar reactivity of our model system with the standard conditions using **Pyr** (Table S11 and p. S48[†]). Next, we investigated the reactivity of **AcP-Pyr** in homogenous anhydrous conditions. Tetrabutylammonium dihydrogen phosphate (**TBAP**) was used instead of H_3PO_4 to avoid water contamination, and **TEA** was preferred over **Py** to avoid precipitation (Table S12 and p. S52[†]).

When **TEA** (0.12 mmol) and **TBAP** (0.06 mmol) were pre-mixed in CD_3CN and **Pyr-Cl** (0.06 mmol) was added to the mixture, 3% yield of **PEP** was obtained *via* its cyclic form **cPEP** (Fig. 8, Pathway A₀, ESI, Section V.F.5 and p. S58[†]) along with 1% of **P3-pyr**. To identify the formation of the acyl phosphate

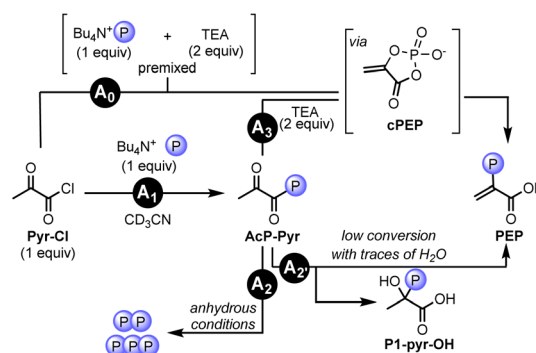


Fig. 8 Study of the phosphoryl transfer in aprotic solvent (CD_3CN). Reaction A₀: reaction of premixed base and phosphate with **Pyr-Cl** leading to the formation of **PEP** through its cyclic form **cPEP**. Reaction A₁: the activated carboxylic acid **Pyr-Cl** is transformed to the corresponding acyl-phosphate (**AcP-Pyr**), leading to the formation of polyphosphates in anhydrous conditions (A₂) or to the formation of **PEP** by either subsequent addition of **TEA** (A₃) or with trace amount of H_2O (along with the formation of the side product **P1-pyr-OH** (A_{2'})).



intermediate, stepwise experiments were performed. When mixing **TBAP** and **Pyr-Cl** in CD_3CN , the formation of **AcP-Pyr** was detected by ^1H and ^{31}P NMR along with the formation of pyrophosphate (Fig. 8, Pathway A1; Table S12, entry 5 and p. S52†). In the absence of a base, **AcP-Pyr** was converted to pyrophosphate and triphosphate in roughly 30 min without formation of **PEP** (Fig. 8, Pathway A2, Table S12, entry 5 and p. S54–S55†). When trace amounts of H_2O were present in the mixture, **AcP-Pyr** underwent intramolecular phosphoryl transfer to form the corresponding hydroxyphosphate **P1-pyr-OH** (Fig. 8, Pathway A2', ESI, Section V.F.4 and p. S57–S58†), accompanied by traces of **PEP**. When the base **TEA** (2 equiv.) was added after a reaction time of 5 min, trace amounts (<1%) of **PEP** along with multiple polyphosphate species were observed (Fig. 8, Pathway A3; Table S12, entry 6 and p. S56†). The low yield in **PEP** can be explained by the formation of the **P1-pyr-OH** side product prior to the addition of base. These experiments show that the formation of **PEP** from **AcP-Pyr** (through enolization and intramolecular phosphoryl transfer) requires the presence of a base and is in competition with the formation of polyphosphates and the side product **P1-pyr-OH** (i.e., by a nucleophilic attack of a phosphate or a water molecule on the activated carboxylic acid or on the carbonyl group of **AcP-Pyr**, respectively).

DFT study of the proposed pathways

DFT calculations were performed to evaluate some key aspects of the reaction mechanism. First, we examined the ability of different sites within pyruvate to undergo direct phosphorylation by phosphoryl pyridine (Fig. 9A). *N*-Phosphoryl pyridine was chosen as the model phosphorylating agent based on our NMR experiments (cf. Fig. 4) and the recent studies of Cummins.⁴⁶ As shown in Fig. 9A, the computed barrier for the phosphorylation of the carboxylic acid moiety is approx. 60 kJ mol^{-1} lower than the barrier for the direct

phosphorylation of the enol, in line with our experimental observations. Next, we computed the thermochemistry of the overall phosphorylation sequence (Fig. S7 and p. S146†) by analyzing the intermediates identified in aqueous solution (Fig. 9B, **P1-pyr-im** and **P1-pyr-py**), along with several possible intermediates (Fig. 9B, **Int. A–C** and **P1-pyr-OH**). As expected, without an activated phosphorylating agent, direct phosphorylation of pyruvic acid to **AcP-Pyr** with phosphoric acid is endergonic by 42.9 kJ mol^{-1} (ESI, Fig. S7 and p. S146†). From **AcP-Pyr**, the intramolecular phosphoryl transfer was studied by considering three main pathways by going through nucleophilic addition of the base (**Int. A**), enolization (**Int. B**), and addition of H_2O (**Int. C**, Fig. 9B). The latter being unlikely to occur in our paste conditions due to the presence of P_4O_{10} , which also acts as a dehydrating agent. Nucleophilic addition of 1-methylimidazole (**Im**) to the **AcP-Pyr** ketone was calculated to be favorable by 10.0 kJ mol^{-1} (**Int. A1**), whereas the addition of pyridine (**Py**) or water is slightly disfavored by 15.1 (**Int. A2**) and 7.5 kJ mol^{-1} (**Int. C**), respectively. However, the subsequent intramolecular phosphoryl transfers from the imidazolium hemiaminal (**Int. A1**), pyridinium hemiaminal (**Int. A2**), enol (**Int. B**), or hydrate (**Int. C**) intermediates derived from **AcP-Pyr** were all calculated to be exergonic by 16.7 , 9.4 , 14.6 , and 28.6 kJ mol^{-1} , respectively. In line with the experimental observation of intermediates **P1**, the nucleophile addition is favorable for **Im**, and subsequent intramolecular phosphoryl transfers from the carboxylate to the enol or alcohol are thermochemically favorable. Interestingly, a pentacoordinate P intermediate (cf. Fig. 2, **cPEP·H₂O**) was not found to be a stable minimum but rather corresponds to the transition state of phosphoryl transfer. However, the computed barrier for the concerted transfer is relatively high, with ≈ 129 – 146 kJ mol^{-1} using explicit molecules of water as a model base for the deprotonation of the enolate and a hydronium ion as a model acid for the protonation of the carboxylate. As the employed

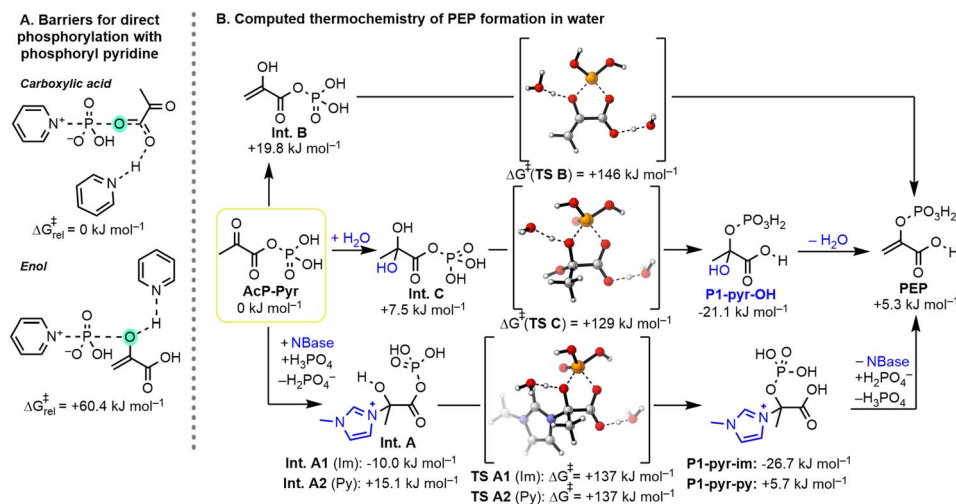


Fig. 9 (A) Computational study comparing the phosphorylation of pyruvate at its carboxylate vs. its enol. (B) Computational study showing the thermochemistry of key steps and the barriers for intramolecular phosphoryl transfer. Calculations were performed at the SMD(H_2O)/M06-2X/def2-TZVPD//SMD(H_2O)/M06-2X/def2-SVP level of theory.



not contain sulfur, thiol-based leaving groups in metabolism, such as CoA, generally contain phosphoryl groups. Fourth, in water, the formation of acyl phosphates from carboxylates is arguably chemically simpler than the conversion of carboxylates to thioesters – the former requires only a single component (an activated phosphate), whereas the latter requires two (a thiol and an activating agent for the carboxylate). All these points suggest that an early stage of protometabolism was more likely an “Acyl Phosphate World” than a “Thioester World”. Consequently, nonenzymatic carboxylate phosphorylation should be a target of the highest priority for prebiotic chemistry using life as a guide.⁵

Data availability

Everything is in the ESI.†

Author contributions

JZ performed the experiments and wrote the first draft of the paper. RJM performed the DFT calculations. JM conceptualized the project and acquired funding. All authors edited the paper.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This work was supported by the Interdisciplinary Thematic Institute ITI-CSC via the IdEx Unistra (ANR-10-IDEX-0002) within the program Investissement d'Avenir. J. Z. thanks the French government for an MRT fellowship. R. J. M. thanks the Deutsche Forschungsgemeinschaft (DFG, German Research Foundation) for a fellowship (MA 9687/1-1). This project has received funding from the European Research Council (ERC) under the European Union's Horizon 2020 research and innovation program (grant agreement no. 101001752). J. M. thanks the VW Foundation (no. 96_742) for generous support. Computations were performed at the High-Performance Computing Center of the University of Strasbourg. The authors would like to particularly thank Dr Jesus Raya for his help and expertise in solid-state NMR analyses. Dr Maurice Coppe, and Dr Bruno Vincent are thanked for assistance with NMR analyses. Wahnyalo Kazöne is thanked for MS analyses. Dr Quentin Dherbassy and Emilie Werner are thanked for helpful discussions.

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