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Photo-isomerization of the isolated photoactive yellow protein chromophore: what comes before the primary step?[†]

 Cate S. Anstöter,[‡] Basile F. E. Curchod[‡] and Jan R. R. Verlet^{‡*}

Photoactive proteins typically rely on structural changes in a small chromophore to initiate a biological response. While these changes often involve isomerization as the “primary step”, preceding this is an ultrafast relaxation of the molecular framework caused by the sudden change in electronic structure upon photoexcitation. Here, we capture this motion for an isolated model chromophore of the photoactive yellow protein using time-resolved photoelectron imaging. It occurs in <150 fs and is apparent from a spectral shift of ~70 meV and a change in photoelectron anisotropy. Electronic structure calculations enable the quantitative assignment of the geometric and electronic structure changes to a planar intermediate from which the primary step can then proceed.

Photoactive proteins are common in nature and are necessary for various biological functions including vision, phototaxis, signaling, and photoprotection.^{1–3} The absorption of a photon occurs by a relatively small chromophore that is embedded in the protein and typically leads to a local structural change in the chromophore which subsequently triggers a response in the protein. Building a detailed understanding of the dynamics of the primary photophysical changes occurring in these chromophores has been a major goal of ultrafast photobiology, both experimentally and computationally. Gas-phase spectroscopy is particularly useful as it probes the intrinsic dynamics of the chromophore and is amenable to high level computational methods.⁴ As the transduction of light to mechanical motion typically involves isomerization of the chromophore, interest has focused on this process, which has been described as the “primary” or “first” step in the photocycle.^{5–9} However, there are important dynamical processes that precede this initial step.¹⁰ Specifically, photon absorption *via* a $\pi^* \leftarrow \pi$ transition is expected to result in dynamical changes in structure

associated with an alteration of the C–C bonds and, indeed, this motion is necessary for the subsequent isomerization. However, the nuclear and electronic dynamics of this initial impulsive motion have yet to be clearly observed in isolated systems. Here, we resolve these dynamics using time-resolved photoelectron imaging and computational chemistry.

One particular class of photoactive proteins contain chromophores based on *para*-substituted phenolate anions. This includes both the green fluorescent protein (GFP) and the photoactive yellow protein (PYP). PYP in particular has served as an important test-case for new experimental methods.^{11–17} The chromophore in PYP is a deprotonated *para*-coumaric thioester, which links to the protein *via* the thioester group.^{18,19} A common model system for the chromophore is the deprotonated *para*-coumaric ketone (*pCK*[−], Fig. 1). This anion has previously been studied by time-resolved photoelectron imaging,¹⁵ where the isomerization dynamics could be clearly identified through changes in the photoelectron kinetic energy (eKE) distribution as well as through the photoelectron angular distributions (PADs). Combined, these changes are sensitive to ultrafast evolution of nuclear and electronic structure. Here, we focus on the dynamics taking place *before* the isomerization in *pCK*[−], that are induced by the $\pi^* \leftarrow \pi$ transition as shown in Fig. 1.

We have recorded the time-resolved photoelectron images of mass-selected gas-phase *pCK*[−] excited at 2.79 eV (444 nm) and probed at 1.55 eV (800 nm).^{15,20,21} See ESI[†] for the full

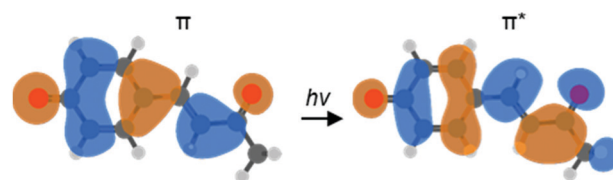


Fig. 1 Computed molecular orbitals (MOs) involved in the vertical $\pi\pi^*$ transition of *pCK*[−] that leads to subsequent isomerization on the *S*₁ excited state (LR-TDDFT/TDA/ ω B97X-D/aug-cc-pVDZ).

Department of Chemistry, Durham University, Durham DH1 3LE, UK.

E-mail: j.r.r.verlet@durham.ac.uk

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[‡] Present address: Department of Chemistry, Temple University, Philadelphia, 19122, USA.

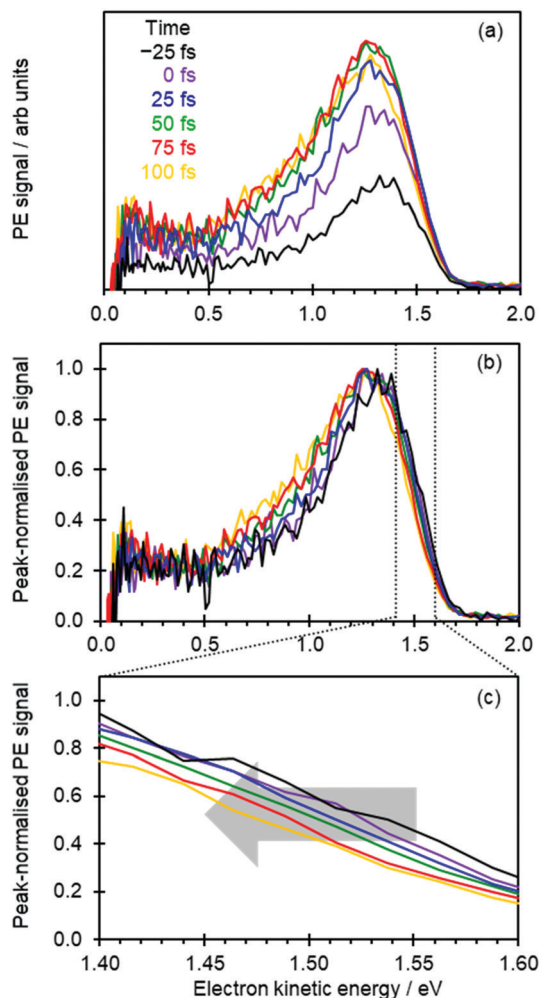


Fig. 2 (a) Time-resolved photoelectron spectra of pCK^- following excitation at 2.79 eV and probed at 1.55 eV. (b) Same data as in (a) but with each spectrum normalized to its maximum intensity. (c) Same data as in (b) but focusing on the high kinetic energy edge of the spectra, showing the spectral shift over the first 100 fs.

experimental details. The pump was chosen to coincide with the $S_1 \leftarrow S_0$ transition and to be just below the detachment continuum (electron affinity is 2.87 ± 0.05 eV).^{15,22} The spectral resolution of the experiment is $\sim 5\%$ of the eKE. The cross-correlation between pump and probe pulses is 100 ± 10 fs (full width at half maximum). In terms of measuring exponential decays, the experiment offers a time-resolution of ~ 50 fs, but this can be significantly better when probing spectral shifts.²³ Conversely, to measure small spectral shifts requires appropriate spectral resolution. The current experiments offer a balance between the timescale of the dynamics and the required spectral resolution of any changes. Here we focus on the dynamics occurring within the first 200 fs.

Fig. 2(a) shows the time-resolved photoelectron spectra, in which photoelectron spectra with $t \ll 0$ have been subtracted to leave only the transient signals. The total photoelectron signal at high eKE rises rapidly and corresponds to the temporal resolution of the experiment. The positive peak with a

maximum around eKE ~ 1.3 eV is a measure of the S_1 excited state population that has been projected onto the neutral ground state (D_0) continuum.^{8,15} For $t > 100$ fs, the photoelectron peak significantly broadens with signal peaking around eKE ~ 0.7 eV rising: this spectral change corresponds to isomerization on the S_1 excited state.^{8,15}

To capture the dynamics preceding isomerization, we focus here on the spectra shown in Fig. 2(a). The changing intensities in Fig. 2(a) makes it difficult to appreciate any spectral changes occurring on the sub-100 fs timeframe. Therefore, in Fig. 2(b) the same time-resolved photoelectron spectra are presented, where each spectrum has been normalized to its maximum intensity.

At first sight, Fig. 2(b) suggests that there is little change in the photoelectron spectra over the first 100 fs. However, upon closer inspection, there is a clear shift towards lower eKE as time evolves. This shift is noticeable at both the high-eKE and low-eKE edge of the peak. We focus here on the former as the low-eKE side will involve some motions related to initial isomerisation dynamics which we seek to avoid in our consideration here and was considered in detail elsewhere.¹⁵ The high-eKE edge of the pump-probe signal is shown more clearly in Fig. 2(c), where we have expanded the spectral region between $1.40 < \text{eKE} < 1.60$ eV. The rate of the spectral shift can be roughly determined by taking the eKE at the half maximum, $\text{eKE}_{1/2\text{max}}$, which we have done by fitting the rising edge of the signal (roughly between intensities of 0.25 and 0.75) to a linear function. The R^2 for all these fits is ≥ 0.99 except for the point at $t = -25$ fs, for which $R^2 = 0.98$ due to the lower signal intensity (see Fig. 2(a)). The $\text{eKE}_{1/2\text{max}}$ determined in this manner is shown in Fig. 3(a) as a function of time. By 150 fs the spectral shift is complete. Fig. 3(a) also suggests that the shift is roughly linear. A linear fit to the first 125 fs shows that the rate

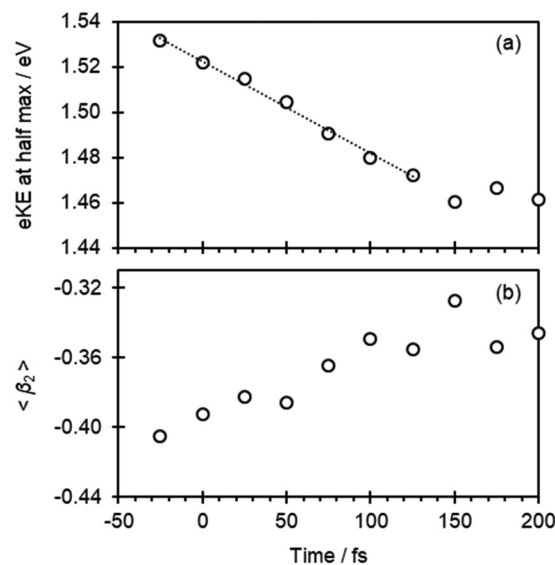


Fig. 3 (a) The electron kinetic energy at half maximum of the high energy edge determined from a linear fit, plotted as a function of pump-probe delay. The dashed line is a linear fit to these data. (b) The weighted average of β_2 as a function of pump-probe delay.

of the eKE shift amounts to $\sim 0.4 \text{ meV fs}^{-1}$, whereas the total shift, $\Delta e\text{KE} = 69 \pm 10 \text{ meV}$.

In addition to the photoelectron spectra, we have also analyzed the PADs. These are sensitive to the orbital from which the electron is detached (π^* in Fig. 1) and therefore a measure of the electronic structure.^{24,25} The PADs are generally quantified by an anisotropy parameter, β_2 , which range from +2 to -1 for photoelectrons leaving predominantly parallel and perpendicular to the probe light polarization axis, respectively.^{24–26} In Fig. 3(b), the weighted average of β_2 in the $1.30 < e\text{KE} < 1.60 \text{ eV}$ range, $\langle \beta_2 \rangle$, is plotted as a function of pump-probe delay. This is seen to change from $\langle \beta_2 \rangle$ about -0.41 to -0.35 in the first 100 fs.

To relate experimental observations to changes in $p\text{CK}^-$, we have computed critical points on the S_1 potential energy surface that are expected to be important in the early dynamics. Specifically, we focused on the S_0 minimum (DFT/ $\omega\text{B97X-D/aug-cc-pVDZ}$), which corresponds to the Franck–Condon geometry, FC, and the S_1 planar minimum, $S_{1,\text{PM}}$, (LR-TDDFT/TDA/ $\omega\text{B97X-D/aug-cc-pVDZ}$). The vertical excitation energies of the S_0 , S_1 and D_0 states were then refined using multi-state extended multi-configurational quasi-degenerate perturbation theory (MS-XMCQDPT2).²⁷ An (aug)-cc-pVTZ basis set was used, in which the augmented function was affixed to the oxygen atoms only.²⁸ Detachment to the neutral (D_0) plus free electron continuum was mimicked through addition of an extremely diffuse p-function ($\alpha = 10^{-10}$).^{29–31} Finally, we calculated the Dyson orbitals using EOM-EE/IP-CCSD/6-31+G** and the PADs were modelled using ezDyson v4.^{32,33} See ESI† for the full computational details.

The results from the calculations are shown in Table 1. The time-resolved photoelectron spectra arise from differences in energies between the S_1 state and the final D_0 neutral ground state. The energy difference on the D_0 state between the geometries at the FC and $S_{1,\text{PM}}$ is only 3.4 meV. Hence, changes in the observed eKE will predominantly reflect changes in the S_1 energy. The energy difference between the FC and $S_{1,\text{PM}}$ geometries on the S_1 state is 77.6 meV. Taken together with the changes in D_0 , the calculations predict that $\Delta e\text{KE} = 74.2 \text{ meV}$, in excellent agreement with the observed overall shift determined from Fig. 3 ($69 \pm 10 \text{ meV}$). We can therefore have confidence that the computed structures are representative of the dynamics observed experimentally.

Table 1 Calculated vertical energies of electronic states of the $p\text{CK}^-$ anion and neutral states for the Franck–Condon (FC) and S_1 planar minimum ($S_{1,\text{PM}}$) geometries^a

Geometry	$E(S_0)$	$E(S_1)$	$E(D_0)$	$\Delta(S_1-D_0)$	β_2
FC	0.000	2.790	2.940	0.150	-0.58
$S_{1,\text{PM}}$	0.050	2.713	2.937	0.224	-0.49

^a All energies are in eV and are relative to the anion ground state. The D_0 energies have been shifted by -0.340 eV to coincide with the experimental adiabatic energy.¹⁵ The computed β_2 are obtained by averaging over the same range as used in the experiment.

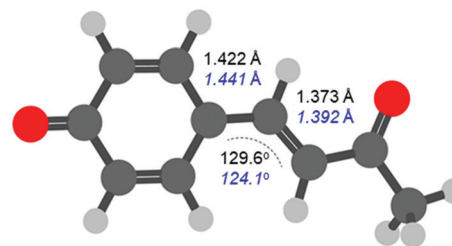


Fig. 4 Calculated structure of $p\text{CK}^-$. The distances and angles between the three bridging carbon atoms connecting the phenyl ring and ketone are indicated for the FC structure (black) and the $S_{1,\text{PM}}$ (blue, italics), obtained at the DFT/ $\omega\text{B97X-D/aug-cc-pVDZ}$ and LR-TDDFT/TDA/ $\omega\text{B97X-D/aug-cc-pVDZ}$ levels of theory, respectively.

The FC structure is shown in Fig. 4 where the dominant geometric changes between FC and $S_{1,\text{PM}}$ are indicated. As expected, following the $\pi^* \leftarrow \pi$ transition shown in Fig. 1, the C–C bond distances increase along the 3 bridging C atoms connecting the phenyl ring and ketone. The structural changes are small with an elongation of $\sim 2 \text{ pm}$ ($\sim 1.3\%$ increase). Additionally, there is also a change in the C–C=C bond angle, which decreases from 129.6° to 124.1° (change of $\sim 4\%$). Other bonds also change but to a significantly smaller extent and our results are in general agreement with previous studies.³⁴ Despite the very small changes, the photoelectron spectra can clearly track these structural changes that precede the primary isomerization event.

The measured $\langle \beta_2 \rangle$ also show changes in the first 100 fs. A negative $\langle \beta_2 \rangle$ is generally associated with electron emission from a π orbital,²⁵ in agreement with expectation from Fig. 1. The computed β_2 averaged over the same range as in the experiment is shown in Table 1. The shift in anisotropy is reproduced in the computationally modelled PADs, with β_2 becoming less negative by 0.09 in progressing from FC to $S_{1,\text{PM}}$, compared to a shift of 0.06 observed experimentally. The absolute computed β_2 are slightly more negative than experiment, but this mostly reflects the fact that the computed structures are considered at 0 K, while the experimental ion temperature is $\sim 300 \text{ K}$.³⁵ Overall, the computed electronic structure is consistent with that measured experimentally and demonstrates the remarkable sensitivity of β_2 to the small changes in electronic structure (see ESI† for Dyson orbitals).

The extent of the changes is perhaps smaller than one might anticipate. However, most of the orbital (and structural) changes take place over the 3 C atoms shown in Fig. 4. Within a simple Hückel picture,^{29,36,37} these 3 C atoms can be approximated as the allyl radical. Within this picture, even though the transition is formally of $\pi^* \leftarrow \pi$ character, the highest occupied molecular orbital (HOMO) can be viewed as the non-bonding allyl orbital so that the transition character can be considered to be of $\pi^* \leftarrow \pi$ character, with smaller associated structural changes.

The data in Fig. 2 also suggest that the nuclear wavepacket is relatively well-localized over the first 100 fs: a rapidly spreading wavepacket would lead to broadening in the photoelectron spectrum. Instead, the gradient of the high-eKE

edge remains essentially unchanged as the dynamics proceed (the broadening at the low-eKE edge involves initial rotation about the single-bond at the *para*-position on the phenolate ring¹⁵). Martínez and coworkers³⁸ have previously computed the excited state molecular dynamics of isolated *p*CK⁻ in the S₁ state and showed that, although the dominant motion following excitation is isomerization about the single bond, several trajectories exhibited a delay before this motion was activated. These trajectories therefore appear to correlate well with the motion observed here connecting FC to S_{1,PM}. However, several computed trajectories also led to immediate isomerization. Our experiments point to a more sequential process, where the genuine primary step can be resolved from the isomerization step in *p*CK⁻, although there is evidence that a fraction undergoes more direct isomerization based on the spread in the low-eKE edge. Dynamics on a few 100 fs timescale have also been observed in aqueous chromophore and in the PYP protein using Ultraviolet Resonance Femtosecond Stimulated Raman Spectroscopy. For *para*-coumaric acid, a deformation was observed similar to the current work where the chromophore remains planar.³⁹ In the protein, the early dynamics lead to a weakening of the hydrogen-bonding at the phenolate.⁴⁰ The dynamics may of course be different in other biochromophores. For the retinal chromophore, for example, the initially excited skeletal vibrations coexist with the isomerization dynamics⁴¹ and the two motions may therefore not be as separable as in *p*CK⁻. Nevertheless, with the right experimental conditions, the step before the primary isomerization step in other isolated bio-chromophores may be observable. This genuine first step and how strongly it is coupled to the isomerization coordinate is likely to be critically important as a driver for the isomerization process and therefore may offer a new handle to control the overall process.

Conclusions

In conclusion, we have captured the dynamics preceding the “primary” motion in the PYP chromophore. The structural changes on the S₁ state immediately following photoexcitation – both geometric and electronic – have been resolved using experiment and computation. The $\pi^* \leftarrow \pi$ transition launches a nuclear wavepacket that leads to small changes between the 3 C atoms at the *para*-position of the phenolate. These changes can be viewed as the “zeroth” step that enable the subsequent isomerization of the chromophore commonly regarded as the “first” step in a photoreceptor bio-cycle.

Conflicts of interest

There are no conflicts to declare.

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References

- M. A. van der Horst and K. J. Hellingwerf, *Acc. Chem. Res.*, 2004, **37**, 13–20.
- C. Dugave and L. Demange, *Chem. Rev.*, 2003, **103**, 2475–2532.
- E. A. Rodriguez, R. E. Campbell, J. Y. Lin, M. Z. Lin, A. Miyawaki, A. E. Palmer, X. Shu, J. Zhang and R. Y. Tsien, *Trends Biochem. Sci.*, 2017, **42**, 111–129.
- Photophysics of Ionic Biochromophores*, S. B. Nielsen and J. A. Wyer, ed., Springer-Verlag, Berlin Heidelberg, 2013.
- R. W. Schoenlein, L. A. Peteanu, R. A. Mathies and C. V. Shank, *Science*, 1991, **254**, 412–415.
- R. Kort, H. Vonk, X. Xu, W. D. Hoff, W. Crielaard and K. J. Hellingwerf, *FEBS Lett.*, 1996, **382**, 73–78.
- P. Kukura, D. W. McCamant, S. Yoon, D. B. Wandschneider and R. A. Mathies, *Science*, 2005, **310**, 1006–1009.
- I.-R. Lee, W. Lee and A. H. Zewail, *Proc. Natl. Acad. Sci. U. S. A.*, 2006, **103**, 258–262.
- D. Polli, P. Altoe, O. Weingart, K. M. Spillane, C. Manzoni, D. Brida, G. Tomasello, G. Orlandi, P. Kukura, R. A. Mathies, M. Garavelli and G. Cerullo, *Nature*, 2010, **467**, 440–U88.
- M. Ben-Nun and T. J. Martinez, *Chem. Phys.*, 2000, **259**, 237–248.
- F. Schotte, H. S. Cho, V. R. I. Kaila, H. Kamikubo, N. Dashdorj, E. R. Henry, T. J. Graber, R. Henning, M. Wulff, G. Hummer, M. Kataoka and P. A. Anfinrud, *Proc. Natl. Acad. Sci. U. S. A.*, 2012, **109**, 19256–19261.
- Y. O. Jung, J. H. Lee, J. Kim, M. Schmidt, K. Moffat, V. Šrajcar and H. Ihee, *Nat. Chem.*, 2013, **5**, 212–220.
- J. Tenboer, S. Basu, N. Zatssepina, K. Pande, D. Milathianaki, M. Frank, M. Hunter, S. Boutet, G. J. Williams, J. E. Koglin, D. Oberthuer, M. Heymann, C. Kupitz, C. Conrad, J. Coe, S. Roy-Chowdhury, U. Weierstall, D. James, D. Wang, T. Grant, A. Barty, O. Yefanov, J. Scales, C. Gati, C. Seuring, V. Šrajcar, R. Henning, P. Schwander, R. Fromme, A. Ourmazd, K. Moffat, J. J. V. Thor, J. C. H. Spence, P. Fromme, H. N. Chapman and M. Schmidt, *Science*, 2014, **346**, 1242–1246.
- K. Pande, C. D. M. Hutchison, G. Groenhof, A. Aquila, J. S. Robinson, J. Tenboer, S. Basu, S. Boutet, D. P. DePonte, M. Liang, T. A. White, N. A. Zatssepina, O. Yefanov, D. Morozov, D. Oberthuer, C. Gati, G. Subramanian, D. James, Y. Zhao, J. Koralek, J. Brayshaw, C. Kupitz, C. Conrad, S. Roy-Chowdhury, J. D. Coe, M. Metz, P. L. Xavier, T. D. Grant, J. E. Koglin, G. Ketawala, R. Fromme, V. Šrajcar, R. Henning, J. C. H. Spence, A. Ourmazd, P. Schwander, U. Weierstall, M. Frank, P. Fromme, A. Barty, H. N. Chapman, K. Moffat, J. J. van Thor and M. Schmidt, *Science*, 2016, **352**, 725–729.
- C. S. Anstöter, B. F. E. Curchod and J. R. R. Verlet, *Nat. Commun.*, 2020, **11**, 2827.
- H. Kuramochi, S. Takeuchi, H. Kamikubo, M. Kataoka and T. Tahara, *Sci. Adv.*, 2019, **5**, eaau4490.
- J. N. Bull, C. S. Anstöter and J. R. R. Verlet, *Nat. Commun.*, 2019, **10**, 1–9.
- W. D. Hoff, P. Dux, K. Hard, B. Devreese, I. M. Nugteren-Roodzant, W. Crielaard, R. Boelens, R. Kaptein, J. Van Beeumen and K. J. Hellingwerf, *Biochemistry*, 1994, **33**, 13959–13962.

- 19 E. D. Getzoff, K. N. Gutwin and U. K. Genick, *Nat. Struct. Mol. Biol.*, 2003, **10**, 663–668.
- 20 J. Lecointre, G. M. Roberts, D. A. Horke and J. R. R. Verlet, *J. Phys. Chem. A*, 2010, **114**, 11216–11224.
- 21 L. H. Stanley, C. S. Anstöter and J. R. R. Verlet, *Chem. Sci.*, 2017, **8**, 3054–3061.
- 22 T. Rocha-Rinza, O. Christiansen, J. Rajput, A. Gopalan, D. B. Rahbek, L. H. Andersen, A. V. Bochenkova, A. A. Granovsky, K. B. Bravaya, A. V. Nemukhin, K. L. Christiansen and M. B. Nielsen, *J. Phys. Chem. A*, 2009, **113**, 9442–9449.
- 23 D. A. Horke, Q. Li, L. Blancafort and J. R. R. Verlet, *Nat. Chem.*, 2013, **5**, 711–717.
- 24 K. L. Reid, *Annu. Rev. Phys. Chem.*, 2003, **54**, 397–424.
- 25 A. Sanov, *Annu. Rev. Phys. Chem.*, 2014, **65**, 341–363.
- 26 J. Cooper and R. N. Zare, *J. Chem. Phys.*, 1968, **48**, 942–943.
- 27 A. A. Granovsky, *J. Chem. Phys.*, 2011, **134**, 214113.
- 28 T. H. Dunning, *J. Chem. Phys.*, 1989, **90**, 1007–1023.
- 29 A. V. Bochenkova, B. Klærke, D. B. Rahbek, J. Rajput, Y. Toker and L. H. Andersen, *Angew. Chem., Int. Ed.*, 2014, **53**, 9797–9801.
- 30 A. V. Bochenkova, C. R. S. Mooney, M. A. Parkes, J. L. Woodhouse, L. Zhang, R. Lewin, J. M. Ward, H. C. Hailes, L. H. Andersen and H. H. Fielding, *Chem. Sci.*, 2017, **8**, 3154–3163.
- 31 C. S. Anstöter, T. E. Gartmann, L. H. Stanley, A. V. Bochenkova and J. R. R. Verlet, *Phys. Chem. Chem. Phys.*, 2018, **20**, 24019–24026.
- 32 C. M. Oana and A. I. Krylov, *J. Chem. Phys.*, 2009, **131**, 124114.
- 33 S. Gozem and A. I. Krylov, *WIREs Comput. Mol. Sci.*, 2021, e1546.
- 34 M. Boggio-Pasqua and G. Groenhof, *Comput. Theor. Chem.*, 2014, **1040–1041**, 6–13.
- 35 C. S. Anstöter and J. R. R. Verlet, *J. Phys. Chem. A*, 2021, **125**, 4888–4895.
- 36 K. B. Bravaya, B. L. Grigorenko, A. V. Nemukhin and A. I. Krylov, *Acc. Chem. Res.*, 2012, **45**, 265–275.
- 37 C. S. Anstöter, C. R. Dean and J. R. R. Verlet, *Phys. Chem. Chem. Phys.*, 2017, **19**, 29772–29779.
- 38 A. M. Virshup, J. Chen and T. J. Martínez, *J. Chem. Phys.*, 2012, **137**, 22A519.
- 39 H. Kuramochi, S. Takeuchi and T. Tahara, *J. Phys. Chem. Lett.*, 2012, **3**, 2025–2029.
- 40 H. Kuramochi, S. Takeuchi, K. Yonezawa, H. Kamikubo, M. Kataoka and T. Tahara, *Nat. Chem.*, 2017, **9**, 660–666.
- 41 P. J. M. Johnson, M. H. Farag, A. Halpin, T. Morizumi, V. I. Prokhorenko, J. Knoester, T. L. C. Jansen, O. P. Ernst and R. J. D. Miller, The Primary Photochemistry of Vision Occurs at the Molecular Speed Limit, <http://pubs.acs.org/doi/full/10.1021/acs.jpcc.7b02329>, (accessed 29 September 2021).