



 Cite this: *RSC Adv.*, 2020, 10, 37168

Network pharmacology-based study of the mechanisms of action of anti-diabetic triterpenoids from *Cyclocarya paliurus*†

 Zixin Lin,^{ab} Yingpeng Tong,^{bc} Na Li,^b Ziping Zhu^b and Junmin Li *^{bc}

Diabetes is a complex illness requiring long-term therapy. *Cyclocarya paliurus*, a recently confirmed new food resource, shows significant hypoglycemic and hypolipidemic effects in type II diabetes. Triterpenoid saponins are considered as the effective medicinal components of *C. paliurus* and are useful for the treatment of diabetes mellitus. However, little is known regarding their specific mechanism of actions. In this study, we used active ingredient screening and target prediction techniques to determine the components of *C. paliurus* responsible for its anti-diabetic effects as well as their targets. In addition, we used bioinformatics technology and molecular docking analysis to determine the mechanisms underlying their anti-diabetic effects. A total of 39 triterpenes were identified through a literature search and 1 triterpene compound by experiments. In all, 33 potential target proteins associated with 36 pathways were predicted to be related to diabetes. Finally, 7 compounds, 15 target proteins, and 15 signaling pathways were found to play important roles in the therapeutic effects of *C. paliurus* against diabetes. These results provide a theoretical framework for the use of *C. paliurus* against diabetes. Moreover, molecular docking verification showed that more than 90% of the active ingredients had binding activity when tested against key target proteins, and a literature search showed that the active ingredients identified had anti-diabetic effects, indicating that the results were highly reliable.

 Received 8th August 2020
 Accepted 24th September 2020

DOI: 10.1039/d0ra06846b

rsc.li/rsc-advances

1. Introduction

Diabetes is a disorder of glucose metabolism that leads to high blood sugar levels and to a series of complications which reduce the quality of life of patients, increasing their mortality.¹ Based on its pathogenesis, it can be divided into type 1 diabetes (T1DM) and type 2 diabetes (T2DM), with T2DM accounting for more than 90% of cases.² At present, diabetes is treated with drugs, such as sulfonylureas, DPP-4 inhibitors, oral antidiabetic agents, sodium-glucose co-transport inhibitors, and glucokinase activators,^{3–7} but many drugs have side effects.⁸ Therefore, it is urgent to find new, safe and effective early intervention medicinal compounds for diabetes.⁹

Traditional botanical and herbal medicines have been widely used for the treatment of diabetes,^{10,11} e.g. *Lobelia chinensis* Lour.,⁷ *Hydrangea macrophylla* var. *thunbergii*,¹² and *Cyclocarya paliurus* (Batal.) Iljinskaja.¹³ *C. paliurus* is a medicinal plant native to southern China. Its leaves have been traditionally used in the form of herbal tea for their beneficial effects against

diabetes. Various bioactive compounds have been extracted from the leaves of *C. paliurus*, including flavonoids, triterpenoids, polysaccharides, and organic acids, which may contribute to its antihyperglycemic, antihyperlipidemic, and antihypertensive effects.^{13–24} Although the mechanisms underlying the anti-diabetic effects of the ethanol and aqueous extracts of *C. paliurus* leaves have been explored,¹⁸ the specific bioactive constituents of *C. paliurus* and their targets are still unknown. Due to their safety and efficacy, plant-derived triterpenoids have attracted attention for the treatment of diabetes.^{25–27} For example, new cucurbitane-type triterpenoids have shown potential for the prevention and management of diabetes by improving insulin sensitivity and glucose homeostasis.²⁸ Triterpenoids, such as cyclocaric acid B and cyclocarioside H, extracted from *C. paliurus* leaves, promote glucose uptake in the absence of insulin, and ameliorate inflammation by inhibiting the insulin receptor substrate 1 (IRS-1)/phosphoinositide 3-kinase (PI3K)/protein kinase B (Akt) pathway.²⁹ However, no *C. paliurus* triterpenoids with anti-diabetic effects or their specific targets have been described.

Network pharmacology is a new systematic method to study target-drug interactions and their influence on disease. Combined with pharmacology and pharmacodynamics, network pharmacology has been successfully applied to explore the mechanisms of action of traditional botanical or herbal medicines at the molecular level,³⁰ to dissect the relationship

^aSchool of Life Science, Shanghai Normal University, Shanghai 200234, China

^bZhejiang Provincial Key Laboratory of Evolutionary Ecology and Conservation, Taizhou University, Taizhou 318000, China. E-mail: lijmtzc@126.com

^cSchool of Advanced Study, Taizhou University, Taizhou 318000, China

† Electronic supplementary information (ESI) available. See DOI: 10.1039/d0ra06846b



2.4 Component-target network

To elucidate the relationship between *C. paliurus* chemical constituents and diabetes targets, a “component-target-disease” network was analyzed by linking the selected chemical constituents, component-related targets, diabetes-related targets, and the corresponding targets of interactive proteins. The networks were visually analyzed and screened using Cytoscape 3.6.1 software. The screening criteria were as follows: potential targets for the treatment of diabetes with *C. paliurus* had a node value (degree) and BC (betweenness centrality) greater than the median values of all points, and the closeness centrality was greater than the median values of all points. The “degree” and “betweenness” were calculated using the analysis function of Cytoscape 3.6.1 to evaluate its topological features.³⁶

2.5 Protein-protein interaction (PPI) network

To explain interactions between target proteins of related *C. paliurus* compounds, these were uploaded to the STRING database (<https://string-db.org/>). The reliability of protein interactions analyzed with the STRING database is divided into multiple levels, with interaction scores between 0.15 and 0.4 considered of low reliability, between 0.4 and 0.7 of medium reliability, and greater than 0.7 of high reliability.³⁷ Targets showing interactions greater than medium reliability were selected to construct the interaction network of *C. paliurus* anti-diabetic targets. The node interaction relationship parameters were uploaded into Cytoscape 3.6.1 software to draw the interaction network, and the internet was used to obtain the final protein interaction diagram.

2.6 Gene ontology (GO) and Kyoto encyclopedia of genes and genomes (KEGG) enrichment analysis of target proteins

To elucidate the role played by target proteins interacting with the active ingredients of *C. paliurus* in terms of gene function and signaling pathways, the Database for Annotation, Visualization and Integrated Discovery (DAVID, <http://david.ncifcrf.gov/>) 6.8 was used to analyze GO function and KEGG pathway enrichment. The biological process (BP), molecular function (MF) and cell components (CC) of the total targets of *C. paliurus* were analyzed by GO enrichment, and the column graphs were drawn according to the *P* value. The KEGG pathway was also analyzed using DAVID6.8, setting $P \leq 0.05$ and introducing Gene ID from the total targets of *C. paliurus*. According to the enrichment degree and *P* value, the column graphs of the KEGG pathway were drawn. The main signaling pathways of diabetes mellitus were selected and imported into Cytoscape 3.6.1 software for the construction and visualization of the “component-target-pathway” interaction network.

2.7 Component-target-pathway network

After obtaining all the interaction information of the active ingredient compounds, the target proteins, and the pathways, the “Component-Target-Pathway” network was constructed using Cytoscape 3.6.1 software. In this network, nodes represent components, targets, and pathways, and edges represent

interactions between each other. A hypothetical schematic diagram of the target proteins involved in the pathways was drawn. Based on the network model map, a pathway involving active components and target proteins for this disease is initially explored to provide a preliminary theoretical basis for the design of targeted drugs.

2.8 Molecular docking

Target proteins were modified using Pymol software, including hydrogenation, water content, optimization, and repair of amino acid residues. The target proteins' docking pockets were constructed to test the ligands. Interaction between active ingredients and targets was evaluated based on binding energy values by means of AutoDock. The smaller the docking binding energy, the better the molecule will bind to the target protein. When the binding energy is less than 0, the ligand and receptor can bind spontaneously. It is generally believed that energy values below 0 indicate there is a certain binding activity between the target and the ligand; energy values below -5.0 kJ mol^{-1} indicate that the molecule has a good binding activity for the target, and values below $-10.0 \text{ kJ mol}^{-1}$ indicate a strong binding activity.³⁸ The 3D structures of the prepared compounds were used as ligands for molecular docking, and the receptors were the target proteins.^{39–44} A size of CASP3' grid box was set as *x, y, z* center coordinate *i.e.* 41.391, 14.109, 77.896 and *x, y, z* size coordinate *i.e.* 15, 15, 15. A size of MAPK14' grid box was set as *x, y, z* center coordinate *i.e.* 20.647, 13.217, 31.550 and *x, y, z* size coordinate *i.e.* 15, 15, 15.

3. Results

3.1 Screening of potential targets

Based on an extensive survey of the scientific literature, a total of 39 triterpenoids were identified; this includes one compound

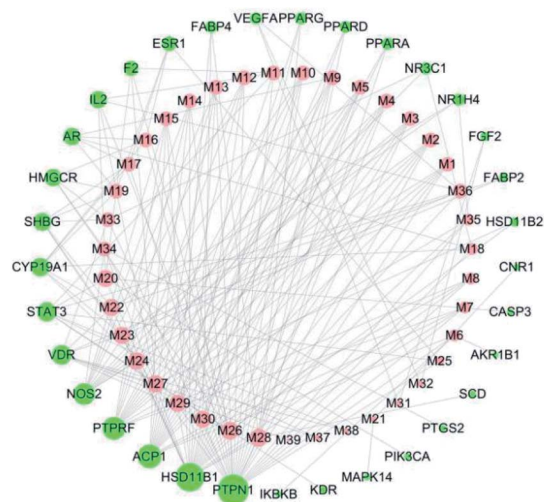


Fig. 2 *Cyclocarya paliurus* component-target network. The red node represents the active ingredients, the green node represents the target proteins, and the size of the nodes represent the size of the degree value.



Table 2 The information of the target proteins used in the manuscript

| UniProt IDs | Protein name | Gene name | Betweenness centrality | Degree |
|-------------|--|-----------|------------------------|--------|
| P24666 | Low molecular weight phosphotyrosine protein phosphatase | ACP1 | 0.04820819 | 16 |
| P15121 | Aldose reductase | AKR1B1 | 0.00271300 | 2 |
| P10275 | Androgen receptor | AR | 0.01510033 | 6 |
| P42574 | Caspase-3 | CASP3 | 0.00164939 | 2 |
| P21554 | Cannabinoid receptor 1 (by homology) | CNR1 | 0.01369327 | 2 |
| P11511 | Cytochrome P450 19A1 | CYP19A1 | 0.02352865 | 8 |
| P03372 | Estrogen receptor alpha | ESR1 | 0.01221787 | 5 |
| P00734 | Thrombin | F2 | 0.00971275 | 5 |
| P12104 | Fatty acid binding protein intestinal | FABP2 | 0.00305104 | 3 |
| P15090 | Fatty acid binding protein adipocyte | FABP4 | 0.00487387 | 4 |
| P09038 | Basic fibroblast growth factor | FGF2 | 0.02471924 | 3 |
| P04035 | HMG-CoA reductase | HMGCR | 0.01564161 | 6 |
| P28845 | 11-Beta-hydroxysteroid dehydrogenase 1 | HSD11B1 | 0.14538756 | 24 |
| P80365 | 11-Beta-hydroxysteroid dehydrogenase 2 | HSD11B2 | 0.00419333 | 3 |
| O14920 | Inhibitor of nuclear factor kappa B kinase beta subunit | IKKBK | 0.00271300 | 2 |
| P60568 | Interleukin-2 | IL2 | 0.01315293 | 6 |
| P35968 | Vascular endothelial growth factor receptor 2 | KDR | 0.00265707 | 2 |
| Q16539 | MAP kinase p38 alpha | MAPK14 | 0.01369327 | 2 |
| P35228 | Nitric oxide synthase, inducible | NOS2 | 0.03106597 | 13 |
| Q96RI1 | Bile acid receptor FXR | NR1H4 | 0.00625265 | 4 |
| P04150 | Glucocorticoid receptor | NR3C1 | 0.00661281 | 4 |
| P42336 | PI3-kinase p110-alpha subunit | PIK3CA | 0.00164939 | 2 |
| Q07869 | Peroxisome proliferator-activated receptor alpha | PPARA | 0.00487387 | 4 |
| Q03181 | Peroxisome proliferator-activated receptor delta | PPARD | 0.00487387 | 4 |
| P37231 | Peroxisome proliferator-activated receptor gamma | PPARG | 0.00487387 | 4 |
| P35354 | Cyclooxygenase-2 | PTGS2 | 0.00176559 | 2 |
| P18031 | Protein-tyrosine phosphatase 1B | PTPN1 | 0.20514580 | 28 |
| P10586 | Receptor-type tyrosine-protein phosphatase F (LAR) | PTPRF | 0.03748757 | 15 |
| O00767 | Acyl-CoA desaturase | SCD | 0.00139742 | 2 |
| P04278 | Testis-specific androgen-binding protein | SHBG | 0.01774612 | 7 |
| P40763 | Signal transducer and activator of transcription 3 | STAT3 | 0.04628878 | 9 |
| P11473 | Vitamin D receptor | VDR | 0.07454926 | 10 |
| P15692 | Vascular endothelial growth factor A | VEGFA | 0.03145277 | 4 |

with multiple compounds. Network values for active components and target proteins are listed in Tables 1 and 2.

3.3 Analysis of target protein PPI network

The interaction network for the anti-diabetic target proteins of *C. paliurus*, including 33 nodes and 153 edges, was constructed based on an interaction reliability greater than mid-range (Fig. 3). In protein interaction networks, the position of node proteins is not the same. The most important nodes are key nodes. The network node degree parameter is a common method for evaluating key nodes of the network. The PPI network showed that the median value of betweenness centrality and closeness centrality was greater than the median with high degree for a total of 15 target proteins. These 15 target proteins (Table 3) can be considered as core target proteins for the treatment of diabetes and may play an important role in the treatment of diabetes. Among them, PTGS2, VEGFA, CASP3, PPARG, STAT3, and NR3C1 are centrally located in the PPI network (Fig. 3), indicating that these proteins are involved in the pathogenesis of diabetes mellitus.

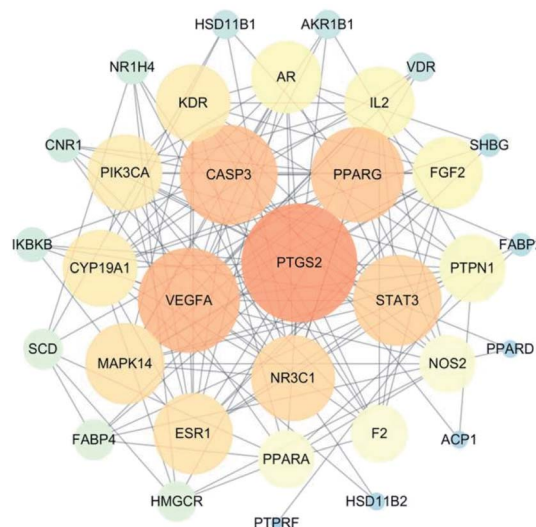


Fig. 3 *Cyclocarya paliurus*'s potential target protein interaction network for treatment of diabetes. The size and color of the node represent the size of the degree value. The darker the color, the larger the node and the more important it is in the network.



Table 3 Features of *Cyclocarya paliurus*'s protein-protein interaction network

| Gene symbol | Name | Betweenness centrality | Closeness centrality | Degree |
|-------------|--|------------------------|----------------------|--------|
| PTGS2 | Prostaglandin-endoperoxide synthase 2 | 0.15607784 | 0.76190476 | 22 |
| VEGFA | Vascular endothelial growth factor A | 0.05924204 | 0.71111111 | 19 |
| CASP3 | Caspase 3 | 0.04400629 | 0.68085106 | 18 |
| PPARG | Peroxisome proliferator activated receptor gamma | 0.14458119 | 0.68085106 | 17 |
| STAT3 | Signal transducer and activator of transcription 3 | 0.02315961 | 0.66666667 | 16 |
| NR3C1 | Nuclear receptor subfamily 3 group C member 1 | 0.07062907 | 0.62745098 | 15 |
| MAPK14 | Mitogen-activated protein kinase 14 | 0.01666107 | 0.64000000 | 14 |
| ESR1 | Estrogen receptor 1 | 0.04629238 | 0.62745098 | 14 |
| KDR | Kinase insert domain receptor | 0.04561908 | 0.60377358 | 13 |
| PIK3CA | Phosphatidylinositol-4,5-bisphosphate 3-kinase catalytic subunit alpha | 0.01214958 | 0.60377358 | 13 |
| CYP19A1 | Cytochrome P450 family 19 subfamily a member 1 | 0.07381888 | 0.60377358 | 13 |
| IL2 | Interleukin 2 | 0.01306559 | 0.57142857 | 12 |
| AR | Androgen receptor | 0.01695036 | 0.58181818 | 12 |
| PTPN1 | Protein tyrosine phosphatase non-receptor type 1 | 0.09420179 | 0.59259259 | 11 |
| PPARA | Peroxisome proliferator activated receptor alpha | 0.03537093 | 0.54237288 | 9 |

3.4 GO analysis of target proteins

GO enrichment analysis results (Fig. 4) showed that the numbers of target proteins involved in the BP, MF, and CC categories were 116 (70.7%), 37 (22.5%), and 11 (6.7%), respectively. In the BP category, the target proteins were mainly involved in positive regulation of transcription from RNA polymerase II promoter (13, 39.4%), signal transduction (9, 29.3%), and transcription, DNA-templated (9, 29.3%). In the MF category, the target proteins were mainly involved in protein binding (23, 69.7%), zinc ion binding (9, 27.3%) and transcription factor activity, sequence-specific DNA binding (9, 27.3%). In the CC category, target proteins were mainly associated with the nucleus (17, 51.5%), cytoplasm (14, 42.4%), nucleoplasm (12, 36.4%) and cytosol (12, 36.4%). These results demonstrate that *C. paliurus* acts on diabetes probably by engaging the above mentioned pathways.

3.5 KEGG classification of target proteins

KEGG pathway annotation showed that 27 of the 33 (81.8%) potential target proteins were enriched and involved in 41

pathways; of these, 35 pathways were significantly correlated with the target proteins ($P \leq 0.05$). The following pathways included the largest number of proteins: cancer pathways (11, 40.7%), proteoglycans in cancer (8, 29.6%), insulin resistance (6, 22.2%), acute myeloid leukemia (6, 22.2%), PI3K-Akt signaling pathway (6, 22.2%), PPAR signaling pathway (6, 22.2%), and so on. The top 10 pathways with the largest number of proteins involved are shown in Fig. 4, and the complete KEGG classification results are shown in Table 4. The target proteins involved in cancer pathways included

AR, CASP3, PPARG, PTGS2, VEGFA, PPARG, PIK3CA, NOS2, IKKKB, FGF2 and STAT3; the target proteins involved in insulin resistance included PPARA, PTPRF, PIK3CA, PTPN1, IKKKB and STAT3; and the target proteins involved in the PI3K-Akt signaling pathway included VEGFA, PIK3CA, IKKKB, FGF2, KDR and IL2. As can be seen, there are multiple target proteins in a pathway and the same target protein exists in multiple pathways. These results suggest that *C. paliurus* may exert its effects on diabetes by regulating these pathways *via* core target proteins.

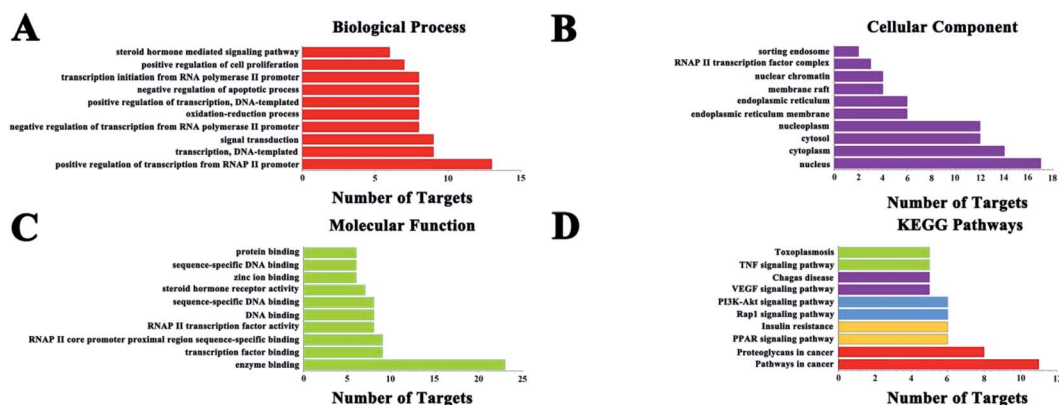


Fig. 4 GO and KEGG pathways analyses of the target proteins by using the DAVID database. Data show the top 10 remarkably enriched items in the biological process (A), cell component (B), molecular function (C), and KEGG pathways (D).



Table 4 The result of KEGG Classification of target proteins^a

| Term | Count | Percent% | P value | Genes |
|--|-------|----------|---------|---|
| hsa05200:Pathways in cancer | 11 | 33.33 | 0.00 | AR, CASP3, PPARD, PTGS2, VEGFA, PPARG, PIK3CA, NOS2, IKBKB, FGF2, STAT3 |
| hsa05205:Proteoglycans in cancer | 8 | 24.24 | 0.00 | CASP3, MAPK14, VEGFA, ESRI, PIK3CA, FGF2, STAT3, KDR |
| hsa03320:PPAR signaling pathway | 6 | 18.18 | 0.00 | PPARA, PPARD, SCD, PPARG, FABP4, FABP2 |
| hsa04931:Insulin resistance | 6 | 18.18 | 0.00 | PPARA, PTPRF, PIK3CA, PTPN1, IKBKB, STAT3 |
| hsa04015:Rap1 signaling pathway | 6 | 18.18 | 0.00 | MAPK14, CNR1, VEGFA, PIK3CA, FGF2, KDR |
| hsa04151:PI3K-Akt signaling pathway | 6 | 18.18 | 0.02 | VEGFA, PIK3CA, IKBKB, FGF2, KDR, IL2 |
| hsa04370:VEGF signaling pathway | 5 | 15.15 | 0.00 | PTGS2, MAPK14, VEGFA, PIK3CA, KDR |
| hsa05142:Chagas disease | 5 | 15.15 | 0.00 | MAPK14, PIK3CA, NOS2, IKBKB, IL2 |
| hsa04668:TNF signaling pathway | 5 | 15.15 | 0.00 | CASP3, PTGS2, MAPK14, PIK3CA, IKBKB |
| hsa05145:Toxoplasmosis | 5 | 15.15 | 0.00 | CASP3, MAPK14, NOS2, IKBKB, STAT3 |
| hsa05160:Hepatitis C | 5 | 15.15 | 0.00 | PPARA, MAPK14, PIK3CA, IKBKB, STAT3 |
| hsa04014:Ras signaling pathway | 5 | 15.15 | 0.02 | VEGFA, PIK3CA, IKBKB, FGF2, KDR |
| hsa05206:MicroRNAs in cancer | 5 | 15.15 | 0.03 | CASP3, PTGS2, VEGFA, IKBKB, STAT3 |
| hsa05221:Acute myeloid leukemia | 4 | 12.12 | 0.00 | PPARD, PIK3CA, IKBKB, STAT3 |
| hsa05212:Pancreatic cancer | 4 | 12.12 | 0.00 | VEGFA, PIK3CA, IKBKB, STAT3 |
| hsa04917:Prolactin signaling pathway | 4 | 12.12 | 0.00 | MAPK14, ESR1, PIK3CA, STAT3 |
| hsa05222:Small cell lung cancer | 4 | 12.12 | 0.01 | PTGS2, PIK3CA, NOS2, IKBKB |
| hsa04066:HIF-1 signaling pathway | 4 | 12.12 | 0.01 | VEGFA, PIK3CA, NOS2, STAT3 |
| hsa04660:T cell receptor signaling pathway | 4 | 12.12 | 0.01 | MAPK14, PIK3CA, IKBKB, IL2 |
| hsa05169:Epstein-Barr virus infection | 4 | 12.12 | 0.02 | MAPK14, PIK3CA, IKBKB, STAT3 |
| hsa04152:AMPK signaling pathway | 4 | 12.12 | 0.02 | HMGCR, SCD, PPARG, PIK3CA |
| hsa04380:Osteoclast differentiation | 4 | 12.12 | 0.02 | MAPK14, PPARG, PIK3CA, IKBKB |
| hsa04068:FoxO signaling pathway | 4 | 12.12 | 0.02 | MAPK14, PIK3CA, IKBKB, STAT3 |
| hsa04910:Insulin signaling pathway | 4 | 12.12 | 0.02 | PTPRF, PIK3CA, PTPN1, IKBKB |
| hsa04550:Signaling pathways regulating pluripotency of stem cells | 4 | 12.12 | 0.02 | MAPK14, PIK3CA, FGF2, STAT3 |
| hsa05161:Hepatitis B | 4 | 12.12 | 0.02 | CASP3, PIK3CA, IKBKB, STAT3 |
| hsa04932:Non-alcoholic fatty liver disease (NAFLD) | 4 | 12.12 | 0.03 | PPARA, CASP3, PIK3CA, IKBKB |
| hsa05152:Tuberculosis | 4 | 12.12 | 0.04 | VDR, CASP3, MAPK14, NOS2 |
| hsa04923:Regulation of lipolysis in adipocytes | 3 | 9.09 | 0.02 | PTGS2, PIK3CA, FABP4 |
| hsa00140:Steroid hormone biosynthesis | 3 | 9.09 | 0.03 | HSD11B1, HSD11B2, CYP19A1 |
| hsa04210:Apoptosis | 3 | 9.09 | 0.03 | CASP3, PIK3CA, IKBKB |
| hsa05120:Epithelial cell signaling in <i>Helicobacter pylori</i> infection | 3 | 9.09 | 0.03 | CASP3, MAPK14, IKBKB |
| hsa04920:Adipocytokine signaling pathway | 3 | 9.09 | 0.04 | PPARA, IKBKB, STAT3 |
| hsa05140:Leishmaniasis | 3 | 9.09 | 0.04 | PTGS2, MAPK14, NOS2 |
| hsa05133:Pertussis | 3 | 9.09 | 0.04 | CASP3, MAPK14, NOS2 |

^a Bold text means the important pathways reported and involved in diabetes.

3.6 Component-target-pathway network

The component-target-pathway network was constructed to visualize all interactions between target proteins and the anti-diabetic-related pathways. Based on the previous KEGG enrichment analysis, a component-target-pathway network was generated by connecting compounds, targets and pathways (Fig. 5). This network included 106 nodes (39 active compound nodes, 33 composite target protein nodes and 34 pathways nodes) and 745 edges. The component-target-pathway network results are shown in ESI Table 2 and ESI Fig. 1.†

The network analysis showed that the median value of betweenness centrality and closeness centrality was greater than the median with high degree for a total of 15 pathways, including cancer (hsa05200), insulin resistance (hsa04931), HIF-1 signaling pathway (hsa04066), PI3K-Akt signaling pathway (hsa04151) *etc.* These 15 pathways with several diabetes-associated target proteins can be considered as core

pathways for the treatment of diabetes. Among these pathways, insulin resistance, HIF-1 signaling, and PI3K-Akt signaling pathways have been shown to have a clear link with the occurrence of diabetes. As shown in Fig. 6, the active anti-inflammatory and anti-diabetic components present in *C. pal- iurus* can synergize with multiple target proteins within these pathways to form a multi-component-multi-target-multi-pathway mechanism.

3.7 Docking analysis

The protein structure was set to a rigid macromolecule, and the algorithm to local search parameters. Then, the object of molecular docking visualization was selected according to the lowest binding energy for docking. The core compounds and diabetes target proteins were ranked according to the docking binding energy. The top 10 core target proteins in the previous ranking were docked with 7 key medicinal components (Table



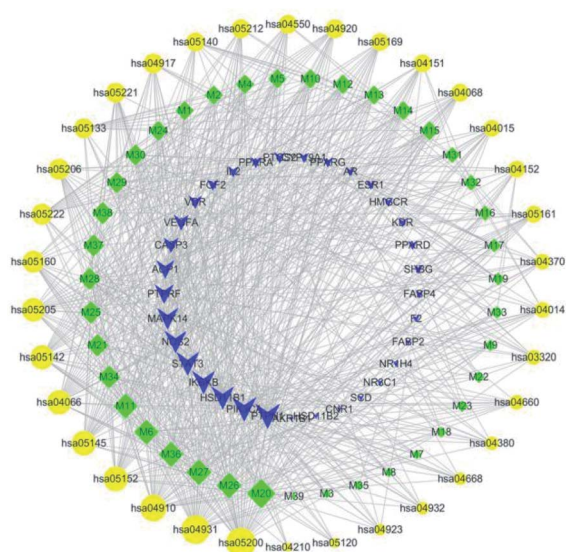


Fig. 5 *Cyclocarya paliurus*'s component-target-pathway network. The green diamond-shaped node represents the active ingredient, the blue triangular node represents the target, the yellow circular node represents the pathway, and the size of the node represent the degree value. Lines represent the relationships between the compounds, targets, and pathways.

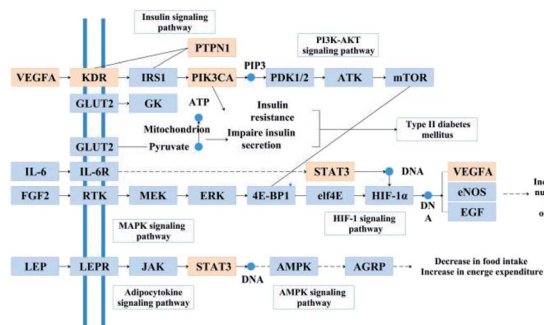


Fig. 6 Distribution of the target proteins of *Cyclocarya paliurus* on the predicted pathway. The orange boxes are potential target proteins of *C. paliurus*, while the blue boxes are relevant targets in the pathway.

5). The docking results showed that the lowest binding energy for the 7 core medicinal components and CASP3 were all below -5.0 kJ mol^{-1} , indicating that the 7 key medicinal components bound better to the CASP3 target protein. With the exception of $2\alpha,3\beta,23$ -trihydroxyoleana-11,13(18)-dien-28-oic acid, the other 6 components showed the lowest docking binding energies with MAPK14 target protein when compared with the other 9 target proteins. Interactions between the 7 core components of *C. paliurus* and the previously mentioned 10 core target proteins with the lowest binding energy and the smallest binding energy were visualized using Pymol (Fig. 7 and 8).

4. Discussion

Used as a health tea, *C. paliurus* has a long history in the treatment of diabetes. In this study, 39 triterpenoids were

selected to create a library of compounds, and network pharmacology technology was employed to explore the multiple pharmacological effects of triterpenoids from *C. paliurus* in the treatment of diabetes. We found that seven core ingredients found in the leaves of *C. paliurus*, including arjunglucoside II, α -boswellic acid, $3\beta,23$ -dihydroxy-1,12-dioxo-olean-28-oic acid, 23 -trihydroxyurs-11-oxo-12-ene-28-oic acid, $3\beta,23,27$ -trihydroxy-1-oxo-olean-12-ene-28-oic acid, $2\alpha,3\beta,23$ -trihydroxyoleana-11,13(18)-dien-28-oic acid and cyclocarioside K, showed potential for the treatment of diabetes.

Among them, α -boswellic acid has been shown to exert some anti-diabetic effects by suppressing the expression of proinflammatory cytokines.⁴⁵ Cyclocarioside K is a new epoxydammarane triterpenoid saponin isolated from the ethanol leaf extracts of *C. paliurus*.⁴⁶ Arjunglucoside II is a common glucoside derivative of arjunic acid, found in *Terminalia arjuna*.⁴⁷ The DPP-IV inhibitory activity of arjunic acid and its derivatives may have demonstrable therapeutic benefits in diabetes patients with cardiovascular comorbidities.⁴⁸ $3\beta,23$ -Dihydroxy-1,12-dioxo-olean-28-oic acid and $3\beta,23,27$ -trihydroxy-1-oxo-olean-12-ene-28-oic acid are new triterpenoid saponins isolated from the CH_3Cl -soluble extract of *C. paliurus* leaves, whereas 23 -trihydroxyurs-11-oxo-12-ene-28-oic acid and $2\alpha,3\beta,23$ -trihydroxyoleana-11,13(18)-dien-28-oic acid are also known triterpenoid saponins isolated from the CH_3Cl -soluble extract of the leaves of *C. paliurus*.⁴⁹ These triterpenoid saponins may have an effect on diabetes by inhibiting apolipoprotein B48 overproduction.^{49,50} Based on the interaction network of *C. paliurus*'s anti-diabetes target proteins, we found that STAT3, PTPN1, PTGS2, VEGFA, CASP3, etc. were the key target proteins. These targets may play an important role in *C. paliurus*'s anti-diabetes effects. According to the results of the component-target and PPI networks, STAT3, CASP3, ACPI1, PTPN1, HSD11B1 and other targets were regulated by multiple components, such as arjunglucoside II, cyclocarioside K, quadranoside IV, corosolic acid, cyclocariosides I and so on. Some of these ingredients have been proven to have strong anti-diabetic effects. For example, corosolic acid can reduce glucose levels in human hepatocellular carcinoma cells, as well as in zebrafish and in rats.⁵¹ Molecular docking is one of the most widely used methods to investigate binding of a compound to a target protein.³⁹⁻⁴² We used AutoDock software to analyze molecular docking. Compounds with docking energies below -5 kJ mol^{-1} were considered to bind well to their targets. In the PPI network diagram, betweenness is defined as the number of shortest paths between pairs of nodes that run through nodes, and degree centrality is the most direct measure of the importance of a network node. The larger the node degree value, the more important the node is in the network. We used two indicators (betweenness centrality and closeness centrality) as the criteria for screening targets. Only target proteins with betweenness centrality and closeness centrality values greater than the median for all target proteins were selected. Then, all target proteins were sorted according to the degree value. Based on the docking results between the top ten target proteins and core compounds, we observed that although the docking energy of some core compounds to the target proteins was very small



Table 5 The lowest binding energy of the active ingredients of *Cyclocarya paliurus* to the ten core target proteins for treating diabetes (unit: kcal mol⁻¹)^a

| No. | Compound name | PTGS2 | VEGFA | CASP3 | PPARG | STAT3 | NR3C1 | MAPK14 | ESR1 | KDR | PIK3CA |
|-----|--|-------|-------|-------|-------|-------|-------|--------|-------|------|--------|
| 1 | 3β,23-Dihydroxy-1,12-dioxo-olean-28-oic acid | -5.2 | -4.5 | -6.1 | 2.9 | -5.8 | -2.4 | -6.7 | -4.7 | -5 | -5.3 |
| 2 | 3β,23,27-Trihydroxy-1-oxo-olean-12-ene-28-oic acid | -5.2 | -4.5 | -6.1 | 3 | -5.8 | -2.4 | -6.7 | -4.7 | -4.9 | -5.3 |
| 3 | 2α,3β,23-Trihydroxyurs-11-oxo-12-ene-28-oic acid | -4.9 | -4.6 | -5.2 | 0 | -5.2 | -3.6 | -8.7 | -2.9 | -5 | -6.1 |
| 4 | Arjunglucoside II | -4.8 | -4.6 | -5.5 | 2.3 | -5.4 | 8.2 | -6.7 | -4.4 | -4.6 | -5.1 |
| 5 | α-Boswellic acid | -5.7 | -5.3 | -6.1 | -1.7 | -6 | -6.3 | -9.1 | -6.2 | -5.2 | -5.4 |
| 6 | 2α,3β,23-Trihydroxyoleana-11,13(18)-dien-28-oic acid | -4.5 | -4 | -6 | 1.7 | -5.3 | 16.5 | -0.3 | 3.1 | -4.7 | -2.4 |
| 7 | Cyclocarioside K | -4.3 | -4.9 | -4.9 | -2.9 | -5.7 | 4.5 | -8 | -5.4 | -4.1 | -6.7 |
| 8 | Ligand | -3.1 | -3.2 | -3.4 | -5.5 | -6.1 | -11.9 | -12.7 | -11.4 | -3 | -7.9 |

^a Ligand means a substance that has the ability to recognize the receptor and can bind to it. The docking method is rigid docking. Algorithm is all local search parameters.

and the docking results were good, there were also poor docking results. Only STAT3 showed docking energies for all compounds below -5.0 kJ mol⁻¹. STAT3 is an important signal transduction factor that participates in the signal transduction process of various cytokines, such as interferon, interleukins and growth factors, and it forms part of the JAK2/STAT3 signaling pathway.^{52,53} Many studies have shown that the JAK2/STAT3 signaling pathway is an important pathway that mediates the signal transduction process of various cytokines and growth factors, and can regulate cell growth, differentiation, migration, apoptosis, autophagy, immunity, and metabolism.^{54,55} It is considered to play an important role in the development of diabetes.^{56,57} STAT3 may regulate diabetes by acting on SOCS3 and thereby affecting the activity of insulin receptor substrate 1 (IRS-1).⁵⁸ SOCS3 can mediate central leptin resistance in obese patients.⁵⁹ Leptin signal transduction in the hypothalamus can regulate liver glucose and lipid metabolism⁶⁰ as well as liver gluconeogenesis.⁶¹ It seems evident that STAT3 plays an important role regulating disorders of glucose metabolism. In network pharmacology analysis, component-target networks are mainly used to screen core compounds and targets. Betweenness centrality and closeness centrality values greater than the median are also the main criteria for screening core target proteins. All eligible target proteins were sorted according to the degree value, and the one with the highest value was PTPN1. In component-target networks, the higher the degree of a target protein, the more the compounds that regulate it. The more active compounds bind to a protein target, the greater the possibility of producing a drug effect, and the higher the possibility of becoming the key node. PTPN1 is a non-transmembrane protein tyrosine phosphatase which functions mainly by regulating the levels of protein tyrosine phosphorylation in cells, and is involved in the regulation of multiple cell signaling pathways modulating cell proliferation, growth and migration.⁶² Many studies have found correlations between DNA sequence variations in the PTPN1 gene, SNP and T2DM.⁶³ PTPN1 can regulate insulin signaling by modulating the

expression of the PTP1B enzyme. PTP1B is a key factor that regulates various metabolic processes in the body and is closely associated with the occurrence and development of diseases, especially type 2 diabetes.⁶⁴ Studies have shown that inhibiting the expression of the PTP1B enzyme can regulate insulin sensitivity in diabetic mice and improve insulin resistance, producing a therapeutic effect in diabetes.⁶⁵ Therefore, we can study the potential pharmacological effects of *C. paliurus* on diabetes by analyzing the relationship between active compounds and core targets. According to the results of the component-target-protein network analysis, the insulin resistance pathway, the PI3K-Akt signaling pathway and the HIF-1 pathway may play an important role in the anti-diabetes effects of *C. paliurus*. As shown in Fig. 5, the active anti-diabetes ingredients of *C. paliurus* can synergize with various target proteins in these pathways, resulting in a multi-component, multi-target and multi-pathway mechanism of action. Insulin resistance, a condition in which cells are resistant to the action of insulin, is often found in obese and diabetic patients and is closely related with the occurrence of diabetes.⁶⁶ It is associated with increased activity of phosphatases, including PTPs, PTEN, and PP2A, and decreased activation of signaling molecules, such as PI3K/AKT, resulting in reduced GLUT4 translocation, glucose uptake and glycogen synthesis in skeletal muscle, as well as increased hepatic gluconeogenesis and decreased glycogen synthesis in the liver.^{67,68} Many studies have shown that this pathway is activated by changes in the levels of enzymes such as IL-6, Akt and IRS-1, or other abnormalities.^{69,70}

HIF-1, a transcription factor which functions as a major regulator of oxygen homeostasis,⁷¹ is associated with diabetic nephropathy. HIF-1 expression indirectly affects renal oxygen metabolism.^{72,73} In hypoxic conditions, it can reduce damage to the body by regulating the expression of target genes coding for downstream factors related with the hypoxic response, and by modulating cell energy metabolism, glucose metabolism and apoptosis.⁷⁴ Studies have shown that oxidative stress and



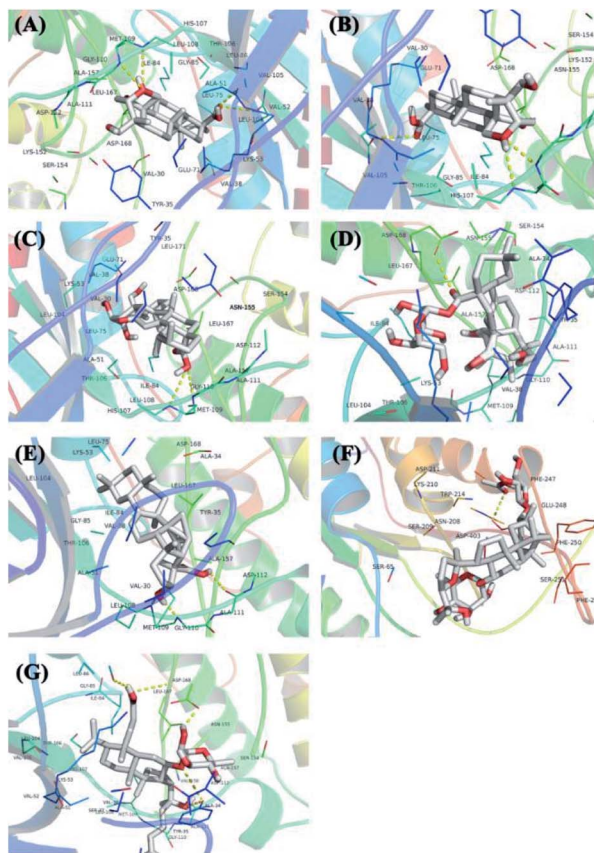


Fig. 7 The 3D docking results of ten core protein molecules and seven core components are visualized. The structure of the compound is represented by a stick, the different branches of the protein are represented by different colors, and the yellow dotted line represents its hydrogen bond, which marks the position of the hydrogen bond and the compound in the compound. (A) Interaction between $3\beta,23$ -dihydroxy-1,12-dioxo-olean-28-oic acid and MAPK14; (B) interaction between $3\beta,23,27$ -trihydroxy-1-oxo-olean-12-ene-28-oic acid and MAPK14; (C) interaction between $2\alpha,3\beta,23$ -trihydroxyurs-11-oxo-12-ene-28-oic acid and MAPK14; (D) interaction between arjungucoside II and MAPK14; (E) interaction between α -boswellic acid and MAPK14; (F) interaction between $2\alpha,3\beta,23$ -trihydroxyoleana-11,13(18)-dien-28-oic acid and CASP3; (G) interaction between cyclocarioside K and MAPK14.

microcirculatory disorders are important mechanisms underlying the development of diabetic nephropathy and other diseases.⁷⁵ Antioxidant therapy can increase the expression of medullary HIF in diabetic kidneys and improve renal oxidative damage.⁷⁶ HIF activation can attenuate changes in renal oxygen metabolism and mitochondrial function caused by diabetes, thereby reducing proteinuria and improving renal tubular interstitial damage.⁷⁷ The PI3K-Akt signaling pathway plays an important role maintaining insulin stability.⁷⁵ This signaling pathway is activated by many types of cellular stimuli or toxic insults and regulates fundamental cellular functions, such as transcription, translation, proliferation, growth, and survival.⁷⁶ At present, there are studies showing that selective activation of the PI3K/AKT signaling pathway can modulate neuroprotection, angiogenesis, and islet β cell survival in diabetic rats.⁷⁵ Many

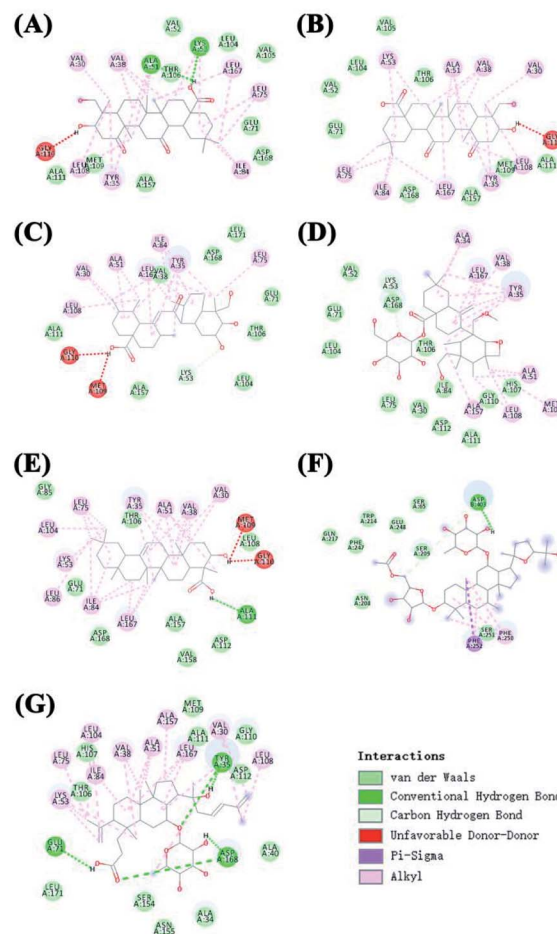


Fig. 8 The 2D docking results of ten core protein molecules and seven core components are visualized. (A) Interaction between $3\beta,23$ -dihydroxy-1,12-dioxo-olean-28-oic acid and MAPK14; (B) interaction between $3\beta,23,27$ -trihydroxy-1-oxo-olean-12-ene-28-oic acid and MAPK14; (C) interaction between $2\alpha,3\beta,23$ -trihydroxyurs-11-oxo-12-ene-28-oic acid and MAPK14; (D) interaction between arjungucoside II and MAPK14; (E) interaction between α -boswellic acid and MAPK14; (F) interaction between $2\alpha,3\beta,23$ -trihydroxyoleana-11,13(18)-dien-28-oic acid and CASP3; (G) interaction between cyclocarioside K and MAPK14.

studies indicate that after insulin binds to its receptor, it will self-phosphorylate and phosphorylate IRS-2 tyrosine sites.⁷⁷ Phosphorylated IRS-2 binds to the p85 subunit of PI3K to further activate PI3K and to activate Akt after phosphorylation.⁷⁸ Akt2 is a subtype of Akt, a serine/threonine kinase and an important signaling molecule located downstream of PI3K^{79,80} which can be regulated by modulating Gsk-3 β , GLUT-4, *etc.* A series of downstream molecules promote glycogen synthesis, glucose transport, and other pathways to regulate glucose metabolism, thereby regulating diabetes.⁷⁹

Receptor theory is the basic theory of pharmacodynamics. It postulates that the combination of drugs and target proteins is the basis of pharmacodynamics. Molecular docking is a method to evaluate binding of active drug ingredients to target proteins. In network pharmacology research, molecular docking is commonly used to study interactions between small molecules and key target proteins of the network, and to validate key target



- 12 H. Zhang, H. Matsuda, A. Kumahara, Y. Ito, S. Nakamura and M. Yoshikawa, New Type of Anti-Diabetic Compounds From the Processed Leaves of *Hydrangea Macrophylla* Var. *Thunbergii* (Hydrangeae Dulcis Folium), *Bioorg. Med. Chem. Lett.*, 2007, **17**, 4972–4976.
- 13 L. Zhai, Z. Ning, T. Huang, B. Wen, C. Liao, C. Lin, L. Zhao, H. Xiao and Z. Bian, *Cyclocarya paliurus* Leaves Tea Improves Dyslipidemia in Diabetic Mice: A Lipidomics-Based Network Pharmacology Study, *Front. Pharmacol.*, 2018, **9**, 973.
- 14 Z. Yang, K. Ouyang, J. Zhao, H. Chen, L. Xiong and W. Wang, Structural Characterization and Hypolipidemic Effect of *Cyclocarya Paliurus* Polysaccharide in Rat, *Int. J. Biol. Macromol.*, 2016, **91**, 1073–1080.
- 15 H. Kurihara, H. Fukami, A. Kusumoto, Y. Toyoda, H. Shibata, Y. Matsui, S. Asami and T. Tanaka, Hypoglycemic Action of *Cyclocarya paliurus* (Batal.) Iljinskaja in Normal and Diabetic Mice, *Biosci., Biotechnol., Biochem.*, 2003, **67**, 877–880.
- 16 S. Li, J. Li, X. Guan, J. Li, S. Deng, L. Li, M. Tang, J. Huang, Z. Chen and R. Yang, Hypoglycemic Effects and Constituents of the Barks of *Cyclocarya Paliurus* and their Inhibiting Activities to Glucosidase and Glycogen Phosphorylase, *Fitoterapia*, 2011, **82**, 1081–1085.
- 17 Y. Ma, C. Jiang, N. Yao, Y. Li and Q. Wang, Antihyperlipidemic Effect of *Cyclocarya Paliurus* (Batal.) Iljinskaja Extract and Inhibition of Apolipoprotein B48 Overproduction in Hyperlipidemic Mice, *J. Ethnopharmacol.*, 2015, **166**, 286–296.
- 18 Q. Wang, C. Jiang, S. Fang, J. Wang, Y. Ji, X. Shang, Y. Ni, Z. Yin and J. Zhang, Antihyperglycemic, Antihyperlipidemic and Antioxidant Effects of Ethanol and Aqueous Extracts of *Cyclocarya Paliurus* Leaves in Type 2 Diabetic Rats, *J. Ethnopharmacol.*, 2013, **150**, 1119–1127.
- 19 J. Xie, D. Cai-jun, N. Shao-Ping, L. Feng, W. Zhi-Jun, S. Ming-yue and M. Xie, Extraction, Chemical Composition and Antioxidant Activity of Flavonoids From *Cyclocarya Paliurus* (Batal.) Iljinskaja Leaves, *Food Chem.*, 2015, **186**, 186–197.
- 20 J. Xie, M. Xie, S. Nie, M. Shen, Y. Wang and C. Li, Isolation, Chemical Composition and Antioxidant Activities of a Water-Soluble Polysaccharide From *Cyclocarya Paliurus* (Batal.) Iljinskaja, *Food Chem.*, 2010, **119**, 1626–1632.
- 21 H. Yoshitomi, R. Tsuru, L. Li, J. Zhou, M. Kudo, T. Liu and M. Gao, *Cyclocarya Paliurus* Extract Activates Insulin Signaling Via Sirtuin1 in C2C12 Myotubes and Decreases Blood Glucose Level in Mice with Impaired Insulin Secretion, *PLoS One*, 2017, **12**, e183988.
- 22 C. Jiang, Q. Wang, Y. Wei, N. Yao, Z. Wu, Y. Ma, Z. Lin, M. Zhao, C. Che, X. Yao, J. Zhang and Z. Yin, Cholesterol-Lowering Effects and Potential Mechanisms of Different Polar Extracts From *Cyclocarya Paliurus* Leave in Hyperlipidemic Mice, *J. Ethnopharmacol.*, 2015, **176**, 17–26.
- 23 W. Lin, Y. Li, Q. Lu, H. Lu and J. Li, Combined Analysis of the Metabolome and Transcriptome Identified Candidate Genes Involved in Phenolic Acid Biosynthesis in the Leaves of *Cyclocarya Paliurus*, *Int. J. Mol. Sci.*, 2020, **21**, 1337.
- 24 C. Sun, X. Shang, H. Ding, Y. Cao and S. Fang, Natural Variations in Flavonoids and Triterpenoids of *Cyclocarya Paliurus* Leaves, *J. For. Res.*, 2020, **31**, DOI: 10.1007/s11676-020-01139-1.
- 25 J. G. A. W. Du, M. Zhang and Li Chen, Compound K, a Final Intestinal Metabolite of Ginsenosides, Enhances Insulin Secretion in MIN6 Pancreatic β -Cells by Upregulation of GLUT2, *Fitoterapia*, 2013, **87**, 84–88.
- 26 A. E. Ardiles, Á. González-Rodríguez, M. J. N. Ez, N. R. Perestelo, V. Pardo, I. A. Jiménez, Á. M. Valverde and I. L. Bazzocchi, Studies of Naturally Occurring Friedelane Triterpenoids as Insulin Sensitizers in the Treatment Type 2 Diabetes Mellitus, *Phytochemistry*, 2012, **84**, 116–124.
- 27 D. K. Burdi, S. N. Qureshi and A. B. Ghanghro, An Overview of Available Hypoglycemic Triterpenoids and Saponins to Cure Diabetes Mellitus, *Adv. Life Sci.*, 2014, **1**, 119–128.
- 28 J. Han, N. Tuan, M. Park, K. Quan, J. Oh, K. Heo, M. Na and C. Myung, Cucurbitane Triterpenoids From the Fruits of *Momordica Charantia* Improve Insulin Sensitivity and Glucose Homeostasis in Streptozotocin-Induced Diabetic Mice, *Mol. Nutr. Food Res.*, 2018, **62**, 1700769.
- 29 K. N. Zhu, C. H. Jiang, Y. S. Tian, N. Xiao, Z. F. Wu, Y. L. Ma, Z. Lin, S. Z. Fang, X. L. Shang, K. Liu, J. Zhang, B. L. Liu and Z. Q. Yin, Two Triterpenoids From *Cyclocarya Paliurus* (Batal) Iljinsk (Juglandaceae) Promote Glucose Uptake in 3T3-L1 Adipocytes: The Relationship to AMPK Activation, *Phytomedicine*, 2015, **22**, 837–846.
- 30 S. Yue, L. Xin, Y. Fan, S. Li, Y. Tang, J. Duan, H. Guan and C. Wang, Herb Pair Danggui-Honghua: Mechanisms Underlying Blood Stasis Syndrome by System Pharmacology Approach, *Sci. Rep.*, 2017, **7**, 40318.
- 31 M. U. Kakar, M. Naveed, M. Saeed, S. Zhao, M. Rasheed, S. Firdoos, R. Manzoor, Y. Deng and R. Dai, A Review On Structure, Extraction, and Biological Activities of Polysaccharides Isolated From *Cyclocarya Paliurus* (Batalin) Iljinskaja, *Int. J. Biol. Macromol.*, 2020, **156**, 420–429.
- 32 Z. Wu and T. Ding, *Seed Plants of China*, Kunming: Yunnan Science and Technology Press., 1999.
- 33 Z. Li, X. Ma, D. Wang, Y. Li, C. Wang and X. Jin, Evolution of Plastid Genomes of *Holcoglossum* (Orchidaceae) with Recent Radiation, *BMC Evol. Biol.*, 2019, **19**, 63.
- 34 J. Ru, P. Li, J. Wang, W. Zhou, B. Li, C. Huang, P. Li, Z. Guo, W. Tao, Y. Yang, X. Xu, Y. Li, Y. Wang and L. Yang, TCMSP: A Database of Systems Pharmacology for Drug Discovery From Herbal Medicines, *J. Cheminf.*, 2014, **6**, 13.
- 35 D. Gfeller, O. Michielin and V. Zoete, Shaping the Interaction Landscape of Bioactive Molecules, *Bioinformatics*, 2013, **29**, 3073–3079.
- 36 X. Liu, J. Wu, D. Zhang, K. Wang, X. Duan and X. Zhang, A Network Pharmacology Approach to Uncover the Multiple Mechanisms of *Hedyotis Diffusa* Willd. On Colorectal Cancer, *J. Cheminf.*, 2018, **2018**, 6517034.
- 37 C. von Mering, L. J. Jensen, B. Snel, S. D. Hooper, M. Krupp, M. Foglierini, N. Jouffre, M. A. Huynen and P. Bork, STRING: Known and Predicted Protein-Protein Associations, Integrated and Transferred Across Organisms, *Nucleic Acids Res.*, 2004, **33**, D433–D437.
- 38 R. Yu, L. Chen, R. Lan, R. Shen and P. Li, Computational Screening of Antagonists Against the SARS-CoV-2 (COVID-



- 19) Coronavirus by Molecular Docking, *Int. J. Antimicrob. Agents*, 2020, **56**, 106012.
- 39 X. G. Liu, M. C. Lv, M. Y. Huang, Y. Q. Sun, P. Y. Gao and D. Q. Li, A Network Pharmacology Study On the Triterpene Saponins from *Medicago Sativa* L. For the Treatment of Neurodegenerative Diseases, *J. Food Biochem.*, 2019, **43**, e12955.
- 40 J. Li, X. Ma, C. Liu, H. Li, J. Zhuang, C. Gao, C. Zhou, L. Liu, K. Wang and C. Sun, Exploring the Mechanism of Danshen against Myelofibrosis by Network Pharmacology and Molecular Docking, *Evid. base Compl. Alternative Med.*, 2018, **2018**, 1–11.
- 41 R. Liu, C. Su, Y. Xu, K. Shang, K. Sun, C. Li and J. Lu, Identifying Potential Active Components of Walnut Leaf that Action Diabetes Mellitus through Integration of UHPLC-Q-Orbitrap HRMS and Network Pharmacology Analysis, *J. Ethnopharmacol.*, 2020, **253**, 112659.
- 42 Q. Luo, X. Shi, J. Ding, Z. Ma, X. Chen, Y. Leng, X. Zhang and Y. Liu, Network Pharmacology Integrated Molecular Docking Reveals the Antiosteosarcoma Mechanism of *Biochanin A*, *Evid. base Compl. Alternative Med.*, 2019, **2019**, 1–10.
- 43 Y. Xiong, Y. Yang, W. Xiong, Y. Yao, H. Wu and M. Zhang, Network Pharmacology-Based Research On the Active Component and Mechanism of the Antihepatoma Effect of *Rubia Cordifolia* L., *J. Cell. Biochem.*, 2019, **120**, 12461–12472.
- 44 C. Mu, Y. Sheng, Q. Wang, A. Amin and Y. Xie, Potential Compound From Herbal Food of *Rhizoma Polygonati* for Treatment of COVID-19 Analyzed by Network Pharmacology and Molecular Docking Technology, *J. Funct. Foods*, 2020, 104149.
- 45 H. P. T. Ammon, Boswellic Extracts and 11-Keto- β -Boswellic Acids Prevent Type 1 and Type 2 Diabetes Mellitus by Suppressing the Expression of Proinflammatory Cytokines, *Phytomedicine*, 2019, **63**, 153002.
- 46 B. Cui and S. Li, New Triterpenoid Saponins From the Leaves of *Cyclocarya Paliurus*, *Chin. Chem. Lett.*, 2015, **26**, 585–589.
- 47 T. Honda, T. Murae, T. Tsuyuki, T. Takahashi and M. Sawai, Arjungenin, Arjunglucoside I, and Arjunglucoside II. A New Triterpene and New Triterpene Glucosides From *Terminalia Arjuna*, *Bull. Chem. Soc. Jpn.*, 1976, **49**, 3213–3218.
- 48 I. R. Mohanty, M. Borde, S. Kumar C and U. Maheshwari, Dipeptidyl Peptidase IV Inhibitory Activity of *Terminalia Arjuna* Attributes to its Cardioprotective Effects in Experimental Diabetes: In Silico, in Vitro and in Vivo Analyses, *Phytomedicine*, 2019, **57**, 158–165.
- 49 Z. Wu, F. Meng, L. Cao, C. Jiang, M. Zhao, X. Shang, S. Fang, W. Ye, Q. Zhang, J. Zhang and Z. Yin, Triterpenoids From *Cyclocarya Paliurus* and their Inhibitory Effect On the Secretion of Apolipoprotein B48 in Caco-2 Cells, *Phytochemistry*, 2017, **142**, 76–84.
- 50 C. Phillips, G. Murugasu, D. Owens, P. Collins, A. Johnson and G. H. Tomkin, Improved Metabolic Control Reduces the Number of Postprandial Apolipoprotein B-48-containing Particles in Type 2 Diabetes, *Atherosclerosis*, 2000, **148**, 283–291.
- 51 S. Xu, G. Wang, W. Peng, Y. Xu, Y. Zhang, Y. Ge, Y. Jing and Z. Gong, Corosolic Acid Isolated From *Eriobotrya Japonica* Leaves Reduces Glucose Level in Human Hepatocellular Carcinoma Cells, Zebrafish and Rats, *Sci. Rep.*, 2019, **9**, 4388.
- 52 J. Jiang, C. Yu, X. Guo, H. Zhang, S. Tian, K. Cai, Z. He and C. Sun, G Protein-Coupled Receptor GPR87 Promotes the Expansion of PDA Stem Cells through Activating JAK2/STAT3, *Mol. Ther.–Oncolytics*, 2020, **17**, 384–393.
- 53 S. J. Thomas, J. A. Snowden, M. P. Zeidler and S. J. Danson, The Role of JAK/STAT Signalling in the Pathogenesis, Prognosis and Treatment of Solid Tumours, *Br. J. Cancer*, 2015, **113**, 365–371.
- 54 J. Zhou, Y. Jiang, J. Zhao, H. Zhang, J. Fu, P. Luo, Y. Ma, D. Zou, H. Gao, J. Hu, Y. Zhang and Z. Jing, Dp44mT, an Iron Chelator, Suppresses Growth and Induces Apoptosis Via RORA-mediated NDRG2-IL6/JAK2/STAT3 Signaling in Glioma, *Cell. Oncol.*, 2020, **43**, 461–475.
- 55 G. L. Zhao, L. M. Yu, W. L. Gao, W. X. Duan, B. Jiang, X. D. Liu, B. Zhang, Z. H. Liu, M. E. Zhai, Z. X. Jin, S. Q. Yu and Y. Wang, Berberine Protects Rat Heart From Ischemia/Reperfusion Injury Via Activating JAK2/STAT3 Signaling and Attenuating Endoplasmic Reticulum Stress, *Acta Pharmacol. Sin.*, 2016, **37**, 354–367.
- 56 C. M. Taniguchi, B. Emanuelli and C. R. Kahn, Critical Nodes in Signalling Pathways: Insights Into Insulin Action, *Nat. Rev. Mol. Cell Biol.*, 2006, **7**, 85–96.
- 57 Y. Gao, M. Zhang, R. Zhang, L. You, T. Li and R. H. Liu, Whole Grain Brown Rice Extrudate Ameliorates the Symptoms of Diabetes by Activating the IRS1/PI3K/AKT Insulin Pathway in Db/Db Mice, *J. Agric. Food Chem.*, 2019, **67**, 11657–11664.
- 58 L. Rui, M. Yuan, D. Frantz, S. Shoelson and M. F. White, SOCS-1 and SOCS-3 Block Insulin Signaling by Ubiquitin-Mediated Degradation of IRS1 and IRS2, *J. Biol. Chem.*, 2002, **277**, 42394–42398.
- 59 A. L. Rodrigues, E. G. de Moura, M. C. F. Passos, I. H. Trevenzoli, E. P. S. da Coceição, I. T. Bonono, J. F. N. Neto and P. C. Lisboa, Postnatal Early Overfeeding Induces Hypothalamic Higher SOCS3 Expression and Lower STAT3 Activity in Adult Rats, *J. Nutr. Biochem.*, 2011, **22**, 109–117.
- 60 G. J. Morton and M. W. Schwartz, Leptin and the Central Nervous System Control of Glucose Metabolism, *Physiol. Rev.*, 2011, **91**, 389–411.
- 61 L. Cheng, Y. Yu, A. Szabo, Y. Wu, H. Wang, D. Camer and X. Huang, Palmitic Acid Induces Central Leptin Resistance and Impairs Hepatic Glucose and Lipid Metabolism in Male Mice, *J. Nutr. Biochem.*, 2015, **26**, 541–548.
- 62 C. E. Nunes-Xavier, O. Aurteneixe, L. Zaldumbide, R. López-Almaraz, A. Erramuzpe, J. M. Cortés, J. I. López and R. Pulido, Protein Tyrosine Phosphatase PTPN1 Modulates Cell Growth and Associates with Poor Outcome in Human Neuroblastoma, *Diagn. Pathol.*, 2019, **14**, 134.
- 63 J. Xu, L. Li, Z. Qian, J. Hong, S. Shen and W. Huang, Reduction of PTP1B by RNAi Upregulates the Activity of Insulin Controlled Fatty Acid Synthase Promoter, *Biochem. Biophys. Res. Commun.*, 2005, **329**, 538–543.
- 64 S. Legeay, P. Fautrat, J. B. Norman, G. Antonova, S. Kennard, T. Bruder-Nascimento, V. S. Patel, S. Faure and E. J. B. de



- Chantemèle, Selective Deficiency in Endothelial PTP1B Protects From Diabetes and Endoplasmic Reticulum Stress-Associated Endothelial Dysfunction Via Preventing Endothelial Cell Apoptosis, *Biomed. Pharmacother.*, 2020, **127**, 110200.
- 65 C. M. Rondinone, J. M. Trevillyan, J. Clampit, R. J. Gum, C. Berg, P. Kroeger, L. Frost, B. A. Zinker, R. Reilly, R. Ulrich, M. Butler, B. P. Monia, M. R. Jirousek and J. F. Waring, Protein Tyrosine Phosphatase 1B Reduction Regulates Adiposity and Expression of Genes Involved in Lipogenesis, *Diabetes*, 2002, **51**, 2405–2411.
- 66 A. Janus, E. Szahidewicz-Krupska, G. Mazur and A. Doroszko, Insulin Resistance and Endothelial Dysfunction Constitute a Common Therapeutic Target in Cardiometabolic Disorders, *Mediators Inflammation*, 2016, **2016**, 3634948.
- 67 D. Leto and A. R. Saltiel, Regulation of Glucose Transport by Insulin: Traffic Control of GLUT4, *Nat. Rev. Mol. Cell Biol.*, 2012, **13**, 383–396.
- 68 P. R. Shepherd and B. B. Kahn, Glucose Transporters and Insulin Action-Implications for Insulin Resistance and Diabetes Mellitus, *N. Engl. J. Med.*, 1999, **341**, 248–257.
- 69 S. Polsky and S. L. Ellis, Obesity, Insulin Resistance, and Type 1 Diabetes Mellitus, *Curr. Opin. Endocrinol. Diabetes Obes.*, 2015, **22**, 277.
- 70 Y. Gao, M. Zhang, R. Zhang, L. You, T. Li and R. H. Liu, Whole Grain Brown Rice Extrudate Ameliorates the Symptoms of Diabetes by Activating the IRS1/PI3K/AKT Insulin Pathway in Db/Db Mice, *J. Agric. Food Chem.*, 2019, **67**, 11657–11664.
- 71 E. S. Popravka, N. S. Linkova, S. V. Trofimova and V. K. Khavinson, HIF-1 as a Marker of Age-Related Diseases Associated with Tissue Hypoxia, *Biol. Bull.*, 2018, **8**, 497–508.
- 72 M. T. Coughlan, A. L. Mibus and J. M. Forbes, Oxidative Stress and Advanced Glycation in Diabetic Nephropathy, *Ann. N. Y. Acad. Sci.*, 2008, **1126**, 190–193.
- 73 L. Nordquist, M. Friederich-Persson, A. Fasching, P. Liss, K. Shoji, M. Nangaku, P. Hansell and F. Palm, Activation of Hypoxia-Inducible Factors Prevents Diabetic Nephropathy, *J. Am. Soc. Nephrol.*, 2015, **26**, 328.
- 74 C. Rosenberger, M. Khamaisi, Z. Abassi, V. Shilo, S. Zangen, M. Goldfarb, A. Shina, F. Zibertrest, K. Eckardt, S. Rosen and S. Heyman, Adaptation to Hypoxia in the Diabetic Rat Kidney, *Kidney Int.*, 2008, **73**, 34–42.
- 75 R. U. Ostrovskaya, S. V. Ivanov, T. A. Gudasheva and S. B. Seredenin, A Novel Dipeptide NGF Mimetic GK-2 Selectively Activating the PI3K/AKT Signaling Pathway Promotes the Survival of Pancreatic β -Cells in a Rat Model of Diabetes, *Acta Naturae*, 2019, **11**, 48–57.
- 76 X. Li, H. Liu, J. Wang, J. Qin, Z. Bai, B. Chi, W. Yan and X. Chen, Curcumol Induces Cell Cycle Arrest and Apoptosis by Inhibiting IGF-1R/PI3K/Akt Signaling Pathway in Human Nasopharyngeal Carcinoma CNE-2 Cells, *Phytother Res.*, 2018, **32**, 2214–2225.
- 77 J. F. Wojtaszewski, B. F. Hansen, B. Kiens and E. A. Richter, Insulin Signaling in Human Skeletal Muscle: Time Course and Effect of Exercise, *Diabetes*, 1997, **46**, 1775–1781.
- 78 L. R. Pearce, D. Komander and D. R. Alessi, The Nuts and Bolts of AGC Protein Kinases, *Nat. Rev. Mol. Cell Biol.*, 2010, **11**, 9–22.
- 79 G. Xi, X. Shen, C. Wai, M. White and D. Clemmons, Hyperglycemia Induces Vascular Smooth Muscle Cell Dedifferentiation by Suppressing Insulin Receptor Substrate-1-Mediated p53/KLF4 Complex Stabilization, *J. Biol. Chem.*, 2018, **294**, A118–A5398.
- 80 C. Kim, J. Lee, M. Kim and J. Hwang, Hypoglycemic Effect of Whole Grain Diet in C57BL/KsJ-db/db Mice by Activating PI3K/Akt and AMPK Pathways, *J. Food Sci. Biotechnol.*, 2019, **28**, 895–905.
- 81 W. Feng, H. Ao, S. Yue and C. Peng, Systems Pharmacology Reveals the Unique Mechanism Features of Shenzhu Capsule for Treatment of Ulcerative Colitis in Comparison with Synthetic Drugs, *Sci. Rep.*, 2018, **8**, 16160.

